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Chapter

Interactive Effect of Organic and Inorganic Amendments along with Plant Growth Promoting Rhizobacteria on Ameliorating Salinity Stress in *Maize*

Sajid Rashid Ahmad, Sana Ashraf and Humaira Nawaz

Abstract

Saline soil is one of the common environmental issues that negatively affects the soil quality of agricultural lands. It reduces the plant growth and productivity worldwide. Soil Salinity and sodicity affecting land about 1128 million hectares globally determined by recent researches. The most important salt-sensitive cereal crops in the world are Maize (*Zea mays* L.) For food security, its need of hour to securing attainable production of maize crop in the salt affected soils. To reduce negative impacts of saline soil on plant growth, sustainable approaches such as organic amendments like press mud and inorganic amendments like silicon can be applied. For increasing crop productivity, plant growth promoting rhizobacteria (PGPR) which are salt-tolerant in saline agriculture can also be applied. In this book chapter interactive effect of different organic and inorganic amendments and plant growth-promoting rhizobacteria to reduce salinity stress on maize has been discussed.

Keywords: Salinity stress, Maize, Food security, Organic amendments, Inorganic amendments

1. Introduction

In the Arid and semi-arid areas salt affected soil poses immense threats to the agriculture industry worldwide [1]. Researchers have reported that about 1128 million ha of land is affected by salinity and sodicity globally [2]. Soil salinization at global level has caused food insecurity in several countries during last decade. In Pakistan approximately 6.8 million hectares of land is affected by salinity [3]. Saline soil is characterized by the presence of high level of sodium and its chlorides and sulphates [4]. Soil having 4 dS m⁻¹ or more electrical conductivity of the saturation soil paste is considered as salt affected soil [5–7]. Due to high concentrations of Na⁺ and Cl⁻ in the plant, Sodium chloride can reduce crop productivity by making the roots water uptake more difficult that can cause plant toxicity [8]. Research studies proved that approximately 20% of the world's cultivated land is affected by salinity [9].

Due to inappropriate management of irrigation and drainage, soil salinity is gradually increasing in irrigated lands [10]. Global warming as an environmental

issue has greatly affected arid regions of the world that are at highest risk of soil salinization. Salt redistribution in the soil profile is due to climatic factor such as precipitation. Agricultural productivity is affected when salt is added by wind to coastal agricultural lands [11]. Soil Biodiversity and microbial activity is affected by the high salt concentration [12].

Under saline soil plant growth is negatively affected by osmotic effects and hormonal imbalance. It also causes nutritional disorder and specific ion toxicity [13]. The adverse effects of saline soil on plants include: (1) Osmotic potential is decreases due to excessive soluble salts in the soil solutions. It also causes physiological drought by decreasing plants ability to absorb water (2) toxicity due to salt ions inside the plant cells. The Growth inhibition is caused by sodium and chloride ions as sodium ions are retained in the roots and stems and only chloride ions become concentrated in the shoot in some plants which is causing negative affects to the plants [14, 15]. (3) Secondary stresses which is mainly caused by osmotic and ionic pressure. It includes high concentration of toxic compounds such as ROS and nutrient imbalance in plants. Sodium ions compete with potassium ions under saline condition and causing reproductive disorders by calcium ions in the cell membrane [16, 17]. The maize (*Zea mays* L). is a major food crop in the world food. The productivity of maize crop is declined as it moderately salt-sensitive plant [18].

In this scenario, it is the need of time that agronomists and environmentalists should develop eco-friendly, cost effective and sustainable methods to reclaim saline soils [19]. Currently, various physico-chemical processes are in practice for the reclamation of saline soils. To some extent these methods are unsustainable and inefficient at high salt concentration [20]. The traditional breeding and biotechnological methods for the production of salinity-tolerant crops is a time-consuming process. By using chemical neutralizers and sustainable approaches sustainable crop yield in saline soils must be secured. It can also be secured by using salt-tolerant varieties or amelioration methods.

For plant growth and development, microorganisms play an important role under different environmental conditions [21, 22]. For enhancing crop productivity in saline soil, the application of plant growth-promoting rhizobacteria has become sustainable approach [23, 24]. Inoculation with PGPR leads towards abiotic stress regulation which can cause systemic tolerance directly or indirectly [25]. Many PGPR have been applied for their positive role in improving plant-water relations and for ion homeostasis. It is also used for photosynthetic efficiency in plants under salt stress. Plants can effectively protect from many stresses by PGPR that produce IAA and ACC deaminase. IAA accumulation increase transcription of ACC synthase genes. It is resulted an increases ACC concentration that can lead to the production of ethylene. Excess ACC are broken by PGPR that produce ACC deaminase. It also decreases plant ethylene levels under harsh environmental conditions. It permits IAA to encourage the growth of the plants [26].

Bacteria secrete exopolysaccharides which can bind soil particles into aggregates. These are helpful in regulating soil structures. It also increases water holding as well as cation exchange capacity of soil [27]. An enclosed matrix of microcolonies is formed by EPS which provide protection against environmental changings. It also leads towards water as well as nutrient retention and epiphytic colonization [28]. The exopolysaccharide secretion by PGPR binds sodium ions and reduces its uptake in plants which is determined by researches studies [29].

In saline soil a diversity of salt-tolerant PGPR such as *Azospirillum*, *Burkholderia*, *Rhizobium*, *Pseudomonas*, *Acetobacter* and *Bacillus* have been applied. These are also tested for promoting plant growth under salt stress [30, 31]. Thus, it has been demonstrating by different researches that use of PGPR is a beneficial approach to increase plant performance in saline soil [32, 33]. The physiological drought is

caused by salt in soil environment which reduces the ability of plants to remove water. Many biotic and physical stresses on plants can be reduced by application of Si fertilizer which can change the negative effects of saline soil [34, 35]. By improving sodium ions and potassium ions homeostasis, silicon may increase salinity tolerance in plants. It also improves nutritional status and photosynthetic efficiency of plants under stress conditions [36–38]. Many laboratory and greenhouse experiments have determined that under saline conditions, Si reduced the uptake of sodium ions and chloride ions [39, 40]. The use of organic matter increases the physico-chemical and biological properties of salt-affected soils [41]. Organic matter also plays an important role by improving roots to grow more uniformly. The soil CO₂ concentration is increased by decaying organic matter. It also releases H⁺ and enhances CaCO₃ dissolution. It can release more calcium for sodium exchange [42] Application of press mud is very effective in reclaiming saline sodic soils [43, 44].

2. Impacts of soil salinity

2.1 Impact of soil salinity on plant growth and development

Saline soil affects plant growth, development and process of photosynthesis. It also affects protein synthesis and lipid metabolism [45]. Osmotic stress reduces photosynthetic efficiency which is resulted in partial closure of stomata [46]. The nutrient imbalance and membrane destabilization are caused by soil salinity [47]. The cell growth and development are decreased in plants in responses to osmotic stress. It resulted in decreased leaf area and chlorophyll content [48].

The nutritional imbalances are also caused by decrease in the uptake of calcium ions and potassium ions in leaves and an increase in the uptake of sodium ions. In some cases, there is a requirement of low sodium ions and high potassium ions or calcium ions are required for optimum function, but increased sodium ions resulted in metabolic disturbances. Cell swelling in plants is caused by accumulation of sodium and chloride which can affect plant enzymes. It can also result in physiological changes and reduced energy production [49]. The photosynthetic function is disturbed by nitrate reductase activity due to chloride ions [50]. There are competitive interactions with nutrient ions for binding sites. It can also affect transfer of protein in root cells under excessive sodium and chloride ions in rhizosphere. It also affects processes like movement of material, deposition, and partitioning within plants [51]. Salts can increase in intercellular spaces resulted in cell dehydration [52]. Oxidative stress increases due to the accumulation of reactive oxygen species which has negative impact on cell membranes, proteins, enzymes, and nucleic acids [53] Both antioxidant enzymes and non-enzymatic antioxidants are produced by plants to protect against oxidative stress [54].

2.2 Impacts of soil salinity on rhizosphere microbial diversity

Microbial biomass is an important parameter as it functions as an agent transformation and plays its role as the recycling of the organic matter by providing soil nutrients. In the first few centimeters of the soil surface, there are microbial biomass and organic matter. Microbiological activity is affected by the salinization process [55]. Microbial diversity, functions, and compositions are negatively affected by salinity [56]. Total bacteria and actinobacteria are reduced by a 5% increase in salinity. The attachment of *Azospirillum brasilense* to maize roots was observed to reduce due to salinity [57]. Due to increase salinity in the rhizosphere, the plant root secretion and organic matter decomposition by microorganisms are adversely affected [58].

3. Application of organic amendments to agricultural crops to mitigate salinity effects

3.1 Organic matter

There is an excess of salts in water which is used for irrigation purposes. It can reduce the crop yield due to its increased salt concentration [59]. Soil electrical conductivity is being increased due to the continuous increase of salts in it [60]. Water which is used for irrigation having excess salts in it resulted in negative impacts on plant physiology, soil water plant relationships, and limits the production of crops [61]. By application of organic manure in soil, the toxicity of salts can be minimized, and soil properties can be improved as cost-effective approaches [62].

There are agricultural practices that are used for the management of saltaffected soil [63]. Addition of organic martial is beneficial as a fertilizer which can modify and improve the soil characteristic. For recovery of saline soil, organic amendments like organic manure and compost are being tested as efficient methods [64]. Application of organic matter for the reclamation of sandy soil is an effective method to improve the physical properties of soil [65]. Researchers determined that poultry manure, farmyard manure (FYM), crop residues as compost are being used for the addition of nutrients in the soil. It is beneficial for improving plants' health. It can also modify physiochemical properties of plants [66]. Farmyard manure is the most commonly and easily available source of organic matter. There are different factors which can affect the efficiency of farmyard manure such as nature of feed consumed by the animal, type of animal and waste management methods [67].

3.2 Biochar

There are different long-term and short-term methods for reclamation of salt-affected soil, but short-term management approaches are useful as a management strategy that are cost effective and high-income generating methods [68]. The biochar is an effective method for organic amendment of salt-affected soil that results in

- Soil physicochemical and biological properties are improved
- Stomatal conductance and phytohormones can be regulated
- · Reduction in oxidative stress
- Increase in mineral nutrient uptake
- Effects on plant growth, photosynthesis and biomass
- Na ion toxicity in plants is reduced

4. Application of inorganic amendments to agricultural crops to mitigate salinity effects

4.1 Exogenous application of sulfur

Salt affected soil has many salts in it and each salt has a differential contribution to salt stress. There are different salts such as Na₂SO4, NaCl, Na2CO3, CaSO4, MgCl2, KCl but the most important of these is NaCl [69–73]. For the regulation of cell metabolism and hormone signaling pathway, Sulfur plays a very important role. For regulating seed germination its acts as a biochemical agent [74, 75]. For the

synthesis of protein, chlorophyll, vitamins, and glutathione which are helpful to tolerate various stresses, sulfur plays a very important role [76]. Sulfur compounds are also present in many amino acids and their composition changes by the application of sulfur [77]. To improve plant growth by improving its cellular functions especially in saline soil, the addition of sulfur is beneficial [78]. Different approaches are being applied to mitigate the deleterious effects of salinity on health of plants. The exogenous application of inorganic salts and osmo-protectants are cost efficient approach to reduce the negative effects of salt stress on plant growth [79, 80].

4.2 Use of silicon nutrition to alleviate the salt stress in maize

In contrast to Na⁺ and Cl⁻ toxicity, silicon (Si) has ameliorative features. It can help plants to grow on saline soil. For industrialized counties, it can prove costeffective. Under biotic stress, silicon can improve plant growth also reduces radiation effects on it. It is helpful in reducing water loss up to 30% [81]. The exogenous application of Si for different salt-tolerant plant species has been reported [82, 83]. Under saline environment, Si uptake by plats increases root activity and inhibits transpiration. But in the plasma membrane, it increases the activity of ATPase and PPase. This can result in decrease in Na uptake and an increase in K uptake [84, 85].

4.2.1 Silicon-mediated mechanisms underlying increased crops tolerance to salinity

Si application can directly influence growth of plants by diminishing the transport of Na⁺ ions while indirectly activating physiological processes under saline conditions.

4.2.1.1 Reduced Na⁺ uptake by plant roots due to Si application

Due to high concentration of Cl⁻ and Na⁺ and low concentration of K⁺ and Ca⁺² in the saline environment, Na⁺/K⁺ ratio vary in plants [84]. Due to elevated level of Na⁺ and overproduction of ROS, plant metabolism is being changed [85] Research studies demonstrated that Si can reduce ion toxicity which is resulted from the saline condition. It is also helpful in increasing K⁺ and decreasing Na⁺ uptake [86]. Thus, research studies determined that Si application resulted in reduced Na⁺ buildup in the roots [86]. Si as phytolith, accumulates different parts of plant bodies. Si deposits underneath cell walls of roots to bind the Na⁺ and reduces Na⁺ toxicity by decreasing the Na⁺ transport in upper regions and increasing the K⁺ uptake.

4.2.1.2 Stimulation of antioxidant defense system in crops

Under the saline conditions, studies have determined the enhanced production of antioxidant due to the application of Si [87]. Effects of Si on the antioxidants depend upon different factors like the severity of saline stress, time, plant species, and the concentration of Si. Thus, studies determined that application of Si can regulate antioxidant defense system by reducing salinity effects. This also resulted in decrease lipid peroxidation and regulate membrane integrity. It also can decrease permeability of plasma membrane. The research studies determine that non-Sitreated and Si-treated plants show different responses under saline conditions. Application of Si plays a protective role to improve antioxidant activity.

5. Role of PGPRs in alleviation of salinity stress in maize crop

In the semi-arid environment, salinity pose negative effects on the growth and production of various crops. It also affects aggregate stability of soil. Soil structure

stability has important for improvement of soil properties. The soil microbial communities such as free-living or symbiotic organisms play an immense role to improve soil structure. It is proved that the activities which microbes performed to soil aggregate stability are very advantageous [88]. It can efficient solution for saline soil and make it fit for agricultural practices. PGPRs can help in inducing plant tolerance to various abiotic stresses including salt stress. In saline environments, PGPR-crop interactions improved the plant growth. It can also promote plant survival in adverse conditions [89]. PGPR promote the growth and development of plants by providing nitrogen, phytohormones soluble phosphates, and iron [90]. The plant is being protected against various soil-borne diseases, and it is known that most of these diseases are caused by pathogenic fungi [91].

5.1 Various attributes of PGPsRs in mitigating negative effects of salt stress in maize crop

5.1.1 Enhanced root proliferation and plant vigor

PGPRs can promote the growth of the plants by means of PGPRs which colonize the rhizosphere [92]. The co-inoculation of seeds of different PGPR species is a beneficial strategy to remediate salt-stressed soil. This approach has improved the plant tolerance towards abiotic stresses and the structure of root hairs.

5.1.2 Phytohormones produced by bacteria

The physiological response in plants is increased by phytohormones produced by microbes in root zone. Production of indoleacetic acid and gibberellins promote the root length. It also increases number of tips, surface area of roots and uptake of nutrients thus promoting the plant vigor exposed to saline conditions [93–96]. Indole acetic acid production is a common characteristic of PGPR. This bacterium is observed to reduce salinity stress in plants.

5.1.3 Role of PGPR as a sink for 1-aminocyclopropane-1-carboxylate (ACC)

Increase in ACC levels can result in higher ethylene production under saline environment. It can also increase plant injuries [97, 98]. Cobalt ions and amino ethoxy vinyle glycine as chemical inhibitors of ethylene synthesis is often used to control salinity problems. These chemicals are expensive and have harmful effects on the environment. PGPR play a role of sink for ACC which can be hydrolyzed to generate a-ketobutyrate and ammonia to reduce the ethylene production.

5.1.4 PGPR-mediated ion homeostasis

Plants inoculated with PGPR have showed high concentration of K⁺ which led to high Na⁺/K⁺ ratio and ultimately improved tolerance towards salt stress [99–101]. Salinity can damage the cell-membrane in plants which can enhance its permeability and electrolyte leakage. In maize, Lower the electrolyte leakage has been determined the inoculation with Rhizobium [102–104].

5.1.5 Accumulation of osmolytes

The functioning of photosynthetic structures and maintaining water homeostasis are essential for reducing salinity impact on plants. Excessive production of various compatible organic solutes (such as glycine betaine and proline) has been

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observed as stress responses in plants [105]. Accumulation of proline is a physiological response of plants to saline conditions [106]. It also maintains high leaf water potential and protects the plants from negative effects of oxidative stress. Researchers have determined that PGPRs contribute to accumulation of osmolytes to increase plant tolerance towards stress.

5.1.6 Antioxidative enzymes

Reactive oxygen species (ROS) damage the nucleic acids, proteins and lipids. Limited photosynthetic activity under salinity promotes the excessive production of ROS [107]. Antioxidants have been found to greatly reduce the oxidative damage. Under saline conditions, the activities of the enzymatic antioxidants such as guaicol peroxidase, catalase and superoxide dismutase increased [108]. Researchers determined that the application of PGPR caused a significant increase in polyphenol oxidase, superoxide dismutase and other enzymes involved in plant defense system. It also increases in enzymes such as peroxidase, phenyl alanine ammonialyase, catalase, phenolics and lipoxygenase [109–111]. These PGPR-stimulated enzymes are playing important role in removing hydrogen peroxide from stressed roots [112].

5.1.7 Ameliorating effects of bacterial extracellular polymeric substances (EPS)

Researchers determined that inoculation with EPS-producing PGPR have significantly increased the volume of soil macropores, rhizospheric soil aggregation, improved fertilizer as well as water availability. This approach can help plants to survive in salt-stressed soils. Different studies have shown positive effects of EPS-producing PGPR on the rhizospheric soil aggregation [113]. As bacterial EPS can sequester the cations, there may be an opportunity to eliminate the salinity stress by increasing the EPS-producing PGPR strains [114].

5.1.8 Enhancement of plant nutrient uptake

It is obvious that PGPR can regulate the availability of plant nutrients. So, employing PGPR can cut down the use of chemical fertilizers. Various PGPR strains are involved in solubilizing the inorganic phosphate and mineralization of organic phosphate, thus providing nutrients to plants [115]. However, the former activity of PGPR is the key role of PGPR in providing nutrients to plants.

5.1.9 PGPR-mediated disease suppression

Many rhizobacteria are known to produce antifungal metabolites like phenazines, HCN, pyrrolnitrin, tensin, pyoluteorin, 2,4-diacetylphloroglucinol, and viscosinamide [116]. However, various PGPR strains can control the pathogen of plants grown under salt stress.

6. Interactive techniques to ameliorate salinity stress in maize

6.1 Silicon and PGPR to mitigate salt stress in maize

An environment-friendly and cost-effective approach for lessening salinity in crop plants is the co-application of silicon and PGPR [117]. Different studies have shown that by improving photosynthetic efficiency, and scavenging enzyme

activity soil salinity tolerance can be enhanced. It also determined that this approach can improve the plant tolerance towards salinity, ROS and Na^+/K^+ ratio [118]. PGPR promote the growth of plants via synthesis of phytohormones, exopolysaccharides, volatile organic compounds and different other mechanisms [118]. Recently, it has been found that both Si and PGPR can enhance plants tolerance to saline environment to improve growth and yield of plants [118].

6.2 Combined effects of biochar and plant growth-promoting bacterial endophytes on alleviating salt stress in maize

Employing the salt tolerant PGPR to enhance crop productivity has been a sustainable and efficient method [119-122]. Researchers have documented that PGPR produced the exopolysaccharide (EPSs) that prevent the uptake of Na + ions by sequestering these ions [123, 124]. Studies demonstrated that few PGPR have an important enzyme, ACC- deaminase, which can reduce ethylene production by metabolizing ACC into ammonia. ACC is the precursor of ethylene and a-ketobutyrate [125–127]. Unlike PGPR, plant growth-promoting bacterial endophytes colonize the internal tissues of plants without causing any harm to the plants [128]. It can lead to several physiological modifications that contribute to plant growth and development [129–131]. These, plant growth-promoting bacterial endophytes may promote plant growth by adopting the similar mechanisms as observed in PGPR [132]. Thus, it is proved that plant growth-promoting bacterial endophytes are more effective in promoting plant growth even under severe stresses as compared to PGPR. Different researchers have demonstrated that for reducing soil salinity addition of biochar along with endophytic bacteria is an efficient and environment friendly approach [133].

For enhancing crop growth and yield, use of biochar is cost effective and eco-friendly option to boost water and nutrient-holding capacity of soil [134–137]. Application of biochar has positive effects on physicochemical properties of soil. Moreover, Biochar can also improve a variety of soil microbes by providing them a favorable habitat and nourishment [138]. Thus, it is an excellent solution for recycling organic waste and solution to environmental pollution.

There are three important mechanisms underlying biochar-mediated reduction of salt stress in plants. These include:

- a. High adsorption of Na⁺ on biochar resulting in reduced availability of Na⁺ in soil solution
- b. Regulation of ions concentration in soil solution by liberating mineral nutrients
- c. Dilution of soil solution via increasing available moisture contents of soil to reduce the osmotic stress [139].

7. Conclusion

Reclamation of saline soils is mainly achieved by employing various physicochemical processes. However, these processes are not sustainable and considered inefficient in the case of high salt concentration. PGPR contain a vital enzyme, 1-aminocyclopropane-1-carboxylate deaminase that can decrease salinity induced ethylene production. Silicon and elemental sulfur can also be applied to reduce the negative effects of soil salinity on plants. The organic matter such as press mud usually contains about 70% lime, 15–20% organic matter and 23% sugar. This organic

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matter is highly soluble and readily available to the microbial activity and soil. Due to microbial activity more carbon dioxide is produced that may increase the solubility of lime and hence reclaim the saline soils. Hence, the combine application of organic amendments (like press mud), inorganic amendments (like silicon and elemental sulfur) and PGPR can ameliorate the saline soil in an environmentally sustainable way.

Conflicts of interest

The authors declare no conflict of interest.



Author details

Sajid Rashid Ahmad*, Sana Ashraf* and Humaira Nawaz College of Earth and Environmental Sciences, University of the Punjab, Lahore, Pakistan

*Address all correspondence to: sajidpu@yahoo.com and sana.cees@pu.edu.pk

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References

- [1] Muchate, N.S., G.C. Nikalje, N.S. Rajurkar, P. Suprasanna and T.D. Nikam. 2016. Plant salt stress: adaptive responses, tolerance mechanism and bioengineering for salt tolerance. Bot Rev. doi:10.1007/s12229-016-9173-y.
- [2] Wicke, B., E. Smeet, V. Dornburg, B. Vashev, T. Gaiser, W. Turkenburg and A. Faaij. 2011. The global technical and economic potential of bioenergy from salt-affected soils. Energy Environment Science, 4: 2669-2681.
- [3] QURESHI, R.H., M. Aslam and A. Javaid. 2003. Productivity enhancement in the salt affected lands of Joint Satiana Pilot Project Area of Pakistan. J Crop Prod 7, 277-297.
- [4] Rajaravindran, M. and S. Natarajan. 2012. Effects of salinity stress on growth and antioxidant enzymes of the halophyte *Sesuvium portulacastrum*. International Journal Research Plant Science, 2: 23-28.
- [5] Munns, R. and M. Tester. 2008. Mechanism of salinity tolerance. Annu. Rev. Plant Biol., 59: 651-681 Tester M, Davenport R (2003) Na+ tolerance and Na+ transport in higher plants. Ann Bot 91(5):503-27. doi:10.1093/aob/mcg058.
- [6] Tester M, Davenport R (2003) Na+ tolerance and Na+ transport in higher plants. Ann Bot 91(5):503-27. doi:10.1093/ aob/mcg058.
- [7] Hanin, M., C.Ebel, M. Ngom, L. Laplaze and K. Masmoudi. 2016. New insights on plant salt tolerance mechanisms and their potential use for breeding. Front Plant Sci. 7:1787
- [8] Deinlein, U., A.B. Stephan, T. Horie, W. Luo, G. Xu and J.I. Schroeder. 2014. Plant salt-tolerance mechanisms. Trends Plant Sci. 19:371-379.
- [9] FAO. 2000. Global network on integrated soil management for

- sustainable use of salt-affected soils. Available in: http://www.fao.org/ag/AGL/agll/spush/intro.htm (28. Jan. 2015)
- [10] WWAP. 2012. World Water Assessment Programme. The United Natins World Water Development Report 4: Managing Water under Uncertainity and Risk. Paris: UNESCO.
- [11] FAO. 2008. Land and Plant Nutrition Management Service. http:// www.fao.org/ag/agl/agll/. Accessed on November/ 15/2012.
- [12] Schirawski, J., and M.H. Perlin. 2018. Plant- microbe interaction 2017-the good, the bad and the diverse. Int. J. Mol. Sci. 19:1374.
- [13] Panuccio, M.R., S.E. Jacobsen, S.S. Akhtar and A. Muscolo. 2014. Effect of saline water irrigation on seed germination and early seedling growth of the halophyte quinoa. AoB Plants 6, plu047
- [14] Mager, P., M. Gerth and J.I. Schreoeder. 2002. Molecular mechanisms of potassium and sodium uptake in plant. Plant Soil 247, 43-54.
- [15] Tester, M., and R. Davenport. 2003. NaC tolerance and NaC transport in higher plants. Ann. Bot. 91, 503-527.
- [16] Zhu, J.K. 2016. Abiotic stress signaling and responses in plants. Cell, 167, 313-324.
- [17] Yang, Y. and Y. Guo. 2018. Elucidating the molecular mechanisms mediating plant salt-stress responses. New Phytol. 217:523-539.
- [18] Fu, Q., C. Liu, N. Ding, Y. Lin and B. Guo. 2010. Ameliorative effects of inoculation with the plant growth promoting rhizobacterium Pseudomonas sp. DW1 ongrowth of eggplant (*Solanum melongena* L.)

- seedlingsunder salt stress, Agr. Water Manage., 97: 1994-2000.
- [19] Ma, Y., M. Rajkumar, C. Zhang and H. Freitas. 2016. Beneficial role of bacterial endophytes in heavy metal phytoremediation. J. Environ. Manag. 174, 14-25.
- [20] Ayyam, V., S. Palanivel and S. Chandrakasan. 2019. "Approaches in land degradation management for productivity enhancement," in Coastal Ecosystems of the Tropics Adaptive Management, eds V. Ayyam, S. Palanivel, and S. Chandrakasan (Singapore: Springer)
- [21] ugtenberg, B. and F. Kamilova. 2009. Plant-growth-promoting rhizobacteria. Annu. Rev. Microbiol. 63, 541-556.
- [22] Qin, Y., I.S. Druzhinina, X. Pan and Z. Yuan. 2016. Microbially mediated plant salt tolerance and microbiomebased solutions for saline agriculture. Biotechnol Adv. 34:1245-1259.
- [23] Mayak, S., T. Tirosh and B.R. Glick. 2004. Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. Plant Physiology and Biochemistry 42, 565-572.
- [24] Ahmad, M., Z.A. Zahir, M. Khalid, F. Nazli and M. Arshad. 2013. Efficacy of Rhizobium and Pseudomonas strains to improve physiology, ionic balance and quality of mung bean under salt-affected conditions on farmer's fields. Plant Physiology and Biochemistry 63, 170-176.
- [25] Yang, J., J.W. Kloepper and C.M. Ryu. 2009. Rhizosphere bacteria help plants tolerate abiotic stress. Trends Plant Sci. 14:1-4.
- [26] Glick BR, Penrose DM, Li J (1998) A model for the lowering of plant ethylene concentrations by plant growth-promoting bacteria. Journal of

- Theoretical Biology 190, 63-68. doi:10.1006/jtbi.1997.0532.
- [27] Upadhyay SK, Singh JS, Singh DP (2011) Exopolysaccharide-producing plant growth-promoting rhizobacteria under salinity condition.
- [28] Balsanelli, E., V.A. de Baura, F.D. Pedrosa, E.M. de Souza and R.A. Monteiro. 2014. Exopolysaccharide biosynthesis enables mature biofilm formation on abiotic surfaces by Herbaspirillum seropedicae. PLOS ONE 9: 110-392.
- [29] Ashraf, M., S.H. Berge and O.T. Mahmood. 2004. Inoculating wheat seedlings with exopolysaccharide-producing bacteria restricts sodium uptake and stimulates plant growth under salt stress. Biol. Fertil. Soils 40, 157-162.
- [30] Chatterjee, P., S. Samaddar, R. Anandham, Y. Kang, K. Kim and G. Selvakumar. 2017. Beneficial soil bacterium Pseudomonas frederiksbersgensis OS261augments salt tolerance and promotes red pepper plant growth. FrontPlant Sci. 8:1-9.
- [31] Egamberdieva, D., D. Jabborova and A. Hashem. 2015. Pseudomonas induces salinity tolerance in cotton (*Gossypium hirsutum*) and resistance to Fusariumroot rot through the modulation of indole-3-acetic acid. Saudi J BiolSci. 22(6):773-9.
- [32] Glick BR. 2014. Bacteria with ACC deaminase can promote plant growth and help to feed the world. Microbiol Res. 169:30-39.
- [33] Numan, M., S. Bashir, Y. Khan, R. Mumtaz, Z.K. Shinwari, A.L. Khan, A. Khan and A. Al-Harrasi. 2018. Plant growth promoting bacteria as an alternative strategy for salt tolerance in plants: a review. Microbiol Res. 209:21-32.
- [34] Alzahrani, Y., A. Kuşvuran, H.F. Alharby, S. Kuşvuran and M.M. Rady.

- 2018. The defensive role of silicon in wheat against stress conditions induced by drought, salinity or cadmium. Ecotoxicol Environ Saf.154:187-196.
- [35] Etesami, H. 2018. Can interaction between silicon and plant growth promoting rhizobacteria benefit in alleviating abiotic and biotic stresses in crop plants? Agric Ecosyst Environ. 253:98-112.
- [36] Garg, N. and P. Bhandari. 2016. Interactive effects of silicon and arbuscular mycorrhiza in modulating ascorbate-glutathione cycle and antioxidant scavenging capacity in differentially salt-tolerant *Cicer arietinum* L. genotypes subjected to long-term salinity. Protoplasma. 253:1325-1345.
- [37] Li, Y.T., W.J. Zang, J.J. Cui, D.Y. Lang, M. Li, Q.P. Zhao and X.H. Zhang. 2016. Silicon nutrition alleviates the lipid peroxidation and ion imbalance of Glycyrrhiza uralensis seedlings under salt stress. Acta Physiol Plant. 38:96-105.
- [38] Rios, J.J., M.C. Martínez-Ballesta, J.M. Ruiz, B. Blasco and M. Carvajal. 2017. Silicon-mediated improvement in plant salinity tolerance: the role of aquaporins. Front Plant Sci. 8:948.
- [39] Abbas, T., R.M. Balal, M.A. Shahid, M.A. Pervez, C.M. Ayyub, M.A. Aqueel and M.M. Javaid. 2015. Silicon-induced alleviation of NaCl toxicity in okra (*Abelmoschus esculentus*) is associated with enhanced photosynthesis, osmoprotectants and antioxidant metabolism. Acta Physiol Plant. 37:6.
- [40] Garg, N. and P. Bhandari. 2015. Silicon nutrition and mycorrhizal inoculations improve growth, nutrient status, K+/Na+ ratio and yield of *Cicer arietinum* L. genotypes under salinity stress. Plant Growth Regul. 78, 371-387.
- [41] Clark, G.J., N. Dodgshun, P.W.G. Sale and C. Tang. 2007. Changes in chemical and biological properties of a

- sodic clay subsoil with addition of organic amendments. Soil Biology and Biochemistry, 39: 2806-2817.
- [42] Ghafoor, A., G. Murtaza, B. Ahmad and T.M. Boers. 2008. Evaluation of amelioration treatments and economic aspects of using saline-sodic water for rice and wheat production on saltaffected soils under arid land conditions. Irrigation and Drainage, 57: 424-434.
- [43] Wong, V.N., R.C. Dalal and R.S. Greene. 2009. Carbon dynamics of sodic and saline soils following gypsum and organic material additions: a laboratory incubation. Applied Soil Ecology, 41: 29-40.
- [44] Cha-um, S. and C. Kirdmanee. 2011. Remediation of salt-affected soil by the addition of organic matter: an investigation into improving glutinous rice productivity. Scientia Agricola, 68: 406-410.
- [45] Parida AK, Das AB (2005) Salt tolerance and salinity effects on plants: a review. Ecotox Environ Safe 60(3):324-49. doi:10.1016/j.ecoenv.2004.06.010
- [46] Meloni DA, Oliva MA, Martinez CA, Cambraia J (2003) Photosynthesis and activity of superoxide dismutase, peroxidase and glutathione reductase in cotton under salt stress. Environ Exp Bot 49(1):69-76.doi:10.1016/S0098-8472(02)00058-8
- [47] Hasegawa PM, Bressan RA, Zhu JK, Bohnert HJ (2000) Plant cellular and molecular responses to high salinity. Annu Rev Plant Physiol Plant Mol Biol 51:463-99. doi:10.1146/annurev. arplant.51.1.463
- [48] Shannon MC, Grieve CM (1999) Tolerance of vegetable crops to salinity. Sci Hortic 78(1-4):5-38. doi:10.1016/ S0304-4238(98)00189-7
- [49] Larcher W. (1980) Physiological plant ecology: ecophysiology and stress

- physiology of functional groups, 2nd edn. Springer-Verlag, Berlin.
- [50] Xu ZH, Saffigna PG, Farquhar GD, Simpson JA, Haines RJ, Walker S et al (2000) Carbon isotope discrimination and oxygen isotope composition in clones of the F (1) hybrid between slash pine and Caribbean pine in relation to tree growth, water-use efficiency and foliar nutrient concentration. Tree Physiol 20(18):1209-17. doi:10.1093/treephys/20.18.1209
- [51] Tester, M., and R. Davenport. 2003. NaC tolerance and NaC transport in higher plants. Ann. Bot. 91, 503-527
- [52] White PJ, Broadley MR (2001) Chloride in soils and its uptake and movement within the plant: a review. Ann Bot 88(6):967-88. doi:10.1006/anbo.2001.1540.
- [53] Ruiz-Lozano, J. M., Porcel, R., Azcón, C., & Aroca, R. (2012). Regulation by arbuscular mycorrhizae of the integrated physiological response to salinity in plants: new challenges in physiological and molecular studies. *Journal of Experimental Botany*, 63(11), 4033-4044.
- [54] Hasegawa PM, Bressan RA, Zhu JK, Bohnert HJ (2000) Plant cellular and molecular responses to high salinity. Annu Rev Plant Physiol Plant Mol Biol 51:463-99. doi:10.1146/annurev. arplant.51.1.463.
- [55] ietz DN, Haynes RJ (2003) Effects of irrigation-induced salinity and sodicity on soil microbial activity. Soil Biol Biochem 35(6):845-54.doi:10.1016/S0038-0717(03)00125-1
- [56] Borneman J, Skroch PW, O'Sullivan KM, Palus JA, Rumjanek NG, Jansen JL et al (1996) Molecular microbial diversity of an agricul-tural soil in Wisconsin. Appl Environ Microbiol 62(6):1935-43.
- [57] Jofre E, Fischer S, Rivarola V, Balegno H, Mori G (1998) Saline stress

- affects the attachment of Azospirillum brasilense Cd to maize and wheat roots. Can J Microbiol 44(5):416-22. doi: 10.1139/w98-024.
- [58] Ondrasek G, Rengel Z, Romic D, Savic R (2010) Environmental salinisation processes in agro-ecosystem of neretva river estuary. Novenytermeles 59:223-226.
- [59] Fuller, M.P., J.H. Hamza, H.Z. Rihan and M. Al-Issawi. 2012. Germination of primed seed under NaCl stress in wheat. Int. Sch. Res. Netw. Bot.,12: 1-5. https://doi.org/10.5402/2012/167804
- [60] Kim, H., H. Jeong, J. Jeon and S. Bae. 2016. Effects of irrigation with saline water on crop growth and yield in greenhouse cultivation. Water, 8(4): 127-135. https://doi.org/10.3390/w8040127.
- [61] Plaut, Z., M. Edelstein and M. Ben-Hur. 2013. Overcoming salinity barriers to crop production using traditional methods. Crit. Rev. Plant Sci.,.32(4): 250291. https://doi.org/10.1080/07352689.2012.752236.
- [62] Shaaban, M., M. Abid and R.A.I. Abou-Shanab. 2013. Amelioration of salt affected soils in rice paddy system by application of organic and inorganic amendments. Plant Soil Environ., 59(5): 227-233. https://doi.org/10.17221/881/2012-PSE
- [63] Amezketa, E.A., R. Aragues and R. Gazol. 2005. Efficiency of sulfuric acid, mined gypsum and two gypsum by-products in soil crusting prevention and sodic soil reclamation. Agron. J., 97: 983-989. https://doi.org/10.2134/agronj2004.0236
- [64] Wahid, A., S. Akhtar, I. Ali and E. Rasul. 2015. Amelioration of saline-sodic soils with organic matter and their use for wheat growth. Commun. Soil Sci. Plant Anal., 29(15-16):

- 2307-2318.https://doi.org/10.1080/ 0010362980937011
- [65] Mamo, M., J.F. Moncrief, C.J. Rosen and T.R.Halbach. 2000. Municipal solid waste compost application on soil water and water stress in irrigated corn. Compost Sci. Util., 8(3):236-246. https://doi.org/10.1080/1065657X. 2000.10701996
- [66] Ahmad, M., Z.A. Zahir, M. Khalid, F. Nazli and M. Arshad. 2013. Efficacy of Rhizobium and Pseudomonas strains to improve physiology, ionic balance and quality of mung bean under salt-affected conditions on farmer's fields. Plant Physiology and Biochemistry 63, 170-176.
- [67] Iqbal, M., A. Hassan and M. Ibrahim. 2008. Effects of tillage systems and mulch on soil physical quality parameters and maize (*Zea mays* L.) yield in semi-arid Pakistan. Biol. Agric. Hortic.,25(4): 311-325. https://doi.org/10.1080/01448
- [68] Qadir M, Quillérou E, Nangia V, Murtaza G, Singh M, Thomas RJ, Drechsel P, Noble AD (2014) Economics of salt-induced land deg-radation and restoration. Nat Res Forum 38:282-295
- [69] Rengasamy, P. 2002. Transient salinity and subsoil constraints to dryland farming in Australian sodic soils: An overview. Aust. J. Exp. Agric., 42: 351-61.
- [70] Munns, R. and M. Tester. 2008. Mechanisms of salinity tolerance. Annu. Rev. Plant Biol. 59, 651-681.
- [71] Tavakkoli, E., P. Rengasamy and G.K. McDonald. 2010. High concentrations of Na+ and Cl– ions in soil solution have simultaneous detrimental effects on growth of faba bean under salinity stress. J. Exp. Bot., 61: 4449-4459.
- [72] Shahzad, M., K. Witzel, C. Zörb, and K.H. Mühling. 2012.

- Growth-related changes in subcellular ion patterns in maize leaves (*Zea mays* L.) under salt stress. J. Agron. Crop Sci.,198: 46-56.
- [73] Abbasi, G.H., J. Akhtar, M. Anwarul-Haq, S. Ali, Z. Chen, and W. Malik. 2014. Exogenous potassium differentially mitigates salt stress in tolerant and sensitive maize hybrids. Pak. J. Bot., 46: 135-146.
- [74] Lauchli, A. and E. Epstein. 1990. Plant responses to saline and sodic conditions, in: (Ed.): Tanji K.K. agricultural salinity assessment and management, American Society of Civil Engineering, New York, p. 113-137.
- [75] Johnson HE, Broadhurst D, Goodacre R, Smith AR (2003) Metabolic fingerprinting of salt-stressed tomatoes. Phytochemistry 62(6):919–28. doi:10.1016/S0031-9422(02)00722-7.
- [76] Spadaro, D., B.W. Yun, S.H. Spoel, C. Chu, Y.Q. Wang and G.J. Loake. 2010. The redox switch: dynamic regulation of protein function by cysteine modifications. Physiol. Plant.,138: 360-371.
- [77] Singh, B.R. 2003. Sulfur and crop quality-agronomical strategies for crop improvement. Abstracts of COST Action 829 Meetings, Braunschweig, Germany. p. 35-36.
- [78] Taiz, L. and E. Zeiger. 2006. Plant Physiology. 4th Edition.Sinauer Associates Inc. Sunderland, Massachusetts.
- [79] Ashraf M, Afzal M, Ahmed R, Mujeeb F, Sarwar A, Ali L (2010). Alleviation of detrimental effects of NaCl by silicon nutrition in salt Esensitive and Etolerant genotypes of sugarcane (*Saccharum officinarum* L.). Plant Soill 326(12):381-391.
- [80] Ashraf M, Ozturk M, Ahmad MSA, Aksoy A (2012). Crop production for

- agricultural improvement. Springer Science+Business Media, NY.
- [81] Dionisio-Sese, M. L., & Tobita, S. (1998). Antioxidant responses of rice seedlings to salinity stress. *Plant Science*, *135*(1), 1-9.
- [82] Tuna, A. L., Kaya, C., Higgs, D., Murillo-Amador, B., Aydemir, S., and Girgin, A.R. (2008). Silicon improves salinity tolerance in wheat plants. Environ. Exp. Bot. 62, 10-16. doi: 10.1016/j.envexpbot.2007.06.006
- [83] Liang Y, Sun W, Zhu YG, Christie P (2007). Mechanisms of silicon-mediated alleviation of abiotic stresses in higher plants: A review.Environmental Pollution 147(2):422-428.
- [84] Khan, M.A. and I.A. Ungar. 1997. Effect of thermo period on recovery of seed germination of halophyte from saline conditions. Am. J. Bot., 84: 279-283.
- [85] Mahajan, S., & Tuteja, N. (2005). Cold, salinity and drought stresses: an overview. *Archives of biochemistry and biophysics*, 444(2), 139-158.
- [86] Tuna, A. L., Kaya, C., Higgs, D., Murillo-Amador, B., Aydemir, S., and Girgin, A.R. (2008). Silicon improves salinity tolerance in wheat plants. Environ. Exp. Bot. 62, 10-16. doi: 10.1016/j.envexpbot.2007.06.006
- [87] Li YQ, Zhao HL, Yi XY, Zuo XA, Chen YP (2006) Dynamics of carbon and nitrogen storages in plant–soil system during desert-ification process in horqin sandy land. Huan Jing Ke Xue 27(4):635-40.
- [88] Jastrow JD, Miller RM (1991) Methods for assessing the effects of biota on soil structure. Agric Ecosyst Environ 34(1-4):279-303. doi:10.1016/ 0167-8809(91)90115-E.
- [89] Dimkpa C, Weinand T, Asch F (2009) Plant-rhizobacteria interactions

- alleviate abiotic stress conditions. Plant Cell Environ 32(12):1682-94. doi: 10.1111/j.1365-3040.2009.02028.x.
- [90] Hayat R, Ali S, Amara U, Khalid R, Ahmed I (2010) Soil beneficial bacteria and their role in plant growth promotion: a review. Ann Microbiol 60 (4):579-98. doi:10.1007/s13213-010-0117-1.
- [91] Lugtenberg B, Kamilova F (2009) Plant-growth-promoting rhizobacteria. Annu Rev Microbiol 63:541-56. doi:10.1146/annurev.micro.62.081307. 162918.
- [92] Diby P, Bharathkumar S, Sudha N (2005a) Osmotolerance in biocontrol strain of pseudomonas pseudoalcaligenes MSP-538: a study using osmolyte, protein and gene expression profiling. Ann Microbiol55(4):243-47
- [93] Diby P, Sarma YR, Srinivasan V, Anandaraj M (2005b) *Pseudomonas fluorescens* mediated vigour in black pepper (*Piper nigrum* L.) undergreen house cultivation. Ann Microbiol 55(3):171-74.
- [94] Egamberdieva D, Kucharova Z (2009) Selection for root colonizing bacteria stimulating wheat growth in saline soils. Biol Fertil Soils45(6):563-71. doi:10.1007/s00374-009-0366-y
- [95] Egamberdieva D (2012) Pseudomonas chlororaphis: a salttolerant bacte-rial inoculant for plant growth stimulation under saline soil conditions. Acta Physiol Plant 34(2): 751-56. doi:10.1007/s11738-011-0875-9
- [96] Egamberdieva D (2011) Survival of pseudomonas extremorientalis TSAU20 and P. Chlororaphis TSAU13 in the rhizosphere of com-mon bean (Phaseolus.
- [97] Botella MA, del Amor FM, Amoros A, Serrano M, Martinez V, Cerda A(2000) Polyamine, ethylene and

other physico-chemical parameters in tomato (*Lycopersicon esculentum*) fruits as affected by salinity. Physiol Plant 109(4):428-34. doi:10.1034/j.1399-3054.2000.100409.x

[98] Botella MA, Martinez V, Pardines J, Cerdá A (1997) Salinity induced potassium deficiency in maize plants. J Plant Physiol 150(1-2):200-05. doi:10.1016/S0176-1617(97)80203-9.

[99] Serraj R, Sinclair TR (2002)
Osmolyte accumulation: can it really help increase crop yield under drought conditions? Plant Cell Environ 25(2):333-41. doi:10.1046/j.1365-3040.2002.00754.x

[100] Nadeem SM, Shaharoona B, Arshad M, Crowley DE (2012) Population density and functional diversity of plant growth promoting rhizobacteria associated with avocado trees in saline soils. ApplSoil Ecol 62:147-54. doi:10.1016/j.apsoil. 2012.08.005

[101] Nadeem SM, Zahir ZA, Naveed M, Arshad M (2007) Preliminary investigations on inducing salt tolerance in maize through inoculation with rhizobacteria containing ACC deaminase activity. Can J Microbiol 53(10):1141-9. doi:10.1139/W07-081.

[102] Kohler J, Caravaca F, Roldan A (2010) An AM fungus and a PGPR intensify the adverse effects of salinity on the stability of rhizosphere soil aggregates of *Lactuca sativa*. Soil Biol Biochem 42(3):429-34.doi:10.1016/j. soilbio.2009.11.021

[103] Kohler J, Hernandez JA, Caravaca F, Roldan A (2009) Induction of antioxidant enzymes is involved in the greater effectiveness of a PGPR versus AM fungi with respect to increasing the tolerance of lettuce to severe salt stress. Environ Exp Bot 65(2-3):245-52. doi:10.1016/j.envexpbot. 2008.09.008. [104] Rojas-Tapias D, Moreno-Galvan A, Pardo-Diaz S, Obando M, Rivera D, Bonilla R (2012) Effect of inoculation with plant growth-promoting bacteria (PGPB) on amelioration of saline stress in maize (Zeamays). Appl Soil Ecol 61:264-72. doi:10.1016/j.apsoil.2012.01.006.

[105] Bano A, Fatima M (2009) Salt tolerance in *Zea mays* (L) following inoculation with Rhizobium and Pseudomonas. Biol Fertil Soils 45(4):405-13. doi:10.1007/s00374-008-0344-9.

[106] Peng YL, Gao ZW, Gao Y, Liu GF, Sheng LX, Wang DL (2008) Ecophysiological characteristics of alfalfa seedlings in response to various mixed salt-alkaline stresses. J Integr Plant Biol 50(1):29-39.doi:10.1111/j.1744-7909.2007.00607.x

[107] Johnson HE, Broadhurst D, Goodacre R, Smith AR (2003) Metabolic fingerprinting of salt-stressed tomatoes. Phytochemistry 62(6):919–28. doi:10.1016/S0031-9422(02)00722-7.

[108] Mittova V, Tal M, Volokita M, Guy M (2003) Up-regulation of the leaf mitochondrial and peroxisomal antioxidative systems in response to salt-induced oxidative stress in the wild salt-tolerant tomato species Lycopersicon pennellii. Plant Cell Environ 26(6):845-56. doi:10.1046/j.1365-3040.2003.01016.x.

[109] Liang Y, Sun W, Zhu YG, Christie P (2007). Mechanisms of silicon-mediated alleviation of abiotic stresses in higher plants: A review.Environmental Pollution 147(2):422-428.

[110] Nautiyal CS, Govindarajan R, Lavania M, Pushpangadan P (2008) Novel mechanism of modulating natural antioxidants in functional foods: Involvement of plant growth promoting rhizobacteria NRRL B-30488. J Agr Food Chem 56(12):4474-81. doi:10.1021/ jf073258i.

[111] Chakraborty N, Ghosh R, Ghosh S, Narula K, Tayal R, Datta A, Chakraborty S (2013) Reduction of oxalate levels in tomato fruit 748 D. Paul, H. Lade and consequent metabolic remodeling following overexpression of a fungal oxalate decarboxylase1[W]. Plant Physiol 162(1):364-78. doi:10.1104/pp. 112.209197.

[112] Kim SY, Lim JH, Park MR, Kim YJ, Park TI, Se YW, Choi KG, Yun SJ (2005) Enhanced antioxidant enzymes are associated with reduced hydrogen peroxide in barley roots under saline stress. J Biochem Mol Biol 38(2):218-24. doi:10.5483/BMBRep.2005.38.2.218.

[113] Alami Y, Achouak W, Marol C, Heulin T (2000) Rhizosphere soil aggregation and plant growth promotion of sunflowers by an exopolysaccharide-producing rhizobium sp. strain isolated from sunflower roots. Appl Environ Microbiol 66(8):3393-8. doi:10.1128/AEM.66.8.3393-3398.2000.

[114] Geddie JL, Sutherland IW (1993) Uptake of metals by bacterial polysaccharides. J Appl Bacteriol 74(4):467-72. doi:10.1111/j.1365-2672.1993.tb05155.x.

[115] Dobbelaere S, Vanderleyden J, Okon Y (2003) Plant growth-promoting effects of diazotrophs in the rhizosphere. Crit Rev Plant Sci 22(2): 107-49. doi:10.1080/713610853.

[116] Bhattacharyya PN, Jha DK (2011) Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. World J Microbiol Biotechnol28(4):1327-1350. doi:10.1007/s11274-011-0979-9

[117] Khan, A.; Khan, A.L.; Muneer, S.; Kim, Y.H.; Al-Rawahi, A.; Al-Harrasi, A. Silicon and salinity: Crosstalk in cropmediated stress tolerance mechanisms. Front. Plant Sci. 2019, 10, 1429. [CrossRef]

[118] Adhikari, A.; Khan, M.A.; Lee, K.E.; Kang, S.M.; Dhungana, S.K.; Bhusal, N.; Lee, I.J. The halotolerant rhizobacterium-pseudomonas

[119] Mahmood, S.; Daur, I.; Al-Solaimani, S.G.; Ahmad, S.; Madkour, M.H.; Yasir, M.; Hirt, H.; Ali, S.; Ali, Z. Plant growth promoting

[120] Al-Garni, S.M.S.; Khan, M.M.A.; Bahieldin, A. Plant growth-promoting bacteria and silicon fertilizer enhance plant growth and salinity tolerance in *Coriandrum sativum*. J. Plant Interact. 2019, 14, 386-396. [CrossRef]

[121] Mayak S, Tirosh T, Glick BR (2004) Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. Plant Physiology and Biochemistry 42, 565-572. doi:10.1016/j.plaphy.2004. 05.009

[122] Ahmad M, Zahir ZA, Khalid M, Nazli F, Arshad M (2013) Efficacy of Rhizobium and Pseudomonas strains to improve physiology, ionic balance and quality of mung bean under salt-affected conditions on farmer's fields. Plant Physiology and Biochemistry 63, 170-176. doi:10.1016/j.plaphy.2012.11.024

[123] Ashraf M, Wu L (1994) Breeding for salinity tolerance in plants. Critical Reviews in Plant Sciences 13, 17-42. doi:10.1080/07352689409701906

[124] Ashraf M, Hasnain S, Berge O, Mahmood T (2004) Inoculating wheat seedlings with exopolysaccharide-producing bacteria restricts sodium uptake and stimulates plant growth under salt stress. Biology and Fertility of Soils 40, 157-162. doi:10.1007/s00374-004-0766-y

[125] Glick BR (2012) Plant growth-promoting bacteria: mechanisms and applications. Scientifica 15, 963401.

[126] Nadeem SM, Zahir ZA, Naveed M, Arshad M (2009) Rhizobacteria containing ACC-deaminase confer salt tolerance in maize grown on saltaffected fields. Canadian Journal of Microbiology 55, 1302-1309. doi:10.1139/W09-092

[127] Nadeem S, Zahir Z, NaveedM, Nawaz S (2013) Mitigation of salinityinduced negative impact on the growth and yield of wheat by plant growthpromoting rhizobacteria in naturally saline conditions. Annals of Microbiology 63, 225-232. doi:10.1007/ s13213-012-0465-0

[128] Ma Y, Prasad MNV, Rajkumar M, Freitas H (2011) Plant growth promoting rhizobacteria and endophytes accelerate phytoremediation of metalliferous soils. Biotechnology Advances 29, 248-258. doi:10.1016/j.biotechadv.2010.12.001

[129] Schulz B, Boyle C (2006) What are endophytes? In 'Microbial root endophytes'. (Eds BJE Schulz, CIC Boyle, TN Sieber) pp. 1-13. (Springer-Verlag: Berlin)

[130] Pillay VK, Nowak J (1997) Inoculum density, temperature, and genotype effects on in vitro growth promotion and epiphytic and endophytic colonization of tomato (*Lycopersicon esculentum* L.) seedlings inoculated with a pseudomonad bacterium. Canadian Journal of Microbiology 43, 354-361. doi:10.1139/m97-049

[131] Ma Y, Prasad MNV, Rajkumar M, Freitas H (2011) Plant growth promoting rhizobacteria and endophytes accelerate phytoremediation of metalliferous soils. Biotechnology Advances 29, 248-258. doi:10.1016/j.biotechadv.2010.12.001

[132] Ryan RP, Germaine K, Franks A, Ryan DJ, Dowling DN (2008) Bacterial endophytes: recent developments and applications. FEMS Microbiology Letters 278, 1-9. doi:10.1111/j.1574-6968. 2007.00918.x

[133] Thomas SC, Frye S, Gale N, Garmon M, Launchbury R, Machado N, Melamed S, Murray J, Petroff A, Winsborough C (2013) Biochar mitigates negative effects of salt additions on two herbaceous plant Biochar and microbes alleviate salinity stress Functional Plant Biology K species. Journal of EnvironmentalManagement 129, 62-68. doi:10.1016/j.jenvman.2013.05.057

[134] Cantrell KB, Hunt PG, Uchimiya SM, Novak JM, Ro KS (2012) Impact of pyrolysis temperature and manure source on physicochemical characteristics of biochar. Bioresource Technology 107, 419-428.doi:10.1016/j. biortech.2011.11.084

[135] Akhtar SS, Li G, Andersen MN, Liu F (2014) Biochar enhances yield and quality of tomato under reduced irrigation. Agricultural Water Management 138, 37-44. doi:10.1016/j. agwat.2014.02.016

[136] Akhtar SS, Andersen MN, Liu F (2015b) Biochar mitigates salinity stress in potato. Journal of Agronomy and Crop Science doi:10.1111/jac.12132

[137] Xu CY, Hosseini-Bai S, Hao Y, Rachaputi RC, Wang H, Xu Z, Wallace H (2014) Effect of biochar amendment on yield and photosynthesis of peanut on two types of soils. Environmental Science and Pollution Research International 22, 6112-6125. doi:10.1007/s11356-014-3820-9

[138] Lehmann J, Rillig MC, Thies J, Masiello CA, Hockaday WC, Crowley D (2011) Biochar effects on soil biota – a review. Soil Biology & Biochemistry 43, 1812-1836. doi:10.1016/j.soilbio.2011. 04.022.

[139] Akhtar SS, Andersen MN, Liu F (2015a) Residual effects of biochar on improving growth, physiology and yield of wheat under salt stress. Agricultural Water Management 158, 61-68. doi:10.1016/j.agwat.2015.04.010