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Hypersaline Actinomycetes and Their Biological Applications

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Abstract

Actinomycetes are the potential sources of novel metabolites, therapeutic compounds, enzymes, and other chemicals. Among them, the applications of halophilic actinomycetes toward the medically and industrially important metabolites and enzymes are gaining increasing attention by the scientific community. A large number of novel compounds and enzymes from halophilic actinomycetes have been isolated and characterized from various geographic regions around the world. In this chapter, occurrence, characterization, halotolerant mechanisms, medical importance, metabolites, enzymes, and industrial applications of halophilic actinomycetes are discussed. Halophilic actinomycetes may also serve as good models for the production of important metabolites and enzymes with respect to stress response.

Keywords: Halophilic actinomycetes, occurrence, metabolites, enzymes, applications

1. Introduction

Actinomycetes are the most valuable microorganisms for the production and synthesis of economically important therapeutic compounds and antibiotics. They are the source for the production of about more than 50% of discovered bioactive compounds, including antitumor agents, antibiotics, enzymes, and immunosuppressive agents [1, 2]. Most of these bioactive secondary metabolites were isolated from terrestrial actinomycetes; however, in recent

years, the rate of discovery of new bioactive compounds has decreased. Hence, it is crucial that new groups of actinomycetes from unexplored or underexploited environments should be exploited optimally to obtain novel bioactive secondary metabolites [3]. Unexplored regions of high salt deposited areas, conditions closely related to the marine water regions, regions with increased levels of pH, and low oxygen were the suitable places to isolate halophilic actinomycetes. Actinomycetes recovered from halophilic and unexplored regions attracted the interest of researchers because of their applications in the medical and biotechnological fields [4]. This chapter deals with the characterization methods for the identification of halophilic actinomycetes and their applications

2. Occurrence of halophilic actinomycetes

Marine sediment, soil, water, contaminated regions on the seashore, salt lakes, saline soils, alkaline-saline habitats, brines, and other regions are good sources for the selection of novel halophilic actinomycetes. Recent literature claimed that the saline regions contain many significant uncultured actinomycetes [5]. In particular, actinomycetes such as *Streptomyces pharamarensis*, *Prauserella halophila*, *Pseudonocardia* sp., *Salinispora arenicola*, *Aeromicrobium* sp., *Micromonospora* sp., *Marinactinospora thermotolerans*, *Microbacterium* sp., *Nocardiosis xinjiangensis*, *Salinibacterium* sp., *Salinactinospora qingdaonensis*, *Rhodococcus* sp., *Actinomadura* sp., *Saccharopolyspora* sp., *Streptosporangium* sp., *Actinopolyspora algeriensis*, *Marinophilus* sp., *Streptomonospora halophila*, *Verrucosisporea* sp., *Gordonia* sp., and *Nonomuraea* species were recovered and characterized from hypersaline regions [6–10]. Also, it was quoted that the characterization of novel halotolerant actinomycetes was specific with respect to their morphological, biochemical, physiological, and molecular level identification.

3. Identification and characterization of halophilic actinomycetes

In general, different pretreatment methods such as incorporation of specific antibiotics were used to prevent the growth of bacteria and fungi, and alteration in the composition of the cultivation medium was specific for the isolation of rare actinomycetes. However, humic acid and different concentrations of vitamin agar medium were the best base for the selection of rare halophilic actinomycetes. Humic acid strengthens the morphological identification and characterizes the spore chain of the actinomycetes by preventing the production of diffusible pigments in the cultivation medium. A substantial part of the microbes in environmental samples are not cultivable and therefore cannot be identified by methods based on culturing of the strains. However, protocols based on molecular techniques are not dependent on the viability of the microbes, and molecular methods for the amplification of specific genes such as 16S rRNA and *rec A* can be designed to work at different levels of specificity for the detection of whole groups of actinomycetes. Polymerase chain reaction (PCR)-based methods for the detection and identification of microbes are widely used in the cultivation of industrially important microbes [11]. Applications in environmental microbi-

ology, especially in the soil environment, are also increasing [12]. With currently used cultivation methods, only a small part of the halophilic actinomycetes diversity is detected. The cultivability values ranged from 0.001% to 15%, depending on the cultivation area and the method of cultivation medium [13]. On the other hand, PCR amplification of 16S rRNA genes from environmental samples has revealed that 7–64% of the amplified sequences originated from uncultured actinomycetes [14]. Comparisons of the amounts of total and cultivable microbes in the indoor environment have shown that the cultivable part of the microbial community ranges from 1% to 10% [15]. Therefore, careful attention must be paid in the identification of the rare halophilic actinomycetes.

4. Halotolerance mechanism

Halophilic actinomycetes require higher concentrations of salt for their growth and are classified into moderate (15% NaCl) and extreme (30% NaCl) halophiles. They survive through two mechanisms – “high-salt-in” and “low-salt, organic-solutes-in” – for the protection of intercellular proteins in the presence of salts similar to potassium chloride and production of organic acids, which will directly alter the intracellular enzyme levels. To adjust to the hyper-salt conditions, halophilic cell membranes are composed of main adaptive systems that block NaCl from penetrating into the cells, such as the accumulation of inorganic salts and the water-soluble low-molecular-weight organic compounds. Inorganic salts in the form of (Na^+ , K^+ , Cl^-) are mainly involved in adjusting the osmotic potentials, and the organic salts in the form of small solutes or electrolytes are concerned with the balance of the intracellular salt levels [16]. These two mechanisms are mainly involved in the maintenance of the cell structure; therefore, the cells thrive in stress conditions such as high salt [17].

5. Importance of halophilic actinomycetes

Actinomycetes identified in the halophilic regions such as salt lakes, salterns, solar salts, and subsurface salt formation have to cope up with the osmotic stress were an infinite pool of novel chemical molecules with increasing importance for several biotechnological applications in different fields. Traditionally, halophiles have been used in the food and nutraceutical industries for the fermentation of soy and fish sauces and β -carotene production; also they have been recently used in many novel and unique molecules such as compatible solutes, biopolymers or carotenoids, enzymes, biodegradable plastics, biosurfactants, bioemulsifiers, and bacteriorhodopsins for molecular biotechnology applications [18, 19]. Among the valuable products, enzymes obtained from these organisms, mainly involved in the processing of food, act as a catalyst in the bioremediation process and other biosynthetic processes. In addition to the enzymes, highly sensitive chromoproteins derived from halophiles act as a biocomputing and light-sensitive neurological probe for the treatment of blindness [20]. Ectoine and hydroxyectoine obtained from the halophilic actinomycetes are commercially used as protective and stabilizing agents for mammalian cells [21]. Ectoine derived from

Halomonas boliviensis fermentation reduces the DNA lesions induced by visible and UVA-visible lights. Besides, halophiles attracted many researchers for the development of sustainable energy production to minimize the effects of global warming. Because halophiles have superior qualities such as high tolerance levels of the enzymes toward salts and temperatures, stability in the presence of organic solvents, secretion of therapeutic compounds, and antimicrobial properties, in many applications, studies of halophilic bacteria have developed rapidly and significant advances have been made recently [22].

6. Metabolites produced by halophilic actinomycetes

In the last two decades, halophilic actinomycetes have gained significant importance as a new, promising source for novel bioactive compounds that can be used for drug development. Halophilic actinomycetes may produce a variety of bioactive compounds, which have a wide range of biological activities, including antibacterial, antifungal, antiviral, and other bioactive compounds [23]. The vast majority of bioactive compounds that isolated from halophilic actinomycetes are derived from the genus *Streptomyces*, whose species are very widely distributed in nature and cover around 80% of the total produced antibiotics [24].

7. Antibacterial

An antibacterial is an agent that may either inhibit the bacterial growth or kill the bacteria. The urgent need to find novel antibacterial agents has increased during the last decade owing to the increase of emerging multidrug-resistant bacteria. The discovery of novel antibiotics that have a potent effect against resistant pathogenic bacteria is an important aspect of antibiotics research today. The diversity of natural products makes it one of the most important sources for novel structures, which have been noticed to possess useful biological activities. Several studies are oriented toward the isolation of new actinomycetes from different habitats in the context of the search for novel antibiotics from new sources representing one of these habitats [25].

Generally, the antibacterial activity of halophilic actinomycetes from marine environments is extensively studied. Nevertheless, the exploitation of halophilic actinomycetes as a source for the discovery of novel antibiotics is still at an early stage, despite that numerous novel antibacterial compounds were isolated during the last few years (Table 1). Arenimycin (Fig. 1a) is a novel antibacterial compound isolated from the extreme halophilic actinomycete *Salinispora arenicola*. This compound was classified as a new antibiotic based on the novel structure, which belongs to the benzo [α] naphthacene quinone class of antibiotics. Arenimycin has proven to show a potent antibacterial activity against a panel of drug-resistant human pathogens such as rifampin- and methicillin-resistant *Staphylococcus aureus*. Arenimycin is a representative of the first report of this class of antibiotics from marine actinomycetes [26]. Abyssomicin C (Fig. 1b) is another example for novel antibacterial compounds extracted

from the halophilic actinomycete *Verrucosispora* sp. [27]. Abyssomicin C was derived with other two novel structures that belong to the polycyclic polyketides abyssomicin B and abyssomicin D. Among these three compounds, abyssomicin C strongly exhibits antibacterial activity against Gram-positive bacteria, including multiresistant clinical isolates of *Staphylococcus aureus*. The mode of action of this new antibiotic is based on the inhibition of *para*-aminobenzoic acid biosynthesis resulting in the inhibition of the folic acid biosynthesis pathway.

Figure 1. Chemical structure of some novel antibacterial compounds produced by halophilic actinomycetes [26, 27].

Compound	Source
1,4-Dihydroxy-2-(3-hydroxybutyl)-9,10-anthraquinone 9,10-anthrac	<i>Streptomyces</i> sp.
1-Hydroxy-1-norresistomycin	<i>Streptomyces chinaensis</i>
Abyssomicins	<i>Verrucosispora</i> sp.
Arenimycin	<i>Salinispora arenicola</i>
Bisanthraquinone	<i>Streptomyces</i> sp.
Bonactin	<i>Streptomyces</i> sp.
Caboxamycin	<i>Streptomyces</i> sp.
Chloro-dihydroquinones	<i>Streptomyces</i> sp.
Diazepinomicin	<i>Micromonospora</i> sp.
Essramycin	<i>Streptomyces</i> sp.
Frigocyclinone	<i>Streptomyces griseus</i>
Glaciapyrroles	<i>Streptomyces</i> sp.
Gutingimycin	<i>Streptomyces</i> sp.

Compound	Source
Helquinoline	<i>Janibacter limosus</i>
Himalomycins	<i>Streptomyces</i> sp.
Lajollamycin	<i>Streptomyces nodosus</i>
Lincomycin	<i>Streptomyces lincolnensis</i>
Lynamicins	<i>Marinispora</i> sp.
Marinomycins	<i>Marinispora</i> sp.
Marinopyrroles	<i>Streptomyces</i> sp.
Proximicins	<i>Verrucosispora</i> sp.
Resistoflavin methyl ether)	<i>Streptomyces</i> sp.
Tirandamycins	<i>Streptomyces</i> sp.
TP-1161	<i>Nocardioopsis</i> sp.

The listed compounds and the strains were obtained from the report of Subramani and Aalbersberg, (2012) [36].

Table 1. Novel antibacterial compounds produced by halophilic actinomycetes

8. Antifungal

The number of antifungal agents available for controlling fungal infections is still limited in comparison to antibacterial agents, and their use is still risky owing to toxicity and side effects [28]. Therefore, studies are still being conducted to isolate and identify novel antifungals that are potentially effective against pathogenic fungi [23]. Halophilic actinomycetes from marine environments are useful biological tools for the discovery of novel antifungal substances against fungi. Few studies reported the isolation of antifungal agents from halophilic actinomycetes. *Streptomyces* is the main source for antifungal agents (Table 2) which have been isolated from marine habitats.

Compound	Source
Azalomycin F4a 2-ethylpentyl ester	<i>Streptomyces</i> sp.
Bonactin	<i>Streptomyces</i> sp.
Chandrananimycin	<i>Actinomadura</i> sp.
Daryamides	<i>Streptomyces</i> sp.
N-(2-hydroxyphenyl)-2-phenazinamine (NHP)	<i>Nocardia dassonvillei</i>

The listed compounds and the strains were obtained from the report of Subramani and Aalbersberg, (2012) [36].

Table 2. Novel antifungal agents produced by halophilic actinomycetes

Chandrananimycin A (Fig. 2a) is a novel antifungal substance produced by marine *Actinomadura* sp. strain M048. Chandrananimycin A exhibited strong, effective antifungal activity against *Mucor miehei*. It exhibits other biological activities as an antibacterial, anticancer, and antialgal agent against the microalgae [29]. *N*-(2-hydroxyphenyl)-2-phenazamine (NHP) (Fig. 2b) is a new antibiotic produced by halophilic actinomycetes isolated from the sediment sample *Nocardia dassonvillei* collected from the Arctic Ocean [30]. The new antibiotics possess potent antifungal activity against *Candida albicans*, with an MIC of 64 µg /ml.

Figure 2. Chemical structure of novel antifungal isolated from halophilic actinomycetes [30].

9. Antiviral

Natural products represent the main source for discovering new/novel chemical structures that are used for the treatment of infections. The vast majority of secondary metabolites produced by microbes that have been developed for controlling microbial infections are directed against bacterial and fungal infections but not against viral infections. To date, antiviral agents have been isolated from natural products that are limited, and the studies in this field are few [31]. The antiviral agents available commercially in pharmacological markets are over 40 compounds, including those being tested as promising antiviral agents or alternative antiviral medicines [32]. Potential substances that possess antiviral activity have been isolated from halophilic microorganisms from marine environments. EPS-1 and EPS-2 are antiviral agents produced by *Bacillus licheniformis* and *Geobacillus thermodenitrificans*, respectively, targeting the replication of herpes simplex virus type 2 (HSV-2) [33]. Cyanovirin-N was obtained from the halophilic cyanobacteria *Nostoc ellipsosporum*. Cyanovirin-N has antiviral activity against HIV-1 and HIV-2 by targeting the replication [34]. However, halophilic actinomycetes produce several bioactive compounds, the compounds derived from halophilic actinomycetes as antiviral agents are limited. Antimycin A is an antiviral substance produced by *Streptomyces kaviengensis* isolated from marine sediments collected from the coast of New Ireland, Papua New Guinea. Antimycin A has potent antiviral activity against western equine encephalitis virus and a wide range of RNA viruses in cultured cells, includ-

ing members of the *Togaviridae*, *Flaviviridae*, *Bunyaviridae*, *Picornaviridae*, and *Paramyxoviridae* families. Benzastatin C (Fig. 3) is a 3-chloro-tetrahydroquinolone alkaloid extracted from the halophilic actinomycete *Streptomyces nitrosporeus* [35]. This compound exhibited antiviral activity against herpes simplex virus type 1 (HSV-1), herpes simplex virus type 2 (HSV-2), and vesicular stomatitis virus (VSV), respectively, in a dose-dependent manner with EC₅₀ values of 1.92, 0.53, and 1.99 µg/mL.

Figure 3. Chemical structure of benzastatin C produced by *Streptomyces nitrosporeus* [35].

10. Therapeutic compounds

Diseases constitute a significant threat in the life of humans; over 30,000 diseases have been described clinically. Nevertheless, only less than one third of these can be treated [36]. There is an urgent need to obtain new therapeutic agents to cover this medical requirement. Natural products are able to fulfill medical needs through the discovery of novel therapeutic compounds [37]. In this regard, investigations on halophilic actinomycetes during recent years have led to the development of numerous isolated therapeutic substances (Table 3), including anticancer, antitumor, anti-inflammatory, antioxidant, and antimalarial substances [38]. Salinosporamide A (Fig. 4a) is a novel anticancer substance isolated from the marine actinomycete *Salinispora tropica*. Its chemical structure is bicyclic beta-lactone gamma-lactam. The mode of action of salinosporamide A is the inhibition of proteasome, which leads to induced apoptosis in multiple myeloma cells [39]. Nereus Pharmaceuticals, Inc., has developed salinosporamide A under the name NPI-0052 for the treatment of cancer in humans, which represents the first clinical anticancer agent candidate for the treatment of cancer produced by halophilic actinomycetes [36]. Lodopyridone (Fig. 4b) is an anticancer agent obtained from the marine actinomycete *Saccharomonospora* sp. [40]. Lodopyridone exhibited anticancer activity against the human colon adenocarcinoma cell line HCT-116 with an IC₅₀ of 3.6 µM. Cyclomarin A is a novel anti-inflammatory agent isolated from halophilic *Streptomyces* sp. It is a cyclic heptapeptide and possesses potent anti-inflammatory activity in both

in vivo and in vitro assays. Trioxacarcin is a new antimalarial substance isolated from marine *Streptomyces* sp. [41]. Trioxacarcin has been tested for antimalarial activity. Results showed extremely high antiplasmodial activity against the pathogen of malaria in comparison to the artemisinin drug, the most effective drug against malaria. Actinosporins C and D are novel antioxidants produced by the sponge-associated actinomycete *Actinokineospora* sp. strain EG49 isolated from the marine sponge *Spheciospongia vagabunda* [42]. Actinosporins C and D showed (at 1.25 μ M) a significant antioxidant and protective capacity from the genomic damage induced by hydrogen peroxide in the human promyelocytic (HL-60) cell line.

Compound	Source
Biological activity: anticancer	
1-Hydroxy-1-norresistomycin	<i>Streptomyces chinaensis</i>
3,6-Disubstituted indoles	<i>Streptomyces</i> sp.
Caprolactones	<i>Streptomyces</i> sp.
Chinikomycins	<i>Streptomyces</i> sp.
IB-00208	<i>Actinomadura</i> sp.
Lodopyridone	<i>Saccharomonospora</i> sp.
Marinomycins A-D	<i>Marinispora</i>
Mechercharmycins	<i>Thermoactinomyces</i> sp.
Salinosporamide A	<i>Salinispora tropica</i>
ZHD-0501	<i>Actinomadura</i> sp.
Biological activity: antitumor	
1,8-Dihydroxy-2-ethyl-3-methylanthraquinone)	<i>Streptomyces</i> sp.
Arenicolides	<i>Salinispora arenicola</i>
Aureolic acid	<i>Streptomyces</i> sp.
Aureoverticillactam	<i>Streptomyces aureoverticillatus</i>
Butenolides	<i>Streptoverticillium luteoverticillatum</i>
Chalcomycin	<i>Streptomyces</i> sp.
Daryamides	<i>Streptomyces</i> sp.
Elaiomycins B and C	<i>Streptomyces</i> sp.
Glyciapyrroles	<i>Streptomyces</i> sp.

Compound	Source
Mitomycin C	<i>Streptomyces lavendulae</i>
Piericidins	<i>Streptomyces</i> sp.
Staurosporinone	<i>Streptomyces</i> sp.
Streptokordin	<i>Streptomyces</i> sp.
Biological activity: anti-inflammatory	
Cyclomarins	<i>Streptomyces</i> sp. <i>Salinispora</i> <i>Arenicola</i>
Salinamides A and B	<i>Streptomyces</i> sp.
Biological activity: antioxidant	
Dermacozines A-G	<i>Dermacoccus</i>
Actinosporins C-D	<i>Actinokineospora</i> sp.
Biological activity: antimalarial	
Trioxacarin A, B, and C	<i>Streptomyces ochraceus</i> <i>Streptomyces bottropensis</i>
The listed compounds and the strains were obtained from the report of Subramani and Aalbersberg, (2012) [36].	

Table 3. Novel therapeutic substances isolated form halophilic actinomycetes

Figure 4. Chemical structure of some therapeutic compounds produced by halophilic actinomycetes [5].

11. Biodegradable chemicals

Saline environments, particularly waters and soils, are always exposed to contamination by heavy metals or other toxic chemical substances due to anthropogenic activities [43]. The degradation of different substances in the environment is a continuous process due to the continued activities of microorganisms. Microorganisms degrade toxic compounds to non-toxic materials such as H_2O , CO_2 , or other inorganic compounds [44]. Halophiles play a significant role in the biodegradation of the materials that contaminate marine environments. Actinomycetes have been reported to be one of those microorganisms that contribute in the degradation of organic compounds in the nature and play a role in the mineralization of organic matter [36]. Petroleum substances constitute one of the materials that contaminate marine habitats [45]. In a study carried out on oil-utilizing halophilic bacteria, results of phylogenetic studies showed that 30% of all isolates belonged to actinomycetes [46]. Extremely halophilic actinomycetes, *Streptomyces albiacialis*, were reported to be able to grow in extreme environment by using crude oil as the sole carbon source in a salinity level up to 10% NaCl. Halophilic actinomycetes isolated from the oil-polluted soil in Russia were identified as *Rhodococcus erythropolis*. It was grown in a medium containing crude oil as a unique source of carbon. Results showed degradation of n-alkanes and iso-alkanes with chain lengths of C_{11} – C_{30} and C_{14} – C_{18} [47]. A new extremely halophilic actinomycete strain that belongs to *Actinopolyspora* sp. was isolated from saline and arid surroundings of an oil field in the Sultanate of Oman. This strain exhibited the capability of degrading alkanes up to C_{15} and, at a slower rate, up to C_{25} [48].

12. Industrially important enzymes

The extracellular enzymes produced by halophilic actinomycetes have significant advantages in biotechnological applications, such as biosynthetic processes, environmental bioremediation, and food processing. Most of the enzymes are active and stable at high temperatures and pH values and have ionic strength in the presence of organic solvents [49]. They have the capability of secreting extracellular enzymes such as lipase, DNase, protease, esterase, pullulanase, galactosidase, nuclease, xylanase, inulinase, cellulose, pectinase, gelatinase, alpha-glucosidase, beta-glucosidase, alpha-mannosidase, beta-mannosidase, chitinases, xylose isomerases, fructokinases, ribokinases, etc., which exhibit higher activity in alkaline pH and stability in high concentrations of organic solvents in their environment which have been reported in the last few years [50–52]. Among the industrially important enzymes, alkaline proteases and cellulase are produced by a number of microorganisms, but limited research was done on bulk production of alkaline proteases and cellulases by alkaliphilic actinomycetes. Cellulases and proteases are the most wanted extracellular enzymes and constitute 60% of global enzymes sales [53]. Reports claimed that a certain number of alkaliphilic and halophilic actinomycetes produce extracellular protease enzymes [54]. The halophilic protease secreted by the *Nocardiopsis prasina* HA-4 withstands a wide range of pH (7–10) and temperature (20–42°C). *Nocardiopsis halotolerans* and *Saccharomonospora halophila*

recovered from Kuwait showed better keratinolytic activity under high salt concentration [55]. *Streptomyces psammoticus* that secreted lignin degrading enzymes under alkaline conditions confirmed that the enzymes have applications in delignification of pulp, textile dye decolorization, and effluent treatment [53]. Cellulases are mainly involved in the production of second-generation bioethanol and textile processing industry [56]. Halophilic cellulases obtained from actinomycetes and from metagenomic library of some marine bacteria were reported to be halostable, thermostable, and alkalostable, all favorable for textile and laundry industries [57].

13. Advantage of using halophilic actinomycetes in the industry

Halophilic actinomycetes were suitable candidates for the expression of soluble recombinant proteins [58]. A broad range of plasmid vectors obtained from halophilic actinomycetes are used for cloning native promoters and heterologous promoters for the stable expression of gene for the production of the industrially important, compatible solute ectoine [59]. Since halophilic actinomycetes have antimicrobial activity and other extracellular enzymes production capability, there is less chance of contamination during fermentation. Therefore, the major cost for the sterilization of the cultivation media is reduced.

14. Conclusions and future prospects

Several halophilic actinomycetes have special biological and chemical defense systems such as pH tolerance, and stress-tolerant metals present one of the main sources for the discovery of novel metabolites, biosurfactants, several other chemicals, and commercially important enzymes with various applications. Despite the huge demand of synthetic molecules with effective antimicrobial properties, novel methods and technologies for discovering novel natural products from microbial sources from halophilic regions should be studied. The antimicrobial metabolites producing novel actinomycetes are good bugs for unsterile cultivation and continuous bioprocessing using seawater and low-cost-contributing media components. It will be interesting to identify the mechanism of the stable properties of halophilic enzymes, which may lead to significant novel biotechnological applications. Additionally, the stability of recombinant vectors in harsh conditions such as high salt, multiple metal ions, or organic solvents should be investigated. In conclusion, halophilic actinomycetes will be a useful host for the production of commodity chemicals, antibiotics, enzymes, and biofuels in bulk with low cost.

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References

- [1] Arasu MV, Duraipandiyar V, Ignacimuthu S. Antibacterial and antifungal activities of polyketide metabolite from marine *Streptomyces* sp. AP-123 and its cytotoxic effect. *Chemosphere* 2013; 90:479–487.
- [2] Balachandran C, Duraipandiyar V, Emi N, Ignacimuthu S. Antimicrobial and cytotoxic properties of *Streptomyces* sp. (ERINLG-51) isolated from Southern Western Ghats. *South Indian Journal of Biological Sciences* 2015; 1: 7-14.
- [3] Lam KS. Discovery of novel metabolites from marine actinomycetes. *Curr Opin Microbiol* 2006; 9:245–251.
- [4] Gorajana A, Vinjamuri S, Kurada BV, Peela S, Jangam P, Poluri, E. Resistoflavine, cytotoxic compound from a marine actinomycete, *Streptomyces chibaensis* AUBN 1/7. *Microbiol Res* 2007; 162:322–327.
- [5] Tiwari K, Gupta KR. Rare actinomycetes: a potential storehouse for novel antibiotics. *Crit Rev Biotechnol* 2012; 32:108–132.
- [6] Magarvey NA, Keller JM, Bernan V, Dworkin M, Sherman DH. Isolation and characterization of novel marine-derived actinomycete taxa rich in bioactive metabolites. *Appl Environ Microbiol* 2004; 70:7520–7529.
- [7] Riedlinger J, Reicke A, Krismer B, Zahner H, Bull AT, Maldonado LA. Abyssomicins, inhibitors of para-aminobenzoic acid pathway produced by the marine *Verrucosisspora* strain AB-18-032. *J Antibiot* 2004; 57:271–279.
- [8] Maldonado LA, Fenical W, Jensen PR, Kauffman CA, Mincer TJ, Ward AC. *Salinispora arenicola* gen. nov., sp. nov. and *Salinispora tropica* sp. nov., obligate marine actinomycetes belonging to the family *Micromonosporaceae*. *Int J Syst Evol Microbiol* 2005a; 55:1759–1766.
- [9] Jensen PR, Mincer TJ, Williams PG, Fenical W. Marine actinomycete diversity and natural product discovery. *Antonie Van Leeuwenhoek* 2005; 87:43–48.

- [10] Mincer TJ, Fenical W, Jensen PR. Culture-dependent and culture-independent diversity within the obligate marine actinomycete genus *Salinispora*. *App Environ Microbiol* 2005; 71:7019–7028.
- [11] Tang YW, Persing DH. Molecular detection and identification of microorganisms. In: Murray PR (editor in chief), *Manual of Clinical Microbiology*. Washington DC, USA: ASM Press; 1999, 215–244.
- [12] Ranjard L, Poly F, Nazaret S. Monitoring complex bacterial communities using culture-independent molecular techniques: application to soil environment. *Res Microbiol* 2000; 151:167–177.
- [13] Amann RI, Ludwig W, Schleifer KH. Phylogenetic identification and in situ detection of individual microbial cells without cultivation. *Bicrobiol Rev* 1995; 59: 143–169.
- [14] Kuske CR, Barns SM, Busch JD. Diverse uncultivated bacterial groups from soils of the arid south western United States that are present in many geographic regions. *Appl Environ Microbiol* 1997; 63:3614–3621.
- [15] Toivola M, Alm S, Reponen T, Kolari S, Nevalainen A. Personal exposures and microenvironmental concentrations of particles and bio-aerosols. *J Environ Monit* 2002; 4:166–174.
- [16] Quillaguamán J, Guzmán H, Van-Thuoc D, Hatti-Kaul R. Synthesis and production of polyhydroxyalkanoates by halophiles: current potential and future prospects. *Appl Microbiol Biotechnol* 2010; 85:1687–1696.
- [17] Vargas C, Argandona M, Reina-Bueno M, Rodriguez-Moya J, Fernandez-Aunion C, Joaquin JN. Unravelling the adaptation responses to osmotic and temperature stress in *Chromohalobacter salexigens*, a bacterium with broad salinity tolerance. *Saline Biosystems* 2008; 4:14.
- [18] DasSarma P, Coker JA, Huse V, DasSarma S. Halophiles, biotechnology. In: Flickinger MC (ed.), *Encyclopedia of Industrial Biotechnology, Bioprocess, Bioseparation, and Cell Technology*. John Wiley & Sons Ltd; 2010: 2769–2777.
- [19] Oren A. Industrial and environmental applications of halophilic microorganisms. *Environ Technol* 2010; 31:825–834.
- [20] Busskamp V, Duebel J, Balya D, Fradot M, Viney TJ, Siegert S, Groner AC, Cabuy E, Forster V, Seeliger M, Biel M, Humphries P, Paques M, Mohand-Said S, Trono D, Deisseroth K, Sahel JA, Picaud S, Roska B. Genetic reactivation of cone photoreceptors restores *visual responses in retinitis pigmentosa*. *Science* 2010; 329:413–417.
- [21] Pastor J, Salvador M, Argandoña M, Bernal V, Reina-Bueno M, Csonka L. Ectoines in cell stress protection: uses and biotechnological production. *Biotechnol Adv* 2010; 28:782–801.

- [22] Gomez J, Steiner W. The biocatalytic potential of extremophiles and extremozymes. *Food Technol Biotech* 2004; 2:223–235.
- [23] Manivasagan P, Kang KH, Sivakumar K, Li-Chan ECY, Oh HM, Kim SK. Marine actinobacteria: an important source of bioactive natural products. *Environ Toxicol Pharmacol* 2014; 38:172–188.
- [24] Procópio RE, Silva IR, Martins MK, Azevedo JL, Araújo JM. Antibiotics produced by *Streptomyces*. *Braz J Infect Dis* 2012; 16:466–471.
- [25] Sujatha KVS, Raju B, Ramana T. Studies on a new marine *Streptomyces* BT-408 producing polyketide antibiotic SBR-22 effective against methicillin resistant *Staphylococcus aureus*. *Microbiol Res* 2005; 160:119–126.
- [26] Asolkar RN, Kirkland TN, Jensen PR, Fenical W. Arenimycin, an antibiotic effective against rifampin- and methicillin-resistant *Staphylococcus aureus* from the marine actinomycete *Salinispora arenicola*. *J Antibiot* 2010; 63:37–39.
- [27] Riedlinger J, Reicke A, Zähler H, Krismer B, Bull AT, Maldonado LA. Abyssomicins, inhibitors of the para-aminobenzoic acid pathway produced by the marine *Verrucospora* strain AB-18-032. *J Antibiot* 2004; 57:271–279.
- [28] Ambavane V, Tokdar P, Parab R, Sreekumar ES, Mahajan G, Mishra PD, D'Souza, L, Ranadive P. Caerulomycin A-an antifungal compound isolated from marine actinomycetes. *Advan Microb* 2014; 4: 567–578.
- [29] Maskey RP, Li F, Qin S, Fiebig HH, Laatsch H. Chandrananimycins A approximately C: production of novel anticancer antibiotics from a marine *Actinomadura* sp. isolate M048 by variation of medium composition and growth conditions. *J Antibiot* 2003b; 56:622–34.
- [30] Gao X, Lu Y, Xing Y, Ma Y, Lu J, Bao W. A novel anticancer and antifungus phenazine derivative from a marine actinomycete BM-17. *Microbiol Res* 2012; 167: 616–622.
- [31] Raveh A, Delekta PC, Dobry CJ, Peng W, Schultz PJ, Blakely PK, Tai AW, Matainaho T, Irani DN, Sherman DH, Miller DJ. Discovery of potent broad spectrum antivirals derived from marine actinobacteria. *PLoS One* 2013; 8:1–19.
- [32] Yasuhara-Bell J, Lu Y. Marine compounds and their antiviral activities. *Antivir Res* 2010; 86:231–240.
- [33] Arena A, Gugliandolo C, Stassi G, Pavone B, Iannello D, Bisignano G, Maugeri TL. An exopolysaccharide produced by *Geobacillus thermodenitrificans* strain B3–72: antiviral activity on immune competent cells. *Immunol Lett* 2009; 123:132–137.
- [34] Boyd M R, Gustafson KR, McMahon JB, Shoemaker RH, O'Keefe BR, Mori T, Gulkowski RJ., Wu L, Rivera MI, Lauricott CM, Currens MJ, Cardellina JH, Buckheit RW, P. Nara L, Pannell LK, Sowder RC, Henderson LE. Discovery of cyanovirin-N, a novel human immunodeficiency virus-inactivating protein that binds viral surface

- envelope glycoprotein gp120: potential applications to microbicide development. *Antimicrob. Agents Chemother* 1997; 41:1521–1530.
- [35] Lee JG, Yoo ID, Kim WG. Differential antiviral activity of benzastatin C and its dechlorinated derivative from *Streptomyces nitrosporeus*. *Biol Pharm Bull* 2007; 30:795–807.
- [36] Subramani R, Aalbersberg W. Marine actinomycetes: an ongoing source of novel bioactive metabolites. *Microbiol Res* 2012; 167:571–580.
- [37] Zhang L. Integrated approaches for discovering novel drugs from microbial natural products. In: Zhang L, Demain A (eds). *Natural Products: Drug Discovery and Therapeutics Medicines*. Totowa, NJ: Human Press. 2005; 33–56.
- [38] Abdel-Mageed WM, Milne BF, Wagner M, Schumacher M, Sandor P, Pathom-aree W. Dermacozines, a new phenazine family from deep-sea dermacocci isolated from a Mariana Trench sediment. *Org Biomol Chem* 2010; 8: 2352–2362.
- [39] Jensen PR, Williams PG, Oh DC, Zeigler L, Fenical W. Species-specific secondary metabolite production in marine actinomycetes of the genus *Salinispora*. *Appl Environ Microbiol* 2007; 73:1146–1152.
- [40] Maloney KN, MacMillan JB, Kauffman CA, Jensen PR, DiPasquale AG, Rheingold AL. Lodopyridone, a structurally unprecedented alkaloid from a marine actinomycete. *Org Lett* 2009; 11:5422.
- [41] Maskey RP, Helmke E, Kayser O, Fiebig HH, Maier A, Busche A. Anti-cancer and antibacterial trioxacarcins with high anti-malaria activity from a marine *Streptomyces* and their absolute stereochemistry. *J Antibiot* 2004; 57:771–779.
- [42] Grkovic T, Abdelmohsen UR, Othman EM, Stopper H, Edrada-Ebel R, Hentschel U, Quinn RJ. Two new antioxidant actinosporin analogues from the calcium alginate beads culture of sponge-associated *Actinokineospora* sp. strain EG49. *Bioorg Med Chem Lett* 2014; 24:5089–5092.
- [43] Ventosa A, Nieto JJ, Oren A. Biology of moderately halophilic aerobic bacteria. *Microbiol Mol Biol Rev* 1998; 62:504–544.
- [44] Philip JC, Bamforth SM, Singleton I, Atlas R M. Environmental pollution and restoration: a role for bioremediation. In: R. M. Atlas, J. Philip (eds.), *Bioremediation: Applied Microbial Solutions for Real World Environmental Cleanup*. Washington, DC: ASM Press; 2005: 1–48.
- [45] Fathepure BZ. Recent studies in microbial degradation of petroleum hydrocarbons in hypersaline environments. *Front Microbiol* 2014; 5:173. doi: 10.3389/fmicb.2014.00173
- [46] Hamed J, Mohammadipanah F, Ventosa A. Systematic and biotechnological aspects of halophilic and halotolerant actinomycetes. *Extremophiles* 2013; 17: 1–13. doi: 10.1007/s00792-012-0493-5.

- [47] Zvyagintseva IS, Poglasova MN, Gotoeva MT, Belyaev SS. Effect of the medium salinity on oil degradation by *Nocardioform* bacteria. *Microbiology* 2001; 70: 652–656.
- [48] Al-Mueini R, Al-Dalali M, Al-Amri IS, Patzelt H. Hydrocarbon degradation at high salinity by a novel extremely halophilic actinomycete. *Environ Chem* 2007; 4: 5–7.
- [49] Chen YG, Wang YX, Zhang YQ, Tang SK, Liu ZX, Xiao HD, Xu LH, Cui XL, W. Li J. *Nocardiopsis litoralis* sp. nov., a halophilic marine actinomycete isolated from a sea anemone. *Int J Syst Evol Microbiol* 2009; 59:2708–2713.
- [50] Essghaeir B, Rouassi M, Boudabous A, Jijkali H, Sadfi-Zouaoui N. Production and partial characterization of chitinase from a halotolerant *Planococcus rifitoensis* strain M2-26. *World J Microbiol Biotechnol* 2010; 26:977–84.
- [51] Kakhki MA, Amoozegar MA, Khaledi EM. Diversity of hydrolytic enzymes in haloarchaeal strains isolated from salt lake. *Int J Environ Sci Technol* 2011; 8:705–14.
- [52] Debananda SN, Kshetri P, Sanasam S, Nimaichand S. Screening, identification of best producers and optimization of extracellular proteases from moderately halophilic alkali thermo tolerant indigenous actinomycetes. *World Appl Sci J* 2009; 7: 907–916.
- [53] Kizhekkedathu NN, Prema P. Mangrove actinomycetes as the source of ligninolytic enzymes. *Actinomycetologica* 2005; 19:40–47.
- [54] Mital SD, Joshi RH, Patel RK, Singh SP. Characterization and stability of extracellular alkaline proteases from halophilic and alkaliphilic bacteria isolated from saline habitat of coastal Gujarat, India. *Braz J Microbiol* 2006; 37:276–282.
- [55] Zarban S, Al-Musallam AA, Abbas I, Stackebrandt E, Kroppenstedt RM. *Saccharomonospora halophila* sp. nov., a novel halophilic actinomycete isolated from marsh soil in Kuwait. *Int J Syst Evol Microbiol* 2002; 52:555–558.
- [56] Wang CY, Hsieh YR, Ng CC, Chan H, Lin HT, Tzeng WS, et al. Purification and characterization of a novel halostable cellulase from *Salinivibrio* sp. strain NTU-05. *Enzyme Microb Technol* 2009; 44:373–379.
- [57] Shivanand P, Mugeraya G, Kumar A. Utilization of renewable agricultural residues for the production of extracellular halostable cellulase from newly isolated *Halomonas* sp strain PS47. *Ann Microbiol* 2013; 63:1257–63.
- [58] Tokunaga H, Arakawa T, Tokunaga M. Novel soluble expression technologies derived from unique properties of halophilic proteins. *Appl Microbiol Biotechnol* 2010a; 88:1223–31.
- [59] Bursy J, Kuhlmann AU, Pittelkow M, Hartmann H, Jebbar M, Pierik AJ, et al. Synthesis and uptake of the compatible solutes ectoine and 5-hydroxyectoine by *Streptomyces coelicolor* A3(2) in response to salt and heat stresses. *Appl Environ Microbiol* 2008; 74:7286–96.