We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

186,000

200M

Downloads

154

Our authors are among the

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Chapter

Vitamins D and B₁₂, Altered Synaptic Plasticity and Extracellular Matrix

Marcela Bermudez Echeverry, Silvia Honda Takada, Bruna Petrucelli Arruda, Debora Sterzeck Cardoso, Pamela Pinheiro Martins, Juliane Midori Ikebara, Aline V. Sousa-Santos and Victor R.C. Torres da Silva

Abstract

Brain plasticity is regulated through dynamic interactions between perineuronal nets, matrix metalloproteases (MMPs) and the extracellular matrix (ECM). Several studies have identified a crucial role for vitamins D and B_{12} in brain development and a deficiency in these vitamins may contribute to the emergence of cognitive deficits, as well as the onset of both autism spectrum disorder and schizophrenia. However, the mechanisms underlying the interplay between ECM, MMPs, vitamins and these neuropsychiatric conditions are poorly understood. In this chapter, we seek to understand how the risk of neurodegeneration in vulnerable individuals and the aetiology of specific neuropsychiatric disorders are affected by vitamin D and B_{12} deficiency, in conjunction with low levels of the antioxidant glutathione, impaired GABAergic inhibition, and alterations in the permanent ECM.

Keywords: vitamin deficiency, perineuronal nets, matrix metalloproteases, parvalbumin interneurons, GABA, neurodevelopment

1. Introduction

A proteoglycan-rich matrix, the perineuronal net (PNN) is a dense structure within the extracellular matrix (ECM), whose synapses form through gaps around many neuronal bodies and dendrites at a late stage in brain development. PNNs are formed at the end of a critical period of neurodevelopment, following the transformation of the central nervous system (CNS) from an environment conducive to neuronal growth and motility to one that is more restrictive, in response to several sensory inputs from both neurons and glia driving increased neuroplasticity [1]. The main components of the PNN matrix include several chondroitin sulfate proteoglycans (CSPGs), such as hyaluronan, link proteins, and tenascin-R and -C.

During mammalian development, hyaluronan binds to members of the lectican family originally produced in neurons, including versican V0 and V1 and neurocan [2], whereas aggrecan seems to be expressed by astroglial cells in the juvenile matrix [3]. Other lecticans include versican 2, brevican, phosphacan, tenascin-R and the

link proteins HAPLN2/Bral1 and HAPLN4/Bral2 are only observed in more mature matrix environments approximately 2 weeks after birth [4–6], in contrast to the composition of the juvenile matrix. Following this period, shifts in brevican expression occur at the end of myelination, leading to white-matter precursor changes from an oligodendroglial to an astrocytic lineage [7], and resulting in a compact extracellular matrix forming the PPN [8].

PNNs have been observed 2–5 weeks after birth around parvalbumin (PV⁺)-expressing GABAergic interneurons in pyramidal cortex, and around large motor neurons of the brainstem and spinal cord. This period coincides with the end of experience-dependent refinement of the synaptic network [8], but marked by a still critical period of matrix turnover and proteoglycan degradation by ADAMTS metalloendopeptidases and matrix metalloproteinases (MMPs) [8, 9]. PNN formations can also be observed in several distinct areas of the CNS, such as other regions of the cerebral cortex, the hippocampus (HPC), thalamus, and cerebellum [8].

PNNs in the adult CNS secrete hyaluronan through the action of membranebound HA synthase, an enzyme linked to the action of link proteins, lecticans, tenascin-R and chondroitin sulphate proteoglycans (CSPGs), creating supramolecular aggregates on the surface of neurons [1]. Other relevant glycoproteins besides CSPGs include Reelin, mainly secreted by Cajal-Retzius cells and involved in the control of neuronal migration and the establishment of cell aggregation and dendrite formation during the embryonic and early postnatal stages of development [10]. In adulthood, Reelin signalling is involved in the modulation of synaptic function and binds to very-low-density lipoprotein receptors and apolipoprotein E receptor 2 [11]. Increased clustering of Reelin receptors leads to a build-up of DAB1 proteins on the neuron membrane, greater activation of Src/SFK family kinases, and tyrosine phosphorylation of N-methyl-D-aspartate receptors (NMDARs), resulting in a net increase of receptor activity (**Figure 1**) [12]. Reelin insufficiency may lead to alterations in NMDAR clustering and LTP, such as in dysfunctional GABA-ergic transmission in the cerebral cortex and hippocampus observed among the morphofunctional signalling changes in schizophrenia (SZ) [12].

In fact, alterations of GABAergic signalling within a prenatal stress period have been identified as important factors in the development of SZ [13], autism spectrum disorder (ASD) [14], and epilepsy [15], often leading to an altered density of GABAergic cells and aberrant oscillatory activity. However, one functional model of brain development has proposed that prenatal stress involves DNA methylation, possibly inducing methylation of the gene responsible for Reelin promoter, with the consequent down-expression of Reelin resulting in abnormalities within the neuronal architecture of the prefrontal cortex, a reduction in dendritic complexity and a decreased number of GABAergic neurons, leading to altered developmental neuronal connectivity [13].

Animal studies have demonstrated that DNA methylation in the BDNF gene controls its expression during forebrain development in mice [16]. Furthermore, binding of BDNF and nerve growth factor (NGF) neurotrophins to their respective receptors (TrkB/A) triggers the PI-3kinase/AKT pathway, with activation of the mammalian target of rapamycin (mTOR) [17] and Akt-dependent inhibition of the serine/threonine kinase Gsk3 β , resulting in decreased transcription of pro-inflammatory cytokines (IL-1 β , TNF- α , and IL-6) [18]. Since Reelin/lipoprotein receptors do not contain a cytoplasmic kinase domain, the core Reelin signalling pathway seems to be associated with tyrosine kinase receptor (RTK or Trk) activity [19], the most likely coreceptor candidate [20]. As such, Reelin signalling from the ECM in collaboration with TrkB/A receptor activation, leads to increased phosphorylation of

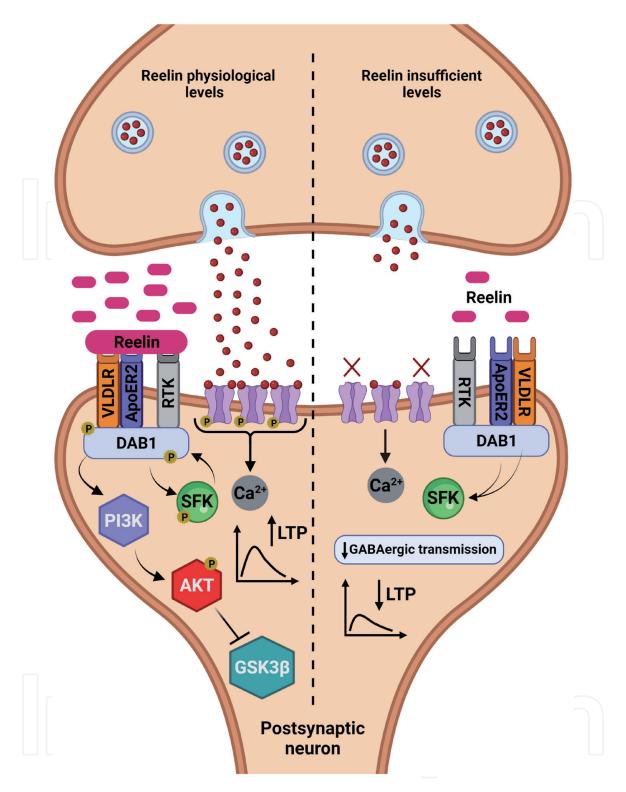


Figure 1.

Effect of Reelin concentrations in the ECM on NMDA signalling. Reelin activates adaptor protein disabled 1 (DAB1) by binding with its very-low-density lipoprotein receptor (VLDLR) and apolipoprotein E receptor type 2 (APOER2). DAB1 is phosphorylated by Src family kinases (SFK) at different sites on the protein - this phosphorylation occurs mainly through the action of a co-receptor, tyrosine kinase receptor (RTK or Trk), implicated in a variety of cellular processes including growth, differentiation, and regulation of energy metabolism in the neuron. DAB1 phosphorylation leads to inhibition of the serine/threonine kinase Gsk3 β via protein kinase B (Akt), where a decrease in AKT phosphorylation levels with subsequent high levels of GSK-3 β phosphorylation, has been observed in lymphocytes and brains of individuals with schizophrenia (SZ). Clustering of Reelin receptors via SKF activation also leads to greater tyrosine phosphorylation of N-methyl-D-aspartate receptors (NMDARs), resulting in a net increase of receptor activity following the induction of long-term potentiation via Ca2 $^+$ regulation. This signalling cascade appears to be an essential process for neurobiological regulation during neurodevelopment (modified from [12]). Created with BioRender.

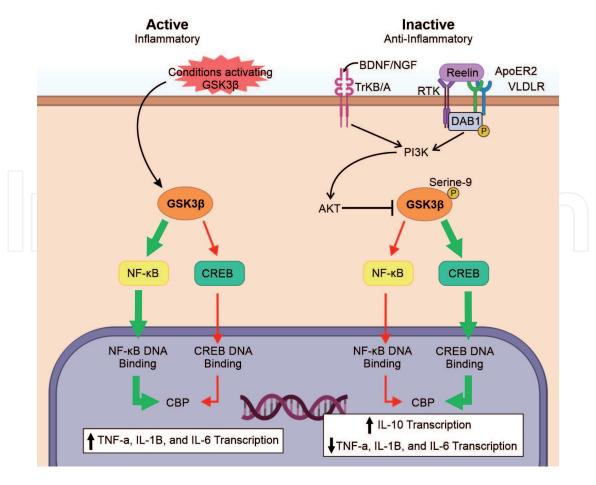


Figure 2.

Hypothetical signalling cascades for GSK3 β modulation of the expression of pro-inflammatory and anti-inflammatory cytokines in glial cells. Receptor crosstalk between receptors TrkB/A and Reelin in the ECM (see **Figure 1**), increase the phosphorylation of serine-9 GSK3 β leading to GSK3 β inhibition, an increase in the translocation of CREB from the cytoplasm to the nucleus, and an increase in the transcription of anti-inflammatory cytokine (IL-10). In fact, GSK3 β can modulate the expression of both pro-inflammatory and anti-inflammatory cytokines. GSK3 β activation in glial cells triggers the NF- κ B pathway and the translocation of NF- κ B from the cytoplasm to the nucleus, leading to an increase in the transcription of pro-inflammatory cytokines ((IL-1 β , TNF- α , and IL-6), via the action of CREB-binding protein (CBP). Inhibition of GSK3 β results in an increase of CREB translocation from the cytoplasm to the nucleus. Phosphorylated CREB binds specifically to the nuclear CBPs at transcriptional sites, resulting in increased transcription of anti-inflammatory cytokines such as IL-10 (modified from [21]).

serine-9 GSK3 β , with the inhibition of GSK3 β in glial cells and leukocytes resulting in more CREB being translocated from the cytoplasm to the nucleus, and an increase in the transcription of anti-inflammatory cytokines (IL-10) (**Figure 2**) [21].

PNN development and the maturation of PV⁺ inhibitory cells, as well as processes such as myelination, mark the end of the critical period of human neurodevelopment [22]. Disruption or delay to the formation of the PNN results in the resumption or extension of the time window for neuroplasticity in the brain [23], wherein the nervous system is more sensitive to epigenetic, physical, biochemical, environmental, and nutritional factors. The effects of nutrition on individuals during gestational and early development have been extensively researched, leading many researchers to conclude that nutritional factors such as vitamins, folate and iodine can cause long-lasting impacts in neurodevelopment [24, 25]. As the foetus' and newborn's acquisition of vitamins like B₁₂ and D, depends to a great extent on maternal diet, such research has increasingly focussed on the impact of the mothers' vitamin deficiency on their offspring's brain development during the foetal and exclusive breastfeeding stages.

S-adenosyl methionine (SAM) is a universal methyl donor for some of the main methylation reactions. Vitamin B_{12} is an important cofactor in the one-carbon cycle

and is involved in the formation of SAM. Vitamin B_{12} supplements have been shown to improve pregnancy outcomes and reduce the risk of neurodevelopmental disorders in the developing child [26]. In rats, dose-dependent vitamin B_{12} supplementation was able to maintain the levels of docosahexaenoic acid (DHA) and BDNF in the hippocampus and cortex in pups at birth, and BDNF in the hippocampus at 3 months of age [17]. In addition, the combination of omega-3 fatty acid and vitamin B_{12} administration maintained spatial memory performance in neonates [17]. Experimental evidence suggests that DHA, together with greater levels of physical exercise, increases activated forms of CREB and synapsin I, reducing oxidative stress in the hippocampus [18].

Vitamin D deficiency may also reduce the integrity of PNNs and synaptic plasticity in neuropsychiatric disorders through the modulation of MMPs. Vitamin D deficiency has been associated with vulnerability to SZ [27], as well as ASD [28] and attention deficit and hyperactivity disorder (ADHD) [29], the two most common neurodevelopmental disorders. As mentioned earlier, ADAMTS and MMPs are two families of endogenous zinc-dependent proteases, secreted as inactive proenzymes that cleave ECM components. Alterations in the genes that encode MMP-16 and MMP-9 have been observed in patients with SZ [30]. High levels of MMP-9 can support the proteolytic cleavage of ECM with permissive synaptic plasticity but also lead to abnormal aggrecan degradation, abnormal development and neural excitability [30]. Chronic stress and neurological trauma can enhance MMP-9 levels in the brain [31, 32], and consequently raise the risk of SZ. A plausible proposal has been made that vitamin D deficiency leads to PNN degradation in patients with SZ [27]. In fact, vitamin D

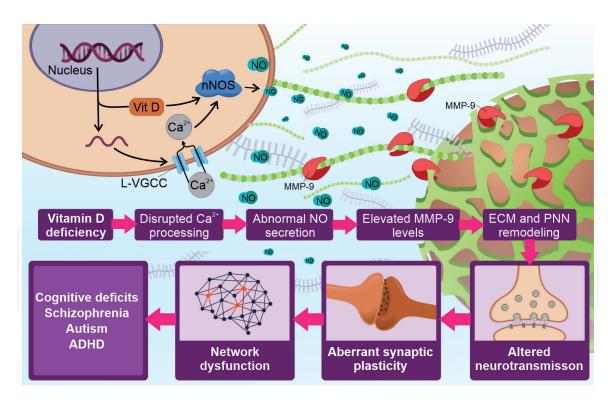


Figure 3.

Vitamin D deficiency and PNN formation during neurodevelopment. The figure above shows a neuron enveloped by a PNN. Vitamin D deficiency may induce a deficit in ECM organisation over the course of neurodevelopment, leading to the PNN loss and network-wide dysfunction in GABAergic, glutamatergic, and dopaminergic neurotransmission. Vitamin D deficiency is also linked to altered transcription of calcium channels (L-VGCC) potentially increasing the level of calcium input into the neuron, and to altered neuronal nitric oxide synthase (nNOS) activity, resulting in an increase of nitric oxide (NO) secretion into the extracellular space and elevated levels of MMP-9. This enhanced MMP-9 expression induces increased aggrecan synthesis, resulting in disruptions to the network of several neurotransmission systems important for normal cognitive function (spatial learning deficits), and SZ, autism and ADHD. Abbreviations: L-VGCC: L-type voltage-gated calcium channel; ECM: Extracellular matrix; MMP-9: Matrix metalloproteinase-9; nNOS: Neuronal nitric oxide synthase; NO: Nitric oxide; PNN: Perineuronal net; SZ: Schizophrenia; ADHD: Attention deficit and hyperactivity disorder (modified from [34]).

deficiency is associated with increased MMP-9 production [33] and calcium activity on the neuronal membrane, leading to increased nitric oxide (NO) formation and higher MMP-9 levels, and further appears to modulate its endogenous inhibitor TIMP1 (tissue inhibitor of MMP) [34]. Aggrecan-rich PNNs undergo restructuring leading to the occurrence of more synaptic anomalies and greater network dysfunction in GABAergic, glutamatergic, and dopaminergic neurotransmission, as evidenced by some forms of SZ (**Figure 3**) [34]. Cognitive deficits, such as spatial learning deficits, have been observed in adult mice with vitamin D deficiencies, with reduced density of PNNs and neural networks within the hippocampus [35].

2. Vitamin B₁₂ in neurodevelopment

Vitamin B_{12} is a member of the cobalamin family and can usually be obtained in sufficient quantities from meat, eggs, and dairy products. It is critical for the production of red blood cells, and the growth and maintenance of the nervous system.

Major causes of vitamin B_{12} deficiency include autoimmune pernicious anaemia, gastrectomy, ileal resection, pancreatic insufficiency, and malabsorption syndromes [36]. The human body is incapable of endogenous B_{12} production so it must be obtained from external sources in the individual's diet [37, 38]. Currently, a nutritional B_{12} deficiency is common in vegans, a fast-growing eating trend in many Western countries [39]. Vitamin B_{12} deficiency is also common in developing countries, with more widespread occurrence over more widespread sections of society beginning in early life and persisting throughout adulthood [40].

Pregnant women require a greater amount of vitamin B_{12} (2.5 g/day) compared to the general adult population (2.4 g/day), to adequately meet the nutritional needs of the foetus via absorption through the placenta [41]. The lack of sufficient vitamin B_{12} ingestion by the mother during pregnancy results in its deficiency in breast milk and the foetal bloodstream [42, 43]. Indeed, several studies have linked maternal deficiency in vitamin B_{12} concentration to developmental complications, including spontaneous abortion [44], low birth weight [45, 46], intrauterine growth restriction [45], and neural tube defects [47]. Lack of sufficient vitamin B_{12} in the mother's diet is reflected in similarly low concentrations of B_{12} in the bloodstream of breastfed babies during the period of exclusive breastfeeding, when the baby is dependent on breast milk for vitamin absorption, despite the relatively short window of this development period [48].

As in adults, vitamin B_{12} is mainly stored in the newborn's liver and is used on demand. However, as newborns have limited hepatic reserves, even if they are born at a healthy weight and size, symptoms resulting from vitamin B_{12} deficiency may appear from as early as 2 months of age [49]. Such symptoms imply a clinical pattern including abnormal pigmentation, hypotonia, liver and spleen enlargement, anorexia and growth failure associated with poor brain growth, which were first described by Jadhav and colleagues in 1962 [50].

Cellular deficiency of vitamin B_{12} results in slower proliferation and a faster differentiation of neuroblastoma cells [51], which point to its pivotal role in cell division and differentiation. In addition, deficiency of vitamin B_{12} in rodents is associated with selective brain damage, vascular and cognitive impairment [52], long-lasting functional disabilities in exploratory behaviour, learning and memory functions, and a mild decrease in hippocampal neurogenesis [53]. Moreover, vitamin B_{12} seems to play an important role in myelination as its deficiency leads to demyelination and may cause severe retardation of myelination during prenatal development [54].

Different mechanisms have been proposed to account for the effects of vitamin B_{12} deficiency on general neural function, most especially in relation to

neurodevelopment. The most well-understood mechanisms are related to the metabolic reactions in which vitamin B_{12} is the exclusive cofactor for mammalian cells [55], and the importance of the coenzyme forms of vitamin B_{12} , methylcobalamin (MeCbl) and adenosylcobalamin (AdoCbl) to the methionine and methyl groups that are used for DNA synthesis, epigenetic and cellular division.

When vitamin B_{12} is in MeCbl form it is a cofactor for methionine synthetase, which promotes the methylation of homocysteine (HCY) to methionine required in cases of a reduced methionine status of dietary ingestion. The synthesised form of methionine is subsequently condensed into SAM, which is finally demethylated into S-adenosylhomocysteine (SAH) and is a methyl donor for the conversion of phosphatidylethanolamine to phosphatidylcholine [56]. The altered SAM:SAH ratio may be the result of B_{12} deficiency as SAH and HCY levels increase while SAM levels decrease. This decreased SAM:SAH ratio may impair the methylation that is necessary for the synthesis of proteins, lipids, and neurotransmitters [57, 58] and leads to inhibition of DNA synthesis and cell division, since folate is not being recycled [59]. These results show that without methionine, the myelin and neurotransmitters considered essential for neurodevelopment cannot be produced. Methionine

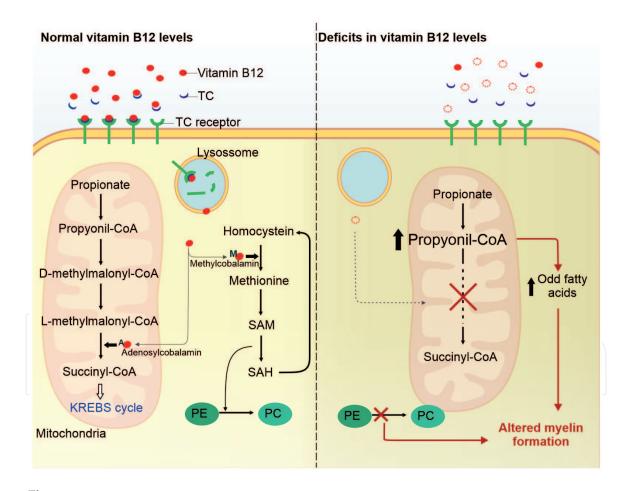


Figure 4.

Cellular processing of vitamin B in normal and deficiency conditions. Transcobalamin (TC) binds to vitamin B12 with high affinity and transports it by TC receptor-mediated endocytosis. The TC undergoes degradation into the lysosome, liberating the vitamin B12, that can be in two forms, methylcobalamin (MeCbl) and adenosylcobalamin (AdoCbl). When the vitamin B is in MeCbl form, it participates in HCY cascade as a cofactor, promoting the methylation of HCY to methionine that is subsequently condensed into SAM, which is finally demethylated into S-adenosylhomocysteine (SAH) and it a methyl donor for the conversion of phosphatidylethanolamine (PE) to phosphatidylcholine. The propionate (propionic acid) reacts with coenzyme A yielding propionyl CoA that is converted first to D-methylmalonyl-CoA and then to L-methylmalonyl-CoA and finally to Succinyl-CoA with the participation of AdoCbl form of vitamin B12 in this step. The deficiency of vitamin B12 leads to an inappropriate conversion of methylmalonyl CoA to succinyl CoA resulting in excessive production of the propionyl CoA, and consequently an increase of odd-chain fatty acid synthesis, and the inefficient conversion of PE to PC may impair

propionyl CoA production by the mitochondria, resulting in an altered myelin formation.

is synthesised from homocysteine, increased levels of which are observed in many neurodegenerative diseases, and which are functionally linked to brain injury and cognitive impairment [60].

When vitamin B_{12} is found in adenosylcobalamin (AdoCbl) form, it is required as the cofactor of the enzyme L-methylmalonyl-CoA mutase (EC 5.4.99.2) in the mitochondria and promotes the conversion of methylmalonyl CoA to succinyl CoA. This pathway is important for the mitochondria to reuse propionyl CoA and the energy obtained through the Krebs cycle. In other instances, this pathway may be recruited in response to increased levels of SAM in which the elimination of HCY is necessary [61]. The inappropriate conversion of methylmalonyl CoA to succinyl CoA leads to excessive production of the precursor propionyl CoA, resulting in odd-chain fatty acid synthesis and subsequent incorporation of these abnormal fatty acids into the nerve sheaths, which leads to altered myelin formation, reduced quantities of ethanolamine, phospholipids, and sphingomyelin (Figure 4) [56]. In addition, the inefficient conversion of phosphatidylethanolamine to phosphatidylcholine may impair propionyl CoA production by the mitochondria and the myelination, or lead to demyelination [62]. Myelination, together with synaptic refinement and the physiological maturation of inhibitory neural networks, are key processes of brain development that seem to coincide with the appearance of PNNs. In fact, alterations in these processes are well described in disorders such as SZ and ASD [63, 64].

3. Vitamin D and neurodevelopment

Vitamin D is known as the "neglected neurosteroid", a term proposed by McGrath and colleagues in 2001 [65], and exerts a great variety of effects during brain development.

In 2005, a study conducted the first description of the distribution of 1,25-dihydroxyvitamin D3 receptors (VDR) and 1α -hydroxylase (1α -OHase), the enzyme responsible for the formation of the active vitamin D, in the human brain. They are found both in neurons and glial cells and show a region- and layer-specific pattern of expression in the brain. VDR receptors are absent in the macrocellular cells within the nucleus basalis of Meynert (NBM) and Purkinje cells in the cerebellum, while both VDRs and 1α -OHase have been identified in the substantia nigra and the hypothalamus [66]. VDRs can also be found in the developing brain during the critical period of cell proliferation, in the temporal lobe, cingulate, thalamus, cerebellum, amygdala and hippocampus [67].

Similarly to vitamin B₁₂ deficiency, vitamin D deficiency during pregnancy appears to have serious consequences for foetal health. In the literature, abortions within the first trimester of gestation have been reported in association with the mothers' lack of sufficient vitamin D concentration [68–70]. Moreover, as foetus are dependent on access to the mother's supply of vitamin D, in conjunction with the large distribution of vitamin D receptors in the brain, there is a high probability that the vitamin D status of pregnant mothers may affect child neurocognitive development [67]. Among other sequelae, researchers have found cognitive and neural deficits in foetus and young infants due to maternal vitamin D deficiency during pregnancy, such as suboptimal neurocognitive development [71] and delays in the development of gross and fine motor function, problem-solving and communication [72]. However, a recent study analysed high-dosage vitamin D supplementation in the third trimester of pregnancy and its effects on child brain development and failed to report any improvements in the neurobiological mechanisms underlying developmental behaviour, such as motor milestones and cognitive and language development [73]. Few studies of the direct effects of vitamin D on

brain development have been conducted to date in humans and many of them are inconclusive, probably because of differences in the timing of vitamin D exposure, the types of assessment used, and the age at which the assessment occurred.

Studies in rats have identified similar critical periods to vitamin B_{12} during neurodevelopment in which maternal vitamin D deficiency can result in neurodevelopmental alterations, such as abnormal cell proliferation and decreased expression of neuronal structure genes in the brain of mice offspring [74]. VDRs are temporally regulated in the rat brain during development, with the first VDR expression occurring in the mesencephalon on day 12 [75]. VDR expression continues across different areas throughout the course of development [75, 76] and is directly correlated to the onset of natural cell elimination [77]. Because of its key role in regulating developmental processes throughout the brain, altered levels of vitamin D or VDR dysfunction can result in long-lasting behavioural disruption in animal models [67]. Consistent with its role in PNN formation, altered vitamin D levels and VDR signalling during development have been associated with neuropsychiatric disorders such as SZ [78] and autism [79, 80].

The mechanisms by which vitamin D influences brain development are diverse. There is evidence that vitamin D modulates neurotrophic factors such as NGF and glial cell line-derived neurotrophic factor (GDNF), suggesting that it may have a neuroprotective function [81, 82]. Other neuroprotective functions of vitamin D include the reduction of the neurotoxicity of glutamate and reactive oxygen species (ROS), the upregulation of antioxidant molecules [83, 84], and the suppression of macrophage activity, leading to decreased neuroinflammation activity [85]. When used as an immune suppressant, vitamin D has been reported to suppress the concentration of proinflammatory cytokines in the brain [86, 87], to reduce blood–brain barrier disruption and macrophage/microglia activation in induced autoimmune encephalomyelitis [88], and to attenuate proinflammatory processes and up-regulate anti-inflammatory processes such as the M2 microglia phenotype, in a mouse model of Parkinson's disease [89].

Vitamin D is known to reduce calcium levels in the brain, preventing cell death via excitotoxicity, and to downregulate or modulate L-type voltage-gated calcium channels (L-VGCCs) [90] (**Figure 5**). L-VGCCs are expressed in great quantity in the developing brain and have a critical role in synaptic plasticity and in regulating basal and burst firing activity in dopaminergic neurons within the ventral tegmental area [91]. Interestingly, disruption of L-VGCC function in hippocampal PV⁺ interneurons during development leads to significant morphological changes, such as reduced cell number and a decrease in dendritic arbour complexity [92, 93].

The association between vitamin D and L-VGCC function in the dopaminergic system and PV⁺ interneurons strengthen the hypothesis that vitamin D (dys) regulation plays a role in the onset of SZ [94]. Both epidemiological research and rodent models have shown a correlation between vitamin D deficiency and SZ. Alterations in the dopaminergic system have been frequently reported in response to vitamin D deficiency, consistent with increased VDR expression in brain areas primarily innervated by dopaminergic projections [95].

Associations have also been made between vitamin D receptors, calcium channels, PV⁺ interneurons and PNNs. L-VGCCs regulate PV⁺ expression and interneuron development [92] which are significantly disrupted in SZ. Curiously, the appearance of PNNs coincides with the synaptic refinement, myelination and maturation of inhibitory networks, and there is therefore strong evidence that PNNs have a pivotal role in the pathogenesis of SZ [63].

Following this same reasoning, vitamin D deficiency is considered a potential candidate for the development of several key alterations in ASD pathophysiology, most of them present mid to early development. John Cannell has been the main

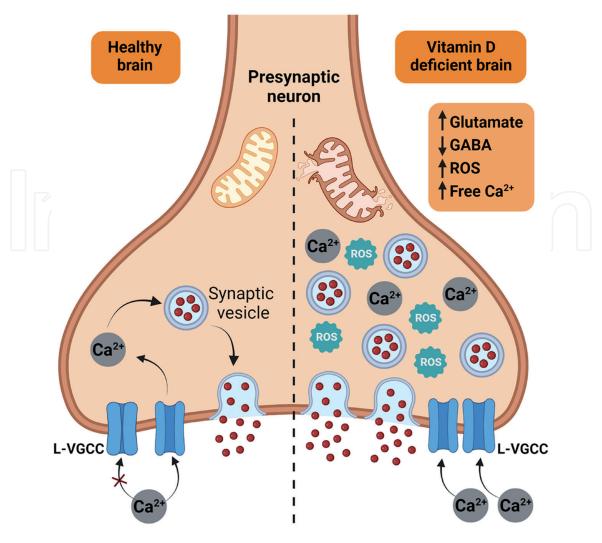


Figure 5.

Effects of vitamin D in presynaptic transmission Vitamin D plays an important role in several neurodevelopmental processes, such as neurotransmission and calcium signalling, preventing cell damage, especially mitochondrial dysfunction. As mentioned above, vitamin D acts by regulating L-VGCC in order that, for example, prevent the accumulation of intracellular calcium and the concomitant activation of pathways that can lead to cell death. With vitamin D deficiency, L-VGCC becomes functionality altered, leading to increased intracellular calcium, mitochondrial dysfunction, and the consequent generation of free radicals such as reactive oxygen species (ROS) and inhibitory/excitatory synaptic imbalance, illustrated by the increase in glutamate release and GABA neurotransmitter decrease, resulting in excitotoxicity and neuroinflammation (modified from [91]). Created with BioRender.

proponent of this hypothesis [96] and since its inception a number of studies have contributed to this line of research. The pathophysiology of ASD is multifactorial with solid evidence for both a genetic background and an environmental component. Some epidemiological findings have raised the additional hypothesis of specific genes which are environmentally responsive to the ASD, as epidemiological observations have pointed to changes in genotype expression. Cannell and others have cited neurosteroid pathway genes as good examples of environmentally responsive genes, since alterations in neurosteroid concentration may affect the genetic expression of the neural proteins regulated by steroids.

4. The link between vitamins B_{12} and D, PNNs and neurodevelopmental disorders: a mechanistic hypothesis

As already mentioned before, PNNs appear during postnatal development and surround cortical inhibitory GABA neurons expressing PV⁺, which control

the output of mainly cortical and hippocampal neurons and are necessary for fast rhythmic neuronal synchrony during information processing in cognitive tasks [97, 98]. PNNs enveloping PV⁺-inhibitory interneurons are known to be vital for cognition [99]. These cortical PV⁺-inhibitory interneurons express specific NMDA receptor (NMDAR) subunits such as NR2A, and are targets of the NMDA receptor antagonist MK-801. MK-801 is the main component for studying an important hypothesis for our understanding of the aetiology of SZ, the cortical hypofunction of NMDA receptors (i.e. the glutamatergic hypothesis) [100]. This hypothesis is compatible with the concurrent descriptions provided by the neurodevelopmental hypothesis, with respect to the disruptions to the central nervous system over the course of development [101].

An animal model using MK-801 treatment revealed a critical postnatal period, specifically from 7 to 14 days after birth (P7–14), resulting in SZ-like behaviour during adulthood, with a significant reduction in PV⁺-expressing cells in the PFC but not in the hippocampus, for mice treated with MK-801 from P7-14 compared to matched controls [102]. In addition, mice in the treated group showed changes in performance on cognitive, social and behavioural tasks, while electrophysiological recordings from brain slices within the PFC showed significantly reduced frequency of upstates in MK-801-treated mice, but increased gamma activity in MK-801-treated mice compared to saline-treated mice [102]. Recent memory reactivation has been shown to occur in the presence of slow oscillatory up-states, contributing to memory consolidation [103]. This electrophysiological profile is altered in humans with SZ, with increased delta oscillations and irregular gamma rhythm [104, 105]. Moreover, PNN removal from visual cortex during a critical postnatal period alters the balance between excitatory and inhibitory PV⁺ spiking activity, inducing greater potentiation of gamma activity and restoring the neural network to an immature or juvenile ECM state, suggesting that the maturation of GABAergic fast spiking and PV⁺-inhibitory neurons suppresses the spontaneous activity of excitatory neurons [106].

Together with these electrophysiological findings, the metabolic requirements of PNN function have also been explored in fast-spiking cells, as well as their intrinsic vulnerability. In fact, PNNs appear to promote interneuron maturation and network stability and may also protect neurons against iron sequestration and oxidative stress [107, 108]. Oxidative stress has been observed in the PFC of SZ patients in conjunction with decreased levels of glutathione (GSH), an endogenous antioxidant and redox regulator [109]. This outcome can in turn be aggravated by the overexpression of truncated DISC1 that is associated with SZ in humans, causing mitochondrial dysfunction with decreased mitochondrial NADH dehydrogenase activity, diminished cellular ATP contents, and overactivity of mitochondrial Ca2⁺ mechanisms [110].

As mentioned previously with regards to the role of GSH, there are two metabolically active forms of vitamin B₁₂, AdoCbl, and MeCbl: essential as a cofactor for the reaction necessary folate-dependent methylation of HCY by methionine synthase (MS) in the cytoplasm. As MS levels determine the ratio of methyl donor SAM to the endogenous methylation inhibitor SAH, the MeCbl reaction can influence SAM-dependent methylation reactions mainly through methylation of DNA and histones [111], an effect previously described with in relation to BDNF and epigenetic control. Zhang et al. 2016 analysed the postmortem human frontal cortex of autistic and schizophrenic individuals and 80-year-old individuals and found that the MeCbl form of vitamin B₁₂ decreases with age, as well as to age of onset of ASD and SZ, as compared to age-matched controls. Additionally, they also observed an abnormally lower total cobalamin Cbl and MeCbl concentration in ASD and SZ subjects, leading the authors to propose a "Redox/Methylation Hypothesis of Autism" in light of the impaired GSH-dependent synthesis observed in the brains of autistic individuals [111]. In the same study, the authors further found that certain

brain regions, as cerebellum and temporal cortex, in ASD, showed synthesis of reduced and oxidised glutathione (GSH/GSSG) (stable redox status), while other regions maintained SAM/SAH production (stable methylation status) [111], suggesting regional differences for metabolic disorders.

Concomitantly, several neurobiological changes found in ASD subjects or ASD animal models are consistent with an account of vitamin D deficiency. The main neurobiological processes mediated by vitamin D in neurodevelopment include neural cell proliferation [74], GABA, glutamate and serotonin neurotransmission [92], calcium signalling, mitochondrial regulation, oxidative stress and neuroinflammation (for review see [112]). Most of these processes are altered in ASD, resulting in the reduction of brain volume and changes in the number of glial and neuron cells, excitatory/inhibitory imbalance, disrupted calcium signalling, increased oxidative stress, the overactivation of microglia, and immune system dysregulation [113].

One of the best-studied hypotheses about the pathophysiology of ASD is the occurrence of changes in brain connectivity during development, which posits a reduced number of connections between distal brain regions and an increase in connections between proximal brain regions [114]. However, this hypothesis shows some inconsistencies in the light of several findings, such as reports of hyperconnectivity over long axonal fibres in autism or even mixed patterns of hypo- and hyper-connectivity [115–118].

Another hypothesis regarding the pathophysiology of ASD is one of excitatory-inhibitory imbalance, caused both by the probable hyperactivity of excitatory cells and a reduction in the number, activity or even delay of inhibitory interneurons in the maturation process [119–121].

PV⁺-expressing interneurons are the main regulators of inhibitory/excitatory balance and orchestrate the coordinative function of brain microcircuitry via their fast-spiking inhibitory inputs onto pyramidal neurons [97, 122]. As such, the healthy maturation of PV⁺-interneurons is crucial for the establishment of optimal neural and behavioural development. Disruption to PV⁺ expression in PFC caused by early-life adversity leads to altered social interactions [123] and anxiety-related behaviour in rodents [124, 125]. Both forms of altered behaviour can be found in patients diagnosed with ASD [126–128].

As mentioned previously, the maturation of PV^+ interneurons are mostly regulated by PNNs [129, 130]. Despite the protective actions of the PNN- PV^+ , the interneurons are very susceptible to oxidative stress [131, 132], and as such vitamin D deficiency represents a threat to the integrity of PNN function and consequently to the development of the GABAergic system.

Lastly, it is important to highlight the dysregulation of the immune system that is observed in ASD. ASD patients suffer from chronic systemic inflammation with a disbalance in cytokine expression, leading to increased production of proinflammatory cytokines. Vitamin D has been shown to contain immunomodulatory properties and may be an alternative treatment for slowing or minimising behavioural alterations in ASD (for review see [133]). Chronic systemic inflammation in early life may disrupt PV⁺ interneuron maturation and consequently lead to changes in PNNs. Taken together, these findings reveal the important role of vitamin D in maintaining the integrity of PNNs and the efficiency of synaptic transmission as a whole.

5. Conclusion

The ECM is one component of the tetrapartite synapse, together with the network of glial cells. The PNN is a dense ECM structure that enwraps inhibitory fast-spiking parvalbumin (PV⁺) interneurons, serving both as a protective barrier

and to regulate synaptic plasticity. The destruction of those PNNs during the critical postnatal period of brain development alters the balance between excitatory and inhibitory PV⁺ spiking activity, inducing a greater potentiation of gammaband activity, and reverting the firing pattern of the neural network to a so-called immature or juvenile ECM state. Studies have shown that a vitamin D deficiency in early development may lead to a reduction in the PNN integrity and synaptic plasticity through modulation of MMPs, contributing to increased risk of the onset of neuropsychiatric disorders, as SZ, ASD, and ADHD. Also, in preclinical studies, dose-dependent vitamin B₁₂ supplements were sufficient to maintain the levels of DHA and BDNF in the hippocampus and cortex in neonates, and BDNF levels in the hippocampus at 3 months of age, considered a sensitive window or critical period during neurodevelopment. In addition, the ingestion and metabolism of vitamin B₁₂ methylcobalamin (MeCbl) variants can influence S- adenosyl methionine and SAM-dependent methylation reactions, mainly through the methylation of DNA and histones, and the MeCbl deficiency can result in impairment of GSHdependent synthesis, inducing oxidative stress in the PFC of schizophrenic patients, or with autism diagnosis. Thus, adequate levels of vitamin B₁₂ and D appear to contribute to maintaining the integrity of PNNs, consequently lead to PV⁺ interneuron maturation.

Acknowledgements

The authors wish to thank IntechOpen for their support and payment of the Open Access Publishing Fee.

Conflict of interest

The authors declare no conflict of interest.

Author details

Marcela Bermudez Echeverry^{1,2*}, Silvia Honda Takada¹, Bruna Petrucelli Arruda¹, Debora Sterzeck Cardoso¹, Pamela Pinheiro Martins¹, Juliane Midori Ikebara¹, Aline V. Sousa-Santos¹ and Victor R.C. Torres da Silva¹

- 1 Universidade Federal do ABC (UFABC), Centro da Matemática, Computação e Cognição, São Bernardo do Campo, SP, Brazil
- 2 Neuroscience Laboratory, School of Medicine, University of Santander (UDES), Bucaramanga, Santander, Colombia

*Address all correspondence to: marcela.echeverry@ufabc.edu.br

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. CC BY

References

- [1] Tsien, R.Y., Very long-term memories may be stored in the pattern of holes in the perineuronal net. Proc Natl Acad Sci U S A, 2013. **110**(30): p. 12456-12461.
- [2] Engel, M., et al., Chondroitin sulfate proteoglycans in the developing central nervous system. I. cellular sites of synthesis of neurocan and phosphacan. J Comp Neurol, 1996. **366**(1): p. 34-43.
- [3] Domowicz, M.S., et al., *Aggrecan is expressed by embryonic brain glia and regulates astrocyte development.* Dev Biol, 2008. **315**(1): p. 114-124.
- [4] Bekku, Y., et al., Molecular cloning of Bral2, a novel brain-specific link protein, and immunohistochemical colocalization with brevican in perineuronal nets. Mol Cell Neurosci, 2003. **24**(1): p. 148-159.
- [5] Hirakawa, S., et al., *The brain link protein-1 (BRAL1): cDNA cloning, genomic structure, and characterization as a novel link protein expressed in adult brain.* Biochem Biophys Res Commun, 2000. **276**(3): p. 982-989.
- [6] Meyer-Puttlitz, B., et al., Chondroitin sulfate and chondroitin/keratan sulfate proteoglycans of nervous tissue: developmental changes of neurocan and phosphacan. J Neurochem, 1995. **65**(5): p. 2327-2337.
- [7] Ogawa, H., et al., Lp3/Hapln3, a novel link protein that co-localizes with versican and is coordinately up-regulated by platelet-derived growth factor in arterial smooth muscle cells. Matrix Biol, 2004. 23(5): p. 287-298.
- [8] Zimmermann, D.R. and M.T. Dours-Zimmermann, Extracellular matrix of the central nervous system: from neglect to challenge. Histochem Cell Biol, 2008. **130**(4): p. 635-653.
- [9] Flannery, C.R., *MMPs and ADAMTSs: functional studies*. Front Biosci, 2006. **11**: p. 544-569.

- [10] Kohno, T., et al., *Importance of Reelin C-terminal region in the development and maintenance of the postnatal cerebral cortex and its regulation by specific proteolysis.* J Neurosci, 2015. **35**(11): p. 4776-4787.
- [11] Ishii, K., K.I. Kubo, and K. Nakajima, *Reelin and Neuropsychiatric Disorders*. Front Cell Neurosci, 2016. **10**: p. 229.
- [12] Dityatev, A., M. Schachner, and P. Sonderegger, *The dual role of the extracellular matrix in synaptic plasticity and homeostasis*. Nat Rev Neurosci, 2010. **11**(11): p. 735-746.
- [13] Negron-Oyarzo, I., et al., Schizophrenia and reelin: a model based on prenatal stress to study epigenetics, brain development and behavior. Biol Res, 2016. **49**: p. 16.
- [14] Pizzarelli, R. and E. Cherubini, Alterations of GABAergic signaling in autism spectrum disorders. Neural Plast, 2011. **2011**: p. 297153.
- [15] Trevino, M., C. Vivar, and R. Gutierrez, Beta/gamma oscillatory activity in the CA3 hippocampal area is depressed by aberrant GABAergic transmission from the dentate gyrus after seizures. J Neurosci, 2007. 27(1): p. 251-259.
- [16] Dennis, K.E. and P. Levitt, Regional expression of brain derived neurotrophic factor (BDNF) is correlated with dynamic patterns of promoter methylation in the developing mouse forebrain. Brain Res Mol Brain Res, 2005. **140**(1-2): p. 1-9.
- [17] Rathod, R., et al., Maternal omega-3 fatty acid supplementation on vitamin B12 rich diet improves brain omega-3 fatty acids, neurotrophins and cognition in the Wistar rat offspring. Brain Dev, 2014. **36**(10): p. 853-863.

- [18] Wu, A., Z. Ying, and F. Gomez-Pinilla, *Docosahexaenoic acid dietary supplementation enhances the effects of exercise on synaptic plasticity and cognition*. Neuroscience, 2008. **155**(3): p. 751-759.
- [19] Cooper, J.A. and B.W. Howell, Lipoprotein receptors: signaling functions in the brain? Cell, 1999. **97**(6): p. 671-674.
- [20] Bock, H.H. and P. May, *Canonical and Non-canonical Reelin Signaling*. Front Cell Neurosci, 2016. **10**: p. 166.
- [21] Maixner, D.W. and H.R. Weng, *The Role of Glycogen Synthase Kinase 3 Beta in Neuroinflammation and Pain.* J Pharm Pharmacol (Los Angel), 2013. **1**(1): p. 001.
- [22] Takesian, A.E. and T.K. Hensch, *Balancing plasticity/stability across brain development*. Prog Brain Res, 2013. **207**: p. 3-34.
- [23] Beurdeley, M., et al., *Otx2 binding to perineuronal nets persistently regulates plasticity in the mature visual cortex*. J Neurosci, 2012. **32**(27): p. 9429-9437.
- [24] Colombo, J., K.M. Gustafson, and S.E. Carlson, *Critical and Sensitive Periods in Development and Nutrition*. Ann Nutr Metab, 2019. **75 Suppl 1**: p. 34-42.
- [25] Scott, J.A., The first 1000 days: A critical period of nutritional opportunity and vulnerability. Nutr Diet, 2020. 77(3): p. 295-297.
- [26] Pawlak, R., et al., How prevalent is vitamin B(12) deficiency among vegetarians? Nutr Rev, 2013. **71**(2): p. 110-117.
- [27] Chiang, M., R. Natarajan, and X. Fan, *Vitamin D in schizophrenia: a clinical review*. Evid Based Ment Health, 2016. **19**(1): p. 6-9.
- [28] Fernell, E., et al., Autism spectrum disorder and low vitamin D at birth: a

- *sibling control study.* Mol Autism, 2015. **6**: p. 3.
- [29] Sharif, M.R., et al., *The Relationship between Serum Vitamin D Level and Attention Deficit Hyperactivity Disorder.* Iran J Child Neurol, 2015. **9**(4): p. 48-53.
- [30] Wen, T.H., et al., Genetic Reduction of Matrix Metalloproteinase-9 Promotes Formation of Perineuronal Nets Around Parvalbumin-Expressing Interneurons and Normalizes Auditory Cortex Responses in Developing Fmr1 Knock-Out Mice. Cereb Cortex, 2018. 28(11): p. 3951-3964.
- [31] Meyer-Lindenberg, A. and H. Tost, *Neural mechanisms of social risk for psychiatric disorders.* Nat Neurosci, 2012. **15**(5): p. 663-668.
- [32] Zhang, H., et al., *Matrix* metalloproteinases and neurotrauma: evolving roles in injury and reparative processes. Neuroscientist, 2010. **16**(2): p. 156-170.
- [33] Kim, S.H., et al., Vitamin D Inhibits Expression and Activity of Matrix Metalloproteinase in Human Lung Fibroblasts (HFL-1) Cells. Tuberc Respir Dis (Seoul), 2014. 77(2): p. 73-80.
- [34] Mayne, P.E. and T.H.J. Burne, Vitamin D in Synaptic Plasticity, Cognitive Function, and Neuropsychiatric Illness. Trends Neurosci, 2019. **42**(4): p. 293-306.
- [35] Al-Amin, M.M., et al., Adult vitamin D deficiency disrupts hippocampal-dependent learning and structural brain connectivity in BALB/c mice. Brain Struct Funct, 2019. **224**(3): p. 1315-1329.
- [36] Watanabe, F., et al., Biologically active vitamin B12 compounds in foods for preventing deficiency among vegetarians and elderly subjects. J Agric Food Chem, 2013. **61**(28): p. 6769-6775.
- [37] Anonymous, Dietary reference values for food energy and nutrients for the

- United Kingdom. Report of the Panel on Dietary Reference Values of the Committee on Medical Aspects of Food Policy. Rep Health Soc Subj (Lond), 1991. 41: p. 1-210.
- [38] FAO/WHO, Evaluation of certain veterinary drug residues in food. Thirty-second report of the Joint FAO/WHO Expert Committee on Food Additives.
 World Health Organ Tech Rep Ser, 1988.
 763: p. 1-40.
- [39] Simcikas, S.J.A.C.E., Is the percentage of vegetarians and vegans in the US increasing. 2018.
- [40] Allen, L.H., *Causes of vitamin B12 and folate deficiency.* Food Nutr Bull, 2008. **29**(2 Suppl): p. S20-34; discussion S35-7.
- [41] Graber, S.E., et al., *Placental transport of vitamin B12 in the pregnant rat.* J Clin Invest, 1971. **50**(5): p. 1000-1004.
- [42] Baker, S.J. and H.J. Sinn, *Megaloblastic anaemia of infancy; report of a case*. Med J Aust, 1952. **1**(22): p. 750-752.
- [43] Deegan, K.L., et al., Breast milk vitamin B-12 concentrations in Guatemalan women are correlated with maternal but not infant vitamin B-12 status at 12 months postpartum. J Nutr, 2012. **142**(1): p. 112-116.
- [44] Kharb, S., et al., Prospective study on role of folic acid and vitamin B12 in early pregnancy and spontaneous abortion. 2018. **2**(4): p. 265-268.
- [45] Muthayya, S., et al., Low maternal vitamin B12 status is associated with intrauterine growth retardation in urban South Indians. Eur J Clin Nutr, 2006. **60**(6): p. 791-801.
- [46] Yajnik, C.S., et al., *Maternal total homocysteine concentration and neonatal size in India*. Asia Pac J Clin Nutr, 2005. **14**(2): p. 179-181.

- [47] Molloy, A.M., et al., Maternal vitamin B12 status and risk of neural tube defects in a population with high neural tube defect prevalence and no folic Acid fortification. Pediatrics, 2009. **123**(3): p. 917-923.
- [48] Rosenblatt, D.S. and V.M. Whitehead, *Cobalamin and folate deficiency: acquired and hereditary disorders in children*. Semin Hematol, 1999. **36**(1): p. 19-34.
- [49] Michaud, J.L., et al., *Nutritional* vitamin B12 deficiency: two cases detected by routine newborn urinary screening. Eur J Pediatr, 1992. **151**(3): p. 218-220.
- [50] Jadhav, M., et al., *Vitamin B12 deficiency in Indian infants. A clinical syndrome.* Lancet, 1962. **2**(7262): p. 903-907.
- [51] Battaglia-Hsu, S.F., et al., Vitamin B12 deficiency reduces proliferation and promotes differentiation of neuroblastoma cells and up-regulates PP2A, proNGF, and TACE. Proc Natl Acad Sci U S A, 2009. **106**(51): p. 21930-21935.
- [52] Troen, A.M., et al., *B-vitamin* deficiency causes hyperhomocysteinemia and vascular cognitive impairment in mice. Proc Natl Acad Sci U S A, 2008. **105**(34): p. 12474-12479.
- [53] Daval, J.L., S. Blaise, and J.L. Gueant, *Vitamin B deficiency causes neural cell loss and cognitive impairment in the developing rat.* Proc Natl Acad Sci U S A, 2009. **106**(1): p. E1; author reply E2.
- [54] Lovblad, K., et al., Retardation of myelination due to dietary vitamin B12 deficiency: cranial MRI findings. Pediatr Radiol, 1997. **27**(2): p. 155-158.
- [55] Pons, L., et al., Transcytosis and coenzymatic conversion of [(57)Co] cobalamin bound to either endogenous transcobalamin II or exogenous intrinsic factor in caco-2 cells. Cell Physiol Biochem, 2000. **10**(3): p. 135-148.

- [56] Stollhoff, K. and F.J. Schulte, *Vitamin B12 and brain development*. Eur J Pediatr, 1987. **146**(2): p. 201-205.
- [57] Briddon, A., Homocysteine in the context of cobalamin metabolism and deficiency states. Amino Acids, 2003. **24**(1-2): p. 1-12.
- [58] Weir, D.G., et al., Correlation of the ratio of S-adenosyl-L-methionine to S-adenosyl-L-homocysteine in the brain and cerebrospinal fluid of the pig: implications for the determination of this methylation ratio in human brain. Clin Sci (Lond), 1992. **82**(1): p. 93-97.
- [59] Guerra-Shinohara, E.M., et al., Low ratio of S-adenosylmethionine to S-adenosylhomocysteine is associated with vitamin deficiency in Brazilian pregnant women and newborns. Am J Clin Nutr, 2004. **80**(5): p. 1312-1321.
- [60] Lipton, S.A., et al., Neurotoxicity associated with dual actions of homocysteine at the N-methyl-D-aspartate receptor. Proc Natl Acad Sci U S A, 1997. **94**(11): p. 5923-5928.
- [61] Hoffer, L.J., *Homocysteine remethylation and trans-sulfuration*. Metabolism, 2004. **53**(11): p. 1480-1483.
- [62] Grattan-Smith, P.J., et al., *The* neurological syndrome of infantile cobalamin deficiency: developmental regression and involuntary movements. Mov Disord, 1997. **12**(1): p. 39-46.
- [63] Bitanihirwe, B.K. and T.U. Woo, *Perineuronal nets and schizophrenia: the importance of neuronal coatings.* Neurosci Biobehav Rev, 2014. **45**: p. 85-99.
- [64] Singh, V.K., et al., Antibodies to myelin basic protein in children with autistic behavior. Brain Behav Immun, 1993. 7(1): p. 97-103.
- [65] McGrath, J.J., et al., *Vitamin D* insufficiency in south-east Queensland. Med J Aust, 2001. **174**(3): p. 150-151.

- [66] Eyles, D.W., et al., Distribution of the vitamin D receptor and 1 alphahydroxylase in human brain. J Chem Neuroanat, 2005. **29**(1): p. 21-30.
- [67] Eyles, D.W., T.H. Burne, and J.J. McGrath, Vitamin D, effects on brain development, adult brain function and the links between low levels of vitamin D and neuropsychiatric disease. Front Neuroendocrinol, 2013. 34(1): p. 47-64.
- [68] Urrutia, R.P. and J.M. Thorp, *Vitamin D in pregnancy: current concepts.* Curr Opin Obstet Gynecol, 2012. **24**(2): p. 57-64.
- [69] Andersen, L.B., et al., Vitamin D depletion aggravates hypertension and target-organ damage. J Am Heart Assoc, 2015. **4**(2).
- [70] Li, N., et al., Women with recurrent spontaneous abortion have decreased 25(OH) vitamin D and VDR at the fetal-maternal interface. Braz J Med Biol Res, 2017. **50**(11): p. e6527.
- [71] Darling, A.L., et al., Association between maternal vitamin D status in pregnancy and neurodevelopmental outcomes in childhood: results from the Avon Longitudinal Study of Parents and Children (ALSPAC). Br J Nutr, 2017. 117(12): p. 1682-1692.
- [72] Dhamayanti, M., et al., Association of maternal vitamin D deficiency and infants' neurodevelopmental status: A cohort study on vitamin D and its impact during pregnancy and childhood in Indonesia. J Paediatr Child Health, 2020. 56(1): p. 16-21.
- [73] Sass, L., et al., High-Dose Vitamin D Supplementation in Pregnancy and Neurodevelopment in Childhood: A Prespecified Secondary Analysis of a Randomized Clinical Trial. JAMA Netw Open, 2020. **3**(12): p. e2026018.
- [74] Feron, F., et al., Developmental Vitamin D3 deficiency alters the adult rat

- *brain*. Brain Res Bull, 2005. **65**(2): p. 141-148.
- [75] Veenstra, T.D., et al., Zinc-induced conformational changes in the DNA-binding domain of the vitamin D receptor determined by electrospray ionization mass spectrometry. J Am Soc Mass Spectrom, 1998. **9**(1): p. 8-14.
- [76] Cui, X., et al., Maternal vitamin D depletion alters neurogenesis in the developing rat brain. Int J Dev Neurosci, 2007. **25**(4): p. 227-232.
- [77] Ko, P., et al., Maternal vitamin D3 deprivation and the regulation of apoptosis and cell cycle during rat brain development. Brain Res Dev Brain Res, 2004. **153**(1): p. 61-68.
- [78] Schoenrock, S.A. and L.M. Tarantino, *Developmental vitamin D deficiency and schizophrenia: the role of animal models*. Genes Brain Behav, 2016. **15**(1): p. 45-61.
- [79] Currenti, S.A., *Understanding and determining the etiology of autism.* Cell Mol Neurobiol, 2010. **30**(2): p. 161-171.
- [80] Schmidt, R.J., et al., Selected vitamin D metabolic gene variants and risk for autism spectrum disorder in the CHARGE Study. Early Hum Dev, 2015. **91**(8): p. 483-489.
- [81] Kalueff, A.V. and P. Tuohimaa, Neurosteroid hormone vitamin D and its utility in clinical nutrition. Curr Opin Clin Nutr Metab Care, 2007. **10**(1): p. 12-19.
- [82] Brown, J., et al., 1,25-dihydroxyvitamin D3 induces nerve growth factor, promotes neurite outgrowth and inhibits mitosis in embryonic rat hippocampal neurons. Neurosci Lett, 2003. 343(2): p. 139-143.
- [83] Garcion, E., et al., 1,25-dihydroxyvitamin D3 regulates the synthesis of gamma-glutamyl

- transpeptidase and glutathione levels in rat primary astrocytes. J Neurochem, 1999. **73**(2): p. 859-866.
- [84] Ibi, M., et al., Protective effects of 1 alpha,25-(OH)(2)D(3) against the neurotoxicity of glutamate and reactive oxygen species in mesencephalic culture. Neuropharmacology, 2001. **40**(6): p. 761-771.
- [85] Garcion, E., et al., Expression of inducible nitric oxide synthase during rat brain inflammation: regulation by 1,25-dihydroxyvitamin D3. Glia, 1998. **22**(3): p. 282-294.
- [86] Moore, D.D., et al., International Union of Pharmacology. LXII. The NR1H and NR1I receptors: constitutive androstane receptor, pregnene X receptor, farnesoid X receptor alpha, farnesoid X receptor beta, liver X receptor alpha, liver X receptor beta, and vitamin D receptor. Pharmacol Rev, 2006. 58(4): p. 742-759.
- [87] van Etten, E. and C. Mathieu, *Immunoregulation by* 1,25-dihydroxyvitamin D3: basic concepts. J Steroid Biochem Mol Biol, 2005. 97(1-2): p. 93-101.
- [88] de Oliveira, L.R.C., et al., Calcitriol Prevents Neuroinflammation and Reduces Blood-Brain Barrier Disruption and Local Macrophage/Microglia Activation. Front Pharmacol, 2020. **11**: p. 161.
- [89] Cianciulli, A., et al., Folic Acid Is Able to Polarize the Inflammatory Response in LPS Activated Microglia by Regulating Multiple Signaling Pathways. Mediators Inflamm, 2016. **2016**: p. 5240127.
- [90] Zanatta, L., et al., 1alpha,25-dihydroxyvitamin D(3) mechanism of action: modulation of L-type calcium channels leading to calcium uptake and intermediate filament phosphorylation in cerebral cortex of young rats. Biochim Biophys Acta, 2012. **1823**(10): p. 1708-1719.

- [91] Liu, Y., et al., Cav1.2 and Cav1.3 L-type calcium channels regulate dopaminergic firing activity in the mouse ventral tegmental area. J Neurophysiol, 2014. **112**(5): p. 1119-1130.
- [92] Jiang, M. and J.W. Swann, A role for L-type calcium channels in the maturation of parvalbumin-containing hippocampal interneurons. Neuroscience, 2005. **135**(3): p. 839-850.
- [93] Kasatkina, L.A., et al., Vitamin D deficiency induces the excitation/inhibition brain imbalance and the proinflammatory shift. Int J Biochem Cell Biol, 2020. **119**: p. 105665.
- [94] Chung, G.E., et al., *The serum* vitamin D level is inversely correlated with nonalcoholic fatty liver disease. Clin Mol Hepatol, 2016. **22**(1): p. 146-151.
- [95] Cui, X., et al., The vitamin D receptor in dopamine neurons; its presence in human substantia nigra and its ontogenesis in rat midbrain. Neuroscience, 2013. **236**: p. 77-87.
- [96] Cannell, J.J., *Autism and vitamin D.* Med Hypotheses, 2008. **70**(4): p. 750-759.
- [97] Sohal, V.S., et al., *Parvalbumin* neurons and gamma rhythms enhance cortical circuit performance. Nature, 2009. **459**(7247): p. 698-702.
- [98] Whittington, M.A., et al., *Multiple origins of the cortical gamma rhythm*. Dev Neurobiol, 2011. **71**(1): p. 92-106.
- [99] Murray, A.J., et al., *Parvalbumin-positive interneurons of the prefrontal cortex support working memory and cognitive flexibility*. Sci Rep, 2015. 5: p. 16778.
- [100] Nakazawa, K. and K. Sapkota, *The origin of NMDA receptor hypofunction in schizophrenia*. Pharmacol Ther, 2020. **205**: p. 107426.

- [101] Owen, M.J., et al., *Neurodevelopmental hypothesis of schizophrenia*. Br J Psychiatry, 2011. **198**(3): p. 173-175.
- [102] Plataki, M.E., et al., Effect of Neonatal Treatment With the NMDA Receptor Antagonist, MK-801, During Different Temporal Windows of Postnatal Period in Adult Prefrontal Cortical and Hippocampal Function. Front Behav Neurosci, 2021. 15: p. 689193.
- [103] Tatsuno, M., et al., Memory reactivation in rat medial prefrontal cortex occurs in a subtype of cortical UP state during slow-wave sleep. Philos Trans R Soc Lond B Biol Sci, 2020. 375(1799): p. 20190227.
- [104] Narayanan, B., et al., Multivariate genetic determinants of EEG oscillations in schizophrenia and psychotic bipolar disorder from the BSNIP study. Transl Psychiatry, 2015. 5: p. e588.
- [105] Uhlhaas, P.J. and W. Singer, Oscillations and neuronal dynamics in schizophrenia: the search for basic symptoms and translational opportunities. Biol Psychiatry, 2015. 77(12): p. 1001-1009.
- [106] Lensjo, K.K., et al., Removal of Perineuronal Nets Unlocks Juvenile Plasticity Through Network Mechanisms of Decreased Inhibition and Increased Gamma Activity. J Neurosci, 2017. 37(5): p. 1269-1283.
- [107] Morawski, M., et al., *Perineuronal nets potentially protect against oxidative stress*. Exp Neurol, 2004. **188**(2): p. 309-315.
- [108] Suttkus, A., et al., Neuroprotection against iron-induced cell death by perineuronal nets an in vivo analysis of oxidative stress. Am J Neurodegener Dis, 2012. **1**(2): p. 122-129.
- [109] Do, K.Q., et al., Schizophrenia: glutathione deficit in cerebrospinal fluid

- *and prefrontal cortex in vivo*. Eur J Neurosci, 2000. **12**(10): p. 3721-3728.
- [110] Park, Y.U., et al., Disrupted-in-schizophrenia 1 (DISC1) plays essential roles in mitochondria in collaboration with Mitofilin. Proc Natl Acad Sci U S A, 2010. **107**(41): p. 17785-17790.
- [111] Zhang, Y., et al., *Decreased Brain Levels of Vitamin B12 in Aging, Autism and Schizophrenia*. PLoS One, 2016. **11**(1): p. e0146797.
- [112] Berridge, M.J., Vitamin D, reactive oxygen species and calcium signalling in ageing and disease. Philos Trans R Soc Lond B Biol Sci, 2016. **371**(1700).
- [113] Siracusano, M., et al., *Vitamin D Deficiency and Autism Spectrum Disorder*. Curr Pharm Des, 2020. **26**(21): p. 2460-2474.
- [114] Belmonte, M.K., et al., *Autism and abnormal development of brain connectivity*. J Neurosci, 2004. **24**(42): p. 9228-9231.
- [115] Courchesne, E. and K. Pierce, *Brain overgrowth in autism during a critical time in development: implications for frontal pyramidal neuron and interneuron development and connectivity.* Int J Dev Neurosci, 2005. **23**(2-3): p. 153-170.
- [116] Maximo, J.O., E.J. Cadena, and R.K. Kana, *The implications of brain connectivity in the neuropsychology of autism.* Neuropsychol Rev, 2014. **24**(1): p. 16-31.
- [117] Mizuno, A., et al., *Partially* enhanced thalamocortical functional connectivity in autism. Brain Res, 2006. **1104**(1): p. 160-174.
- [118] Shih, P., et al., Atypical network connectivity for imitation in autism spectrum disorder. Neuropsychologia, 2010. **48**(10): p. 2931-2939.
- [119] Markicevic, M., et al., *Cortical Excitation:Inhibition Imbalance Causes*

- Abnormal Brain Network Dynamics as Observed in Neurodevelopmental Disorders. Cereb Cortex, 2020. **30**(9): p. 4922-4937.
- [120] Lacaille, H., et al., *Impaired Interneuron Development in a Novel Model of Neonatal Brain Injury.* eNeuro, 2019. **6**(1).
- [121] Trakoshis, S., et al., *Intrinsic* excitation-inhibition imbalance affects medial prefrontal cortex differently in autistic men versus women. Elife, 2020. **9**.
- [122] Ferguson, B.R. and W.J. Gao, PV Interneurons: Critical Regulators of E/I Balance for Prefrontal Cortex-Dependent Behavior and Psychiatric Disorders. Front Neural Circuits, 2018. 12: p. 37.
- [123] Holland, F.H., et al., Early life stress disrupts social behavior and prefrontal cortex parvalbumin interneurons at an earlier time-point in females than in males. Neurosci Lett, 2014. **566**: p. 131-136.
- [124] Lee, M.J., et al., Investigation of anxiety and depressive disorders and psychiatric medication use before and after cancer diagnosis. Psychooncology, 2021. **30**(6): p. 919-927.
- [125] Page, C.E., et al., Prefrontal parvalbumin cells are sensitive to stress and mediate anxiety-related behaviors in female mice. Sci Rep, 2019. **9**(1): p. 19772.
- [126] Gillis, R.F. and G.A. Rouleau, *The ongoing dissection of the genetic architecture of autistic spectrum disorder*. Mol Autism, 2011. **2**(1): p. 12.
- [127] Coffey, B.J., et al., *Anxiety disorders and tic severity in juveniles with Tourette's disorder.* J Am Acad Child Adolesc Psychiatry, 2000. **39**(5): p. 562-568.
- [128] Howlin, K.J., et al., Evidence for electroneutral sodium chloride transport in rat proximal convoluted tubule. Am J

Vitamins D and B₁₂, Altered Synaptic Plasticity and Extracellular Matrix DOI: http://dx.doi.org/10.5772/intechopen.100055

Physiol, 1986. **250**(4 Pt 2): p. F644-F648.

[129] Hartig, W., K. Brauer, and G. Bruckner, Wisteria floribunda agglutinin-labelled nets surround parvalbumin-containing neurons. Neuroreport, 1992. 3(10): p. 869-872.

[130] Enwright, J.F., et al., Reduced Labeling of Parvalbumin Neurons and Perineuronal Nets in the Dorsolateral Prefrontal Cortex of Subjects with Schizophrenia. Neuropsychopharmacology, 2016. 41(9): p. 2206-2214.

[131] Cabungcal, J.H., et al., *Perineuronal* nets protect fast-spiking interneurons against oxidative stress. Proc Natl Acad Sci U S A, 2013. **110**(22): p. 9130-9135.

[132] Brenhouse, H.C. and J.M. Schwarz, *Immunoadolescence: Neuroimmune development and adolescent behavior.* Neurosci Biobehav Rev, 2016. **70**: p. 288-299.

[133] Vegelin, M., G. Teodorowicz, and H. Savelkoul, *Vitamin D and Autism Spectrum Disorder*. 2021.

