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# Bean Genome Diversity Reveals the Genomic Consequences of Speciation, Adaptation, and Domestication

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## Abstract

Here we review whether genomic islands of speciation are repeatedly more prone to harbor within-species differentiation due to genomic features, such as suppressed recombination, smaller effective population size, and increased drift, across repeated hierarchically nested levels of divergence. Our discussion focuses on two species of *Phaseolus* beans with strong genepool and population substructure and multiple independent domestications each. We overview regions of species-associated divergence, as well as divergence recovered in within-species between-genepool comparisons and in within-genepool wild-cultivated comparisons. We discuss whether regions with overall high relative differentiation coincide with sections of low SNP density and with between-species pericentric inversions, since these convergences would suggest that shared variants are being recurrently fixed at replicated comparisons, and in a similar manner across different hierarchically nested levels of divergence, likely as the result of genomic features that make certain regions more prone to accumulate islands of speciation as well as within-species divergence. We conclude that neighboring signatures of speciation, adaptation, and domestication in *Phaseolus* beans seem to be influenced by ubiquitous genomic constrains, which may continue shaping, fortuitously, genomic differentiation at various other scales of divergence. This pattern also suggests that genomic regions important for adaptation may frequently be sheltered from recombination.

**Keywords:** genomic islands of speciation, genomic signatures of selection, adaptation, domestication syndrome, convergent evolution, gene flow, genomics constrains, GBS-derived SNP markers

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## 1. Introduction: A strategy to discern among confounding causes of genomic divergence

Genomic signatures associated with species, genepools, and ecotypes' divergence can result from causes other than reduced gene flow, for example, random genetic drift and selection [1]. Moreover, the origin of the outlier variants from novel or standing genetic variation leads to distinctively different patterns of genomic divergence [2–4]. One approach that can help to distinguish these underlying causes of divergence is carrying out a replicated sampling of contrasting populations [5, 6]. If genetic drift rather than selection is responsible for the divergence, it is unlikely that signals of differentiation reappear consistently across replicates [5]. On the other hand, if selection acted on the same genetic variants at the replicated contrasting pairs, genomic regions with comparatively high divergence between individuals from contrasting populations should be identical at each of the replicated populations. Parallel selection on shared genetic variation should therefore lead to low divergence within populations and across replicates, in the exact genomic regions where equivalent variants are selected at each contrasting population [6]. Discerning among gene flow, genetic drift and selection as the cause of parallel genomic divergence are possible as long as there is some degree of replication considered in the sampling of contrasting populations.

The genomic landscape of divergence can also be influenced by differences in ancestral variation and recombination in the genome [7, 8]. Lineage sorting may be enhanced relative to background levels by a reduction in the effective population size ( $N_e$ ) due to processes other than gene flow, like low recombination [8–10]. Since differentiation is further speeded up in low-recombining regions because of linked selection [11–13], the imprint caused by genomic features on the differentiation landscape should be ubiquitous across different levels of divergence. Therefore, besides a replicated sampling of contrasting populations, a hierarchical nested sampling across various scales of divergence is advisable in order to examine whether genomic islands of divergence may display differentiation due to suppressed recombination, smaller effective population size, and increased drift.

In order to discern among confounding causes of genomic divergence in a system with strong population structure and subjected to domestication, we suggest conducting the following analyses by taking advantage of a replicated hierarchical nested sampling across various scales of divergence:

- A. Analyze whether  $F_{ST}$  outliers between species coincide with high  $F_{ST}$  values at within-species comparisons. This pattern is expected if genomic islands of speciation are repeatedly more prone to harbor within-species divergence as a result of limited recombination [8].
- B. Assess whether the within-species between-genepool divergence  $F_{ST}$  profiles are similar among four available comparisons. This trend is expected if the same variants were selected as the result of similar selective pressures at multiple domestication events, but not if divergence outliers were due to population divergence, that is, genetic drift [5].
- C. Assess whether the within-genepool wild-cultivated divergence  $F_{ST}$  profiles are similar among the available comparisons. This coincidence is expected if the same variants are selected as the result of parallel domestication but not if divergence is due to genetic drift [5].

Finally, we suggest exploring if regions of high  $F_{ST}$  co-localized with regions of low  $F_{ST}$  in within-population comparisons.  $\Delta_{Div}$  can be used to analyze the difference between these two  $F_{ST}$  values in each window. Peaks in the  $\Delta_{Div}$  statistic point to genomic regions that diverged as a result of parallel divergence from shared variation rather than due to novel variation evolving at each site [6].

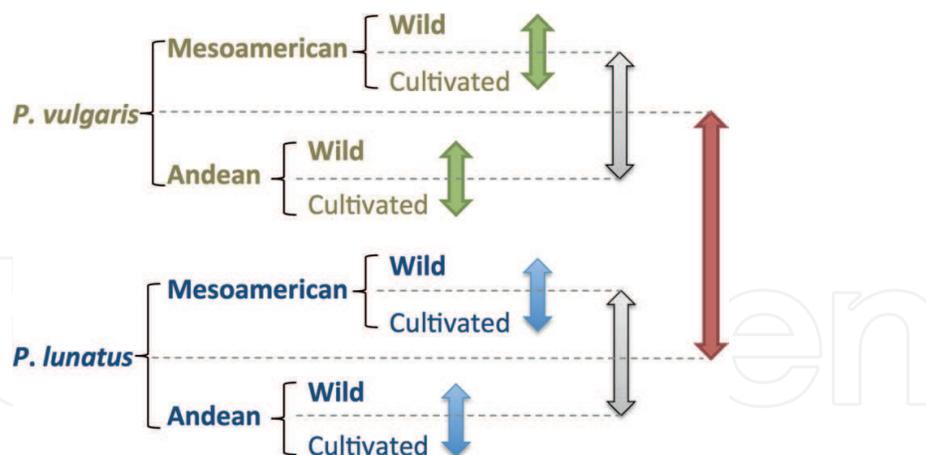
## 2. Beans as a model system to study divergence across various scales of divergence in a replicated hierarchical nested framework

*Phaseolus* beans, with their striking genepool structure and multiple domestications, constitute an excellent model system [14, 15] to the approach described in the previous section and to explore to what extent genomic features, besides reduced gene flow and divergent selection, may lead to genomic divergence between (i.e., speciation islands) and within (i.e., during the natural colonization of new habitats as well as part of the domestication syndromes) species [16]. Common and lima beans are the only bean species with multiple domestications among the five domesticated species of *Phaseolus* [14]. Wild common bean (*P. vulgaris* L.) diverged from its sister species in the tropical Andes [17] and colonized South and Central America from its original distribution in Central America, originating what nowadays is known as the Andean and Mesoamerican genepools. Independent domestications in each genepool gave rise to the Andean and Mesoamerican cultivars [18–20]. On the other hand, wild lima bean (*P. lunatus* L.) diverged from common bean, after which natural spread also led to a strong genepool structure, with two Andean and two Mesoamerican genepools. Further independent domestications happened in one Andean and one Mesoamerican genepools [21].

With this in mind, in this chapter we discuss how the recurrent phylogeographic splits and nested domestication events of common and lima beans help understand whether genomic islands of speciation in *Phaseolus* species are more prone to harbor within-species divergence due to reduced recombination and increased drift (**Figure 1**). We concretely focus our discussion by asking the following questions:

1. Are between-species  $F_{ST}$  outliers recovered in within-species comparisons?
2. Is there any parallelism in the within-species divergence  $F_{ST}$  profiles?
3. Are low-recombining regions (i.e., centromeres) more prone to exhibit divergence across repeated and hierarchically nested scales of divergence?

If there were some parallelisms in the genetic adaptations to the Mesoamerican and Andean environments or in the genetic consequences of the domestication syndromes, then there would be matching signals of differentiation in the within-species between-genepool divergence  $F_{ST}$  profiles and in the within-genepool wild-cultivated divergence  $F_{ST}$  profiles, respectively. These patterns of repeatability would not be observed if between-genepool and wild-cultivated divergence outliers were due to genetic drift [5], if selection pressures were different [22] or if equivalent selective forces did not act on the same shared variation [6, 23].



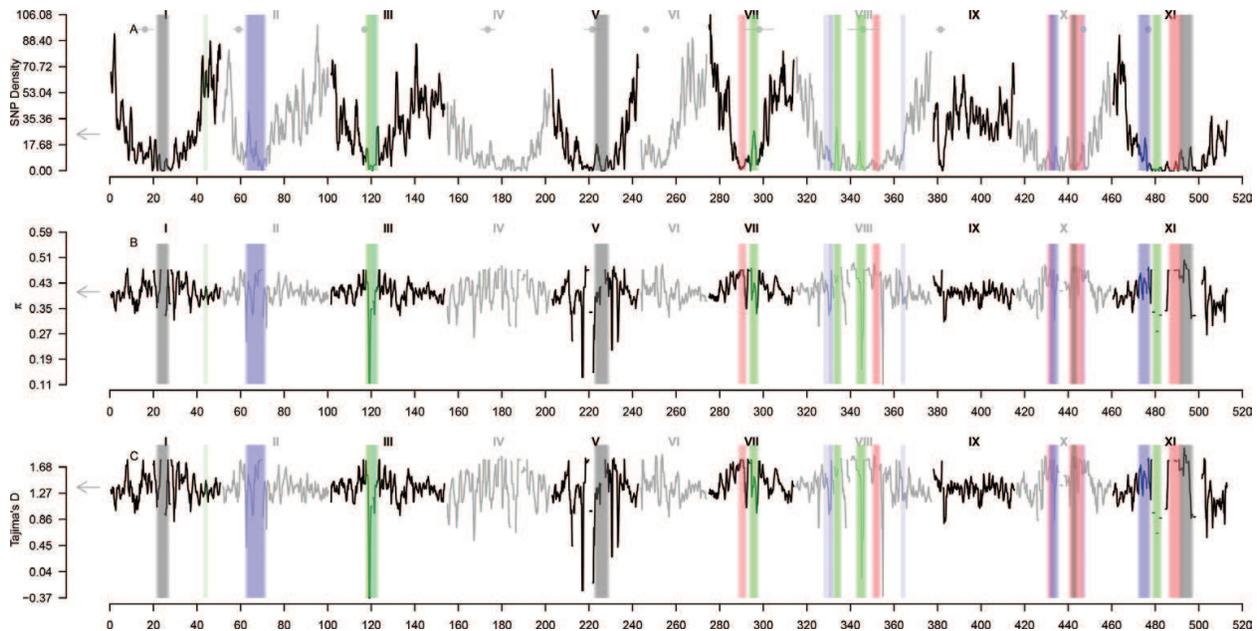
**Figure 1.** Schematic representation of a sampling across hierarchically nested sampling levels of divergence.

Yet, genomic constrains, rather than true signals of convergent adaptation and domestication, could still be the reason for these parallelisms. If genomic features were indeed constraining divergence, then genomic islands of differentiation would coincide with low-recombining regions regardless the nature and the scale of divergence.

### 3. Evidence that genomic features constrain divergence across scales

By looking at the genomic diversity patterns in common and lima beans [24–30], there is evidence that differentiation across repeated and hierarchically nested levels of divergence always co-occurs with regions of low SNP density (**Figure 2**). Increased lineage sorting, and consequently rapid differentiation, is a common phenomenon in low-recombining regions because of linked selection and a reduction in the effective population size [8–10]. Likewise, low-recombining regions also tend to exhibit a decline in diversity due to background selection and, to a lower extent, because of genetic hitchhiking [11]. This can be understood as evidence that regions with low SNP diversity are enriched for contiguous signatures of differentiation between bean species, between genepools, and as part of the multiple domestication syndromes. These concurring signatures could be a by-product of genomic constrains inherent to low-recombining regions.

One of the regions that repeatedly exhibit high differentiation across hierarchically nested levels of divergence in the presence of low SNP density is the centromeric section of chromosome Pv11. The wild-cultivated divergence peak in this chromosome is shared by three domestication syndromes and is located beside the outlier peak detected for all within-species between-genepool comparisons, which in turn coincides with a major between-species peak. In this wide section of chromosome Pv11, there are indications that convergent divergence is consistently correlated with very low SNP density, as expected because of combined effects of linked and background selection in low-recombining regions [8–10, 22]. The observation that genomic constrains are biasing divergence across scales in this section of chromosome Pv11



**Figure 2.** Patterns of genome-wide diversity in common bean and lima beans based on 13,213 GBS-derived SNP markers. A sliding window analysis (window size =  $1 \times 10^7$  bp, step size = 500 kb) was used to compute (A) SNP density, (B) nucleotide diversity as measured by  $\pi$ , and (C) Tajima's D. Vertical translucent boxes highlight the 1 Mb flanking region of each  $F_{ST}$ -based outlier window midpoint when  $F_{ST}$  was computed as follows: (red boxes) between species (*P. lunatus* versus *P. vulgaris*), (gray boxes) between genepools (average of four within-species between-genepool comparisons), (green boxes) between domestication statuses for *P. vulgaris* (average of two within-genepool wild-cultivated comparisons), and (blue boxes) between domestication statuses for *P. lunatus* (average of three within-genepool wild-cultivated comparisons). Results of all windowed analyses are plotted against window midpoints in millions of base pairs (Mb). Black and gray colors highlight different common bean (Pv) chromosomes. Gray arrows on the vertical axes indicate genome-wide averages. Horizontal gray lines with a central-filled gray dot at the top of the figure mark the centromeres [from 20] (figure modified from [16]).

is reinforced by the fact that previous genomic scans did not attribute to this region a consistent outstanding role during the domestication syndromes [20, 21] or in conferring adaptation to different environments and latitudes across the Americas [31]. The only exception is the candidate gene influencing plant size (*Phvul.011G213300*) as part of the Mesoamerican domestication syndrome of common bean [20], but then this pattern has not been consistently reported for the other domestication events as to explain its steady repeatability across hierarchically nested levels of divergence in windows with low SNP density.

Other “hotspots” for spurious divergence due to genomic constrains may be the regions with low SNP density in chromosomes Pv8 and Pv10 that exhibit signatures of between-species divergence as well as repeated between-genepool and within-genepool wild-cultivated divergence (Figure 2). The region in chromosome Pv8 was previously reported to be highly divergent during the domestication of the Andean common bean, but then there were not candidate genes in this region associated with that domestication syndrome in particular [20], despite that the same region is known for being involved in plant and seed growth (i.e., *Phvul.008G168000*) during the Mesoamerican domestication of the same species. This paradox may then be a consequence of genomic constrains obscuring genuine anthropic selection and repeatedly forcing divergence in this region. Similarly, the wide divergent region

in chromosome Pv10, characterized by two outlier peaks split by a “high valley,” actually matches a pericentric inversion between species [32], exemplifying how genomic features inexorably condition differentiation across scales of divergence.

The observation that low-recombining regions are enriched for differentiation across repeated and hierarchically nested levels of divergence in *Phaseolus* beans opposes the profiles of the genome-wide selection scans carried out in common bean. While low-recombining regions are more prone to exhibit signatures of divergence, regions toward the arms of the chromosomes with high SNP density more often harbor adaptive variation [31]. This trend follows expectations because low-recombining regions are more liable to display divergence because of linked selection [11, 33, 34], whereas recombination hotspots usually exhibit higher SNP density and are enriched with functional genes [11, 35]—an already well-described relationship for common bean [36, 37]. Also, adaptive divergent selection usually homogenizes haplotypes within the same niche and fixes polymorphisms in different populations, so that few haplotypes with high frequency remain. This selective process leads to high values of nucleotide diversity and Tajima’s *D* and low values of the Watterson’s  $\theta$  estimator [38], a tendency that was corroborated in wild common bean when looking for adaptive variants [31] but that was lacking in the present study while retrieving the genomic landscape of divergence between species, gene pools, and domestication statuses.

#### 4. Signatures of shared within-species parallel divergence

There is some evidence of some parallelisms in the genetic adaptations to the Mesoamerican and Andean environments in common and lima beans (**Figure 2**). The landscape of genomic adaptation has remained largely unexplored in *Phaseolus* beans. Among the few other studies addressing this question, a panel of wild common bean sampled across the Andean and Mesoamerican ranges revealed that regardless the strength of the bottlenecks [39], the signatures of divergent adaptation are widespread along the genome and coincided with regions of elevated SNP density [31], frequent recombination, and high gene content [36]. However, these surveys have not explicitly addressed the colonization of the Andes by lineages coming from Central America and the corresponding change in selection pressures associated with different altitudes, latitudes, and microenvironments. Topographically complex mountainous systems, such as the Andes, harbor an impressive heterogeneity of climates at a small scale [40–43]. The ridges and valleys constitute physical barriers that limit dispersal and cause local variation in rainfall, resulting in genetic isolation and variation in habitats. Both processes have likely speeded up the evolution of high species diversity in this region [44–48]. Yet, the relative effects of geographic isolation [49–51], environmental variation at a small scale [52–58], and their potential interactions across gene pools remain poorly understood in wild beans. Therefore, characterizing the genomic consequences associated with the colonization of heterogeneous environments may ultimately disclose further cases of genetic parallelism in the adaptation of beans.

The genomic consequences of multiple domestication events are also moderately recurrent as revealed by our survey. From the twelve regions putatively differentiated as the result of the domestication syndrome, only five (42%) appear in more than one comparison but none appears in all. Two peaks in chromosome Pv3 and Pv10 are repeated across three different comparisons

of all five profiles of the domestication syndromes. At least the region in chromosome Pv3 has been reported to be involved in the vernalization pathway (i.e., *Phvul.003G033400*) as part of the Mesoamerican domestication of common bean [20]. Two other divergence peaks in chromosomes Pv8 and Pv11 are consistent across all three genomic profiles of the Mesoamerican domestication syndrome. The region in chromosome Pv8 is known for being related with the encoding of the nitrate reductase (i.e., *Phvul.008G168000*), a critical element for plant and seed growth, during the Mesoamerican domestication of common bean [20]. Also as part of this domestication event, the region in chromosome Pv8 is associated with increased plant size through the ubiquitin ligase degradation pathway (i.e., *Phvul.011G213300*) that controls flower and stem size [20]. More loosely, a peak at chromosome Pv2 in the Mesoamerican common bean domestication  $F_{ST}$  profile is recovered in the profiles of all three lima bean domestications. This region has been linked with the domestication syndrome of lima bean since it is involved in the regulation of seed germination (i.e., *Phvul.002G033500*) and leaf size (i.e., *Phvul.002G041800*) and is enriched by inflated linkage disequilibrium scores [21]. Although scattered, some of these few regions may reveal true parallelisms in the domestication syndromes, whereas others may still be constrained by genomic features.

Also striking is the rarity of regions putatively involved in domestication and shared by several domestication events. This trend, mostly expected for quantitative traits with complex genetic architectures [59–61], had already been noticed for the common bean [20]—potentially applying for lima bean as well [21], and so does not necessarily speak for a prevalent role of drift. Since divergence in the lack of repeatability is a liable result of lineage sorting, caution must be undertaken while interpreting these signals. Singularities may result from different adaptive pressures across the Americas unique to each species, distinctive adaptation to the Mesoamerican microenvironments, dissimilar selection as part of each domestication event [22], equivalent selective forces acting on different genetic variants [6, 23], or genetic drift [5]. Discerning among these causes requires further genotyping in an extended panel specifically addressing each comparison. At least for the divergence peak at chromosome Pv7 in the wild-cultivated Mesoamerican common bean comparison, other drivers besides the domestication itself are an unlikely reason for divergence because a wide region in chromosome Pv7 region is known for being associated with increased seed weight (i.e., *Phvul.007G094299-Phvul.007G.99700*) during the Mesoamerican domestication of common bean [20], as well as with flowering regulation (i.e., *Phvul.007G096500* and *Phvul.007065600*) as part of the domestication of lima bean [21] and both common bean gene pools [20].

## 5. Take-home message

Genomic islands of speciation are not necessarily more prone to harbor within-species divergence, yet subjacent genomic constrains could still be shaping parallel divergence at broader genomic scales. With that in mind, we first discussed how genomic features and linked selection could enhance convergent differentiation in low-recombining regions. Later, we reviewed cases of moderate repeatability in the genomic consequences of multiple adaptation and domestication events. This chapter emphasizes that differentiation across repeated and hierarchically nested levels of divergence co-occurs with regions of low SNP density, and these concurring

signatures may be a by-product of genomic constraints inherent to low-recombining regions. We advise a more systematic use of repeated and hierarchically nested samplings in order to improve our understanding of the underlying causes of the genomic landscape of divergence. Because certain regions are more prone to accumulate islands of divergence as the result of genomic constraints, we advocate that studies of genomic divergence should consider more systematically a dual-purpose sampling, such as the one we described in the first section. In the first place, using replicated populations under presumably similar selection pressures helps accounting for lineage sorting and characterizing the nature of the selected variants, i.e., novel versus standing [6]. Second, a hierarchically nested sampling across various levels of divergence allows for further assessments on the processes, which like genomic constraints, may give rise to parallel divergence patterns [2–4, 62]. Finally, some of these examinations must be verified with genomic features and estimates of the recombination rate [63–65]. We foresee that as the evidence of pervasive genomic constraints shaping genomic differentiation across species and at countless scales of divergence accumulates, replicated samplings of contrasting populations in a hierarchically nested framework of divergence will become indispensable.

In the long run, we are looking forward to see more coherent and systematic samplings of replicated contrasting populations across hierarchically nested levels of divergence in of genomic divergence has always been challenging, but the field is now moving forward toward a more cohesive framework. New ways [66, 67] to characterize obscuring genomic features promise aiding our understanding on how the genomic landscape of divergence is shaped.

Among the five domesticated species in the *Phaseolus* genus, common and lima beans are the only ones exhibiting range expansions toward South American and multiple domestications [14]. However, exploring the landscape of divergence in other domesticated *Phaseolus* species is equally insightful because of their overlapping distribution ranges, nested phylogenetic relationships, and divergent adaptations. For instance, year (*P. dumosus*) and runner (*P. coccineus*) beans are Mesoamerican and well adapted to humid habitats, which makes them a potential source of resistance to biotic stresses. On the other hand, tepary bean (*P. acutifolius*) is also Mesoamerican but is well known for growing in desert and semiarid environments, which makes it a likely source of tolerance to abiotic stresses. These species also possess well-established genomic resources [68] that could speed up newer genome-wide comparisons. *Phaseolus* species that never underwent domestication are also abundant (ca. 70) and could enrich our understanding of genomic divergence in this intricate complex. Considering the *Phaseolus* species complex as a whole will ultimately reinforce beans as a model for understanding speciation, adaptation, and crop evolution [14, 15, 69–72].

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## References

- [1] Nei M. The neutral theory of molecular evolution in the genomic era. *Annual Review of Genomics and Human Genetics*. 2010;**11**:265-289
- [2] Barrett RD, Schluter D. Adaptation from standing genetic variation. *Trends in Ecology & Evolution*. 2008;**23**(1):38-44
- [3] Pritchard JK, Pickrell JK, Coop G. The genetics of human adaptation: Hard sweeps, soft sweeps, and polygenic adaptation. *Current Biology*. 2010;**20**(4):R208-R215
- [4] Hermisson J, Pennings PS. Soft sweeps: Molecular population genetics of adaptation from standing genetic variation. *Genetics*. 2005;**169**(4):2335-2352
- [5] Lotterhos KE, Whitlock MC. The relative power of genome scans to detect local adaptation depends on sampling design and statistical method. *Molecular Ecology*. 2015;**24**(5):1031-1046
- [6] Roesti M et al. The genomic signature of parallel adaptation from shared genetic variation. *Molecular Ecology*. 2014;**23**(16):3944-3956

- [7] Strasburg JL et al. What can patterns of differentiation across plant genomes tell us about adaptation and speciation? *Philosophical Transactions of the Royal Society, B: Biological Sciences*. 2011;**367**(1587):364-373
- [8] Wolf JB, Ellegren H. Making sense of genomic islands of differentiation in light of speciation. *Nature Reviews. Genetics*. 2017;**18**(2):87-100
- [9] Jones FC et al. The genomic basis of adaptive evolution in three spine sticklebacks. *Nature*. 2012;**484**(7392):55-61
- [10] Zhou L, Bawa R, Holliday JA. Exome resequencing reveals signatures of demographic and adaptive processes across the genome and range of black cottonwood (*Populus trichocarpa*). *Molecular Ecology*. 2014;**23**(10):2486-2499
- [11] Ellegren H, Galtier N. Determinants of genetic diversity. *Nature Reviews. Genetics*. 2016;**17**(7):422-433
- [12] Kelleher CT, Wilkin J, Zhuang J, Cortés AJ, Quintero ÁLP, Gallagher TF, et al. SNP discovery, gene diversity, and linkage disequilibrium in wild populations of *Populus tremuloides*. *Tree Genetics & Genomes*. 2012;**8**(4):821-829. DOI: 10.1007/s11295-012-0467-x
- [13] Cortés AJ. On The Big Challenges of a Small Shrub: Ecological Genetics of *Salix herbacea* L. Uppsala: Acta Universitatis Upsaliensis; 2015
- [14] Bitocchi E et al. Beans (*Phaseolus* spp.) as a model for understanding crop evolution. *Frontiers in Plant Science*. 2017;**8**:722
- [15] Broughton WJ et al. Beans (*Phaseolus* spp.)—model food legumes. *Plant and Soil*. 2003;**252**(1):55-128
- [16] Cortés AJ et al. Does the genomic landscape of species divergence in *Phaseolus* beans reveal parallel signatures of adaptation and domestication? *Frontiers in Plant Science*. In revision
- [17] Rendon-Anaya M et al. Genomic history of the origin and domestication of common bean unveils its closest sister species. *Genome Biology*. 2017;**18**(1):60
- [18] Gepts P, Debouck D. Origin, domestication and evolution of the common bean (*Phaseolus vulgaris* L.). In: Van Shoonhoven A, Voysest O, editors. *Common Beans: Research for Crop Improvement*. Wallingford: Commonwealth Agricultural Bureau; 1991. pp. 7-53
- [19] Kwak M, Gepts P. Structure of genetic diversity in the two major gene pools of common bean (*Phaseolus vulgaris* L., *Fabaceae*). *Theoretical and Applied Genetics*. 2009;**118**(5): 979-992
- [20] Schmutz J et al. A reference genome for common bean and genome-wide analysis of dual domestications. *Nature Genetics*. 2014;**46**(7):707-713
- [21] Chacon-Sanchez MI, Martinez-Castillo J. Testing domestication scenarios of lima bean (*Phaseolus lunatus* L.) in Mesoamerica: Insights from genome-wide genetic markers. *Frontiers in Plant Science*. 2017;**8**:1551

- [22] Ravinet M et al. Interpreting the genomic landscape of speciation: A road map for finding barriers to gene flow. *Journal of Evolutionary Biology*. 2017;**30**(8):1450-1477
- [23] Ravinet M et al. Shared and nonshared genomic divergence in parallel ecotypes of *Littorina saxatilis* at a local scale. *Molecular Ecology*. 2016;**25**(1):287-305
- [24] Cortés AJ, Chavarro MC, Blair MW. SNP marker diversity in common bean (*Phaseolus vulgaris* L.). *Theoretical and Applied Genetics*. 2011;**123**(5):827-845
- [25] Blair MW, Soler A, Cortés AJ. Diversification and population structure in common beans (*Phaseolus vulgaris* L.). *Plos One*. 2012;**7**(11):e49488
- [26] Cortés AJ, Chavarro MC, Madriñán S, This D, Blair MW. Molecular ecology and selection in the drought-related *Asr* gene polymorphisms in wild and cultivated common bean (*Phaseolus vulgaris* L.). *BMC Genetics*
- [27] Cortés AJ, This D, Chavarro C, Madriñán S, Blair MW. Nucleotide diversity patterns at the drought-related *DREB2* encoding genes in wild and cultivated common bean (*Phaseolus vulgaris* L.). *Theoretical and Applied Genetics*. 2012;**125**(5):1069-1085
- [28] Galeano CH, Cortés AJ, Fernandez AC, Soler A, Franco-Herrera N, Makunde G, et al. Gene-based single nucleotide polymorphism markers for genetic and association mapping in common bean. *BMC Genetics*. 2012;**13**(1):48
- [29] Blair MW, Cortés AJ, Penmetza RV, Farmer A, Carrasquilla-Garcia N, Cook DR. A high-throughput SNP marker system for parental polymorphism screening, and diversity analysis in common bean (*Phaseolus vulgaris* L.). *Theoretical and Applied Genetics*. 2013;**126**(2):535-548
- [30] Blair MW, Cortés AJ, This D. Identification of an *ERECTA* gene and its drought adaptation associations with wild and cultivated common bean. *Plant Science*. 2016;**242**:250-9
- [31] Cortés AJ, Blair MW. Genotyping by sequencing and genome—Environment associations in wild common bean predict widespread divergent adaptation to drought. *Frontiers in Plant Science*. 2018;**9**:128
- [32] Bonifácio EM et al. Comparative cytogenetic mapping between the lima bean (*Phaseolus lunatus* L.) and the common bean (*P. vulgaris* L.). *Theoretical and Applied Genetics*. 2012;**124**(8):1513-1520
- [33] Ellegren H, Wolf JBW. Parallelism in genomic landscapes of differentiation, conserved genomic features and the role of linked selection. *Journal of Evolutionary Biology*. 2017;**30**(8):1516-1518
- [34] Ellegren H et al. The genomic landscape of species divergence in *Ficedula* flycatchers. *Nature*. 2012;**491**(7426):756-760
- [35] Rodgers-Melnick E et al. Recombination in diverse maize is stable, predictable, and associated with genetic load. *Proceedings of the National Academy of Sciences*. 2015;**112**(12):3823-3828

- [36] Blair MW et al. Uneven recombination rate and linkage disequilibrium across a reference SNP map for common bean (*Phaseolus vulgaris* L.). PLoS One. 2018;**13**(3):e0189597
- [37] Bhakta MS, Jones VA, Vallejos CE. Punctuated distribution of recombination hotspots and demarcation of pericentromeric regions in *Phaseolus vulgaris* L. PLoS One. 2015;**10**(1): e0116822
- [38] Wakeley J. Coalescent Theory: An Introduction. Cambridge: Harvard University; 2008
- [39] Ariani A, Berny Mier YTJ, Gepts P. Spatial and temporal scales of range expansion in wild *Phaseolus vulgaris*. Molecular Biology and Evolution. 2018;**35**(1):119-131
- [40] Cortés AJ, Blair MW. Naturally available genetic adaptation in common bean and its response to climate change. In: Srinivasarao C, Shanker AK, Shanker C, editors. Climate Resilient Agriculture—Strategies and Perspectives. Rijeka, Croatia: InTech; 2018
- [41] Cortés AJ, Wheeler JA. The environmental heterogeneity of mountains at a fine scale in a changing world. In: Hoorn C, Perrigo A, Antonelli A, editors. Mountains, Climate, and Biodiversity. NY: Wiley; 2018
- [42] Cortés AJ. Environmental Heterogeneity at a Fine Scale: Ecological and Genetic Implications in a Changing World. Saarbrücken, Germany: LAP Lambert Academic Publishing; 2016. 60 p
- [43] Cortés AJ. Local scale genetic diversity and its role in coping with changing climate. In: Bitz L, editor. Genetic Diversity. Rijeka, Croatia: InTech; 2017. p. 140 pp
- [44] Hoorn C et al. Amazonia through time: Andean uplift, climate change, landscape evolution, and biodiversity. Science. 2010;**330**(6006):927-931
- [45] Antonelli A et al. Tracing the impact of the Andean uplift on Neotropical plant evolution. Proceedings of the National Academy of Sciences. 2009;**106**(24):9749-9754
- [46] Madriñán S, Cortés AJ, Richardson JE. Páramo is the world's fastest evolving and coolest biodiversity hotspot. Frontiers in Genetics. 2013;**4**:192
- [47] Zizka A, Antonelli A. Mountains of diversity. Nature. 2018;**555**:173-174
- [48] Arnegard ME et al. Genetics of ecological divergence during speciation. Nature. 2014;**511**(7509):307-311
- [49] Cortés AJ, Waeber S, Lexer C, Sedlacek J, Wheeler JA, van Kleunen M, et al. Small-scale patterns in snowmelt timing affect gene flow and the distribution of genetic diversity in the alpine dwarf shrub *Salix herbacea*. Heredity. 2014;**113**:233-239
- [50] Cortés AJ, Liu X, Sedlacek J, Wheeler JA, Lexer C, Karrenberg S. Maintenance of female-bias in a polygenic sex determination system is consistent with genomic conflict. On The Big Challenges of a Small Shrub: Ecological Genetics of *Salix herbacea* L. Uppsala: Acta Universitatis Upsaliensis; 2015

- [51] Cortés AJ, Wheeler JA, Sedlacek J, Lexer C, Karrenberg S. Genome-wide patterns of microhabitat-driven divergence in the alpine dwarf shrub *Salix herbacea* L. On The Big Challenges of a Small Shrub: Ecological Genetics of *Salix herbacea* L. Uppsala: Acta Universitatis Upsaliensis; 2015
- [52] Sedlacek J, Bossdorf O, Cortés AJ, Wheeler JA, van-Kleunen M. What role do plant-soil interactions play in the habitat suitability and potential range expansion of the alpine dwarf shrub *Salix herbacea*? *Basic and Applied Ecology*. 2014;**15**(4):305-315
- [53] Wheeler JA, Hoch G, Cortés AJ, Sedlacek J, Wipf S, Rixen C. Increased spring freezing vulnerability for alpine shrubs under early snowmelt. *Oecologia*. 2014;**175**(1):219-229. DOI: <https://doi.org/10.1007/s00442-013-2872-8>
- [54] Sedlacek J, Wheeler JA, Cortés AJ, Bossdorf O, Hoch G, Lexer C, et al. The response of the alpine dwarf shrub *salix herbacea* to altered snowmelt timing: Lessons from a multi-site transplant experiment. *PloS One*. 2015;**10**(4):e0122395
- [55] Wheeler JA, Schnider F, Sedlacek J, Cortés AJ, Wipf S, Hoch G, et al. With a little help from my friends: community facilitation increases performance in the dwarf shrub *Salix herbacea*. *Basic and Applied Ecology*. 2015;**16**:202-229
- [56] Little CJ, Wheeler JA, Sedlacek J, Cortés AJ, Rixen C. Small-scale drivers: the importance of nutrient availability and snowmelt timing on performance of the alpine shrub *Salix herbacea*. *Oecologia*. 2016;**180**(4):1015-1024
- [57] Sedlacek J, Cortés AJ, Wheeler JA, Bossdorf O, Hoch G, Klapste J, et al. Evolutionary potential in the Alpine: Trait heritabilities and performance variation of the dwarf willow *Salix herbacea* from different elevations and microhabitats. *Ecology and Evolution*. 2016;**6**(12):3940-3952
- [58] Wheeler JA, Cortés AJ, Sedlacek J, Karrenberg S, van Kleunen M, Wipf S, et al. The snow and the willows: Accelerated spring snowmelt reduces performance in the low-lying alpine shrub *Salix herbacea*. *Journal of Ecology*. 2016;**104**(4):1041-1050
- [59] Meyer RS, DuVal AE, Jensen HR. Patterns and processes in crop domestication: An historical review and quantitative analysis of 203 global food crops. *The New Phytologist*. 2012;**196**(1):29-48
- [60] Blair MW, Iriarte G, Beebe S. QTL analysis of yield traits in an advanced backcross population derived from a cultivated Andean x wild common bean (*Phaseolus vulgaris* L.) cross. *Theoretical and Applied Genetics*. 2006;**112**(6):1149-1163
- [61] Cortés AJ, Monserrate F, Ramírez-Villegas J, Madriñán S, Blair MW. Drought tolerance in wild plant populations: The case of common beans (*Phaseolus vulgaris* L.). *Plos One*. 2013;**8**(5):e62898
- [62] Pereira RJ et al. Transcriptome-wide patterns of divergence during allopatric evolution. *Molecular Ecology*. 2016;**25**(7):1478-1493

- [63] Maynard Smith J, Haigh J. The hitch-hiking effect of a favourable gene. *Genetics Research*. 1974;**23**(01):23-35
- [64] Nielsen R. Molecular signatures of natural selection. *Annual Review of Genetics*. 2005;**39**: 197-218
- [65] Storz JF. Using genome scans of DNA polymorphism to infer adaptive population divergence. *Molecular Ecology*. 2005;**14**:671-688
- [66] Cortés AJ. On how role versatility boosts an STI. *Journal of Theoretical Biology*. 2018;**440**: 66-69
- [67] Cortés AJ. Prevalence in MSM Is Enhanced by Role Versatility. In: Mazari A, editor. *Big Data Analytics in HIV/AIDS Research*. Hershey, USA: IGI Global; 2018
- [68] Guerra-Garcia A et al. Domestication genomics of the open-pollinated scarlet runner bean (*Phaseolus coccineus* L.). *Frontiers in Plant Science*. 2017;**8**:1891
- [69] Cortés AJ. On the origin of the common bean (*Phaseolus vulgaris* L.). *American Journal of Plant Sciences*. 2013;**4**(10):1998-2000
- [70] Irwin DE et al. Recurrent selection explains parallel evolution of genomic regions of high relative but low absolute differentiation in a ring species. *Molecular Ecology*. 2016;**25**(18):4488-4507
- [71] Nachman MW, Payseur BA. Recombination rate variation and speciation: Theoretical predictions and empirical results from rabbits and mice. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*. 2012;**367**(1587):409-421
- [72] Cruickshank TE, Hahn MW. Reanalysis suggests that genomic islands of speciation are due to reduced diversity, not reduced gene flow. *Molecular Ecology*. 2014;**23**(13):3133-3157