

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

7,000

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Photosynthetic Response and Adaptation of Plants in Perspective of Global Climate Change

Mohammad Javad Ahmadi-Lahijani and Saeed Moori

Abstract

The intense agricultural and human being activities, especially after the industrialization era, have increased the CO₂ concentration, which led to changes in the global climate. Climate change and its consequences, that is, elevated CO₂, water stress, and extreme temperatures, have induced many biotic and abiotic stresses and have caused alterations in plant physiology, leading to a reduced photosynthetic capacity of plants. Photosynthesis is the most crucial biochemical process in plants that determines the final dry matter production and productivity of plants. The efficiency and status of the photosynthetic apparatus can be measured by the measurement of chlorophyll fluorescence. Measurements of chlorophyll fluorescence are easy, non-destructive, and quick, and it reflects changes in the general bioenergy status of a plant. Studies have indicated that abiotic stresses emerging from climate changes cause changes in the biological processes of plants and damage the internal structure of photosynthesis and control of the cellular process. Chlorophyll fluorescence, meanwhile, is an effective parameter and an indicator of photosynthetic status and its mechanisms under stressful conditions. Therefore, the photosynthetic changes and adaptation and the role of chlorophyll fluorescence in determining its status under climate change are discussed in this chapter.

Keywords: abiotic stress, chlorophyll fluorescence, drought, elevated CO₂, extreme temperatures, leaf physiology

1. Introduction

Food production is required to be increased by ~70% to feed the global population of 9 billion by 2050 [1], since the food demand, especially in developing countries, will be immensely enhanced. During the last 160,000 years, the concentration of atmospheric carbon dioxide has been varying between 170 and 300 $\mu\text{mol mol}^{-1}$. But with the beginning of the industrial revolution in Western Europe (between 1750 and 1800), the concentration of CO₂ increased from 280 to 385 $\mu\text{mol mol}^{-1}$ [2]. According to predictions, with the rapid increase in world population, consumption of fossil

fuels, industrial development, and deforestation, the concentration of carbon dioxide, which is $\sim 400 \mu\text{mol mol}^{-1}$, will reach 700 micromoles by the end of this century [3].

Climate change and global warming have been one of the most controversial issues in the recent decade. Intense agricultural and industrial activities since the industrial revolution have hastened the process of global warming. The chemistry of the climate has been changed by agricultural and human being activities and consequently, many abiotic and biotic stresses have emerged and negatively affected plants' physiology and biochemistry. Crops resistant to environmental stresses should be the focus of agricultural plant development under the increased global temperatures and climate changes.

Due to continuously increasing the greenhouse gases, such as CO_2 , in the atmosphere, climate change is happening rapidly. Climate change by increasing temperatures and reducing precipitations imposes abiotic stress exposure in many areas. Abiotic stresses, such as drought, salinity, cold, heat, UV radiation, and heavy metals, are the major limitations in agricultural products and adversely influence plant growth. It is estimated that abiotic stresses reduce crop yield by approximately 50% [4]. Drought, salinity, and extreme temperatures are among the most dreadful abiotic stresses in modern agriculture.

One of the most vital processes of plants that are affected by global climate change is photosynthesis. Photosynthesis is a vital biochemical process in plants that supplies the carbon and energy required for the biosynthesis of organic compounds and controls plant growth and development [5]. Photosynthesis is particularly sensitive to environmental constraints [6]. The environmental stresses adversely affect the photosynthetic capacity of plants. The increasing global population and climate change over the coming decades require enhanced photosynthetic efficiency to ensure food security. Thus, an understanding of the photosynthetic response and optimization under future climate uncertainties will be required for an improvement in crop production to meet future food requirements.

Chlorophyll fluorescence is one of the effective, non-destructive, and quick methods for evaluating the photochemical status of the plant photosynthetic system. Chlorophyll fluorescence is a useful parameter for the measurement of environmental

Plant species	Environmental conditions	Parameters	Reference
Potato (<i>Solanum tuberosum</i>)	Elevated CO_2	g_m , T_D , g_s ↓ A_n , C_i , R_D ↑	[10, 11]
Tomato (<i>Solanum lycopersicum</i> L.)	Elevated CO_2	A_n , V_{cmax} , J_{max} , f_v/f_m , ETR, $\text{NADP}^+/\text{NADPH}$ ↑ NPQ, RL ↓	[12]
<i>Fagus sylvatica</i>	Elevated CO_2	A_n , R_D ↑ g_s , V_{cmax} ↓	[13]
<i>Yucca</i> (<i>Y. brevifolia</i> and <i>Y. schidigera</i>)	Elevated CO_2	A_n , f_v/f_m , Φ_{PSII} ↑ g_s ↓	[14]
Cotton (<i>Gossypium hirsutum</i> L.)	Elevated CO_2	F_o' , F_m' , Φ_{CO_2} ↑ f_v/f_m' , qP , ETR, Φ_{PSII} , Φ_{PSII}/Φ_{CO_2} , ETR/ A_n ↓	[15]
Grape (<i>Vitis vinifera</i> L.)	Elevated CO_2	qP , Φ_{PSII} , ETR ↑ f_v/f_m , NPQ ↓	[16]
Oak (<i>Picea abies</i>) and (<i>Quercus petraea</i>)	Elevated CO_2	A_n , g_s , T_D , WUE ↑	[17]

Plant species	Environmental conditions	Parameters	Reference
Pea (<i>Pisum sativum</i> L.)	High temperatures	$A_n, g_s \downarrow$	[18]
Wheat (<i>Triticum aestivum</i>)	High temperatures	WUE $\downarrow$	[19]
Barley (<i>Hordeum vulgare</i> L.)	High temperatures	$f_v/f_m, \Phi_{PSII} \downarrow$	[20]
Tomato (<i>S. lycopersicum</i> L.)	High temperatures	ETR $\downarrow$	[21]
Alfalfa (<i>Medicago sativa</i>)	High temperatures	Chl $\downarrow$ $F_o, F_m \uparrow$	[22]
Tomato (<i>S. lycopersicum</i> L.)	High temperatures	$A_n, V_{c_{max}}, J_{max}, f_v/f_m, ETR, NADP^+ / NADPH \downarrow$ NPQ $\uparrow$	[12]
Lentil (<i>Lens culinaris</i>)	Low temperatures	$f_v'/f_m', f_q'/F_m' \downarrow$	[23]
<i>Salvia leriifolia</i> Benth, <i>Visia faba</i>	Low temperatures	$f_v'/f_m' \downarrow$	[24–26]
Faba bean (<i>Vicia faba</i> L.)	Low temperatures	$g_m, A_n, T_D, g_s, C_i, C_i:C_a \downarrow$	[27]
Chickpea (<i>Cicer arietinum</i> L.)	Low temperatures	$f_v'/f_m', f_q'/F_m' \downarrow$	[28, 29]
Barley (<i>H. vulgare</i> L.)	Low temperatures	$\Phi_{PSII}, ETR \downarrow$ NPQ $\uparrow$	[30]
Oats (<i>Avena sativa</i>)	Low temperatures	$f_v/f_m \downarrow$	[31]
Barley (<i>H. vulgare</i> L.)	Drought	Chl, $F_o, f_v/f_o, f_v/f_m, ETR \downarrow$	[32]
Maize (<i>Zea mays</i> L.)	Drought	Rubisco $\downarrow$	[33]
Black-eyed pea (<i>Vigna unguiculata</i>)	Drought	$A_n, f_v'/f_m' \downarrow$	[34]
Barley (<i>H. vulgare</i> L.)	Drought	NPQ $\uparrow$	[35]
Castor bean (<i>Ricinus communis</i>)	Drought	$A_n, C_i \downarrow$	[36]
Wheat (<i>T. aestivum</i>)	Drought	$g_m, A_n, T_D, g_s \downarrow$	[37]
Oak (<i>P. abies</i>) and (<i>Q. petraea</i>)	Drought	$A_n, g_s, T_D, WUE, V_C, J \downarrow$	[17]
Sweet corn (<i>Z. mays</i> L.)	Drought	$f_v/f_m \downarrow$	[38]

Increase ($\uparrow$), decrease ($\downarrow$).

Table 1.
Effect of climate changes induced stresses on photosynthetic and chlorophyll fluorescence parameters.

stress effects on photosynthetic apparatus and an effective indicator of photosynthesis limiting factors. The photochemical efficiency of photosystem II (PSII) is strongly influenced by the climate change consequences such as elevated CO₂, extreme temperatures, and water stress, and a reduction in leaf relative water content and the accumulation of carbohydrates in leaves decreases the quantum efficiency of PSII [7].

More food must be produced by global agriculture to sustain a growing human population in the twenty-first century [8]. Producing more food, however, is threatened by the climate change constraints that limit plant productivity [9]. Under natural conditions, plants are exposed to many adverse environmental stresses that disrupt the photosynthetic apparatus, causing a decrease in plant productivity and overall yield. In the present chapter, the impacts of changing climatic conditions on photosynthesis, with an emphasis on the main consequences of climate change, that is, elevated CO₂, extreme temperatures, and drought are discussed (**Table 1**).

2. Climate change consequences and photosynthetic response

2.1 Elevated CO₂

Carbon dioxide, like other important factors, such as light, water, and nutrients, is one of the determinant factors in plant production. Carbon dioxide is the key substrate for photosynthesis and the source of carbon for plants; however, high, or low CO₂ concentration diversely affects plant growth and productivity [39]. Carbon dioxide stimulates photosynthesis, inhibits photorespiration, and increases the efficiency of water and nitrogen use, which leads to more biomass production and changes in plant composition. Increasing CO₂ concentration by preventing photorespiration in C₃ plants increases the efficiency of photosynthesis because, in the current CO₂ concentration the carboxylation capacity of Ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) does not reach the saturation limit (Drake et al., 1997). The increase in growth and yield of crop species due to doubling the CO₂ concentration was primarily due to the faster photosynthetic rate and secondarily due to less photorespiration [40].

Photosynthesis of C₃ plants is not completely saturated at the current CO₂ concentration. Increasing CO₂ concentration stimulates the rate of photosynthesis and has a positive effect on the growth and performance of plants [41]. Idso and Idso [42] believe that by doubling the current CO₂ concentration, biomass production, and yield of plants will increase by one-third or more if other factors are not limiting. However, plant species differ in response to CO₂ concentration. Faster-growing species are more stimulated and produce more biomass than slow-growing species. Also, plants growing in better nutritional conditions respond more to increased CO₂ concentration than those that are exposed to nutritional stress [43]. Apart from the indirect effects of atmospheric elevated CO₂ concentration, CO₂ concentration directly affects C₃ plants if other factors are not limiting [44]. In research on potatoes in an open-growth chamber, it was found that the photosynthesis of plants grown under elevated CO₂ concentration (720 ppm) was 10 to 40% higher than those grown under ambient CO₂ concentration (400 ppm) [45]. In addition, leaf starch and sucrose content were higher in plants grown under CO₂ concentration conditions, especially in young leaves. This shows that the response of plants to the CO₂ concentration also depends on leaf age.

In general, increasing CO₂ concentration as a substrate for photosynthesis increases leaf area, biomass, and CO₂ fixation. The main reason for the increase in photosynthesis and subsequent increase in growth is the competitive effect of the Rubisco enzyme, which increases the carboxylation of this enzyme [46]. The results of the experiments showed that the rate of photosynthesis was significantly increased under elevated CO₂ concentration in two potato cultivars [10, 11, 47]. Chen and Setter [48]

reported that cell division in physiological sinks is an important factor in increasing the photosynthesis of C_3 plants under CO_2 concentration. Increasing CO_2 concentration to $720 \mu\text{mol mol}^{-1}$ increased cotton canopy photosynthesis by 40% [49]. Also, the increased CO_2 concentration delayed the aging of sugarcane leaves [50]. Elevated CO_2 concentration also increased wheat production [51].

Potato plant leaves showed an 80–100% increase in photosynthetic rate when exposed to elevated CO_2 concentration [52]. However, long-term growth under elevated CO_2 concentration conditions led to plant acclimation to this environment and a relative decrease in photosynthesis [53]. Sicher and Bunce [54] reported that this acclimation is reversible by shifting plants to lower CO_2 concentration. Sicher and Bunce [55] stated that the acclimation response to higher CO_2 concentration is mainly due to a decrease in Rubisco activity than a decrease in the amount of this enzyme. In contrast, Schapendonk [56] found that photosynthetic acclimation, under elevated CO_2 concentration, was accompanied by a decrease in Rubisco and concluded that the acclimation is a complex mechanism resulting from the negative feedback of source-sink disequilibrium induced by high CO_2 concentration. In a study on two model tree species—coniferous Norway spruce and broadleaved sessile oak, A_n was increased in oak saplings under elevated CO_2 concentration ($700 \mu\text{mol CO}_2 \text{ mol}^{-1}$), whereas in Norway spruce, A_{max} remained unchanged or slightly declined; indicating a down-regulation of photosynthesis. Such acclimation was associated with the acclimation of both J and V_c .

Transpiration rate and g_s were decreased with increasing CO_2 concentration, while WUE was increased [57]. Therefore, the beneficial effects of increased CO_2 concentration on yield may be due to changes in either A_n or WUE or both; on the other hand, the reduction of g_s can increase the temperature of the leaf, which further increases the speed of the developmental stages and shortens the grain filling period [58]. The increase in growth due to elevated CO_2 concentration has been attributed to the improvement of plant water relations or the increase of cell expansion [59]. An increase in C_i due to an increase in CO_2 concentration can trigger partial stomatal closure, although the process of how stomata respond to CO_2 signals remained uncertain [60].

An increase in CO_2 concentration accelerates aging in plants. One of the reasons for this is the effect of CO_2 on reducing g_s and increasing leaf temperature. Another reason is the increase in the demand for underground parts for nitrogen and the reduction of N supply to aerial organs [61]. Nitrogen redistribution from chlorophyll-binding proteins has been proposed as the main factor in chlorophyll degradation [62]. Chlorophyll is known as the first electron donor in the process of electron transfer and the photosynthesis apparatus and plays a fundamental role in absorbing light energy in the photosynthesis apparatus [63]. The results of various studies show that elevated CO_2 concentration causes a decrease [64, 65], an increase [66], or no change [52] in the chlorophyll content of potato leaves. Bindi [66] reported that the chlorophyll content of potato leaves under conditions of increased CO_2 concentration was on average 9.3% lower than that of plants under normal conditions.

Reducing g_s , oxidative stress, and decreasing the activity of Rubisco affect photosynthesis under environmental stresses [67]. In addition, PSI and PSII, ETR, and Chl biosynthesis are negatively influenced by abiotic stresses [68, 69]. The quantum efficiency of PSII is considered a quantitative indicator of electron transfer through PSII, which is related to the photochemical efficiency of PSII [69]. Non-photochemical quenching indicates how much excess energy is released as heat by the plant relative to linear electron transport. Under unfavorable conditions, *that is*, environmental stresses, more energy is required to be dissipated since qP is disrupted. Therefore, NPQ

is strongly enhanced when physiological sinks are few and leaf physiology and biochemistry are adversely affected by environmental stresses [70]. Working on tomato and grape plants showed that elevated CO₂ concentration decreased NPQ of leaves, while qP was enhanced, indicating that higher CO₂ concentration probably stimulates the photosynthetic efficiency and improves the photochemistry of leaves [12, 16].

There are different reports on the effect of elevated CO₂ concentration on chlorophyll fluorescence. Hao [71] stated that the increase in CO₂ concentration increased the rate of photosynthesis and $J_{\max}$ with an increase in f_v/f_m , the efficiency of photoreceptors, and the transfer energy of PSII reaction centers (RC). Also, qP was reduced under those conditions. On the other hand, Pérez [72] and Ge [73] reported reduced leaf Chl content and factors related to chlorophyll fluorescence, including the photochemical efficiency of PSII and the ETR due to an increase in CO₂ concentration. Taub [74] also reported that in most of the species in their study, the efficiency of photosystem II (f_v/f_m) was significantly higher in plants grown under elevated CO₂ concentration. They stated that this higher efficiency was due to both higher F_m and lower F_o fluorescence. The results of a study showed that elevated CO₂ concentration (800 mmol mol⁻¹) improved leaf A_n , $V_{c\max}$, $J_{\max}$, and f_v/f_m of tomato (*Solanum lycopersicum* L.) plants at a 24 h recovery [12]. Furthermore, the elevated CO₂ concentration also increased the absorption flux, trapped energy flux, ETR, energy dissipation per PSII cross-section, the concentration of NADP⁺ and ratio of NADP⁺/NADPH, and decreased photoinhibition, damage to PSs and ROS accumulation.

2.2 Extreme temperatures

Plants are exposed to frequent low and high-temperature stresses during their life [75]. Global warming induces temperature stress on plants and limits productivity and biomass production. Climate change is likely to increase extreme temperatures beyond the optimum temperatures for the growth of plants. Temperature above or below the optimal threshold disrupts plant cellular homeostasis, which further slows down plant growth, development, and metabolism [76]. The ideal temperature for plant growth and development is in the range of 10 to 35°C. Rising temperature to a specific point enhances plants to generate excess energy; however, heat stress adversely affects plant growth and diminishes the photosynthetic rate [77]. Elevated temperature increases respiration levels in plants. Raising the temperature from 15 to 40°C elevated the respiration rate and disturbed the morphological features of crop species [78].

Heat tolerance is directly related to the ability of plants to maintain the CO₂ assimilation rate. Stomatal conductance and transpiration rate are closely related to leaf temperature [79]. Stomatal conductance, substomatal CO₂ concentration, and leaf water status are affected by the temperature above the optimum levels for plant growth [80]. The concentration of substomatal CO₂ is altered at high temperatures upon stomatal closure and inhibits net photosynthesis [81]. Moreover, high temperatures directly affect the vapor pressure deficit that alters the plant's hydraulic conductance and water supply of the leaves [82]. Studies indicated that the net CO₂ assimilation rate in soybean decreased with an increase in temperature mainly due to the reduction in g_s and C_i and lower biomass accumulation [83]. A reduction in photosynthetic ET diminished ATP production and A_n under high temperatures [84]. A significant decrease in the photosynthetic electron transport chain, ATP production, and NADPH under high temperatures led to a decrease in photosynthesis [85].

The negative effect of heat stress on photosynthesis might be due to the reduced Rubisco content and activity [86]. The reduced Rubisco thermal stability decreases its activation under higher temperatures [87]. Rubisco is activated by the RA at an optimum temperature. The catalytic activity of Rubisco is stimulated by an increase in temperature, but the RA fluctuates in response to high temperature [87]. While Rubisco is stable even at 50°C, the activity of RA is decreased at temperatures beyond the optimum [88]. The first step in photosynthetic and photorespiration pathways is catalyzed by Rubisco. The carboxylation efficiency of Rubisco is decreased at high temperatures because of the temperature sensitivity of the RA protein. An elevation in temperature leads to the deactivation of the Rubisco enzyme by the generation of inhibitory compounds such as xylulose-1,5-bisphosphate. Also, the RA breakdown at high temperatures causes the Rubisco disruption [89]. The RA is the main enzyme in the CO₂ fixation process in plants, but at higher temperatures, it is not sufficiently able to keep the balance of the inactivation [90].

Chlorophyll pigments are important for light harvesting; however, temperature stress negatively affects their biosynthesis in plastids [91]. High temperatures degrade the chlorophyll molecule due to different enzymatic impairments; the first enzyme in pyrrole biosynthesis (5-aminolevulinate dehydratase (ALAD)) is negatively affected by high temperature [92]. The decreased chlorophyll biosynthesis in celery leaves at high-temperature stress was likely due to the mRNA down-regulation of 15 genes involved in chlorophyll biosynthesis [93].

Plant productivity is restricted by temperature stress in different ways [94]. The photosynthetic apparatus is the first site of inhibition and is highly sensitive to heat stress. High-temperature alter the reduction-oxidation capacity of PSII acceptors and reduce the photosynthetic electron transport (ET) efficiency of both photosystems [76]. The important components of photosynthetic apparatus are the PSI and PSII, CO₂ reduction pathways, photosynthetic pigments, and ETR and any impairment inhibits overall photosynthesis [92].

High temperatures increase the permeability of membranes, damage PSII subunits, and the manganese complex, and limit ET. The increased permeability of thylakoid membranes leads to peroxidation of membranes, membrane protein changes, the opening of ionic channels, redistribution of specific lipids in thylakoid membranes, and the formation of single-layered membranes [76, 92]. The oxygen-evolving complex of plants grown at high temperatures is partially damaged. Kalaji [6] found that low and high temperatures decreased the reduced PSII electron acceptors pool (mainly Q_A) in barley seedlings. The Φ_{PSII} and the qP were decreased at high temperatures in oak leaves [95].

Kalaji [7] believed that the PI_{ABS} is the most sensitive indicator of various stressors including extreme temperatures. Damage to thylakoid membranes and a decrease in the PSII activity can be the reason for decreased fluorescence in response to high-temperature stress [89]. PSII thermostability is often calculated with the use of fluorescence methods by determining the relationships between F_o and leaf temperature. The fast fluorescence kinetics (JIP-test parameters) can also use to determine the effects of critical temperatures, which are often affected by a much lower temperature than the F_o [7, 96].

One of the crucial factors in predicting future global warming is the response of photosynthesis to temperature. Plant CO₂ assimilation is impaired under environmental stress conditions, such as temperature, while light absorption remains unaffected. Excessive light energy absorption leads to the production of ROS and the photosynthetic machinery, mainly PSII, which is highly sensitive to photodamage, is severely

damaged. Although plants have various mechanisms to protect the PSII, photoinhibition occurs when the photodamage rate is exceeded the PSII repairment rate, leading to reduced photosynthetic efficiency [97].

High night temperature stress is increasing due to climate change, and it suppresses the net CO₂ assimilation rate in both C₃ and C₄ plants. The ratio of reduced plastoquinone (Q_B) to (Q_A) and the ratio of Q_A to RC is reduced under high night temperatures. Furthermore, f_v/f_m was decreased, and F_o was increased under high night temperatures [98]. High night temperature reduces qP , Φ_{PSII} , and ETR, increases NPQ, and inhibits the donation of electrons by the oxygen-evolving complex (OEC). Pan [12] observed that high temperature reduced tomato (*S. lycopersicum* L.) leaves photosynthesis by reducing the energy fluxes limitations, ET, and redox homeostasis. They observed that $V_{c_{max}}$, J_{max} , and f_v/f_m were diminished by high temperature (42°C for 24 h).

The saturation of fatty acids and membrane fluidity is induced by low temperatures, and it affects the efficiency of photosynthetic ET. Previous studies on various plant species elucidated that the leaf photosynthetic activity is affected by short-term or long-term high and low temperatures [7]. Plants by stimulating thermal energy dissipation and increasing the hydrophobic protein PsbS content, which participates in the thermal energy dissipation, try to reduce the generation of ROS and adapt to low temperatures [99]. Low temperatures inhibit sucrose synthesis, reduce photosynthetic ET, increase photoinhibition, and disturb the photophosphorylation process. Rapacz [100] found that mild frosts initially disturbed the energy transfer to the primary quinone electron acceptor of PSII, Q_A in wheat plants; however, lower temperatures, that is, freezing, may cease energy flow between the PSII RC, Chl, and Q_A, which these primary injuries could only be partially repaired. Consequently, further freezing hinders the ET between the PSII RCs and Q_A and the secondary damage may lead to PSII deactivation. They concluded that both primary and secondary freezing damages resulted in a decreased PI_{ABS}. Strauss [101] also observed that the PI_{ABS} was decreased at low temperatures in soybean plants. Working on faba bean (*Vicia faba* L.) landraces revealed that gas exchange variables are promising criteria for screening freezing-tolerant landraces at early growth stages [27]. The physiological, biochemical, and molecular modifications of chickpea (*Cicer arietinum* L.) seedlings were studied under freezing stress, and it was found that f_v'/f_m' and $t\Phi_{PSII}$ of the cold-tolerant genotype recovered faster compared to the cold-sensitive genotype [28, 29]. They found that f_v'/f_m' and Φ_{PSII} were significantly lower in freezing compared with higher temperatures. In a study on lentil (*Lens culinaris* Medik.) genotypes under freezing stress, Nabati [23] found that F_m' , f_v'/f_m' , and Φ_{PSII} were decreased at freezing temperatures. They concluded that the freezing-tolerant genotypes showed a high potential to restore PSII performance and survival rate.

2.3 Drought stress

Global climate change and lower availability of underground water induce a water crisis worldwide. The constant rise in the atmospheric global temperature induces frequent droughts around the world, which further impacts the biological systems [102]. Plants may experience different forms of abiotic stresses, such as drought during their life, which adversely affect plant growth, survival, and productivity [103]. Drought is a serious problem in arid and semiarid environments with precipitation deficiency [104].

Plant photosynthesis, growth, and yield are impaired by drought stress [105]. Photosynthesis is highly sensitive to drought stress and is the first-line process that is altered by drought stress. Lower photoassimilate production reduces leaf growth and

crop yield [37]. Impaired photosynthesis under water deficit relates to either stomatal or non-stomatal limitations. Plants enhance their tolerance levels to survive under such a harsh environment by adopting different strategies, such as stomata closure and osmotic adjustment [106]. Closure of stomata as the primary response of leaves to drought conditions prevents water loss and decreases Tr and increases WUE of plants [92]. The primary response of plants to drought stress is closing the stomata. CO_2 and water exchange in plants are regulated by stomatal openings. Although stomatal closure limits water loss, CO_2 absorption and transportation of non-structural carbon (NSC) are also hindered by stomatal closure, leading to carbon starvation which further affects further processes [107].

Nonstomatal limitations of photosynthesis might be due to lower synthesis and supply of Rubisco and/or other metabolic responses [108]. The proteins D_1 and D_2 can also be damaged by drought stress [109]. Since the PSII is quite resistant to water stress, the photochemical reactions may only be influenced by severe water stress [110]. Lauriano [111] found that changes in the values of chlorophyll fluorescence parameters in peanut leaves were more pronounced under severe drought. Decreased leaf CO_2 transport rate under prolonged and severe water stress reduces CO_2 concentration in chloroplasts, thus weakening photosynthesis. The decrease in the cells CO_2 concentration reduces the activity of sucrose phosphate synthase, nitrate reductase, and capacity for ribulose biphosphate (RuBP) regeneration, and deactivates Rubisco [49]. The chloroplast thylakoid membrane is degraded under water stress and adversely affects photosynthetic pigment and reduces the photosynthetic rate [112].

Water stress induces oxidative stress. Under water stress, a reduction in chloroplastic CO_2 concentration due to the stomatal closure leads to the impairment of the Calvin cycle and reduces the production of $NADP^+$, leading to excessive electron transport chain (ETC) reduction and directing the electrons to O_2 via Mehler reaction to form singlet O_2 , and consequently, ROS [113]. Under drought conditions, triplet chlorophyll stages ($^3Chl^*$) may be overproduced if too much energy is delivered to antenna complexes. This promotes singlet oxygen (1O_2) production, which is a highly reactive form of oxygen that can photo-oxidase chlorophyll (mainly P680) and cause peroxidation of membrane lipids [111]. Partial closure of the stomatal reduces CO_2 assimilation and might lead to an imbalance between PSII photochemical activity and NADPH demand, which in turn, the generation of ROS can be stimulated and lead to higher sensitivity to photodestruction. Under stressful conditions such as low water availability and high irradiance and temperature, photosynthetic efficiency decreases due to a probable high chronic photoinhibition [7].

Studies of the alterations in the chlorophyll fluorescence kinetics provide an in-depth understanding of the structure and functions of the photosynthetic apparatus, particularly PSII [114]. Drought can change the kinetics of chlorophyll fluorescence by affecting PSII. The photochemical efficiency of PSII is strongly influenced by the relative water content of the leaf. The reduction of photosynthesis and the accumulation of carbohydrates in the leaf decrease the quantum efficiency of PSII [7]. One of the consequences of drought is stomatal closure which reduces the heat exchange of leaves. High temperature affects PSII, photosynthetic ET, and ATP synthesis [7]. A decrease in f_v/f_m and yield are indicators of photoinhibition in plants under stressful conditions, indicating lower efficiency of photosynthetic conversion of PAR photon energy [108]. The f_v/f_m is decreased at advanced stages of stress. The f_v/f_m is directly related to chlorophyll activity in the PSs RC. Working on maize plants, Karvar [38] found that deficit irrigation decreased the f_v/f_m . A decrease in leaf Chl content was the likely reason for the diminished f_v/f_m . Carotenoids are non-enzymatic antioxidants

that prevent Chl photooxidation under stressful conditions [103]. The stability of carotene and xanthophyll cycle pigments significantly contributed to the protection mechanism of PSII RCs. Furthermore, the cyclic electrons flow around PSI significantly contributed to the dissipation of excess energy in some plant species under water stress [111].

The PSII Φ_{PSII} and ETR_{PSII} are also important parameters to measure drought stress effects on leaves, which provide estimation for both stomatal and non-stomatal effects of drought stress. However, the relative fluorescence decreases ratio (Rfd) proposed by Lichtenthaler [115] as a more sensitive parameter correlated with photosynthetic assimilation than the PSII Φ_{PSII} or ETR_{PSII} . In sunflower plants, it was observed that water potential (Ψ), g_s , A_n , Φ_{PSII} , f_v/f_m , and daily accumulation of total non-structural carbohydrates (TNC) was decreased under drought, but NPQ, malondialdehyde concentration (MDA), and soluble carbohydrates content was increased [116]. The PI_{ABS} was also positively correlated with the water availability for plants. Van Heerden [104] found that a higher water supply increased PI_{ABS} in *Augea capensis* and *Zygophyllum prismatocarpum*.

3. Conclusions

Increasing greenhouse gases emission have led to global warming and climate change worldwide. The global climate change consequences, *that is*, elevated CO_2 concentration, water stress, and extreme temperatures, are serious problems affecting the photosynthetic efficiency and adaptation of plants and adversely affecting agricultural yields. Studies suggest that most plants will be more stressed and less productive in the future in response to climate change. Climate change reduces photosynthetic capacity directly by damaging photosynthetic structures and processes. The changes and modifications of the photosynthetic machinery under different stressful conditions can be evaluated by the chlorophyll fluorescence analysis. Analyses of chlorophyll fluorescence seem to be a promising tool for breeding crops with improved tolerance under stressful conditions. Therefore, the application of chlorophyll fluorescence can be useful to identify which part of the photosynthetic apparatus is affected by the stress and it might help identify good-performing genes by chlorophyll fluorescence to be used in breeding programs.

Abbreviations

A_n	net assimilation rate
Chl	chlorophyll
C_i	sub-stomatal CO_2 concentration
ETR	electron transport rate
ETR/A_n	photorespiration
F_m	maximal fluorescence
F_o	minimal fluorescence
f_q'/F_m'	light-adapted operational efficiency of photosystem II
F_v'	light-adapted variable fluorescence
f_v'/f_m'	light-adapted maximum efficiency of photosystem II
g_m	mesophyll conductance
g_s	stomatal conductance

J	electron transport
$J_{\max}$	maximum ribulose-1,5-bisphosphate (RuBP) regeneration rate
NPQ	non-photochemical quenching
OEC	oxygen-evolving complex
PI_{ABS}	performance index
PSI	photosystems I
PSII	photosystems II
qP	photochemical quenching
RA	Rubisco activase
R_D	dark respiration
R_L	photorespiration rate
ROS	reactive oxygen species
T_r	transpiration rate
V_C	Rubisco carboxylation rate
$V_{C_{\max}}$	mximum carboxylation rate
WUE	water use efficiency
Φ_{CO_2}	quantum yield of CO_2 assimilation
Φ_{PSII}	effective quantum yield

Author details

Mohammad Javad Ahmadi-Lahijani^{1*} and Saeed Moori²

1 Ferdowsi University of Mashhad, Iran

2 Lorestan University, Iran

*Address all correspondence to: mjahmadi@um.ac.ir

IntechOpen

© 2023 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Hussain S, Liu T, Iqbal N, Brestic M, Pang T, Mumtaz M, et al. Effects of lignin, cellulose, hemicellulose, sucrose and monosaccharide carbohydrates on soybean physical stem strength and yield in intercropping. *Photochemical & Photobiological Sciences*. 2020;**19**(4):462-472
- [2] Schimel DS. Terrestrial ecosystems and the carbon cycle. *Global change biology*. 1995;**1**(1):77-91
- [3] Solomon S, Qin D, Manning M, Averyt K, Marquis M. *Climate Change 2007-the Physical Science Basis: Working Group I Contribution to the Fourth Assessment Report of the IPCC*. Switzerland: Cambridge University Press; 2007
- [4] Rodziewicz P, Swarcewicz B, Chmielewska K, Wojakowska A, Stobiecki M. Influence of abiotic stresses on plant proteome and metabolome changes. *Acta Physiologiae Plantarum*. 2014;**36**(1):1-19
- [5] Simkin AJ, López-Calcano PE, Raines CA. Feeding the world: Improving photosynthetic efficiency for sustainable crop production. *Journal of Experimental Botany*. 2019;**70**(4):1119-1140
- [6] Kalaji HM, Carpentier R, Allakhverdiev SI, Bosa K. Fluorescence parameters as early indicators of light stress in barley. *Journal of Photochemistry and Photobiology B: Biology*. 2012;**112**:1-6
- [7] Kalaji MH, Goltsev VN, Žuk-Gołaszewska K, Zivcak M, Brestic M. *Chlorophyll Fluorescence: Understanding Crop Performance—Basics and Applications*. United States: CRC Press; 2017. p. 237
- [8] Beddington JR, Asaduzzaman M, Clark ME, Fernández Bremauntz A, Guillou M, Howlett D, et al. What next for agriculture after Durban? *Science*. 2012;**335**(6066):289-290
- [9] Lobell DB, Burke MB, Tebaldi C, Mastrandrea MD, Falcon WP, Naylor RL. Prioritizing climate change adaptation needs for food security in 2030. *Science*. 2008;**319**(5863):607-610
- [10] Ahmadi-Lahijani MJ, Kafi M, Nezami A, Nabati J, Erwin JE. ABA and BAP improve the accumulation of carbohydrates and alter carbon allocation in potato plants at elevated CO₂. *Physiology and Molecular Biology of Plants*. 2021;**27**(2):313-325
- [11] Ahmadi-Lahijani MJ, Kafi M, Nezami A, Nabati J, Zare Mehrjerdi M, Shahkoomahally S, et al. Variations in assimilation rate, photoassimilate translocation, and cellular fine structure of potato cultivars (*solanum Tuberosum* L.) exposed to elevated CO₂. *Plant Physiology and Biochemistry: PPB*. 2018;**130**:303-313
- [12] Pan C, Ahammed GJ, Li X, Shi K. Elevated CO₂ improves photosynthesis under high temperature by attenuating the functional limitations to energy fluxes, electron transport and redox homeostasis in tomato leaves. *Frontiers in plant science*. 2018;**9**:1739
- [13] Urban O, Klem K, Holišová P, Šigut L, Šprtová M, Teslová-Navrátilová P, et al. Impact of elevated CO₂ concentration on dynamics of leaf photosynthesis in *Fagus sylvatica* is modulated by sky conditions. *Environmental Pollution*. 2014;**185**:271-280
- [14] Huxman TE, Hamerlynck EP, Loik ME, Smith S. Gas exchange and chlorophyll fluorescence responses of

three south-western yucca species to elevated CO₂ and high temperature. Plant, Cell & Environment. 1998;**21**(12):1275-1283

[15] Singh SK, Reddy VR. Combined effects of phosphorus nutrition and elevated carbon dioxide concentration on chlorophyll fluorescence, photosynthesis, and nutrient efficiency of cotton. Journal of Plant Nutrition and Soil Science. 2014;**177**(6):892-902

[16] Zhao X, Li W-F, Wang Y, Ma Z-H, Yang S-J, Zhou Q, et al. Elevated CO₂ concentration promotes photosynthesis of grape (*Vitis vinifera* L. cv. pinot noir') plantlet in vitro by regulating RbcS and Rca revealed by proteomic and transcriptomic profiles. BMC plant biology. 2019;**19**(1):1-16

[17] Ofori-Amanfo KK, Klem K, Veselá B, Holub P, Agyei T, Marek MV, et al. Interactive effect of elevated CO₂ and reduced summer precipitation on photosynthesis is species-specific: The case study with soil-planted Norway spruce and sessile oak in a mountainous forest plot. Forests. 2020;**12**(1):42

[18] Abdulmajeed AM, Derby SR, Strickland SK, Qaderi MM. Interactive effects of temperature and UVB radiation on methane emissions from different organs of pea plants grown in hydroponic system. Journal of Photochemistry and Photobiology B: Biology. 2017;**166**:193-201

[19] Sattar A, Sher A, Ijaz M, Ul-Allah S, Rizwan MS, Hussain M, et al. Terminal drought and heat stress alter physiological and biochemical attributes in flag leaf of bread wheat. PLoS One. 2020;**15**(5):e0232974

[20] Jedmowski C, Ashoub A, Momtaz O, Brüggemann W. Impact of drought, heat, and their combination on

chlorophyll fluorescence and yield of wild barley (*Hordeum spontaneum*). Journal of Botany. 2015;**2015**:1-9. DOI: 10.1155/2015/120868

[21] Camejo D, Rodríguez P, Morales MA, Dell'Amico JM, Torrecillas A, Alarcón JJ. High temperature effects on photosynthetic activity of two tomato cultivars with different heat susceptibility. Journal of Plant Physiology. 2005;**162**(3):281-289

[22] Wassie M, Zhang W, Zhang Q, Ji K, Chen L. Effect of heat stress on growth and physiological traits of alfalfa (*Medicago sativa* L.) and a comprehensive evaluation for heat tolerance. Agronomy. 2019;**9**(10):597

[23] Nabati J, Nezami A, Mirmiran SM, Hasanfard A, Ahmadi Lahijani MJ. The chlorophyll fluorescence parameters response of lentil (*Lens culinaris* Medik.) genotypes to freezing stress. Iranian journal of field. Crop Science. 2021;**53**(1):79-93

[24] Dashti M, Kafi M, Tavakoli H, Mirza M, Nezami A. Effects of freezing stress on Morpho-physiological indices and chlorophyll fluorescence of salvia leriifolia Benth. in seedling stage. Journal of Plant Research (Iranian Journal of Biology). 2016;**28**(5):962-973

[25] Nezami A, Khazaei H, Eshghizadeh H, Riahinia S. Evaluation of freezing temperature tolerance of lentil (*Lens culinaris* Medik.) genotypes with using chlorophyll fluorescence parameters. Agronomy Journal (Pajouhesh & Sazandegi). 2011;**99**:24-33

[26] Zhou R, Hyldgaard B, Yu X, Rosenqvist E, Ugarte RM, Yu S, et al. Phenotyping of faba beans (*Vicia faba* L.) under cold and heat stresses using chlorophyll fluorescence. Euphytica. 2018;**214**(4):1-13

- [27] Nabati J, Nezami A, Hasanfard A, Haghghat SZ. The trend of changes in chlorophyll fluorescence parameters in two *Vicia faba* ecotype during freezing stresses. Iranian journal pulses. Research. 2018;**9**(2):139-150
- [28] Karimzadeh Soureshjani H, Nezami A, Nabati J, Oskoueian E, Ahmadi-Lahijani MJ. The physiological, biochemical, and molecular modifications of chickpea (*Cicer arietinum* L.) seedlings under freezing stress. Journal of Plant Growth Regulation. 2022;**41**(3):1109-1124
- [29] Karimzadeh Soureshjani H, Nezami A, Nabati J, Oskueian E, Ahmadi-Lahijani MJ. Genetic variations in antioxidant content and chlorophyll fluorescence of chickpea (*Cicer arietinum* L.) genotypes exposed to freezing temperatures. Acta Physiologiae Plantarum. 2022;**44**(12):1-13
- [30] Dai F, Zhou M, Zhang G. The change of chlorophyll fluorescence parameters in winter barley during recovery after freezing shock and as affected by cold acclimation and irradiance. Plant Physiology and Biochemistry. 2007;**45**(12):915-921
- [31] Rizza F, Pagani D, Stanca A, Cattivelli L. Use of chlorophyll fluorescence to evaluate the cold acclimation and freezing tolerance of winter and spring oats. Plant breeding. 2001;**120**(5):389-396
- [32] Li R-h, Guo P-G, Michael B, Stefania G, Salvatore C. Evaluation of chlorophyll content and fluorescence parameters as indicators of drought tolerance in barley. Agricultural Sciences in China. 2006;**5**(10):751-757
- [33] Zhang R, Zhang X, Camberato J, Xue J. Photosynthetic performance of maize hybrids to drought stress. Russian Journal of Plant Physiology. 2015;**62**(6):788-796
- [34] Singh SK, Reddy KR. Regulation of photosynthesis, fluorescence, stomatal conductance and water-use efficiency of cowpea (*Vigna unguiculata* [L.] Walp.) under drought. Journal of Photochemistry and Photobiology B: Biology. 2011;**105**(1):40-50
- [35] Oukarroum A, Schansker G, Strasser RJ. Drought stress effects on photosystem I content and photosystem II thermotolerance analyzed using Chl a fluorescence kinetics in barley varieties differing in their drought tolerance. Physiologia Plantarum. 2009;**137**(2):188-199
- [36] Santos CM, Endres L, Ferreira VM, Silva JV, Rolim EV, Wanderley HC. Photosynthetic capacity and water use efficiency in *Ricinus communis* (L.) under drought stress in semi-humid and semi-arid areas. An. Acad. Brasil. Ciênc. 2017;**89**:3015-3029
- [37] Ahmadi-Lahijani MJ, Emam Y. Post-anthesis drought stress effects on photosynthesis rate and chlorophyll content of wheat genotypes. Journal of Plant Physiology and Breeding. 2016;**6**(1):35-52
- [38] Karvar M, Azari A, Rahimi A, Maddah-Hosseini S, Ahmadi-Lahijani MJ. Titanium dioxide nanoparticles (TiO₂-NPs) enhance drought tolerance and grain yield of sweet corn (*Zea mays* L.) under deficit irrigation regimes. Acta Physiologiae Plantarum. 2022;**44**(2):1-14
- [39] Sage RF, Coleman JR. Effects of low atmospheric CO₂ on plants: More than a thing of the past. Trends in plant science. 2001;**6**(1):18-24
- [40] Drake BG, González-Meler MA, Long SP. More efficient plants: A

consequence of rising atmospheric CO₂. Annual Review of Plant Biology. 1997;**48**:609-639

[41] Reddy AR, Rasineni GK, Raghavendra AS. The impact of global elevated CO₂ concentration on photosynthesis and plant productivity. Current Science. 2010;**99**(1):46-57

[42] Idso KE, Idso SB. Plant responses to atmospheric CO₂ enrichment in the face of environmental constraints: A review of the past 10 years' research. Agricultural and Forest Meteorology. 1994;**69**(3-4):153-203

[43] Poorter H. Do slow-growing species and nutrient-stressed plants respond relatively strongly to elevated CO₂? Global Change Biology. 1998;**4**(6):693-697

[44] Lawlor D, Mitchell R. The effects of increasing CO₂ on crop photosynthesis and productivity: A review of field studies. Plant, Cell & Environment. 1991;**14**(8):807-818

[45] Katny MAC, Hoffmann-Thoma G, Schrier AA, Fangmeier A, Jäger H-J, van Bel AJ. Increase of photosynthesis and starch in potato under elevated CO₂ is dependent on leaf age. Journal of Plant Physiology. 2005;**162**(4):429-438

[46] Lawlor DW, Mitchell RA. Crop ecosystem responses to climatic change: Wheat. Climate change and global crop productivity. 2000;**57**:80

[47] Ahmadi Lahijani MJ, Kafi M, Nezami A, Nabati J, Erwin J. Effect of CO₂ enrichment on gas exchanges, biochemical traits, and Minituber yield in potato (*Solanum tuberosum* L.) Cultivars. Journal of Agricultural Science and Technology. 2019;**21**(4):883-894

[48] Chen C-T, Setter TL. Response of potato dry matter assimilation and

partitioning to elevated CO₂ at various stages of tuber initiation and growth. Environmental and experimental botany. 2012;**80**:27-34

[49] Reddy KR, Zhao D. Interactive effects of elevated CO₂ and potassium deficiency on photosynthesis, growth, and biomass partitioning of cotton. Field Crops Research. 2005;**94**(2-3):201, 213

[50] Vu JC, Allen LH Jr. Growth at elevated CO₂ delays the adverse effects of drought stress on leaf photosynthesis of the C₄ sugarcane. Journal of Plant Physiology. 2009;**166**(2):107-116

[51] Högy P, Fangmeier A. Effects of elevated atmospheric CO₂ on grain quality of wheat. Journal of Cereal Science. 2008;**48**(3):580-591

[52] Donnelly A, Craigan J, Black CR, Colls JJ, Landon G. Elevated CO₂ increases biomass and tuber yield in potato even at high ozone concentrations. New Phytologist. 2001;**149**(2):265-274

[53] Vandermeiren K, Black C, Lawson T, Casanova M, Ojanperä K. Photosynthetic and stomatal responses of potatoes grown under elevated CO₂ and/or O₃—Results from the European CHIP-programme. European Journal of Agronomy. 2002;**17**(4):337-352

[54] Sicher RC, Bunce JA. Adjustments of net photosynthesis in *Solanum tuberosum* in response to reciprocal changes in ambient and elevated growth CO₂ partial pressures. Physiologia Plantarum. 2001;**112**(1):55-61

[55] Sicher RC, Bunce JA. Photosynthetic enhancement and conductance to water vapor of field-grown *Solanum tuberosum* (L.) in response to CO₂ enrichment. Photosynthesis Research. 1999;**62**(2):155-163

- [56] Schapendonk AH, van Oijen M, Dijkstra P, Pot CS, Jordi WJ, Stoopen GM. Effects of elevated CO₂ concentration on photosynthetic acclimation and productivity of two potato cultivars grown in open-top chambers. *Functional Plant Biology*. 2000;**27**(12):1119-1130
- [57] Donnelly A, Jones MB, Burke JI, Schnieders B. Elevated CO₂ provides protection from O₃ induced photosynthetic damage and chlorophyll loss in flag leaves of spring wheat (*Triticum aestivum* L., cv. 'Minaret'). *Agriculture, ecosystems & environment*. 2000;**80**(1-2):159-168
- [58] Amthor JS. Effects of atmospheric CO₂ concentration on wheat yield: Review of results from experiments using various approaches to control CO₂ concentration. *Field Crops Research*. 2001;**73**(1):1-34
- [59] Mateos-Naranjo E, Redondo-Gómez S, Álvarez R, Cambrollé J, Gandullo J, Figueroa ME. Synergic effect of salinity and CO₂ enrichment on growth and photosynthetic responses of the invasive cordgrass *Spartina densiflora*. *Journal of Experimental Botany*. 2010;**61**(6):1643-1654
- [60] Robredo A, Pérez-López U, de la Maza HS, González-Moro B, Lacuesta M, Mena-Petite A, et al. Elevated CO₂ alleviates the impact of drought on barley improving water status by lowering stomatal conductance and delaying its effects on photosynthesis. *Environmental and experimental botany*. 2007;**59**(3):252-263
- [61] Miglietta F, Magliulo V, Bindi M, Cerio L, Vaccari F, Loduca V, et al. Free air CO₂ enrichment of potato (*Solanum tuberosum* L.): Development, growth and yield. *Global change biology*. 1998;**4**(2):163-172
- [62] Matile P, Hortensteiner S, Thomas H, Krautler B. Chlorophyll breakdown in senescent leaves. *Plant Physiology*. 1996;**112**(4):1403
- [63] Murray JW. Photosynthesis: Light and life. *The Biochemist*. 2013;**35**(5):4-7
- [64] Bunce JA. Direct and acclimatory responses of stomatal conductance to elevated carbon dioxide in four herbaceous crop species in the field. *Global change biology*. 2001;**7**(3):323-331
- [65] Lawson T, Craigon J, Tulloch A-M, Black CR, Colls JJ, Landon G. Photosynthetic responses to elevated CO₂ and O₃ in field-grown potato (*Solanum tuberosum*). *Journal of Plant Physiology*. 2001;**158**(3):309-323
- [66] Bindi M, Hacour A, Vandermeiren K, Craigon J, Ojanperä K, Sellden G, et al. Chlorophyll concentration of potatoes grown under elevated carbon dioxide and/or ozone concentrations. *European Journal of Agronomy*. 2002;**17**(4):319-335
- [67] Kohli SK, Handa N, Sharma A, Kumar V, Kaur P, Bhardwaj R. Synergistic effect of 24-epibrassinolide and salicylic acid on photosynthetic efficiency and gene expression in *Brassica juncea* L. under Pb stress. *Turkish Journal of Biology*. 2017;**41**(6):943-953
- [68] Sharma A, Thakur S, Kumar V, Kanwar MK, Kesavan AK, Thukral AK, et al. Pre-sowing seed treatment with 24-epibrassinolide ameliorates pesticide stress in *Brassica juncea* L. through the modulation of stress markers. *Frontiers in plant science*. 2016;**7**:1569
- [69] Kalaji HM, Jajoo A, Oukarroum A, Brestic M, Zivcak M, Samborska IA, et al. Chlorophyll a fluorescence as a tool to monitor physiological status of plants under abiotic stress conditions. *Acta Physiologiae Plantarum*. 2016;**38**(4):1-11

- [70] Myers D, Thomas R, DeLucia E. Photosynthetic capacity of loblolly pine (*Pinus taeda* L.) trees during the first year of carbon dioxide enrichment in a forest ecosystem. *Plant, Cell & Environment*. 1999;22(5):473-481
- [71] Hao X, Li P, Feng Y, Han X, Gao J, Lin E, et al. Effects of fully open-air CO₂ concentration elevation on leaf photosynthesis and ultrastructure of *Isatis indigotica* fort. *PLoS One*. 2013;8(9):e74600
- [72] Pérez P, Morcuende R, del Molino IMN, Martínez-Carrasco R. Diurnal changes of rubisco in response to elevated CO₂, temperature and nitrogen in wheat grown under temperature gradient tunnels. *Environmental and experimental botany*. 2005;53(1):13-27
- [73] Ge Z-M, Zhou X, Kellomäki S, Wang K-Y, Peltola H, Martikainen P. Responses of leaf photosynthesis, pigments and chlorophyll fluorescence within canopy position in a boreal grass (*Phalaris arundinacea* L.) to elevated temperature and CO₂ under varying water regimes. *Photosynthetica*. 2011;49(2):172-184
- [74] Taub DR, Seemann JR, Coleman JS. Growth in elevated CO₂ protects photosynthesis against high-temperature damage. *Plant, Cell & Environment*. 2000;23(6):649-656
- [75] Mckersie BD, YaY L. *Oxidative Stress. Stress and Stress Coping in Cultivated Plants*. Switzerland: Springer Nature; 1994. pp. 15-54
- [76] Mathur S, Agrawal D, Jajoo A. Photosynthesis: Response to high temperature stress. *Journal of Photochemistry and Photobiology B: Biology*. 2014;137:116-126
- [77] Raza A, Razzaq A, Mehmood SS, Zou X, Zhang X, Lv Y, et al. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants*. 2019;8(2):34
- [78] Jan SA, Bibi N, Shinwari ZK, Rabbani MA, Ullah S, Qadir A, et al. Impact of salt, drought, heat and frost stresses on morpho-biochemical and physiological properties of brassica species: An updated review. *Journal of Rural Development and Agriculture*. 2017;2(1):1-10
- [79] Carmo-Silva AE, Gore MA, Andrade-Sanchez P, French AN, Hunsaker DJ, Salvucci ME. Decreased CO₂ availability and inactivation of rubisco limit photosynthesis in cotton plants under heat and drought stress in the field. *Environmental and Experimental Botany*. 2012;83:1-11
- [80] Greer DH, Weedon MM. Modelling photosynthetic responses to temperature of grapevine (*Vitis vinifera* cv. Semillon) leaves on vines grown in a hot climate. *Plant, Cell & Environment*. 2012;35(6):1050-1064
- [81] Hussain M, Khan TA, Yusuf M, Fariduddin Q. Silicon-mediated role of 24-epibrassinolide in wheat under high-temperature stress. *Environmental Science and Pollution Research*. 2019;26(17):17163-17172
- [82] Yang Z, Sinclair TR, Zhu M, Messina CD, Cooper M, Hammer GL. Temperature effect on transpiration response of maize plants to vapour pressure deficit. *Environmental and Experimental Botany*. 2012;78:157-162
- [83] Kumari VV, Roy A, Vijayan R, Banerjee P, Verma VC, Nalia A, et al. Drought and heat stress in cool-season food legumes in sub-tropical regions: Consequences, adaptation, and mitigation strategies. *Plants*. 2021;10(6):1038

- [84] Cen Y-P, Sage RF. The regulation of rubisco activity in response to variation in temperature and atmospheric CO₂ partial pressure in sweet potato. *Plant Physiology*. 2005;**139**(2):979-990
- [85] Wise R, Olson A, Schrader S, Sharkey T. Electron transport is the functional limitation of photosynthesis in field-grown Pima cotton plants at high temperature. *Plant, Cell & Environment*. 2004;**27**(6):717-724
- [86] Jajoo A, Allakhverdiev SI. High-temperature stress in plants: Consequences and strategies for protecting photosynthetic machinery. *Plant Stress Physiology*. 2017;**2017**:138-154
- [87] Salvucci ME, Crafts-Brandner SJ. Inhibition of photosynthesis by heat stress: The activation state of rubisco as a limiting factor in photosynthesis. *Physiologia Plantarum*. 2004;**120**(2):179-186
- [88] Yamori W, von Caemmerer S. Effect of rubisco activase deficiency on the temperature response of CO₂ assimilation rate and rubisco activation state: Insights from transgenic tobacco with reduced amounts of rubisco activase. *Plant Physiology*. 2009;**151**(4):2073-2082
- [89] Weng J-H, Lai M-F. Estimating heat tolerance among plant species by two chlorophyll fluorescence parameters. *Photosynthetica*. 2005;**43**(3):439-444
- [90] Yamori W, Hikosaka K, Way DA. Temperature response of photosynthesis in C₃, C₄, and CAM plants: Temperature acclimation and temperature adaptation. *Photosynthesis Research*. 2014;**119**(1):101-117
- [91] Efeoğlu B, Terzioğlu S. Photosynthetic responses of two wheat varieties to high temperature. *EurAsian Journal of BioSciences (elektronik)*. 2009;**3**:97-106. DOI: 10.5053/ejobios.2009.3.0.13
- [92] Ashraf M, Harris PJ. Photosynthesis under stressful environments: An overview. *Photosynthetica*. 2013;**51**(2):163-190
- [93] Huang W, Ma HY, Huang Y, Li Y, Wang GL, Jiang Q, et al. Comparative proteomic analysis provides novel insights into chlorophyll biosynthesis in celery under temperature stress. *Physiologia Plantarum*. 2017;**161**(4):468-485
- [94] Allakhverdiev SI, Los DA, Mohanty P, Nishiyama Y, Murata N. Glycinebetaine alleviates the inhibitory effect of moderate heat stress on the repair of photosystem II during photoinhibition. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*. 2007;**1767**(12):1363-1371
- [95] Haldimann P, Feller U. Inhibition of photosynthesis by high temperature in oak (*Quercus pubescens* L.) leaves grown under natural conditions closely correlates with a reversible heat-dependent reduction of the activation state of ribulose-1, 5-bisphosphate carboxylase/oxygenase. *Plant, Cell & Environment*. 2004;**27**(9):1169-1183
- [96] Srivastava A, Strasser RJ. How do land plants respond to stress temperature and stress light. *Archives des Sciences*. 1995;**48**:135-146
- [97] Muñoz P, Munné-Bosch S. Photo-oxidative stress during leaf, flower and fruit development. *Plant Physiology*. 2018;**176**(2):1004-1014
- [98] Brestic M, Zivcak M. PSII fluorescence techniques for measurement of drought and high temperature stress signal in crop plants: Protocols and applications. In: *Molecular Stress Physiology of Plants*. Switzerland: Springer Nature; 2013. pp. 87-131
- [99] Demmig-Adams B, Adams WW III, Barker DH, Logan BA,

- Bowling DR, Verhoeven AS. Using chlorophyll fluorescence to assess the fraction of absorbed light allocated to thermal dissipation of excess excitation. *Physiologia Plantarum*. 1996;**98**(2):253-264
- [100] Rapacz M. Chlorophyll a fluorescence transient during freezing and recovery in winter wheat. *Photosynthetica*. 2007;**45**(3):409-418
- [101] Strauss A, Krüger G, Strasser R, Van Heerden P. Ranking of dark chilling tolerance in soybean genotypes probed by the chlorophyll a fluorescence transient OJIP. *Environmental and Experimental Botany*. 2006;**56**(2):147-157
- [102] Parry ML, Canziani O, Palutikof J, Van der Linden P, Hanson C. *Climate Change 2007-Impacts, Adaptation and Vulnerability: Working Group II Contribution to the Fourth Assessment Report of the IPCC*. Switzerland: Cambridge University Press; 2007
- [103] Farooq M, Wahid A, Kobayashi N, Fujita D, Basra S. Plant drought stress: Effects, mechanisms and management. In: *Sustainable Agriculture*. Springer; 2009. pp. 153-188
- [104] Van Heerden P, Swanepoel J, Krüger G. Modulation of photosynthesis by drought in two desert scrub species exhibiting C₃-mode CO₂ assimilation. *Environmental and Experimental Botany*. Netherlands. 2007;**61**(2):124-136
- [105] Meng L-L, Song J-F, Wen J, Zhang J, Wei J-H. Effects of drought stress on fluorescence characteristics of photosystem II in leaves of *Plectranthus scutellarioides*. *Photosynthetica*. 2016;**54**(3):414-421
- [106] Sharma A, Shahzad B, Kumar V, Kohli SK, Sidhu GPS, Bali AS, et al. Phytohormones regulate accumulation of osmolytes under abiotic stress. *Biomolecules*. 2019;**9**(7):285
- [107] Sevanto S. Phloem transport and drought. *Journal of Experimental Botany*. 2014;**65**(7):1751-1759
- [108] Yin C, Berninger F, Li C. Photosynthetic responses of *Populus przewalski* subjected to drought stress. *Photosynthetica*. 2006;**44**(1):62-68
- [109] Oukarroum A, El Madidi S, Schansker G, Strasser RJ. Probing the responses of barley cultivars (*Hordeum vulgare* L.) by chlorophyll a fluorescence OLKJIP under drought stress and re-watering. *Environmental and Experimental Botany*. 2007;**60**(3):438-446
- [110] Souza R, Machado E, Silva J, Lagôa A, Silveira J. Photosynthetic gas exchange, chlorophyll fluorescence and some associated metabolic changes in cowpea (*Vigna unguiculata*) during water stress and recovery. *Environmental and Experimental Botany*. 2004;**51**(1):45-56
- [111] Lauriano J, Ramalho J, Lidon F. Mechanisms of energy dissipation in peanut under water stress. *Photosynthetica*. 2006;**44**(3):404-410
- [112] Bertoli DJ, Cannon SB, Froenicke L, Huang G, Farmer AD, Cannon EK, et al. The genome sequences of *Arachis duranensis* and *Arachis ipaensis*, the diploid ancestors of cultivated peanut. *Nature genetics*. 2016;**48**(4):438-446
- [113] Noctor G, Mhamdi A, Foyer CH. The roles of reactive oxygen metabolism in drought: Not so cut and dried. *Plant Physiology*. 2014;**164**(4):1636-1648
- [114] Longenberger PS, Smith C, Duke S, McMichael B. Evaluation of chlorophyll fluorescence as a tool for

the identification of drought tolerance
in upland cotton. *Euphytica*.
2009;**166**(1):25-33

[115] Lichtenthaler H, Buschmann C,
Knapp M. How to correctly determine
the different chlorophyll fluorescence
parameters and the chlorophyll
fluorescence decrease ratio RFd of
leaves with the PAM fluorometer.
Photosynthetica. 2005;**43**(3):379-393

[116] Correia MJ, Osório ML,
Osório J, Barrote I, Martins M, David MM.
Influence of transient shade periods on
the effects of drought on photosynthesis,
carbohydrate accumulation and lipid
peroxidation in sunflower leaves.
*Environmental and Experimental
Botany*. 2006;**58**(1-3):75-84