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Medicinal Plants Used by Indigenous Communities of Oaxaca, Mexico, to Treat Gastrointestinal Disorders

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Abstract

The use of medicinal plants for the treatment of gastrointestinal disorders and ethnodiseases such as diarrhea, stomachache, dysentery, “empacho” (blockage), and bile is a common strategy among indigenous communities. It is estimated that approximately 34% of medicinal plants are used to treat diseases of the digestive tract. In Mexico, gastrointestinal infections caused by bacteria, parasites, or viruses represent one of the main causes of death in children in rural populations. Our objective was to document the use of medicinal plants used by the indigenous groups of Oaxaca, Mexico, for the treatment of gastrointestinal disorders, based on previous studies, experiences, and field observations in indigenous communities and supplemented with bibliographic references. In Oaxaca, there are 16 indigenous groups, the largest being the speakers of the Zapoteco, Mixteco, Mazateco, Mixe, Chinanteco, Amuzgo, Tacuate, Chatino, and Cuicateco languages. In this review of the medicinal plants used for gastrointestinal disorders, 186 species were grouped into 147 genera and 71 botanical families, among which the largest number of species belonged to Asteraceae (29), Fabaceae (15), Euphorbiaceae (9), Solanaceae (9), and Lamiaceae (9). Different pharmacological studies showed potential for preventing microbial and fungal pathogens that cause gastrointestinal disease.

Keywords: ethnodiseases, bioculture, indigenous knowledge, plant diversity

1. Introduction

Medicinal plants are key elements of traditional medicine because they are part of the collection of knowledge and cultural heritage of sociocultural communities. Communities are often repositories and users of medicinal plants, and both rural and urban social groups with rural or indigenous origins have knowledge of these plants. Based on ethnobotanical, ethnomedicinal, and ethnopharmacological studies, the plants that are used by different healers and families of the Tarahumara, Yaqui, Chontal, Nahuatl, Mazahua, Otomi, Mixteco, Zapoteco, Mixe, Maya and Tzotzil groups, and all Mexican indigenous groups are symbolic [1].

In Mexico, gastrointestinal diseases are common in communities that are highly marginalized and poverty-stricken and are often transmitted through the fecal-oral route and the consumption of contaminated water and food. The infant population is the most vulnerable, both in terms of incidence and vulnerability [2]. In 2012, approximately 20% of the population of Chiapas, Guerrero, and Oaxaca did not have access to an adequate quality of water. This problem was exacerbated because only between 65 and 75% of houses have adequate waste management systems. Gastrointestinal diseases in children under 5 years of age directly affect mortality rates (19.2–19.4%) and mainly occur in marginalized communities [3]. These observations suggest that minimizing gastrointestinal diseases is dependent on the social environment of the community, access to clean water and food safety. As such, the symptoms and syndromes of diarrhea, stomachache, gastric atrophy, and enteric fever are associated with bacterial, fungal, parasitic, and rotavirus agents [2], and the frequency of these symptoms among children varies from community to community.

Medicinal plants are commonly used by communities for the treatment of gastrointestinal disorders through plant infusions, maceration, chewing, poultices, and different types of extracts (alcoholic or aqueous) [1]. Some studies have documented the traditional uses of medicinal plants by healers, their conservation, and their cultural importance in different indigenous groups [4–9]. Other studies have contributed ethnobotanical knowledge of medicinal plants [10–12]. The evaluation of phytochemical compounds with medicinal activity is also a subject of interest [13–16]. One aspect in which different authors have devoted a great amount of attention to is the antiparasitic and antimicrobial effects of medicinal plants for the treatment of diarrhea and other gastrointestinal disorders [17–25].

Research in natural products is often based on ethnobotanical information. One goal of ethnopharmacology is to improve the understanding of the pharmacological effects of plants on health, especially for indigenous communities that are highly marginalized and poverty-stricken. In this study, we examined plants used for the treatment of gastrointestinal disorders such as diarrhea, dysentery, and abdominal pain. The objective of this work was to document the use of medicinal plants used by the indigenous groups of Oaxaca, Mexico for the treatment of gastrointestinal disorders, based on previous unreported studies, experiences, and field observations in indigenous communities. Our results are supplemented with bibliographic reviews.

2. Sociocultural and ethnographic context of Oaxaca, Mexico

Mexico has an indigenous population of 12.25 million (10.1% of the total), of which 7.38 million speak an indigenous language [26]. It is estimated that there are 1.2 million speakers in Oaxaca over 3 years of age that speak an indigenous language. Oaxaca is the state with the largest indigenous population (32.2% of its total population) and has 245 municipalities where more than 40% of the population speaks indigenous languages [26–28]. However, in Mexico, 24.4 million people consider themselves indigenous [26, 29], a figure that is higher than the total number of inhabitants of several European countries and is of great importance in the preservation of culture, biodiversity, and biocultural heritage. Indigenous peoples are also associated with the use of medicinal plants.

Based on the vegetation, ecosystem, biome, and indigenous settlement maps, Toledo et al. [30] identified 26 indigenous regions in Mexico. These regions coincide with the zones of greatest biodiversity and pluriculturality or indigenous settlements and are also where protected natural areas are located. In Oaxaca, eight

indigenous regions were identified: Mixteca, Cañada, Papaloapan, Sierra Norte, Istmo, Costa, Sierra Sur, and Valles Centrales. Different indigenous communities can be found in each of these regions. The Mixteca region is dominated by the Mixteco, Triqui, Chocho and Nahuatl groups, the Cañada by the Cuicateco and Mazateco groups, and so on. The topography of the mountainous areas and valleys of Oaxaca is rugged, with contrasting climates ranging from humid temperate at 2000–3000 masl, to temperate to subtemperate in intermediate zones from 1000 to 2000 masl, to tropical and subtropical regions (<1000 masl) [31]. Thus, indigenous communities are located in various climates, altitudes and vegetation, and consequently have access to and knowledge of different medicinal plants. We should note that among Oaxaca indigenous regions, the differences are not extreme in terms of climatic conditions, flora, fauna, topography, crops, handicrafts, and the use of traditional medicinal plants (**Table 1**); however, there are sociocultural differences in the knowledge associated with the use of plants for treating ethnodiseases and diseases with clinical diagnoses.

Among indigenous communities from distant ethnic regions and origins, and occupying different geographical territories, the use of plants for the treatment of gastrointestinal disorders is different, even though the symbolic or cosmological meanings may be the same. Thus, there is similar understanding of how plants are

Indigenous region (n ¹)	Main indigenous groups	Climate and precipitations	Altitude (m)
Mixteca (155)	Mixtecos, Triquis, Nahuas	Tropical subhumid, semitropical subhumid to temperate subhumid, annual precipitation from 550 to 2177 mm. Includes a semi-dry region	1200–2800
Cañada (45)	Mazatecos, Cuicatecos, Ixcatecos, Nahuas	Semidesert with variations of very warm, semiwarm and temperate in high areas, annual precipitation from 372.8 to 643.7 mm	1180–2700
Papaloapan (20)	Chinantecos, Zapotecos, Mazatecos, Mixes	Tropical humid, semitropical humid to humid temperate, and precipitation from 2000 to 4500 mm. The rainiest region of Oaxaca	0–2000
Sierra Norte (68)	Zapotecos de la Sierra, Mixes, Chinantecos, Mazatecos, Cuicatecos	Tropical subhumid, semitropical subhumid to temperate subhumid, annual precipitation from 1000 to 3000 mm	1000–3200
Valles Centrales (121)	Zapotecos de Valles, Mixtecos, Mixes	Subtropical to semidry, annual precipitation from 600 to 800 mm	1000–2000
Istmo (41)	Zapotecos, Huaves, Zoques, Chontales	Tropical subhumid to semitropical subhumid, annual precipitation from 800 to 2500 mm.	0–1600
Sierra Sur (70)	Zapotecos de la Sierra Sur, Amuzgos, Mixtecos, Chatinos, Triquis	Tropical subhumid, semitropical subhumid to humid temperate, annual precipitation from 800 to 2000 mm	1000–2600
Costa (50)	Zapotecos de la Costa, Mixtecos, Amuzgos, Chatinos, Chontales	Tropical subhumid, semitropical subhumid to temperate, delimited region by Sierra Madre del Sur, annual precipitation from 731.9 to 2050 mm	0–2000

¹Number of municipalities; sources: Arellanes et al. [31].

Table 1.
Ecogeographic descriptions of eight indigenous regions of Oaxaca, Mexico.

used to treat stomach, intestinal, diarrhea, or psychosomatic diseases (e.g., “susto” (fright), stomachaches, “mal aire” (bad air) with vomiting, and “empacho” (blockage), but the species of plant used, form of use, and application differ significantly between ethnic groups. In severe cases of illness, indigenous peoples resort to the community healers or alternate among medical professionals, pharmaceutical medicine, and the use of medicinal plants recommended by the family or healers [32, 33].

3. Relationship between indigenous groups and endemic medicinal plants used for gastrointestinal disorders

Among rural and indigenous communities, the concept of health and disease is holistic. As such, the human body develops its organic and metabolic functions without physical deterioration and includes mental, mystical-spiritual, and psychological aspects (symbolic reality). Social, cultural, and ethnic concepts are implicit in the perceptions of health and disease, and based on these concepts, the healing-therapeutic benefits of medicinal plants are sought. From the indigenous perspective, the causal explanations of illness are complex. The body suffers symptomatic imbalances resulting from physical, emotional, or mental deterioration, which are consequences of “susto” (fright), anger, interpersonal conflicts, cold currents, harmful foods (cold, hot, types of meat, cravings, etc.), sexual relations, witchcraft, etc. [34–38]. Consequently, the factors that produce imbalances in the body, in this case stomach problems, are to be avoided.

The knowledge and use of medicinal plants in indigenous communities is changing due to the introduction of pharmaceutical products recommended by health professionals. Giovannini et al. [39] note two trends in support of this fact: the complete displacement of medicinal plants by pharmaceutical products and the coexistence between clinical medicine and the use of medicinal plants. Thus far, local indigenous knowledge for the treatment of cardiovascular diseases, cancers, diabetes, and other prevalent diseases has been disseminated, which is why patient medicine is used. However, there are some examples of healing via the use of medicinal plants. In cases of chronic diarrhea caused by pathogens, the use of plants has been beneficial, such as plants used to treat diarrhea caused by *Salmonella* spp. [24]. In Oaxaca, acute diarrhea and acute respiratory infections are the main causes of mortality in children under 5 years of age [40].

The prevalence among Oaxacan children of microbial or parasitic gastrointestinal diseases, which cause acute diarrhea and other problems, has forced communities to resort to medicinal plants. Consequently, under these circumstances, there is a set of common species among indigenous communities that are used similarly, provided that their ecological conditions are also similar. It should also be noted that knowledge may be treated differently between different ethnic communities, especially when faced with common health problems. For example, the Mixe, Mixteco, and Zapoteco groups of the Sierras, located in temperate regions, commonly use *Chenopodium graveolens*, *Lantana achyranthifolia*, *Baccharis salicifolia*, and *Miconia mexicana*, although the local names used by each region are different. Additionally, the Chinanteco groups and Zapoteco of the Istmo use similar plant species (Table 2).

3.1 Healers and medicinal plants

Within Oaxacan indigenous groups, the healers play essential roles as therapists or ethnopractitioners and are not referred to as shamans, witch doctors, or herbalists. Here, we only refer to the cures or treatments using plants recommended by healers for the treatment of gastrointestinal disorders. The indigenous healers have

Indigenous group (climate at the communities)	Species	Spanish name	References
Mixtecos, Mixe, Zapotecos de las Sierras (temperate)	<i>Chenopodium graveolens</i> Willd.	Epazote de zorrillo	[14, 17]
Mixtecos, Mixe, Zapotecos de las Sierras (temperate)	<i>Lantana achyranthifolia</i> Desf.	Cinco negritos, Hierba mariposa	[8, 16]
Mixtecos, Mixe, Zapotecos de las Sierras (temperate)	<i>Baccharis salicifolia</i> (Ruiz & Pavón) Pers.	Jara, cacho de venado	[1, 4, 5, 13]
Mixtecos, Mixe, Zapotecos de las Sierras (temperate)	<i>Miconia mexicana</i> (Bonpl.) Naudin	Itswa	[21]
Mixe, Mixtecos (temperate)	<i>Loeselia mexicana</i> (Lam.) Brand.	Espinosilla	[21, 41]
Chinantecos, Zapotecos del Istmo (tropical and subtropical)	<i>Gomphrena serrata</i> L.	Cabezona, amor seco, amor de soltero	[1, 42, 43]
Chinantecos, Zapotecos del Istmo (tropical and subtropical)	<i>Byrsonima crassifolia</i> (L.) Kunth	Nanche amarillo (corteza)	[6, 14, 24, 41]
Chinantecos, Zapotecos del Istmo (tropical and subtropical)	<i>Cuphea pinetorum</i> Benth.	Cenicilla, Hierba de gallina	[18, 25]
Chinantecos, Mixe, Zapotecos del Istmo (tropical and subtropical)	<i>Gouania lupuloides</i> (L.) Urb.	Bejuco de reuma	[17, 44, 45]

Table 2.
Common plants used by indigenous groups for the treatment of gastrointestinal disorders in Oaxaca, Mexico.

social recognition in the communities as persons of knowledge, skills, and healing practices. These healers have the facilities to explain the physical and nonphysical causes of diseases through a symbolic, verbal and corporal language, and resort to deities to exercise healing techniques. The body of knowledge is a legacy inherited or acquired from their ethnic medical culture and becomes a depository with the capacity to incorporate medical experiences regarding the descriptions of diseases, anatomy, and physiology of the human body [1, 34, 38, 46, 47].

The healer’s diagnosis is based on the spirit, soul, and body, which are closely interconnected, to define health and disease. In Oaxacan communities, Mixe and Mazateco healers use psychotropic or hallucinogenic plants and mushrooms for successful diagnoses and healing. These hallucinogens are used as a mechanism to access supernatural forces and as interlocutors to ward off malignant agents [34, 46, 47]. The remedies or treatments recommended by healers for gastrointestinal problems vary from diets, teas, maceration or poultices of medicinal plants, changes in eating habits such as avoiding foods considered “hot” or “cold,” avoiding animals or people, sun exposure, or environmental changes in temperature, such as avoiding cold water, etc.

María Sabina was a well-known healer among the Mazateco Indians of the Sierra de Huautla de Jiménez in Oaxaca and throughout Mexico. After her death, she was recognized as a symbolic character of the Mexican healers. She used mushrooms (e.g., *Psilocybe zapotecorum* and *Psilocybe mexicana*) and hallucinogenic plants (e.g., *Salvia divinorum*) to obtain divine powers both to diagnose and to restore the health of the patient. In the case of gastrointestinal diseases, all recommendations are preventive/curative and are accompanied by psychospiritual rituals of protection, reintegration, and cleanliness of the soul. She is known to have said, “The health services provided by healers and Western medicine should not be lucrative,” [48, 49].

3.2 Plants used in indigenous communities for the treatment of gastrointestinal disorders

Medicinal plants are essential natural resources in the indigenous communities of Oaxaca, Mexico; they are easily accessible and there is a high diversity of species and forms of use for the treatment of diseases of the digestive tract. The region has the greatest level of diversity and endemism of species of phytotherapeutic use [50]. Based on a bibliographical compilation and field notes of visits to the indigenous communities of Oaxaca, a short list of medicinal plants used for gastrointestinal disorders was obtained (**Table 1A**). The compilation consists of 71 botanical families, among which the greatest number of species is in the families Asteraceae (29), Fabaceae (15), Euphorbiaceae (9), Solanaceae (9), Lamiaceae (9), Verbenaceae (6), Myrtaceae (5), Malvaceae (5), and Fagaceae (5). These families included 147 genera and 186 endemic species [50]. The genera used most frequently were *Croton* (5), *Quercus* (4), *Piper* (4), *Psidium* (3), *Ocimum* (3), and *Tagetes* (3). The medicinal species introduced to Mexico were excluded from this list, despite being widely used by Oaxacan indigenous groups.

Among indigenous communities, plants are grouped according to gastrointestinal physiological alterations, among which the most common refer to diarrhea (112 plants), stomach pain (90 plants), and dysentery (79 plants). Acute diarrhea is a symptom of gastrointestinal tract infection, which is commonly caused by pathogenic bacteria (e.g., *Escherichia coli*, *Salmonella* spp., *Vibrio cholerae*, *Clostridium perfringens*, *Bacillus cereus*, *Staphylococcus aureus*, *Vibrio parahaemolyticus*, *Campylobacter jejuni*, *Campylobacter coli*, *Shigella* spp., and *Aeromonas* spp.), viruses (rotavirus, adenovirus, enterovirus, and norovirus), or parasites (*Giardia lamblia* and *Entamoeba histolytica*) and is frequently accompanied by abdominal pain, stomach pain, fever, and vomiting [51]. For the treatment of vomiting, constipation and parasites, between 28 and 31 plant species are used. For the condition known locally as “empacho” (gastrointestinal disease) as well as for indigestion, ulcers and gastritis, between 17 and 39 species are utilized (**Table 1A**). It is important to note that we refer to the local descriptions of symptoms and the disease names that are used by Oaxacan indigenous communities.

The indigenous knowledge of medicinal plants includes the different phases of use, correct identification of the species (although they do not have systematic botanical studies), wild or cultivated origin, collection time (morning, noon, afternoon, or night), plant part to be used, and processing required for use. Each of these elements affects the effectiveness of the plant [8, 17, 51]. The leaves, stems and flowers are the most used parts, and the bark, roots and seeds are used less often (**Table 1A**). The form of use or extraction preparation ranges from infusion and cooking in water, maceration in ethyl alcohol (*Dorstenia drakena* L. and *Barkleyanthus salicifolius* (Kunth) H. Rob. & Brettell) or cane alcohol (*Saccharum officinarum* L.) and consumed as tea (oral), or as a topical application between the stomach and intestines. Additionally, the fresh crushed leaves or leaves macerated in ethyl alcohol can be ingested or applied topically (rubbed onto the affected part). Plants for treating dysentery are used as purgatives or for treating constipation as rectal washes and include *Eryngium foetidum* L., *Capraria biflora* L., *Prosopis laevigata*, and *Solanum rostratum* Dunal (**Table 1A**).

Bark is often cooked or used as an infusion in hot or cold water; it is a common treatment for diarrhea, dysentery and related symptoms. The bark of *Semialarium mexicanum* (Miers) Mennega, *Hymenaea courbaril* L., *Quercus oleoides*, *Hintonia latiflora*, and *Guaiacum coulteri* is used to treat ulcers or gastritis, and the bark of *Amphipterygium adstringens* is used to treat stomach cancer. The roots of some plants have shown to be beneficial to treat diarrhea, stomach pain, and intestinal

infection. In addition, the use of fruits and seeds to treat diarrhea, dysentery, constipation, and “empacho” is mentioned. For example, the fruits and seeds of the chili pepper (*Capsicum annuum* L.) are used for the treatment of dyspepsia (inflammation of the digestive tract), diarrhea, and dysentery. An infusion of guava leaves and roots (*Psidium* spp.) is used to treat diarrhea, dysentery, stomach pain, flatulence, and vomiting. The peel of the fruits of *Curatella americana* L. and *Persea americana* Mill. and the stigmas of the corn flower (*Zea mays* L.) are made into an infusion to treat stomachaches. The seeds of *Carica papaya* L. are used to treat diarrhea, vomiting, fever, intestinal inflammation, and parasites (**Table 1A**).

“Empacho” is a digestive ethnodisease recognized by traditional Mexican people. This disorder primarily occurs in children and does not correspond to a specific clinical diagnosis, but is culturally recognized in all Mexican rural communities. Empacho is characterized by discomfort caused by the intake of food that is difficult to unfold, and the healer or mother of the child indicates that food is “stuck” to the stomach or intestine. Symptoms include abdominal pain, lack of appetite, diarrhea, flatulence, and vomiting. The treatment includes the use of ash- or salt-containing oral plant infusions, purgatives, and massages [52]. Some traditional treatments recommend infusions made from the bark and/or leaves of *Guazuma ulmifolia* Lam. complemented with spoonful of castor oil and massages (sobadas) that “release the empacho” [53].

Another ethnodisease associated with gastrointestinal disorders is “bile or bile leakage.” The healer and adults report “pounding” (vibrations) from an area near the stomach, which is accompanied by pain in the esophagus. Symptoms include loss of appetite, headache, feeling of “bitterness” in the mouth upon waking up, and fatigue. *Pseudognaphalium attenuatum* DC., *Agastache mexicana* spp. *Xolocotziana*, *Hintonia latiflora*, *Loeselia mexicana*, *Tecoma stans*, *Zornia thymifolia*, *Anoda cristata*, *Oenothera rosea*, *Verbena litoralis*, and *Calea ternifolia* var. *Ternifolia* are used to treat bile (**Table 1A**). For gastrointestinal intestinal disorders in children, *Psidium guajava*, *Byrsonima crassifolia*, and *Quercus* spp. are used [53]. Mixe communities use fruit and bark infusions of *G. ulmifolia* to treat diarrhea and hemorrhages [14], whereas *Cestrum nocturnum* is used among the Zapotecas from the southern highlands and the Chinantecos of Oaxaca [53, 54].

Several studies have shown the importance of flavonoids, tannins, terpenoids, and alkaloids present in medicinal plants used for the treatment of gastrointestinal diseases. Other mechanisms of action include antispasmodic activity, delaying of intestinal transit, suppression of intestinal motility, stimulation of the adsorption of water, and a reduction in the secretion of electrolytes [20]. The compounds most frequently reported are terpenoids (monoterpenes, sesquiterpenes, di-, and triterpenes), flavonoids (flavones, flavonols), tannins (condensed and hydrolysable), and volatile compounds, which are derived from the essential oils in aromatic plants.

Root extracts of *Tagetes erecta* have shown to have high efficacy against Gram-positive bacteria (*Staphylococcus aureus*, *Bacillus subtilis*, and *Micrococcus luteus*), Gram-negative bacteria (*Escherichia coli* and *Pseudomonas aeruginosa*), and fungi (*Candida albicans* and *Aspergillus niger*). Flowers of *T. erecta* have been shown to contain thiophene derivatives, terpenoids, alkaloids, flavonoids, and carotenoids [55]. Similarly, methanol extracts of *Tagetes lucida* yielded coumarins with high antimicrobial activity against Gram-positive and Gram-negative bacterial strains, with greater inhibition of *V. cholerae* as well as antifungal activity. The antifungal activity of *T. lucida* results from the presence of dimethoxy: 6,7-dimethoxy-4-methylcoumarin and scoparone (6,7-dimethoxycoumarin) [56] and is effective against *Helicobacter pylori* [57]. In traditional Oaxacan medicine, the anthelmintic effects of *Chenopodium ambrosioides* (epazote) along with the epimastigotes of *Trypanosoma cruzi* have demonstrated efficacy against *Entamoeba histolytica* infections due to the effect of limonene [58].

4. Antipathogenic efficacy of medicinal plants

A natural question about the use of medicinal plants for treating gastrointestinal disorders is: how effective are the uses of plants in controlling or preventing infections by pathogenic agents? Several *in vitro* studies have been conducted that aimed to evaluate the biological activity of plant extracts against enteropathogenic bacteria (*Escherichia coli*, *Shigella sonnei*, *Shigella flexneri*, *Salmonella*, and *Campylobacter*) and parasites (*Giardia lamblia* and *Entamoeba histolytica*) responsible for diarrhea, dysentery and/or gastric disorders [59–61]. Several chemical compounds (alkaloids, tannins, flavonoids, and terpenes) responsible for the pharmacological effects of these plants have been identified and isolated [62, 63]. Examples of biological activity of native plants of Oaxaca, Mexico, used for the prevention and control of gastrointestinal disorders are listed in **Table 3**.

The experiences in the communities of Oaxaca indicate that the roots and aerial parts of *Geranium mexicanum* Kunth and the flowers of *Chiranthodendron pentadactylon* are effective to treat diarrhea, dysentery, and stomach pain (**Table 3**). The methanolic or aqueous extracts of these plants showed high antibacterial activity against *Shigella flexneri* and *Shigella sonnei* at minimum inhibitory concentrations of 8 mg/mL. Both extracts can be considered an alternative strategy to treat enteric pathogens resistant to common drugs [60]. In biological experiments using mice, it was demonstrated that the molecule (–)-epicatechin, which is isolated from *Geranium mexicanum*, has high antiprotozoal activity (*Giardia lamblia*) in concentrations of 0.072 $\mu\text{mol/kg}$, which is the dose required to kill 50% of microorganisms [59].

Calzada et al. [69], found that (–)-epicatechin and tiliroside, isolated from extracts of *C. pentadactylon* flowers, were effective against *E. histolytica*, *G. lamblia*, *E. coli*, *S. sonnei*, *S. flexneri*, *Salmonella* sp., and *Vibrio cholerae*. In another work, Calzada et al. [11] determined that root extracts of *G. mexicanum* had a greater hyperperistaltic effect than the extracts of *Lygodium venustum*, *Chenopodium ambrosoides*, and *C. pentadactylon*, which was similar to the effect of the drug loperamide, which is used to control acute and constant diarrhea associated with intestinal inflammation. In experiments with mice, the methanolic extracts of *G. mexicanum*, *Bocconia frutescens*, and *C. pentadactylon* in a concentration of 300 mg/kg of body weight had high inhibitory or intestinal antisecretory activity against *V. cholerae*. The activity of these extracts exceeds the effect of the drug loperamide [72]. A similar effect was determined with extracts of *C. pentadactylon* flowers [73]. *B. frutescens* also showed inhibitory activity against *S. aureus* and *E. coli* [60]. These results show that *G. mexicanum*, *C. pentadactylon*, and *B. frutescens* have medicinal properties and are effective for the treatment of diarrhea, among other gastrointestinal problems (**Table 3**).

The methanolic extracts *Ocimum basilicum* and *Artemisia ludoviciana* have inhibitory effects against *V. cholerae*, and the minimum bactericidal concentration varies from 0.5 to 3.0 mg/mL. The extracts have a degradative effect on the cellular membranes of *V. cholerae*, which increases membrane permeability, decreases cytoplasmic pH, hyperpolarizes the membrane, decreases the cellular ATP concentration, and consequently causes cell death [65]. This result indicates a chemical modification of the growth medium and induction of the death of the pathogen. For example, Hussain et al. [62] reported that basil contains essential oils (linalool, epi- α -cadinol, α -bergamotene, and γ -cadinene) that exert antimicrobial activity against *S. aureus* and *B. subtilis*. Similarly, the essential oil of *A. ludoviciana* contains camphor, 1,8-cineole, and camphene, which were effective against *Acanthamoeba castellanii*, *Leishmania infantum*, and *Trichomonas vaginalis* [66].

Plant species	Gastro-intestinal disorders (parts ¹)	Chemical compounds	Antimicrobial activity	References
<i>Ambrosia artemisiifolia</i> L.	Stomachache and intestinal parasites (All)	Isabelin (germacranolide sesquiterpene dilactone)	<i>S. aureus</i> <i>C. albicans</i>	[64]
<i>Artemisia ludoviciana</i> Nutt	Chronic active gastritis and peptic ulcer (St, L)	Camphor, 1,8-cineole and camphene, sesquiterpene lactones, and flavonoids	<i>V. cholerae</i> <i>H. pylori</i> <i>A. castellanii</i> , <i>L. infantum</i> <i>T. vaginalis</i>	[57, 65–67]
<i>Geranium mexicanum</i> Kunth	Diarrhea, dysentery, and stomachache (R, St, L)	(–)-Epicatechin	<i>S. flexneri</i> <i>S. sonnei</i> , <i>E. histolytica</i> <i>G. lamblia</i>	[59–61]
<i>Ocimum basilicum</i>	Diarrhea, dysentery, stomachache and vomit (St, L)	Linalool, epi- α -cadinol, α -bergamotene, and γ -cadinene	<i>V. cholerae</i> <i>S. aureus</i> and <i>B. subtilis</i>	[62, 65]
<i>Lantana achyranthifolia</i> Desf.	Gastro-intestinal disease (AP)	Carvacrol, isocaryophyllene, α -bisabolol, bisabolene, and 1,8-cineole	<i>V. cholerae</i> , <i>S. boydii</i> , <i>Y. enterocolitica</i> <i>F. moniliforme</i>	[8, 68]
<i>Chiranthodendron pentadactylon</i>	Antimicrobial and antidiarrheal activities (F)	Epicatechin and tiliroside	<i>S. flexneri</i> <i>S. sonnei</i> <i>E. histolytica</i> <i>G. lamblia</i> <i>E. coli</i> <i>Salmonella</i> spp. <i>V. cholerae</i> .	[60, 69]
<i>Anoda cristata</i>	Stomachache (L)	Acacetin and diosmetin	<i>H. pylori</i>	[63, 70]
<i>Bocconia frutescens</i>	Diarrhea, dysentery stomachache ulcers (L)		<i>E. coli</i> and <i>S. aureus</i> .	[63]
<i>Lippia graveolens</i> H.B.K.	Diarrhea, dysentery, indigestion (AP)	Carvacrol, α -terpinyl acetate, m-cymene, and thymol	<i>R. solani</i> <i>E. coli</i> <i>P. aeruginosa</i> <i>S. aureus</i>	[68, 71]
<i>Guaiacum coulteri</i> A. Gray	Chronic active gastritis and peptic ulcer (OB)	Alkaloids	<i>Helicobacter pylori</i>	[57]

¹All, all parts; AP, aerial parts; L, leaf; St, stem; and OB, outer bark.

Table 3.
Antimicrobial activity of native medicinal plants of Oaxaca, Mexico.

Aqueous extracts of *A. ludoviciana ssp. Mexicana* and methanolic extracts of *Guaiacum coulteri* showed high growth inhibitory activity against *Helicobacter pylori* at minimum inhibitory concentrations of 125 and < 15.6 $\mu\text{g/mL}$, respectively [57].

H. pylori is the etiological agent of chronic active gastritis and peptic ulcer and is related to gastric carcinoma. It was also shown that *A. ludoviciana* has activity against *Campylobacter jejuni* and *C. coli* at minimum bactericidal concentrations of 0.5 mg/mL [74]. Ruiz-Cancino et al. [67] determined that the sesquiterpene lactones (douglanin, ludovicin A, 1 α , 3 α -dihydroxyarbusculin B, santamarin, arglanin, artemorin, chrysartemin B, armefolin, ridentin, eudesmanolide 3 α -hydroxyreynosin, etc.) and flavonoids (eupatilin and jaceosidin) of *A. ludoviciana* spp. *mexicana* are the molecules responsible for the inhibitory properties against the nuclear transcription factor kappa B, NF- κ B [14]. NF- κ B is involved in critical mechanisms related to the development of cancer. The signaling cascades of NF- κ B may be the main malignant gastrointestinal mediators that favor esophageal, gastric, and colon cancer [75].

The essential oils of *Lantana achyranthifolia* consist of monoterpenes and sesquiterpenes (carvacrol, isocaryophyllene, α -bisabolol, α -bisabolene, and 1,8-cineole), have antibacterial activities against Gram-positive and Gram-negative bacteria, *V. cholerae*, *Shigella boydii*, and *Yersinia enterocolitica* [76]. The antibacterial activity results from the compounds carvacrol, 1,8-cineole and linalool [77]. The essential oils of *L. achyranthifolia* and *Lippia graveolens* have antifungal activity [68] and contain carvacrol, α -terpinyl acetate, β -caryophyllene, geranyl acetate, terpinyl acetate, bornyl acetate, and limonene [77, 78]. More research is required on other medicinal species with similar potential.

The extracts of *Cnidoscolus aconitifolius*, *Crotalaria pumila*, and *Anoda cristata* (Table 3) inhibit bacterial growth. The flavonoids acacetin and diosmetin from *A. cristata* have been shown to inhibit up to 90% of *H. pylori* growth. These data suggest that, through the use of nutraceutical food plants, *H. pylori* infections can be prevented [70]. *Ambrosia artemisiifolia* L. is used to treat stomach pain and contains isabelin, a germacranolide sesquiterpene dilactone with antimicrobial activity against *S. aureus* and *Candida albicans* [64].

5. Remarks

Medicinal plants continue to play an indispensable role in the daily life of rural and urban communities. However, their use is controversial, and they are often only used in teas or hot drinks; they are not associated with healing properties. Through interactions with the indigenous communities of Oaxaca, Mexico, it was found that all households use the inherited knowledge of medicinal plants. In the regions furthest from the urban centers where the hospitals are located, medicinal plants are part of the survival strategy.

In the communities of Oaxaca and other regions of Mexico, healers (the generic name for traditional practitioners) are called “yerberos” (herbalists) because they only use plants. The consultation or intervention of the healers occurs when the symptoms of the disease or ethnodisease continue after the patient takes the “remedies” (teas, crushed, chewed, potions, etc.), which are prepared by the adults of the family. We define ethnodisease as a disorder that has no somatic symptoms, description of organic or metabolic dysfunction, or association with clinical symptomatology. Thus, “empacho” is often associated with indigestion, but has a broader sociocultural description than a gastroenterological disorder.

Based on our field notes and previous documentation of the indigenous communities in Oaxaca, Mexico, as well as our bibliography, a brief list of medicinal plants used for gastrointestinal disorders was compiled. This list comprises 71 botanical families, among which the most speciose were Asteraceae (29), Fabaceae (15), Euphorbiaceae (9), Solanaceae (9), Lamiaceae (9), Verbenaceae (6), Myrtaceae (5),

Malvaceae (5), and Fagaceae (5). The families included 147 genera and 186 endemic species. The most frequently used genera were *Croton* (5), *Quercus* (4), *Piper* (4), *Psidium* (3), *Ocimum* (3), and *Tagetes* (3) (**Table 1A**). The medicinal species introduced to Mexico were excluded from this list, despite being widely used by Oaxacan indigenous groups.

The use of antibiotics for infectious gastrointestinal diseases is unconscionable among the users of pharmacological medicine and has generated collateral damage, including antibiotic resistance of pathogenic microorganisms. Cases of antibiotic resistance are increasingly frequent, and thus, medicinal plants will fill an important niche once their antibacterial, antiprotozoal, antisecretory, spasmolytic, and anti-inflammatory protective effects are demonstrated. In addition to these benefits, antiradical and antioxidant activity effective against gastroenterological disorders have been shown after the frequent consumption of teas. Advances have been made in the knowledge of antipathogenic effects of medicinal plants and their association with specific compounds. However, not all plants used by the indigenous groups of Mexico and Latin America are being studied and documented.

Acknowledgements

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A. Appendix

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
Acanthaceae			
<i>Justicia spicigera</i> Schltdl.	Stomachache, diarrhea, dysentery, nausea, cramps (L, St, F)	Phenolics, flavonoids, lignans	[14, 17, 51, 53, 79–81]
Amaranthaceae			
<i>Dysphania ambrosioides</i> (L.)	Diarrhea, vomiting, stomach pain and inflammation, parasites (AP, L)	Terpenoids, flavonoids	[4, 11, 53, 54, 60, 82–85]
<i>Dysphania graveolens</i> (Willd.) Mosyakin & Clemants	Stomach pain, diarrhea, parasites, dysentery, indigestion, <i>empacho</i> , vomiting (AP)	Terpenoids, flavonoids	[17, 53, 57, 86]
Anacardiaceae			
<i>Amphipterygium adstringens</i> (Schltdl.) Standl.	Ulcers, stomach cancer, gastritis, stomach pain, intestinal infection, and inflammation (Se, St, Bark)	Triterpenes, phenolic, lipids	[41, 53, 57, 87, 88]
<i>Spondias mombin</i> L.	Diarrhea, stomach pain, dysentery, <i>empacho</i> (OB)	Phenols, flavonoids, saponins	[53, 89–91]
<i>Spondias purpurea</i> L.	Diarrhea in children (OB, R)	Phenols, flavonoids, tannins, phenolic acid derivatives, terpenoids	[14, 17, 41, 92, 93]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
Annonaceae			
<i>Annona glabra</i> L.	Dysentery, diarrhea (R, L, Sh)	Flavonoids, diterpenoids	[53, 94, 95]
<i>A. reticulata</i> L.	Diarrhea, stomach pain, intestinal pain (bark, L, Sh)	Acetogenins, terpenoids, alkaloids, phenols	[14, 53, 92, 96–100]
Apiaceae			
<i>Donnellsmithia juncea</i> (Spreng.) Mathias & Constance	Diarrhea, vomiting (All)		[53]
<i>Eryngium foetidum</i> L.	Diarrhea, dysentery, stomach pain (L)	Tannins, saponins, terpenoids, flavonoids, phenols, eryngial	[53, 54, 92, 101–103]
Apocynaceae			
<i>Plumeria rubra</i> L.	Intestinal parasites, diarrhea, purgative (Steam latex)	Terpenoids, iridoids, phenols, flavonoids	[53, 104, 105]
<i>Thevetia ahouai</i> (L.) A. DC.	Ulcers, purgative (All)	Cardenolide glycosides	[53, 106]
Araceae			
<i>Anthurium schlechtendalii</i> Kunth ssp. Jimenezii	Diarrhea (All)		[92]
<i>A. schlechtendalii</i> Kunth ssp. <i>Schlechtendalii</i>	Diarrhea (All)		[92]
Aristolochiaceae			
<i>Aristolochia leuconeura</i> Linden	Diarrhea, colics (All)		[84]
Asclepiadaceae			
<i>Asclepias curassavica</i> L.	Intestinal parasites, purgative (AP)	Cardenolides, glycosides, protease	[14, 92, 107]
Asteraceae			
<i>Ageratum conyzoides</i> L.	Stomach pain (All)	Terpenoids, flavonoids, benzofuranes, and coumarins	[92, 108, 109]
<i>Ambrosia artemisiifolia</i> L.	Stomach pain, intestinal parasites (L)	Terpenoids, sterols	[53]
<i>Artemisia ludoviciana</i> ssp. <i>mexicana</i> Nutt.	Stomach pain, vomiting, dysentery, colic, parasites, indigestion, diarrhea, flatulence (L, St)	Sesquiterpene lactones, flavonoids	[4–6, 10, 11, 14, 53, 57, 60, 67, 72, 84]
<i>Baccharis conferta</i> Kunth	Diarrhea, vomiting, indigestion, colic, and stomach pain (All)	Flavonoids, terpenoids	[10, 41, 43, 53, 110]
<i>B. salicifolia</i> (Ruiz & Pav.) Pers.	Stomach infection, stomach pain; diarrhea, dysentery (All)	Flavonoids, terpenoids	[5, 9, 13, 53]
<i>Barkleyanthus salicifolius</i> (Kunth) H. Rob. & Brettell	Vomiting, diarrhea, fever (St, L)	Terpenoids, alkaloids, flavonoids	[9, 53, 111]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>Bidens pilosa</i> L.	Diarrhea, vomiting, stomach pain, and inflammation, ulcers (F, L, B)	Terpenoids	[53, 112]
<i>Calea ternifolia</i> Kunth var. <i>ternifolia</i>	Stomach pain, diarrhea, indigestion, malaria, <i>bilis</i> (L)	Terpenoids	[10, 53, 113]
<i>C. urticifolia</i> (Miller) DC.	Dysentery, diarrhea, malaria, stomach pain, heartburn (All)	Terpenoids	[14, 17, 53, 92, 113]
<i>Chaptalia nutans</i> L. Pollak	Dysentery, intestinal parasites (R, L, St)	7-O-beta-D-glucopyranosyl-nutanocoumarin	[17, 41, 53, 114]
<i>Chrysactinia mexicana</i> A. Gray	Diarrhea, dysentery, colic (AP)	Terpenoids	[11, 53, 60, 115–117]
<i>Dyssodia papposa</i> (Vent.) Hitchc.	Diarrhea, stomach pain, vomiting (All)		[53]
<i>Heterotheca inuloides</i> Cass	Gastritis, ulcers (AP)	Terpenoids, flavonoids, phenolics, steroids	[53, 57, 118–120]
<i>Koanophyllon albicaule</i> (Sch. Bip. ex Klatt) R.M. King & H. Rob.	Diarrhea (L)	Sterols, flavonoids, tannins	[53]
<i>Melampodium divaricatum</i> (Rich.) DC	Dysentery, vomiting, nausea (All)	Terpenoids, coumarins, glycoside derivatives	[53, 121]
<i>Mikania houstoniana</i> (L.) B.L. Rob.	Stomach pain (St)		[53]
<i>Parthenium hysterophorus</i> L.	Stomach pain, fever, <i>empacho</i> , malaria, parasites (All)	Resin, alkaloids	[53]
<i>Pinaropappus roseus</i> (Less.) Less.	Constipation (L, R, F)		[53]
<i>Piqueria trinervia</i> Cav.	Intestinal infections, diarrhea, typhoid, <i>empacho</i> , stomach pain, purgative, parasites, malaria (F, L, R)	Terpenoids	[53, 122]
<i>Pluchea odorata</i> (L.) Cass.	Vomiting, stomach pain (L)	Flavonoids, terpenoids	[41, 53, 123]
<i>Porophyllum macrocephalum</i> DC.	Laxative (R, L)	Terpenoids, sulfur compounds	[10, 53]
<i>Pseudognaphalium attenuatum</i> DC.	Stomach pain, gastritis, <i>bilis</i> (AP)		[53, 92]
<i>Sanvitalia procumbens</i> Lam.	Diarrhea, dysentery, indigestion, vomiting, stomach pain (All)	Terpenoids	[53]
<i>Tagetes erecta</i> L.	Stomach pain, <i>empacho</i> , diarrhea, colic, vomiting, indigestion, parasites (L, F)	Thiophene derivative, terpenoids, flavonoids, carotenoids	[53, 54, 124–126]
<i>T. filifolia</i> Lag.	Stomach pain, flatulence (All)	Terpenoids	[41, 53, 127]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>T. lucida</i> Cav.	Stomach pain, colic, diarrhea, dysentery, <i>empacho</i> , typhoid, vomiting (L, F)	Coumarins, terpenoids, flavonoids	[53, 56, 57, 92, 124]
<i>Tithonia diversifolia</i> (Hemsl.) A. Gray	Vomiting, stomach pain, diarrhea, malaria (L)	Terpenoids, flavonoids, phenols	[5, 10, 57, 84, 128]
<i>Vernonanthura patens</i> (Kunth) H. Rob.	Dysentery (L, Sh, B)	Terpenoids	[53, 129, 130]
<i>Zinnia peruviana</i> L.	Diarrhea, stomach pain (All)		[53, 131]
Bignoniaceae			
<i>Dolichandra uncata</i> (Andrews) L.G. Lohmann	Intestinal inflammation, fever (L, B9)		[53]
<i>Parmentiera aculeata</i> (Kunth) Seem.	Dysentery, <i>empacho</i> (Bark, Fr)	Flavonoids, sterols, tannins	[7, 41, 53, 132]
<i>Tabebuia rosea</i> (Bertol.) A. DC.	Dysentery, fever, stomach inflammation (Bark)		[41, 43, 53]
<i>Tecoma stans</i> (L.) Juss ex Kunth	Stomach pain, dysentery, <i>bilis</i> , gastritis, poor digestion, <i>empacho</i> , intestinal atony (L, St, B, OB)	Alkaloids, terpenoids, flavonoids, phenolic acids	[6, 22, 41, 53, 133, 134]
Brassicaceae			
<i>Nasturtium officinale</i> R. Br.	Indigestion (All)		[53]
Burseraceae			
<i>Bursera microphylla</i> A. Gray	Stomach pain (B, gum)	Lignans, terpenoids, flavonoids	[53, 135]
<i>Bursera simaruba</i> L. Sarg.	Dysentery, stomach pain, diarrhea, intestinal infection, purgative (OB)	Tannins, flavonoids, saponins	[41, 53, 135]
Caricaceae			
<i>Carica papaya</i> L.	Diarrhea, dysentery, colic, intestinal parasites, constipation (Fr, L, Se, Latex)	Papain	[6, 11, 53, 72, 92]
Celastraceae			
<i>Semialarium mexicanum</i> (Miers) Mennega	Ulcers, stomach pain, colic, diarrhea, dysentery (R)	Terpenoids	[11, 53, 60]
Chrysobalanaceae			
<i>Chrysobalanus icaco</i> L.	Diarrhea, dysentery (Se, Fr)		[53]
Convolvulaceae			
<i>Ipomoea bracteata</i> Cav.	Diarrhea (T)		[53, 54]
<i>Ipomoea pes-caprae</i> (L.)	Dysentery (L)	Aromatic compounds	[41, 53]
Cruciferae			

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>Lepidium virginicum</i> L.	Diarrhea, dysentery, stomach pain, flatulence, colic, vomiting, indigestion, <i>empacho</i> , purgative, intestinal parasites (All)		[53]
Cucurbitaceae			
<i>Cucurbita pepo</i> L.	Intestinal parasites (Se)	Steroids, alkaloids, flavonoids, terpenoids, glycosides, pyrrolidine, sterols	[53]
<i>Sechium edule</i> (Jacq.) Sw.	Intestinal parasites, vomiting, constipation (L)		[6, 92]
Dilleniaceae			
<i>Curatella americana</i> L.	Stomach pain, diarrhea (Fr, L)	Flavonoids, terpenoids	[41, 53]
Eduphorbiaceae			
<i>Phyllanthus niruri</i> L.	Diarrhea, stomach pain (All)		[53]
Ericaceae			
<i>Arctostaphylos pungens</i> Kunth	Diarrhea, stomach pain, <i>empacho</i> (L, Fr)	Pyrocatechin, resin, and tannins	[53]
Euphorbiaceae			
<i>Acalypha alopecuroides</i> Jacq.	Diarrhea, ulcers (All)	Flavonoids, polyphenols, saponins, tannins	[53]
<i>Croton ciliatoglandulifer</i> Ortega	<i>Empacho</i> , intestinal inflammation, purgative (All)	Isoquinoline derivatives	[53]
<i>C. draco</i> Schltdl. & Cham.	Diarrhea, vomiting, stomach pain, <i>empacho</i> (OB)		[41, 53, 96]
<i>C. glandulosus</i> L.	Stomach pain	Terpenoids, alkaloids	[92]
<i>C. repens</i> Schlecht.	Diarrhea, dysentery (R)		[53, 92]
<i>C. soliman</i> Cham. & Schltdl.	Intestinal inflammation, parasites (R)		[53]
<i>Euphorbia tithymaloides</i> L.	Intestinal parasites, purgative, gastritis (Latex)	Alkaloids, steroids, sterols, terpenoids	[53, 92]
<i>Hura polyandra</i> Baill.	Stomach pain, constipation, intestinal parasites, purgative (Se)		[10, 53, 92, 96]
<i>Jatropha curcas</i> L.	Diarrhea, vomiting, constipation (R)		[53, 136]
Fabaceae			
<i>Chamaecrista hispidula</i> (Vahl) H.S. Irwin & Barneby			[84]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>Crotalaria longirostrata</i> Hook. & Arn.	Indigestion (L, B)		[41, 53]
<i>Desmodium incanum</i> DC.	Diarrhea, stomach pain (L)		[53]
<i>Diphysa carthagenensis</i> Jacq.	Diarrhea, dysentery (OB)		[53]
<i>Enterolobium cyclocarpum</i> (Jacq.) Griseb.	Diarrhea, purgative, indigestion (Fr, gum, bark)	Terpenoids	[53]
<i>Gliricidia sepium</i> (Jacq.) Kunth ex Walp.	<i>Empacho</i> , parasites (L, St)	Flavonoids	[41, 53, 96]
<i>Hymenaea courbaril</i> L.	Dysentery, diarrhea, ulcers (OB, L)	Tannins flavonoids, terpenoids	[41, 53, 92]
<i>Leucaena diversifolia</i> (Schltdl.) Benth.	Parasites (All)		[53]
<i>Machaerium floribundum</i> Benth.	Diarrhea (All)		[43, 53, 92]
<i>Mimosa albida</i> Humb. & Bonpl. Ex Willd.	Diarrhea, dysentery (B)		[41, 53]
<i>Pithecellobium dulce</i> (Roxb.) Benth.	Diarrhea, stomach pain, dysentery, constipation, indigestion (OB)	Flavonoids, glucoside derivatives, sterols, terpenoids	[53]
<i>Prosopis juliflora</i> (Sw.) DC.	Stomach pain, dysentery, indigestion, purgative, parasites (L)	Alkaloids, terpenoids, flavonoids	[53]
<i>P. laevigata</i> (Humb. & Bonpl. ex Willd.) M.C. Johnst.	Colic, intestinal inflammation, dysentery, stomach pain (OB)		[53]
<i>Senna skinneri</i> (Benth.) H.S. Irwin & Barneby	Dysentery, <i>empacho</i> , fever (L)		[53]
<i>Vachellia farnesiana</i> (L.) Wight & Arn.	<i>Empacho</i> , dysentery, dyspepsia, diarrhea, typhoid (All)	Sterols, alkaloids, flavonoids, phenols	[53]
<i>Zornia thymifolia</i> Kunth	Stomach pain, <i>bilis</i> , ulcer (St, L)		[53, 92, 96]
Fabaceae			
<i>Quercus crassipes</i> Bonpl.	Diarrhea (OB)		[53]
<i>Q. glaucescens</i> Bonpl.	Diarrhea (OB)	Tannins	[92]
<i>Q. oleoides</i> Schltdl. & Cham.	Diarrhea, gastritis, <i>empacho</i> (OB)	Tannins	[17, 53, 92]
<i>Q. sapotifolia</i> Liebm.	Diarrhea (OB)	Tannins	[92]
Geraniaceae			
<i>Geranium mexicanum</i> Kunth	Diarrhea, dysentery, stomach pain (R, AP)	Sterols, flavonoids, tyramine	[11, 53, 60, 61, 72]
Gesneriaceae			
<i>Sinningia incarnata</i> (Aubl.) D.L. Denham	Diarrhea, dysentery (T)		[14, 92]
Heliconiaceae			

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>Heliotropium indicum</i> L.			[84]
Krameriaceae			
<i>Krameria paucifolia</i> (Rose) Rose	Diarrhea (AP)		[137]
Lamiaceae			
<i>Agastache mexicana</i> Bye, E.L. Linares and Ramam.	Colic, stomach pain, <i>bilis</i> (F, L, St)	Terpenoids, oleic acid, flavonoids	[138, 139]
<i>A. mexicana</i> ssp. <i>Mexicana</i>	Colic, stomach pain, intestinal pain, <i>empacho</i> , indigestion (L)	Terpenoids, flavonoids	[138, 140–142]
<i>Clinopodium macrostemum</i> var. <i>laevigatum</i> (Moc. & Sessé ex Benth.) Kuntze	Stomach pain, dysentery, hangover (L, F)	Terpenoids	[14, 53, 143]
<i>Ocimum basilicum</i> L.	Diarrhea, dysentery, stomach pain, vomiting, <i>empacho</i> (AP)		[9, 11, 41, 43, 53, 57, 60, 72, 140, 144]
<i>O. campechianum</i> Mill.	Intestinal inflammation, ulcers, gastritis, fever, dysentery, <i>empacho</i> , vomiting, stomach pain vermifuge (L, B)	Terpenoids	[53, 54, 145, 146]
<i>O. carnosum</i> (Spreng.) Link & Otto ex Benth	Stomach pain, flatulence, diarrhea, dysentery, abdominal pain and intestinal parasites (L)	Phenols, terpenoids	[53]
<i>Salvia hispanica</i> L.	Diarrhea (Se)	Unsaturated fatty acids, flavonoids	[53, 147, 148]
<i>S. microphylla</i> Kunth	Diarrhea, dysentery, <i>empacho</i> , infection and inflammation of the stomach, vomiting (B)	Terpenoids, alkaloids, tannins	[53, 146]
<i>Vitex mollis</i> Kunth	Colic, intestinal inflammation, diarrhea, dysentery, stomach pain (L, Sh)		[53]
Lauraceae			
<i>Litsea glaucescens</i> Kunth	Stomach pain, <i>empacho</i> , children's diarrhea, indigestion (L)	Terpenoids	[53, 54, 149, 150]
<i>Persea americana</i> Mill.	Stomach pain, parasites, diarrhea, dysentery, constipation, flatulence, vomiting (Fr, L)	Terpenoids	[41, 57, 92, 151–153]
Liliaceae			
<i>Milla biflora</i> Cav.	Vomiting, stomach pain, nausea, diarrhea (F)		[53]
Loganiaceae			
<i>Buddleja americana</i> L.	Stomach pain, stomach infection, ulcers (L)	Lignans, flavonoids, alkaloids, tannins	[14, 53, 92]
Lygodiaceae			

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>Lygodium venustum</i> Sw.	Diarrhea, dysentery, nausea (AP)		[11, 53, 60, 72]
Lythraceae			
<i>Cuphea hyssopifolia</i> Kunth	Stomach pain (All)		[53]
Magnoliaceae			
<i>Chiranthodendron pentadactylon</i> Larreat	Diarrhea, dysentery, colic (F)	Flavonoids, steroids, phenols	[11, 53, 60, 72, 133, 144]
<i>Magnolia mexicana</i> DC.	Stomach pain, parasites (F)		[6, 53]
<i>Magnolia schiedeana</i> Schltdl.	Stomach pain (F)		[6]
Malpighiaceae			
<i>Byrsonima crassifolia</i> (L.) Kunth	Diarrhea, dysentery, <i>empacho</i> , stomach pain, indigestion, constipation (OB)	Tannins, proanthocyanidines, phenolic acids, terpenoids	[4, 6, 10, 41, 53, 84, 92]
<i>Malpighia mexicana</i> A. Juss.	<i>Empacho</i> , diarrhea, dysentery (OB)		[53]
Malvaceae			
<i>Anoda cristata</i> (L.) Schltdl.	Stomach pain, <i>empacho</i> , intestinal inflammation, <i>bilis</i> (AP, R)		[53]
<i>Malvaviscus arboreus</i> Cav.	Dysentery, diarrhea, stomach pain (AP)	Flavonoids, sterols, tannins	[14, 17, 53, 92, 96]
<i>Pavonia schiedeana</i> Steud.	<i>Empacho</i> , diarrhea (L, St)	Sterols, tannins	[53]
<i>Sida acuta</i> Burm. f.	Diarrhea, dysentery, <i>empacho</i> (L)		[53]
<i>Sida rhombifolia</i> L.	Stomach pain, gastritis, ulcers, diarrhea, dysentery (B)	Alkaloids	[41, 53, 146]
Melastomataceae			
<i>Conostegia xalapensis</i> (Bonpl.) D. Don	Diarrhea (L)		[41, 53]
Meliaceae			
<i>Swietenia humilis</i> L.	<i>Empacho</i> , stomach pain, diarrhea, fever, amoebic dysentery (Se, OB)		[41, 53]
<i>Swietenia macrophylla</i> G. King	Diarrhea, fever (Se)		[53]
Menispermaceae			
<i>Cissampelos pareira</i> L.	Dysentery, diarrhea, snake bite (L, St, R)	Alkaloids, sterols	[14, 17, 53, 54, 92]
Molluginaceae			
<i>Mollugo verticillata</i> L.	Intestinal inflammation, dysentery, diarrhea, <i>empacho</i> , colic, and stomach pain (B, L)		[41, 53]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
Moraceae			
<i>Dorstenia drakena</i> L.	Diarrhea, dysentery, stomach pain (R)		[6, 53, 137]
<i>Ficus cotinifolia</i> Kunth	Malaria, ulcers (L, latex)		[53]
Muntingiaceae			
<i>Muntingia calabura</i> L.	Diarrhea, stomach pain, <i>empacho</i> , vomiting, dysentery, intestinal infection (OB)	Flavonoids	[53, 96]
Myrtaceae			
<i>Eugenia acapulcensis</i> Steud.	Diarrhea, dysentery (L, OB)	Tannins	[14, 17, 92]
<i>E. capuli</i> (Schltdl. & Cham.) Hook. & Arn	Diarrhea, dysentery (B)		[53]
<i>Psidium guajava</i> L.	Diarrhea, stomach pain, dysentery (All)	Sterols, terpenoids, flavonoids, tannins	[4, 6, 14, 43, 53, 54, 84, 92]
<i>P. guineense</i> Sw.	Diarrhea, dysentery, flatulence, vomiting (L, Fr, R)		[53, 92]
<i>P. salutare</i> (Kunth) O. Berg	Diarrhea (Fr, L, R)		[4, 6, 84]
Nyctaginaceae			
<i>Boerhavia coccinea</i> Mill	Purgative (L)		[53]
<i>Mirabilis jalapa</i> L.	Stomach pain, purgative (All)	Terpenoids, sterols, flavonoids, alkaloids	[53, 57]
Oleaceae			
<i>Fraxinus uhdei</i> (Wenz.) Lingelsh.	Diarrhea, infection intestinal, purgante (All)		[53]
Onagraceae			
<i>Oenothera rosea</i> L'Hér. ex	Stomach pain, inflammation or infection, colic, diarrhea, <i>bilis</i> , <i>empacho</i> , constipation (AP)		[53]
Papaveraceae			
<i>Argemone mexicana</i> L.	Purgative (Se)	Alkaloids, flavonoids y glycosides	[53]
<i>Bocconia frutescens</i> L	Diarrhea, dysentery, stomach pain, ulcers (AP)		[11, 53, 60, 72]
Passifloraceae			
<i>Passiflora exsudans</i> Zucc.	Dysentery (R)		[53]
Phytolaccaceae			
<i>Petiveria alliacea</i> L.	Stomach pain, dyspepsia, <i>susto</i> (R)	Alkaloids, sterols	[53]
Pinaceae			
<i>Pinus oocarpa</i> Schiede ex Schltdl.	Dysentery (L)		[53]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
Piperaceae			
<i>Piper aduncum</i> L.	Diarrhea (All)		[53]
<i>P. auritum</i> Kunth	Stomach pain, lack of appetite and constipation (L)	Terpenoids	[53, 54, 84, 96]
<i>P. sanctum</i> (Miq.) Schlttl. ex C. DC.	Stomach pain, diarrhea (L)	Aromatic compounds	[53, 54, 154]
<i>Piper schiedeianum</i> Steudel	<i>Empacho</i> (All)		[53]
Plantaginaceae			
<i>Scoparia dulcis</i> L.	Diarrhea, colic, stomach pain, dysentery (AP)	Flavonoid, glucosides, terpenoids	[53, 92]
Poaceae			
<i>Lasiacis ruscifolia</i> (Kunth) Hitchc.	Diarrhea (All)		[53, 54]
<i>Zea mays</i> L.	Diarrhea, stomach pain, constipation, dysentery, vomiting (cob, tassel)	Polysaccharides	[6, 17, 41, 53, 92]
Polemoniaceae			
<i>Loeselia mexicana</i> (Lam.) Brand	<i>Bilis</i> , dysentery, stomach pain or inflammation, indigestion, typhoid, vomiting, laxative (AP)		[14, 21, 41, 53, 92]
Polygalaceae			
<i>Polygala longicaulis</i> Kunth	Stomach pain (All)		[53]
<i>P. variabilis</i> Kunth	Abdominal pain (All)		[92]
Portulacaceae			
<i>Portulaca oleracea</i> L.	Intestinal infection, constipation, parasites (All)	Tannins, alkaloids, steroids	[53, 155]
Primulaceae			
<i>Myrsine juergensenii</i> (Mez) Ricketson & Pipoly	Stomach pain (All)		[53]
Rhamnaceae			
<i>Karwinskia humboldtiana</i> (Schult.) Zucc.	Dysentery (L)		[53]
Rhizophoraceae			
<i>Rhizophora mangle</i> L.	Dysentery, diarrhea (R, OB)	Phenolics, tannins	[53]
Rubiaceae			
<i>Galium mexicanum</i> Kunth	Stomach pain, <i>empacho</i> , diarrhea (All)		[53]
<i>Hintonia latiflora</i> (Sessé & Moc. ex DC.) Bullock	<i>Bilis</i> , gastric ulcer, <i>empacho</i> , parasites, malaria, gastroenteritis (OB)	Phenylcoumarins, cucurbitacins, glucocucurbitacins	[53, 156]
<i>Randia echinocarpa</i> Sessé & Moc. ex DC.	Diarrhea, malaria (All)		[53]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
Sapindaceae			
<i>Serjania triquetra</i> Radlk.			[53, 84]
Sapotaceae			
<i>Pouteria sapota</i> (Jacq.) H.E. Moore & Stearn	Diarrhea, <i>empacho</i> , stomach pain (All)	Phenolics, carotenoids, δ-tocopherol	[41, 53, 96, 157]
Scrophulariaceae			
<i>Capraria biflora</i> L.	Dysentery, inflammation of the stomach, gastroenteritis, intestinal fever (L)	Alkaloids, terpenoids, naphthoquinone	[53, 158, 159]
<i>Russelia sarmentosa</i> Jacq.	Stomach pain (L)		[14, 53, 92]
Selaginellaceae			
<i>Selaginella pallescens</i> (Presl) Spring	Diarrhea (All)		[53, 54]
Smilacaceae			
<i>Smilax laurifolia</i> L.	Diarrhea (All)	Taninos ausentes, 1B	[92]
Solanaceae			
<i>Calibrachoa parviflora</i> (A. Juss.) D'Arcy	Flatulence (All)		[53]
<i>Capsicum annum</i> L.	Dyspepsia, diarrhea, dysentery (Fr, S)	Alkaloids, capsaicinoids, carotenoids, tocopherols, sapogenins, phenolic acids	[53, 160]
<i>Cestrum dumetorum</i> Schltdl.	Intestinal inflammation (All)	Sterols, tannins	[53]
<i>C. nocturnum</i> L.	Stomach pain (L, St, F)	Sapogenins, coumarins, sitosterols, flavonoids	[41, 43, 53, 54, 161]
<i>Physalis coztomatl</i> Moc. & Sessé ex Dunal	Diarrhea with blood from amoebic infection or other parasites, diarrhea, stomach pain, dysentery (R, B)	Alkaloids, glycosides	[53]
<i>P. lagascea</i> R. & S.	Stomach pain (L)		[53]
<i>Solanum rostratum</i> Dunal	Purgative, stomach pain, diarrhea (L, F)		[53]
<i>S. amictum</i> Moric. ex Dunal.	Stomach pain (L)		[53]
<i>S. torvum</i> Sw.	Diarrhea, dolor de estómago (All)	Sapogenins, alkaloids	[53]
Sterculiaceae			
<i>Guazuma ulmifolia</i> Lam.	Diarrhea, intestinal infection, dysentery, <i>empacho</i> (All)	Procyanidins, Tannins	[6, 10, 14, 41, 53, 92]
<i>Melochia pyramidata</i> L.	Purgative (AP)		[53]
<i>Waltheria indica</i> L.	Diarrhea, dysentery, <i>empacho</i> , fever (All)	Flavonoids, esters, alkaloids	[53]
Verbenaceae			
<i>Lantana achyranthifolia</i> Desf.	Diarrhea, dysentery, <i>empacho</i> (All)	Terpenoids, flavonoids, and phenylpropanoids	[8, 16, 53, 68]

Family/ species ¹	Gastrointestinal disorders (used parts ²)	Bioactive compounds	References
<i>L. camara</i> L.	Stomach pain or inflammation, intestinal pain, parasites, vomiting (L, St)	Terpenoids, flavonoids, phenols	[53, 162–164]
<i>Lippia alba</i> (Mill.) N.E. Br. ex Britton & P. Wilson	Diarrhea, dysentery, stomach pain, colic, indigestion (AP)	Terpenoids	[4, 11, 53, 60, 61, 72, 92, 133, 146]
<i>L. graveolens</i> Kunth	Diarrhea, dysentery, <i>empacho</i> , indigestion (AP)	Terpenoids, flavonoids	[53, 165, 166]
<i>Stachytarpheta jamaicensis</i> (L.) Vahl	Stomach pain, intestinal inflammation (L)		[53, 92]
<i>Verbena litoralis</i> Kunth	Stomach pain, <i>bilis</i> , vomiting, malaria (L)		[53, 146]
Zygophyllaceae			
<i>Guaiacum coulteri</i> A. Gray	Ulcers (OB)	Alkaloids	[41, 53, 57]

¹Checklist of native vascular plants of Mexico [50].
²L, leaves; St, stem; F, flower; AP, aerial parts; Se, seeds; R, root; All, whole plant; Sh, shoots; B, branches; Fr, Fruit; T, tuber; OB, outer bark.

Table 1A.
List of plants used by the indigenous groups of Oaxaca for the treatment of gastrointestinal disorders.

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References

- [1] UNAM. Biblioteca Digital de la Medicina Tradicional Mexicana [Internet]. 2009. Available from: <http://www.medicinatradicionalmexicana.unam.mx/index.php> [Accessed: 2018-09-27]
- [2] Hernández C, Aguilera MG, Castro G. Situación de las enfermedades gastrointestinales en México. *Enfermedades Infecciosas y Microbiología*. 2011;**31**(4):137-151 <http://www.medigraphic.com/pdfs/micro/ei-2011/ei114f.pdf>
- [3] Secretaria de Salud. Informe sobre la Salud de los Mexicanos 2015: Diagnóstico general de la salud poblacional. [Internet]. 2015. Available from: https://www.gob.mx/cms/uploads/attachment/file/64176/INFORME_LA_SALUD_DE_LOS_MEXICANOS_2015_S.pdf [Accessed: 2018-09-27]
- [4] Heinrich M, Ankli A, Frei B, Weimann C, Sticher O. Medicinal plants in Mexico: Healers' consensus and cultural importance. *Social Science & Medicine*. 1998;**47**(11):1859-1871. DOI: 10.1016/S0277-9536(98)00181-6
- [5] Heinrich M, Robles M, West JE, Ortiz-de-Montellano BR, Rodriguez E. Ethnopharmacology of Mexican Asteraceae (Compositae). *Annual Review of Pharmacology and Toxicology*. 1998;**38**:539-565. DOI: 10.1146/annurev.pharmtox.38.1.539
- [6] Frei B, Sticher O, Viesca C, Heinrich M. Medicinal and food plants: Isthmus sierra Zapotec criteria for selection. *Angewandte Botanik*. 1998;**72**:82-86 http://discovery.ucl.ac.uk/1381118/1/Heinrich_Applied%20Botany_Frei_1998.pdf
- [7] Ankli A, Heinrich M, Bork P, Wolfram L, Bauerfeind P, Brun R, et al. Yucatec Mayan medicinal plants: Evaluation based on indigenous uses. *Journal of Ethnopharmacology*. 2002;**79**(1):43-52. DOI: 10.1016/S0378-8741(01)00355-5
- [8] Hernández T, Canalez M, Caballero J, Durán A, Lira R. Análisis cuantitativo del conocimiento tradicional sobre plantas utilizadas para el tratamiento de enfermedades gastrointestinales en Zapotitlán de Las Salinas, Puebla, México. *Interciencia*. 2005;**30**(9): 529-535 <http://www.redalyc.org/articulo.oa?id=33910803>
- [9] Valdés-Cobos A. Conservación y uso de plantas medicinales: El caso de la región de la Mixteca Alta Oaxaqueña, México. *Ambiente y Desarrollo*. 2013;**17**(33):87-97 <http://revistas.javeriana.edu.co/index.php/ambienteydesarrollo/article/view/7044>
- [10] Heinrich M. Ethnobotany and natural products: The research for new molecules, new treatments of old diseases or a better understanding of indigenous cultures? *Current Topics in Medicinal Chemistry*. 2003;**3**(2):29-42. DOI: 10.2174/1568026033392570
- [11] Calzada F, Arista R, Pérez H. Effect of plants used in Mexico to treat gastrointestinal disorders on charcoal-gum acacia-induced hyperperistalsis in rats. *Journal of Ethnopharmacology*. 2010;**128**(1):49-51. DOI: 10.1016/j.jep.2009.12.022
- [12] Juárez-Vázquez MC, Carranza-Álvarez C, Alonso-Castro AJ, González-Alcaraz VF, Bravo-Acevedo E, Chamarro-Tinajero FJ, et al. Ethnobotany of medicinal plants used in Xalpatlahuac, Guerrero, México. *Journal of Ethnopharmacology*. 2013;**148**(2):521-527. DOI: 10.1016/j.jep.2013.04.048
- [13] Loayza I, Abujder D, Aranda R, Jakupovic J, Collin G, Deslauriers

H, et al. Essential oils of *Baccharis salicifolia*, *B. latifolia* and *B. dracunculifolia*. *Phytochemistry*. 1995;**38**(2):381-389. DOI: 10.1016/0031-9422(94)00628-7

[14] Bork PM, Scmitz LL, Kuhtnt M, Escher C, Heinrich M. Sesquiterpene lactone containing Mexican Indian medicinal plants and pure sesquiterpene lactones as potent inhibitors of transcription facto NF-kB. *FEBS Letters*. 1997;**402**(1):85-90. DOI: 10.1016/S0014-5793(96)01502-5

[15] Savithramma N, Rao ML, Ankanna S. Screening of traditional medicinal plants for secondary metabolites. *International Journal of Research in Pharmaceutical Sciences*. 2011;**2**(4):643-647 https://www.researchgate.net/profile/S_Ankanna/publication/267690135_Screening_of_Medicinal_Plants_for_Secondary_Metabolites/links/55ed339808aeb6516268d1e2/Screening-of-Medicinal-Plants-for-Secondary-Metabolites.pdf

[16] Sousa EO, Costa JGM. Genus *lantana*: Chemical aspects and biological activities. *Brazilian Journal of Pharmacognosy*. 2012;**22**(5):1155-1180. DOI: 10.1590/S0102-695X2012005000058

[17] Heinrich M, Kuhnt M, Wright CW, Rimpler H, Phillipson JD, Schandelmaier A, et al. Parasitological and microbiological evaluation of Mixe indian medicinal plantas (Mexico). *Journal of Ethnopharmacology*. 1992;**36**(1):81-85. DOI: 10.1016/0378-8741(92)90063-W

[18] Calzada F. Additional antiprotozoal constituents from *Cuphea pinetorum*, a planta used in Mayan traditional medicine to treat diarrhea. *Phytotherapy Research*. 2005;**19**(8): 725-727. DOI: 10.1002/ptr.1717

[19] Atta AH, Mouneir SM. Evaluation of some medicinal plant extracts for

anitidiarrhoeal activity. *Phytotherapy Research*. 2005;**19**(6):481-485. DOI: 10.1002/ptr.1639

[20] Palombo EA. Phytochemicals from traditional medicinal plants used in the treatment of diarrhea: Modes of action and effects on intestinal function. *Phytotherapy Research*. 2006;**20**(9): 717-724. DOI: 10.1002/ptr.1907

[21] Navarro-García VM, Rojas G, Zepeda LG, Aviles M, Fuentes M, Herrera A, et al. Antifungal and antibacterial activity of four selected Mexican medicinal plants. *Pharmaceutical Biology*. 2006;**44**(4):297-300. DOI: 10.1080/13880200600715837

[22] Robles-Zepeda RE, Velázquez-Contreras CA, Garibay-Escobar A, Gálvez-Ruiz JC, Ruiz-Bustos E. Antimicrobial activity of northwestern Mexican plants against *Helicobacter pylori*. *Journal of Medicinal Food*. 2011;**14**(10):1280-1283. DOI: 10.1089/jmf.2010.0263

[23] Cushnie TP, Lamb AJ. Recent advances in understanding the antibacterial properties of flavonoids. *International Journal of Antimicrobial Agents*. 2011;**38**(2):99-107. DOI: 10.1016/j.ijantimicag.2011.02.014

[24] Arrieta-Baez D, de Esparza RR, Jiménez-Estrada M. Mexican plants used in the salmonellosis treatment. In: Kumar Y, editor. *Salmonella—A Diversified Superbug*. Rijeka, Croatia: Intech; 2012. pp. 169-184. DOI: 10.5772/31291

[25] Ramírez-Moreno E, Soto-Sanchez J, Rivera G, Marchat LA. Mexican medicinal plants as an alternative for the development of new compounds against protozoan parasites. In: Khater H, Govindarajan M, Benelli G, editors. *Natural Remedies in the Fight against Parasites*. Rijeka, Croatia: Intech; 2017. pp. 62-91. DOI: 10.5772/67259

- [26] Instituto Nacional de Estadística, Geografía e Informática (INEGI). Resultados de Encuesta Intercensal 2015. Instituto Nacional de Estadística, Geografía e Informática, Aguascalientes, México. [Internet]. 2015. Available from: <http://www.beta.inegi.org.mx/proyectos/enchogares/especiales/intercensal/?init=1> [Accessed: 2018-09-27]
- [27] García-Vargas LA. Radiografía demográfica de la población indígena en Oaxaca. *Oaxaca Población Siglo XXI*. 2018;**41**:7-20 <http://www.digepo.oaxaca.gob.mx/recursos/revistas/revista42.pdf>
- [28] Instituto Nacional de Lenguas Indígenas (INALI). Catálogo de las lenguas indígenas nacionales. Variantes lingüísticas de México con sus autodenominaciones y referencias geoestadísticas. México DF: INALI; 2010. 371 p. Available from: https://site.inali.gob.mx/pdf/catalogo_lenguas_indigenas.pdf [Accessed: 2018-09-27]
- [29] Grupos H-DJ, de Oaxaca I. Situación sociodemográfica. Oaxaca, México: Universidad Autónoma Benito Juárez de Oaxaca; 2005. 139 p
- [30] Toledo VM, Alarcón-Chaires P, Moguel P, Olivo M, Cabrera A, Leyequien E, et al. El atlas etnoecológico de México y Centroamérica: Fundamentos, métodos y resultados. *Etnoecología*. 2001;**6**(8):7-41 https://ccp.ucr.ac.cr/bvp/pdf/cambiodemografico/atlas_etnologico.pdf
- [31] Arellanes A, de la Cruz V, Romero MA, Sánchez C, Ruiz FJ, Martínez VR, et al. Historia y Geografía de Oaxaca. Carteles: Oaxaca, México; 2006. 207 p
- [32] Heinrich M. Herbal and symbolic medicines of the lowland Mixe (Oaxaca, Mexico). Disease concepts, healer's roles, and plant use. *Anthropos*. 1994;**89**:73-83 <https://www.jstor.org/stable/40463843>
- [33] Pérez-Nicolas M, Vibrans H, Romero-Manzanares A, Saynes-Vásquez A, Luna-Cavazos M, Flores-Cruz M, et al. Patterns of knowledge and use of medicinal plants in Santiago Camotlán, Oaxaca, Mexico. *Economic Botany*. 2017;**71**(3):209-223. DOI: 10.1007/s12231-017-9384-0
- [34] Rubel AJ, Browner CH. Antropología de la salud en Oaxaca. *Alteridades*. 1999;**9**(17):85-94 <http://alteridades.izt.uam.mx/index.php/Alte/article/view/461/460>
- [35] Ugent D. Medicine, myths and magic the folk healers of a Mexican market. *Economic Botany*. 2000;**54**(4):427-438. DOI: 10.1007/BF02866542
- [36] Berenzon-Gorn S, Hernández-Hernández J, Saavedra-Solano N. Percepciones y creencias en torno a la salud-enfermedad mental, narradas por curanderos urbanos de la ciudad de México. *Gazeta de Antropología*. 2001;**17**:21 <http://digibug.ugr.es/handle/10481/7481>
- [37] Berenzon-Gorn S, Ito-Sugiyama E, Vargas-Guadarrama LA. Enfermedades y padeceres por los que se recurre a terapeutas tradicionales de la Ciudad de México. *Salud Pública de México*. 2006;**48**(1):45-56 <http://saludpublica.mx/index.php/spm/article/view/6670/8292>
- [38] Lámbarri-Rodríguez A, Flores-Palacios F, Berzon-Gorn S. Curanderos, malestar y daños: una interpretación social. *Salud Mental*. 2012;**35**(2): 123-128 http://revistasaludmental.mx/index.php/salud_mental/article/view/1464/1462
- [39] Giovannini P, Reyes-García V, Waldstein A, Heinrich M. Do pharmaceuticals displace local knowledge and use of medicinal plants? Estimates from a cross-sectional study in a rural indigenous community,

Mexico. Social Science & Medicine. 2011;**72**(6):928-936. DOI: 10.1016/j.socscimed.2011.01.007

[40] Instituto Nacional de Salud Pública (INSP). Encuesta Nacional de Salud y Nutrición 2012. Resultados Nacionales. 2nd ed. Cuernavaca, México: Instituto Nacional de Salud Pública. 2013. 190 p. Available from: <https://ensanut.insp.mx/informes/ENSANUT2012ResultadosNacionales2Ed.pdf> Accessed: 2018-09-27

[41] Luna-José AL, Rendón-Aguilar B. Recursos vegetales útiles en diez comunidades de la Sierra Madre del Sur, Oaxaca, Mexico. Polibotánica. 2008;**26**:193-242 <http://www.scielo.org.mx/pdf/polib/n26/n26a11.pdf>

[42] Salas-Morales SH, Saynes-Vásquez A, Schibli L. Flora de la costa de Oaxaca, México: Lista florística de la región de Zimatán. Boletín de la Sociedad Botánica de México. 2003;**72**:21-58 <http://www.redalyc.org/pdf/577/57707202.pdf>

[43] Aguilar-Stoen M, Moe SR, Camargo-Ricalde SL. Home gardens sustain crop diversity and improve farm resilience in Candelaria Loxicha, Oaxaca, Mexico. Human Ecology. 2009;**37**(1):55-77. DOI: 10.1007/s10745-008-9197-y

[44] Pool A. Taxonomic revision of *Gouania* (Rhamnaceae) for North America. Annals of the Missouri Botanical Garden. 2014;**99**(3):490-552. DOI: 10.3417/2013016

[45] Meave JA, Rincón-Gutiérrez A, Ibarra-Maríquez G, Gallardo-Hernández C, Romero-Romero MA. Checklist of the vascular flora of a portion of the hyper-humid region of La Chinantla, Northern Oaxaca range, Mexico. Botanical Sciences. 2017;**95**(4):722-759. DOI: 10.17129/botsci.1812

[46] Zacharias S. Mexican Curanderismo as ethnopschotherapy: A qualitative

study on treatment practices, effectiveness, and mechanisms of change. International Journal of Disability, Development and Education. 2006;**53**(4):381-400. DOI: 10.1080/10349120601008522

[47] Montiel-Tafur M, Crowe TK, Torres E. A review of curanderismo and healing practices among Mexicans and Mexican Americans. Occupational Therapy International. 2009;**16**(1): 82-88. DOI: 10.1002/oti.265

[48] Ciofalo N. Indigenous women's ways of knowing and healing in Mexico. Women & Therapy. 2018;**41**(1-2):52-68. DOI: 10.1080/02703149.2017.1323478

[49] Maqueda AE. The use of *Salvia divinorum* from a Mazatec perspective. In: Labate BC, Cavnar C, editors. Plant Medicines, Healing and Psychedelic Science. Cham, Switzerland: Springer International Publishing AG; 2018. pp. 55-70. DOI: 10.1007/978-3-319-76720-8_4

[50] Villaseñor JL. Checklist of the native vascular plants of Mexico. Revista Mexicana de Biodiversidad. 2016;**87**(3):559-902. DOI: 10.1016/j.rmb.2016.06.017

[51] Sepúlveda-Jiménez G, Reyna-Aquino C, Chaires-Martínez L, Bermúdez-Torres K, Rodríguez-Monroy M. Antioxidant activity and content of phenolic compounds and flavonoids from *Justicia spicigera*. Journal of Biological Sciences. 2009;**9**(6):629-632. DOI: 10.3923/jbs.2009.629.632

[52] UNAM. Diccionario enciclopédico de la medicina tradicional mexicana. [Internet]. 2009. Biblioteca Digital de la Medicina Tradicional Mexicana. Available from: <http://www.medicinatradicionalmexicana.unam.mx/alfa.php?opcion=D&p=a> [Accessed: 2018-09-27]

[53] Argueta A, Gallardo VM, editors. Atlas de las plantas de la medicina

tradicional mexicana. 1st ed. México: Instituto Nacional Indigenista; 1994. 1786 p

[54] Lipp FJ. Ethnobotany of the Chinantec indians, Oaxaca, Mexico. *Economic Botany*. 1971;**25**:234-244. DOI: 10.1007/BF02860760

[55] Li W, Gao Y, Zhao J, Wang Q. Phenolic, flavonoid, and lutein ester content and antioxidant activity of 11 cultivars of Chinese marigold. *Journal of Agricultural and Food Chemistry*. 2007;**55**(21):8478-8484. DOI: 10.1021/jf071696j

[56] Céspedes CL, Avila JG, Martínez A, Serrato B, Calderón-Mugica JC, Salgado-Garciglia R. Antifungal and antibacterial activities of Mexican tarragon (*Tagetes lucida*). *Journal of Agricultural and Food Chemistry*. 2006;**54**(10):3521-3527. DOI: 10.1021/jf053071w

[57] Castillo-Juárez I, González V, Jaime-Aguilar H, Martínez G, Linares E, Bye R, et al. Anti-*Helicobacter pylori* activity of plants used in Mexican traditional medicine for gastrointestinal disorders. *Journal of Ethnopharmacology*. 2009;**122**(2):402-405. DOI: 10.1016/j.jep.2008.12.021

[58] Kiuchi F, Itano Y, Uchiyama N, Honda G, Tsubouchi A, Nakajima-Shimada J, et al. Monoterpene hydroperoxides with trypanocidal activity from *Chenopodium ambrosioides*. *Journal of Natural Products*. 2002;**65**(4):509-512. DOI: 10.1021/np010445g

[59] Barbosa E, Calzada F, Campos R. *In vivo* anti-giardial activity of three flavonoids isolated of some medicinal plants used in Mexican traditional medicine for the treatment of diarrhea. *Journal of Ethnopharmacology*. 2007;**109**(3):552-554. DOI: 10.1016/j.jep.2006.09.009

[60] Alanis AD, Calzada F, Cervantes JA, Torres J, Ceballos GM. Antibacterial properties of some plants used in Mexican traditional medicine for the treatment of gastrointestinal disorders. *Journal of Ethnopharmacology*. 2005;**100**(1-2):153-157. DOI: 10.1016/j.jep.2005.02.022

[61] Calzada F, Cervantes-Martínez JA, Yépez-Mulia L. *In vitro* antiprotozoal activity from the roots of *Geranium mexicanum* and its constituents on *Entamoeba histolytica* and *Giardia lamblia*. *Journal of Ethnopharmacology*. 2005;**98**(1-2):191-193. DOI: 10.1016/j.jep.2005.01.019

[62] Hussain AI, Anwar F, Sherazi STH, Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chemistry*. 2008;**108**(3):986-995

[63] Sánchez-Arreola E, Hernández-Molina LR, Sánchez-Salas JL, Martínez-Espino G. Alkaloids from *Bocconia frutescens* and biological activity of their extracts. *Pharmaceutical Biology*. 2006;**44**(7):540-543. DOI: 10.1080/13880200600883106

[64] Molinaro F, Tyc O, Beekwilder J, Cankar K, Berteau CM, Negre M, et al. The effect of isabelin, a sesquiterpene lactone from *Ambrosia artemisiifolia* on soil microorganisms and human pathogens. *FEMS Microbiology Letters*. 2018;**365**(4):fny001. DOI: 10.1093/femsle/fny001

[65] Sánchez E, García S, Heredia N. Extracts of edible and medicinal plants damage membranes of *Vibrio cholerae*. *Applied and Environmental Microbiology*. 2010;**76**(20):6888-6894. DOI: 10.1128/AEM.03052-09

[66] Baldemir A, Karaman Ü, Ilgun S, Kacmaz G, Demirci B. Antiparasitic efficacy of *Artemisia ludoviciana* Nutt. (Asteraceae) essential oil for

Acanthamoeba castellanii, *Leishmania infantum* and *Trichomonas vaginalis*. Indian Journal of Pharmaceutical Education and Research. 2018; **52**(3):416-425. DOI: 10.5530/ijper.52.3.48

[67] Ruiz-Cancino A, Cano AE, Delgado G. Sesquiterpene lactones and flavonoids from *Artemisia ludoviciana* ssp. *mexicana*. Phytochemistry. 1993;**33**(5):1113-1115. DOI: 10.1016/0031-9422(93)85032-M

[68] Hernández T, Canales M, García AM, Duran Á, Meráz S, Dávila P, et al. Antifungal activity of the essential oils of two verbenaceae: *Lantana achyranthifolia* and *Lippia graveolens* of Zapotitlán de las Salinas, Puebla (México). Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas. 2008;**7**(4):202-206 <https://www.blacpma.usach.cl/sites/blacpma/files/007-004.pdf>

[69] Calzada F, Juárez T, García-Hernández N, Valdes M, Ávila O, Mulia LY, et al. Antiprotozoal, antibacterial and antidiarrheal properties from the flowers of *Chiranthodendron pentadactylon* and isolated flavonoids. Pharmacognosy Magazine. 2017;**13**(50):240-244 <http://www.phcog.com/text.asp?2017/13/50/240/204564>

[70] Gomez-Chang E, Uribe-Estanislao GV, Martinez-Martinez M, Gálvez-Mariscal A, Romero I. Anti-*Helicobacter pylori* potential of three edible plants known as Quelites in Mexico. Journal of Medicinal Food. 2018. DOI: 10.1089/jmf.2017.0137

[71] Arana-Sánchez A, Estarrón-Espinosa M, Obledo-Vázquez EN, Padilla-Camberos E, Silva-Vázquez R, Lugo-Cervantes E. Antimicrobial and antioxidant activities of Mexican oregano essential oils (*Lippia graveolens* HBK) with different composition when microencapsulated in β -cyclodextrin. Letters in Applied

Microbiology. 2010;**50**(6):585-590. DOI: 10.1111/j.1472-765X.2010.02837.x

[72] Velázquez C, Calzada F, Torres J, González F, Ceballos G. Antisecretory activity of plants used to treat gastrointestinal disorders in Mexico. Journal of Ethnopharmacology. 2006;**103**(1):66-70. DOI: 10.1016/j.jep.2005.06.046

[73] Velázquez C, Calzada F, Esquivel B, Barbosa E, Calzada S. Antisecretory activity from the flowers of *Chiranthodendron pentadactylon* and its flavonoids on intestinal fluid accumulation induced by *Vibrio cholerae* toxin in rats. Journal of Ethnopharmacology. 2009;**126**(3):455-458. DOI: 10.1016/j.jep.2009.09.016

[74] Castillo SL, Heredia N, Contreras JF, García S. Extracts of edible and medicinal plants in inhibition of growth, adherence, and cytotoxin production of *Campylobacter jejuni* and *Campylobacter coli*. Journal of Food Science. 2011;**76**(6):M421-M426. DOI: 10.1111/j.1750-3841.2011.02229.x

[75] Gambhir S, Vyas D, Hollis M, Aekka A, Vyas A. Nuclear factor kappa B role in inflammation associated gastrointestinal malignancies. World Journal of Gastroenterology. 2015;**21**(11):3174-3183. DOI: 10.3748/wjg.v21.i11.3174

[76] Hernández T, Canales M, Avila JG, García AM, Martínez A, Caballero J, et al. Composition and antibacterial activity of essential oil of *Lantana achyranthifolia* Desf. (Verbenaceae). Journal of Ethnopharmacology. 2005;**96**(3):551-554. DOI: 10.1016/j.jep.2004.09.044

[77] Cimanga K, Apers S, de Bruyne T, Van Miert S, Hermans N, Totté J, et al. Chemical composition and antifungal activity of essential oils of some aromatic medicinal plants growing in the Democratic Republic

of Congo. Journal of Essential Oil Research. 2002;**14**(5):382-387. DOI: 10.1080/10412905.2002.9699894

[78] Deena MJ, Thoppil JE. Antimicrobial activity of the essential oil of *Lantana camara*. Fitoterapia. 2000;**71**(4):453-455. DOI: 10.1016/S0367-326X(00)00140-4

[79] Euler KL, Alam M. Isolation of kaempferitrin from *Justicia spicigera*. Journal of Natural Products. 1982;**45**(2):220-221. DOI: 10.1021/np50020a020

[80] Vega-Avila E, Espejo-Serna A, Alarcón-Aguilar F, Velasco-Lezama R. Cytotoxic activity of four Mexican medicinal plants. Proceedings of the Western Pharmacology Society. 2009;**52**:78-82

[81] Corrêa GM, Alcântara AF. Chemical constituents and biological activities of species of *Justicia*: A review. Revista Brasileira de Farmacognosia. 2012;**22**(1):220-238. DOI: 10.1590/S0102-695X2011005000196

[82] Kliks MM. Studies on the traditional herbal anthelmintic *Chenopodium ambrosioides* L.: Ethnopharmacological evaluation and clinical field trials. Social Science & Medicine. 1985;**21**(8):879-886. DOI: 10.1016/0277-9536(85)90144-3

[83] Ávila-Blanco ME, Rodríguez MG, Moreno-Duque JL, Muñoz-Ortega M, Ventura-Juárez J. Amoebicidal activity of essential oil of *Dysphania ambrosioides* (L.) Mosyakin & Clemants in an amoebic liver abscess hamster model. Evidence-based Complementary and Alternative Medicine. 2014:1-7. DOI: 10.1155/2014/930208

[84] Geck MS, Réyes-García AJ, Casu L, Leonti M. Acculturation and ethnomedicine: A regional comparison of medicinal plant knowledge among the Zoque of southern Mexico. Journal

of Ethnopharmacology. 2016;**187**:146-159. DOI: 10.1016/j.jep.2016.04.036

[85] Barros L, Pereira E, Calhella RC, Dueñas M, Carvalho AM, Santos-Buelga C, et al. Bioactivity and chemical characterization in hydrophilic and lipophilic compounds of *Chenopodium ambrosioides* L. Journal of Functional Foods. 2013;**5**(4):1732-1740. DOI: 10.1016/j.jff.2013.07.019

[86] Álvarez-Ospina H, Rivero Cruz I, Duarte G, Bye R, Mata R. HPLC determination of the major active flavonoids and GC-MS analysis of volatile components of *Dysphania graveolens* (Amaranthaceae). Phytochemical Analysis. 2013;**24**(3):248-254. DOI: 10.1002/pca.2405

[87] Oviedo-Chávez I, Ramírez-Apan T, Soto-Hernández M, Martínez-Vázquez M. Principles of the bark of *Amphipterygium adstringens* (Julianaceae) with anti-inflammatory activity. Phytomedicine. 2004;**11**(5):436-445. DOI: 10.1016/j.phymed.2003.05.003

[88] Rivero-Cruz I, Acevedo L, Guerrero JA, Martínez S, Bye R, Pereda-Miranda R, et al. Antimycobacterial agents from selected Mexican medicinal plants. The Journal of Pharmacy and Pharmacology. 2005;**57**(9):1117-1126. DOI: 10.1211/jpp.57.9.0007

[89] Igwe CU, Onyeze GO, Onwuliri VA, Osuagwu CG, Ojiako AO. Evaluation of the chemical compositions of the leaf of *Spondias mombin* Linn from Nigeria. Australian Journal of Basic and Applied Sciences. 2010;**4**(5):706-710. <http://www.ajbasweb.com/old/ajbas/2010/706-710.pdf>

[90] Alves-Brito S, Ferreira de Almeida CL, Italo de Santana T, Da Silva AR, Bezerra do Nascimento JC, Torres-Souza I, et al. Antiulcer activity and potential mechanism of action of the leaves of

- Spondias mombin* L. Oxidative Medicine and Cellular Longevity. 2018;1-20. DOI: 10.1155/2018/1731459
- [91] Cristofoli NL, Lima CA, Vieira MM, Andrade KS, Ferreira SR. Antioxidant and antimicrobial potential of cajazeira leaves (*Spondias mombin*) extracts. Separation Science and Technology. 2018;12:1-1. DOI: 10.1080/01496395.2018.1508233
- [92] Heinrich M, Rimpler H, Barrera NA. Indigenous phytotherapy of gastrointestinal disorders in a lowland Mixe community (Oaxaca, Mexico): Ethnopharmacologic evaluation. Journal of Ethnopharmacology. 1992;36:63-80. DOI: 10.1016/0378-8741(92)90062-V
- [93] Sameh S, Al-Sayed E, Labib RM, Singab AN. Genus *Spondias*: A phytochemical and pharmacological review. Evidence-based Complementary and Alternative Medicine. 2018;1-13. DOI: 10.1155/2018/5382904
- [94] Galvão SDL, Monteiro AD, Siqueira EP, Bomfim MR, Dias-Souza MV, Ferreira GF, et al. *Annona glabra* flavonoids act as antimicrobials by binding to *Pseudomonas aeruginosa* cell walls. Frontiers in Microbiology. 2016;21(7):1-9. DOI: 10.3389/fmicb.2016.02053
- [95] Nhiem NX, Hien NT, Tai BH, Anh HL, Hang DT, Quang TH, et al. New ent-kauranes from the fruits of *Annona glabra* and their inhibitory nitric oxide production in LPS-stimulated RAW264.7 macrophages. Bioorganic & Medicinal Chemistry Letters. 2015;25(2):254-258. DOI: 10.1016/j.bmcl.2014.11.059
- [96] Abe F, Nagafuji S, Yamauchi T, Okabe H, Maki J, Higo H, et al. Trypanocidal constituents in plants. 1. Evaluation of some Mexican plants for their trypanocidal activity and active constituents in Guaco, roots of *Aristolochia taliscana*. Biological & Pharmaceutical Bulletin. 2002;25(9):1188-1191. DOI: 10.1248/bpb.25.1188
- [97] Chang FR, Wu YC, Duh CY, Wang SK. Studies on the acetogenins of Formosan annonaceous plants. II. Cytotoxic acetogenins from *Annona reticulata*. Journal of Natural Products. 1993;56:1688-1694. DOI: 10.1021/np50100a005
- [98] Chavan SS, Shamkuwar PB, Damale MG, Pawar DP. A comprehensive review on *Annona reticulata*. International Journal of Pharmaceutical Sciences and Research. 2014;5(1):45-50. DOI: 10.13040/IJPSR.0975-8232.5(1).45-50
- [99] Jamkhande PG, Wattamwar AS, Kankudte AD, Tidke PS, Kalaskar MG. Assessment of *Annona reticulata* Linn. leaves fractions for in vitro antioxidative effect and antimicrobial potential against standard human pathogenic strains. Alexandria Journal of Medicine. 2016;52:19-25. DOI: 10.1016/j.ajme.2014.12.007J
- [100] Quílez AM, Fernández-Arche MA, García-Giménez MD, De la Puerta R. Potential therapeutic applications of the genus *Annona* local and traditional uses and pharmacology. Journal of Ethnopharmacology. 2018;225:244-270. DOI: 10.1016/j.jep.2018.06.014
- [101] Paul JH, Seaforth CE, Tikasingh T. *Eryngium foetidum* L.: A review. Fitoterapia. 2011;82(3):302-308. DOI: 10.1016/j.fitote.2010.11.010
- [102] Rojas-Silva P, Graziose R, Vesely B, Poulev A, Mbeunkui F, Grace MH, et al. Leishmanicidal activity of a daucane sesquiterpene isolated from *Eryngium foetidum*. Pharmaceutical Biology. 2014;52(3):398-401. DOI: 10.3109/13880209.2013.837077
- [103] Forbes WM, Gallimore WA, Mansingh A, Reese PB, Robinson RD. Eryngial (trans-2-dodecenal), a bioactive compound from *Eryngium*

- foetidum*: Its identification, chemical isolation, characterization and comparison with ivermectin in vitro. *Parasitology*. 2014;**141**(2):269-278. DOI: 10.1017/S003118201300156X
- [104] Hassan EM, Shahat AA, Ibrahim NA, Vlietinck AJ, Apers S, Pieters L. A new monoterpene alkaloid and other constituents of *Plumeria acutifolia*. *Planta Medica*. 2008;**74**(14):1749-1750. DOI: 10.1055/s-0028-1088317
- [105] Srivastava A, Gupta AK, Rajendiran A. Phytochemical screening and in-vitro anthelmintic activity of methanolic extract from the stem bark of *Plumeria rubra* Linn. *International Journal of Pharmaceutical Sciences and Research*. 2017;**8**(12):5336-5341. DOI: 10.13040/IJPSR.0975-8232.8(12).5336-41
- [106] Endo H, Warashina T, Noro T, Castro VH, Mora GA, Poveda LJ, et al. Cardenolide glycosides from *Thevetia ahouai* (LINN.) a. DC. *Chemical & Pharmaceutical Bulletin*. 1997;**45**(9):1536-1538. DOI: 10.1248/cpb.45.1536
- [107] Li JZ, Qing C, Chen CX, Hao XJ, Liu HY. Cytotoxicity of cardenolides and cardenolide glycosides from *Asclepias curassavica*. *Bioorganic & Medicinal Chemistry Letters*. 2009;**19**(7): 1956-1959. DOI: 10.1016/j.bmcl.2009.02.045
- [108] Shirwaikar A, Bhilegaonkar PM, Malini S, Kumar JS. The gastroprotective activity of the ethanol extract of *Ageratum conyzoides*. *Journal of Ethnopharmacology*. 2003;**86**(1):117-121. DOI: 10.1016/S0378-8741(03)00050-3
- [109] Kouame BF, Toure D, Kablan L, Bedi G, Tea I, Robins R, et al. Chemical constituents and antibacterial activity of essential oils from flowers and stems of *Ageratum conyzoides* from Ivory Coast. *Records of Natural Products*. 2018;**(2)**:160-168. DOI: 10.25135/rnp.22.17.06.040
- [110] Weimann C, Göransson U, Pongprayoon-Claeson U, Claeson P, Bohlin L, Rimpler H, et al. Spasmolytic effects of *Baccharis conferta* and some of its constituents. *The Journal of Pharmacy and Pharmacology*. 2002;**54**(1):99-104. DOI: 10.1211/0022357021771797
- [111] Romo de Vivar A, Pérez-Castorena AL, Arciniegas A, Villaseñor JL. Secondary metabolites from Mexican species of the tribe Senecioneae (Asteraceae). *Journal of the Mexican Chemical Society*. 2007;**51**(3):160-172 <http://www.scielo.org.mx/pdf/jmcs/v51n3/v51n3a7.pdf>
- [112] Shen Y, Sun Z, Shi P, Wang G, Wu Y, Li S, et al. Anticancer effect of petroleum ether extract from *Bidens pilosa* L and its constituent's analysis by GC-MS. *Journal of Ethnopharmacology*. 2018;**217**:126-133. DOI: 10.1016/j.jep.2018.02.019
- [113] Lima TC, de Jesus Souza R, da Silva FA, Biavatti MW. The genus *Calea* L.: A review on traditional uses, phytochemistry, and biological activities. *Phytotherapy Research*. 2018;**32**(5):769-795. DOI: 10.1002/ptr.6010
- [114] Torrado-Truiti MD, Sarragiotto MH, Abreu-Filho BA, Vataru-Nakamura C, Dias-Filho BP. In vitro antibacterial activity of a 7-O-beta-D-glucopyranosyl-nutanocoumarin from *Chaptalia nutans* (Asteraceae). *Memórias do Instituto Oswaldo Cruz*. 2003;**98**(2):283-286. DOI: 10.1590/S0074-02762003000200020
- [115] Cárdenas-Ortega NC, Zavala-Sánchez MA, Aguirre-Rivera JR, Pérez-González C, Pérez-Gutiérrez S. Chemical composition and antifungal activity of essential oil of *Chrysactinia mexicana* Gray. *Journal of Agricultural*

and Food Chemistry. 2005;**53**(11): 4347-4349. DOI: 10.1021/jf040372h

[116] Guerra-Boone L, Alvarez-Román R, Salazar-Aranda R, Torres-Cirio A, Rivas-Galindo VM, Waksman de Torres N, et al. Chemical compositions and antimicrobial and antioxidant activities of the essential oils from *Magnolia grandiflora*, *Chrysactinia mexicana*, and *Schinus molle* found in Northeast Mexico. Natural Product Communications. 2013;**8**(1):135-138

[117] Zavala-Mendoza D, Grasa L, Zavala-Sánchez MÁ, Pérez-Gutiérrez S, Murillo MD. Antispasmodic effects and action mechanism of essential oil of *Chrysactinia mexicana* A. Gray on rabbit ileum. Molecules. 2016;**21**(6):783. DOI: 10.3390/molecules21060783

[118] Sagrero-Nieves L, Bartley JP. Volatile components from the leaves of *Heterotheca inuloides* Cass. Flavour and Fragrance Journal. 1996;**11**(1):49-51. DOI: 10.1002/(SICI)1099-1026(199601)11:1<49::AID-FFJ538>3.0.CO;2-J

[119] Egas V, Salazar-Cervantes G, Romero I, Méndez-Cuesta CA, Rodríguez-Chávez JL, Delgado G. Anti-*Helicobacter pylori* metabolites from *Heterotheca inuloides* (Mexican arnica). Fitoterapia. 2018;**127**:314-321. DOI: 10.1016/j.fitote.2018.03.001

[120] García-Pérez JS, Cuéllar-Bermúdez SP, Arévalo-Gallegos A, Rodríguez-Rodríguez J, Iqbal H, Parra-Saldivar R. Identification of bioactivity, volatile and fatty acid profile in supercritical fluid extracts of Mexican arnica. International Journal of Molecular Sciences. 2016;**17**(9):1528. DOI: 10.3390/ijms17091528

[121] Borges-Del-Castillo J, Martinez-Martir AI, Rodriguez-Luis F, Rodriguez-Ubis JC, Vazquez-Bueno P. Isolation and synthesis of two coumarins from *Melampodium divaricatum*.

Phytochemistry. 1984;**23**(4):859-861. DOI: 10.1016/S0031-9422(00)85043-8

[122] Rufino-González Y, Ponce-Macotela M, Jiménez-Estrada M, Jiménez-Fragoso CN, Palencia G, Sansón-Romero G, et al. *Piqueria trinervia* as a source of metabolites against *Giardia intestinalis*. Pharmaceutical Biology. 2017;**55**(1):1787-1791. DOI: 10.1080/13880209.2017.1325912

[123] Wollenweber E, Mann K, Arriaga FJ, Yatskievych G. Flavonoids and terpenoids from the leaf resin of *Pluchea odorata*. Zeitschrift für Naturforschung. Section C. 1985;**40**:321-324. DOI: 10.1515/znc-1985-5-607

[124] Vasudevan P, Kashyap S, Sharma S. Tagetes: A multipurpose plant. Bioresource Technology. 1997;**62**:29-35. DOI: 10.1016/S0960-8524(97)00101-6

[125] Pérez-Gutiérrez RO, Hernández-Luna HE, Hernández-Garrido SE. Antioxidant activity of *Tagetes erecta* essential oil. Journal of the Chilean Chemical Society. 2006;**51**(2):883-886. DOI: 10.4067/S0717-97072006000200010

[126] Xu LW, Juan CH, Qi HY, Sshi YP. Phytochemicals and their biological activities of plants in *Tagetes* L. Chinese Herbal Medicines. 2012;**4**(2):103-117. DOI: 10.3969/j.issn.1674-6384.2012.02.004

[127] Camarillo G, Rodríguez C. Biological activity of *Tagetes filifolia* (Asteraceae) on *Trialeurodes vaporariorum* (Hemiptera: Aleyrodidae). Revista Colombiana de Entomología. 2009;**35**(2):177-184 <http://www.scielo.org.co/pdf/rcen/v35n2/v35n2a12.pdf>

[128] Tagne AM, Marino F, Cosentino M. *Tithonia diversifolia* (Hemsl.) A. Gray as a medicinal plant: A comprehensive review of its ethnopharmacology, phytochemistry, pharmacotoxicology

and clinical relevance. *Journal of Ethnopharmacology*. 2018;**220**:94-116. DOI: 10.1016/j.jep.2018.03.025

[129] Santana PI, Osorio MS, Sterner O, García EL. Phytochemical studies of fractions and compounds present in *Vernonanthura patens* with antifungal bioactivity and potential as antineoplastic. In: Rao V, editor. *Phytochemicals—A Global Perspective of their Role in Nutrition and Health*. InTech; 2012. pp. 503-518. DOI: 10.5772/28961

[130] Manzano PI, Miranda M, Abreu-Payrol J, Silva M, Sterner O, Peralta EL. Pentacyclic triterpenoids with antimicrobial activity from the leaves of *Vernonanthura patens* (Asteraceae). *Emirates Journal of Food and Agriculture*. 2013;**1**:539-543. DOI: 10.9755/ejfa.v25i7.15989

[131] Satorres SE, Chiaramello AI, Tonn CE, Laciár AL. Antibacterial activity of organic extracts from *Zinnia peruviana* (L.) against gram-positive and gram-negative bacteria. *Emirates Journal of Food and Agriculture*. 2012;**24**(4): 344-347 <https://www.ejfa.me/index.php/journal/article/view/894>

[132] Lim TK. *Parmentiera aculeata*. In: *Edible Medicinal and Non-Medicinal Plants*. Dordrecht: Springer; 2012. pp. 508-511. DOI: 10.1007/978-90-481-8661-7_67

[133] Abad MJ, Bermejo P, Villar A, Sanchez Palomino S, Carrasco L. Antiviral activity of medicinal plant extracts. *Phytotherapy Research*. 1997;**11**(3):198-202. DOI: 10.1002/(SICI)1099-1573(199705)11:3<198::AID-PTR78>3.0.CO;2-L

[134] Mohan SC, Anand T, Priya RM. Protective effect of *Tecoma stans* flowers on gentamicin-induced nephrotoxicity in rats. *Asian Journal of Biochemistry*. 2016;**11**(1):59-67. DOI: 10.3923/ajb.2016.59.67

[135] Marcotullio M, Curini M, Becerra J. An ethnopharmacological, phytochemical and pharmacological review on lignans from Mexican *Bursera* spp. *Molecules*. 2018;**23**:1976. DOI: 10.3390/molecules23081976

[136] Mujumdar AM, Misar AV. Anti-inflammatory activity of *Jatropha curcas* roots in mice and rats. *Journal of Ethnopharmacology*. 2004;**90**(1):11-15. DOI: 10.1016/j.jep.2003.09.019

[137] Frei B, Heinrich M, Bork PM, Herrmann D, Jaki B, Kato T, et al. Multiple screening of medicinal plants from Oaxaca, Mexico: Ethnobotany and bioassays as a basis for phytochemical investigation. *Phytomedicine*. 1998;**5**(3):177-186. DOI: 10.1016/S0944-7113(98)80025-1

[138] Estrada-Reyes R, Aguirre-Hernández E, García-Argáez A, Soto-Hernández M, Linares E, Bye R, et al. Comparative chemical composition of *Agastache mexicana* subsp. *mexicana* and *A. mexicana* subsp. *xolocotziana*. *Biochemical Systematics and Ecology*. 2004;**32**(7):685-694. DOI: 10.1016/j.bse.2004.01.005

[139] Juárez ZN, Hernández LR, Bach H, Sánchez-Arreola E, Bach H. Antifungal activity of essential oils extracted from *Agastache mexicana* ssp. *xolocotziana* and *Porophyllum linaria* against post-harvest pathogens. *Industrial Crops and Products*. 2015;**74**:178-182. DOI: 10.1016/j.indcrop.2015.04.058

[140] Méndez-Hernández A, Hernández-Hernández AA, López-Santiago MD, Morales-López J, editors. *Herbolaria Oaxaqueña para la Salud*. 1st ed. México; 2009. pp. 1-143

[141] Hernández-Abreu O, Durán-Gómez L, Best-Brown R, Villalobos-Molina R, Rivera-Leyva J, Estrada-Soto S. Validated liquid chromatographic method and analysis of content of tilianin on several extracts obtained

from *Agastache mexicana* and its correlation with vasorelaxant effect. *Journal of Ethnopharmacology*. 2011;**138**(2):487-491. DOI: 10.1016/j.jep.2011.09.041

[142] Zielińska S, Matkowski A. Phytochemistry and bioactivity of aromatic and medicinal plants from the genus *Agastache* (Lamiaceae). *Phytochemistry Reviews*. 2014;**13**(2):391-416. DOI: 10.1007/s11101-014-9349-1

[143] Villa-Ruano N, Pacheco-Hernández Y, Cruz-Durán R, Lozoya-Gloria E. Volatiles and seasonal variation of the essential oil composition from the leaves of *Clinopodium macrostemon* var. *laevigatum* and its biological activities. *Industrial Crops and Products*. 2015;**77**:741-747. DOI: 10.1016/j.indcrop.2015.09.050

[144] Calzada F, Yépez-Mulia L, Aguilar A. In vitro susceptibility of *Entamoeba histolytica* and *Giardia lamblia* to plants used in Mexican traditional medicine for the treatment of gastrointestinal disorders. *Journal of Ethnopharmacology*. 2006;**108**(3):367-370. DOI: 10.1016/j.jep.2006.05.025

[145] Caamal-Herrera IO, Muñoz-Rodríguez D, Madera-Santana T, Azamar-Barrios JA. Identification of volatile compounds in essential oil and extracts of *Ocimum micranthum* Willd leaves using GC/MS. *International Journal of Applied Research in Natural Products*. 2016;**9**(1):31-40 <http://www.ijarnp.org/index.php/ijarnp/article/view/331/pdf>

[146] Pérez-Nicolás M, Vibrans H, Romero-Manzanares A. Can the use of medicinal plants motivate forest conservation in the humid mountains of northern Oaxaca, México? *Botanical Sciences*. 2018;**96**(2):267-285. DOI: 10.17129/botsci.1862

[147] Reyes-Caudillo E, Tecante A, Valdivia-López MA. Dietary fibre

content and antioxidant activity of phenolic compounds present in Mexican chia (*Salvia hispanica* L.) seeds. *Food Chemistry*. 2008;**107**(2):656-663. DOI: 10.1016/j.foodchem.2007.08.062

[148] Porras-Loaiza P, Jiménez-Munguía MT, Sosa-Morales ME, Palou E, López-Malo A. Physical properties, chemical characterization and fatty acid composition of Mexican chia (*Salvia hispanica* L.) seeds. *International Journal of Food Science and Technology*. 2014;**49**(2):571-577. DOI: 10.1111/ijfs.12339

[149] Jiménez-Pérez ND, Lorea-Hernández FG, Jankowski CK, Reyes-Chilpa R. Essential oils in Mexican bays (*Litsea* spp., Lauraceae): Taxonomic assortment and ethnobotanical implications. *Economic Botany*. 2011;**65**(2):178-189. DOI: 10.1007/s12231-011-9160-5

[150] Guzmán-Gutiérrez SL, Gómez-Cansino R, García-Zebadúa JC, Jiménez-Pérez NC, Reyes-Chilpa R. Antidepressant activity of *Litsea glaucescens* essential oil: Identification of β -pinene and linalool as active principles. *Journal of Ethnopharmacology*. 2012;**143**(2):673-679. DOI: 10.1016/j.jep.2012.07.026

[151] Sagrero-Nieves L, Bartley JP. Volatile components of avocado leaves (*Persea americana* mill) from the Mexican race. *Journal of the Science of Food and Agriculture*. 1995;**67**(1):49-51. DOI: 10.1002/jsfa.2740670109

[152] Abe F, Nagafuji S, Okawa M, Kinjo J, Akahane H, Ogura T, et al. Trypanocidal constituents in plants. 5. Evaluation of some Mexican plants for their trypanocidal activity and active constituents in the seeds of *Persea americana*. *Biological & Pharmaceutical Bulletin*. 2005;**28**(7):1314-1317. DOI: 10.1248/bpb.28.1314

[153] Granados-Echegoyen C, Pérez-Pacheco R, Alonso-Hernández N,

- Vásquez-López A, Lagunez-Rivera L, Rojas-Olivos A. Chemical characterization and mosquito larvicidal activity of essential oil from leaves of *Persea americana* mill (Lauraceae) against *Culex quinquefasciatus* (say). Asian Pacific Journal of Tropical Disease. 2015;5(6):463-467. DOI: 10.1016/S2222-1808(15)60816-7
- [154] Mata R, Morales I, Pérez O, Rivero-Cruz I, Acevedo L, Enriquez-Mendoza I, et al. Antimycobacterial compounds from *Piper sanctum*. Journal of Natural Products. 2004;67(12):1961-1968. DOI: 10.1021/np0401260
- [155] Catap ES, Kho MJ, Jimenez MR. In vivo nonspecific immunomodulatory and antispasmodic effects of common purslane (*Portulaca oleracea* Linn.) leaf extracts in ICR mice. Journal of Ethnopharmacology. 2018;215:191-198. DOI: 10.1016/j.jep.2018.01.009
- [156] Argotte-Ramos R, Ramírez-Avila G, Rodríguez-Gutiérrez MD, Ovilla-Muñoz M, Lanz-Mendoza H, Rodríguez MH, et al. Antimalarial 4-phenylcoumarins from the stem bark of *Hintonia latiflora*. Journal of Natural Products. 2006;69(10):1442-1444. DOI: 10.1021/np060233p
- [157] Yahia EM, Gutiérrez-Orozco F, Arvizu-de Leon C. Phytochemical and antioxidant characterization of mamey (*Pouteria sapota* Jacq. HE Moore & Stearn) fruit. Food Research International. 2011;44(7):2175-2181. DOI: 10.1016/j.foodres.2010.11.029
- [158] Vasconcellos MC, Montenegro RC, Militão GC, Fonseca AM, Pessoa OD, Lemos TL, et al. Bioactivity of biflorin, a typical o-naphthoquinone isolated from *Capraria biflora* L. Zeitschrift für Naturforschung. Section C. 2005;60(5-6):394-398. DOI: 10.1515/znc-2005-5-605
- [159] Collins DO, Gallimore WA, Reynolds WF, Williams LA, Reese PB. New skeletal sesquiterpenoids, caprariolides A–D, from *Capraria biflora* and their insecticidal activity. Journal of Natural Products. 2000;63(11):1515-1518. DOI: 10.1021/np000280w
- [160] Imran M, Butt MS, Suleria HA. *Capsicum annuum* bioactive compounds: Health promotion perspectives. In: Jean-Michel M, Ramawat KG, editors. Bioactive Molecules in Food. Cham: Springer; 2018. pp. 1-22. DOI: 10.1007/978-3-319-54528-8_47-1
- [161] Khaled RN, Ćirić A, Glamočlija J, Calhelha RC, Ferreira ICFR, Soković M. Identification of the bioactive constituents and the antibacterial, antifungal and cytotoxic activities of different fractions from *Cestrum nocturnum* L. Jordan Journal of Biological Sciences. 2018;11(3):273-279
- [162] Zoubiri S, Baaliouamer A. GC and GC/MS analyses of the Algerian *Lantana camara* leaf essential oil: Effect against *Sitophilus granarius* adults. Journal of Saudi Chemical Society. 2012;16(3):291-297. DOI: 10.1016/j.jscs.2011.01.013
- [163] Hernández T, García-Bores AM, Serrano R, Ávila G, Dávila P, Cervantes H, et al. Fitoquímica y actividades biológicas de plantas de importancia en la medicina tradicional del Valle de Tehuacán-Cuicatlán. TIP. Revista Especializada en Ciencias Químico-Biológicas. 2015;18(2):116-121. DOI: 10.1016/j.recqb.2015.09.003
- [164] Sousa EO, Miranda CM, Nobre CB, Boligon AA, Athayde ML, Costa JG. Phytochemical analysis and antioxidant activities of *Lantana camara* and *Lantana montevidensis* extracts. Industrial Crops and Products. 2015;70:7-15. DOI: 10.1016/j.indcrop.2015.03.010
- [165] Lin LZ, Mukhopadhyay S, Robbins RJ, Harnly JM. Identification

and quantification of flavonoids of Mexican oregano (*Lippia graveolens*) by LC-DAD-ESI/MS analysis. Journal of Food Composition and Analysis. 2007;**20**(5):361-369. DOI: 10.1016/j.jfca.2006.09.005

[166] Hernández T, Canales M, Avila JG, García AM, Meraz S, Caballero J, et al. Composition and antibacterial activity of essential oil of *Lippia graveolens* HBK (Verbenaceae). Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromaticas. 2009;**8**(4):295-300

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