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Chapter

Chemical Biology Toolsets for Drug Discovery and Target Identification

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Abstract

Chemical biology is the scientific discipline that deals with the application of chemical techniques and often small molecules produced through synthetic chemistry, to the manipulation and study of biological systems. Its working framework ranges from simple chemical entities to complex drugs by employing the principles of biological origin. This chapter particularly focuses on the principles and working models of chemical biology to discover new drug leads. Drug discovery is an extensive and multifaceted complex process. Chemical biology uses both natural and synthetic compounds with the best therapeutic potential and verifies them by employing the best possible chemical toolsets. Screening of compounds is done by the use of phenotypic as well as the target-based screening to identify and characterize the potent hits. After the identification of target, it is characterized, and validated by extensive testing. The next step is the validation of hits obtained, and lead compounds are tested in clinical trials before introducing them for commercial application.

Keywords: chemical biology, drug discovery, target identification, target validation, phenotypic screening

1. Introduction to chemical biology and history

Chemical biology flourished as a discipline of science which makes use of several aspects of chemistry to understand biology [1]. Chemical biology includes a wide range of fundamental problems related to the understanding of complex biological processes by the development of synthetic frameworks to generate selective and active lead compounds [2].

The roots of history of chemical biology lie in the emergence of chemistry and biology as separate disciplines. Chemical biology flourished as a separate discipline of science because of newer challenges and questions for the study of chemical methods employed on living bodies. This branch of study is concerned with advanced molecular concepts of biology harnessed to the use of chemical entities. In spite of the newness of this concept, the history of chemical biology extends up to two centuries, considering the foundations of chemistry and biology. Here only a brief account of history of chemical biology is discussed. Joseph Priestley

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discovered nitrous oxide gas in 1772 and incubated the mice with "airs" (the gases discovered till that time). He used 10 gases including nitrous oxide on experimental mice. His experiments on mice faced a strong mass discontent from Americans who showed a sympathetic behavior towards animal rights. Thus, the first chemical biologist fell a prey to angry mob due to his experiment on mice [3].

Afterwards, another chemist, Humphry Davy, worked (1778–1829) on the newly isolated and unfamiliar gases at that time. Frightened by the previous experiment, Humphry completely omitted the use of mice and decided to carry out the research on himself. It was not a matter of surprise that one of the gases, carbon monoxide, proved fatal for the scientist, but the pleasant effect of nitrous oxide made him name this gas, "the laughing gas." He also investigated the use of this gas in medical surgeries. Samuel Taylor also documented this gas as a pleasure-making gas [4], but the practical use of this gas in medicine was described in 1844 by an American



Figure 1. History of chemical biology with its eminent events.

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dentist, Horace Wells [5]. In 1998, three scientists, namely, Ferid Murad, Robert Furchgott, and Louis Ignarro, won Nobel Prize for the demonstration of significant role of nitric acid in cell signaling [6].

Wöhler is a well-known scientist in the history of chemical biology. He attempted to lay the basis of chemical biology by carrying out his research on vitalism. He prepared urea from inorganic chemicals and rejected the famous "vital force theory" in 1828 [7]. The next important event in the history of chemical biology "cellular imaging" was revolutionized by utilizing the chemical approaches during the nine-teenth century. John Hershel invented the cyanotype process which was brought into practice by Anna Atkins to prepare delicate botanical specimens. This noble lady also published her book entitled as *Photographs of British Algae: Cyanotype Impressions* [8].

Ehrlich (1854–1915) is thought to be the pioneer of the earliest forms of chemotherapy and drug therapy. He carried out numerous experiments on aniline based dyes and proposed the idea of "magic bullets." He said that these magic bullets are capable of targeting specific pathogens. He discovered a chemical compound Salvarsan, a drug used against syphilis. This compound is also called as Ehrlich's 606th compound, it was named so because of the successful compound he discovered after 605 failed target compounds. The discovery of this compound paved a way for the discovery of new chemical entities or the new "magic bullets" [6, 9] (**Figure 1**).

Chemical biology flourished as an eminent scientific discipline due to significant contributions of Koehler (pioneer of various chemical screening approaches), Saghatelian (discovery and characterization of lipids and peptides), Wang (use of chemoproteomics in determination of electrophilically lipidated cellular proteins), and Patti and Northen (metabolomics analysis) [1].

2. Chemical biology tools

2.1 Chemical probes

Chemical probes are the small molecules which bind to the specific targeted sites and initiate their cellular activities. These archetypal tools act as highly valued reagents for molecular- and genetic-level biological research. Chemical probes are helpful in the accurate investigation of biological pathways and their associated targets [10].

2.2 Antisense and RNAi technologies

Many tools have been involved in target validation since the 1980s. Target identification and validation are long procedures. They were mainly based on structureactivity relationship. The drug discovery system becomes the most important approach towards the targeted cells [11]. Traditional antisense and RNA interference (RNAi) technologies are the robust tools used in multidimensional phases to discover and validate the potential drug targets. This approach elaborates the potentially selective cleavage of a targeted messenger RNA. This targeting technique enables the researchers to explore the protein-based expression on phenotypes [12].

2.3 Protein degradation strategies

2.3.1 Induced protein degradation

Induced protein degradation is an event-driven approach which depends on drug binding and eliminating the target protein after tagging it. This approach is gaining attention in recent times because of the selective degradation of the target proteins. Drug discovery based on small molecules focuses on the loss of function of proteins due to the already-occupied binding sites ultimately making the proteins unable to target. In this approach, there is a need of high drug exposure in vivo to avoid target inhibition conditions which may lead to potentially harmful side effects of that drug. Proteolysis-targeting chimeras (PROTACS) use the cellular quality control setup to degrade the selective proteins as their targets. This protein degradation system reduces the quantity of drug to be exposed to the living systems which are to be used for halting the protein functions. These proteins may belong to regulatory proteins, transcription factors, and scaffolding proteins [13, 14].

2.3.2 Chemoproteomics

Chemoproteomics is employed as a chemical tool for target identification. It can be used to investigate the signal transductions. This particular field of study has flourished as a key technology to characterize the action mechanism of chemical probes and drugs which can act as pharmacological modulators, hence validating the cellular targets of several therapeutic drug candidates. Chemoproteomics can be further characterized as affinity- and activity-based chemical proteomics [15]. In some cases when probe development is a difficult task, multiple kinase inhibitors are used for targeting the kinome effectively [16].

3. Drug discovery

Drug discovery is a hectic multistep procedure comprising of highly systematic approaches to identify, and characterize different compounds leading towards the development of hits and validate them extensively via utilization of chemical toolsets to attain the status of a commercial therapeutic drug status. The important steps of drug discovery are mentioned in **Figure 2**.

3.1 Screening

There are two fundamental approaches which can be used for the purpose of drug discovery, namely, phenotypic screening and target-based screening.



Figure 2.

A diagram representing the summary of key notes regarding drug discovery from natural products.

The first one looks at the effects of phenotype that the compound induces on cell, tissue or whole organism, and the second one evaluates the effects of a compound on a purified target protein.

3.1.1 Phenotypic screening

In the early twentieth century, drug development started with the advancements in pharmacology and synthetic and therapeutic chemistry. In the 1950s and 1960s, enzyme kinetics has provided methods for accurate computation of compound's effectiveness and enzyme competence [17].

Between 1999 and 2008, the US Food and Drug Administration (FDA) approved new drug discovery approaches. During this period, 75 small molecules were discovered and analyzed. Out of these, 28 drugs were discovered through phenotypic selection, and 17 drugs were identified by target dependent selection [18].

"Alemtuzumab" was the first antibody that was been obtained by using hybridoma technology in combination with phenotypic identification. It was previously reported against relapse of multiple sclerosis and chronic lymphocytic leukemia (CLL). The CD44 antigen (cell surface glycoprotein) antagonist, RG7356, was isolated with the help of function F.I.R.S.T[™] platform. Therefore, functional assays antibodies were used to check effects on cell signaling, proliferation, and programmed cell death [19].

Large combinatorial antibody libraries are the sources of human monoclonal antibodies, successfully used in medical and phenotypic screening. For example, BI-505 was isolated by using F.I.R.S.T[™] platform. Improved versions of antibodies were ultimately used in simulation studies of tumor cell death assay and for selective B-lymphoma cell surface binding. Soon after the isolation of BI-505, its molecular target was identified as ICAM-1, which were found to be involved in apoptosis of B-lymphoma cells. BI-505 has a broad antimyeloma activity [20].

By using phenotypic screening technology, patients can increase their effective antibody response like B-cell repertoire. For example, from a healthcare worker, anti-respiratory syncytial virus (RSV) antibody, D25, was isolated. On the virus coat, D25 neutralizes RSV, and perfusion structure of the F protein was expressed which was not identified by target-based screening [21]. The use of phenotypic screening in various experiments is outlined in **Table 1**.

Disease	Cells	Assay type	Time duration	References
Breast cancer	MCF7-RFP MDA-RFP	Cytochemical and immunohistochemical staining analyses	8–10 days	[22]
Idiopathic pulmonary fibrosis	Alveolar epithelial type II cells	Immunofluorescence staining for in vitro, Western blot, FACs, ELISA, in vitro biochemical kinase assay, migration assay	13 days	[23]
Respiratory papillomatosis	Lung tumor cells	Cell viability assay	48 hours	[24]
Cystic fibrosis	Bronchial epithelial cell	Western blots	18–24 days	[25]
Huntington's disease	PC12	Protease release assay	48 hours	[26]
Familial dysautonomia	Neural crest precursors	RT-PCR assay	48 hours	[27]

Table 1.

Phenotypic screening used in some experiments.

3.1.2 Target-based screening

Target-based screening of natural compounds and synthetic chemicals is being considered as a significant innovation for anticancer drug development [28]. In 2007, Lysine demethylase 5B (KDM5B) and Histone demethylase were recognized, which are liable for the removal of H3K4me2/3 activation marker. Thus, for cancer therapy, KDM5B is regarded as a promising drug target, but the elevated levels of KDM5B were found in many human cancers [29].

The respiratory chain of *Streptococcus agalactiae* consists of two enzymes; type 2-NADH dehydrogenase (NDH-2) and cytochrome *bd* oxygen reductase. *S. agalactiae* is considered as the primary cause of sepsis and meningitis in neonates as well as considerable cause of pneumonia and urinary tract infection [30]. The difference between phenotypic and target-based screening is shown in **Figure 3**.

Some of the target-based screening methods are mentioned as follows.

3.1.2.1 Mass spectrometry-based method

Mass spectrometry is known to be a highly efficient technique for the identification and structural characterization of natural products derived from herbal medicine [31].

Target-based method relies on mass spectrometry to search for active compounds, and this technology can be used for identification, structural characterization, quantitative elemental analysis, tracking of key intermediate compounds in a chemical reaction, analysis of pharmaceuticals and metabolites, and elucidation of unknown structures in drug development. All these achievements can be finally used in various applications like pharmaceutics (drug developments, pharmacokinetics, metabolic pathways), clinical screening, etc. On the basis of MS data information of compounds, the UniFi[™] platform has been built for more detailed analysis of structures [32].

3.1.2.2 Liquid chromatography-mass spectrometry (LC-MS)

LC-MS is an analytical technique for separating different complex mixtures into their components using liquid chromatography. These assays check the correct synthesis, purity, various physical and chemical properties like their volatility and active functionalities present in the newly synthesized chemical entities [33]. During drug discovery, LC-MS hyphenated technique is used for seperation and structural characterization of compounds [34].



Figure 3.

The action potential of phenotypic as well as target-based screening of compounds to validate the hits and leads from natural and synthetic compounds.

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3.1.2.3 Gas chromatography-mass spectrometry (GC-MS)

GC-MS is another hyphenated technique for the identification and structure elucidation of unknown compounds derived from natural products [35]. For example, by using GC-MS technique, comprising a gas chromatograph (GC) coupled to a mass spectrometer (MS), complex components of natural oils mixtures may be separated, identified, and quantified, e.g., oils extracted from Apiaceae family (*Anethum* graveolens, Carum carvi, Cuminum cyminum, Coriandrum sativum, Pimpinella anisum, Daucus carota, Apium graveolens, Foeniculum vulgare, and Ammi visnaga). As a result of this separation technique, petroselinic acid was the major fatty acid from all other palmitic, palmitoleic, stearic, petroselinic, linoleic, linolinic, and arachidic acids [36].

3.1.2.4 Ultra-performance liquid chromatography-mass spectrometry (UPLC-MS)

Currently, UPLC-MS is one of the most adaptable hyphenated techniques. Proteomics and metabolomics have proved to be useful concepts for understanding the causes of different diseases. This technology aims to seperate and identify proteins and metabolites for cellular signaling pathways and to discover biomarkers for screening and diagnosis as well as determining response to a specific treatment [37]. For example, vancomycin (VCM) is clinically used for the treatment of human intracranial infections. The treatment concentration of vanomycin greatly varies among the patients. UPLC-MS technique was developed and used for the analysis of VCM in human cerebrospinal fluid [38].

3.1.2.5 Nuclear magnetic resonance spectroscopy (NMR)

Among the common techniques of metabolomics, NMR has evolved the most. Unlike mass spectroscopy, NMR is also used for quantitative analysis, but it does not require extra steps for sample preparation [39]. It is commonly used to analyze the 3D structures of biomacromolecules and their interactions. It has been proved a valuable tool for the reliable identification of small molecules that bind to proteins and for hit-to-lead optimization. Mainly, NMR spectroscopy is suitable for the analysis of bulk metabolites [40]. NMR has been used for analyzing the structure of protein, nucleic acid, and small molecule [41]. NMR has been proven to be a useful tool in target-based drug discovery in the step of hit identification and lead optimization [42]. For example, NMR spectroscopy is used to understand the structure of G-quadruplexes, which are noncanonical, four standard nucleic acids with consecutive sequences of guanines [43].

3.1.2.6 Thermal shift or calorimetry-based method

Isothermal titration calorimetry (ITC) is the only technique which is currently available for the direct determination of enthalpy, ΔH , of a ligand binding to a protein [44]. Thermodynamic evaluation might be useful to provide information about specificity, agonist versus antagonist effects of ligands, and other important properties [45]. Fragment-based drug discovery (FBDD) is an approach of particular interest and relevance here. Fragments are molecules smaller than typical drugs, and they generally bind with lower affinity than conventional drug screening hits [46]. Measuring the contributions of enthalpy and entropy to the free energy of binding provides information that can be useful in fragment elaboration and subsequent medicinal chemistry work [47]. ITC is a uniquely powerful tool for characterization of the thermodynamics of test compounds binding to target proteins. Interaction between the compound and protein leads to release or uptake of small amounts of heat, while the mixture is held at a close approximation to

Target base screening		(Features		Phenoty	pic screening
Advantage	Dis Advantage		Molecular		Advantage	Dis Advantage
Known	Have to know	ta	arget of a disea	se	Do not need to know	Unknown
Higher, Relatively easy to set up	Assay may be less biologically relevant		Screening throu put and assay	gh /	Medium or low, biologically relevant	Could be low; could have higher cost
Known at onset, which can accelerate preclinical drug development	Limited possibility of identifying a new mechanism	of	Mechanism of action	(Multiple targets and signaling pathways can be targeted	Unknown at onset
Direct binding assay, modeling, X-ray crystallography	Need to be confirmed in cell-based and phenotypic assays with native target	ł	Methods for confirmation	\wedge	Can move to in vivo study quickly	Target identification; can be complicated and time consuming
Readily available and direct			Methods for SAR optimization		Additional assays may need to support SAR	May need to develop a more targeted assay
Direct if it is relevant	May be not disease -relevant		Disease relevance		Usually disease- relevant	
	Limited by the hypothesis		Hypothesis limitation		Less hypothesis -restricted	

Figure 4.

Comparison between the advantages and disadvantages of target-based and phenotypic screening based upon the different features such as molecular target of disease, its mechanism of action, confirmation methods, SAR optimization methods, and hypothesis limitation.

constant temperature [48]. Thermal shift screening methods has allowed to identify compounds that interact with *Trypanosoma brucei* choline kinase (TBCK) and inhibit TBCK, a validated drug target against African sleeping sickness [49].

3.1.2.7 Affinity-based methods

The methods regarding affinity-based immobilized proteins have vital role in understanding the connections between small molecules and their biological targets [50]. Affinity-based technologies are divided into two groups: (1) direct detection of noncovalent macromolecule-ligand complex and (2) indirect detection of noncovalent macromolecule-ligand complex. The negative aspect of this approach is that it recognizes chemical entities basically based on their binding affinities for a target irrespective of whether or not the biological function of the target is affected. In the late 1980s, matrix-assisted laser desorption ionization (MALDI) and electrospray ionization (ESI) techniques were used to analyze proteins and nucleic acids. Both phenotypic screening and target-based screening are comparable to each other in terms of benefits and drawbacks. This fact has been illustrated in **Figure 4**.

4. Target identification and characterization

Target identification and elucidation of its action mechanism have played vital roles in probing small molecules and drug discovery. Target identification has been based on biological and technologically advanced cell-based assays [51].

4.1 Disease association and target validation

Identification of the molecules and their underlying pathophysiological mechanisms contribute towards the discovery of targets that can be modulated therapeutically [52]. Each drug target is linked to a disease using integrated genome-wide data from a broad range of data sources. The target validation reveals the evidence that associates a target with a disease [53].

4.2 Bioactive small molecules

Bioactive small molecules are preferred as lead structures for the target validation. These small molecules isolated from phenotypic screen play a crucial role in chemical biology [54, 55]. Many genomic, proteomic, and bioinformatic technologies have been developed for validation of the drugs.

4.3 Protein interactions

To identify the selective potent drugs, the first step is to find the protein interference. In signal transductions, protein-protein interactions are involved in the complex cellular networks that govern the different processes [56]. The deregulated transcription factors are involved in playing significant roles in human pathological abnormalities, but the complicated nature of protein-protein networks has made the transcription-targeted therapeutics impractical. Recent technological advancements are the ray of hope regarding the modulation of protein interaction networks [57].

4.4 Cell-based models and target validation

Exosomes are highly adequate for drug carriers as a cell-based model. Due to the association of multiple proteins with cellular membranes, the exosomes are well-known in cell to cell communication, and they are the novel approach for the delivery of potent drugs. Exosome-based drug technique is applied for a variety of disorders such as cancer and various neurodegenerative disorders [58].

5. Target validation

Drug target discovery and validation demand complicated and expensive frameworks which may pose heavy financial load on pharmaceutical industry. Target validation is referred to as the direct involvement of a certain molecular target in pathological conformity; hence, its reversal or inflection may have a therapeutic effect [12].

5.1 Approaches to target validation

The following approaches are used in target validation during the discovery and development of drug.

5.1.1 Antibodies

Firstly access the antibody fitness towards a specific target. Then, standardized procedures are obligatory to ensure the quality of the sample in test procedures; hence, utilizing only a single approach will not work in all situations [59]. Mass spectrometry is used to identify the validation of the antibody. This type of technique confirms the validity for antibodies or their fragments against the targets. The antibody is able to bind to its natural antigen in cell lysates among thousands of other proteins, DNA, RNA, and other cellular components [60].

5.1.2 Cellular thermal shift assay (CETSA)

CETSA is used to assess the capability of a ligand to bind with its targets (cells or tissue samples). The basis of this method lies on the ligand-induced thermodynamic

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stabilization of target proteins. The compound-treated cell lysates and intact cells were heated to different temperatures, and in the soluble fractions, the target protein was separated from destabilized protein and detected by Western blotting. SPROX is a method of target validation based on identification of ligand-induced stabilization of target proteins. It evaluates the levels of methionine oxidation of target proteins [61].

5.1.3 Drug affinity responsive target stability (DARTS)

DARTS has been used for the identification of the targeted proteins. It is based on ligand binding interaction with proteins forming a complex which changes the structural stability of target protein. There alteration is measured by SDS page/ liquid chromatography. DARTS is also involved in the analysis of the low affinity interactions [61].

6. Hit generation

Hit identification is considered as the significant bottleneck for lead generation success and for new medicines. An example for random hit identification is physical and biochemical testing [62]. The journey of a compound from the hit status to lead status follows a series of steps which have been briefly illustrated in **Figure 5**. The figure describes a note of possible techniques which could be utilized for the selection of lead compounds and proceeding them through lead optimization preclinical and clinical phase trials.



Figure 5.

A diagram elaborating the significant steps of lead optimization proceeding to clinical phase of natural compounds.

7. Development of lead drug

Pharmaceutical companies are facing constant economic pressure to bring efficacy in drug discovery and development process. Lists of compounds obtained after hit optimization are further subjected to refining process in order to find out the lead compounds that can be analyzed for production at commercial scale. During this "hitto-lead" refining process, many compounds are dropped out due to inadequate absorption, distribution, metabolism, excretion, and toxicity/ADMET characteristics [63].

Refining of hit compounds to lead compound is done through the process of secondary screening. Almost 50% of all drug candidates thin out during optimization and preclinical and clinical trials [64].

There are many approaches available for the discovery and development of drug which might follow different pathways to optimize the compounds into bioavailable drugs. All these pathways must have a common origin; they all begin with a lead compound. It is necessary to go through the phylogeny study of all the compounds because there are some properties like solubility, target affinity, toxicity, ease of synthesis, and bioavailability, all of which are highly dependent on the initial lead selection and the method of identification [65].

7.1 Techniques of lead selection

A rational approach is used to select lead drug candidate after optimization of hit compounds. There are many methods which can be used for screening of compounds. Selection of techniques depends upon the source of hit compounds and types of their solvents as well. The following techniques are useful in selection.

7.1.1 QSAR model development

Quantitative structure-activity relationship model is used to compare chemical structures by using database of prior selected active compounds. Different software like ChemBioOffice Ultra 1.11 is used to generate two-dimensional and three-dimensional structures. The results of QSAR can be validated by using statistical approaches like correlation coefficient and regression coefficient [66].

7.1.2 Visualization of SAR activity

It is called as Bayesian approach. It provides with proficient understanding of shape features, hydrophobic nature, and electrostatic properties of the compounds. All of these features lie under the structure–activity relationship of selected compounds from hits. Structure data analysis of SAR is obtained in 3D form. Other results are obtained in diverse type of interrelated biochemical data, i.e., average of activities and region explored analysis. The results obtained from average activity show a common part in active compounds, and region explored data exhibit the areas of fully explored compounds [67].

7.1.3 Fragment-based drug discovery

It is a powerful method which is used to find out the proportion of ligands with high affinity to target proteins. The compounds which are found to have low ligand binding ability are eliminated, and the compounds with high ligand ability move forward to the precision of compounds. FBDD consists of the techniques such as NMR, SAR, X-ray crystallography, and surface plasmon resonance (SPR).

7.1.3.1 X-ray crystallography

It can ascertain the binding sites and modes of ligand binding to protein [68].

7.1.3.2 Surface plasmon resonance (SPR)

Surface plasmon resonance is known as a nonlabel technology that can identify, screen, and quantify intermolecular interactions in actual time. It is applied to quantify binding affinities. SPR-dependent biosensors work by detecting the ligands and immobilized target molecular interactions and supply appropriate information on kinetics of biomolecular interactions. The output information can be utilized to provide comprehensive functional data on binding actions such as specificity, kinetics, concentration, and affinity [69]. Scientific literature study revealed Biacore tools as mainly used SPR technology at commercial levels [70].

7.2 Preclinical trials

In the last 2 years, different methodologies based on high-throughput screening and their combinations with chemistry have been developed in order to manufacture versatile compounds by limiting the resources. Among these methodologies, several other in vitro and in silico supplementary approaches have also come forward for the identification and potential evaluation of these compounds as lead candidate validation. Those compounds which are selected as "hits" during this screening procedure are further analyzed and subjected to in vivo toxicity and efficacy profiling. During preclinical stage of drug development, simple formulation approaches are favored. Combinatorial chemistry and high-throughput approaches have been appraised in several publications [71].

PLOTs are preclinical lead optimization technologies that should be rapid enough to edge with high-throughput discovery screenings without causing further delay and should be predictive and cost-effective. PLOT platform usually comprised of in vitro systems, small and acquiescent to mechanization, and that is why it is easy to achieve the mandatory throughput with minimum use of compound use [72].

7.2.1 Tools of preclinical drug development

Selection of methodology and tools for selection of preclinical drug candidates is a rigorous process. Sequential approach of preclinical to clinical is practiced to sort out the long list of target selected compounds. This streamline strategy provides with deeper understanding of action of the drug prior to its progress to the next steps [73].

7.2.2 Pharmacokinetics and pharmacodynamics (PK/PD) during preclinical drug evaluation

Pharmacodynamics involves the study of effect of drug in dose- and timedependent manner. Pharmacokinetics is the study of absorption, metabolization, distribution, and excretion of a drug over time. PK/PD is a program at early phase of lead drug development which acts as a bridge between drug discovery and preclinical drug development. This stage set aims for further development activities, and information obtained at this stage act as a key to subsequent steps. It is necessary because of the following reasons:

- a. It provides potency-based intrinsic activity of the compound rather than dose.
- b. It characterizes the compounds on the basis of dose concentration and effect relationship.

c. It allows the investigation of tolerance phenomenon of compounds on the basis of physiological parameters [74].

7.3 Lead optimization

Optimization of a drug is a multifaceted process. It usually involves various types of screening methods which tend to find out the metabolism and pharmaco-kinetic properties of selected compounds or drugs [75].

7.3.1 ADME

This is the final stage of preclinical trials; after this the optimized drug is further processed towards the clinical trial. Absorption, distribution, metabolism, and excretion screening is performed at this stage. The primary goal of ADME is to develop a competitive drug with adequate safety avoiding PK failure in clinical phase.

7.3.2 ADME properties

Ideal properties of a drug in ADME testing involve the good oral bioavailability, blood clearance and volume of convenient dosing, and low potential of drug-drug interaction. All of these properties are assessed at early stage of drug discovery [76].

7.3.3 DRUGeff

Drug effect is a parameter which determines the concentration of a drug which do not cause any harm at the site of action. In other words at this stage, toxicity of a drug is tested to find out the minimum safe dosage potency. In vitro DRUGeff testing of all compounds show interaction with the target treatment, until a small portion of dose gets to select according to biophase levels. Concentration of treatment dose maximization per unit of biophase acts as a key objective for lead optimization. The drugs qualifying this test enter into the clinical phase [77].

7.4 Clinical phase of drug discovery

The final step of drug discovery and development is referred to as the clinical trial. At this stage, the data regarding safety and efficacy of the new drug must be proven by application to humans directly in different phases. After the successful trials, research data is sent to the FDA for approval for commercial manufacturing and marketing (**Figure 6**) [78].

7.4.1 Clinical phase I

The first phase of clinical trial normally takes several weeks to some months. At this stage application of optimized drug is tested on a small group of volunteers.



Figure 6. *The journey of potential leads from preclinical to clinical trials.*

They may or may not get paid for their participation in drug trial studies. This mini trial is useful in determining the absorption and side effect of drug in relation to its dose concentration [17].

7.4.2 Clinical phase II

The second phase of clinical trial may last up to 2 years. It is a totally randomized study which involves the application of drug on a relatively large group of patients. This trial study is divided into two groups of patients, one receiving experimental drug and the other receiving placebo. Sometimes it may be named as a blind application trial. This type of random application of drug allows investigators and pharmaceutics to prove the success and safety of drug to the FDA with comparative information [79].

7.4.3 Clinical phase III

It is a large-scale testing of drugs on hundreds of patients. This third stage testing provides with a more thorough understanding and effectiveness of useful drugs to the FDA and pharmaceutical companies. The pharmaceutical company can request for the approval for commercial synthesis of drug after phase III is completed [80].

7.4.4 Clinical phase IV

After the approval of a drug for commercial consumption, clinical phase IV trials are used as post marketing surveillance trials. This trial system is based upon the various objectives at commercial levels, i.e., the comparison of newly approved and already-available drugs in market, to evaluate the chronic effects on patients' quality of life and to estimate the economical comparison of newly approved and already-present drugs as well as the traditional system of medication [81].

8. Conclusion and future perspectives

Chemical biology is an emerging field of science which particularly focuses on the research in biological systems by employing the chemicals and related chemoinformatic tools. This field of study is working well in combination with medicinal and combinatorial chemistry to seek the cure of incurable and life-threatening human pathologies. This chapter illustrated the significant techniques and chemical setups which can be employed to testify the chemical as well as biological aspects of natural and synthetic compounds before introducing them as therapeutic drugs in the field of medicine. There is an ultimate need of the hour to seek for the newer and better drugs which are safer, cheaper, and more effective than the already existing therapeutics. This field of study is flourishing at a very fast pace, and it is anticipated that it will provide better treatment options and strategies in future for the medical practitioners to use the best among the rest drugs discovered.

Conflict of interest

Authors have no conflict of interest.

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References

[1] Saghatelian A, Nomura DK, Weerapana E. Editorial overview: Omics: The maturation of chemical biology. Current Opinion in Chemical Biology. 2016;**30**:v-vi

[2] Ostler EL. Chemical biology is. Chemistry Central Journal. 2007;**1**:5

[3] Priestley J. Experiments and Observations on Different Kinds of Air: And Other Branches of Natural Philosophy, Connected with the Subject. In Three Volumes: Being the Former Six Volumes Abridged and Methodized, with Many Additions. United Kingdom: Thomas Pearson; 1790

[4] Hoover SR. Coleridge, Humphry Davy, and some early experiences with a consciousness-altering drug. Bulletin of Research in the Humanities. 1978;**81**:9-27

[5] Wright AJ. Davy comes to America: Woodhouse, Barton, and the nitrous oxide crossing. Journal of Clinical Anesthesia. 1995;7:347-355

[6] Morrison KL, Weiss GA. The origins of chemical biology. Nature Chemical Biology. 2006;**2**:3-6

[7] Wöhler F. Poggendorff's. Annals of Physical Chemistry. 1828;**12**:253

[8] Atkins A, Chuang J, Schaaf L. Anna Atkins: Photographs of British Algæ. Germany: Gerhard Steidl Druckerei und Verlag; 2020

[9] Miescher F. Die histochemischen und physiologischen Arbeiten von Friedrich Miescher. Leipzig, Germany: Vogel; 1897

[10] Blagg J, Workman P. Choose and use your chemical probe wisely to explore cancer biology. Cancer Cell. 2017;**32**:9-25

[11] Duarte Y, Marquez-Miranda V, Miossec MJ, Gonzalez-Nilo F. Integration of target discovery, drug discovery and drug delivery: A review on computational strategies. Wiley Interdisciplinary Reviews. Nanomedicine and Nanobiotechnology. 2019;**11**:e1554

[12] Lavery KS, King TH. Antisense and RNAi: Powerful tools in drug target discovery and validation. Current Opinion in Drug Discovery & Development. 2003;**6**:561-569

[13] Lai AC, Crews CM. Induced protein degradation: An emerging drug discovery paradigm. Nature Reviews. Drug Discovery. 2017;**16**:101-114

[14] Toure M, Crews CM. Smallmolecule PROTACS: New approaches to protein degradation. Angewandte Chemie. 2016;55:1966-1973

[15] Drewes G, Knapp S. Chemoproteomics and chemical probes for target discovery. Trends in Biotechnology. 2018;**36**:1275-1286

[16] Yao Z, Petschnigg J, Ketteler R, Stagljar I. Application guide for omics approaches to cell signaling. Nature Chemical Biology. 2015;**11**:387-397

[17] King TA, Stewart HL, Mortensen KT, North AJP, Sore HF, Spring DR. Cycloaddition strategies for the synthesis of diverse heterocyclic spirocycles for fragment-based drug discovery. European Journal of Organic Chemistry. 2019;**2019**:5219-5229

[18] Swinney DC, Anthony J. How were new medicines discovered? Nature Reviews. Drug Discovery. 2011;**10**:507-519

[19] Menke-van der Houven van Oordt CW, Gomez-Roca C, van Herpen C, Coveler AL, Mahalingam D, Verheul HM, et al. First-in-human phase I clinical trial of RG7356, an anti-CD44 humanized antibody, in patients with advanced, CD44-expressing *Chemical Biology Toolsets for Drug Discovery and Target Identification* DOI: http://dx.doi.org/10.5772/intechopen.91732

solid tumors. Oncotarget. 2016;**7**:80046-80058

[20] Fransson J, Tornberg UC, Borrebaeck CA, Carlsson R, Frendeus B. Rapid induction of apoptosis in B-cell lymphoma by functionally isolated human antibodies. International Journal of Cancer. 2006;**119**:349-358

[21] McLellan JS, Chen M, Leung S, Graepel KW, Du X, Yang Y, et al. Structure of RSV fusion glycoprotein trimer bound to a perfusion-specific neutralizing antibody. Science. 2013;**340**:1113-1117

[22] Khan GN, Kim EJ, Shin TS, Lee SH. Heterogeneous cell types in single-cell-derived clones of MCF7 and MDA-MB-231 cells. Anticancer Research. 2017;**37**:2343-2354

[23] Fujino N, Kubo H, Maciewicz RA. Phenotypic screening identifies Axl kinase as a negative regulator of an alveolar epithelial cell phenotype. Laboratory Investigation. 2017;**97**:1047-1062

[24] Yuan H, Myers S, Wang J, Zhou D, Woo JA, Kallakury B, et al. Use of reprogrammed cells to identify therapy for respiratory papillomatosis. The New England Journal of Medicine. 2012;**367**:1220-1227

[25] Fulcher ML, Gabriel SE, Olsen JC, Tatreau JR, Gentzsch M, Livanos E, et al. Novel human bronchial epithelial cell lines for cystic fibrosis research. The American Journal of Physiology-Lung Cellular and Molecular Physiology. 2009;**296**:L82-L91

[26] Titus SA, Southall N, Marugan J, Austin CP, Zheng W. High-throughput multiplexed quantitation of protein aggregation and cytotoxicity in a Huntington's disease model. Current Chemical Genomics. 2012;**6**:79-86

[27] Lee G, Ramirez CN, Kim H, Zeltner N, Liu B, Radu C, et al. Largescale screening using familial dysautonomia induced pluripotent stem cells identifies compounds that rescue IKBKAP expression. Nature Biotechnology. 2012;**30**:1244-1248

[28] Yamori T. Chemical evaluation by cancer cell line panel and its role in molecular target-based anticancer drug screening. Cancer Chemotherapy. 2004;**31**:485-490

[29] Rotili D, Mai A. Targeting histone demethylases: A new avenue for the fight against cancer. Genes & Cancer. 2011;**2**:663-679

[30] Yamamoto Y, Pargade V, Lamberet G, Gaudu P, Thomas F, Texereau J, et al. The group B streptococcus NADH oxidase Nox-2 is involved in fatty acid biosynthesis during aerobic growth and contributes to virulence. Molecular Microbiology. 2006;**62**:772-785

[31] Henke MT, Kelleher NL. Modern mass spectrometry for synthetic biology and structure-based discovery of natural products. Natural Product Reports. 2016;**33**:942-950

[32] Wang X, Zhang A, Han Y, Wang P, Sun H, Song G, et al. Urine metabolomics analysis for biomarker discovery and detection of jaundice syndrome in patients with liver disease. Molecular & Cellular Proteomics. 2012;**11**:370-380

[33] Lee MS, Kerns EH. LC/MS applications in drug development. Mass Spectrometry Reviews. 1999;**18**:187-279

[34] Yu S, Li S, Yang H, Lee F, Wu JT, Qian MG. A novel liquid chromatography/ tandem mass spectrometry based depletion method for measuring red blood cell partitioning of pharmaceutical compounds in drug discovery. Rapid Communications in Mass Spectrometry. 2005;**19**:250-254

[35] Spanik I, Machynakova A. Recent applications of gas chromatography

with high-resolution mass spectrometry. Journal of Separation Science. 2018;**41**:163-179

[36] Nguyen T, Aparicio M, Saleh MA. Accurate mass GC/LC-quadrupole time of flight mass spectrometry analysis of fatty acids and triacylglycerols of spicy fruits from the Apiaceae Family. Molecules. 2015;**20**:21421-21432

[37] Zhao YY, Lin RC. UPLC-MS(E) application in disease biomarker discovery: The discoveries in proteomics to metabolomics. Chemico-Biological Interactions. 2014;**215**:7-16

[38] Mei S, Wang J, Zhu L, Chen R, Li X, Chen K, et al. A UPLC-MS/MS method for analysis of vancomycin in human cerebrospinal fluid and comparison with the chemiluminescence immunoassay. Biomedical Chromatography. 2017;**31**:e3939

[39] Emwas AH. The strengths and weaknesses of NMR spectroscopy and mass spectrometry with particular focus on metabolomics research. Methods in Molecular Biology. 2015;**1277**:161-193

[40] Harner MJ, Frank AO, Fesik SW. Fragment-based drug discovery using NMR spectroscopy. Journal of Biomolecular NMR. 2013;**56**:65-75

[41] Billeter M, Wagner G, Wuthrich K. Solution NMR structure determination of proteins revisited. Journal of Biomolecular NMR. 2008;**42**:155-158

[42] Pellecchia M, Sem DS, Wuthrich K. NMR in drug discovery. Nature Reviews. Drug Discovery. 2002;**1**:211-219

[43] Lin C, Dickerhoff J, Yang D. NMR studies of G-Quadruplex structures and G-Quadruplex-interactive compounds. Methods in Molecular Biology. 2019;**2035**:157-176

[44] Wiseman T, Williston S, Brandts JF, Lin LN. Rapid measurement of binding constants and heats of binding using a new titration calorimeter. Analytical Biochemistry. 1989;**179**:131-137

[45] Chaires JB. Calorimetry and thermodynamics in drug design. Annual Review of Biophysics. 2008;**37**:135-151

[46] Hopkins AL, Groom CR, Alex A. Ligand efficiency: A useful metric for lead selection. Drug Discovery Today. 2004;**9**:430-431

[47] Ward WH, Holdgate GA. Isothermal titration calorimetry in drug discovery. Progress in Medicinal Chemistry. 2001;**38**:309-376

[48] Recht MI, De Bruyker D, Bell AG, Wolkin MV, Peeters E, Anderson GB, et al. Enthalpy array analysis of enzymatic and binding reactions. Analytical Biochemistry. 2008;**377**:33-39

[49] Major LL, Denton H, Smith TK. Coupled enzyme activity and thermal shift screening of the Maybridge rule of 3 fragment library against Trypanosoma brucei choline kinase; a genetically validated drug target. In: El-Shemy HA, editor. Drug Discovery. Rijeka (HR): IntechOpen; 2013. pp. 413-431

[50] Temporini C, Brusotti G, Pochetti G, Massolini G, Calleri E. Affinity-based separation methods for the study of biological interactions: The case of peroxisome proliferatoractivated receptors in drug discovery. Methods. 2018;**146**:12-25

[51] Schenone M, Dancik V, Wagner BK, Clemons PA. Target identification and mechanism of action in chemical biology and drug discovery. Nature Chemical Biology. 2013;**9**:232-240

[52] Floris M, Olla S, Schlessinger D, Cucca F. Genetic-driven druggable target identification and validation. Trends in Genetics. 2018;**34**:558-570

[53] Koscielny G, An P, Carvalho-Silva D, Cham JA, Fumis L, Gasparyan R, et al.

Chemical Biology Toolsets for Drug Discovery and Target Identification DOI: http://dx.doi.org/10.5772/intechopen.91732

Open targets: A platform for therapeutic targetidentificationandvalidation.Nucleic Acids Research. 2017;**45**:D985-D994

[54] Velagapudi SP, Gallo SM, Disney MD. Sequence-based design of bioactive small molecules that target precursor microRNAs. Nature Chemical Biology. 2014;**10**:291-297

[55] Jung HJ, Kwon HJ. Target deconvolution of bioactive small molecules: The heart of chemical biology and drug discovery. Archives of Pharmacal Research. 2015;**38**:1627-1641

[56] Rimbault C, Maruthi K, Breillat C, Genuer C, Crespillo S, Puente-Munoz V, et al. Engineering selective competitors for the discrimination of highly conserved protein-protein interaction modules. Nature Communications. 2019;**10**:4521

[57] Mapp AK, Pricer R, Sturlis S. Targeting transcription is no longer a quixotic quest. Nature Chemical Biology. 2015;**11**:891

[58] Batrakova EV, Kim MS. Using exosomes, naturally-equipped nanocarriers, for drug delivery. Journal of Controlled Release: Official Journal of the Controlled Release Society.
2015;219:396-405

[59] Taussig MJ, Fonseca C, Trimmer JS. Antibody validation: A view from the mountains. New Biotechnology. 2018;**45**:1-8

[60] Persson H, Preger C, Marcon E, Lengqvist J, Graslund S. Antibody validation by immunoprecipitation followed by mass spectrometry analysis. Methods in Molecular Biology. 2017;**1575**:175-187

[61] Chang J, Kim Y, Kwon HJ. Advances in identification and validation of protein targets of natural products without chemical modification. Natural Product Reports. 2016;**33**:719-730 [62] Bleicher KH, Bohm HJ, Muller K, Alanine AI. Hit and lead generation: Beyond high-throughput screening. Nature Reviews. Drug Discovery. 2003;**2**:369-378

[63] Nyunoya H, Lusty CJ. The carB gene of Escherichia coli: A duplicated gene coding for the large subunit of carbamoyl-phosphate synthetase. Proceedings of the National Academy of Sciences of the United States of America. 1983;**80**:4629-4633

[64] Lofas S. Optimizing the hit-tolead process using SPR analysis. Assay and Drug Development Technologies. 2004;**2**:407-415

[65] Fejzo J, Lepre CA, Peng JW, Bemis GW, Ajay, Murcko MA, et al. The SHAPES strategy: An NMR-based approach for lead generation in drug discovery. Chemistry & Biology. 1999;**6**:755-769

[66] Floresta G, Rescifina A, Marrazzo A, Dichiara M, Pistara V, Pittala V, et al. Hyphenated 3D-QSAR statistical model-scaffold hopping analysis for the identification of potentially potent and selective sigma-2 receptor ligands. European Journal of Medicinal Chemistry. 2017;**139**:884-891

[67] Alam S, Khan F. QSAR and docking studies on xanthone derivatives for anticancer activity targeting DNA topoisomerase II alpha. Drug Design, Development and Therapy. 2014;**8**:183-195

[68] Erlanson DA, Davis BJ, Jahnke W. Fragment-based drug discovery: Advancing fragments in the absence of crystal structures. Cell Chemical Biology. 2019;**26**:9-15

[69] Liu C, Yang Y, Wu Y. Recent advances in exosomal protein detection via liquid biopsy biosensors for cancer screening, diagnosis, and prognosis. The AAPS Journal. 2018;**20**:41 [70] Kukanskis K, Elkind J, Melendez J, Murphy T, Miller G, Garner H. Detection of DNA hybridization using the TISPR-1 surface plasmon resonance biosensor. Analytical Biochemistry. 1999;**274**:7-17

[71] Ramstrom O, Lehn JM. Drug discovery by dynamic combinatorial libraries. Nature Reviews. Drug Discovery. 2002;**1**:26-36

[72] Atterwill CK, Wing MG. In vitro preclinical lead optimisation technologies (PLOTs) in pharmaceutical development. Toxicology Letters. 2002;**127**:143-151

[73] Boger E, Friden M. Physiologically based pharmacokinetic/ pharmacodynamic modeling accurately predicts the better bronchodilatory effect of inhaled versus oral salbutamol dosage forms. Journal of Aerosol Medicine and Pulmonary Drug Delivery. 2019;**32**:1-12

[74] Ekblom M, Hammarlund-Udenaes M, Paalzow L. Modeling of tolerance development and rebound effect during different intravenous administrations of morphine to rats. Journal of Pharmacology and Experimental Therapeutics. 1993;**266**:244-252

[75] Cheng KC, Korfmacher WA, White RE, Njoroge FG. Lead optimization in discovery drug metabolism and pharmacokinetics/case study: The hepatitis C virus (HCV) protease inhibitor SCH 503034. Perspectives in Medicinal Chemistry. 2007;**1**:1-9

[76] Balani SK, Miwa GT, Gan LS, Wu JT, Lee FW. Strategy of utilizing in vitro and in vivo ADME tools for lead optimization and drug candidate selection. Current Topics in Medicinal Chemistry. 2005;**5**:1033-1038

[77] Braggio S, Montanari D, Rossi T, Ratti E. Drug efficiency: A new concept to guide lead optimization programs towards the selection of better clinical candidates. Expert Opinion on Drug Discovery. 2010;**5**:609-618

[78] Swann PG, Shapiro MA. Regulatory considerations for development of bioanalytical assays for biotechnology products. Bioanalysis. 2011;**3**:597-603

[79] Sartori SB, Singewald N. Novel pharmacological targets in drug development for the treatment of anxiety and anxiety-related disorders. Pharmacology & Therapeutics. 2019;**204**:107402

[80] Regan D, Garcia K, Thamm D. Clinical, pathological, and ethical considerations for the conduct of clinical trials in dogs with naturally occurring cancer: A comparative approach to accelerate translational drug development. ILAR Journal. 2018;**59**:99-110

[81] Stephenson N, Shane E, Chase J, Rowland J, Ries D, Justice N, et al. Survey of machine learning techniques in drug discovery. Current Drug Metabolism. 2019;**20**:185-193

