

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Tracking *Salmonella* Enteritidis in the Genomics Era: Clade Definition Using a SNP-PCR Assay and Implications for Population Structure

Dele Ogunremi, Ruimin Gao, Rosemarie Slowey, Shu Chen, Olga Andrievskaia, Sadjia Bekal, Lawrence Goodridge and Roger C. Levesque

Abstract

Salmonella enterica serovar Enteritidis (or *Salmonella* Enteritidis, SE) is one of the oldest members of the genus *Salmonella*, based on the date of first description and has only gained prominence as a significant bacterial contaminant of food over the last three or four decades. Currently, SE is the most common *Salmonella* serovar causing foodborne illnesses. Control measures to alleviate human infections require that food isolates be characterized and this was until recently carried out using Pulsed-Field Gel Electrophoresis (PFGE) and phage typing as the main laboratory subtyping tools for use in demonstrating relatedness of isolates recovered from infected humans and the food source. The results provided by these analytical tools were presented with easy-to-understand and comprehensible nomenclature, however, the techniques were inherently poorly discriminatory, which is attributable to the clonality of SE. The tools have now given way to whole genome sequencing which provides a full and comprehensive genetic attributes of an organism and a very attractive and superior tool for defining an isolate and for inferring genetic relatedness among isolates. A comparative phylogenomic analysis of isolates of choice provides both a visual appreciation of relatedness as well as quantifiable estimates of genetic distance. Despite the considerable information provided by whole genome analysis and development of a phylogenetic tree, the approach does not lend itself to generating a useful nomenclature-based description of SE subtypes. To this end, a highly discriminatory, cost-effective, high throughput, validated single nucleotide based genotypic polymerase chain reaction assay (SNP-PCR) was developed focussing on 60 polymorphic loci. The procedure was used to identify 25 circulating clades of SE, the largest number so far described for this organism. The new subtyping test, which exploited whole genome sequencing data, displays the attributes of an ideal subtyping test: high discrimination, low cost, rapid, highly reproducible and epidemiological concordance. The procedure is useful for identifying the subtype designation of an isolate, for defining the population structure of the organism as well as for surveillance and outbreak detection.

Keywords: *Salmonella* Enteritidis, clades, WGS, SNP-PCR, PFGE, phage typing, nomenclature, population structure

1. Introduction

The genus *Salmonella* contains a large number of Gram-negative bacteria primarily found in the gastrointestinal tract of vertebrate organisms including humans, cattle, pigs, horses, companion animals, avian, reptiles and fish [1]. There are two species of *Salmonella*, namely *Salmonella enterica* and *S. bongori* [2]. *Salmonella enterica* is the species of relevance in food safety, and consists of five subspecies of varying importance in human health. *Salmonella enterica* subspecies *enterica* has received the greatest attention because of its large number of constituent organisms, now estimated at about 2,600, each defined as a serovar based on the Kauffman-White classification [1]. *Salmonella enterica* serovar Enteritidis (commonly written as *Salmonella* Enteritidis or SE) is the most prominent. The organism was originally described as a distinct species and named as *Salmonella enterica* alongside two other species namely *Salmonella choleraesuis* and *Salmonella typhi*. Since those early days, the taxonomy of *Salmonella* has changed to reflect two species and hundreds of serovars. Curiously, a limited number of *S. enterica* serovars is associated with foodborne illnesses of which SE has emerged over the last few decades as the most prevalent cause of foodborne salmonellosis in humans worldwide [3]. However, this has not always been the case and prior to the 1970s there was only the occasional report of foodborne salmonellosis attributable to SE.

The earliest reports of foodborne illnesses caused by *Salmonella* were attributed to duck egg sources as summarized by Scott [4]. Subsequently, the organisms were found in live chicks, ducks and ducklings [5, 6]. Although these early reports came from different countries, SE did not become a common cause of foodborne illnesses until the 1980s [7]. By 1994, SE was the most commonly reported *Salmonella* serotype, with an incidence of 110 laboratory-confirmed infections per 100,000 population in the Northeast of US, and shell eggs from hens were identified as the major vehicle for SE infection in humans [8], in contrast to the earlier reports incriminating duck eggs. A 2010 outbreak of egg-related SE infections in the US resulted in an estimated 1,939 illnesses and a recall of over 500 million eggs, which ranked as the largest egg recall in history and one of the most expensive food recalls ever [9]. Similar events occurred in other parts of the world and were severe enough to warrant a warning of a new pandemic [7]. Together with two other serovars namely, Typhimurium and Heidelberg, the three most common serovars alone account for 59% of *Salmonella* outbreaks in humans in Canada, while the 10 most commonly observed *Salmonella* serovars account for about 76% of the total *Salmonella* infections reported. Establishing epidemiological linkages between contaminated products and human disease for *Salmonella* serovars has been particularly difficult for a number of reasons. One of the historically important reasons has been the clonal nature of many of the dominant serovars, especially Enteritidis which makes discrimination of strains difficult and an attribution of a particular strain linked with illness to a food source particularly challenging.

One resource that has been used by researchers to study SE is the strain P125109 phage type 4 (PT4) which was isolated from an outbreak of human food poisoning in the United Kingdom, and traced back to a poultry farm. The strain is highly virulent in newly hatched chickens and is also invasive in laying hens, resulting in egg contamination [10, 11]. The complete genome sequences of the host-promiscuous SE PT4 isolate P125109 was determined by Thomson *et al.* in 2008 [12].

Next generation sequencing (NGS) and especially whole genome sequencing (WGS) has emerged in recent years and has made it possible to sequence bacterial genomes within hours, a remarkable feat that is revolutionizing the field of microbiology. With the advent of microbial WGS, new light is shed on the nature of pathogens and our understanding of the biology of *Salmonella* is steadily increasing as *Salmonella* genomes are generated increasingly at a rapid rate and are deposited in public databases. Further understanding of genome diversity and variation of bacterial pathogens has the potential to improve quantitative risk assessment and assess the evolution of *Salmonella*, relationship among strains and serovars, emergence of new strains and the role of mobile genetic elements especially plasmids and bacteriophages in *Salmonella* [13]. The recent development of the *Salmonella* SystOmics database (SalFoS <https://salfos.ibis.ulaval.ca/>), a rich collection of over 3000 *Salmonella* genomes and their metadata represents a milestone and an important resource for future approaches to mitigate the burden of foodborne salmonellosis [14].

Food safety which is significantly impacted by *Salmonella* has gained from the advent of microbial genomics. Subspecies characterization including serovar identification and strain differentiation can now be done using genomics approach. As will soon be evident to the reader, there is much work yet to be done as the new capacity is yet to translate to tangible benefits to the consumer. Outbreaks caused by SE have remained at a high level or even increasing and there is a need to evaluate the efficacy of procedures used to detect the organism in food as well as approaches used in tracking the organism through the entire spectrum of the food chain, from farm to fork.

2. Laboratory culture and identification of organism

2.1 Culture procedures for *Salmonella*

Culture-based methods are commonly employed to detect pathogens in food, and in clinical and environmental samples. The Compendium of Analytical Methods (<https://www.canada.ca/en/health-canada/services/food-nutrition/research-programs-analytical-methods/analytical-methods/compendium-methods.html>) and the Bacteriological Analytical Manual (<https://www.fda.gov/food/laboratory-methods-food/bacteriological-analytical-manual-bam>) are compilations of laboratory procedures developed by the food safety regulatory agencies in Canada and the United States, respectively and each contains a catalog of official and recommended methods for isolating and detecting *Salmonella*. Briefly, *Salmonella* detection in food relies on a series of culture steps in broth formulations optimized to resuscitate *Salmonella* following injury caused by food handling, processing and storage and to reduce the abundance of competing bacteria [15]. In many enrichment protocols, broth and culture plates have been described for the isolation of *Salmonella* in different types of samples and matrices [16–18]. Typically, the first step is to culture a suspect food sample in a non-selective pre-enrichment broth, examples of which are lactose broth, buffered peptone water, trypticase soy, brilliant green water, powdered milk with brilliant green and universal pre-enrichment [16]. Following an overnight incubation commonly performed at 37°C, the culture material is subsequently transferred into a selective enrichment broth which suppresses and inhibits the growth of non-salmonellae while expanding the *Salmonella* population, facilitating isolation by plating on the appropriate media plates [19, 20]. Tetrathionate (TT) and Rappaport-Vassiliadis (RV) broths and RV semi-solid medium are the most commonly used selective culture conditions, performed at 37° or 42°C overnight for several days [15, 19].

When used to detect the presence of a microorganism in a food sample, laboratory culture procedures are slow and time consuming, requiring the sequential use of non-selective and selective enrichment broths and could take a week or longer. Another disadvantage is the documented inherent bias in the performance of selective broths which results in the preferential recovery of certain *Salmonella* serovars and not others [17, 21, 22]. For instance, different *Salmonella* serotypes are recovered by culture procedures performed on non-clinical, non-human sources when compared to samples tested in hospitals and other clinical settings from patients experiencing symptoms. Experimental results show that members of some *Salmonella* serogroups are unable to effectively compete with other serovars leading to a reduced efficiency of recovery of some *Salmonella* organisms including SE, from contaminated food [21]. The use of culture-independent procedures that can lead to rapid and sensitive detection of *Salmonella* [23] may in time eclipse the routine use of culture methods for detection. Nevertheless, the recovery of *Salmonella* in food is currently required to establish risk to the consumer and in support of a regulatory action. For this reason, and for the purpose of building inventories of microbial organisms for clinical and regulatory food microbiology, culture procedures are expected to remain in use. A wide variety of selective plating media are available for the isolation of *Salmonella* and a number of them will now be examined.

2.1.1 Xylose lysine desoxycholate (XLD) agar

XLD agar is a selective growth medium originally shown to facilitate the isolation of *Shigella* but was demonstrably useful for *Salmonella* isolation and has been further modified since its first description [24, 25]. At pH 7.4, the XLD agar appears bright pink or red as a result of the phenol red indicator. *Salmonella* ferments xylose, a sugar molecule, to produce acid and the bacterial colony turns yellow. In time, xylose is consumed and lysine is in turn utilized which upon decarboxylation produces an acidic environment and colonies turn back to red. In contrast, *Shigella* cannot ferment xylose and the colony remains red. *Salmonella* is able to metabolize thiosulfate to produce hydrogen sulphide, leading to the formation of colonies with black centres, which is an important feature in differentiating *Salmonella* colonies from *Shigella*. XLD agar is capable of supporting other members of *Enterobacteriaceae* such as *Escherichia coli* however the colonies and media turns yellow because of the fermentation of lactose which is also present in the agar. *Pseudomonas aeruginosa* is also able to grow on XLD plates as pink, flat, rough colonies but will not metabolize thiosulfate nor turn black. *Proteus* organisms can grow on XLD to give rose colored colonies and can sometimes metabolize thiosulfate to render the colonies black which will be readily confused with *Salmonella*. In addition, *Salmonella* strains have been described that do not metabolize thiosulfate and will grow as pink colonies which will be readily confused with *Shigella*. Thus, XLD agar is a moderately selective medium for isolating *Salmonella* and for differentiating it from other organisms.

2.1.2 Xylose lysine Tergitol-4 (XLT-4) agar

Similar to XLD agar, XLT-4 agar is also a selective culture medium which is used to isolate and identify *Salmonella* in food and environmental samples. Compared to XLD agar, XLT-4 is supplemented with a surfactant, 7-ethyl-2-methyl-4-undecanol hydrogen sulfate commonly referred to as Tergitol 4 while lacking sodium chloride and sodium desoxycholate. The surfactant is responsible for the inhibition of *Proteus* spp. and other non-salmonellae. XLT-4 agar is

clearly one of the most stringent of all selective culture plates used for isolating *Salmonella* with positive colonies growing up as red and eventually turning black starting from the centre as a result of hydrogen sulfide production. However, *Salmonella* strains that fail to produce hydrogen sulfide appear as yellow colonies on XLT-4 agar [26, 27].

2.1.3 XA medium - modified XLD agar by adding D-arabinose

XA medium is an improved selective and differential medium over XLD agar following its supplementation with arabinose, a sugar that is fermented by *Citrobacter* and *Proteus* but not by *Salmonella* [28]. The sensitivity of isolation of *Salmonella* using the XA and XLD media are equally high, however, the specificity of XA medium (92.0%) is superior to that of XLD (73.0%) [28]. Many *Salmonella* organisms appear as black colonies on XA agar whereas non-salmonellae will either not grow or appear as pink colonies. The use of arabinose to differentiate *Salmonella* from other closely related organisms represents a cost-effective approach, especially when compared to chromogenic plates (see Section 2.1.7).

2.1.4 Hektoen enteric (HE) agar

HE agar is a selective and differential medium for isolating and distinguishing members of the genera of *Salmonella* and *Shigella* from the other *Enterobacteriaceae*. HE agar has a blue appearance and contains indicators of lactose fermentation and hydrogen sulfide production while inhibiting the growth of Gram-positive bacteria. Species belonging to *Enterobacteriaceae* that are capable of fermenting one or more carbohydrates produces yellow or salmon-orange colored colonies, e.g., *Klebsiella pneumoniae* which ferments lactose. Non-fermenters produce blue-green colonies. Organisms that reduce sulfur to hydrogen sulfide such as *Salmonella* will produce black colonies or blue-green colonies with a black center. In contrast, colonies of *Shigella* remain green and do not turn black because of inability to metabolize sulfur.

2.1.5 MacConkey agar

MacConkey agar is used for the isolation of Gram-negative enteric bacteria which represents a large group of bacteria prominent among which includes *Salmonella*, *E. coli*, *Proteus*, *Citrobacter*, *Klebsiella*, *Pseudomonas*, *Shigella*, *Enterobacter* and *Yersinia*. These organisms grow on the agar because of the selective property conferred by crystal violet and bile salts to inhibit the growth of Gram-positive bacteria. The indicator system is the neutral red dye which turns red at a pH below 6.8 but is colorless at higher pH. Thus, lactose fermenters such as *E. coli*, *Klebsiella* and *Enterobacter* which contain the *lac* operon form red or pink colonies on MacConkey agar. In contrast, the other organisms including *Salmonella* which are generally non-lactose fermenters do not change color. Because *Salmonella* produce colonies similar to other non-lactose fermenters on MacConkey, the medium does not allow for identification of *Salmonella*, an objective that has to be achieved by employing other more selective agars. At the same time, lactose fermenting *Salmonella* have historically been shown to be causes of severe infections and outbreaks in humans [29] which is attributable to the presence of the *lac* operon carried in the chromosome or on plasmids [30] and leading to colonies that appear pink or reddish on MacConkey agar. Despite its limitations, the MacConkey agar can still be a very useful addition to the collection of media needed to comprehensively isolate and identify *Salmonella* in contaminated samples.

2.1.6 Brilliant green sulfa (BGS) agar

The selectivity of the BGS agar is due to the presence of brilliant green and sulfadiazine, two components that individually inhibit Gram-positive and most Gram-negative bacilli. Phenol red is the pH indicator that detects changes in pH due to the fermentation of sucrose and/or lactose. *Salmonella* colonies range from reddish or pink to nearly white in color with a red zone. Lactose or sucrose fermenters occasionally grow on this medium and appear as yellow-green colonies surrounded by a yellow-green zone. The presence of sulfadiazine in the media is effective in inhibiting the growth of *E. coli* and *Proteus* and to a large extent *Shigella* species [31]. In a latter modification of the BGS agar, the replacement of lactose with glucose and of sulfadiazine with novobiocin to create the novobiocin-brilliant green agar (NBG), led to a higher recovery of *Salmonella* but the medium could not differentiate it from hydrogen sulfide-positive *Citrobacter* organism [32].

2.1.7 Salmonella chromogenic agar

Chromogenic plates have been developed for *Salmonella* as an improved alternative to procedures that rely on the ability of the organism to produce hydrogen sulfide or their inability to ferment lactose, attributes that are not fully diagnostic of *Salmonella*. This often results in *Citrobacter* and *Proteus* species being mistakenly identified as *Salmonella* while some atypical *Salmonella* are missed entirely, using agar plates described above. There are a number of commercially available chromogenic culture media which incorporate different chromogenic substrates and result in different colors of *Salmonella* colonies. Using the *Salmonella* chromogenic agar marketed by Oxoid (United Kingdom) as an example, the medium contains the substrate, Magenta-cap (5-bromo-6-chloro-3-indolylcaprylate) which is hydrolyzed by *Salmonella* species to give magenta colonies. The second substrate, X-Gal (5-bromo-4-chloro-3-indolyl-D-galactopyranoside), is hydrolyzed by many non-*Salmonella* species including *Citrobacter* and *Proteus* to give blue colonies [33, 34]. The selection for *Salmonella* is further enhanced by the presence of bile salts which inhibit Gram-positive bacteria, and of two antibiotics namely, novobiocin and cefsulodin which inhibit *Proteus* and *Pseudomonas*, respectively.

The isolation of *Salmonella* colonies in contaminated food demonstrates the presence of live organisms that can potentially cause harm. As indicated above, the procedure requires a combination of culture conditions, and takes time. Molecular procedures that can rapidly detect *Salmonella* are often used to accelerate the process, to improve on sensitivity of detection and also to confirm colonies as *Salmonella* because of the challenges with the isolation of the bacteria as outlined above. Many molecular techniques are now available for serotype-specific identification of SE.

2.2 Identification of *Salmonella* Enteritidis

Many laboratory diagnostic platforms have been applied to detect and identify *Salmonella* contamination in food and these include the PCR, enzyme-linked immunosorbent assay and the lateral flow assay [35–37]. Examples are available as commercial products. Currently, the most popular platform is the PCR and the most frequently used gene target is the *invA* gene. Nevertheless, many commercial offers do not disclose their target for proprietary reasons. PCR assays have also been developed with other gene targets present either in the chromosome, e.g., *flagellin* [38], *OriC* [39] *hilA* [40], *ttr* [41] or on plasmids, e.g., *SpvR*

operon [42]. Multiplex PCR assays that are able to detect and distinguish among multiple serovars have also been developed by including serovar-specific gene targets such as STM4449 (Typhimurium [43]), STM 4497 (Typhimurium [44]), *fliC* (Typhimurium [45]), *sdfI* (Enteritidis [46]) and *sefA* [29]. Recent work by Nadin-Davis and colleagues showed that many of the previously identified serovar specific markers were shared by other serovars especially *sefA* and *fliC* while highlighting the limitation with the use of a plasmid encoded target [47].

A multiplex PCR method which is capable of detecting all *Salmonella* spp., while identifying and distinguishing SE from the other two most prevalent serovars namely Typhimurium [48] and Heidelberg (Ogunremi et al., unpublished) is now available. The PCR was designed to amplify DNA fragments from four *Salmonella* genes, namely, *invA* gene (211-bp fragment), *iroB* gene (309-bp fragment), Typhimurium STM 4497 (523-bp fragment), and Enteritidis SE147228 (612-bp fragment) and has lately incorporated a 124-bp Heidelberg-specific fragment.

The identification of members of genus *Salmonella* to the subspecies level i.e., serovar is pivotal in tracking these pathogens along the food chain and the above molecular methods are very promising replacements to replace the traditional biochemical tests because of ease of application and high specificity for identifying SE and the other serotypes.

3. Typing of *Salmonella* Enteritidis

3.1 Serotyping

Serotyping has consistently been the basis of public health surveillance of *Salmonella* and has retained this primary role, as a first-line typing method, in the era of WGS based on the development of novel bioinformatics tools (see Section 3.3). Serotypes of *Salmonella* are defined by the presence of two types of antigens, namely, a heat stable, somatic O antigen, a component of the lipopolysaccharide envelope covering the organism which is an important virulence factor, and the H antigen which is present on the flagella of the organism [49]. The antigenic properties of the O antigen are depicted as numerals, e.g., 1,9,12 for SE. In contrast, the H antigens are described using one or a few letters for the phase I antigen (e.g., g, m for SE) or as a combination of letters and numbers for antigens that are expressed should the flagella bear a phase II antigen (e.g., r and 1, 2 for Heidelberg).

Agglutination assays are performed on the organisms using antibodies that are able to recognize specific antigenic molecules developed through laborious cross-absorption process against other serovars [50]. The result is an elaborate classification scheme, developed by Kauffman and White [51, 52] and which has now led to the identification of some 2,600 serotypes of *Salmonella*. The complexity has been further enhanced by the ability of plasmids and prophages to alter the expression of some of the antigens, and this had led to a frequent re-evaluation of some serovar designations. Fortunately, these alterations are fairly rare and the serotyping scheme has served well since first proposed by Schüte in 1920 [53]. Of the large number of *Salmonella* serovars identified so far, only a relatively small numbers, perhaps no more than 100 serovars are commonly associated with foodborne illnesses [54, 55].

3.2 Traditional subtyping procedures for *Salmonella* Enteritidis

There are two approaches for the subspecies characterization of SE. Phenotypic tests rely on the biochemical properties of the live organism and the most

prominent example is phage typing. More recently, DNA based approaches or genotypic tests have dominated the field. The most widely used genotypic test being the Pulsed-Field Gel Electrophoresis. Whole genome sequencing of the DNA of SE, has over the last few years, become the dominant subtyping method in the developed world.

3.2.1 Pulsed-field gel electrophoresis (PFGE)

The PFGE can be used to characterize bacteria isolates based on the pattern of distribution of restriction enzyme sites present in the organism's DNA. For *Salmonella*, the electrophoretic mobility of DNA fragments digested by the restriction enzyme *Xba*I or *Bln*I produces a characteristic fingerprinting pattern that is used to subtype the isolate. During the period between 2009 and 2019, the Canadian Food Inspection Agency used the PFGE for outbreak investigations as one of the two subtyping tests for SE, the other being the phage type. Despite the presence of hundreds of different PFGE types among field isolates of SE only two PFGE types predominated and each consisted of thousands of isolates in the Canadian PulseNet database. The two commonest Canadian primary PFGE types, namely SEN.XAI 0003 and SEN.XAI 0006, were responsible for 33.8 and 19.2% of Canadian SE isolates documented in the PulseNet database between 2012 and 2017 (Ogunremi, Allain and Nadon, unpublished). The predominance of only a few PFGE SE types was long recognized as a consequence of the poor discriminatory ability of the technique for analyzing the relatedness of SE isolates (**Table 1**) rather than a reflection of an evolutionary dominance of a few circulating strains [56]. These observations led to the pursuance of WGS as an alternative approach [57].

3.2.2 Phage typing

In contrast to the PFGE, phage typing is a phenotypic test that exploits the ability of certain bacteriophages, i.e., viruses that infect bacteria, to differentially attach and gain entrance into strains of bacteria. Phage typing of SE is the outcome of the pattern of susceptibility of different strains to a bacteriophage or a combination of bacteriophages, resulting in lysis of the bacterial cell [58]. A large number of phage types of SE have been described in Canada and elsewhere, however phage types 8, 13 and 13a were observed to predominate in Canada [59]. This observation may not reflect the presence of a few, circulating dominant strains of SE in Canada, but instead may be a consequence of the inadequacy of phage typing as a discriminatory tool that can accurately delineate the population structure of SE in Canada, similar to the PFGE as discussed above (see Section 3.2.2 and **Table 1**). The plasticity of phage types also diminishes its use as a subtyping tool. Factors such as the restriction system within the bacteria, ability of lipopolysaccharides and outer membranes to adsorb the bacteriophage, and the immune system of the vertebrate host infected by the bacteria can alter the phage type of an organism [60]. The reagents used for phage typing require very rigorous quality control and yet, test performance can be remarkably different among laboratories [61]. Changes occurring within an organism such as the acquisition or loss of IncN plasmid [62, 63], transfer of IncX plasmid [64] or loss of the lipopolysaccharide layer [65] have been shown to lead to poor test reproducibility. Thus, two isolates with the same phage type may in fact be unrelated and conversely, two isolates that show distinct phage types may be closely related. As a result of these factors, phage typing shows inadequate discriminatory power, partial typeability and poor reproducibility [66].

Clade	Strain identification	Source description	Phage type	PFGE type SENXAI, SENBNI	EnteroBase	
					MLST (7 gene)	cgMLST_v2 + HierCC_v1
1	2007-MI-0187-0006	Poultry environment	Atypical	0214, 0225	814	259062
2	08OTH012 6–4	Poultry environment	9b	0214, 0225	814	259068
3	06-1472	Animal feed	13a	0006, N/A	639	273915
4	OLF 10012–1	Sea food, clams	13, 1b	0009, 0013	11	5485
5	ID094888	Clinical case	6a	N/A, 0011	11	259098
6	dart-1997-742-B2	Cheese lunchables	8	0003, 0003	11	259481
7	S-MBS4754A	Chicken ceacum	51	N/A	8471	259064
8	SE974-OLF-2015-NSub	Bovine, heifer	N/A	N/A	11	260728
9	S-MBS1982A	Chicken thigh	N/A	N/A	11	259069
10	10OTH025 7–14	Poultry environment	13	0038, 0016	11	259063
11	S-MBS0737R	Chicken carcass	13a	N/A	11	259067
12	05–3936	Chicken breast	13a	0068, N/A	11	259480
13	07–1474	Chicken nuggets	8	0003, N/A	11	30959
14	S-MBS3492A	Chicken breast	N/A	N/A	11	259071
15	S-MBS7608A	Chicken carcass	8	N/A	11	259072
16	10SU010 19–1	Poultry environment	8	0003, 0003	11	5490
17	07–1485	Chicken nuggets	14b	0003, 0003	11	30959
18	S-MBS3006A	Chicken ceacum	8	N/A	11	259070
19	11OTH025 11-5	Poultry environment	8	0003, 0003	11	273916
20	S-MBS8825A	Chicken ceacum	8	N/A	11	259066
21	SA20100239	Bovine liver	2	N/A	11	14029
22	00D989 83–4	Poultry environment	23	0003, 0009	11	5498
23	SE972-OLF-2015-NSub112-S19	Water treatment plant	8	N/A	11	259100
24	ID112184	Human	8	0007, 0212	11	259479
25	EN1811	Food processing equipment	13	0076, 0003	11	233056

The single nucleotide-polymorphism chain reaction (SNP-PCR) was used to test Salmonella Enteritidis (SE) isolates and a representative strain for each designated clade (from 1 to 25) is shown in comparison to traditional and whole genome sequence based subtyping results. Only the SNP-PCR and EnteroBase core-genome multi-locus sequence typing (cg-MLST) supplemented with Hierarchical level analysis (HierCC) showed distinct resolution of the representative strains. All other methods including 7 gene MLST, phage typing and pulsed-field gel electrophoresis (PFGE) did not provide adequate discriminatory ability relevant for strain differentiation, outbreak investigation or tracking SE from farm to fork. N/A: Not available.

Table 1.
Clade designation of Salmonella Enteritidis organisms depicting a representative strain for each clade and comparison with the results of traditional and new subtyping assays.

3.2.3 Multiple locus variable-number tandem repeat analysis (MLVA) assay

MLVA is a molecular typing method that is based on PCR amplification of polymorphic regions of the DNA containing variable numbers of tandemly repeated sequences. The method has been standardized by PulseNet International and applied to the epidemiological investigations of SE either as a supplement or substitute for PFGE subtyping [67, 68]. An advantage of the MLVA is the designation of the typing results with a numeric sequence of tandem repeats. This represents a simple, easy-to-understand nomenclature which facilitated the reporting and exchange of test results between laboratories, and translated to a reliable tracking of an organism during epidemiological investigations. The discriminative ability of the MLVA has been variously shown to be superior [69], equivalent [70] or poorer than the PFGE [71].

Detailed genetic studies of SE have consistently shown the underlying causes of the poor discriminatory abilities of available subtyping tools, namely: isolates of SE are extremely similar (i.e., are highly clonal) and this poses a difficulty in finding a definitive, distinguishing trait that could be used to track lineages [70, 72, 73]. The timely arrival and increasing adoption of WGS has altered the analytical landscape.

3.3 Application of whole genome sequencing (WGS) in *Salmonella* Enteritidis: identification and characterization

The development of WGS procedure has heralded the application of a powerful technology for the identification and characterization of SE [57] which has been used for outbreak investigations [74], trace back procedures [75] and surveillance [76]. Furthermore, WGS analysis of SE has provided insights into phylogenetic relatedness of isolates, presence and prevalence antimicrobial resistance genes, novel mobile elements, virulence markers and bacteriophages in strains of the organism isolated from humans, food animals, production facilities and environmental sources [77–79]. Relevant to developing long term control and intervention strategies are the insights to be gained from the increasing application of WGS to the understanding of transmission dynamics of SE as was done in Chile to infer possible transmission of SE between gulls, poultry, and humans [80]. Bioinformatics approaches that allow useful information to be mined from genome sequences will now be discussed.

3.3.1 Whole genome-based serotyping

Serovar prediction can now be done on *Salmonella* isolates if the whole genome sequence is available by replacing the laborious agglutination assay (see Section 3.1) with an *in silico* analysis of the nucleotide sequence of the organism. Effectively, the traditional gold standard of traditional serology based on the Kauffmann-White Scheme has been replaced in the developed economies with *in silico* approaches [81]. Two of the mostly widely tools for this purpose are the *Salmonella In Silico* Typing Resource (SISTR) software and the SeqSero2 software [82, 83].

SISTR is an open, web-based bioinformatics platform capable of rapid *in silico* analyses of minimally processed draft assemblies of *Salmonella* genomes to generate accurate serovar designations. A collection of markers previously developed for the various *Salmonella* serovars formed the basis of the new tool [84]. The performance of SISTR is enhanced by the integration of additional multilocus sequence typing tools (see Section 3.3.2) which as a separate platform has been suggested as a replacement for the use of serotypes to define taxonomic as well as evolutionary groups of *Salmonella* [55]. SeqSero, which was launched in 2015 was developed to

employ the use of the *rfb* cluster, *fliC* and *flixB* to categorize *Salmonella* according to serovar using draft genome assemblies [83]. A subsequent improvement of the software, released as SeqSero2 included addition of markers at the level of the genus, species, subspecies as well as certain serotypes. Furthermore, a kmer-based algorithm was included that ensured a genome can be analyzed and the result available within seconds [85].

3.3.2 Multilocus sequence typing

Multilocus sequence typing (MLST) evaluates the nucleotide sequences of multiple housekeeping genes of an organism as a means of establishing similarities or differences among isolates [86]. Based on the sequences, each housekeeping gene is assigned an allele which can be stringed together in a nomenclature that defines the organism. Although the MLST scheme was developed using the bacterium *Neisseria meningitidis* [86], the advantage of electronic portability of sequence data and ease of incorporation of additional genes found a good synergy in the advent of WGS and has gained application in food safety. This has birthed the widely used EnteroBase (<https://enterobase.warwick.ac.uk/>) [87], an integrated web-based platform that permits the upload and analysis of short read Illumina sequences. This has allowed the expansion of the MLST scheme which was based on the initial six housekeeping genes [86] to a series of flexible applications and expansions for *Salmonella* including seven genes (legacy MLST), 3002 genes identified as the core genome of *Salmonella*, to produce core genome MLST (cgMLST) and 21,065 orthologous genes detected in a set of 537 *Salmonella* genomes, regarded as whole genome MLST (wgMLST). Despite the adoption of the wgMLST by PulseNet International [88], an influential international body which overlooks regulatory subtyping procedures for foodborne bacteria, EnteroBase's Sequence Type, ST, of *Salmonella* became a widely adopted subtype descriptor for *Salmonella*. However, ST does not provide adequate resolution for epidemiological concordance and outbreak level discrimination [89], and in addressing the challenge EnteroBase has additionally provided the core genome ST, cgSTs, complemented with a newly described 11 levels of genetic resolution hierarchies or HierCC for *Salmonella* (Table 1) [87, 90]. The result is a tool that appears to provide the needed resolution for strain differentiation in the context of disease outbreaks.

3.3.3 Single nucleotide polymorphism (SNP) pipelines

Single base substitutions represent one of the commonest variation in genomes and the resulting polymorphism can form the basis for the characterization of a microbe including SE. SNPs are detected as nucleotide changes at a specific location in a genome after aligning or comparing it to a designated reference genome. Bioinformatics pipelines have been developed to automate the aligning and identification of the variants. A number of SNP pipelines are in common use and will now be described. SNVPhyl which was developed at the Public Health Agency of Canada identifies high quality SNPs among a set of selected isolates and is useful for generating phylogenetic trees from these SNPs [91]. Public Health England developed SnapperDB, also a high-quality SNP pipeline which analyzes microbial genomes, evaluates genetic distances among the genomes and infers relatedness of strains [92]. Parsnp detects core genome SNP in bacterial genomes and with the aid of adjunct interactive tool Gingr can be used to display informative overviews for specific sub-clades and genomic regions [93]. The kSNP tool detects SNPs in the pan genome but is uniquely able to carry out comparisons among genomes without a requirement for genome alignment nor the use a reference genome [94].

3.4 Rationale for developing a new reliable, rapid, robust, cost-effective, epidemiologically concordant, easily implementable subtyping tool

A strategy aimed at developing a tool capable of differentiating lineages in the highly clonal *S. Enteritidis* lineages will likely require interrogating a significant amount of the bacterial DNA information. The opportunities provided by the massively parallel sequencing technology [95], which deduces the entire nucleotide sequence of an organism appeared at the onset to be the most viable option in charting a course to address the need. Use of genome sequence for taxonomy including strain differentiation could conceivably work well with strains showing significant genetic diversity, e.g., >5% differences among unrelated strains. However, this may be very difficult for a clonal organism such as SE where diversity between unrelated strains could be as little as 1% and the similar regions of the genome would have to be ignored before focusing on the dissimilar portions to demonstrate an accurate quantitative estimate of relatedness. This may explain the failure to use whole genome sequence to develop a reliable estimation of genetic distance by means of a phylogenetic tree for a group of SE isolates (Ogunremi et al., unpublished data) using a method shown to work for other bacteria [96].

Consequently, this led to an effort to develop, analyze and characterize the genomes of SE. During the early phase of this endeavor involving a select number of SE isolates from Canada, 669 SNPs were detected in the genome of SE [57]. Subsequent analysis of 135 SE genomes present in the GenBank in 2014 led to the identification of a total of 1440 SNPs providing a robust resource that was exploited for a SNP-based strain differentiation and clustering of foodborne SE isolates [57]. Thus, despite the universal acceptance of the usefulness of whole genome sequences for microbes, individual organisms such as the highly clonal SE may pose a unique challenge that might require a more focused analysis on carefully selected targets of the entire genome.

4. Single nucleotide polymorphism-polymerase chain reaction test (SNP-PCR) as a new, nomenclature friendly procedure

4.1 History and development of *Salmonella* Enteritidis lineages/clades and SNP-PCR

The existing molecular methods investigate only very small portions or attributes of the entire bacterial genome. The PFGE, as an example, identifies enzyme restriction patterns in the genome whereas WGS-based procedures have available for analysis detailed information on the entire genome to exploit as a basis for comparison and discrimination. To that end, extremely small differences, such as single nucleotide polymorphisms (SNPs), can be identified and used for subtyping as long as these attributes are consistently preserved in a particular bacterial lineage. Notably, Allard and colleagues [97] carried out bioinformatics analysis of a total of 104 SE genomes belonging, for the most part, to the predominant PFGE pattern (JEGX01.0004). They described a total of 9 clades and found 366 genes that showed variation, i.e., presence or absence, in the SE genome. This observation complemented and expanded on an earlier study by another laboratory which showed that two isolates of SE with the same phage type, PT 13a, were differentiated by a relatively large number of loci, i.e., 250 SNPs [73]. Similarly, by using a specific reference genome, for instance SE strain P125109, the WGS-based sequence reads were mapped to the reference to find SNPs which were used to build maximum-likelihood phylogenetic trees.

Another study involving 55 SE strains selected from clinical and environmental samples in Minnesota and Ohio from 2001 to 2014 showed the existence of only two major groups [98]. Furthermore, WGS based SNPs analysis of 675 SE isolates from 45 countries formed a global epidemic clade and two new clades that were found to be geographically restricted to distinct regions of Africa [99]. Using a closely related serovar - *S. Gallinarum* - as an outgroup, a maximum-likelihood phylogenetic tree was constructed based on the alignment of a total of 42,373 SNPs [99]. In addition, a SNP-based phylogenetic structure of 401 European SE isolates implicated outbreaks correlating with national and international egg distribution network [75].

Thus, genetic variation that could allow the development of a routine subtyping tool for tracking purposes is present and demonstrable within the SE genome but was apparently not fully exploited given the few number of subgroupings in each of the reported, sampled populations, and this presented a need to properly mine the SE genome and develop a very discriminatory subtyping procedure. In exploring this need, our hypothesis was that the use of a large number of SNPs may not necessarily improve the power of discrimination. More is not necessarily better. A large number of uninformative loci may be counterproductive and undesirable for strain differentiation. As a first step to address this need, whole-genome sequences of 11 SE isolates obtained in Canada were developed and compared to SE P125109 reference strain phage type 4 which led to the identification of 1361 loci where the SE genome showed SNP [100]. Subsequent selection of 60 SNPs spread throughout the genome and distributed among different gene types and in intergenic locations led to the development of a rapid, inexpensive fluorescence-based real time PCR subtyping assay [55].

4.2 The SNP-PCR subtyping procedure

The SNP-PCR genotype assay is an allele-specific, single amplification procedure based on the specific binding of one of two, competing forward primers, 18–20 nucleotides long, which differ by one single nucleotide at the locus of interest. The use of a single reverse primer completes the amplification process leading to the accumulation of an amplicon bearing the SNP of interest. Each primer is designed with a specific tail that allows a complementary binding with a commercially provided, customized sequence labeled with a fluorescent dye, FAM or HEX for allele 1 or 2 respectively (LGC Genomics, Beverly, MA). Thus, the first cycle of amplification ensures that the specific forward oligonucleotide present in the primer mix binds to the sequence containing the SNP and excludes the other primer. The reverse primer, also 18–20 nucleotides long, binds and elongates the fragment during amplification ensuring that the tail sequence is present, which then allows the accumulating fragment to contain either the FAM or HEX fluorescent label depending on the initial binding of one of the bi-allelic primers, which is dictated by which of the SNP corresponds to allele 1 or allele 2. Thus, detection is based on the use of fluorescent labeled sequence that assigns the allele number to either of the two nucleotides that may occupy the SNP position. The SNP alleles are compiled for all SE strains at the 60 loci and used as input to carry out evolutionary history analyses using Maximum Parsimony method, which was conducted using Molecular Evolutionary Genetics Analysis on the MEGA-X computing platform [101]. The distinct grouping of the SE isolates are identified as clades and each given a specific numerical description starting from 1.

Following the development of the SNP-PCR procedure, our initial application of the assay to a group of 55 SE isolates obtained in Canada led to the recognition of 12 clades of SE [57].

4.3 Twenty five circulating clades of *Salmonella* Enteritidis

Recently, the laboratory validation of the SNP-PCR assay was completed using 1,127 SE isolates obtained from food, animal, humans, and environmental sources in Canada and Europe and we observed a total of 25 circulating clades of SE (**Table 1**, Ogunremi et al., manuscript under preparation). In addition, 13 other globally distributed isolates identified from published papers [98, 99] as well as the widely used reference SE strain P125109 phage type 4 were also included in a phylogenetic comparison using the Maximum Parsimony method. These strains were distributed across the generated phylogenetic tree and homed to distinct SE clades providing further validation of the SNP-PCR tool to appropriately cluster strains and at the same time, distinguish among different strains (Ogunremi et al., manuscript under preparation). The validation procedure unambiguously demonstrated the robustness of the assay while displaying its prowess in estimating genetic distances and relatedness among and between clades, and its relevance in constructing an evolutionary map of SE following the testing of a large number of isolates.

4.4 Advantages of SNP-PCR: nomenclature and population structure

Previous studies aimed at evaluating the population structure of the highly clonal SE have reported fewer lineages and clades among isolates tested. For instance, a study of 675 very diverse isolates collected over many decades (1948–2013) in 45 countries and 6 continents revealed the presence of only 3 clades; a subgroup of 58 isolates was identified but could not be clustered by the method used by the authors [97]. Yet another study demonstrated 9 clades among a large but PFGE-uniform group of isolates [99]. These studies, which showed a limited diversity among SE populations, served to underscore our contrasting observations, and reinforced the excellent discrimination observed for SE using the validated SNP-PCR assay. The SNP-PCR compares well with cgMLST-HierCC function in EnteroBase in discriminating among strains chosen to represent SE clades from a very diverse SE population from a variety of sources and different continents (**Table 1**; Ogunremi et al., under preparation).

Apart from being a highly discriminatory and robust assay, the SNP-PCR is very cost-effective. Reagents cost are estimated at Can\$0.25 per SNP per isolate and testing 60 SNPs is cheaper than the traditional, less discriminatory subtyping assays (Can\$26 for phage typing and Can\$36 for two-enzyme PFGE analysis in reagent costs) or for WGS (Can\$100). The SNP-PCR validation procedure (described above) showed that only 17 SNP loci needed to be tested to assign an isolate to a clade and the test performed excellently well on crude, boiled bacterial extract, obviating the need for DNA purification and further creating an increased savings of reagents, labour and time.

Another important attribute of the SNP-PCR is its equal adaptability to few samples or a large number of samples. When compared to Illumina WGS which requires a prescribed number of samples per run (e.g., 20 *Salmonella* strains using MiSeq version 3 library kit over 600 cycle sequencing which runs for 65 hours), the SNP-PCR can be used to test one or a few samples with the appropriate controls without any cost implication on the volume of analysis. At the other end, a single PCR sample can handle a 384-well plate loaded with hundreds of samples and machine run completed in 2 hours. The labor costs of running the SE SNP-PCR test (2 h PCR time) and analyzing the results are at least an order of magnitude lower than those of any subtyping approach including traditional molecular tests or WGS. The SNP-PCR test shows very good reproducibility (95%) in tests conducted in six laboratories.

The SNP-PCR impressively satisfies all the seven criteria expected of an ideal subtyping test which includes cost effectiveness, rapid performance, robust results, typeability, high discrimination, reproducibility, and epidemiological concordance [66].

5. Conclusions

The bacterial pathogen, *Salmonella* Enteritidis is one of the most prevalent causes of foodborne illness in humans worldwide, yet tracking a strain of the organism through the food safety system is challenging because of its clonal nature, evident at the genomic level, which historically has resulted in poorly discriminating laboratory typing methods. The current application of genomics has led to the development of comprehensive and highly discriminatory tools however there are still challenges with the interpretation of the outputs and the application of the methods to differentiate between outbreaks and sporadic infections. The effect is a poorly understood population structure of SE.

This chapter illustrates the existence of 25 clades of SE, which should be useful for defining the population structure and tracking the pathogen from farm to fork. The phylogenetic relationships among the 25 clades of SE was obtained using a population of 1127 isolates obtained from a variety of sources in Canada and Europe. The validated SNP-PCR assay displayed the attributes of an ideal subtyping test and can be implemented in resource deprived countries where routine genome sequencing remains unaffordable, as well as in resource rich countries when characterizing a few isolates may not justify the expense of a genome sequencing run or for surveillance where interest in characterizing a large number of lower priority, non-clinical but valuable isolates is a very desirable goal.

Acknowledgements

This work was funded by the Canadian Food Inspection Agency, Ontario Ministry of Agriculture, Food and Rural Affairs, and Genome Research and Development Initiative of the Government of Canada.

Conflict of interest

The authors declare no conflict of interest.

Acronyms and abbreviations

PFGE	Pulsed-field gel electrophoresis
XLD	Xylose lysine desoxycholate
XLT4	Xylose lysine tergitol-4
HE	Hektoen enteric
BGS	Brilliant green sulfa
NTS	Non-typhoidal <i>Salmonella</i>
NGS	Next generation sequencing
SalFoS	<i>Salmonella</i> Foodborne Syst-OMICS database

IntechOpen

Author details

Dele Ogunremi^{1*}, Ruimin Gao^{1*}, Rosemarie Slowey², Shu Chen³,
Olga Andrievskaia¹, Sadjia Bekal⁴, Lawrence Goodridge⁵ and Roger C. Levesque⁶

1 Ottawa Laboratory Fallowfield, Canadian Food Inspection Agency,
Ottawa, Ontario, Canada

2 Central Veterinary Research Laboratory, Department of Agriculture Food and the
Marine, Co. Kildare, Ireland

3 Laboratory Services, University of Guelph, Guelph, Ontario, Canada

4 National Institute of Public Health of Quebec, Quebec, Canada

5 Department of Food Science, University of Guelph, Guelph, Canada

6 Institute for Integrative Systems Biology (IBIS), Université Laval, Quebec City,
Canada

*Address all correspondence to: dele.ogunremi@inspection.gc.ca
and ruimin.gao@inspection.gc.ca

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Özkalp B: **Isolation and identification of *Salmonellas* from different samples.** In: *A Dangerous Foodborne Pathogen* Edited by Mahmoud BSM: InTech; 2012. pp. 123-156.
- [2] Brenner FW, Villar RG, Angulo FJ, Tauxe R, Swaminathan B: ***Salmonella* nomenclature.** *J Clin Microbiol* 2000, **38**(7):2465-2467.
- [3] Vieira A JA, Pires SM, Karlsmose S, Wegener HC, Wong DLF: **WHO Global Foodborne Infections Network Country Databank – a resource to link human and non-human sources of *Salmonella*.** In *Proceeding of the 12th Symposium of the International Society for Veterinary Epidemiology and Economics*, Durban, South Africa 2009.
- [4] Scott WM: **Food poisoning due to eggs.** *Br Med J* 1930, **2**:56-58.
- [5] Garside JS, Gordon RF: ***Salmonella* infections of ducks and ducklings.** *J Comp Pathol* 1943, **53**:80-89.
- [6] Anonymous: **Eggs and *Salmonella* infections.** *Br J Med* 1944:760-761.
- [7] Rodrigue DC, Tauxe RV, Rowe B: **International increase in *Salmonella* Enteritidis: a new pandemic?** *Epidemiol Infect* 1990, **105**(1):21-27.
- [8] Braden CR: ***Salmonella enterica* serotype Enteritidis and eggs: a national epidemic in the United States.** *Clin Infect Dis* 2006, **43**(4):512-517.
- [9] **Multistate outbreak of human *Salmonella* Enteritidis infections associated with shell eggs.** In: Centers for Disease Control and Prevention; 2010. <https://www.cdc.gov/salmonella/2010/shell-eggs-12-2-10.html>.
- [10] Barrow PA: **Experimental infection of chickens with *Salmonella enteritidis*.** *Avian Pathol* 1991, **20**(1):145-153.
- [11] Barrow PA, Lovell MA: **Experimental infection of egg-laying hens with *Salmonella* Enteritidis phage type 4.** *Avian Pathol* 1991, **20**(2):335-348.
- [12] Thomson NR, Clayton DJ, Windhorst D, Vernikos G, Davidson S, Churcher C, Quail MA, Stevens M, Jones MA, Watson M *et al*: **Comparative genome analysis of *Salmonella* Enteritidis PT4 and *Salmonella* Gallinarum 287/91 provides insights into evolutionary and host adaptation pathways.** *Genome Res* 2008, **18**(10):1624-1637.
- [13] Branchu P, Bawn M, Kingsley RA: **Genome variation and molecular epidemiology of *Salmonella enterica* serovar Typhimurium pathovariants.** *Infect Immun* 2018, **86**(8):e00079-18.
- [14] Emond-Rheault JG, Jeukens J, Freschi L, Kukavica-Ibrulj I, Boyle B, Dupont MJ, Colavecchio A, Barrere V, Cadieux B, Arya G *et al*: **A Syst-OMICS approach to ensuring food safety and reducing the economic burden of Salmonellosis.** *Front Microbiol* 2017, **8**:996.
- [15] Daquigan N, Grim CJ, White JR, Hanes DE, Jarvis KG: **Early recovery of *Salmonella* from tood using a 6-hour non-selective pre-enrichment and reformulation of Tetrathionate broth.** *Front Microbiol* 2016, **7**:2103.
- [16] D'Aoust JY: **Update on preenrichment and selective enrichment conditions for detection of *Salmonella* in Foods.** *J Food Prot* 1981, **44**(5):369-374.
- [17] Gorski L: **Selective enrichment media bias the types of *Salmonella enterica* strains isolated from mixed strain cultures and complex enrichment broths.** *PLoS One* 2012, **7**(4):e34722.

- [18] Bailey JS, Cox NA: **Universal preenrichment broth for the simultaneous detection of *Salmonella* and *Listeria* in Foods.** *J Food Prot* 1992, **55**(4):256-259.
- [19] Vassiliadis P: **The Rappaport-Vassiliadis (RV) enrichment medium for the isolation of *Salmonellas*: an overview.** *J Appl Bacteriol* 1983, **54**(1):69-76.
- [20] Fagerberg DJ, Avens JS: **Enrichment and plating methodology for *Salmonella* detection in food.** *Journal Milk Food Technology* 1976, **39**:628-646.
- [21] Singer RS, Mayer AE, Hanson TE, Isaacson RE: **Do microbial interactions and cultivation media decrease the accuracy of *Salmonella* surveillance systems and outbreak investigations?** *J Food Prot* 2009, **72**(4):707-713.
- [22] Harvey RW, Price TH: **The examination of samples infected with multiple salmonella serotypes.** *J Hyg (Lond)* 1967, **65**(3):423-434.
- [23] Ogunremi D, Dupras AA, Naushad S, Gao R, Duceppe MO, Omid K, Marquez IG, Huang H, Goodridge L, Levesque RC *et al*: **A new whole genome culture-independent diagnostic Test (WG-CIDT) for rapid detection of *Salmonella* in lettuce.** *Front Microbiol* 2020, **11**:602.
- [24] Nye KJ, Fallon D, Frodsham D, Gee B, Graham C, Howe S, Messer S, Turner T, Warren RE: **An evaluation of the performance of XLD, DCA, MLCB, and ABC agars as direct plating media for the isolation of *Salmonella enterica* from faeces.** *J Clin Pathol* 2002, **55**(4):286-288.
- [25] Taylor WI: **Isolation of shigellae. I. Xylose lysine agars; new media for isolation of enteric pathogens.** *Am J Clin Pathol* 1965, **44**(4):471-475.
- [26] Miller RG, Tate CR, Mallinson ET, Scherrer JA: **Xylose-lysine-tergitol 4: an improved selective agar medium for the isolation of *Salmonella*.** *Poult Sci* 1991, **70**(12):2429-2432.
- [27] Albert MJ, Al Obaid K, Alfouzan W, Sheikh AR, Udo E, Izumiya H, Bulach DM, Seemann T: **Isolation of *Salmonella enterica* serovar Kentucky strain ST 198 and its H2S-negative variant from a patient: implications for diagnosis.** *J Clin Microbiol* 2014, **52**(11):4090-4093.
- [28] Park SH, Ryu S, Kang DH: **Development of an improved selective and differential medium for isolation of *Salmonella* spp.** *J Clin Microbiol* 2012, **50**(10):3222-3226.
- [29] Pickett G, Agate GH: **Outbreak of salmonellosis due to lactose-fermenting variant of *Salmonella newington*.** In. *Salmonella surveillance: Public Health Service, National Communicable Disease Center, Atlanta; MMWR* 1967, **16**:18.
- [30] Leonard SR, Lacher DW, Lampel KA: **Acquisition of the lac operon by *Salmonella enterica*.** *BMC Microbiol* 2015, **15**:173.
- [31] Moats WA, Kinner JA: **Factors affecting selectivity of brilliant green-phenol red agar for salmonellae.** *Appl Microbiol* 1974, **27**(1):118-123.
- [32] Devenish JA, Ciebin BW, Brodsky MH: **Novobiocin-brilliant green-glucose agar: new medium for isolation of salmonellae.** *Appl Environ Microbiol* 1986, **52**(3):539-545.
- [33] Rambach A: **New plate medium for facilitated differentiation of *Salmonella* spp. from *Proteus* spp. and other enteric bacteria.** *Appl Environ Microbiol* 1990, **56**(1):301-303.
- [34] Cassar R, Cuschieri P: **Comparison of *Salmonella* chromogenic medium with DCLS agar for isolation of *Salmonella* species from stool**

specimens. *J Clin Microbiol* 2003, **41**(7):3229-3232.

[35] Blais BW, Martinez-Perez A: **Detection of group D salmonellae including *Salmonella* Enteritidis in eggs by polymyxin-based enzyme-linked immunosorbent assay.** *J Food Prot* 2008, **71**(2):392-396.

[36] Mozola MA, Peng X, Wendorf M: **Evaluation of the GeneQuence DNA hybridization method in conjunction with 24-hour enrichment protocols for detection of *Salmonella* spp. in select foods: collaborative study.** *J AOAC Int* 2007, **90**(3):738-755.

[37] Seo KH, Valentin-Bon IE, Brackett RE, Holt PS: **Rapid, specific detection of *Salmonella* Enteritidis in pooled eggs by real-time PCR.** *J Food Prot* 2004, **67**(5):864-869.

[38] Song JH, Cho H, Park MY, Na DS, Moon HB, Pai CH: **Detection of *Salmonella* typhi in the blood of patients with typhoid fever by polymerase chain reaction.** *J Clin Microbiol* 1993, **31**(6):1439-1443.

[39] Malorny B, Hoorfar J, Bunge C, Helmuth R: **Multicenter validation of the analytical accuracy of *Salmonella* PCR: towards an international standard.** *Appl Environ Microbiol* 2003, **69**(1):290-296.

[40] Cardona-Castro N, Restrepo-Pineda E, Correa-Ochoa M: **Detection of *hlyA* gene sequences in serovars of *Salmonella enterica* subspecies *enterica*.** *Mem Inst Oswaldo Cruz* 2002, **97**(8):1153-1156.

[41] Gwida MM, Al-Ashmawy MA: **Culture versus PCR for *Salmonella* species identification in some dairy products and dairy handlers with special concern to its zoonotic importance.** *Vet Med Int* 2014, **2014**:502370.

[42] Jenikova G, Pazlarova J, Demnerova K: **Detection of *Salmonella* in food samples by the combination of immunomagnetic separation and PCR assay.** *Int Microbiol* 2000, **3**(4):225-229.

[43] Saeki EK, J. A, Bonfante RC, Hirooka EY, Moreira de Oliveira TCR: **Multiplex PCR (mPCR) for the detection of *Salmonella* spp. and the differentiation of the Typhimurium and Enteritidis serovars in chicken meat.** *J Food Saf* 2013, **33**:25-29.

[44] Shanmugasundaram M, Radhika M, Murali HS, Batra HV: **Detection of *Salmonella enterica* serovar Typhimurium by selective amplification of *fliC*, *fliB*, *iroB*, *invA*, *rfbJ*, *STM2755*, *STM4497* genes by polymerase chain reaction in a monoplex and multiplex format.** *World J Microbiol Biotechnol* 2009, **25**:1385-1394.

[45] Rodpai E, Moongkarndi P, Tunggrugsasut W, Phosannoradej R, Kanarat S: **Comparison of multiplex polymerase chain reaction and immunoassay to detect *Salmonella* spp., *S. Typhimurium*, and *S. Enteritidis* in Thai chicken meat.** *Science Asia* 2013, **39**:150-159.

[46] Agron PG, Walker RL, Kinde H, Sawyer SJ, Hayes DC, Wollard J, Andersen GL: **Identification by subtractive hybridization of sequences specific for *Salmonella enterica* serovar enteritidis.** *Appl Environ Microbiol* 2001, **67**(11):4984-4991.

[47] Nadin-Davis S, Pope L, Ogunremi D, Brooks B, Devenish J: **A real-time PCR regimen for testing environmental samples for *Salmonella enterica* subsp. *enterica* serovars of concern to the poultry industry, with special focus on *Salmonella* Enteritidis.** *Can J Microbiol* 2019, **65**(2):162-173.

- [48] Ogunremi D, Nadin-Davis S, Dupras AA, Marquez IG, Omid K, Pope L, Devenish J, Burke T, Allain R, Leclair D: **Evaluation of a multiplex PCR assay for the identification of *Salmonella* serovars Enteritidis and Typhimurium using retail and abattoir samples.** *J Food Prot* 2017, **80**(2):295-301.
- [49] Grimont PAD, Weill FX: **Antigenic formula of the *Salmonella* serovars.** In., 9 edn: World Health Organization Collaborating Centre for Reference and Research on *Salmonella*; 2007: 1-166.
- [50] Le Minor L, Rohde R: **Guideline for the preparation of *Salmonella* antisera,** 4 edn: World Health Organization Collaborating Centre for Reference and Research on *Salmonella*; 1989.
- [51] Salmonella Subcommittee of the Nomenclature Committee of the International Society for Microbiology: **The Genus *Salmonella* Lignieres, 1900.** *J Hyg (Lond)* 1934, **34**(3):333-350.
- [52] Kauffman F: **Über einen neuen serologischen Formenwechsel der Typhusbacillen.** *Z Hyg Infectkr* 1935, **116**:617.
- [53] Schüte H: **The paratyphoid B group** *Lancet* 1920, **1**:93.
- [54] Guibourdenche M, Roggentin P, Mikoleit M, Fields PI, Bockemuhl J, Grimont PA, Weill FX: **Supplement 2003-2007 (No. 47) to the White-Kauffmann-Le Minor scheme.** *Res Microbiol* 2010, **161**(1):26-29.
- [55] Jackson BR, Griffin PM, Cole D, Walsh KA, Chai SJ: **Outbreak-associated *Salmonella enterica* serotypes and food Commodities, United States, 1998-2008.** *Emerg Infect Dis* 2013, **19**(8):1239-1244.
- [56] Zheng J, Keys CE, Zhao S, Meng J, Brown EW: **Enhanced subtyping scheme for *Salmonella* Enteritidis.** *Emerg Infect Dis* 2007, **13**(12):1932-1935.
- [57] Ogunremi D, Kelly H, Dupras AA, Belanger S, Devenish J: **Development of a new molecular subtyping tool for *Salmonella enterica* serovar Enteritidis based on single nucleotide polymorphism genotyping using PCR.** *J Clin Microbiol* 2014, **52**(12):4275-4285.
- [58] Ward LR, de Sa JD, Rowe B: **A phage-typing scheme for *Salmonella* Enteritidis.** *Epidemiol Infect* 1987, **99**(2):291-294.
- [59] Public Health Agency of Canada. **The National Enteric Surveillance Program (NESP) Annual summary for 2013.** National Microbiology Laboratory, Public Health Agency of Canada, Winnipeg, Manitoba, Canada; 2015.
- [60] Rabsch W: ***Salmonella* Typhimurium phage typing for pathogens.** *Methods Mol Biol* 2007, **394**:177-211.
- [61] Baggesen DL, Sorensen G, Nielsen EM, Wegener HC: **Phage typing of *Salmonella* Typhimurium - is it still a useful tool for surveillance and outbreak investigation?** *Euro Surveill* 2010, **15**(4):19471.
- [62] Frost JA, Ward LR, Rowe B: **Acquisition of a drug resistance plasmid converts *Salmonella* Enteritidis phage type 4 to phage type 24.** *Epidemiol Infect* 1989, **103**(2):243-248.
- [63] Threlfall EJ, Chart H, Ward LR, de Sa JD, Rowe B: **Interrelationships between strains of *Salmonella* Enteritidis belonging to phage types 4, 7, 7a, 8, 13, 13a, 23, 24 and 30.** *J Appl Bacteriol* 1993, **75**(1):43-48.
- [64] Brown DJ, Baggesen DL, Platt DJ, Olsen JE: **Phage type conversion in *Salmonella enterica* serotype Enteritidis caused by the introduction of a resistance plasmid of incompatibility**

group X (IncX). *Epidemiol Infect* 1999, 122(1):19-22.

[65] Chart H, Row B, Threlfall EJ, Ward LR: **Conversion of *Salmonella enteritidis* phage type 4 to phage type 7 involves loss of lipopolysaccharide with concomitant loss of virulence.** *FEMS Microbiol Lett* 1989, 51(1):37-40.

[66] van Belkum A, Tassios PT, Dijkshoorn L, Haeggman S, Cookson B, Fry NK, Fussing V, Green J, Feil E, Gerner-Smidt P *et al*: **Guidelines for the validation and application of typing methods for use in bacterial epidemiology.** *Clin Microbiol Infect* 2007, 13 Suppl 3:1-46.

[67] Nadon CA, Trees E, Ng LK, Moller Nielsen E, Reimer A, Maxwell N, Kubota KA, Gerner-Smidt P, Group MHW: **Development and application of MLVA methods as a tool for inter-laboratory surveillance.** *Euro Surveill* 2013, 18(35):20565.

[68] Bertrand S, De Lamine de Bex G, Wildemauwe C, Lunguya O, Phoba MF, Ley B, Jacobs J, Vanhoof R, Mattheus W: **Multi locus variable-number tandem repeat (MLVA) typing tools improved the surveillance of *Salmonella Enteritidis*: a 6 years retrospective study.** *PLoS One* 2015, 10(2):e0117950.

[69] Boxrud D, Pederson-Gulrud K, Wotton J, Medus C, Lyszkowicz E, Besser J, Bartkus JM: **Comparison of multiple-locus variable-number tandem repeat analysis, pulsed-field gel electrophoresis, and phage typing for subtype analysis of *Salmonella enterica* serotype Enteritidis.** *J Clin Microbiol* 2007, 45(2):536-543.

[70] Ziebell K, Chui L, King R, Johnson S, Boerlin P, Johnson RP: **Subtyping of Canadian isolates of *Salmonella Enteritidis* using Multiple Locus Variable Number Tandem Repeat Analysis (MLVA) alone and in combination with Pulsed-Field Gel**

Electrophoresis (PFGE) and phage typing. *J Microbiol Methods* 2017, 139:29-36.

[71] Nadin-Davis S, Pope L, Chmara J, Duceppe MO, Burke T, Devenish J, Andrievskaia O, Allain R, Ogunremi D: **An unusual *Salmonella Enteritidis* strain carrying a modified virulence plasmid lacking the *prot6e* gene represents a geographically widely distributed lineage.** *Front Microbiol* 2020, 11:1322.

[72] Olson AB, Andrysiak AK, Tracz DM, Guard-Bouldin J, Demczuk W, Ng LK, Maki A, Jamieson F, Gilmour MW: **Limited genetic diversity in *Salmonella enterica* serovar Enteritidis PT13.** *BMC Microbiol* 2007, 7:87.

[73] Guard J, Morales CA, Fedorka-Cray P, Gast RK: **Single nucleotide polymorphisms that differentiate two subpopulations of *Salmonella enteritidis* within phage type.** *BMC Res Notes* 2011, 4:369.

[74] Leekitcharoenphon P, Nielsen EM, Kaas RS, Lund O, Aarestrup FM: **Evaluation of whole genome sequencing for outbreak detection of *Salmonella enterica*.** *PLoS One* 2014, 9(2):e87991.

[75] Dallman T, Inns T, Jombart T, Ashton P, Loman N, Chatt C, Messelhaeusser U, Rabsch W, Simon S, Nikisins S *et al*: **Phylogenetic structure of European *Salmonella Enteritidis* outbreak correlates with national and international egg distribution network.** *Microb Genom* 2016, 2(8):e000070.

[76] den Bakker HC, Allard MW, Bopp D, Brown EW, Fontana J, Iqbal Z, Kinney A, Limberger R, Musser KA, Shudt M *et al*: **Rapid whole-genome sequencing for surveillance of *Salmonella enterica* serovar Enteritidis.** *Emerg Infect Dis* 2014, 20(8):1306-1314.

- [77] Pornsukarom S, van Vliet AHM, Thakur S: **Whole genome sequencing analysis of multiple *Salmonella* serovars provides insights into phylogenetic relatedness, antimicrobial resistance, and virulence markers across humans, food animals and agriculture environmental sources.** *BMC Genomics* 2018, **19**(1):801.
- [78] van den Berg RR, Dissel S, Rapallini M, van der Weijden CC, Wit B, Heymans R: **Characterization and whole genome sequencing of closely related multidrug-resistant *Salmonella enterica* serovar Heidelberg isolates from imported poultry meat in the Netherlands.** *PLoS One* 2019, **14**(7):e0219795.
- [79] Moreno Switt AI, den Bakker HC, Cummings CA, Rodriguez-Rivera LD, Govoni G, Raneiri ML, Degoricija L, Brown S, Hoelzer K, Peters JE *et al*: **Identification and characterization of novel *Salmonella* mobile elements involved in the dissemination of genes linked to virulence and transmission.** *PLoS One* 2012, **7**(7):e41247.
- [80] Toro M, Retamal P, Ayers S, Barreto M, Allard M, Brown EW, Gonzalez-Escalona N: **Whole-genome sequencing analysis of *Salmonella enterica* serovar Enteritidis isolates in Chile provides insights into possible transmission between gulls, poultry, and humans.** *Appl Environ Microbiol* 2016, **82**(20):6223-6232.
- [81] Ibrahim GM, Morin PM: ***Salmonella* serotyping using whole genome sequencing.** *Front Microbiol* 2018, **9**:2993.
- [82] Yoshida CE, Kruczkiewicz P, Laing CR, Lingohr EJ, Gannon VP, Nash JH, Taboada EN: **The *Salmonella* In Silico Typing Resource (SISTR): An open web-accessible tool for rapidly typing and subtyping draft *Salmonella* genome assemblies.** *PLoS One* 2016, **11**(1):e0147101.
- [83] Zhang S, Yin Y, Jones MB, Zhang Z, Deatherage Kaiser BL, Dinsmore BA, Fitzgerald C, Fields PI, Deng X: ***Salmonella* serotype determination utilizing high-throughput genome sequencing data.** *J Clin Microbiol* 2015, **53**(5):1685-1692.
- [84] Franklin K, Lingohr EJ, Yoshida C, Anjum M, Bodrossy L, Clark CG, Kropinski AM, Karmali MA: **Rapid genoserotyping tool for classification of *Salmonella* serovars.** *J Clin Microbiol* 2011, **49**(8):2954-2965.
- [85] Zhang S, den Bakker HC, Li S, Chen J, Dinsmore BA, Lane C, Lauer AC, Fields PI, Deng X: **SeqSero2: Rapid and improved *Salmonella* serotype determination using whole-genome sequencing data.** *Appl Environ Microbiol* 2019, **85**(23).
- [86] Maiden MC, Bygraves JA, Feil E, Morelli G, Russell JE, Urwin R, Zhang Q, Zhou J, Zurth K, Caugant DA *et al*: **Multilocus sequence typing: a portable approach to the identification of clones within populations of pathogenic microorganisms.** *Proc Natl Acad Sci U S A* 1998, **95**(6):3140-3145.
- [87] Zhou Z, Alikhan NF, Mohamed K, Fan Y, Agama Study G, Achtman M: **The Enterobase user's guide, with case studies on *Salmonella* transmissions, *Yersinia pestis* phylogeny, and *Escherichia* core genomic diversity.** *Genome Res* 2020, **30**(1):138-152.
- [88] Nadon C, Van Walle I, Gerner-Smidt P, Campos J, Chinen I, Concepcion-Acevedo J, Gilpin B, Smith AM, Man Kam K, Perez E *et al*: **PulseNet International: Vision for the implementation of whole genome sequencing (WGS) for global food-borne disease surveillance.** *Euro Surveill* 2017, **22**(23):30544.
- [89] Burnett E, Ishida M, de Janon S, Naushad S, Duceppe MO, Gao R,

Jardim A, Chen JC, Tagg KA, Ogunremi D *et al*: **Whole-Genome Sequencing reveals the presence of the *bla*CTX-M-65 gene in extended-spectrum β -Lactamase-producing and multi-drug-resistant clones of *Salmonella* serovar Infantis isolated from broiler chicken environments in the Galapagos islands.** *Antibiotics* 2021, 10(3):267.

[90] Achtman M, Zhou Z, Alikhan NF, Tyne W, Parkhill J, Cormican M, Chiou CS, Torpdahl M, Litrup E, Prendergast DM *et al*: **Genomic diversity of *Salmonella enterica* - The UoWUCC 10K genomes project.** *Wellcome Open Res* 2020, 5:223.

[91] Petkau A, Mabon P, Sieffert C, Knox NC, Cabral J, Iskander M, Iskander M, Weedmark K, Zaheer R, Katz LS *et al*: **SNVPhyl: a single nucleotide variant phylogenomics pipeline for microbial genomic epidemiology.** *Microb Genom* 2017, 3(6):e000116.

[92] Dallman T, Ashton P, Schafer U, Jironkin A, Painset A, Shaaban S, Hartman H, Myers R, Underwood A, Jenkins C *et al*: **SnapperDB: a database solution for routine sequencing analysis of bacterial isolates.** *Bioinformatics* 2018, 34(17):3028-3029.

[93] Treangen TJ, Ondov BD, Koren S, Phillippy AM: **The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes.** *Genome Biol* 2014, 15(11):524.

[94] Gardner SN, Slezak T, Hall BG: **kSNP3.0: SNP detection and phylogenetic analysis of genomes without genome alignment or reference genome.** *Bioinformatics* 2015, 31(17):2877-2878.

[95] Kircher M, Kelso J: **High-throughput DNA sequencing-concepts and limitations.** *Bioessays* 2010, 32(6):524-536.

[96] Xu Z, Hao B: **CVTree update: a newly designed phylogenetic study platform using composition vectors and whole genomes.** *Nucleic Acids Res* 2009, 37(Web Server issue):W174-178.

[97] Allard MW, Luo Y, Strain E, Pettengill J, Timme R, Wang C, Li C, Keys CE, Zheng J, Stones R *et al*: **On the evolutionary history, population genetics and diversity among isolates of *Salmonella Enteritidis* PFGE pattern JEGX01.0004.** *PLoS One* 2013, 8(1):e55254.

[98] Taylor AJ, Lappi V, Wolfgang WJ, Lapierre P, Palumbo MJ, Medus C, Boxrud D: **Characterization of foodborne outbreaks of *Salmonella enterica* serovar Enteritidis with Whole-Genome Sequencing single nucleotide polymorphism-based analysis for surveillance and outbreak detection.** *J Clin Microbiol* 2015, 53(10):3334-3340.

[99] Feasey NA, Hadfield J, Keddy KH, Dallman TJ, Jacobs J, Deng X, Wigley P, Barquist L, Langridge GC, Feltwell T *et al*: **Distinct *Salmonella Enteritidis* lineages associated with enterocolitis in high-income settings and invasive disease in low-income settings.** *Nat Genet* 2016, 48(10):1211-1217.

[100] Ogunremi D, Devenish J, Amoako K, Kelly H, Dupras AA, Belanger S, Wang LR: **High resolution assembly and characterization of genomes of Canadian isolates of *Salmonella Enteritidis*.** *BMC Genomics* 2014, 15:713.

[101] Kumar S, Stecher G, Li M, Knyaz C, Tamura K: **MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms.** *Mol Biol Evol* 2018, 35(6):1547-1549.