

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



---

# Health Effects of Metals in Particulate Matter

---

T.I. Fortoul, V. Rodriguez-Lara, A. Gonzalez-Villalva,  
M. Rojas-Lemus, L. Colin-Barenque,  
P. Bizarro-Nevarés, I. García-Peláez,  
M. Ustarroz-Cano, S. López-Zepeda,  
S. Cervantes-Yépez, N. López-Valdez,  
N. Meléndez-García, M. Espinosa-Zurutuza,  
G. Cano-Gutierrez and M.C. Cano-Rodríguez

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/59749>

---

## 1. Introduction

There is increased evidence of the association of air pollution and deleterious health effects. Particulate matter (PM) has obtained more attention, especially the small size components (PM<sub>10</sub>, PM<sub>2.5</sub>, UFP -ultrafine particles-) that carry on their surface different organic and inorganic elements whose composition differ with local and regional variations [1].

PMs might be identified by source, e.g., primary or secondary; combustion products, traffic, or by particle's size (aerodynamic diameter) (PM<sub>10</sub>, PM<sub>2.5</sub>, UFP -ultrafine particles-). This former parameter is important because the larger the particle the shorter the time it remains suspended in the air and the lower the risk of it being inhaled. Also the smaller the particles the higher the chances of deleterious health effects [1].

### 1.1. Metals in particulate matter, air pollution and sources

Particles toxic components are complex mixtures of solids or liquids with different characteristics (e.g., mass, number, size, shape, surface area, chemical composition, acidity, solubility). Chemical components may be located on the surface or inside the particle. Considering the source there are natural and anthropogenic emissions. Natural sources include sea salt, volcanic ash, pollens, fungal spores, soil particles, forest fires and wind-blow dust [2]. An-

thropogenic sources consist of fossil fuel combustion products, industrial process, mining activities, wood stove burning and cigarette smoking. In urban areas the main source of PM are motor vehicles especially those derived from diesel fuel combustion.

One of the components adhered to the small particles are metals that come from impurities derived from fuel additives and brakes and tires attrition. Transition metals are generated by non-exhaust emissions, impurities in fuel additives, or metallurgic process. Iron (Fe), nickel (Ni), vanadium (V), chromium (Cr) copper (Cu) are considered because its potential to produce Reactive Oxygen Species (ROS) in biological systems [1]. Others such as zinc (Zn) may also exert toxic effects by other mechanisms besides ROS production.

Heavy metals such as cadmium, lead and mercury are some of more common air pollutants emitted by industrial activities, combustion, extraction and processing activities. The diagnosis of health effects for heavy metals might be difficult if there is no previous evidence of the exposure source, while acute exposures usually occur in the workplace and are more easily identified; signs and symptoms differ within metals because each one interacts with different targets such as: specific metal-binding proteins (metallothioneins, transferrin, ferritin, ceruloplasmin) in cases such as Cd, Cu, Hg, Ag, Mn Zn, Al, and Be; membrane carrier-proteins (phosphate and sulphate-transporters), divalent cation-transporters, some examples are V, Cr, Mo, Se. Heavy metals share with transition metals the possibility to exert its toxic effects by the production of ROS [2-4].

## 2. Metals and its effects on health

A variety of health effects associated with PM exposure began with the increased mortality risk in those cities with high particulate concentrations (PM), later myocardial infarction incidence and high particulate matter emitted by internal combustion engines were reported. Also ultrafine particles emitted from the vehicles, especially diesel, induced oxidative stress in the endothelium, and through the nose these particles penetrate the olfactory bulb and reached other structures from the nervous system, inducing increased inflammatory responses in the brain. Reduced lung function and respiratory diseases are frequently reported. Thrombosis, heart rate changes, blood pressure modifications have been associated with exposure to particulate matter also [5] and more recently metabolic abnormalities such as altered glucose metabolism have been reported as well. Low birth-weight, infertility, genotoxicity and cancer [6] are also part of the spectrum of alterations associated with PM. Metals are some of the components adhered to particles surface associated with its toxicity [7, 8]. Here we included the health effects of some metals carried by inhaled particles, and its possible mechanisms of damage.

### 2.1. Respiratory system and metals

Human beings can survive three weeks without food, three days without water but no more than three minutes without air [9]. As Aaron Cohen mentioned, "You can't avoid breathing the air no matter what you are" [10] and this comment is because the recent publications about

the air pollution problems reported in China [11] and the case of a Chinese eight-year-old girl diagnosed with lung cancer, until now the youngest victim of lung cancer [10]. Particulate Matter (PM) is the carrier of metals into the lung structure and its content determines its potential health hazard [7].

The exact mechanisms involved in PM exposures and lung damage have been discussed and mentioned previously, but it seems that oxidative stress, inflammation, and modulation of the immune response are some of the mechanisms proposed. Differences according to the age of the population exposed also have some differences because in children's asthma, reduction in lung growth, allergic rhinitis and respiratory infections are more frequently reported compared with adults. Metals such as Cd, Hg and Au have been associated with autoimmune reactions. Other metals reported on the particles are Fe, Zn, and Ni, and recently with the use of the catalytic converters an increase in the presence of Pt, Pd and Rh in the particles inhaled has been observed. These three elements have been associated with asthma, rhinoconjunctivitis, and dermatitis among occupational exposure workers [7].

Lead has been present in the atmosphere in different concentrations and since tetraethyl lead was reduced as anti-knocking agent in gasolines Pb-concentrations in the air and in children's blood has decreased [12]. Changes in the ultrastructure of the non-ciliated bronchiolar cell (NCBC) after the inhalation of Pb has been reported along with cell hypertrophy, whorl-like structures and mitochondria cristae disarray [13]. The NCBC was also the target of the combined inhalation of Cd and Pb. A decrease in the cell volume with surface irregularities and the presence of sloughed cells on the epithelial surface were noticed in the exposed group; however at 4-week exposure time, clusters of dividing cells were observed [14]. Differences in the lung concentration of each element, compared with the mixture were reported and could explain the morphological differences reported [15].

Cigarette smoke is a source of direct deposit of metals in lung tissue; because metals are not biodegradable remain in the tissue for long periods of time. In a review from Stavrides [16] chromium, cadmium and nickel are mentioned as carcinogenic and genotoxic metals, indicating that the main effect of these metals as carcinogenic agents is because of altered DNA's repair capacity as a consequence of the oxidative stress. In addition the destruction of the cilia by the gaseous phase of the cigarette smoke facilitates the stagnation of the mucus, whose production increases as a consequence of the irritating effect of the smoke components, resulting in longer periods of cells-metals contact with the increasing opportunity to interact with the genetic material and also to interact with DNA, producing cumulative genetic alterations that could result in lung cancer [16]. Cerium and lanthanum are also reported in the environmental tobacco smoke and may produce inflammation and granulomatosis in lung tissue [17]. Cakmak [18] found association with respiratory effects and the content of Cd, Zn and V in particulate matter (PM<sub>2.5</sub>) and the suggested mechanism was oxidative stress resulting in inflammation and tissue damage.

## 2.2. Cardiovascular toxic effects of metals

Particulate Suspended matter (PM) exposure raises the risk of developing cardiovascular diseases, both in the short and long term [19, 20]. The proposed mechanisms include inflam-

mation and the induction of hypercoagulability, oxidative stress and endothelial dysfunction [20, 21]. The best evidence that PM have an important role in cardiovascular morbidity and mortality are the interventional studies, in which measures to reduce the levels of PM are taken and a significant decrease in cardiovascular risk is observed [22, 23].

Fine and ultrafine particles are considered the more toxic compared with bigger particles; particularly Diesel exhaust ultrafine particles are dangerous because of its high metals content, causing oxidative stress that leads to endothelial dysfunction and the development of atherosclerosis increasing the risk of heart infarction or stroke [24]. There are associations of elevated concentrations of some metals such as nickel, copper, arsenic, and selenium in fine or ultrafine particles and markers of cardiovascular disease: higher levels of markers of inflammation (C-reactive protein, interleukin-6, and vascular endothelial growth factor) and reduced levels of circulating endothelial progenitor cells (CEPC) suggesting reduced capacity of endothelial repair [25]. Iron overload is directly associated with hypertension, atherosclerosis progression and increased cardiovascular risk [26] Reports that iron chelation with deferoxamine decreases endothelial dysfunction and has been successfully used to reduce cardiovascular risk in diabetic and non-diabetic patients [27]. Lead causes cardiovascular effects even at low doses and it has been associated with hypertension in animals and humans [28]. Peripheral arterial, coronary heart and cerebral vascular diseases had been also associated with Pb exposure, but further studies are needed to establish its causality [29]. Cadmium affects the cardiovascular system too, demonstrated *in vitro* and in animal studies, in addition of epidemiological evidence that Cd is associated with hypertension, promoting atherosclerosis and myocardial infarction. Mercury promotes atherosclerosis and cardiovascular disease in much lower concentrations than those reported for its neurotoxic effects [30].

### 2.3. Metals and its relationship with cancer

Many epidemiological studies have demonstrated an association between long-term exposure to ambient air pollution and cancer mortality mainly from lung cancer. The risk of cancer is associated with exposure to different pollutants such as nitrogen dioxide (NO<sub>2</sub>), Sulphur dioxide (SO<sub>2</sub>), particulate matter (PM) and several metals attached to its surface. Metals can promote carcinogenesis, through several pathways such as: producing DNA damage, activating different signaling pathways that lead to tumor progression and promoting inflammation. In the following paragraphs the mechanisms by which some metals promote carcinogenesis are detailed.

Experimental animal studies have demonstrated clearly that cadmium and cadmium compounds by multiple routes of exposure generate cancer at various sites in many animal species and in humans. Cadmium exposures of laboratory animals causes leukemia and lymphoma, local sarcoma and cancer of the adrenal gland, liver, lung, kidney, pancreas, pituitary, prostate and testis. Moreover, cadmium and their compounds are classified by the International Agency for Research on Cancer (IARC) as carcinogens in humans. Cadmium exposure is associated with lung adenocarcinoma, also with prostate, kidney, urinary bladder, pancreas and breast cancer in humans. Several studies have shown that Cd carcinogenicity seems to be crucially mediated by the production of ROS such as hydroxyl radicals, superoxide anions, nitric oxide

and hydrogen peroxide and are due to the inactivation of detoxifying enzymes (e.g., catalase, glutathione peroxidase, glutathione reductase and superoxide dismutase) as a consequence of the interaction with thiol groups. Cadmium is also capable of replacing copper and iron in various cytoplasmic and membrane proteins (e.g., ferritin, apoferritin), leading to an increase in the amount of unbound or poorly chelated copper and iron ions inducing oxidative stress via Fenton reactions. Cadmium also produces genotoxicity by the production of DNA single-strand breaks and damage and competes for binding at sites (specifically with a zinc finger motif) that are important in gene regulation, enzyme activity, or maintenance of genomic stability. In addition, this metal modifies the expression of several genes related to carcinogenesis, including intermediate early-response genes such as c-fos, c-jun, and c-myc; stress-response genes such as metallothionein, and heat-shock genes; genes controlling glutathione and related proteins, as well as transcription and translation factors. Also disrupts cell adhesion mediated by E-cadherin and affects the regulation of cell growth and apoptosis causing tumor progression [31-33]. In addition, cadmium has estrogenic effect and may bind to and activate mammary cell estrogen receptors; it also interacts and regulates the transcription of estrogen-dependent genes affecting the synthesis of proteins and/or the activity of cell-signaling pathways in a manner similar to estradiol [34]. In addition to its endocrine effects on mammary tumor cells, cadmium transforms healthy breast epithelial cells into cells with a cancer-like profile through non-hormone-related pathways. Thus, in the presence of cadmium, the cells alter gene expression and DNA changes in DNA methylation (an epigenetic change) that are typical of cells undergoing transformation from healthy to cancerous type [35].

Furthermore, epidemiological data provide increasing evidence that environmental as well as occupational lead exposures may be associated with increased cancer risk. The IARC has classified lead as possible human carcinogen (group 2B) and its inorganic compounds as probable human carcinogens (group 2A). Lead exposure has been associated to increased lung cancer risk. Some studies looking at blood lead levels in the general population have also found a small increased risk of lung cancer in people with higher lead levels. In addition, the majority of the studies found an increased risk of stomach cancer with higher lead exposure, even though the studies did not take into account other factors that could also have been affected stomach cancer risk. There is a stronger association of kidney tumors with lead exposure; brain, lung and bladder cancer have also been linked to lead in different studies, however results are controversial [36]. Several studies have identified the carcinogenic potential of lead, because the genotoxicity of the metal that induces alterations in DNA synthesis, mutations, chromosome aberrations, as well as inhibiting DNA repair or displacing zinc in DNA binding proteins [37]. Lead, also stimulates cell proliferation, induces alterations in gene transcription and causes oxidative damage that promote carcinogenesis [36].

The International Agency for Research on Cancer (IARC) has determined that some nickel compounds are carcinogenic to humans and that metallic nickel may be carcinogenic to humans. The EPA has determined that nickel refinery dust and nickel subsulfide, are human carcinogens. Occupationally exposed people have a higher risk of respiratory tract cancer (nasal sinus and lung cancer mainly) due to inhalation of nickel at their workplace in nickel refineries, nickel-producing or processing plants or using industries. High cancer risk is related

to less soluble oxidic and especially sulfidic nickel species in refinery dust. Earlier studies gave already indications that rats during two years to inhalation exposure of nickel subsulfide developed a significant higher number of lung tumors [38]. Mechanisms of nickel carcinogenicity have not been fully elucidated yet. Ionic nickel ( $\text{Ni}^{2+}$ ) is supposed to be the carcinogenic species because it can bind to cellular components such as nuclear proteins (histones and protamines) and DNA. Nickel also induces chromatin condensation modifications, DNA hypermethylation, histone acetylation and gene silencing, which disturb gene expression. Moreover, there is evidence that nickel ions inhibit enzymes required for DNA repair. Furthermore, nickel modulates gene expression by the induction of DNA methylation and/or suppression of histone acetylation [38-41].

Some studies have shown that serum copper levels are elevated in cancer patients and correlate with the severity of the disease and the response to therapies [42, 43]. Copper-chelating drugs have been reported to have antiangiogenic activity in animal models. Other study has shown that cancer cells express higher levels of the copper transporter Ctr1 and that the tumors were sensitive to the reduction in systemic copper levels compared with normal tissues [44]. Pharmacological suppression of systemic copper impairs oxidative phosphorylation and tumor growth, since copper can modulate the proliferation of cancer cells and associated tumor growth. It has been proposed that copper can be a rate-limiting nutrient for tumors, similar to oxygen and glucose. Copper levels in tumors affect cytochrome c oxidase activity, additional bioavailable copper facilitates increased production of ATP, which is consumed to fuel rapid proliferation of cancer cells. Thus, copper may not initiate transformation, but may stimulate proliferation of transformed cells by providing energy needed for cell-cycle progression as proposed Ishida and coworkers [42]. Additionally, copper ions are well suited to facilitate formation of ROS that can damage biomolecules, including DNA and chromatin. This event occur *in vitro* with isolated DNA or chromatin, or by exposure of cultured mammalian cells to copper complexed with various agents. Whether if this is likely to occur *in vivo* is not well defined. However, copper, can directly bind with high affinity to DNA molecule; this binding can modify the conformational structure of DNA promoting carcinogenesis[45].

Mercury and its compounds mainly methylmercury have been classified as “possibly carcinogenic compounds to humans”. Mercury has been associated with lung cancer, genitourinary tract cancer and probably brain cancer risk in occupational exposed personnel, however these results are in controversy because workers might be also exposed to other metals [46]. Mercury promotes carcinogenesis inducing oxidative stress. In addition, mercury compounds are genotoxic, mainly by inhibiting the mitotic spindle and altering DNA repair processes decreasing the incision step. Lead and aluminum can increase the toxicity of mercury. It has been shown that mercury rapidly depletes the immune system and could decrease immune tumor response. Chronic exposure to relatively low levels of mercury may inhibit antioxidant enzymatic activity due to persistent oxidative stress. This phenomenon might represent an important peripheral target for mercury toxicity in exposed populations [33].

Inhalation exposures to aluminum in several cohort studies reveal a relationship with increased cancer incidence and mortality in the aluminum smelting industry [47]. The IARC has classified occupational exposures during aluminum production as a causal factor, with

sufficient evidence in humans, for cancers of the lung and the bladder. Nowadays, authors conclude that exposure levels to know health hazards associated with the emissions from primary aluminum production should be studied to establish a clear relationship between inhaled aluminum exposure and cancer [48]. The carcinogenic mechanisms of inhaled aluminum exposure are associated to different compounds and not with the metal *per se*, as it is described in different studies [49].

Chromium is widely used in the industry for the production of stainless steel, chromium plating, and spray-painting. The health effects and toxicity/carcinogenicity of Cr inhalation are primarily related to its oxidation state at the time of exposure [50]. According to epidemiological studies, the hexavalent [Cr(VI)] form of this metal, appears to be drastically toxic and carcinogenic, thus it has been classified as carcinogenic to humans by the IARC [51]. The carcinogenicity of the metal is site specific, mainly to the lungs and nasal cavity [50]. The molecular mechanisms of [Cr(VI)]-induced carcinogenesis are well studied and characterized, and the main mechanism of chromium carcinogenesis, is the production of free radicals, resulting in the generation of oxidative stress, This stress causes a series of modifications that are directly linked to the establishment of the cancer phenotype. The genetic changes involve Cr-DNA adducts, formation of DNA-protein cross-links, single and double strand DNA breaks [52, 53].

Studies exploring excessive environmental exposure to iron are often limited by poor characterization of the environmental factors and causal relations to effects other than the chemical properties of the iron [54]. Iron is present primarily in two oxidation states, ferrous ions [Fe(II)] and ferric ions [Fe(III)]. Mechanisms by which iron may contribute to tumor induction or progression, includes oxidative damage-induced changes in genetic material as the initial step involved in Fe-induced mutagenesis and carcinogenesis. Other mechanisms are alterations in gene expression consistent with increased iron requirements in proliferating cells and decreased immune surveillance against cancer [55].

In occupationally exposed individuals, inhalation of Mn is a potential important route of exposure [56]. In general, Mn and its inorganic compounds are considered to possess low mutagenic or carcinogenic potential compared with other heavy metals. The experimental evidence on its carcinogenicity does not provide any clear evidence, while the available occupational and environmental epidemiological evidence is equivocal as to whether exposure to inorganic Mn is associated with a significant cancer risk or not [50]. Apparently from *in vitro* data, Mn and some inorganic Mn compounds are cytotoxic at differing concentrations depending on the test system, generating ROS *in vitro* and *in vivo*, interfering with DNA polymerases, mitochondrial function and activating some cytokines and MAPK cell signaling cascades. These mechanisms should probably contribute to Mn carcinogenesis, but further studies must be made [57].

Finally, V is a major transition element that is released primarily by the burning of fossil fuels, including petroleum, oil, coal, tar, bitumen, and asphaltite. Among V compounds, V pentoxide is highly toxic [58]. The IARC classified vanadium pentoxide as a possible carcinogen to humans (Group 2B) in 2003 [59]. The pentavalent forms, such as V and vanadate have carcinogenic potential via ROS generation, DNA damage, and activation of hypoxia signaling

[51, 60]. There is *in vivo* preclinical data on cancer chemoprevention and therapy, which provide a rationale for its use in human populations [61].

#### 2.4. Genotoxicity and metals

The term genotoxicity refers to any detrimental change in genetic material, regardless of the mechanism by which the change was caused [62]. The DNA lesions can be classified according to the extent and severity of damage. Repairable damage includes single-stranded breaks, oxidized bases, AP-sites and alkali-labile sites. The damages that result from the incorrect repair are chromosomal rearrangements and sister chromatid exchange. Finally, there are irreparable injuries including gains or losses of whole chromosomes or chromosome fragments (chromosomal aberrations) that are the product of clastogenic or aneugenic events. The consequences of DNA damage include alterations in the three-dimensional conformation, blocking the processes of replication and transcription, deletions, chromosomal instability, cell death and mutagenic events. Thus, genotoxic mutagenic events precede and therefore are the source of cellular malignancy [63]. Studies indicate that nickel induces chromosome aberrations in rats [64]; cadmium [65] and lead [66] causes single strand breaks and alkali-labile sites in mouse cells; in rats uranium causes double strand breaks [67]; iron induces in mice chromosome aberrations and micronuclei; in human beings vanadium produces bases oxidation and micronucleus [68], but in mice produces single strand breaks and micronucleus too [69, 70]. Conversely to others metals, titanium showed no adverse effects on DNA in mice cells [71].

#### 2.5. Neurotoxicity of metals

The brain is vulnerable to oxidative stress damage produced by metals, due to its great metabolic activity, high cellular content of lipids, and low levels antioxidants such as catalase and superoxide dismutase. Air pollution is a mixture of gases and metals associated with particulate matter (PM) [72] that induced olfactory dysfunction, neuroinflammation and elevated markers of neurodegeneration [73] and heavy metals in PM are accumulated in brain tissues [74].

Inhaled nanosized particle (NSP) includes ambient spherical particles < 100 nm deposited directly on olfactory dendritic cilia in the olfactory mucosa. Subsequently uptake and translocated along axons of the olfactory nerve by nasal route via neuronal transsynaptic transport and uptake through the blood brain barrier from systemic circulation, and induced oxidative stress and gene expression in central nervous system in human and experimental animals [75, 76].

Occupational and environmental exposure to neurotoxicants such as iron (Fe), copper (Cu), manganese (Mn), aluminum (Al), zinc (Zn) mercury (Hg), lead (Pb) and vanadium induced oxidative stress [77, 78] that generated accumulation of ROS inducing protein, lipid and deoxyribonucleic acid (DNA) oxidation, and produced neurotoxic changes that are risk factors for development neurodegenerative diseases such a Parkinson (PD), Alzheimer (AD), Huntington (HD), amyotrophic lateral sclerosis (ALS) and transmissible spongiform encephalopathy (TSE), [79, 80] [81, 82].

Protein misfolding is implicated in neurodegenerative diseases, such as amyloid- $\beta$  precursor protein, in senile plaques and tau in neurofibrillary tangles of AD,  $\alpha$ -synuclein in Lewy bodies in substantia nigra in PK, prion protein in TSE and huntingtin in HD striatum have been connected to neuronal iron homeostatic control [83].

At a molecular level metal dishomeostasis and mitochondria dysfunction in AD and PD and the cytoplasmic predominance of neuronal 8-hydroxy-Guanine supports mitochondria as the major source of ROS responsible for RNA oxidation and might induce DNA oxidation neuronal damage [84, 85].

Atmospheric Mn is present in gasoline additive methylcyclopentadienyl manganese tricarbonyl (MMT) is a putative modulator of dopamine biology (the primary target of Mn neurotoxicity) and workers exposed to airborne Mn are in risk of developing PD known as manganism, an extrapyramidal neurological disease characterized by rigidity action tremor, bradykinesia, memory and cognitive dysfunction. Mn in blood crosses the blood brain barrier and accumulates inside the neuron disrupting the synaptic transmission and inducing glial activation[86].

In previous studies it has been reported that aluminum inhalation induced altered of expressions of glycogen synthase kinase-3 (GSK3) and protein phosphatase (PP1), which help in the regulation of carbohydrate metabolism in the rat's brain [87]. Mercury exposures in male marmoset monkey showed deposits of Hg in ventral horn motor neuron and atrophy of large myelinated motor axon [88]. In our mice model the inhalation of vanadium pentoxide caused morphological and functional alterations in the central nervous system. In the olfactory bulb we showed dendritic spiny loss of granule cells and ultrastructural changes characterized by swelled organelles, disrupted mitochondria and necrotic and apoptotic neuronal death that might correlate with the olfactory dysfunction. Also, the hippocampal formation showed a decrease in dendritic spines and necrosis of the pyramidal layer of CA1 and granule cell of dentate gyrus that could be related with spatial memory impairment at a 4-week exposure. We also found the loss of immunoreactive- tyrosine hydroxylase + in substantia nigra and a decrease in dendritic spine density in the medium striatal spiny neurons at 8-week exposure time. The blood brain barrier (BBB), after the inhalation of vanadium showed cilia loss, cell sloughing and ependymal epithelium detachment from the basal membrane, and the presence of oxidative damage in the choroid plexus, which was confirmed by the presence of 4-hydroxynonenal. The reported alterations were associated with an increase in MMP-9 and MMP-2 activity in the cortex, the olfactory bulb, hippocampus and striatum [89, 90].

## 2.6. Metals and mental health

Environmental pollution by heavy metals that are produced by the combustion of hydrocarbons is a public health problem, which affects different organs such as central nervous system [91] have resulted in behavior, learning, mental disorders, attention deficit and low mental performance [92-94].

Inhaled metals involved in central nervous damage, mental and behavioral disorders are arsenic, lead, cadmium, mercury, manganese and vanadium [95] [96]. The inhalation of these

metals induces brain damage and resulted in behavioral disorders associated with the severity of the poisoning, the metal involved, the chemical state of the element as well as the exposure route and the age of the exposure [97]. Inhaled arsenic might cause Guillain-Barre similar syndrome with confusion, irritability, cognitive loss, decrease in verbal responses and paralysis [98].

Lead causes an irreversible reduction in cognitive ability in children resulting in an IQ decrease [99]. Oxidative stress is also the mechanism proposed to induce the damage [100]. Other behavioral effects induced by inhaled-Pb are depression, irritability, bipolar states, mental retardation and cognitive deficit [101].

Cadmium has the ability to replace iron and copper, which induces an increase in the production of ROS via the Fenton reaction, which translates into GABA and serotonin systems alteration, causing irritability, depression, amnesia and cognitive disorders [102].

Mercury mechanism of action are not fully known but it seems that interact molecularly with the antioxidant systems such as GSH, cysteine and melatonin [103]. Apoptosis, necrosis, lysis and phagocytosis have been reported in exposed humans [104]. Other neurotoxic effects reported are weakness, inability to concentrate, lethargy, depression, irritability, blindness, coma and death [105].

Excessive manganese exposure during childhood causes hyperactivity and learning disorders [106]. Studies in animals show that inhaled manganese reaches the central nervous system through the olfactory nerve and by the blood, crossing the blood-brain barrier inducing [107] "manganism" that includes tremor, "manganic madness", schizophrenic symptoms, violent behavior, compulsions, emotional instability, hallucinations, and other psychiatric disorders [108].

Vanadium brain accumulation has been related with behavioral and cognitive disorders, [109] [110]. Memory loss, a decrease in the sense of taste and Parkinson's disease has been reported in vanadium exposure [111, 112].

## **2.7. Metals and its toxic effects in the eye**

Despite the fact that the eye is an important air pollutant target because it is directly exposed to the atmosphere, as well as to the elements that enter into the lungs and further are distributed through the systemic circulation [113], until now, only two studies have been found to approach the effect of atmospheric pollutants inhalation on the retina, where phototransduction process takes place. Such studies were performed in a mice model in which the animals were exposed to vanadium (V) [0.02M] inhalation 1 h twice a week, for 4, 8 and 12-week time periods.

In all exposure times, morphological alterations in the photoreceptor layer (PL) and in the inner and outer nuclear layers were observed, as well as a gradual rhodopsin pigment reduction in the PL and an increase of the oxidative stress biomarker -4-hydroxynonenal- in the PL and in the inner and outer plexiform layers [114].

Additionally, the effect of V exposure was evaluated on the Müller glial cell (MGC), which is the predominant radial glia in the retina, for 4 and 8-week time period. Glial fibrillary acidic protein (GFAP) expression increased at four weeks after the exposure, probably as evidence of reactive gliosis, whereas it was observed a gradual reduction in Glutamine synthetase (GS) expression as exposure time passed. Given that GS is an enzyme whose levels are regulated by its substrates -glutamate (Glu) and ammonium- its reduction might suggest that photoreceptors, that produce most of the Glu in the retina, are degenerating in response to the V toxic insult [113]. This is consistent with the rhodopsin pigment reduction that previously was mentioned, because it evidenced damage to the photoreceptors as a consequence of the increase in the oxidative stress induced by the exposure [114].

## 2.8. Metals and glucose metabolism

There is limited information about the role of metals in carbohydrate metabolism and glycemic regulation; however there are some studies showing that metals have hypoglycemic or hyperglycemic effects. The relevance of studying these effects is that the exposure to some metals has been related to increased risk of diabetes, but some other metals have shown a hypoglycemic effect and have been studied as a potential treatment of diabetes (as vanadium).

Iron, mercury, nickel and lead are hyperglycemic metals that are also pollutants and will be discussed in this chapter. Iron is an atmospheric pollutant in both urban and industrial sites [115], near iron or steelmaking industries, near petrochemical areas [116], cement mills and in metro systems of many cities [117]. Mercury is a toxic heavy metal widespread and persistent in the environment and it is considered one of the most relevant atmospheric pollutants (Wang et al, 2006). Nickel is a metal released from many industries and it has proved to be toxic at high concentrations and lead is a heavy metal pollutant because it is released to the atmosphere by the burning fossil fuels, industries and mining activities.

Iron is an essential metal for life, but iron overload is a health risk because it is associated to insulin resistance, hyperglycemia and an elevated risk of type 2 Diabetes Mellitus [26, 118, 119]. Several authors have explored the mechanism of this risk; oxidative stress has been implicated because it is associated with insulin resistance and with direct damage on beta pancreatic cells [120].

There is evidence that mercury can cause hyperglycemia because it can directly damage pancreatic beta cells inducing necrosis or apoptosis [121]. In a follow-up study of young people who were exposed to high levels of mercury a higher risk of developing diabetes after 18 years was found [122]. The majority of the associations are related to water pollution, but it is important to evaluate people who have been exposed by atmospheric pollution. On the other hand, there is no evidence of higher levels of mercury in blood of diabetic type 1 or 2. A causal relationship between mercury and pancreatic dysfunction that leads to hyperglycemia and diabetes has been reported, but not all the diabetics have higher levels of this metal [123, 124].

There are reports about the effect of lead on glycaemia regulation but the effect seems to be related to the dose and the compound. Ibrahim and cols in 2012 [125] had reported hyperglycemic effect on rats exposed to different doses of lead acetate, just as Adham et al. did in birds

in 2011 [126], however other authors have found in rats hypoglycemia, after low doses of lead dissolved in water [127]. It is necessary to understand the mechanism and the reason for this dual effect.

There are multiple reports of hyperglycemia and insulin resistance after exposure to nickel in different animal models [128, 129], so it is important to consider this element as a possible risk factor to glycemic deregulation.

Some metals decrease the levels of blood glucose such as vanadium, chromium, magnesium and zinc. Proposed mechanisms for this effect include: activation of insulin receptors, increasing insulin sensitivity, and function as cofactors or components of the enzymatic systems involved in glucose metabolism or acting as antioxidants to prevent tissue peroxidation. [130].

Vanadium potentiates the action of insulin and lowers blood glucose levels. Some vanadium compounds have been studied as antidiabetic agents [131]. At first it was thought that vanadium exerted an effect on the glycemia because it inhibits appetite at certain concentrations, but hypoglycemia was observed only minutes after its administration, which is not a period of time sufficient to exert its anorectic effects [132]. The hypoglycemic effect is explained because vanadium inhibits some tyrosine protein phosphatases increasing the phosphorylation levels of various insulin pathway intermediaries. Activation of these signaling pathways results in GLUT transporter translocation to the plasma membrane [133]. Another factor that contributes to the hypoglycemic effect of vanadium is its inhibitory effect on gluconeogenesis because it inhibits the expression of the gluconeogenic enzymes PEPCK and GTPase [134]. There are reports of severe hypoglycemia that may threaten life in vanadium acute intoxication [135]. Further studies are needed to evaluate the hypoglycemic effect of vanadium as an air pollutant or in workers occupationally exposed.

Chromium induces hypoglycemia because it is a promoter of glucose catabolism in muscle cells and adipocytes. Also, it functions as a regulator of glycaemia in different animal and human models, and as an inhibitor of glucogenolysis in muscle cells. The trivalent compounds as chromium picolinate increase insulin activity [136]. There are reports that after the consumption of Chromium based compounds; patients suffering from diabetes improved their sensibility to insulin [137]. The mechanisms studied are: 1) increase the concentrations of the messenger RNA for insulin receptor; 2) making a complex with insulin that has a greater activity in the metabolism of glucose than insulin alone; 3) through the decrease in TNF $\alpha$ , resistin and interleukin 6 concentrations; and 4) increasing the sensitivity of pancreatic  $\beta$ -cells [138, 139]. There are no reports of severe hypoglycemia, but further studies are needed to elucidate the effects of chromium in concentrations inhaled as atmospheric pollutants.

Depletion of magnesium is associated to insulin resistance, hyperglycemia and type 2 Diabetes [140]. Low levels of serum magnesium in diabetics is associated with poor glycemic control [141] and foot ulcers [142]. Zinc has insulinomimetic activity *in vitro* and blood glucose lowering effect *in vivo* [143]. In some studies zinc deficiency has been associated with hyperglycemia and diabetes [141]. Also, zinc supplementation in diabetic patients improves glycemic control [144]. However, a higher zinc concentration has promoted metabolic syndrome (overweight, hypertension and dyslipidemia) in Wistar rats [145] Ugwuja et al, in

2014 [141] reported higher zinc levels in complicated diabetic patients compared with diabetic uncomplicated cases. The meaning of these associations is unclear and needs further studies.

## 2.9. Metals and its toxic effects on liver

Inhaled air pollutants that travel through the blood, also produce changes in the integrity of liver parenchyma, which leads to a slowly and irreversible liver damage [146] [147]. Acute or chronic liver damage is the usual consequence in the majority of the toxic agents that enter into the organism, because the liver is the main organ that metabolizes xenobiotic agents such as metals. The progressive deleterious events in the liver starts with steatosis ending in hepatocellular carcinoma, passing through chronic hepatitis, fibrosis and cirrhosis, finalizing in liver failure and death [148].

It is important to emphasize that regardless the etiological agent in all types of liver damage there is overwhelming evidence of an increase in free radicals or a decrease in antioxidant defenses [149]. As well, the reactive oxygen and nitrogen species play a crucial role in the induction and progression of the liver diseases.

Arsenic is a metalloid that has been characterized by causing a variety of alterations in the organism [150]. Arsenic crosses lung alveolar membrane and reaches the blood stream; hence it is transported to all the organs, mainly to the liver, in which is metabolized. It has been reported that arsenic induces liver cancer [151]. This is done through the modulation of transcription factors like NF- $\kappa$ B, AP-1 and p53 that promotes liver tumors. Likewise, Arsenic causes liver lipoperoxidation producing large amounts of ROS [152]. Also a decrease in the levels of the superoxide dismutase enzymes (SOD), catalase (CAT) and glutathione peroxidase has been reported [153].

Lead has been characterized by inducing damage by the production of ROS, that increases lipid peroxidation and decreases antioxidant defenses [154]. It has been reported that lead damages the cell membrane of hepatocytes and its DNA [155].

Cadmium causes liver damage mainly by induction ROS inducing lipoperoxidation via Fenton reaction [156]. The increment of ROS induces DNA damage, proteins oxidation and lipoperoxidation. Cadmium replaces iron and copper in the Fenton reaction. It is also capable of moving to zinc from proteins and changing their structure [157]. Chronic liver exposure to cadmium induces liver failure [158].

Mercury toxicity is the consequence of its high affinity to sulphydric groups in proteins and enzymes involved in cell cycle progression [159] [160]. It induces hepatocyte apoptosis causing acute liver failure [161].

Liver is the main organ for the metabolism of iron and it is also the target of its pathological accumulation, as a consequence of a metabolic disease, such as hemochromatosis or because of an increased exposure [162]. Iron accumulates in the hepatocytes in which induces the formation of hydroxyl radicals ( $\cdot$ OH) from reduced forms of  $O_2$ , ending in oxidative stress [163].

Copper is associated with Wilson's disease resulting in its liver accumulation, because of the reduced metal elimination by bile duct [164]; air pollution exposure also causes liver accumulation and injury [165]. Copper and iron share the same mechanism of damage causing an increase in reactive species in liver parenchyma ending in fibrosis and cirrhosis [166].

Vanadium is other metal that alters liver function, inducing ROS via Fenton reaction, damaging proteins and altering the genetic material [167]. Inhalation of  $V_2O_5$  induces alterations in liver function tests with an increase in ALT y AST, as well as hepatic megalocytosis [168].

### 2.10. Pancreas and metals

Pancreatic parenchyma damage by metals has been poorly analyzed, but some reports mention acute and chronic pancreatitis, and cancer [169]. The pathophysiology and etiology of pancreatic damage are still unknown, and usually are lethal [170] [171] [172]. Vanadium induces an increase in pancreatic enzymes, hypertrophic acinar cells, which results in an acute pancreatitis [173].

### 2.11. Immunotoxicity of metals

Urban populations are often exposed to metals as constituent of particulate matter (PM), one aspect of the myriad toxicities that might arise from these exposures is altered lymphoid system and thus immune responses. Among the metals that when inhaled damage the lymphoid system we can find vanadium, cadmium, mercury, iron, lead, manganese, chromium, copper and arsenic.

Among the main effects of vanadium that our group has reported we can find splenomegaly. Spleens of mice exposed to vanadium showed morphological changes that included an increase in the size of the white pulp, germinal center hyperplasia, and an increase in the size and number of megakaryocytes and CD19+ lymphocytes. In the same study we found a decrease in the mice capacity to star a humoral immune response, when presented to Hepatitis B surface antigen (HBsAg), vanadium exposed mice presented higher antibodies concentrations with lower affinity compared to controls [174]. On the other hand, our group, in the thymus, has reported morphological changes. We found that vanadium exposed mice presented a shift of the normal cortex-medulla relationship, showing a much thinner medulla and the presence of medulla-like areas within the cortex regions. These changes suggest an alteration of the immune response [69, 89]. In addition to these findings we have reported a decrease in the presence of CD11c, a dendritic cell marker, and MHCII an antigen presenting cell marker, in the thymus of vanadium-exposed mice. This study was conducted using two methods, immunohistochemistry and FACS, with similar findings. This hyperplasia downturn could be detrimental for the negative selection of thymocytes, as dendritic cells are closely related to this process, leading to the persistence of self-reactive cells and increased risk of autoimmune diseases [175].

On the other hand, it has been shown that mice exposed to low concentrations of cadmium have an enhanced humoral immune response [176], however, exposing rats to high concentrations of Cd results in the decrease of B and T cell function and impairment of the phagocytic

capacity of NK cells [177]. Chronic exposure to Cd increases serum concentrations of diverse pro-inflammatory cytokines such as IL-1 $\beta$ , TNF- $\alpha$  and IL-6. Chronic exposure has also been associated to splenomegaly, alterations in the histology of the spleen and the appearance of giant cells and fibrosis. In the thymus, atrophy evidenced as a decreased weight of the thymus has been reported with this metal. All these toxic effects can lead to disturbances in the immune selection and response [178].

One of the worst threats mercury inhalation imposes, is the development of autoimmunity in genetically predisposed individuals, chronic exposure is capable of inducing an immunosuppressive state, alongside apoptotic defects that can lead to a syndrome similar to that of Lupus. The mechanism through which this occurs is not entirely understood. It has been proposed that Hg can associate with proteins creating large complexes capable of activating the immune system, this is specially true for molecules present in antigen presenting cells leading to a massive activation of T cells [179]. Hg is not only capable of inducing immunosuppression, it can also induce immunostimulation in both, mice and humans, its exact mechanism is not known yet [180].

Iron is an essential element for metabolic processes occurring in both, human and microbial cells. Therefore its relationship with immune function is evident. There is a hypothesis that the persistence of certain extracellular pathogens in circulation induces an iron restriction in the mononuclear phagocyte system, blocking its phagocytic capacity. As a result changing concentrations of Fe in the system caused by exposure to this metal could alter the immune function [181].

Lead exposure through inhalation can affect the immune system, an increase in circulating concentrations of IL-4 and IFN- $\gamma$  and leukocytosis have been reported in a murine model [182]. The effects of lead on the immune system have been studied using macrophages. Being present in a diverse range of tissues, any adverse effect on them could be associated with several presentations. It has been shown that lead diminishes the phagocytic capacity of macrophages, which plays a central role in innate immunity, and therefore could redirect the response towards a Th2 or antibody producing response which could in turn favor the development of antibody mediated autoimmunity. In the presence of lead NO decreases and with it the macrophages capacity to kill pathogens. Due to Pb the membrane of erythrocytes gets damaged leading to anemia by increasing the rate at which the spleen phagocytes damaged red cells. It has been reported that macrophages can increase their production of TNF- $\alpha$  due to Pb exposure, and with these can damage peripheral tissue. This has been proven in two models; in the first peritoneal macrophages exposed to Pb damaged liver tissue and in the nervous system Pb exposed microglia damaged peripheral neurons [183].

An immunosuppressive state has been documented in workers exposed to Mn inhalation. In an experimental model using rats, Mn decreased circulated populations of CD4<sup>+</sup> and CD8<sup>+</sup> lymphocytes that could explain the immunosuppressive state reported in the work exposure studies [184].

Other metal with immunotoxic properties is Chromium. Occupational exposure to this metal can lead to an imbalance of the humoral and cellular components of the immune response. In

a study conducted on workers exposed to inhaled Cr, it was found that circulating levels of immunoglobulins and complement components were different when compared to their controls. Seric titers of IgG and IgA were lower than those found in the control group, whilst C3 and C4 concentrations were higher. These results suggest that exposure to Cr can produce ill effects on lymphocytes that include inhibition of immunoglobulin secretion and complement activation [185].

Copper inhalation by humans is more frequently found as an occupational exposure. This metal can cause morphological changes in lymphoid tissue. In a murine study Cr inhalation led to an increase in spleen weight and decrease in thymus weight. Morphologically splenomegaly and thymus atrophy were also reported. All of the above could lead to an immunological imbalance [186].

Studies with Arsenic have been conducted using pregnant women that live in rural areas contaminated with this metal. In this study their IgG titers were elevated compared to non-exposed pregnant women from a different region. The study suggest that a higher titer of IgG can increase the pregnant women mortality, however, the overall effect on humoral response that these elevation in IgG can have is still unknown [187].

The immune system is an important and complex system composed of different kind of cells and factors whose function is of the utmost importance in preserving health. This system is on of the most sensible targets of atmospheric pollution. Metals associated with particulate matter can cause an ample specter of immunological disorders. The susceptibility to these disorders, however, can be deeply related to a genetic predisposition in the individual.

## 2.12. Reprotoxicity of metals

Air pollution has been related to adverse effects on female reproductive health such as infertility, miscarriages, delay menarche [188] and an increased risk of hypertensive disorders during pregnancy [189]. Metals contained in particulate matter could be related with these reproductive alterations [190], because it has been proved that some metals like cadmium, lead, mercury, manganese, chromium and nickel have female reprotoxic effects [191].

Cadmium increased the duration of estrous cycle in rats exposed to 1 mg Cd/m<sup>3</sup> (NTP, 1995); as well as after 6 weeks of exposure to 1 mg Cd/m<sup>3</sup> and after 18 weeks after exposure to 0.16 mg Cd/m<sup>3</sup> (5h/5 days/20 weeks) [192]. Female mice exposed to 230 µg Cd/m<sup>3</sup> daily showed a lower incidence of pregnancies, and a lower level of serum 17-β estradiol [193]. Inhalation of 1 mg Cd/m<sup>3</sup> (5 h daily/5 days weekly/5 months) caused a decreased female rats fertility [194].

Mercury vapor has reproductive effects in occupationally exposed women (dental assistants and dentists), reports included abortion, stillbirth and menstrual disorders (irregularity, painful or hemorrhagic menstrual bleeding) [195]. Polymenorrhoea or oligomenorrhoea [196] and reduced fertility [197] it was also observed in women working in a lamp factory and dental assistants, respectively. In animal models, Davis and coworkers [198] observed longer estrous cycles in rats exposed to 2 mg Hg/m<sup>3</sup> (2h/day/11days).

Female workers at a lead smelter showed an increased frequency of abortions and their child showed low birth weight [199]. In pregnant women living in Mexico City, high levels of lead in maternal blood were related with an increased incidence of abortions [200]. Vigeh and coworkers [201] observed high levels of lead and manganese in pregnant women diagnosed with preeclampsia.

MnO<sub>2</sub> dust inhalation augments the number of pups and decreased their body weight gain, when female rats were exposed preconception [202]. In other study rats transfer manganese to their offspring through milk, after preconception exposure to manganese inhalation [203].

Inhaled chromium caused menstrual alterations, postnatal hemorrhage and delivery complication in female workers and near-resident women [204]. In female workers at a nickel refinery plant, it was observed an increased incidence of abortions [205]. In other study, high levels of nickel were found in women with endometriosis [206].

Vanadium inhalation caused an increased length of estrous cycle in females exposed to 4.5 mg V/m<sup>3</sup>, and exposure to 9 mg V/m<sup>3</sup> reduced the number of females with a normal cycle (NTP, 2002). After inhalation of V<sub>2</sub>O<sub>5</sub> 0.02M (1h/week/4 weeks) female mice get into anestrus and showed lower serum levels of estradiol and progesterone and an increase in the width of uterine stroma and myometrium [207]; as well as an increase in the lipidic peroxidation in the ovary and a reduced size of secondary and preovulatory follicles [208].

Bucher and coworkers [209] studied the effect of inhalation of copper sulfate (3 mg/m<sup>3</sup>, 6 h daily, 5 days per week for 13 weeks) in rats; they find alterations in sperm (decreased motility and abnormal sperm increase) and in testicular weight.

In male rats exposed to inhalation of MnSO<sub>4</sub> (3 mg/m<sup>3</sup>) for 6 h/day for 7 days a week, Dorman and coworkers (2001) quantified an increase in the concentration of MnSO<sub>4</sub> in the testes of treated animals (0.79 ± 0.18 µg / mg of dry tissue) compared with the testes of control animals (0.32 ± 0.04 µg / mg of dry tissue).

In mice exposed by inhalation of lead and cadmium has been reported mitochondrial damage in Sertoli cells of mice exposed to inhalation of lead acetate (0.01 M, 1 h / week/4 weeks) and chloride cadmium (0.006 M, 1h / week / 4 weeks); in addition, mitochondrial alterations were more severe and an detected earlier in animals exposed together to both compounds than in controls [210].

In mice exposed to inhaled vanadium pentoxide (0.02 M) for 1 h twice a week, for 12 weeks, alterations were observed in the cells of the seminiferous tubules: necrosis, pseudo-nuclear inclusions and disruption of cellular junctions [211]; alterations were also found in proteins of the, such as decrease of gamma-tubulin [212] and actin [213].

In humans, it has been proposed that exposure to toxic metals is a risk factor in reproductive health. A study by Akinloye and coworkers [214] indicated that the cadmium concentration in serum and seminal fluid from azoospermic men was higher than in oligozoospermic and control men.

In the case of lead, there is evidence of its reproductive toxicity effects in humans. Occupational exposure to lead causes decreased sperm motility and dysfunction of the sex glands [215].

### 2.13. Teratogenesis

During the last decade, epidemiologic studies have researched the connection between air pollution and its adverse effects during pregnancy, being found an increase in preterm birth risk, low birth weight and foetus underdevelopment; however, results are contradictory because of different methodological approaches [216]. A study held in California showed the link between the exposure to PM<sub>2.5</sub> and low birth weight in children born at the end of pregnancy. The particles had sulphide, sulphate, vanadium, iron, manganese, bromine, ammonium and zinc. These particles can affect the fetus weight because of their impact in cardiovascular and respiratory health in the mother, produce oxidative stress and damage the fetus DNA, affecting in its development [217]. An experimental study of mouse in utero exposure to diesel emissions, a pollutant that cause the major number of PM<sub>2.5</sub> suspended particles, demonstrated that there is embryonic reabsorption and placental changes such as hemorrhage, necrosis, swelling and oxidative stress. In adulthood it was found propensity to arterial hypertension and cardiac failure in mice that were exposed to diesel during prenatal development [218].

Waste incinerators produce environmental pollutants such as heavy metals, specifically cadmium, lead, mercury, chromium and arsenic. A study held in Cumbria, UK, found an excessive number of perinatal and child mortality because of spina bifida and cardiac malformations near to incineration places [219].

Heavy metals can produce health problems because of oxidative stress (Cd, Cr, Pb, As), neurological damage (Pb, Hg), DNA damage (As, Cr, Cd), changes in the metabolism of glucose (As) and calcium (Cd, Pb) and interfere with essential elements (Cd, Hg) [220]. It is because of this that these metals have a teratogenic potential, which depends also on its placental transportation. The placenta is an active transporter of essential elements, such as calcium, copper, zinc and iron, as well as toxic elements such as cadmium, lead, mercury and nickel. Heavy metals can go through the placental barrier and accumulate in the fetus tissues and amniotic fluid.

Cadmium accumulates in the placenta, and it has been found a correlation between cadmium levels and the expression of the metallothionein, which retains the cadmium and prevents that it reaches the fetus; but its increase blocks the transportation of Zn to the fetus, decreasing the placental permeability to this essential element. Cadmium also affects the synthesis of the placental hormones such as progesterone and leptin, affects the trophoblastic cells migration and induces an early development on the decidua of the endometrial stroma [220]. The maternal exposure to cadmium during pregnancy is linked to preterm birth, intrauterine growth restriction and low birth weight [220, 221]. In experimental models it has demonstrated that it can affect the embryo development and the implantation. Cadmium increases in rats the oxidative stress and decreases the antioxidant enzymes activity [221]. The exposure of rats to cadmium during the organogenesis period produces external and internal malformations, as well as alterations in the ossification. After birth it was observed an alteration in males and females sexual behavior [222, 223]. The exposure to cadmium in human fetal gonads in culture produces a decrease of germinal cells due to apoptosis, but it does not affect the cells proliferation [223].

Lead can easily go through the placental barrier by simple diffusion. Lead can affect calcium-mediated processes in the syncytiotrophoblast; it accumulates there and reduces the cytochrome oxidase activity, an enzyme of the respiratory chain, reducing therefore the ATP production in this cells. Lead produces oxidative damage and induces preterm birth, abortion, intrauterine growth restriction and congenital abnormalities [220]. Chromium that can be found in the polluted air has been linked to neuroblastoma in children [224].

Arsenic has been linked to miscarriage, low birth weight and malformations in populations working or living near foundries in which emissions arsenic can be found. In animal models arsenic produces toxicity in development, since it produces malformations, intrauterine death and intrauterine growth restriction. These malformations include neural tube defects, gonadal and renal agenesis, eye defects and malformations of the ribs [225].

Manganese can be found in the air due to, essentially, diesel combustion. Manganese goes through the placental barrier by simple diffusion. Manganese is an essential nutrient, and that is why it can be found in the tissues and fluids of both, mother and foetus. The lack or excess of manganese affect the prenatal development. Because of the air pollution an excess of manganese has been linked to intrauterine growth restriction during the foetal period and, postnatally, to hyperactive behavior, decrease of intellectual ability and alteration of the psychomotor development [226]. Experimental studies have showed that the excess of manganese has a teratogenic effect, since that produces growth restriction, embryonic death and bone alterations [227].

High levels of iron during the human embryonic period can be teratogenic. Iron is toxic mainly because it produces oxidative stress. Iron catalyzes the production of hydroxyl-free radicals, which destroy cells by lipid peroxidation, enzymes denaturation, carbohydrates depolymerization and ruptures in DNA. Experimental studies in the mouse show that an increase of iron produces histological alterations in the encephalon, as well as spine and ribs malformations [228].

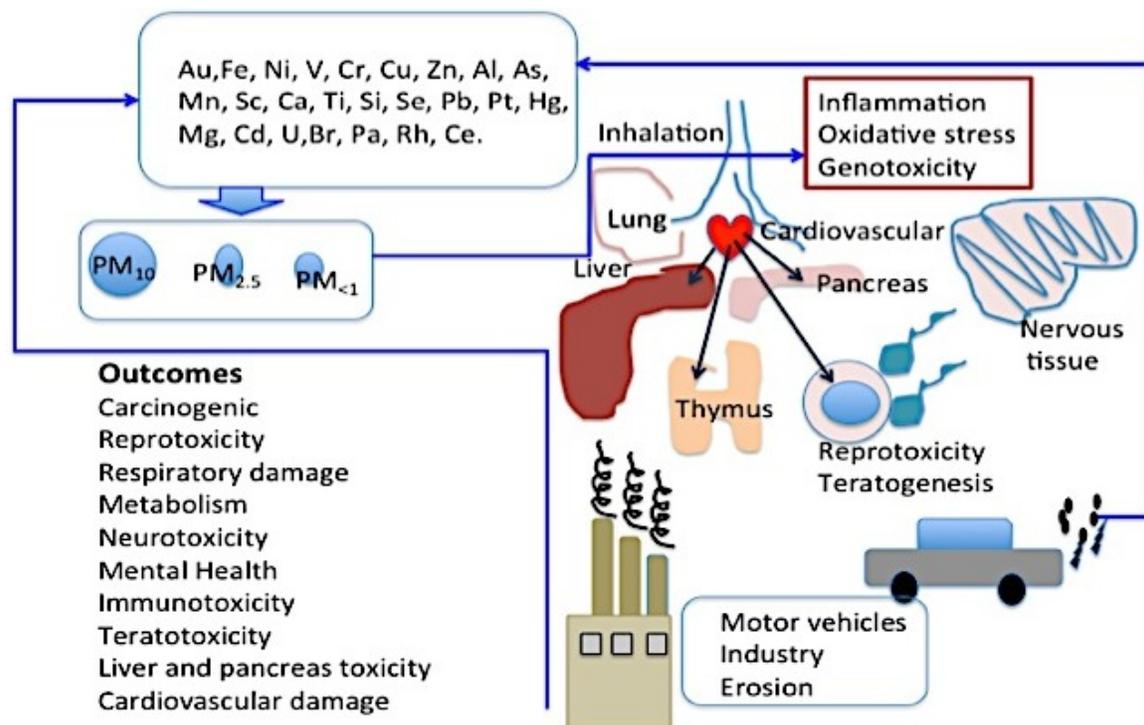
Equally, high levels of nickel affect the mouse embryo development, producing embryonic death, fetal death, and malformations such as hydrocephalus, eyelids alterations, microphthalmia, exophthalmia, clubfoot, umbilical hernia and bone anomalies. The excess in nickel ions can replace other metals required for the structure and functions of enzymes, which get inactivated, and this could be the cause of the embryo and fetus toxicity observed in mice, rats and women [229].

Vanadium causes reproductive damage. It goes through the placenta, constituting complexes with transferrin or albumin, and accumulates in the placenta and the fetal tissues. It has been found that vanadium affects the prenatal development, since it produces embryo mortality, fetus toxicity and teratogenicity in mice, rats, hamsters and chickens [69, 89]. In humans, it has been proved that the exposure to vanadium during pregnancy is linked to low birth weight. In the chicken embryo, the vanadium pentoxide produces embryo mortality and teratogenicity, since it provokes alterations in tubulation and the central nervous system, microphthalmia, abnormalities in the pharyngeal arches and facial development processes or their derivatives, congenital heart defect, limbs malformations and visceral ectopia [89]. Vanadium produces

oxidative damage in proteins, lipids and DNA, interferes with DNA repair and affects cellular signaling pathways and cells proliferation [230].

### 3. Conclusion

Metals enter into the respiratory system adhered to particulate matter, and by this route they reach the systemic circulation. In the blood metals are attached to proteins or ionized entering into the different organs and cells producing a variety of outcomes. The chemical characteristics of the inhaled metals, the length of the exposure, the route, and the physiology of each organ will determinate the metabolism, the affected functions and the possible manifestations, which are resumed in Figure 1. Some metals interact with enzymes inhibiting its actions by the interaction of the metal with the SH group of the enzyme or by displacement of an essential metal cofactor; another interference mechanism is the inhibition of the synthesis of the enzyme, indirectly altering the systemic function; the binding of the metal by certain cytosolic proteins may modify its toxicity. Also metals may interact directly with the components of the cell, and may be accumulated in the lysosomes, or damaging the mitochondria and inhibiting respiratory enzymes leading to cell death. A direct interaction with DNA may produce gene mutations, chromosome aberrations or aneuploidy; these changes could pave the way to proliferation and cancer development [231].



**Figure 1.** Interaction of metals in particulate matter that enters into the respiratory system inducing inflammation, oxidative stress and genotoxicity. The sources and possible outcomes are resumed.

The problem of metals associated with atmospheric particulate matter is not new and there are increasing reports about its health effects, as we have mentioned in the previous sections. Even though the toxic effects of these elements are severe, there are scant specific knowledge about the association of a disease and the inhalation exposure. This problem needs more research in order to count with more information for understanding the mechanisms of damage and to propose measures to control the emissions, decrease the exposure and its adverse effects.

## Acknowledgements

Authors thank Alejandra Núñez-Fortoul for reviewing the final English version.

## Author details

T.I. Fortoul<sup>1\*</sup>, V. Rodriguez-Lara<sup>1</sup>, A. Gonzalez-Villalva<sup>1</sup>, M. Rojas-Lemus<sup>1</sup>, L. Colin-Barenque<sup>2</sup>, P. Bizarro-Nevarés<sup>1</sup>, I. García-Peláez<sup>1</sup>, M. Ustarroz-Cano<sup>1</sup>, S. López-Zepeda<sup>1</sup>, S. Cervantes-Yépez<sup>1</sup>, N. López-Valdez<sup>1</sup>, N. Meléndez-García<sup>1</sup>, M. Espinosa-Zurutuza<sup>3</sup>, G. Cano-Gutierrez<sup>4</sup> and M.C. Cano-Rodríguez<sup>4</sup>

\*Address all correspondence to: fortoul@unam.mx

1 Cellular and Tissular Biology Department, School of Medicine, National Autonomous University of Mexico (UNAM). Mexico City, Mexico

2 Neuromorphology Laboratory, FES Iztacala, National Autonomous University of Mexico (UNAM), Mexico

3 Faculty of Sciences, Biology, National Autonomous University of Mexico (UNAM) Mexico City, Mexico

4 Valle de Mexico University (Coyoacan) School of Health Sciences, Mexico City, Mexico

## References

- [1] Englert N. Fine particles and human health--a review of epidemiological studies. *Toxicology letters*. 2004;149(1-3):235-42.
- [2] Kelly FJ, Fussell JC. Size, source and chemical composition as determinants of toxicity attributable to ambient particulate matter. *Atmos Environ*. 2012; 60:504-26.

- [3] Bu Changming WL HL, Tang Jing.. Evaluation of Health Effects of Fine Particulate PM<sub>2.5</sub>: a Review. *Advanced Materials Research* 2013; 790:441-4.
- [4] Goyer R, Clarkson T. Toxic effects of metals. In: Klaassen C, editor. *Casarett and Doull's Toxicology The basic science of poisons*. 6th Ed ed. New York: McGraw-Hill; 2001.
- [5] Anderson JO, Thundiyil JG, Stolbach A. Clearing the air: a review of the effects of particulate matter air pollution on human health. *Journal of medical toxicology : official journal of the American College of Medical Toxicology*. 2012;8(2):166-75.
- [6] Loomis D, Grosse Y, Lauby-Secretan B, El Ghissassi F, Bouvard V, Benbrahim-Tallaa L, et al. The carcinogenicity of outdoor air pollution. *Lancet Oncol*. 2013;14(13):1262-3.
- [7] Wiseman CL, Zereini F. Airborne particulate matter, platinum group elements and human health: a review of recent evidence. *The Science of the total environment*. 2009;407(8):2493-500.
- [8] Cao S, Duan X, Zhao X, Ma J, Dong T, Huang N, et al. Health risks from the exposure of children to As, Se, Pb and other heavy metals near the largest coking plant in China. *The Science of the total environment*. 2014;472:1001-9.
- [9] Mumford L. *The conduct of life*. Co Ba, editor. New York: Harcord; 1951.
- [10] Kessler R. Air of danger. *Sci Am*. 2014;311(1):S16-7.
- [11] Matus K, Nam KM, Selin NE, Lamsal LN, Reilly JM, Paltsev S. Health damages from air pollution in China. *Global Environ Chang*. 2012;22(1):55-66.
- [12] Soto-Jimenez MF, Flegal AR. Childhood lead poisoning from the smelter in Torreon, Mexico. *Environ Res*. 2011;111(4):590-6.
- [13] Fortoul TI SR, Monca SG, Sanchez IG, Lopez IE, Espejel G, Calderon NL, Saldivar L.. Ultrastructural findings in the murine nonciliated bronchiolar cells (NCBC) after sub-acute inhalation of lead acetate. *Acta Vet Brno* 1999;68:51-5.
- [14] Fortoul TI, Avila-Costa MR, Espejel-Maya G, Mussali-Galante P, Avila-Casado Mdel C, Hernandez-Serrato MI, et al. Metal mixture inhalation (Cd-Pb) and its effects on the bronchiolar epithelium. An ultrastructural approach. *Toxicology and industrial health*. 2004;20(1-5):69-75.
- [15] Fortoul TI, Saldivar OL, Espejel-Maya G, Bazarro NP, Mussali-Galante P, Avila-Casado Mdel C, et al. Inhalation of cadmium, lead or its mixture Effects on the bronchiolar structure and its relation with metal tissue concentrations. *Environmental toxicology and pharmacology*. 2005;19(2):329-34.
- [16] Stavrides JC. Lung carcinogenesis: pivotal role of metals in tobacco smoke. *Free Radic Biol Med*. 2006;41(7):1017-30.

- [17] Bohlandt A, Schierl R, Diemer J, Koch C, Bolte G, Kiranoglu M, et al. High concentrations of cadmium, cerium and lanthanum in indoor air due to environmental tobacco smoke. *Sci Total Environ*. 2012;414:738-41.
- [18] Cakmak S, Dales R, Kauri LM, Mahmud M, Van Ryswyk K, Vanos J, et al. Metal composition of fine particulate air pollution and acute changes in cardiorespiratory physiology. *Environ Pollut*. 2014;189:208-14.
- [19] Bell ML, Zanobetti A, Dominici F. Evidence on vulnerability and susceptibility to health risks associated with short-term exposure to particulate matter: a systematic review and meta-analysis. *Am J Epidemiol*. 2013;178(6):865-76.
- [20] Martinelli N, Olivieri O, Girelli D. Air particulate matter and cardiovascular disease: a narrative review. *Eur J Intern Med*. 2013;24(4):295-302.
- [21] Franchini M, Guida A, Tufano A, Coppola A. Air pollution, vascular disease and thrombosis: linking clinical data and pathogenic mechanisms. *J Thromb Haemost*. 2012;10(12):2438-51.
- [22] Johnston FH, Hanigan IC, Henderson SB, Morgan GG. Evaluation of interventions to reduce air pollution from biomass smoke on mortality in Launceston, Australia: retrospective analysis of daily mortality, 1994-2007. *BMJ*. 2013;346:e8446.
- [23] Pope CA, 3rd, Ezzati M, Dockery DW. Fine-particulate air pollution and life expectancy in the United States. *N Engl J Med*. 2009;360(4):376-86.
- [24] Miller MR, Shaw CA, Langrish JP. From particles to patients: oxidative stress and the cardiovascular effects of air pollution. *Future Cardiol*. 2012;8(4):577-602.
- [25] Niu J, Liberda EN, Qu S, Guo X, Li X, Zhang J, et al. The role of metal components in the cardiovascular effects of PM2.5. *PLoS One*. 2013;8(12):e83782.
- [26] Jomova K, Valko M. Advances in metal-induced oxidative stress and human disease. *Toxicology*. 2011;283(2-3):65-87.
- [27] Kruszewski M. The role of labile iron pool in cardiovascular diseases. *Acta Biochim Pol*. 2004;51(2):471-80.
- [28] Mordukhovich I, Wright RO, Hu H, Amarasiriwardena C, Baccarelli A, Litonjua A, et al. Associations of toenail arsenic, cadmium, mercury, manganese, and lead with blood pressure in the normative aging study. *Environ Health Perspect*. 2012;120(1):98-104.
- [29] Flora G, Gupta D, Tiwari A. Toxicity of lead: A review with recent updates. *Interdiscip Toxicol*. 2012;5(2):47-58.
- [30] Alissa EM, Ferns GA. Heavy metal poisoning and cardiovascular disease. *J Toxicol*. 2011;2011:870125.
- [31] Hartwig A. Cadmium and cancer. *Met Ions Life Sci*. 2013;11:491-507.

- [32] Huff J, Lunn RM, Waalkes MP, Tomatis L, Infante PF. Cadmium-induced cancers in animals and in humans. *Int J Occup Environ Health*. 2007;13(2):202-12.
- [33] Koedrith P, Seo YR. Advances in carcinogenic metal toxicity and potential molecular markers. *Int J Mol Sci*. 2011;12(12):9576-95.
- [34] Silva N, Peiris-John R, Wickremasinghe R, Senanayake H, Sathiakumar N. Cadmium a metalloestrogen: are we convinced? *J Appl Toxicol*. 2012;32(5):318-32.
- [35] Benbrahim-Tallaa L, Tokar EJ, Diwan BA, Dill AL, Coppin JF, Waalkes MP. Cadmium malignantly transforms normal human breast epithelial cells into a basal-like phenotype. *Environ Health Perspect*. 2009;117(12):1847-52.
- [36] Silbergeld EK. Facilitative mechanisms of lead as a carcinogen. *Mutat Res*. 2003;533(1-2):121-33.
- [37] Garcia-Leston J, Mendez J, Pasaro E, Laffon B. Genotoxic effects of lead: an updated review. *Environ Int*. 2010;36(6):623-36.
- [38] Schaumloffel D. Nickel species: analysis and toxic effects. *J Trace Elem Med Biol*. 2012;26(1):1-6.
- [39] Costa M. Molecular mechanisms of nickel carcinogenesis. *Biol Chem*. 2002;383(6):961-7.
- [40] Oller AR, Costa M, Oberdorster G. Carcinogenicity assessment of selected nickel compounds. *Toxicol Appl Pharmacol*. 1997;143(1):152-66.
- [41] Kasprzak KS, Sunderman FW, Jr., Salnikow K. Nickel carcinogenesis. *Mutat Res*. 2003;533(1-2):67-97.
- [42] Ishida S, Andreux P, Poitry-Yamate C, Auwerx J, Hanahan D. Bioavailable copper modulates oxidative phosphorylation and growth of tumors. *Proc Natl Acad Sci U S A*. 2013;110(48):19507-12.
- [43] Linder MC, Moor JR, Wright K. Ceruloplasmin assays in diagnosis and treatment of human lung, breast, and gastrointestinal cancers. *J Natl Cancer Inst*. 1981;67(2):263-75.
- [44] Ishida S, McCormick F, Smith-McCune K, Hanahan D. Enhancing tumor-specific uptake of the anticancer drug cisplatin with a copper chelator. *Cancer Cell*. 2010;17(6):574-83.
- [45] Theophanides T, Anastassopoulou J. Copper and carcinogenesis. *Crit Rev Oncol Hematol*. 2002;42(1):57-64.
- [46] Barregard L, Sallsten G, Jarvholm B. Mortality and cancer incidence in chloralkali workers exposed to inorganic mercury. *Br J Ind Med*. 1990;47(2):99-104.
- [47] Gibbs GW, Labreche F. Cancer risks in aluminum reduction plant workers: a review. *J Occup Environ Med*. 2014;56(5 Suppl):S40-59.

- [48] Martin SC, Lariviere C. Community health risk assessment of primary aluminum smelter emissions. *J Occup Environ Med*. 2014;56(5 Suppl):S33-9.
- [49] Gibbs GW, Labreche F, Busque MA, Duguay P. Mortality and cancer incidence in aluminum smelter workers: a 5-year update. *J Occup Environ Med*. 2014;56(7):739-64.
- [50] Lee JC, Son YO, Pratheeshkumar P, Shi X. Oxidative stress and metal carcinogenesis. *Free Radic Biol Med*. 2012;53(4):742-57.
- [51] Galanis A, Karapetsas A, Sandaltzopoulos R. Metal-induced carcinogenesis, oxidative stress and hypoxia signalling. *Mutat Res*. 2009;674(1-2):31-5.
- [52] Salnikow K, Zhitkovich A. Genetic and epigenetic mechanisms in metal carcinogenesis and cocarcinogenesis: nickel, arsenic, and chromium. *Chem Res Toxicol*. 2008;21(1):28-44.
- [53] Hartwig A. Metal interaction with redox regulation: an integrating concept in metal carcinogenesis? *Free Radic Biol Med*. 2013;55:63-72.
- [54] Beguin Y, Aapro M, Ludwig H, Mizzen L, Osterborg A. Epidemiological and non-clinical studies investigating effects of iron in carcinogenesis--a critical review. *Crit Rev Oncol Hematol*. 2014;89(1):1-15.
- [55] Durackova Z. Some current insights into oxidative stress. *Physiol Res*. 2010;59(4):459-69.
- [56] IEH. Occupational exposure limits: Criteria document for manganese and inorganic manganese compounds. 2004.. Available from: Available at: <http://www.cranfield.ac.uk/health/researchareas/environmenthealth/ieh/ieh%20publications/w17.pdf>.
- [57] Valko M, Morris H, Cronin MT. Metals, toxicity and oxidative stress. *Curr Med Chem*. 2005;12(10):1161-208.
- [58] Beyersmann D, Hartwig A. Carcinogenic metal compounds: recent insight into molecular and cellular mechanisms. *Arch Toxicol*. 2008;82(8):493-512.
- [59] (IARC) IAfRoC. Vanadium pentoxide, IARC Monogr. Eval. Carcinog. Risks Hum.. 2003;83:227-43.
- [60] Assem FL, Levy LS. A review of current toxicological concerns on vanadium pentoxide and other vanadium compounds: gaps in knowledge and directions for future research. *J Toxicol Environ Health B Crit Rev*. 2009;12(4):289-306.
- [61] Bishayee A, Waghay A, Patel MA, Chatterjee M. Vanadium in the detection, prevention and treatment of cancer: the in vivo evidence. *Cancer Lett*. 2010;294(1):1-12.
- [62] ICH. GfiSR. Genotoxicity testing and data interpretation for pharmaceuticals intended for human use. 2012. Available from: <http://www.fda.gov/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/default.htm>.

- [63] Sorsa M. Biological Monitoring. In: La Ferla F, Lauwerys, Robert R., Jeanne Mager Stellman editor. Encyclopedia of Occupational Health and safety. Genova: International Labor Organization,; 2011.
- [64] Chorvatovicova D, Kovacikova Z. Inhalation exposure of rats to metal aerosol. II. Study of mutagenic effect on alveolar macrophages. *J Appl Toxicol.* 1992;12(1):67-8.
- [65] Valverde M, Fortoul TI, Diaz-Barriga F, Mejia J, del Castillo ER. Induction of genotoxicity by cadmium chloride inhalation in several organs of CD-1 mice. *Mutagenesis.* 2000;15(2):109-14.
- [66] Valverde M, Fortoul TI, Diaz-Barriga F, Mejia J, del Castillo ER. Genotoxicity induced in CD-1 mice by inhaled lead: differential organ response. *Mutagenesis.* 2002;17(1):55-61.
- [67] Monleau M, De Meo M, Paquet F, Chazel V, Dumenil G, Donnadiou-Claraz M. Genotoxic and inflammatory effects of depleted uranium particles inhaled by rats. *Toxicol Sci.* 2006;89(1):287-95.
- [68] Ehrlich VA, Nersesyan AK, Atefie K, Hoelzl C, Ferk F, Bichler J, et al. Inhalative exposure to vanadium pentoxide causes DNA damage in workers: results of a multiple end point study. *Environ Health Perspect.* 2008;116(12):1689-93.
- [69] Fortoul TI, Rodriguez-Lara V, Gonzalez-Villalva A, Rojas-Lemus M, Cano-Gutierrez G, Ustarroz-Cano M, et al. Vanadium Inhalation in a Mouse Model for the Understanding of Air-Suspended Particle Systemic Repercussion. *J Biomed Biotechnol.* 2011:1-11.
- [70] Rojas-Lemus M, Altamirano-Lozano M, Fortoul TI. Sex differences in blood genotoxic and cytotoxic effects as a consequence of vanadium inhalation: micronucleus assay evaluation. *Journal of Applied Toxicology.* 2014;34(3):258-64.
- [71] Lindberg HK, Falck GC, Catalan J, Koivisto AJ, Suhonen S, Jarventaus H, et al. Genotoxicity of inhaled nanosized TiO<sub>2</sub> in mice. *Mutat Res.* 2012;745(1-2):58-64.
- [72] MohanKumar SM, Campbell A, Block M, Veronesi B. Particulate matter, oxidative stress and neurotoxicity. *Neurotoxicology.* 2008;29(3):479-88.
- [73] Calderon-Garcidueñas L, Torres-Jardon R, Kulesza RJ, Park SB, D'Angiulli A. Air pollution and detrimental effects on children's brain. The need for a multidisciplinary approach to the issue complexity and challenges. *Front Hum Neurosci.* 2014;8:613.
- [74] Ljubimova JY GP, Portilla-Arias J, Patil R, Konda B, Paff M, Markman JL, Inoue S, Espinoza A, Chesnokova A, Funari V, Kleinman MT, Holler E, Black KL.. Molecular changes in rat brain due to air nano pollution.. *Nanotech.* 2012;3:261-4..
- [75] Oberdorster G, Oberdorster E, Oberdorster J. Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect.* 2005;113(7):823-39.

- [76] Win-Shwe TT, Fujimaki H. Nanoparticles and neurotoxicity. *Int J Mol Sci.* 2011;12(9):6267-80.
- [77] Colin-Barenque L, Fortoul T. Oxidative stress and metals. In: Fortoul T, editor. *Metals and toxicological implications in Health.* Kerala: Research Signpost; 2007. p. 15-25.
- [78] Jellinger KA. The relevance of metals in the pathophysiology of neurodegeneration, pathological considerations. *International review of neurobiology.* 2013;110:1-47.
- [79] Modgil S, Lahiri DK, Sharma VL, Anand A. Role of early life exposure and environment on neurodegeneration: implications on brain disorders. *Transl Neurodegener.* 2014;3:9.
- [80] Cannon JR, Greenamyre JT. The role of environmental exposures in neurodegeneration and neurodegenerative diseases. *Toxicol Sci.* 2011;124(2):225-50.
- [81] Charlet L, Chapron Y, Faller P, Kirsch R, Stone AT, Baveye PC. Neurodegenerative diseases and exposure to the environmental metals Mn, Pb, and Hg. *Coordin Chem Rev.* 2012;256(19-20):2147-63.
- [82] Kozłowski H, Luczkowski M, Remelli M, Valensin D. Copper, zinc and iron in neurodegenerative diseases (Alzheimer's, Parkinson's and prion diseases). *Coordin Chem Rev.* 2012;256(19-20):2129-41.
- [83] Wong BX, Duce JA. The iron regulatory capability of the major protein participants in prevalent neurodegenerative disorders. *Front Pharmacol.* 2014;5:81.
- [84] Braidy N, Poljak A, Marjo C, Rutledge H, Rich A, Jayasena T, et al. Metal and complementary molecular bioimaging in Alzheimer's disease. *Front Aging Neurosci.* 2014;6:138.
- [85] Jomova K, Vondrakova D, Lawson M, Valko M. Metals, oxidative stress and neurodegenerative disorders. *Mol Cell Biochem.* 2010;345(1-2):91-104.
- [86] Aschner M, Erikson KM, Herrero Hernandez E, Tjalkens R. Manganese and its role in Parkinson's disease: from transport to neuropathology. *Neuromolecular Med.* 2009;11(4):252-66.
- [87] Singla N, Dhawan DK. Regulatory role of zinc during aluminium-induced altered carbohydrate metabolism in rat brain. *J Neurosci Res.* 2012;90(3):698-705.
- [88] Roos PM, Dencker L. Mercury in the spinal cord after inhalation of mercury. *Basic Clin Pharmacol Toxicol.* 2012;111(2):126-32.
- [89] Fortoul TI, Rodriguez-Lara V, Gonzalez-Villalva A, Rojas-Lemus M, Cano-Gutierrez G, Ustarroz-Cano M, et al. Inhalation of vanadium pentoxide and its toxic effects in a mouse model. *Inorg Chim Acta.* 2014;420:8-15.
- [90] Colín-Barenque L M-HM, Baiza-Gutman LA, Avila-Costa MR, Ordóñez-Librado, JL, Bizarro-Nevares P, Rodriguez-Lara V, Piñón-Zarate G, Rojas-Lemus M, Mussali-Gal-

- ante P, Fortoul TI. 2008.. Matrix metalloproteinases 2 and 9 in central nervous system and its modification after vanadium inhalation. *J Appl Toxicol* 2008;28(6):718-23.
- [91] Ritchie GD, Still KR, Alexander WK, Nordholm AF, Wilson CL, Rossi J, 3rd, et al. A review of the neurotoxicity risk of selected hydrocarbon fuels. *Journal of toxicology and environmental health Part B, Critical reviews*. 2001;4(3):223-312.
- [92] Terzano C, Di Stefano F, Conti V, Graziani E, Petroianni A. Air pollution ultrafine particles: toxicity beyond the lung. *European review for medical and pharmacological sciences*. 2010;14(10):809-21.
- [93] Julvez J, Grandjean P. Neurodevelopmental toxicity risks due to occupational exposure to industrial chemicals during pregnancy. *Industrial health*. 2009;47(5):459-68.
- [94] Bellinger DC. Children's cognitive health: the influence of environmental chemical exposures. *Alternative therapies in health and medicine*. 2007;13(2):S140-4.
- [95] Landrigan P, Nordberg M, Lucchini R, Nordberg G, Grandjean P, Iregren A, et al. The Declaration of Brescia on prevention of the neurotoxicity of metals June 18, 2006. *American journal of industrial medicine*. 2007;50(10):709-11.
- [96] Jarup L. Hazards of heavy metal contamination. *British medical bulletin*. 2003;68:167-82.
- [97] Trejo-Acevedo A, Diaz-Barriga F, Carrizales L, Dominguez G, Costilla R, Ize-Lema I, et al. Exposure assessment of persistent organic pollutants and metals in Mexican children. *Chemosphere*. 2009;74(7):974-80.
- [98] Sinczuk-Walczak H, Szymczak M, Halatek T. Effects of occupational exposure to arsenic on the nervous system: clinical and neurophysiological studies. *International journal of occupational medicine and environmental health*. 2010;23(4):347-55.
- [99] Needleman HL, Geiger SK, Frank R. Lead and IQ scores: a reanalysis. *Science*. 1985;227(4688):701-2, 4.
- [100] Baranowska-Bosiacka I, Gutowska I, Rybicka M, Nowacki P, Chlubek D. Neurotoxicity of lead. Hypothetical molecular mechanisms of synaptic function disorders. *Neurologia i neurochirurgia polska*. 2012;46(6):569-78.
- [101] Mason LH, Harp JP, Han DY. Pb neurotoxicity: neuropsychological effects of lead toxicity. *BioMed research international*. 2014;2014:840547.
- [102] Schoeters G, Den Hond E, Zuurbier M, Naginiene R, van den Hazel P, Stilianakis N, et al. Cadmium and children: exposure and health effects. *Acta Paediatr Suppl*. 2006;95(453):50-4.
- [103] Murata K, Grandjean P, Dakeishi M. Neurophysiological evidence of methylmercury neurotoxicity. *American journal of industrial medicine*. 2007;50(10):765-71.

- [104] Ceccatelli S, Dare E, Moors M. Methylmercury-induced neurotoxicity and apoptosis. *Chemico-biological interactions*. 2010;188(2):301-8.
- [105] Magos L, Clarkson TW. Overview of the clinical toxicity of mercury. *Annals of clinical biochemistry*. 2006;43(Pt 4):257-68.
- [106] Roels HA, Bowler RM, Kim Y, Claus Henn B, Mergler D, Hoet P, et al. Manganese exposure and cognitive deficits: a growing concern for manganese neurotoxicity. *Neurotoxicology*. 2012;33(4):872-80.
- [107] Racette BA, Aschner M, Guilarte TR, Dydak U, Criswell SR, Zheng W. Pathophysiology of manganese-associated neurotoxicity. *Neurotoxicology*. 2012;33(4):881-6.
- [108] Avila DS, Puntel RL, Aschner M. Manganese in health and disease. *Metal ions in life sciences*. 2013;13:199-227.
- [109] Barceloux DG. Vanadium. *J Toxicol Clin Toxicol*. 1999;37(2):265-78.
- [110] Zwolak I. Vanadium carcinogenic, immunotoxic and neurotoxic effects: a review of in vitro studies. *Toxicology mechanisms and methods*. 2014;24(1):1-12.
- [111] Ngwa HA, Kanthasamy A, Jin H, Anantharam V, Kanthasamy AG. Vanadium exposure induces olfactory dysfunction in an animal model of metal neurotoxicity. *Neurotoxicology*. 2014;43:73-81.
- [112] Afeseh Ngwa H, Kanthasamy A, Anantharam V, Song C, Witte T, Houk R, et al. Vanadium induces dopaminergic neurotoxicity via protein kinase Cdelta dependent oxidative signaling mechanisms: relevance to etiopathogenesis of Parkinson's disease. *Toxicology and applied pharmacology*. 2009;240(2):273-85.
- [113] Cervantes Yépez SE. Efecto de la exposición por inhalación de vanadio en las células gliales de Müller (MGC) de la retina en un modelo murino. México: Universidad Nacional Autónoma de México.; 2014.
- [114] Quezada Maldonado EM. Cambios en la histología de la retina de ratones expuestos a la inhalación de pentóxido de vanadio y la participación del estrés oxidante. México: Universidad Nacional Autónoma de México. ; 2013.
- [115] Zhou S, Yuan Q, Li W, Lu Y, Zhang Y, Wang W. Trace metals in atmospheric fine particles in one industrial urban city: spatial variations, sources, and health implications. *J Environ Sci (China)*. 2014;26(1):205-13.
- [116] dos Anjos Paulino S, Oliveira RL, Loyola J, Minho AS, Arbilla G, Quiterio SL, et al. Trace metals in PM10 and PM 2.5 samples collected in a highly industrialized chemical/petrochemical area and its urbanized surroundings. *Bull Environ Contam Toxicol*. 2014;92(5):590-5.
- [117] Chillrud SN, Grass D, Ross JM, Coulibaly D, Slavkovich V, Epstein D, et al. Steel dust in the New York City subway system as a source of manganese, chromium, and iron exposures for transit workers. *J Urban Health*. 2005;82(1):33-42.

- [118] Fernandez-Real JM, Lopez-Bermejo A, Ricart W. Cross-talk between iron metabolism and diabetes. *Diabetes*. 2002;51(8):2348-54.
- [119] Ferrannini E. Insulin resistance, iron, and the liver. *Lancet*. 2000;355(9222):2181-2.
- [120] Hatunic M, Finucane FM, Brennan AM, Norris S, Pacini G, Nolan JJ. Effect of iron overload on glucose metabolism in patients with hereditary hemochromatosis. *Metabolism*. 2010;59(3):380-4.
- [121] Chang JW, Chen HL, Su HJ, Liao PC, Guo HR, Lee CC. Simultaneous exposure of non-diabetics to high levels of dioxins and mercury increases their risk of insulin resistance. *J Hazard Mater*. 2011;185(2-3):749-55.
- [122] He K, Xun P, Liu K, Morris S, Reis J, Guallar E. Mercury exposure in young adulthood and incidence of diabetes later in life: the CARDIA Trace Element Study. *Diabetes Care*. 2013;36(6):1584-9.
- [123] Serdar MA, Bakir F, Hasimi A, Celik T, Akin O, Kenar L, et al. Trace and toxic element patterns in nonsmoker patients with noninsulin-dependent diabetes mellitus, impaired glucose tolerance, and fasting glucose. *Int J Diabetes Dev Ctries*. 2009;29(1):35-40.
- [124] Forte G, Bocca B, Peruzzu A, Tolu F, Asara Y, Farace C, et al. Blood metals concentration in type 1 and type 2 diabetics. *Biol Trace Elem Res*. 2013;156(1-3):79-90.
- [125] Ibrahim NM, Eweis EA, El-Beltagi HS, Abdel-Mobdy YE. Effect of lead acetate toxicity on experimental male albino rat. *Asian Pac J Trop Biomed*. 2012;2(1):41-6.
- [126] Adham KG, Al-Eisa NA, Farhood MH. Impact of heavy metal pollution on the hemogram and serum biochemistry of the Libyan jird, *Meriones libycus*. *Chemosphere*. 2011;84(10):1408-15.
- [127] Novakova J, Lukacinova A, Lovasova E, Cimbolakova I, Racz O, Nistiar F. Lifetime exposure to low doses of lead in rats: Effect on selected parameters of carbohydrate metabolism. *Toxicol Ind Health*. 2013; 10.1177/0748233713475510
- [128] Xu X, Rao X, Wang TY, Jiang SY, Ying Z, Liu C, et al. Effect of co-exposure to nickel and particulate matter on insulin resistance and mitochondrial dysfunction in a mouse model. *Part Fibre Toxicol*. 2012;9:40.
- [129] Das Gupta A, Dhara PC, Dhundasi SA, Das KK. Effect of garlic (*Allium sativum*) on nickel II or chromium VI induced alterations of glucose homeostasis and hepatic antioxidant status under sub-chronic exposure conditions. *J Basic Clin Physiol Pharmacol*. 2009;20(1):1-14.
- [130] Praveena SP, S. Sameera, K. Trace elements in diabetes mellitus. *J Clin Diagn Res*. 2013;7(9):1863-5.
- [131] Soveid M, Dehghani GA, Omrani GR. Long- term efficacy and safety of vanadium in the treatment of type 1 diabetes. *Arch Iran Med*. 2013;16(7):408-11.

- [132] Yuen VG, Orvig C, McNeill JH. Glucose-lowering effects of a new organic vanadium complex, bis(maltolato)oxovanadium(IV). *Can J Physiol Pharmacol.* 1993;71(3-4):263-9.
- [133] Mehdi MZ, Pandey SK, Theberge JF, Srivastava AK. Insulin signal mimicry as a mechanism for the insulin-like effects of vanadium. *Cell Biochem Biophys.* 2006;44(1):73-81.
- [134] Marzban L, Rahimian R, Brownsey RW, McNeill JH. Mechanisms by which bis(maltolato)oxovanadium(IV) normalizes phosphoenolpyruvate carboxykinase and glucose-6-phosphatase expression in streptozotocin-diabetic rats in vivo. *Endocrinology.* 2002;143(12):4636-45.
- [135] Boulassel B, Sadeg N, Roussel O, Perrin M, Belhadj-Tahar H. Fatal poisoning by vanadium. *Forensic Sci Int.* 2011;206(1-3):e79-81.
- [136] Mertz W. Chromium in human nutrition: a review. *J Nutr.* 1993;123(4):626-33.
- [137] Lai MH. Antioxidant effects and insulin resistance improvement of chromium combined with vitamin C and E supplementation for type 2 diabetes mellitus. *J Clin Biochem Nutr.* 2008;43(3):191-8.
- [138] Jain SK, Kannan K. Chromium chloride inhibits oxidative stress and TNF-alpha secretion caused by exposure to high glucose in cultured U937 monocytes. *Biochem Biophys Res Commun.* 2001;289(3):687-91.
- [139] Qiao W, Peng Z, Wang Z, Wei J, Zhou A. Chromium improves glucose uptake and metabolism through upregulating the mRNA levels of IR, GLUT4, GS, and UCP3 in skeletal muscle cells. *Biol Trace Elem Res.* 2009;131(2):133-42.
- [140] Garg N, Weinberg J, Ghai S, Bradauskaite G, Nuhn M, Gautam A, et al. Lower magnesium level associated with new-onset diabetes and pre-diabetes after kidney transplantation. *J Nephrol* 2014;27:339-344.
- [141] Ugwuja E NA, Ezenkwa U, Oshim A, Nnabu R, Ogiji E, Ogbanshi M. J. Effects of diabetes complications and glycaemic control on some mineral elements in Nigerians patients with diabetes. *Journal of Diabetology*, 2014;27(3):339-344.;In press.
- [142] Ozgur Keskek S, Kirim S, Karaca A, Saler T. Low serum magnesium levels and diabetic foot ulcers. *Pak J Med Sci.* 2013;29(6):1329-33.
- [143] Adachi Y, Yoshikawa Y, Sakurai H. Antidiabetic zinc(II)-N-acetyl-L-cysteine complex: evaluations of in vitro insulinomimetic and in vivo blood glucose-lowering activities. *Biofactors.* 2007;29(4):213-23.
- [144] Capdor J, Foster M, Petocz P, Samman S. Zinc and glycemic control: a meta-analysis of randomised placebo controlled supplementation trials in humans. *J Trace Elem Med Biol.* 2013;27(2):137-42.

- [145] Taneja SK, Mandal R, Girhotra S. Long term excessive Zn-supplementation promotes metabolic syndrome-X in Wistar rats fed sucrose and fat rich semisynthetic diet. *Indian J Exp Biol.* 2006;44(9):705-18.
- [146] Laing S, Wang G, Briazova T, Zhang C, Wang A, Zheng Z, et al. Airborne particulate matter selectively activates endoplasmic reticulum stress response in the lung and liver tissues. *Am J Physiol Cell Physiol.* 2010;299(4):C736-49.
- [147] Tan HH, Fiel MI, Sun Q, Guo J, Gordon RE, Chen LC, et al. Kupffer cell activation by ambient air particulate matter exposure may exacerbate non-alcoholic fatty liver disease. *J Immunotoxicol.* 2009;6(4):266-75.
- [148] Tarantino G, Capone D, Finelli C. Exposure to ambient air particulate matter and non-alcoholic fatty liver disease. *World J Gastroenterol.* 2013;19(25):3951-6.
- [149] Kovacic P, Somanathan R. Unifying mechanism for metals in toxicity, carcinogenicity and therapeutic action: integrated approach involving electron transfer, oxidative stress, antioxidants, cell signaling and receptors. *J Recept Signal Transduct Res.* 2010;30(2):51-60.
- [150] Casale T, Rosati MV, Ciarrocca M, Samperi I, Andreozzi G, Schifano MP, et al. Assessment of liver function in two groups of outdoor workers exposed to arsenic. *Int Arch Occup Environ Health.* 2014;87(7):745-52.
- [151] Dutta M, Ghosh D, Ghosh AK, Bose G, Chattopadhyay A, Rudra S, et al. High fat diet aggravates arsenic induced oxidative stress in rat heart and liver. *Food Chem Toxicol.* 2014;66:262-77.
- [152] Shi X, Wei X, Koo I, Schmidt RH, Yin X, Kim SH, et al. Metabolomic analysis of the effects of chronic arsenic exposure in a mouse model of diet-induced Fatty liver disease. *J Proteome Res.* 2014;13(2):547-54.
- [153] Xu Z, Wang Z, Li JJ, Chen C, Zhang PC, Dong L, et al. Protective effects of selenium on oxidative damage and oxidative stress related gene expression in rat liver under chronic poisoning of arsenic. *Food Chem Toxicol.* 2013;58:1-7.
- [154] Labudda M., Lead hepatotoxicity: selected aspects of pathobiochemistry. *Med Pr.* 2013;64(4):565-8.
- [155] Mudipalli A. Lead hepatotoxicity & potential health effects. *Indian J Med Res.* 2007;126(6):518-27.
- [156] Fowler BA. Monitoring of human populations for early markers of cadmium toxicity: a review. *Toxicol Appl Pharmacol.* 2009;238(3):294-300.
- [157] Lupo S, Hewitt WR, Rush GF. Cadmium toxicity in the isolated perfused rat liver. *Toxicol Lett.* 1986;34(1):5-11.

- [158] Koyuturk M, Yanardag R, Bolkent S, Tunali S. The potential role of combined anti-oxidants against cadmium toxicity on liver of rats. *Toxicol Ind Health*. 2007;23(7):393-401.
- [159] Landrigan PJ, Wright RO, Birnbaum LS. Mercury toxicity in children. *Science*. 2013;342(6165):1447.
- [160] Bernhoft RA. Mercury toxicity and treatment: a review of the literature. *Journal of environmental and public health*. 2012;2012:460508.
- [161] Gibb H, O'Leary KG. Mercury exposure and health impacts among individuals in the artisanal and small-scale gold mining community: a comprehensive review. *Environmental health perspectives*. 2014;122(7):667-72.
- [162] Ackerman Z, Pappo O, Link G, Glazer M, Grozovski M. Liver Toxicity of Thioacetamide is Increased by Hepatocellular Iron Overload. *Biol Trace Elem Res*. 2014; 10.1007/s12011-014-0110-9.
- [163] Pietrangelo A. Iron-induced oxidant stress in alcoholic liver fibrogenesis. *Alcohol*. 2003;30(2):121-9.
- [164] Uriu-Adams JY, Keen CL. Copper, oxidative stress, and human health. *Mol Aspects Med*. 2005;26(4-5):268-98.
- [165] Pietrangelo A. Metals, oxidative stress, and hepatic fibrogenesis. *Seminars in liver disease*. 1996;16(1):13-30.
- [166] Dong W, Simeonova PP, Gallucci R, Matheson J, Flood L, Wang S, et al. Toxic metals stimulate inflammatory cytokines in hepatocytes through oxidative stress mechanisms. *Toxicol Appl Pharmacol*. 1998;151(2):359-66.
- [167] Dafnis E, Sabatini S. Biochemistry and pathophysiology of vanadium. *Nephron*. 1994;67(2):133-43.
- [168] Cano-Gutierrez G, Acevedo-Nava S, Santamaria A, Altamirano-Lozano M, Cano-Rodriguez MC, Fortoul TI. Hepatic megalocytosis due to vanadium inhalation: participation of oxidative stress. *Toxicol Ind Health*. 2012;28(4):353-60.
- [169] Hall TC, Garcea G, Webb MA, Al-Leswas D, Metcalfe MS, Dennison AR. The socio-economic impact of chronic pancreatitis: a systematic review. *J Eval Clin Pract*. 2014;20(3):203-7.
- [170] Tan CR, Yaffee PM, Jamil LH, Lo SK, Nissen N, Pandol SJ, et al. Pancreatic cancer cachexia: a review of mechanisms and therapeutics. *Front Physiol*. 2014;5:88.
- [171] Das SL, Singh PP, Phillips AR, Murphy R, Windsor JA, Petrov MS. Newly diagnosed diabetes mellitus after acute pancreatitis: a systematic review and meta-analysis. *Gut*. 2014;63(5):818-31.
- [172] Tong GX, Geng QQ, Chai J, Cheng J, Chen PL, Liang H, et al. Association between pancreatitis and subsequent risk of pancreatic cancer: a systematic review of epi-

- miological studies. *Asian Pacific journal of cancer prevention* : APJCP. 2014;15(12):5029-34.
- [173] Cano-Gutierrez GF-R, C. Montaña Luis, F. Rodriguez-Lara, V. and Fortoul, TI. Pancreatic changes and vanadium inhalation. *Current Topics in Toxicology*. 2009;6:39-44.
- [174] Pinon-Zarate G, Rodriguez-Lara V, Rojas-Lemus M, Martinez-Pedraza M, Gonzalez-Villalva A, Mussali-Galante P, et al. Vanadium pentoxide inhalation provokes germinal center hyperplasia and suppressed humoral immune responses. *Journal of immunotoxicology*. 2008;5(2):115-22.
- [175] Ustarroz-Cano M, Garcia-Pelaez I, Pinon-Zarate G, Herrera-Enriquez M, Soldevila G, Fortoul TI. CD11c decrease in mouse thymic dendritic cells after vanadium inhalation. *Journal of immunotoxicology*. 2012;9(4):374-80.
- [176] Fujimaki H, Shimizu F, Kubota K. Suppression of antibody response in mice by acute exposure to nitrogen dioxide: in vitro study. *Environ Res*. 1981;26(2):490-6.
- [177] Cifone MG, Alesse E, Di Eugenio R, Napolitano T, Morrone S, Paolini R, et al. In vivo cadmium treatment alters natural killer activity and large granular lymphocyte number in the rat. *Immunopharmacology*. 1989;18(3):149-56.
- [178] Liu J, Liu Y, Habeebu SS, Klaassen CD. Metallothionein-null mice are highly susceptible to the hematotoxic and immunotoxic effects of chronic CdCl<sub>2</sub> exposure. *Toxicol Appl Pharmacol*. 1999;159(2):98-108.
- [179] Jiang Y, Moller G. In vitro effects of HgCl<sub>2</sub> on murine lymphocytes. I. Preferable activation of CD4<sup>+</sup> T cells in a responder strain. *J Immunol*. 1995;154(7):3138-46.
- [180] Tchounwou PB, Ayensu WK, Ninashvili N, Sutton D. Environmental exposure to mercury and its toxicopathologic implications for public health. *Environ Toxicol*. 2003;18(3):149-75.
- [181] Nairz M, Haschka D, Demetz E, Weiss G. Iron at the interface of immunity and infection. *Front Pharmacol*. 2014;5:152.
- [182] Boskabaddy MH, Farkhondeh T. Inhaled lead exposure affects tracheal responsiveness and lung inflammation in guinea pigs during sensitization. *Biol Trace Elem Res*. 2013;154(3):363-71.
- [183] Kasten-Jolly J, Lawrence DA. Lead modulation of macrophages causes multiorgan detrimental health effects. *J Biochem Mol Toxicol*. 2014;28(8):355-72.
- [184] Antonini JM, Zeidler-Erdely PC, Young SH, Roberts JR, Erdely A. Systemic immune cell response in rats after pulmonary exposure to manganese-containing particles collected from welding aerosols. *J Immunotoxicol*. 2012;9(2):184-92.
- [185] Qian Q, Li P, Wang T, Zhang J, Yu S, Chen T, et al. Alteration of Th1/Th2/Th17 cytokine profile and humoral immune responses associated with chromate exposure. *Occup Environ Med*. 2013;70(10):697-702.

- [186] Mitra S, Keswani T, Dey M, Bhattacharya S, Sarkar S, Goswami S, et al. Copper-induced immunotoxicity involves cell cycle arrest and cell death in the spleen and thymus. *Toxicology*. 2012;293(1-3):78-88.
- [187] Ser PH, Banu B, Jebunnesa F, Fatema K, Rosy N, Yasmin R, et al. Arsenic exposure increases maternal but not cord serum immunoglobulin G level in Bangladesh. *Pediatr Int*. 2014.
- [188] Veras MM, Caldini EG, Dolhnikoff M, Saldiva PH. Air pollution and effects on reproductive-system functions globally with particular emphasis on the Brazilian population. *J Toxicol Environ Health B Crit Rev*. 2010;13(1):1-15.
- [189] Xu X, Hu H, Ha S, Roth J. Ambient air pollution and hypertensive disorder of pregnancy. *J Epidemiol Community Health*. 2014;68(1):13-20.
- [190] Mendola P, Messer LC, Rappazzo K. Science linking environmental contaminant exposures with fertility and reproductive health impacts in the adult female. *Fertil Steril*. 2008;89(2 Suppl):e81-94.
- [191] Gerhard I, Monga B, Waldbrenner A, Runnebaum B. Heavy metals and fertility. *J Toxicol Environ Health A*. 1998;54(8):593-611.
- [192] Baranski B, Sitarek K. Effect of oral and inhalation exposure to cadmium on the oestrous cycle in rats. *Toxicol Lett*. 1987;36(3):267-73.
- [193] Blum JL, Xiong JQ, Hoffman C, Zelikoff JT. Cadmium associated with inhaled cadmium oxide nanoparticles impacts fetal and neonatal development and growth. *Toxicol Sci*. 2012;126(2):478-86.
- [194] Baranski B. Effect of exposure of pregnant rats to cadmium on prenatal and postnatal development of the young. *J Hyg Epidemiol Microbiol Immunol*. 1984;29(3):253-62.
- [195] Sikorski R, Juskiewicz T, Paszkowski T, Szprengier-Juskiewicz T. Women in dental surgeries: reproductive hazards in occupational exposure to metallic mercury. *Int Arch Occup Environ Health*. 1987;59(6):551-7.
- [196] De Rosis F, Anastasio SP, Selvaggi L, Beltrame A, Moriani G. Female reproductive health in two lamp factories: effects of exposure to inorganic mercury vapour and stress factors. *Br J Ind Med*. 1985;42(7):488-94.
- [197] Rowland AS, Baird DD, Weinberg CR, Shore DL, Shy CM, Wilcox AJ. The effect of occupational exposure to mercury vapour on the fertility of female dental assistants. *Occup Environ Med*. 1994;51(1):28-34.
- [198] Davis BJ, Price HC, O'Connor RW, Fernando R, Rowland AS, Morgan DL. Mercury vapor and female reproductive toxicity. *Toxicol Sci*. 2001;59(2):291-6.
- [199] Nordstrom S, Beckman L, Nordenson I. Occupational and environmental risks in and around a smelter in northern Sweden. V. Spontaneous abortion among female employees and decreased birth weight in their offspring. *Hereditas*. 1979;90(2):291-6.

- [200] Borja-Aburto VH, Hertz-Picciotto I, Rojas Lopez M, Farias P, Rios C, Blanco J. Blood lead levels measured prospectively and risk of spontaneous abortion. *Am J Epidemiol*. 1999;150(6):590-7.
- [201] Vige M, Yokoyama K, Ramezanzadeh F, Dahaghin M, Sakai T, Morita Y, et al. Lead and other trace metals in preeclampsia: a case-control study in Tehran, Iran. *Environ Res*. 2006;100(2):268-75.
- [202] Lown BA, Morganti JB, D'Agostino R, Stineman CH, Massaro EJ. Effects on the post-natal development of the mouse of preconception, postconception and/or suckling exposure to manganese via maternal inhalation exposure to MnO<sub>2</sub> dust. *Neurotoxicology*. 1984;5(1):119-29.
- [203] Yoon M, Nong A, Clewell HJ, 3rd, Taylor MD, Dorman DC, Andersen ME. Lactational transfer of manganese in rats: predicting manganese tissue concentration in the dam and pups from inhalation exposure with a pharmacokinetic model. *Toxicol Sci*. 2009;112(1):23-43.
- [204] Banu SK, Stanley JA, Lee J, Stephen SD, Arosh JA, Hoyer PB, et al. Hexavalent chromium-induced apoptosis of granulosa cells involves selective sub-cellular translocation of Bcl-2 members, ERK1/2 and p53. *Toxicol Appl Pharmacol*. 2011;251(3):253-66.
- [205] Chashschin VP, Artunina GP, Norseth T. Congenital defects, abortion and other health effects in nickel refinery workers. *Sci Total Environ*. 1994;148(2-3):287-91.
- [206] Silva N, Senanayake H, Waduge V. Elevated levels of whole blood nickel in a group of Sri Lankan women with endometriosis: a case control study. *BMC Res Notes*. 2013;6:13.
- [207] Meléndez-García N. Efecto del vanadio en la morfofisiología del útero de ratones CD.. México: Universidad Nacional Autónoma de México. ; 2014.
- [208] García-Ibarra F. Efecto del ácido ascórbico sobre las alteraciones morfofisiológicas del ovario inducidas por la inhalación de vanadio en un modelo murino. México: Universidad Nacional Autónoma de México.; 2014.
- [209] Bucher JR, Elwell MR, Thompson MB, Chou BJ, Renne R, Ragan HA. Inhalation toxicity studies of cobalt sulfate in F344/N rats and B6C3F1 mice. *Fundam Appl Toxicol*. 1990;15(2):357-72.
- [210] Bizarro P, Acevedo S, Nino-Cabrera G, Mussali-Galante P, Pasos F, Avila-Costa MR, et al. Ultrastructural modifications in the mitochondrion of mouse Sertoli cells after inhalation of lead, cadmium or lead-cadmium mixture. *Reproductive toxicology*. 2003;17(5):561-6.
- [211] Fortoul TI, Bizarro-Nevarés P, Acevedo-Nava S, Pinon-Zarate G, Rodríguez-Lara V, Colin-Barenque L, et al. Ultrastructural findings in murine seminiferous tubules as a consequence of subchronic vanadium pentoxide inhalation. *Reproductive toxicology*. 2007;23(4):588-92.

- [212] Mussali-Galante P, Rodriguez-Lara V, Hernandez-Tellez B, Avila-Costa MR, Colin-Barenque L, Bizarro-Nevarez P, et al. Inhaled vanadium pentoxide decrease gamma-tubulin of mouse testes at different exposure times. *Toxicol Ind Health*. 2005;21(9):215-22.
- [213] Rodriguez-Lara V, Morales-Rivero A, Rivera-Cambas AM, Fortoul TI. Vanadium inhalation induces actin changes in mice testicular cells. *Toxicology and industrial health*. 2013; DOI: 10.1177/0748233713501364.
- [214] Akinloye O, Arowojolu AO, Shittu OB, Anetor JI. Cadmium toxicity: a possible cause of male infertility in Nigeria. *Reprod Biol*. 2006;6(1):17-30.
- [215] Naha N, Manna B. Mechanism of lead induced effects on human spermatozoa after occupational exposure. *Kathmandu Univ Med J (KUMJ)*. 2007;5(1):85-94.
- [216] Bosetti C, Nieuwenhuijsen MJ, Gallus S, Cipriani S, La Vecchia C, Parazzini F. Ambient particulate matter and preterm birth or birth weight: a review of the literature. *Arch Toxicol*. 2010;84(6):447-60.
- [217] Basu R, Harris M, Sie L, Malig B, Broadwin R, Green R. Effects of fine particulate matter and its constituents on low birth weight among full-term infants in California. *Environ Res*. 2014;128:42-51.
- [218] Weldy CS, Liu Y, Liggitt HD, Chin MT. In utero exposure to diesel exhaust air pollution promotes adverse intrauterine conditions, resulting in weight gain, altered blood pressure, and increased susceptibility to heart failure in adult mice. *PLoS One*. 2014;9(2):e88582.
- [219] Dolk H, Vrijheid M. The impact of environmental pollution on congenital anomalies. *Br Med Bull*. 2003;68:25-45.
- [220] Caserta D, Graziano A, Lo Monte G, Bordi G, Moscarini M. Heavy metals and placental fetal-maternal barrier: a mini-review on the major concerns. *Eur Rev Med Pharmacol Sci*. 2013;17(16):2198-206.
- [221] Al-Gubory KH. Environmental pollutants and lifestyle factors induce oxidative stress and poor prenatal development. *Reprod Biomed Online*. 2014;29(1):17-31.
- [222] Salvatori F, Talassi CB, Salzgeber SA, Spinosa HS, Bernardi MM. Embryotoxic and long-term effects of cadmium exposure during embryogenesis in rats. *Neurotoxicol Teratol*. 2004;26(5):673-80.
- [223] Angenard G, Muczynski V, Coffigny H, Pairault C, Duquenne C, Frydman R, et al. Cadmium increases human fetal germ cell apoptosis. *Environ Health Perspect*. 2010;118(3):331-7.
- [224] Heck JE, Park AS, Qiu J, Cockburn M, Ritz B. An exploratory study of ambient air toxics exposure in pregnancy and the risk of neuroblastoma in offspring. *Environ Res*. 2013;127:1-6.

- [225] Golub MS, Macintosh MS, Baumrind N. Developmental and reproductive toxicity of inorganic arsenic: animal studies and human concerns. *J Toxicol Environ Health B Crit Rev.* 1998;1(3):199-241.
- [226] Lin YY, Hwang YH, Chen PC, Chen BY, Wen HJ, Liu JH, et al. Contribution of gestational exposure to ambient traffic air pollutants to fetal cord blood manganese. *Environ Res.* 2012;112:1-7.
- [227] Gerber GB, Leonard A, Hantson P. Carcinogenicity, mutagenicity and teratogenicity of manganese compounds. *Crit Rev Oncol Hematol.* 2002;42(1):25-34.
- [228] Weinberg ED. Can iron be teratogenic? *Biometals.* 2010;23(2):181-4.
- [229] Saini S, Nair N, Saini MR. Embryotoxic and teratogenic effects of nickel in Swiss albino mice during organogenetic period. *Biomed Res Int.* 2013;2013:701439.
- [230] Fortoul TI., Rodriguez-Lara V., Cano-Gutiérrez G., González Villalba A., Colín-Bareñque L., Santamaría A., Ustarroz-Cano M., García-Peláez I., López-Valdez N., Falcón-Rodríguez CI., Pedraza-Chaverri J., Bizarro-Nevares P., Carrillo-Mora P.. Free radicals and health effects. In: Kozyrev D. SV, editor. *Handbook of free radicals: formation, types and effects.* Hauppauge, NY, USA,: Nova Science Publishers, Inc. ; 2010. p. pp. 263-89..
- [231] Cope G, Hodgson E. Classes of toxicants: Use classes. In: Hodgson E, editor. *Modern Toxicology.* Fourth ed. New Jersey: John Wiley & Sons, Inc; 2010. p. 49-50.