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Nicotinic Acetylcholine Receptor Alterations in Autism Spectrum Disorders – Biomarkers and Therapeutic Targets

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1. Introduction

Autism Spectrum Disorders (ASD) are a set of complex neurodevelopmental disorders defined behaviorally by impaired social interaction, delayed and disordered language, repetitive or stereotypic behavior and a restricted range of interest (Fombonne, 1999). ASD affect nearly 1 in 110 children, and disproportionally affect four times as many boys as girls. Comorbid symptoms often include seizures, sleep problems, gastrointestinal disorders, and metabolic deregulation (Coury, 2010). As such, ASD are an enormous challenge for parents, medical professionals, and educators. Their treatments put a significant financial strain on healthcare systems worldwide. There is no pharmacotherapy proven effective for treating the core deficits in ASD. There is also a paucity of biomarkers for autism. Both genetic and environmental factors are thought to contribute to autism susceptibility (Courchesne, 2007; Geschwind, 2009; Südhof, 2008; Ramocki & Zoghbi, 2008), but because only some of the genetic factors have been identified unequivocally thus far (Cook & Scherer, 2008; Levitt & Campbell, 2009), finding effective treatments that target the underlying causes of ASD remains a major challenge.

Identifying endophenotypes and biomarkers for complex and heterogeneous disorders such as ASD are important not only to elucidate their etiologies, but also to identify suitable biochemical molecules and pathways to target the treatment of core deficits. In this review, we present a rationale that neuronal nicotinic acetylcholine receptor (nAChR) alterations are biomarkers for ASD and that specific nAChRs subtypes are likely to be useful therapeutic targets for the treatment of core deficits. This rationale is based on the synthesis of emerging evidence from multiple types of studies, including our own, using postmortem, genetic, functional, and molecular neurobiological methodologies from two disparate areas of research – autism spectrum disorders and nicotine dependence.

2. Neuronal nicotinic acetylcholine receptors

Neuronal nAChRs are a family of ion channels that are permeable to both monovalent (Na^+ and K^+) and divalent (Ca^{++}) cations and are formed by assembly of different combinations of subunits termed $\alpha 2$ to $\alpha 10$ and $\beta 2$ to $\beta 4$. These channels are heteropentamers with the exception of the $\alpha 7$ nAChR, which is usually a homopentamer (Lindstrom, 1996; Lindstrom, 1997; Sargent, 1993). In neurons, nAChRs regulate the release of many different neurotransmitters including acetylcholine, dopamine, γ -aminobutyric acid (GABA), glutamate, and serotonin at presynaptic sites (McGehee & Role, 1996) and mediate fast synaptic transmission at postsynaptic sites (Zhang et al., 1996; Frazier et al., 1998a; Frazier et al., 1998b). These functions have a broad range of physiological effects on reward, analgesia, anxiety, affect, locomotion, attention, mood, learning, memory, and executive function (Miwa et al., 2011). nAChRs can also modulate neurite growth (Pugh & Berg, 1994; Lipton et al., 1988) and cell survival (Pugh & Margiotta, 2000; Messi et al., 1997; Kihara et al., 1997; 1998; 2001). nAChRs have been intensely studied for many decades not only to understand their normal physiological roles, but more importantly to elucidate their pathophysiological role in mediating addiction to nicotine in tobacco, because tobacco use among smokers, in particular, results in greater than 400,000 deaths per year in the U.S. alone. In addition to their role in nicotine addiction, nAChR dysfunctions are also implicated in other disorders, including Alzheimer's disease, Parkinson's disease, schizophrenia, attention deficit-hyperactivity disorder, anxiety disorders, Tourette's syndrome, and depression (Newhouse & Kelton, 2000; Newhouse et al., 2004; Mineur & Picciotto, 2010).

3. Alterations of nAChRs in ASD

3.1 Changes in $\alpha 4\beta 2$ nAChR expression

Examination of postmortem brains of individuals with ASD has identified major nAChR abnormalities in multiple postmortem studies. In the first such study to be undertaken, postmortem tissue from 7 adults with a mean age of 24 years was examined. High-affinity ^3H epibatidine binding was reported to be significantly reduced in the frontal and parietal cortex of these individuals with ASD compared to age-matched controls. Furthermore, immunohistochemical analyses showed that the loss of ^3H epibatidine correlated with reduced expression of the $\alpha 4$ and $\beta 2$ nAChR subunits. Notably, the mRNA for these two nAChR subunits was not significantly decreased, suggesting that the reduction in nAChR subunit levels resulted from an impaired posttranslational mechanism. Also, ^3H pirenzepine binding to M1 and M2 muscarinic AChRs (mAChRs) was not significantly altered, suggesting that the loss of nAChR expression resulted from deregulation of a posttranslational mechanism that specifically affected nAChRs, but not mAChRs (Perry et al., 2001). In a subsequent study, postmortem tissue from 8 adults with a mean age of 24 years was examined. Again, high-affinity ^3H epibatidine binding was reported to be significantly reduced by greater than 50% in the cerebellar cortex of individuals with ASD. High-resolution analyses of the autoradiographic data indicated that the loss of ^3H epibatidine binding occurred in the granule cell layer, the Purkinje layer, and the molecular cell layer of the cerebellum of individuals with ASD compared to age-matched controls. Significant reduction in the expression of the $\alpha 4$ nAChR subunit, but not its mRNA (Lee et al., 2002), was also observed and is consistent with the notion that $\alpha 4\beta 2$ nAChR loss results from an impaired posttranslational mechanism regulating its expression. In a third

study, immunohistochemical analysis of postmortem brains from 3 adults with ASD of mean age 29 years surprisingly showed no changes in the expression of the $\alpha 4$ nAChR subunit in the thalamus compared to age-matched controls. However, reduction of the $\beta 2$ nAChR subunit was observed in the paraventricular nucleus and nucleus reuniens of the thalamus (Martin-Ruiz et al., 2004).

3.2 Changes in $\alpha 7$ nAChR expression

In contrast to the loss of ^3H epibatidine binding and decreased expression of the $\alpha 4$ and $\beta 2$ nAChR subunits, no significant change in the binding of ^{125}I - α -bungarotoxin to the $\alpha 7$ nAChR or immunohistological detection of the $\alpha 7$ nAChR (Perry et al., 2001) was reported in the frontal and parietal cortex. In the cerebellar cortex, however, binding of α -bungarotoxin to the $\alpha 7$ nAChR and immunohistological detection of the $\alpha 7$ nAChR did show a significant increase in the expression of the $\alpha 7$ nAChR in the granule cell layer, but not in the Purkinje cells or the molecular cell layer. Interestingly, similar to the $\beta 2$ nAChR subunit, reduction of the $\alpha 7$ nAChR subunit was also observed in the paraventricular nucleus and nucleus reuniens of the thalamus. Thus, alterations in the expression of both the $\alpha 4\beta 2$ nAChR and the $\alpha 7$ nAChR in individuals with ASD appears to show regional specificity (Perry et al., 2001; Lee et al., 2002; Martin-Ruiz et al., 2004), suggesting that these changes are compensatory and result from altered homeostasis of neural networks, rather than the direct effect of a single molecule in a particular molecular pathway.

Two recent studies on rare genomic microdeletions and copy-number variations (CNVs) revealed a possible involvement of the *CHRNA7* gene in some cases of autism. The first study investigated segmental duplications at breakpoints (BP4–BP5) of chromosome 15q13.2q13.3 from 1441 individuals with autism from 751 families in the Autism Genetic Resource Exchange (AGRE) repository (Miller et al., 2009). This genomic sequence spans over 1.5 Mb and includes *CHRNA7*. From this cohort 10 patients were identified with genomic imbalance at chromosome 15q13.2q13.3, including five with BP4–BP5 microdeletions. Among the 1420 parents and 132 unaffected/unknown siblings no cases of BP4–BP5 microdeletion were found. The second study on genomic CNVs explored the genetic contribution to ASD in a large cohort of families (Simons Simplex Collection consisting of 915 families) with a single autistic child and at least one unaffected sibling (Levy et al., 2011). The contribution of the transmission of “ultrarare” variants to ASD, in particular inherited genomic duplications was also estimated. A transmitted duplication within the *CHRNA7* gene was observed in 8 autistic children and 3 unaffected siblings within 6 families. A further network-based analysis of genetic associations (NETBAG) of that dataset strengthened the involvement of *CHRNA7* as one of the genes affected by rare de novo CNVs in autism (Gilman et al., 2011).

4. nAChRs modulate multiple behaviors deficient in ASD

ASD is defined by three behavioral deficits, impaired social interactions, repetitive behaviors, and delayed language. Multiple studies using animal models implicate a functional role for nAChRs in some of these behavioral deficits in ASD. $\beta 2$ -containing nAChRs regulate executive and social behaviors in studies using $\beta 2$ nAChR subunit knockout mice (Granon et al., 2003). Knockout $\beta 2$ nAChR mice show a decrease in slow exploratory behavior - a measure of cognitive function during which animals slowly and

precisely explore their environment, a lack of sensitization to novel stimuli, and abnormal social behavior during aggressive confrontations with other mice (Granon et al., 2003). Recovery of the slow exploratory behavior was observed by injecting a lentiviral vector expressing the $\beta 2$ nAChR subunit into the ventral tegmental area (VTA) in the knockout mice (Maskos et al., 2005). Re-expressing the $\beta 2$ nAChR subunit in the prefrontal cortex also improves social abnormalities in this knockout mouse. Increased social interaction and decreased novel exploration in a social interaction paradigm with concurrent motivation was ameliorated after stereotactically injecting the $\beta 2$ nAChR subunit into the prelimbic area of the prefrontal cortex (PFC) (Avale et al., 2011).

As previously mentioned, nAChR dysfunction is also implicated in several other neurological disorders with repetitive behavior. We suggest here that similarities in behaviors across those neurological conditions, as well as high prevalence of simultaneity suggest a possible shared underlying mechanism. Moreover, there has been a recent push to redefine repetitive behavior in these neuropsychiatric disorders and instead characterize stereotypies into disorder-related endophenotypes rather than separate disorder-specific symptoms (Kas et al., 2007, Langen et al., 2011). Tourette's syndrome (TS), obsessive compulsive disorder (OCD), and attention deficit hyperactivity disorder (ADHD), all involve disordered cortical-basal ganglia circuitry and all can be successfully treated with drugs acting on nAChRs. The basal ganglia and orbitofrontal cortex, both regions highly innervated by nicotinic acetylcholine receptor rich interneurons are hyperactive during PET/SPECT studies of OCD (Baxter et al., 1988) and hypoactive in studies of ADHD (Zametkin et al., 1990) and TS (Braun et al., 1995). The orbitofrontal cortex controls inhibition and disinhibition of behavior, and lesions in this area are sufficient to cause impulsive and inappropriate behavior. Nicotine or an analog alone has demonstrated potential to treat repetitive behaviors in these disorders. A transdermal nicotine patch, administered as therapy for TS, decreases the severity and frequency of tics, a compulsory symptom of TS (Sanberg, 1997). Nicotine gum administered to OCD patients previously resistant to other treatment clinically improved behavior (Carlsson, 2001; Pasquini et al., 2005). Interestingly, clomipramine, an SSRI commonly prescribed for the treatment of OCD, also acts on nAChRs (Lopez-Valdes, 2002). Lastly, (-)-Nicotine and ABT-418, an $\alpha 4\beta 2$ nAChR agonist (Potter et al., 1999), both successfully treat adults with ADHD (Levin and Simon, 1998; Wilens et al., 1999). It is interesting to note that hyperactivity, tics, and obsessive compulsive disorder are all common comorbid disorders seen in patients with ASD with approximately 59% of ASD patients having impulsivity problems, 8-10% having tics, and 37% having OCD (Levy et al., 2009). Although it is clear that similar neurocircuitry is involved in several disorders with repetitive behavior, further research is needed to determine whether the underlying mechanisms causing this dysfunction overlap in TS, OCD, ADHD, and in ASD.

nAChRs also are involved in several other non-core, but frequently occurring symptoms in ASD. The most common comorbid disorders and symptoms associated with ASD are psychiatric (e.g., depression and anxiety), neurological (e.g., epilepsy), sleep, and sensory (e.g., tactile) disorders. Epilepsy occurs in 5-49% of people with autism (Levy et al., 2009). Genetic abnormalities in CHRN4A and CHRNB2, encoding the α and β nAChR subunits respectively, are sufficient to cause autosomal dominant nocturnal frontal lobe epilepsy (ADNFLE) (De Fusco, 2000; Bertrand, 2002; Steinlein, 2002; Hoda, 2009), however ADNFLE is not associated with ASD. 52-73% of patients with ASD experience sleep disruption and 43-84% experience anxiety disorders. Knocking out the $\alpha 4$ nAChR subunit increases anxiety

in mice (Ross et al., 2000) and the $\beta 2$ nAChR knockout animal shows abnormal sleep pattern (Lena et al., 2004). These studies demonstrate that behaviors regulated by nAChRs are disparate and commonly aberrant in ASD and suggest the potential for nAChR-acting drugs in the treatment of ASD.

Lastly, there is accumulating evidence that the immune system is disrupted in individuals with ASD (Careaga et al., 2010). Elevated levels of chemokines have been detected in the brains and cerebrospinal fluid (Chez et al., 2007; Wills et al., 2009) as well as the plasma (Ashwood et al., 2011a) of individuals with ASD, and this elevation correlated with more impaired behavior (Ashwood et al., 2011b). Furthermore, postmortem studies of individuals with autism also detected presence of activated neuroglial cells in their brain (Vargas et al., 2005). In a recent study, activated microglia were detected in the dorsolateral PFC in 5 out of 13 samples, 2 of which were under the ages of 6 years (Morgan et al., 2010). These results suggest that inflammation of the central nervous system, at least in some individuals, may contribute to the neuropathology of ASD. Thus, suppression of neuroinflammation by targeting $\alpha 7$ nAChRs in ASD may be potentially beneficial.

5. Neurexin and neuroligin deficits in ASD

The neurexins are cell adhesion molecules encoded by three genes corresponding to neurexins 1, 2 and 3 (Missler & Südhof, 1998; Lise & El-Husseini, 2006). As a result of transcriptional initiation from two different promoters, each neurexin gene encodes a longer α -neurexin protein and a shorter β -neurexin protein. The proteins are identical from their intracellular C-termini through their transmembrane domains, glycosylation-rich domains and the sixth LNS domain of α -neurexin, which corresponds to the only LNS domain in β -neurexin. They have divergent N-terminal extracellular domains, which allow for interactions with multiple proteins. Additionally, alternative splicing at multiple splice sites within each gene can give rise to more than one thousand different isoforms, which differ only in their extracellular domains. Neurexins recruit N- and P/Q-type calcium channels to active zones of presynaptic terminals through scaffolding proteins, including calmodulin-associated serine/threonine kinase (CASK) (Hata et al., 1996; Missler et al., 2003; Zhang et al., 2005). α -neurexins were reported to specifically induce GABAergic postsynaptic differentiation (Kang et al., 2008). The enormous structural diversity of the neurexins suggests that they are involved in a multitude of physiological functions yet to be elucidated.

Results from a linkage and copy number variation analysis conducted by the Autism Genome Project Consortium (Szatmari et al., 2007) show that neurexin-1 dysfunction is associated with ASD. This conclusion has been corroborated in multiple linkage analysis studies since (Kim et al., 2008; Marshall et al., 2008) and in analysis of structural variants in the α - and β -neurexin genes (Zahir et al., 2008; Feng et al., 2006; Yan et al., 2008; Gai et al., 2011; Gauthier et al., 2011). Neurexin knock-out animals have provided insights into the functions of the neurexin family. Neurexin 1/2/3 α - triple knock-out animals die perinatally and have reduced spontaneous and evoked neurotransmission at glutamatergic and GABAergic synapses, demonstrating that α -neurexins are necessary for neurotransmitter release at synapses (Missler et al., 2003). Additionally, mice lacking neurexins have impaired neuroendocrine secretion (Dudanov et al., 2006), which may mirror some children with autism that exhibit dysfunction of the hypothalamic-pituitary-adrenocortical system,

possibly due to altered neuroendocrine regulation (Corbett et al., 2006). Similar to the neurexin triple knockout animals, mice lacking neurexin1/2 α or neurexin2/3 α die within 1 month after birth and have reduced neurotransmission. Analyses of brain morphology in α -neurexin knockouts revealed no major impairments in synapse formation, but minor reductions in dendrite branch length and spine numbers were detected, suggesting they are important in synapse maturation more so than formation (Dudanova et al., 2007). None of the single α -neurexin knock-out animals have dramatic phenotypes, with the neurexin-2 α knock-out animals showing the least severe phenotype (Craig & Kang, 2007). In the absence of neurexin-1 α , miniature excitatory postsynaptic currents were reduced in recordings from hippocampal slices. Behaviorally, the neurexin-1 α -deficient mice were identical to wild-type mice in multiple social interactions, but displayed decreased grooming behavior, impaired nest building, decreased pre-pulse inhibition, and improved motor learning in behavioral studies (Etherton et al., 2009). While the neurexin-1 α -deficient mice display behavioral phenotypes similar to what is seen in autism they are not sufficient to explain ASD yet they still provide a useful but limited model of ASD. The β -neurexin and combined α - and β -neurexin knockout animals have not yet been fully evaluated.

The neuroligins are encoded by five differentially spliced genes that encode multiple neuroligin isoforms (Zhang et al., 2005; Boucard et al., 2005). In complementary roles, neuroligins, the postsynaptic binding partners of neurexins, recruit N-methyl-D-aspartate (NMDA) receptors and GABA_A receptors through their interactions with scaffolding proteins such as post-synaptic density 95 (PSD-95) and gephyrin, respectively (Graf et al., 2004; Nam & Chen, 2005; Chih et al., 2006; Pouloupoulos et al., 2009). Thus, bi-directional interactions between neurexins and neuroligins appear to serve a critical function in the assembly and maturation of both glutamatergic and GABAergic synapses through recruitment of the requisite presynaptic and postsynaptic components of neurons (Dean & Dreshbach, 2006; Craig & Kang, 2007; Sudhof, 2008).

Neuroligins are strongly implicated in ASD. Chromosomal rearrangements and copy number variations in neuroligin-1 are linked to autism (Konstantareas & Homatidis, 1999; Ylisaukko-oja et al., 2005; Glessner et al., 2009). There is also evidence that mutations in neuroligin-3 and neuroligin-4 are found in patients with ASD (Laumonnier et al., 2004; Jamain et al., 2003). In addition mouse models support a role for neuroligins in ASD. Neuroligin-1 knock-out mice are viable and fertile, but also have synaptic dysfunctions (Chubykin et al., 2007). At the molecular level, the NMDA/AMPA ratio at corticostriatal synapses is reduced, which is associated with repetitive grooming that may mirror some of the repetitive behaviors seen in autistic patients (Blundell et al., 2010). In contrast to neuroligin-1-deficient mice, which show impaired NMDA receptor signaling, neuroligin-2 knock-out animals have deficits in inhibitory synaptic transmission (Chubykin et al., 2007). Behaviorally, neuroligin-2 knock-out mice exhibit increased anxiety, but normal social interactions (Blundell et al., 2009), similar to the neurexin-1 α -deficient mice. Mutations in neuroligin-3 and neuroligin-4 lead to intracellular retention of the mutant proteins (Chih et al., 2004; Comoletti et al., 2004). The neuroligin-3 R451C mutation is a gain of function mutation. Mice with this point mutation exhibited impaired social interactions and increased inhibitory synaptic transmission (Tabuchi et al., 2007). Mice lacking neuroligin-4 correspond to loss-of-function mutations in human neuroligin-4 and show deficits in reciprocal social interactions and ultrasonic communication (Jamain et al., 2008). Neuroligin 1/2/3 α triple knock-out animals die at birth, but similar to their α -neurexin-deficient

counterparts, do not show dramatic reductions in synapse numbers or brain architecture, but do have severely impaired synaptic transmission (Varoqueaux et al., 2006).

The studies of the neurexin and neuroligin functions indicate a role for them in proper synaptic function but not synapse formation. Although it is clear that the deficits of neurexins and neuroligins play a role in ASD, understanding their interactions with receptors will provide additional insight into their functions.

6. Neurexins associate with multiple receptors, including nAChRs

Accumulating evidence indicates that neurexins interact directly with more than the neuroligins. Our laboratory was the first to provide experimental evidence for direct interactions between neurexins and receptors by showing that neurexin-1 β coimmunoprecipitates with recombinant $\alpha 4\beta 2$ nAChRs when expressed in heterologous cells (Cheng et al., 2009). Functionally, the neurexin-1 β regulates targeting of $\alpha 4\beta 2$ nAChRs to pre-synaptic terminals in neurons (Cheng et al., 2009). Complementary studies report a role for neurexin-1 and neuroligin-1 in recruitment of $\alpha 3$ -containing nAChRs to the post-synaptic density (Conroy et al., 2007; Ross & Conroy, 2008). In addition, recent studies show that neurexins interact with multiple receptors. First, neurexin-1 β interacts with GABA_A receptors; this interaction modulates the cell surface expression levels of the GABA_A receptors but not its functions *per se* (Zhang et al., 2010). Second, leucine-rich repeat transmembrane protein (LRRTM2) binds trans-synaptically to both neurexin-1 α and -1 β and induces presynaptic differentiation at excitatory synapses (Ko et al., 2009; de Wit et al., 2009; Siddiqui et al., 2010). Knock-down of LRRTM2 in the rat dentate gyrus shows a large reduction in AMPAR-mediated EPSCs in *in vivo* recordings from granule cells in hippocampal slices. Furthermore, the association between neurexin-1 and LRRTM2 is a functional interaction. When neurexin-1 is knocked-down in hippocampal neurons, LRRTM2 is unable to induce presynaptic differentiation (de Wit et al., 2009). Finally, neurexins associate with GluR $\delta 2$ receptors via a soluble protein called cerebellin -1 precursor protein (Cbln1) (Uemura et al., 2010). In the Cbln1 knockout mice, the synaptogenic activity of GluR $\delta 2$ receptor is lost. Thus, GluR $\delta 2$ mediates cerebellar synapse formation by interacting with presynaptic neurexins via Cbln1.

7. Genetic variants of neurexin-1 are linked to nicotine dependence

A recent high-density genome-wide association study for nicotine dependence linked single nucleotide polymorphisms (SNP) in the neurexin-1 gene to the development of nicotine dependence and thus smoking behavior (Bierut et al., 2007). A second independent study also showed linkage between a variant of the neurexin-1 gene and nicotine dependence in smokers of European and African-American ancestry (Nussbaum et al., 2008). These results, along with the fact that neurexins functionally target $\alpha 4\beta 2$ nAChRs to synapses, implicate neurexins in the etiology of other neurological diseases typically associated with pathophysiological functions of nAChRs. $\alpha 4\beta 2$ nAChRs mediate essential features of nicotine addiction including reward, tolerance, and sensitization (Tapper et al., 2004). Thus, functions are likely to be affected by changes in the expression levels of neurexin-1. The exact mechanism by which neurexin-1 α and -1 β splicing is regulated to generate the predicted hundreds of neurexin-1 isoforms remains to be elucidated. It is possible that a regulatory SNP in the intron of the neurexin-1 gene could modulate neurexin-1 expression

or splicing efficiency and thus influence nAChR functions by regulating their synaptic targeting efficiency. Because there are hundreds of neurexin-1 α isoforms, the linkage between neurexin-1 gene variants, $\alpha 4\beta 2$ nAChR synaptic targeting, and nicotine dependence requires additional studies. Nevertheless, the functional linkage between neurexin-1 and $\alpha 4\beta 2$ nAChR and their converging roles in nicotine dependence suggests that $\alpha 4\beta 2$ nAChR activity may regulate neurexin-1 gene expression.

8. nAChR modulate excitation-inhibition balance

There is strong evidence that some forms of ASD are caused by an imbalance of excitatory and inhibitory synaptic transmission in neuronal circuits that are responsible for the establishment of language processing and social behavior during prenatal and postnatal brain development. Increased glutamatergic (excitatory) signaling or suppressed GABAergic (inhibitory) signaling is sufficient to disrupt the excitatory/inhibitory balance in local circuit-plasticity (Rubenstein & Merzenich, 2003). A hyperexcitable cortex is poorly differentiated functionally and therefore inherently unstable and susceptible to epilepsy. This might explain why, in addition to the autistic core symptoms, an average of ~30% of individuals with ASD develop clinically apparent seizures (Gillberg & Billstedt, 2000). In several mouse models of autism this lack of homeostasis of excitatory and inhibitory signaling was observed (Tabuchi et al., 2007; Gogolla et al., 2009). In the frontal cortex, cholinergic transmission can modulate cortical tone establishing a homeostasis of excitatory and inhibitory signals (Aracri et al., 2010). In layer V of the prefrontal cortex, nAChR activation increases the threshold for activating glutamatergic synapses (Couey et al., 2007), whereas GABA release is stimulated in several cortical layers by nAChR activation (Alkondon et al., 2000).

We posit that some of the regulatory effects of balancing inhibitory and excitatory synaptic transmission are mediated by synaptic targeting of nAChRs by neurexins. This results in the change of expression levels of nAChRs in various brain regions of autistic individuals. Therefore allosteric modulators or direct agonists targeting nAChRs by might be useful to restore the imbalance of excitatory and inhibitory synaptic transmission caused by deregulated expression of neurexin-1.

9. Nicotinic receptors as biomarkers for ASD

9.1 Positron Emission Tomography ligands for $\alpha 4\beta 2$ nAChRs

The alterations in nAChRs in ASD may also serve as an early molecular biomarker, detectable by imaging tools such as positron emission tomography (PET), the most advanced modality for non-invasive study of receptors. Monitoring the reversal of the loss of $\alpha 4\beta 2$ nAChR in the frontal, parietal, and cerebellar cortex and the upregulation of $\alpha 7$ nAChR in the cerebellar cortex by PET imaging in the brains of individuals with ASD might provide a clinical tool to complement behavioral tests needed to assess the effectiveness of novel pharmacotherapies for autism.

Three radiotracers, [^{11}C]nicotine, (S)-3- (azetidin-2-ylmethoxy)-2-[^{18}F]fluoropyridine (2-[^{18}F]FA) and (S)-5- (azetidin-2-ylmethoxy)-2-[^{18}F]fluoropyridine (6-[^{18}F]FA), have been used for studying $\alpha 4\beta 2$ nAChRs in the human brain using PET. The PET imaging properties of these radioligands are not perfect however. Poor signal-to-noise ratios and other drawbacks of [^{11}C]nicotine suggest that this radiotracer is not well suited for quantitative imaging in

animals and humans. 2-[¹⁸F]FA is the only currently available radioligand for quantitative imaging nAChR in humans. The “slow” brain kinetics of 2-[¹⁸F]FA and 6-[¹⁸F]FA hamper mathematical modeling and reliable kinetic parameter estimation since it takes many hours of PET scanning (5–7 h) for the tracer radioactivity to reach a spatial-temporal steady state (Horti et al., 2010). Another crucial problem with 2-[¹⁸F]FA and 6-[¹⁸F]FA is relatively low binding potential (BP) in extrathalamic regions (BP ≤ 0.6–0.8), including the cortex, which has a lower nAChR density. Altered densities of cortical and striatal nAChRs in neurodegenerative diseases (Pimlott et al., 2004) and schizophrenia (Ochoa & Lasalde-Dominicci, 2007) illustrates the importance of imaging extrathalamic nAChRs.

A variety of radioligands with fast regional brain kinetics have been presented in non-human primates and pigs. Analogs of epibatidine showed “rapid” brain kinetics and improved BP (Gao et al., 2007, 2008). One compound of the series, (-)-2-(6-[¹⁸F]fluoro-2,3'-bipyridin-5'-yl)-7-methyl-7-aza-bicyclo[2.2.1]heptane ([¹⁸F]JHU87522 or [¹⁸F]AZAN) exhibited better imaging properties in animal studies than those of 2-[¹⁸F]FA and 6-[¹⁸F]FA including a greater BP value and faster brain kinetics. In addition, the brain uptake of [¹⁸F]AZAN is greater and its acute toxicity is lower. Most available PET and single photon emission computed tomography (SPECT) imaging agents for nAChR are agonists and these nAChR-agonists are toxic when injected at high doses. Unlike 2-FA that is nAChR agonist, AZAN displays properties of functional antagonist of $\alpha 4\beta 2$ nAChR. Currently, AZAN is undergoing toxicological studies that will determine if this radioligand is sufficiently safe for clinical application as a PET radiotracer. If [¹⁸F]AZAN is safe for human PET studies, there are strong indications that it could become the radiotracer of choice for PET imaging of nAChR in human brain (Horti et al., 2010).

9.2 Positron Emission Tomography ligands for $\alpha 7$ nAChRs

Several radiotracers were developed for selective imaging of the $\alpha 7$ nAChRs in the human brain for PET and SPECT (Dolle et al., 2001; Pomper et al., 2005; Ogawa et al., 2006). Despite these efforts, there have been no clinical studies using these radioligands for $\alpha 7$ nAChRs in the human brain.

Very recently, 4-[¹¹C]methylphenyl 2,5-diazabicyclo[3.2.2]nonane-2-carboxylate ([¹¹C]CHIBA-1001) was developed as a novel PET ligand for $\alpha 7$ nAChRs in the conscious monkey brain. An *in vitro* binding study showed that the IC₅₀ value of CHIBA-1001 for [¹²⁵I] α -bungarotoxin binding to the rat brain homogenates was 45.8 nM. [¹¹C]CHIBA-1001 distribution in the monkey brain measured by PET was consistent with the regional distribution of $\alpha 7$ nAChRs. Moreover, brain uptake of [¹¹C]CHIBA-1001 was dose-dependently blocked by pretreatment with the selective $\alpha 7$ nAChR agonist SSR180711, but was not altered by the selective $\alpha 4\beta 2$ nAChR agonist A-85380 (Hashimoto et al., 2008).

In the human brain, [¹¹C]CHIBA-1001 was found widely distributed in all brain regions. The regional distribution pattern of [¹¹C]CHIBA-1001 is consistent with what is expected *in vitro* (Falk et al., 2003; Court et al., 1999; 2001; Marutle et al., 2001), but different from that of $\alpha 4\beta 2$ nAChRs (Clementi, 2004). However, it is slightly different from the regional distribution in the monkey brain (Hashimoto et al., 2008). In the human brain, remarkable radioactivity accumulation was observed in the cerebellum. These findings suggest that [¹¹C]CHIBA-1001 is a suitable radioligand for imaging $\alpha 7$ nAChRs in the human brain, as it offers acceptable dosimetry and pharmacological safety at the dose required for adequate PET imaging (Toyohara et al., 2009).

These recent advances in the development of new nAChR PET radioligands, like [^{18}F]AZAN for $\alpha 4\beta 2$ nAChRs and [^{11}C]CHIBA-1001 for $\alpha 7$ nAChRs with fast kinetics and low toxicity will provide promising tools for monitoring alterations of brain nAChR especially in young patients with ASD. The principal downside to the use of PET is the unknown risk of using radioactive ligands and sedatives, especially in younger individuals, to perform PET scans.

10. Nicotinic drugs as therapeutic agents for ASD

10.1 Agonists

10.1.1 $\alpha 4\beta 2$ nAChRs

The extensive loss of $\alpha 4\beta 2$ nAChRs in some individuals with ASD provide a rationale for exploratory trials of drugs that can upregulate and activate $\alpha 4\beta 2$ nAChRs and thus compensate for their loss both physically and functionally. The panoply of drugs developed over the last few decades for smoking cessation therapy as well as other disorders with pathophysiological roles for nAChRs (Taly et al., 2009), offers a large selection of drugs that are likely to be specific for $\alpha 4\beta 2$ nAChRs and capable of upregulating them. Varenicline (Chantix), one such drug that has FDA approval for use in smoking cessation therapy is a partial agonist of the $\alpha 4\beta 2$ nAChRs (Coe et al.; 2005) and of interest for treatment of ASD. Although varenicline is also a full agonist of the $\alpha 7$ AChR (Mihalak et al., 2006), its relative specificity for $\alpha 4\beta 2$ nAChRs is thought to be due to differences in its EC_{50} for activation of $\alpha 4\beta 2$ nAChRs versus $\alpha 7$ nAChRs, as well as a function of the low concentrations at which it is used clinically for anti-smoking therapy (Niaura et al., 2006). Thus it has become one of the most widely used smoking cessation drugs with millions of users worldwide and shows little sympathetic and parasympathetic complications from cross activation of ganglionic nAChRs ($\alpha 3\beta 4$ nAChRs). Interestingly, much like nicotine, varenicline can upregulate $\alpha 4\beta 2$ nAChRs *in vitro*. Finally, as a partial agonist, it has the additional benefit of providing chronic low-level activation of $\alpha 4\beta 2$ nAChRs (Papke et al., 2011) and possibly associated downstream intracellular signaling pathways. Varenicline has been shown to change behaviors in some smokers, and a public health advisory from the FDA includes warnings of increased suicidal thoughts and actions. It is important to note, however, that the increase in suicidal thoughts and actions may occur in only a subpopulation of individuals taking varenicline as there is ample evidence that smoking may be more prevalent in those individuals with comorbid neuropsychiatric conditions, including schizophrenia (Adler et al., 1993; Dalack et al., 1999). This may explain behavioral changes reported among smokers using varenicline if individuals have subclinical neuropsychiatric conditions. This idea has been supported by a recent study reporting that there was no clear evidence that varenicline use in itself was associated with an increased risk for depression or suicidal thoughts (Gunnell et al., 2010). Also, unlike in schizophrenia, the prevalence of smoking in individuals with ASD is low (Bejerot & Nylander, 2003), possibly because the loss of $\alpha 4\beta 2$ nAChRs occurs early in development – a clinical feature further strengthening the utility of using $\alpha 4\beta 2$ nAChRs loss as a biomarker for ASD. Nevertheless, any clinical trial of varenicline for individuals with ASD should require close monitoring of possible suicidal ideation given the heterogeneity of causes expected for ASD, some of which may overlap with schizophrenia (Kirov et al., 2009).

10.1.2 $\alpha 7$ nAChRs

It is possible to use $\alpha 7$ nAChR agonists to treat neuroinflammation in ASD. There is strong evidence that activation of the $\alpha 7$ nAChR expressed on monocytes and macrophage, by inhibiting NF-kappaB nuclear translocation, suppresses cytokine release by them (Wang et al., 2003), and that this cholinergic anti-inflammatory pathway that provides a bidirectional link between the nervous and immune system, inhibits the innate immune response (Rosas-Ballina & Tracey, 2009). Hence, a reasonable case can be made for the use of $\alpha 7$ nAChR agonists to treat neuroinflammation in ASD. Individuals could be stratified by monitoring brain inflammation by the uptake of the microglial marker, [^{11}C]PK11195, a PET ligand useful for detecting peripheral benzodiazepine receptors expressed in high amounts in activated microglia (Rojas et al., 2007). However, given that $\alpha 7$ AChR appears to be pathologically upregulated in cerebellum of some individuals with ASD, caution is advocated in the use of $\alpha 7$ AChR agonists to treat ASD. The primary challenge is that the net behavioral benefit from suppressing neuroinflammation mediated by microglia versus over stimulating upregulated $\alpha 7$ AChRs in the granule cell layer, cannot be predicted *a priori*. Two different $\alpha 7$ nAChR agonists have been used to treat schizophrenia; drugs that might be repurposed for use in individuals with ASD and detectable neuroinflammation.

One of these drugs, GTS-21, or 3-(2,4-dimethoxybenzylidene)-anabaseine (DMXB-A) is a partial agonist of $\alpha 7$ nAChRs may have beneficial effects in ASD patients. In healthy control subjects, DMXB-A improves attention, working memory, and episodic memory (Kitagawa et al., 2003). The default network, which has been widely reported to be abnormal in schizophrenia (Garrity et al., 2007), is a functionally connected network of brain regions that includes the posterior cingulate cortex, cuneus/precuneus, medial prefrontal cortex, medial temporal lobe, and inferior parietal cortices (Buckner et al., 2008; Tregellas et al., 2011). Altered default network activity has been shown to be a result of DMXB-A administration to patients with schizophrenia (Tregellas et al., 2011), with decreased expression of $\alpha 7$ nAChRs (Freedman et al., 1995).

A second candidate drug, Tropisetron is a partial agonist of the $\alpha 7$ nAChR. Auditory sensory gating P50 deficits are correlated with neuropsychological deficits in attention, one of the principal cognitive disturbances in schizophrenia. In a clinical trial with 33 schizophrenic patients administration of tropisetron, without placebo, significantly improved auditory sensory gating P50 deficits in non-smoking patients with schizophrenia (Shiina et al., 2010). In mice, the early postnatal period represents a critical time window essential for brain development. The administration of tropisetron from postnatal days 2-12 (P2-P12) in mice did not induce significant cognitive, schizophrenia-like or emotional alterations in tropisetron-treated animals as compared to controls, when tested in multiple behavioral assays (Inta et al., 2011).

10.2 Positive allosteric modulators

Galantamine is an acetylcholinesterase inhibitor that also acts as a positive allosteric modulator at the $\alpha 4\beta 2$ and $\alpha 7$ nAChRs (Dajas-Bailador et al., 2003; Samochocki et al., 2003; Schilström et al., 2007). In two studies with small numbers of subjects it has been reported that galantamine showed potential benefits for attention, memory, and psychomotor speed in schizophrenia (Schubert et al., 2006; Lee et al., 2007). An unpublished study from Johnson and Johnson failed to find an advantage for galantamine on a measure of global cognition

(clinicaltrials.gov, trial number: NCT 00077727). In a 12-week open-label trial of galantamine, thirteen children with autism, previously unmedicated, (mean age, 8.8 +/- 3.5 years) showed a significant reduction in parent-rated irritability and social withdrawal on the Aberrant Behavior Checklist (ABC), as well as significant improvements in emotional lability and inattention on the Conners' Parent Rating Scale—Revised (Nicolson et al., 2006). Similarly, clinical ratings showed reductions in the anger subscale of the Children's Psychiatric Rating Scale. Eight of 13 participants were rated as responders on the basis of their improvement scores on the Clinical Global Impressions scale. The allosteric properties of galantamine could directly lead to increased release of acetylcholine and activation of postsynaptic nAChRs (Samochocki et al., 2003) or act indirectly through its effects on the release of other neurotransmitters, especially glutamate and dopamine (Schilström et al., 2007; Wang et al., 2007).

It has been demonstrated that amyloid- β precursor protein (APP) is upregulated in a mouse model for Fragile X mental retardation (FXS) (Westmark et al., 2008) and two clinical studies have reported higher levels of APP in children with autism. In the first study, affected children expressed sAPP at 2 or more times the levels of children without autism and up to 4 times more than children with mild autism (Sokol et al., 2006). In the second study, elevated plasma sAPP α was found in 60% of known autistic children ($n = 25$) compared to healthy age-matched controls (Bailey et al., 2008). Recent studies showed that galantamine allosterically modulates microglial nAChRs and increases microglial beta-amyloid ($A\beta$) phagocytosis (Wang et al., 2007; Takata et al., 2010).

Collectively, these studies suggest that positive allosteric modulators of $\alpha 4\beta 2$ nAChRs, when used by themselves or in conjunction with agonists, may be beneficial in correcting deficits in the functions of $\alpha 4\beta 2$ nAChRs and thereby core deficits of ASD.

11. Conclusions

This review presents a reasonable rationale based on synthesis of the literature that nAChRs are suitable biomarkers as well as therapeutic targets for addressing core deficits in ASD. Multiple lines of evidence show that nAChRs can modulate many of the functions deficient in individuals with ASD. Furthermore, neuropathological findings, albeit small in numbers, show significant alterations in both $\alpha 4\beta 2$ nAChRs and $\alpha 7$ nAChRs. In the cerebellum, an anatomical area contributing significantly to the etiology of ASD, $\alpha 4\beta 2$ nAChRs are deficient, and $\alpha 7$ nAChRs are upregulated. These findings suggest that well developed PET ligands for both these nAChR subtypes can be used to monitor changes in their expression in response to treatment, behavioral or pharmacological. A novel functional linkage between neurexin-1 and $\alpha 4\beta 2$ nAChR and their converging roles in nicotine dependence suggests that $\alpha 4\beta 2$ nAChR activity may regulate neurexin-1 gene expression. Additionally, agonists and positive allosteric modulators of the $\alpha 4\beta 2$ AChRs are likely to be therapeutic agents that can help restore $\alpha 4\beta 2$ nAChRs expression levels in the brains of individuals with ASD, based on known effects of these agents. A case can be made for the use of $\alpha 7$ nAChRs to reduce neuroinflammation in the brain in those ASD individuals with such clinical pathology. The ultimate hope is that these agents, when administered early in development, by their presumed ability to modulate a number of different neurotransmitter systems and associated signaling pathways, could help correct core deficits associated with ASD.

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13. References

- Adler, L. E., Hoffer, L. D., Wiser, A. & Freedman, R. (1993). Normalization of auditory physiology by cigarette smoking in schizophrenic patients. *Am J Psychiatry* 150: 1856-1861.
- Alkondon, M., Pereira, E. F., Eisenberg, H. M. & Albuquerque, E. X. (2000). Nicotinic receptor activation in human cerebral cortical interneurons: a mechanism for inhibition and disinhibition of neuronal networks. *J Neurosci* 20:66-75.
- Aracri, P., Consonni, S., Morini, R., Perrella, M., Rodighiero, S., Amadeo, A. & Becchetti, A. (2010). Tonic modulation of GABA release by nicotinic acetylcholine receptors in layer V of the murine prefrontal cortex. *Cereb Cortex* 20(7):1539-55.
- Ashwood, P., Krakowiak, P., Hertz-Picciotto, I., Hansen, R., Pessah, I. N. & Van de Water, J. (2011a). Associations of impaired behaviors with elevated plasma chemokines in autism spectrum disorders. *J Neuroimmunol* 232(1-2):196-9.
- Ashwood, P., Krakowiak, P., Hertz-Picciotto, I., Hansen, R., Pessah, I. & Van de Water, J. (2011b). Elevated plasma cytokines in autism spectrum disorders provide evidence of immune dysfunction and are associated with impaired behavioral outcome. *Brain Behav Immun* 25(1):40-5.
- Avale, M. E., Chabout, J., Pons, S., Serreau, P., De Chaumont, F., Olivo-Marin, J. C., Bourgeois, J. P., Maskos, U., Changeux, J. P. & Granon, S. (2011). Prefrontal nicotinic receptors control novel social interaction between mice. *FASEB J* Mar 18. [Epub ahead of print]
- Bailey, A.R., Giunta, B.N., Obregon D., et al. (2008). Peripheral biomarkers in autism: secreted amyloid precursor protein-alpha as a probable key player in early diagnosis. *Int J Clin Exp Med* 1:338-344.
- Baxter, L. R. Jr., Schwartz, J. M., Mazziotta, J. C., Phelps, M. E., Pahl, J. J., Guze, B. H. & Fairbanks, L. (1988). Cerebral glucose metabolic rates in nondepressed patients with obsessive-compulsive disorder. *Am J Psychiatry* 145(12):1560-3.
- Bejerot, S. & Nylander, L. (2003). Low prevalence of smoking in patients with autism spectrum disorders. *Psychiatry Res* 119(1-2):177-82.
- Bencherif, M., Lippiello, P. M., Lucas, R. & Marrero, M. B. (2011). Alpha7 nicotinic receptors as novel therapeutic targets for inflammation-based diseases. *Cell Mol Life Sci* 68(6):931-49.

- Bertrand, D., Picard, F., Le Hellard, S., Weiland, S., Favre, I., Phillips, H., Bertrand, S., Berkovic, S. F., Malafosse, A. & Mulley, J. (2002). How mutations in the nAChRs can cause ADNFLE epilepsy. *Epilepsia* 43 Suppl 5:112-22.
- Bierut, L. J., Madden, P. A., Breslau, N., Johnson, E. O., Hatsukami, D., Pomerleau, O. F., Swan, G. E., Rutter, J., Bertelsen, S., Fox, L., Fugman, D., Goate, A. M., Hinrichs, A. L., Konvicka, K., Martin, N. G., Montgomery, G. W., Saccone, N. L., Saccone, S. F., Wang, J. C., Chase, G. A., Rice, J. P. & Ballinger, D. G. (2007). Novel genes identified in a high-density genome wide association study for nicotine dependence. *Hum Mol Genet* 16(1):24-35.
- Blundell, J., Tabuchi, K., Bolliger, M. F., Blaiss, C. A., Brose, N., Liu, X., Südhof, T. C. & Powell, C. M. (2009). Increased anxiety-like behavior in mice lacking the inhibitory synapse cell adhesion molecule neuroligin 2. *Genes Brain Behav* 8(1):114-26.
- Blundell, J., Blaiss, C. A., Etherton, M. R., Espinosa, F., Tabuchi, K., Walz, C., Bolliger, M. F., Südhof, T. C. & Powell, C. M. (2010). Neuroligin-1 deletion results in impaired spatial memory and increased repetitive behavior. *J Neurosci* 30(6):2115-29.
- Boucard, A. A., Chubykin, A. A., Comoletti, D., Taylor, P. & Südhof, T. C. (2005). A splice code for trans-synaptic cell adhesion mediated by binding of neuroligin 1 to alpha- and beta-neurexins. *Neuron* 48(2):229-36.
- Braun, A. R., Randolph, C., Stoetter, B., Mohr, E., Cox, C., Vadar, K., Sexton, R., Carson, R. E., Herscovitch, P. & Chase, T. N. (1995). The functional neuroanatomy of Tourette's syndrome: an FDG-PET Study. II: Relationships between regional cerebral metabolism and associated behavioral and cognitive features of the illness. *Neuropsychopharmacology* 13(2):151-68.
- Buckner, R., Andrews-Hanna, J. & Schacter, D. (2008). The brain's default network. *Ann N Y Acad Sci* 1124:1-38.
- Careaga, M., Van de Water, J. & Ashwood, P. (2010). Immune dysfunction in autism: a pathway to treatment. *Neurotherapeutics* 7(3):283-92.
- Carlsson, M. L. (2001). On the role of prefrontal cortex glutamate for the antithetical phenomenology of obsessive compulsive disorder and attention deficit hyperactivity disorder. *Prog Neuropsychopharmacol Biol Psychiatry* 25(1):5-26.
- Cheng, S. B., Amici, S. A., Ren, X. Q., McKay, S. B., Treuil, M. W., Lindstrom, J. M., Rao, J. & Anand, R. (2009). Presynaptic targeting of alpha4beta 2 nicotinic acetylcholine receptors is regulated by neurexin-1beta. *J Biol Chem* 284(35):23251-9.
- Chez, M. G., Dowling, T., Patel, P. B., Khanna, P. & Kominsky, M. (2007). Elevation of tumor necrosis factor-alpha in cerebrospinal fluid of autistic children. *Pediatr Neurol* 36(6):361-5.
- Chih, B., Afridi, S. K., Clark, L., Scheiffele, P. (2004). Disorder-associated mutations lead to functional inactivation of neuroligins. *Hum Mol Genet* 13(14):1471-7.
- Chih, B., Gollan, L., Scheiffele, P. (2006). Alternative splicing controls selective trans-synaptic interactions of the neuroligin neurexin complex. *Neuron* 51(2):171-8.
- Chubykin, A. A., Atasoy, D., Etherton, M. R., Brose, N., Kavalali, E. T., Gibson, J. R. & Südhof, T. C. (2007). Activity dependent validation of excitatory versus inhibitory synapses by neuroligin-1 versus neuroligin-2. *Neuron* 54(6):919-31.
- Clementi, G. C. (2004). Neuronal nicotinic receptors: from structure to pathology. *Prog Neurobiol* 74:363-96.

- Coe, J. W., Brooks, P. R., Vetelino, M. G., Wirtz, M. C., Arnold, E. P., Huang, J., Sands, S. B., Davis, T. I., Lebel, L. A., Fox C. B., Shrikhande, A., Heym, J. H., Schaeffer, E., Rollema, H., Lu, Y., Mansbach, R. S., Chambers, L. K, Rovetti, C. C., Schulz, D. W., Tingley, F. D. 3rd & O'Neill, B. T. (2005). Varenicline: an $\alpha 4\beta 2$ nicotinic receptor partial agonist for smoking cessation. *J Med Chem* 48:3474-7.
- Comoletti, D., De Jacobo, A., Jennings, L. L., Flynn, R. E., Gaietta, G., Tsigelny, I., Ellisman, M. H. & Taylor P. (2004). The Arg451Cys-neurologin-3 mutation associated with autism reveals a defect in protein processing. *J Neurosci* 24(20):4889-93.
- Conroy, W. G., Nai, Q., Ross, B., Naughton, G. & Berg, D. K. (2007). Postsynaptic neurologin enhances presynaptic inputs at neuronal nicotinic synapses. *Dev Biol* 307(1):79-91.
- Cook, E. H. Jr. & Scherer, S. W. (2008). Copy-number variations associated with neuropsychiatric conditions. *Nature* 455(7215):919-23.
- Corbett, B. A., Mendoza, S., Abdullah, M., Wegelin, J. A. & Levine, S. (2006). Cortisol circadian rhythms and response to stress in children with autism. *Psychoneuroendocrinology* 31(1):59-68.
- Couey, J. J., Meredith, R. M., Spijker, S., Poorthuis, R. B., Smit, A. B., Brussaard, A. B. & Mansvelder, H. D. (2007). Distributed network actions by nicotine increase the threshold for spike-timing-dependent plasticity in prefrontal cortex. *Neuron* 54(1):73-87.
- Courchesne, E., Pierce, K., Schumann, C. M., Redcay, E., Buckwalter, J. A., Kennedy, D. P., Morgan, J. (2007). Mapping early brain development in autism. *Neuron* 56(2):399-413.
- Court, J., Spurden, D., Lloyd, S., McKeith, I., Ballard, C., Cairns, N., Kerwin, R., Perry, R. & Perry, E. (1999). Neuronal nicotinic receptors in dementia with Lewy bodies and schizophrenia: α -bungarotoxin and nicotinic binding in thalamus. *J Neurochem* 73:1590-7.
- Court, J., Martin-Ruiz, C., Piggott, M., Spurden, D., Griffiths, M. & Perry, E. (2001). Nicotinic receptor abnormalities in Alzheimer's disease. *Biol Psychiatry* 49:175-84.
- Coury, D. (2010). Medical treatment of autism spectrum disorders. *Curr Opin Neurol* 23(2):131-6.
- Craig, A. M. & Kang, Y. (2007). Neurexin-neurologin signaling in synapse development. *Curr Opin Neurobiol* 17(1):43-52.
- Dajas-Bailador, F. A., Heimala, K. & Wonnacott, S. (2003). The allosteric potentiation of nicotinic acetylcholine receptors by galantamine is transduced into cellular responses in neurons: Ca^{2+} signals and neurotransmitter release. *Mol Pharmacol* 64:1217-1226.
- Dalack, G. W., Becks, L., Hill, E., Pomerleau, O. F. & Meador-Woodruff, J. H. (1999). Nicotine withdrawal and psychiatric symptoms in cigarette smokers with schizophrenia. *Neuropsychopharmacology* 21: 195-202.
- De Fusco, M., Becchetti, A., Patrignani, A., Annesi, G., Gambardella, A., Quattrone, A., Ballabio, A., Wanke, E. & Casari, G. (2000). The nicotinic receptor beta 2 subunit is mutant in nocturnal frontal lobe epilepsy. *Nat Genet* 26(3):275-6.
- de Wit, J., Sylwestrak, E., O'Sullivan, M. L., Otto, S., Tiglio, K., Savas, J. N., Yates, J. R. 3rd, Comoletti, D., Taylor, P. & Ghosh, A. (2009). LRRTM2 interacts with Neurexin1 and regulates excitatory synapse formation. *Neuron* 64(6):799-806.

- Dean, C. & Dresbach, T. (2006). Neuroligins and neuroligins: linking cell adhesion, synapse formation and cognitive function. *Trends Neurosci* 29(1):21-9.
- Dolle, F., Valette, H., Hinnen, F., Vaufrey, F., Demphel, S., Coulon, C., et al. (2001). Synthesis and preliminary evaluation of a carbon-11-labelled agonist of the $\alpha 7$ nicotinic acetylcholine receptor. *J Labelled Cpd Radiopharm* 44:785-95.
- Dudanova, I., Sedej, S., Ahmad, M., Masius, H., Sargsyan, V., Zhang, W., Riedel, D., Angenstein, F., Schild, D., Rupnik, M. & Missler, M. (2006). Important contribution of alpha-neurexins to Ca^{2+} -triggered exocytosis of secretory granules. *J Neurosci* 26(41):10599-613.
- Dudanova, I., Tabuchi, K., Rohlmann, A., Südhof, T. C. & Missler, M. (2007). Deletion of alpha-neurexins does not cause a major impairment of axonal pathfinding or synapse formation. *J Comp Neurol* 502(2):261-74.
- Etherton, M. R., Blaiss, C. A., Powell & C. M., Südhof, T. C. (2009). Mouse neuroligin-1alpha deletion causes correlated electrophysiological and behavioral changes consistent with cognitive impairments. *Proc Natl Acad Sci U S A* 106(42):17998-8003.
- Falk, L., Nordberg, A., Seiger, A., Kjældgaard, A. & Hellström-Lindahl, E. (2003). Higher expression of $\alpha 7$ nicotinic acetylcholine receptors in human fetal compared to adult brain. *Dev Brain Res* 142: 151-60.
- Feng, J., Schroer, R., Yan, J., Song, W., Yang, C., Bockholt, A., Cook, E. H. Jr., Skinner, C., Schwartz, C. E. & Sommer, S. S. (2006). High frequency of neuroligin 1beta signal peptide structural variants in patients with autism. *Neurosci Lett* 409(1):10-3.
- Fombonne, E. (1999). The epidemiology of autism: a review. *Psychol Med* 29(4):769-86.
- Frazier, C. J., Buhler, A. V., Weiner, J. L., & Dunwiddie, T. V. (1998a). Synaptic potentials mediated via alpha-bungarotoxin-sensitive nicotinic acetylcholine receptors in rat hippocampal interneurons. *J Neurosci*. 18:8228-8235.
- Frazier, C. J., Rollins, Y. D., Breese, C. R., Leonard, S., Freedman, R. & Dunwiddie, T. V. (1998b). Acetylcholine activates an alpha-bungarotoxin-sensitive nicotinic current in rat hippocampal interneurons, but not pyramidal cells, *J Neurosci* 18:1187-1195.
- Freedman, R., Hall, M., Adler, L. E. & Leonard, S. (1995). Evidence in postmortem brain tissue for decreased numbers of hippocampal nicotinic receptors in schizophrenia. *Biol Psychiatry* 38:22-33.
- Gai, X., Xie, H. M., Perin, J. C., Takahashi, N., Murphy, K., Wenocur, A. S., D'arcy, M., O'Hara, R. J., Goldmuntz, E., Grice, D. E., Shaikh, T. H., Hakonarson, H., Buxbaum, J. D., Elia, J. & White, P. S. (2011). Rare structural variation of synapse and neurotransmission genes in autism. *Mol Psychiatry* Mar 1. [Epub ahead of print].
- Gao, Y., Horti, A. G., Kuwabara, H., Ravert, H. T., Hilton, J., Holt, D. P., et al. (2007). Derivatives of (-)-7-methyl-2-(5-(pyridinyl)pyridin-3-yl)-7-azabicyclo[2.2.1]heptane are potential ligands for positron emission tomography imaging of extrathalamic nicotinic acetylcholine receptors. *J Med Chem* 50(16):3814-3824.
- Gao, Y., Kuwabara, H., Spivak, C. E., Xiao, Y., Kellar, K., Ravert, H. T., et al. (2008). Discovery of (-)-7-methyl-2-exo-[3'-(6-[^{18}F] fluoropyridin-2-yl)-5'-pyridinyl]-7-azabicyclo[2.2.1]heptane, a radiolabeled antagonist for cerebral nicotinic acetylcholine receptor ($\alpha 4\beta 2$ -nAChR) with optimal positron emission tomography imaging properties. *J Med Chem* 51(15):4751-4764.

- Garritty, A. G., Pearlson, G. D., McKiernan, K., Lloyd, D., Kiehl, K. A. & Calhoun, V. D. (2007). Aberrant “default mode” functional connectivity in schizophrenia. *Am J Psychiatry* 164:450 – 457.
- Gauthier, J., Siddiqui, T. J., Huashan, P., Yokomaku, D., Hamdan, F. F., Champagne, N., Lapointe, M., Spiegelman, D., Noreau, A., Lafrenière, R. G., Fathalli, F., Joobar, R., Krebs, M. O., Delisi, L. E., Motttron, L., Fombonne, E., Michaud, J. L., Drapeau, P., Carbonetto, S., Craig, A. M. & Rouleau, G. A. (2011). Truncating mutations in NRXN2 and NRXN1 in autism spectrum disorders and schizophrenia. *Hum Genet.* 2011 Mar 22. [Epub ahead of print].
- Geschwind, D. H. (2009). Advances in autism. *Annu Rev Med* 60:367-80.
- Gillberg, C. & Billstedt, E. (2000). Autism and Asperger syndrome: coexistence with other clinical disorders. *Acta Psychiatr Scand* 102, 321–330.
- Gilman, S., R., Iossifov, I., Levy, D., Ronemus, M., Wigler, M., Vitkup, D. (2011). Rare De Novo Variants Associated with Autism Implicate a Large Functional Network of Genes Involved in Formation and Function of Synapses. *Neuron* 70(5): 898-907.
- Glessner, J. T., Wang, K., Cai, G., Korvatska, O., Kim, C. E., Wood, S., Zhang, H., Estes, A., Brune C. W., Bradfield, J. P., Imielinski, M., Frackelton, E. C., Reichert, J., Crawford, E. L., Munson, J., Sleiman, P. M., Chiavacci, R., Annaiah, K., Thomas, K., Hou, C., Glaberson, W., et al. (2009). Autism genome-wide copy number variation reveals ubiquitin and neuronal genes. *Nature* 459:569–573.
- Gogolla, N., Leblanc, J. J., Quast, K. B., Südhof, T., Fagiolini, M. & Hensch, T. K. (2009). Common circuit defect of excitatory-inhibitory balance in mouse models of autism. *J Neurodev Disord* 1(2):172-181.
- Graf, E. R., Zhang, X., Jin, S. X., Linhoff, M. W. & Craig, A. M. (2004). Neurexins induce differentiation of GABA and glutamate postsynaptic specializations via neuroligins. *Cell* 119: 1013–1026.
- Granon, S., Faure, P. & Changeux, J. P. (2003). Executive and social behaviors under nicotinic receptor regulation. *Proc Natl Acad Sci U S A* 100(16):9596-601.
- Gunnell, D., Irvine, D., Wise, L., Davies, C. & Martin, R. M. (2009). Varenicline and suicidal behaviour: a cohort study based on data from the General Practice Research Database. *BMJ* 339:b3805.
- Hashimoto, K., Nishiyama, S., Ohba, H., Matsuo, M., Kobashi, T., Takahagi, M., et al. (2008). [11C]CHIBA-1001 as a novel PET ligand for $\alpha 7$ nicotinic receptors in the brain: a PET study in conscious monkeys. *PLoS ONE* 3:e3231.
- Hata, Y., Butz, S. & Südhof, T. C. (1996). CASK: a novel dlg/PSD95 homolog with an N-terminal calmodulin-dependent protein kinase domain identified by interaction with neurexins. *J Neurosci* 16(8):2488-94.
- Hoda, J. C., Wanischek, M., Bertrand, D. & Steinlein, O. K. (2009). Pleiotropic functional effects of the first epilepsy-associated mutation in the human CHRNB2 gene. *FEBS Lett* 583(10):1599-604.
- Horti, A. G., Gao, Y., Kuwabara, H. & Dannals, R. F. (2010). Development of radioligands with optimized imaging properties for quantification of nicotinic acetylcholine receptors by positron emission tomography. *Life Sci* 86(15-16):575-84.
- Inta, D., Vogt, M. A., Lima-Ojeda, J. M., Pfeiffer, N., Schneider, M. & Gass, P. (2011). Lack of long-term behavioral alterations after early postnatal treatment with tropisetron:

- Implications for developmental psychobiology. *Pharmacol Biochem Behav* 99(1):35-41.
- Jamain, S., Quach, H., Betancur, C., Råstam, M., Colineaux, C., Gillberg, I. C., Soderstrom, H., Giros, B., Leboyer, M., Gillberg, C., Bourgeron, T.; Paris Autism Research International Sibpair Study. (2003). Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. *Nat Genet* 34(1):27-9.
- Jamain, S., Radyushkin, K., Hammerschmidt, K., Granon, S., Boretius, S., Varoquaux, F., Ramanantsoa, N., Gallego, J., Ronnenberg, A., Winter, D., Frahm, J., Fischer, J., Bourgeron, T., Ehrenreich, H. & Brose, N. (2008). Reduced social interaction and ultrasonic communication in a mouse model of monogenic heritable autism. *Proc Natl Acad Sci U S A* 105(5):1710-5.
- Kang, Y., Zhang, X., Dobie, F., Wu, H. & Craig, A. M. (2008). Induction of GABAergic postsynaptic differentiation by alpha-neurexins. *J Biol Chem* 283(4):2323-34.
- Kas, M.J., Fernandes, C., Schalkwyk, L.C., & Collier, D.A. (2007). Genetics of behavioural domains across the neuropsychiatric spectrum; of mice and men. *Mol Psychiatry* 12(4):324-30.
- Kihara, T., Shimohama, S., Sawada, H., Kimura, J., Kume, T., Kochiyama, H., Maeda, T., & Akaike, A. (1997). Nicotinic receptor stimulation protects neurons against beta-amyloid toxicity. *Ann Neurol* 42:159-63.
- Kihara, T., Shimohama, S., Urushitani, M., Sawada, H., Kimura, J., Kume, T., Maeda, T., and Akaike, A. (1998). Stimulation of alpha4beta2 nicotinic acetylcholine receptors inhibits beta-amyloid toxicity. *Brain Res* 792:331-4.
- Kihara, T., Shimohama, S., Sawada, H., Honda, K., Nakamizo, T., Shibasaki, H., Kume, T., & Akaike, A. (2001). alpha 7 Nicotinic Receptor Transduces Signals to Phosphatidylinositol 3-Kinase to Block A beta -Amyloid-induced Neurotoxicity. *J Biol Chem* 276:13541-6.
- Kim, H. G., Kishikawa, S., Higgins, A. W., Seong, I. S., Donovan, D. J., Shen, Y., Lally, E., Weiss, L. A., Najm, J., Kutsche, K., Descartes, M., Holt, L., Braddock, S., Troxell, R., Kaplan, L., Volkmar, F., Klin, A., Tsatsanis, K., Harris, D. J., Noens, I., Pauls, D. L., Daly, M. J., MacDonald, M. E., Morton, C. C., Quade, B. J. & Gusella, J. F. (2008). Disruption of neurexin 1 associated with autism spectrum disorder. *Am J Hum Genet* 82(1):199-207.
- Kirov, G., Rujescu, D., Ingason, A., Collier, D. A., O'Donovan, M. C. & Owen, M. J. (2009). Neurexin 1 (NRXN1) deletions in schizophrenia. *Schizophr Bull* 35(5):851-4.
- Kitagawa, H., Takenouchi, T., Azuma, R., Wesnes, K., Kramer, W., Clody, D. & Burnett, A. L. (2003). Safety, pharmacokinetics, and effects on cognitive function of multiple doses of GTS-21 in healthy, male volunteers. *Neuropsychopharmacology* 28:542-551.
- Ko, J., Fuccillo, M. V., Malenka, R. C. & Südhof, T. C. (2009). LRRTM2 functions as a neurexin ligand in promoting excitatory synapse formation. *Neuron* 64(6):791-8.
- Konstantareas, M. M. & Homatidis, S. (1999). Chromosomal abnormalities in a series of children with autistic disorder. *J Autism Dev Disord* 29:275-285.
- Langen, M., Durston, S., Kas, M.J., van Engeland, H., & Staal, W.G. (2011). The neurobiology of repetitive behavior: ... and men. *Neurosci and Biobehavioral Rev* 35:356-65.
- Laumonnier, F., Bonnet-Brilhault, F., Gomot, M., Blanc, R., David, A., Moizard, M. P., Raynaud, M., Ronce, N., Lemonnier, E., Calvas, P., Laudier, B., Chelly, J., Fryns, J. P., Ropers, H. H., Hamel, B. C., Andres, C., Barthélémy, C., Moraine, C. & Briault, S.

- (2004). X-linked mental retardation and autism are associated with a mutation in the NLGN4 gene, a member of the neuroligin family. *Am J Hum Genet* 74(3):552-7.
- Lee, M., Martin-Ruiz, C., Graham, A., Court, J., Jaros, E., Perry, R., Iversen, P., Bauman M. & Perry, E. (2002). Nicotinic receptor abnormalities in the cerebellar cortex in autism. *Brain* 125(Pt 7):1483-95.
- Lee, S. W., Lee, J. G., Lee, B. J. & Kim, Y. H. (2007). A 12-week, double-blind, placebo-controlled trial of galantamine adjunctive treatment to conventional antipsychotics for the cognitive impairments in chronic schizophrenia. *Int Clin Psychopharm* 22:63-68.
- Léna, C., Popa, D., Grailhe, R., Escourrou, P., Changeux, J. P. & Adrien, J. (2004). Beta2-containing nicotinic receptors contribute to the organization of sleep and regulate putative micro-arousals in mice. *J Neurosci* 24(25):5711-8.
- Levin, E. D. & Simon, B. B. (1998). Nicotinic acetylcholine involvement in cognitive function in animals. *Psychopharmacology* 138(3-4):217-30.
- Levitt, P. & Campbell, D. B. (2009). The genetic and neurobiologic compass points toward common signaling dysfunctions in autism spectrum disorders. *J Clin Invest* 119(4):747-54.
- Levy, D., Ronemus, M., Yamrom, B., Lee, Y., Leotta, A., Kendall, J., et al. (2011). Rare De Novo and Transmitted Copy-Number Variation in Autistic Spectrum Disorders. *Neuron* 70(5): 886-897.
- Levy, S. E., Mandell, D. S. & Schultz, R. T. (2009). Autism. *Lancet* 374(9701):1627-38.
- Lipton, S. A., Frosch, M. P., Phillips, M. D., Tauck, D. L. & Aizenman, E. (1998). Nicotinic antagonists enhance process outgrowth by rat retinal ganglion cells in culture. *Science* 239:1293-1296.
- Lisé, M. F. & El-Husseini, A. (2006). The neuroligin and neurexin families: from structure to function at the synapse. *Cell Mol Life Sci* 63(16):1833-49.
- Lindstrom, J. (1996). Neuronal nicotinic acetylcholine receptors. *Ion Channels* 4:377-450.
- Lindstrom, J. (1997). Nicotinic acetylcholine receptors in health and disease. *Mol. Neurobiol* 15:193-222.
- López-Valdés, H. E., García-Colunga, J. & Miledi, R. (2002). Effects of clomipramine on neuronal nicotinic acetylcholine receptors. *Eur J Pharmacol* 444(1-2):13-9.
- Marshall CR, Noor A, Vincent JB, Lionel AC, Feuk L, Skaug J, Shago M, Moessner R, Pinto D, Ren Y, Thiruvahindrapduram B, Fiebig A, Schreiber S, Friedman J, Ketelaars CE, Vos YJ, Ficicioglu C, Kirkpatrick S, Nicolson R, Sloman L, Summers A, Gibbons CA, Teebi A, Chitayat D, Weksberg R, Thompson A, Vardy C, Crosbie V, Luscombe S, Baatjes R, Zwaigenbaum L, Roberts W, Fernandez B, Szatmari P, Scherer SW. (2008). Structural variation of chromosomes in autism spectrum disorder. *Am J Hum Genet* 82(2):477-88.
- Martin-Ruiz, C. M., Lee, M., Perry, R. H., Baumann, M., Court, J. A. & Perry, E.K. (2004). Molecular analysis of nicotinic receptor expression in autism. *Brain Res Mol Brain Res* 123(1-2):81-90.
- Marutle, A., Zhang, X., Court, J., Piggott, M., Johnson, M., Perry, R., Perry, E. & Nordberg, A. (2001). Laminar distribution of nicotinic receptor subtypes in cortical regions in schizophrenia. *J Chem Neuroanat* 22:115-26.
- Maskos, U., Molles, B. E., Pons, S., Besson, M., Guiard, B. P., Guilloux, J. P., Evrard, A., Cazala, P., Cormier, A., Mameli-Engvall, M., Dufour, N., Cloëz-Tayarani, I.,

- Bemelmans, A. P., Mallet, J., Gardier, A. M., David, V., Faure, P., Granon, S. & Changeux, J. P. (2005). Nicotine reinforcement and cognition restored by targeted expression of nicotinic receptors. *Nature* 436(7047):103-7.
- McClure, J. B., Swan, G. E., Jack, L., Catz, S. L., Zbikowski, S. M., McAfee, T. A., Deprey, M., Richards, J. & Javitz, H. (2009). Mood, side-effects and smoking outcomes among persons with and without probable lifetime depression taking varenicline. *J Gen Intern Med* 24(5):563-9.
- McGehee, D. S. & Role, L. W. (1996). Presynaptic ionotropic receptors. *Curr Opin Neurobiol* 6:342-349.
- Merikangas, A. K., Corvin, A. P. & Gallagher, L. (2009) Copy-number variants in neurodevelopmental disorders: promises and challenges. *Trends Genet* 25(12):536-44.
- Messi, M. L., Renganathan, M., Grigorenko, E. & Delbono, O. (1997). Activation of alpha7 nicotinic acetylcholine receptor promotes survival of spinal cord motoneurons. *FEBS Lett* 411:32-8.
- Mihalak, K.B., Carroll, F. I. & Luetje, C.W. (2006). Varenicline is a partial agonist at alpha4beta2 and a full agonist at alpha7 neuronal nicotinic receptors. *Mol Pharmacol* 70(3):801-5.
- Miller, D.,T., Shen, Y., Weiss, L., A., Korn, J., Anselm, I., Bridgemohan, C., et al. (2009). Microdeletion/duplication at 15q13.2q13.3 among individuals with features of autism and other neuropsychiatric disorders. *J Med Genet.* 46(4):242-8.
- Mineur, Y. S. & Picciotto, M.R. (2010). Nicotine receptors and depression: revisiting and revising the cholinergic hypothesis. *Trends Pharmacol Sci* 31(12):580-6.
- Missler, M. & Südhof TC. (1998). Neurexins: three genes and 1001 products. *Trends Genet* 14(1):20-6.
- Missler, M., Zhang, W., Rohlmann, A., Kattenstroth, G., Hammer, R. E., Gottmann, K. & Südhof, T.C. (2003). Alpha-neurexins couple Ca²⁺ channels to synaptic vesicle exocytosis. *Nature* 423(6943):939-48.
- Miwa, J. M., Freedman, R. & Lester, H. A. (2011). Neural systems governed by nicotinic acetylcholine receptors: emerging hypotheses. *Neuron.* 70(1):20-33.
- Morgan, J. T., Chana, G., Pardo, C. A., Achim, C., Semendeferi, K., Buckwalter, J., Courchesne, E. & Everall, I. P. (2010) Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biol Psychiatry* 68(4):368-76.
- Nam, C. I. & Chen, L. (2005). Postsynaptic assembly induced by neurexin-neuroligin interaction and neurotransmitter. *Proc Natl Acad Sci USA* 102(17):6137-42.
- Newhouse, P. A. & Kelton, M. (2000). Nicotinic systems in central nervous systems disease: degenerative disorders and beyond. *Pharm Acta Helv* 74(2-3):91-101.
- Newhouse, P., Singh, A. & Potter, A. (2004). Nicotine and nicotinic receptor involvement in neuropsychiatric disorders. *Curr Top Med Chem* 4(3):267-82.
- Niaura, R., Jones, C. & Kirkpatrick, P. (2006). Varenicline. *Nat Rev Drug Discov* 5:537-8.
- Nicolson, R., Craven-Thuss, B. & Smith, J. (2006). A prospective, open-label trial of galantamine in autistic disorder. *J Child Adolesc Psychopharmacol* 16(5):621-9.
- Nussbaum, J., Xu, Q., Payne, T. J., Ma, J. Z., Huang, W., Gelernter, J. & Li, M. D. (2008). Significant association of the neurexin-1 gene (NRXN1) with nicotine dependence in European- and African-American smokers. *Hum Mol Genet* 17(11):1569-77.

- Ochoa, E.L. & Lasalde-Dominicci, J. (2007). Cognitive deficits in schizophrenia: Focus on neuronal nicotinic acetylcholine receptors and smoking. *Cell Mol Neurobiol* 27(5):609–639.
- Ogawa, M., Tatsumi, R., Fujio, M., Katayama, J. & Magata, Y. (2006). Synthesis and evaluation of [125I]I-TSA as a brain nicotinic acetylcholine receptor $\alpha 7$ subtype imaging agent. *Nucl Med Biol* 33:311–36.
- Papke, R. L., Trocmé-Thibierge, C., Guendisch, D., Al Rubaiy, S. A. & Bloom, S.A. (2011). Electrophysiological perspectives on the therapeutic use of nicotinic acetylcholine receptor partial agonists. *J Pharmacol Exp Ther* 337(2):367-79.
- Pasquini, M., Garavini, A. & Biondi, M. (2005). Nicotine augmentation for refractory obsessive-compulsive disorder. A case report. *Prog Neuropsychopharmacol Biol Psychiatry* 29(1):157-9.
- Perry, E. K., Lee, M. L., Martin-Ruiz, C. M., Court, J. A., Volsen, S. G., Merrit, J., Folly, E., Iversen, P. E., Bauman, M. L., Perry, R. H. & Wenk, G. L. (2001). Cholinergic activity in autism: abnormalities in the cerebral cortex and basal forebrain. *Am J Psychiatry* 158(7):1058-66.
- Pimlott, S. L., Piggott, M., Owens, J., Greally, E., Court, J. A., Jaros, E., Perry, R. H., Perry, E. K. & Wyper, D. (2004). Nicotinic acetylcholine receptor distribution in Alzheimer's disease, dementia with Lewy bodies, Parkinson's disease, and vascular dementia: In vitro binding study using 5-[125I]-A-85380. *Neuropsychopharmacology* 29(1):108–116.
- Pomper, M. G., Phillips, E., Fan, H., McCarthy, D. J., Keith, R. A., Gordon, J. C., Scheffel, U., Dannals, R. F. & Musachio, J. L. (2005). Synthesis and biodistribution of radiolabeled $\alpha 7$ nicotinic acetylcholine receptor ligands. *J Nucl Med* 46: 326–34.
- Pouloupoulos, A., Aramuni, G., Meyer, G., Soykan, T., Hoon, M., Papadopoulos, T., Zhang, M., Paarmann, I., Fuchs, C., Harvey, K., Jedlicka, P., Schwarzacher, S. W., Betz, H., Harvey, R. J., Brose, N., Zhang, W. & Varoqueaux, F. (2009). Neuroligin 2 drives postsynaptic assembly at perisomatic inhibitory synapses through gephyrin and collybistin. *Neuron* 63(5):628-42.
- Potter, A., Corwin, J., Lang, J., Piasecki, M., Lenox, R. & Newhouse, P. A. (1999). Acute effects of the selective cholinergic channel activator (nicotinic agonist) ABT-418 in Alzheimer's disease. *Psychopharmacology* 142(4):334-42.
- Pugh, P. C., and Berg, D. K. (1994). Neuronal acetylcholine receptors that bind alpha-bungarotoxin mediate neurite retraction in a calcium-dependent manner, *J Neurosci* 14:889-896.
- Pugh, P. C. & Margiotta, J. F. (2000). Nicotinic acetylcholine receptor agonists promote survival and reduce apoptosis of chick ciliary ganglion neurons. *Mol Cell Neurosci* 15:113-22.
- Ramocki, M.B. & Zoghbi, H.Y. (2008). Failure of neuronal homeostasis results in common neuropsychiatric phenotypes. *Nature* 455(7215):912-8.
- Rojas, S., Martín, A., Arranz, M. J., Pareto, D., Purroy, J., Verdager, E., Llop, J., Gómez, V., Gispert, J.D., Millán, O., Chamorro, A. & Planas, A.M. (2007) Imaging brain inflammation with [(11)C]PK11195 by PET and induction of the peripheral-type benzodiazepine receptor after transient focal ischemia in rats. *J Cereb Blood Flow Metab* 27(12):1975-86.

- Rosas-Ballina, M. & Tracey, K. J. (2009). The neurology of the immune system: neural reflexes regulate immunity. *Neuron* 64(1):28-32.
- Ross, B. S & Conroy, W. G. (2008) Capabilities of neurexins in the chick ciliary ganglion. *Dev Neurobiol.* 68(3):409-19.
- Ross, S. A., Wong, J. Y., Clifford, J. J., Kinsella, A., Massalas, J. S., Horne, M. K., Scheffer, I. E., Kola, I., Waddington, J. L., Berkovic, S. F. & Drago, J. (2000) Phenotypic characterization of an alpha 4 neuronal nicotinic acetylcholine receptor subunit knock-out mouse. *J Neurosci.* 20(17):6431-41.
- Rubenstein, J. & Merzenich, M. (2003). Model of autism: increased ratio of excitation/inhibition in key neural systems. *Genes, Brain, and Behavior* 2(5):255-267.
- Samochocki, M., Höffle, A., Fehrenbacher, A., Jostock, R., Ludwig, J., Christner, C., Radina, M., Zerlin, M., Ullmer, C., Pereira, E. F. R., Lübert, H., Albuquerque, E. X. & Maelicke, A. (2003). Galantamine is an allosterically potentiating ligand of neuronal nicotinic but not of muscarinic acetylcholine receptors. *J Pharmacol Exp Ther* 305:1024-1036.
- Sanberg, P. R., Silver, A. A., Shytle, R. D., Philipp, M. K., Cahill, D. W., Fogelson, H. M., McConville, B. J. (1997) Nicotine for the treatment of Tourette's syndrome. *Pharmacol Ther* 74(1):21-5.
- Sargent, P. B. (1993). The diversity of neuronal nicotinic acetylcholine receptors. *Annu Rev Neurosci* 16:403-443.
- Schilström, B., Ivanov, V. B., Wiker, C. & Svensson, T.H. (2007). Galantamine enhances dopaminergic neurotransmission in vivo via allosteric potentiation of nicotinic acetylcholine receptors. *Neuropsychopharmacology* 32:43-53.
- Schubert, M. X., Young, K. A. & Hicks, P. B. (2006). Galantamine improves cognition in schizophrenic patients stabilized on risperidone. *Biol Psychiatry* 60:530-533.
- Shiina, A., Shirayama, Y., Niitsu, T., Hashimoto, T., Yoshida, T., Hasegawa, T., Haraguchi, T., Kanahara, N., Shiraishi, T., Fujisaki, M., Fukami, G., Nakazato, M., Iyo, M. & Hashimoto, K. (2010). A randomised, double-blind, placebo-controlled trial of tropisetron in patients with schizophrenia. *Ann Gen Psychiatry* 24(9):27.
- Sokol, D. K., Chen, D., Farlow, M. R., Dunn, D. W., Maloney, B., Zimmer, J. A. & Lahiri, D. K. (2006). High levels of Alzheimer beta-amyloid precursor protein (APP) in children with severely autistic behavior and aggression. *J Child Neurol* 21(6):444-9.
- Steinlein, O. K. (2002). Nicotinic acetylcholine receptors and epilepsy. *Curr Drug Targets CNS Neurol Disord* 1(4):443-8.
- Südhof, T. C. (2008) Neuroligins and neurexins link synaptic function to cognitive disease. *Nature* 455(7215):903-11.
- Szatmari P, Paterson AD, Zwaigenbaum L, Roberts W, Brian J, Liu XQ, et al. Autism Genome Project Consortium. (2007). Mapping autism risk loci using genetic linkage and chromosomal rearrangements. *Nat Genet* 39(3):319-28.
- Siddiqui, T. J., Pancaroglu, R., Kang, Y., Rooyakkers, A. & Craig, A. M. (2010). LRRTMs and neuroligins bind neurexins with a differential code to cooperate in glutamate synapse development. *J Neurosci* 30:7495-7506.
- Tabuchi K, Blundell, J., Etherton, M. R., Hammer, R. E., Liu, X., Powell, C. M. & Südhof, T. C. (2007). A neuroligin-3 mutation implicated in autism increases inhibitory synaptic transmission in mice. *Science* 318:71-76.

- Takata, K., Kitamura, Y., Saeki, M., Terada, M., Kagitani, S., Kitamura, R., Fujikawa, Y., Maelicke, A., Tomimoto, H., Taniguchi, T. & Shimohama, S. (2010). Galantamine-induced amyloid- β clearance mediated via stimulation of microglial nicotinic acetylcholine receptors. *J Biol Chem* 285(51):40180-91.
- Taly, A., Corringier, P. J., Guedin, D., Lestage, P. & Changeux, J.P. (2009) Nicotinic receptors: allosteric transitions and therapeutic targets in the nervous system. *Nat Rev Drug Discov* 8(9):733-50.
- Tapper, A. R., McKinney, S. L., Nashmi, R., Schwarz, J., Deshpande, P., Labarca, C., Whiteaker, P., Marks, M. J., Collins, A.C. & Lester, H. A. (2004). Nicotine activation of $\alpha 4^*$ receptors: sufficient for reward, tolerance, and sensitization. *Science* 306(5698):1029-32.
- Toyohara, J., Sakata, M., Wu, J., Ishikawa, M., Oda, K., Ishii, K., et al. (2009). Preclinical and the first clinical studies on [11C]CHIBA-1001 for mapping $\alpha 7$ nicotinic receptors by positron emission tomography. *Ann Nucl Med* 23(3):301-9.
- Tregellas, J. R., Tanabe, J., Rojas, D. C., Shatti, S., Olincy, A., Johnson, L., Martin, L. F., Soti, F., Kem, W. R., Leonard, S. & Freedman, R. (2011). Effects of an $\alpha 7$ -nicotinic agonist on default network activity in schizophrenia. *Biol Psychiatry* 69(1):7-11.
- Uemura, T., Lee, S. J., Yasumura, M., Takeuchi, T., Yoshida, T., Ra, M., Taguchi, R., Sakimura, K. & Mishina, M. (2010). Trans-synaptic interaction of GluRdelta2 and Neurexin through Cbln1 mediates synapse formation in the cerebellum. *Cell* 141(6):1068-79.
- Vargas, D. L., Nascimbene, C., Krishnan, C., Zimmerman, A. W. & Pardo, C. A. (2005). Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann Neurol* 57(1):67-81.
- Varoqueaux, F., Aramuni, G., Rawson, R. L., Mohrmann, R., Missler, M., Gottmann, K., Zhang, W., Südhof, T.C. & Brose, N. (2006). Neuroligins determine synapse maturation and function. *Neuron* 51(6):741-54.
- Wang, D., Noda, Y., Zhou, Y., Mouri, A., Mizoguchi, H., Nitta, A., Chen, W. & Nabeshima, T. (2007). The allosteric potentiation of nicotinic acetylcholine receptors by galantamine ameliorates the cognitive dysfunction in beta amyloid25–35 icv-injected mice: involvement of dopaminergic systems. *Neuropsychopharmacology* 32:1261–1271.
- Wang, H., Yu, M., Ochani, M., Amella, C. A., Tanovic, M., Susarla, S., Li, J. H., Wang, H., Yang, H., Ulloa, L., Al-Abed, Y., Czura, C. J. & Tracey, K. J. (2003). Nicotinic acetylcholine receptor $\alpha 7$ subunit is an essential regulator of inflammation. *Nature* 421(6921):384-8.
- Westmark, C. J., Westmark, P. R., Beard, A. M., Hildebrandt, S. M. & Malter, J. S. (2008). Seizure susceptibility and mortality in mice that over-express amyloid precursor protein. *Int J Clin Exp Pathol.* 1(2):157-68.
- Wilens, T. E., Biederman, J., Spencer, T. J., Bostic, J., Prince, J., Monuteaux, M. C., Soriano, J., Fine, C., Abrams, A., Rater, M. & Polisner, D. (1999) A pilot controlled clinical trial of ABT-418, a cholinergic agonist, in the treatment of adults with attention deficit hyperactivity disorder. *Am J Psychiatry* 156(12):1931-7.
- Wills, S., Cabanlit, M., Bennett, J., Ashwood, P., Amaral, D. G. & Van de Water, J. (2009). Detection of autoantibodies to neural cells of the cerebellum in the plasma of subjects with autism spectrum disorders. *Brain Behav Immun* 23(1):64-74.

- Yan, J., Noltner, K., Feng, J., Li, W., Schroer, R., Skinner, C, Zeng, W., Schwartz, C. E. & Sommer, S. S. (2008). Neurexin 1 α structural variants associated with autism. *Neurosci Lett* 438(3):368-70.
- Ylisaukko-oja, T., Rehnström, K., Auranen, M., Vanhala, R., Alen, R., Kempas, E., Ellonen, P., Turunen, J. A., Makkonen, I., Riikonen, R., Nieminen-von Wendt, T., von Wendt, L., Peltonen, L. & Järvelä, I. (2005). Analysis of four neuroligin genes as candidates for autism. *Eur J Hum Genet* 13:285–292.
- Zahir, F. R., Baross, A., Delaney, A. D., Eydoux, P., Fernandes, N. D., Pugh, T., Marra, M. A., & Friedman, J. M. (2008). A patient with vertebral, cognitive and behavioural abnormalities and a de novo deletion of NRXN1alpha. *J Med Genet* 45(4):239-43.
- Zametkin, A. J., Nordahl, T. E., Gross, M., King, A. C., Semple, W. E., Rumsey, J., Hamburger, S. & Cohen, R. M. (1990) Cerebral glucose metabolism in adults with hyperactivity of childhood onset. *N Engl J Med*. 323(20):1361-6.
- Zhang, Z. W., Coggan, J. S. & Berg, D. K. (1996). Synaptic currents generated by neuronal acetylcholine receptors sensitive to alpha-bungarotoxin. *Neuron* 17:1231-40.
- Zhang, W., Rohlmann, A., Sargsyan, V., Aramuni, G., Hammer, R. E., Südhof, T. C. & Missler, M. (2005). Extracellular domains of alpha-neurexins participate in regulating synaptic transmission by selectively affecting N- and P/Q-type Ca²⁺ channels. *J Neurosci*. 25(17):4330-42.
- Zhang, C., Atasoy, D., Araç, D., Yang, X., Fucillo, M. V., Robison, A.J., Ko, J., Brunger, A.T., & Südhof T.C. (2010). Neurexins physically and functionally interact with GABA(A) receptors. *Neuron* 66(3):403-16.

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The book covers some of the key research developments in autism and brings together the current state of evidence on the neurobiologic understanding of this intriguing disorder. The pathogenetic mechanisms are explored by contributors from diverse perspectives including genetics, neuroimaging, neuroanatomy, neurophysiology, neurochemistry, neuroimmunology, neuroendocrinology, functional organization of the brain and clinical applications from the role of diet to vaccines. It is hoped that understanding these interconnected neurobiological systems, the programming of which is genetically modulated during neurodevelopment and mediated through a range of neuropeptides and interacting neurotransmitter systems, would no doubt assist in developing interventions that accommodate the way the brains of individuals with autism function. In keeping with the multimodal and diverse origins of the disorder, a wide range of topics is covered and these include genetic underpinnings and environmental modulation leading to epigenetic changes in the aetiology; neural substrates, potential biomarkers and endophenotypes that underlie clinical characteristics; as well as neurochemical pathways and pathophysiological mechanisms that pave the way for therapeutic interventions.

How to reference

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