

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Acidosis and Anion Gap

Md. Masudul Hassan

Abstract

The journey of exploring acid and base starts long before, but much advancement was seen in the last century. In 1890, Wilhelm Ostwald electronically measured hydrogen. Svante Arrhenius won the Noble prize in 1903 for the theory of ionization. In 1908, Henderson and Black showed that bicarbonate and phosphate equilibrate with CO_2 at normal body temperature. In 1923, Bronsted first put forward the idea of acid that ionizes in solution and donate hydrogen and the base accepts the hydrogen from the solution. Handerson invented the important bicarbonate buffer system and Hasselbalch First measured the actual blood pH. In 1909, S. P. S. Sorensen developed the pH scale. Later Hasselbalch-Henderson developed an equation that helped in relating pH to the blood bicarbonate and PCO_2 . Acidosis has fatal consequences like CNS damage and death. Acidosis is rapidly stabilized by the body buffer systems. There are equal amounts of cations and anions in blood, but some of them are unmeasured. These unmeasured ions are mostly anions that produce an anion gap. Increased anion gap usually represents metabolic acidosis. Albumin and many other confounding factors influence the anion gap derangements. Accuracy in measuring anion gap is critically important for the evaluation of acidosis.

Keywords: anion gap, acidosis

1. Introduction

The journey of exploring acid and base starts long before, but in the last century the advancement was remarkable. In 1890, Wilhelm Ostwald electronically measured hydrogen [1]. Svante Arrhenius won the Noble prize in 1903 for the theory of ionization [2]. In 1908, Henderson and Black showed that bicarbonate and phosphate equilibrated with CO_2 at normal body temperature in different solution [3]. In 1923, Bronsted first put forward the idea of acid as a substance that ionizes in solution and donate hydrogen and the base accepts the hydrogen from the solution [4]. Bronsted, Henderson and Van Slyke described acid-base balance in the early part of nineteenth century [5]. Handerson invented bicarbonate as the most important buffer system of the body, and Hasselbalch first measured the real blood pH in the early part of nineteenth century [6–8]. In 1909, S. P. S. Sorensen developed the pH scale [8]. Later Hasselbalch-Henderson developed an equation that helped in relating pH to the blood bicarbonate and PCO_2 [7, 9, 10]. In the early 1980s, scientists introduced electrodes specific for each ion. Thereafter, serum electrolyte and the anion gap measurement become routine tools for assessing acidosis.

Acidosis has fatal consequences like CNS damage. Even death is not uncommon. Acidosis is characterized by a decrease in pH, and this change is rapidly

corrected by the body buffer systems. Many clinical conditions develop acidosis, as well as ionic derangements and the only correction of the underlying cause can resolve it. There are equal numbers of cations and anions in the blood and among them there are some unmeasured anions. These unmeasured anions can contribute in the clinically important anion gap. In a healthy individual, there is an acceptable range of normal anion gap. But some conditions can increase or decrease this gap. Increased anion gap usually represents metabolic acidosis. Albumin and many other confounding factors influence the anion gap derangements. Accuracy in measuring anion gap is critically important for the evaluation of acidosis.

2. Normal acid-base balance

The body maintains its normal physiology by the strict balance of acid and base. The body maintains its normal arterial pH close to 7.4 at a range between 7.36–7.44, and the intracellular pH of the human body is 7.2 [11]. Normal acid-base balance is the balance between each hydrogen increase by the intake or production, and that is decreased by elimination. Acid-base balance is measured by measuring pH, CO_2 and HCO_3^- . In general, consuming animal protein add acid in the body, and consuming cereals and vegetables add alkali in the body. In oxidative metabolism, CO_2 is produced in the tissue, and at a similar rate, that is eliminated by the lungs. So, pCO_2 persists at about 5.33 kPa (40 mm of Hg). Different buffer systems of the body play a crucial role in removing excess H^+ . Metabolism of carbohydrate and fat uses O_2 and produce CO_2 and H_2O . Normal lungs efficiently remove most of the CO_2 . In oxidation of amino acids, carbon dioxide and water are produced along with the liberation of nitrogen as ammonia, a toxic material in the body. In the liver, the urea cycle utilizes the ammonia, where this toxic NH_3 combines with CO_2 , and produce urea. In the proximal tubule and other renal epithelial cells, ammonia and bicarbonate are also produced from glutamine metabolism. Some of it returns to the body fluid through the renal veins and is metabolized in the liver. And the rest of the NH_3 excreted in the lumen. So, NH_3 does not exist in the body fluid. Most of the NH_3 is excreted in the urine, and it plays an important role in removing H^+ to maintain normal acid-base balance. In the urine, NH_3 binds hydrogen ion to produce NH_4^+ , and it prevents excessive acidification of urine.

3. Respiratory and renal regulation of acid and base

Excess acid is eliminated from the body by the lungs and the kidneys. In the lungs, acid is eliminated in the form of CO_2 , and in the kidneys, acid is excreted as acid phosphate and ammonium. CO_2 is lipid soluble, and it crosses the cell membranes in the lungs. Most of the CO_2 produced in the tissue is eliminated by alveolar ventilation. Arterial and brain chemoreceptors can sense the acid and base excess, and respiratory system responds with hyper or hypo ventilation. As a result, pH is increased or decreased by increasing and decreasing pCO_2 level. The regulation between CO_2 and H_2CO_3 level is critically maintained when the blood travels through the lung capillaries. When strong acid is added, some HCO_3^- become H_2CO_3 and blood PCO_2 is increased. In acidosis, carbonic acid dissociate to CO_2 and H_2O . As a result, respiratory center is stimulated and it leads to hyperventilation. Hyperventilation eliminates these CO_2 to maintain normal pH. In alkalosis, CO_2 is retained by hypoventilation. This CO_2 combines with H_2O to produce H_2CO_3 , and pH is maintained.

The kidneys excrete acids, both respiratory and nonrespiratory origin and retain HCO_3^- to stabilize the pH of blood. HCO_3^- is predominantly regulated in the kidneys. The nephron reabsorbs all filtered bicarbonate in exchange for H^+ . The kidneys also produce new bicarbonate to neutralize acids. Tubular cells contain carbonic anhydrase, that converts CO_2 and H_2O to HCO_3^- and H^+ . Newly formed HCO_3^- is shunted to peritubular capillaries and H^+ is excreted in tubular lumen. Bicarbonate is also produced from glutamine metabolism along with ammonium. Some NH_4 diffuses to body fluid and converts to urea in the liver. The rest of the them excreted in urine. The tubules are impermeable to bicarbonate, and it cannot be converted back to CO_2 and H_2O . So, the blood HCO_3 level is increased.

In the apical membrane of the kidney tubules, sodium is reabsorbed in exchange for the hydrogen ion. Salts like sulfates, phosphates, ammonia combines the hydrogen ions and excrete it. The kidneys titrate less than half of the excreted acids and the rest is excreted as ammonium [11]. For every ammonium excreted in urine, one HCO_3^- is reabsorbed. HCl and H_2SO_4 are produced during dietary protein metabolism reacts with NaHSO_4 , and produce NaCl and Na_2SO_4 . These Na salts are excreted by the kidneys as NH_4Cl , and $(\text{NH}_4)_2\text{SO}_4$.

The kidneys are largely responsible for K^+ excretion and most of it is reabsorbed in the proximal tubule and in the loop of Henly. In acidosis, K^+ secretion is decreased and K^+ absorption is increased in the collecting duct. In alkalosis, hypokalemia develops from increased K^+ secretion and reduced K^+ absorption in the collecting duct. H^+ and K^+ exchange occur in the tubules. Serum potassium level also influences the renal acid-base balance. In hyperkalemia, potassium is available in an increased amount in the filtrate, and hydrogen will be scarce for exchange with HCO_3 and there will be an imbalance. In hypokalemia, less potassium will be available for H^+ and K^+ exchange and hydrogen will be available to exchange with bicarbonate.

Na^+ , K^+ and NH_4^+ are the principle urinary cations, and the principal urinary anion is chloride. Urinary anion gap helps in estimating renal NH_4^+ excretion, as NH_4^+ is the urinary unmeasured ion. Chloride is an important anion in neutralizing positive ions, reabsorbed in the proximal convoluted tubule and secreted in urine by the collecting duct. Secreted H^+ is also buffered by urinary buffer HPO_4^- to H_2PO_4 , and is excreted in urine.

4. Acidosis and buffer

Acidosis results from a reduction in serum bicarbonate and cause secondary reduction of PaCO_2 resulting in a low blood pH. It develops from the addition of hydrogen or removal of HCO_3 from the body. PaCO_2 in blood is 38 ± 2 mm of Hg and HCO_3 is 24 ± 2 mmol/L. Metabolic acidosis is characterized by the blood pH <7.38 and bicarbonate <22 mmol/L [12].

Acid and base disorders are: respiratory acidosis and respiratory alkalosis, and metabolic acidosis and metabolic alkalosis [13]. In respiratory acidosis, PaCO_2 is increased and it is compensated by renal H^+ excretion, HCO_3 retention and HCO_3 generation. In respiratory alkalosis, decreased PaCO_2 is compensated by renal HCO_3 excretion. In metabolic acidosis, HCO_3 is reduced and it is compensated by hyperventilation and PaCO_2 reduction. HCO_3 is increased in metabolic alkalosis, and it is compensated by increasing PaCO_2 by hypoventilation [14]. Usually, respiratory disorders cause derangements of CO_2 level in the blood, and change in HCO_3 level is developed from metabolic disturbances.

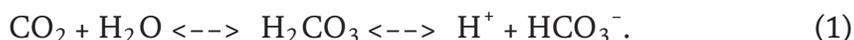
In the blood, Alkali is present mainly in the form of sodium bicarbonate, and bicarbonate is bound to other bases. Increase in BHCO_3 and decrease in H_2CO_3

results in alkalosis, and decrease in BHCO_3 and increase in H_2CO_3 results in acidosis [13]. The body contains many acids. They are hydrochloric acid, carbonic acid, citric acid, lactic acid, phosphoric acid and carboxylic acid. Acute metabolic acidosis is developed by the overproduction of organic acids, like lactic acid and keto acid. Chronic acidosis is caused by bicarbonate wasting and impaired urinary acidification.

Blood cells are more acidic than serum, which influences the distribution of electrolyte and water between them. These transports took place with the oxygenation and reduction of hemoglobin and shift of bases (Na^+ , K^+) due to changes in pH. Under normal environment Na^+ and K^+ do not diffuse through the cell wall. Shifting of water and electrolyte through membrane results from the change in anion (HCO_3^- and Cl^-) and H^+ concentration, and that changes in cell volume. CO_2 , relative electrolyte concentration and weak acid concentrations are three independent variables that regulate blood pH [15].

The body has different buffer systems to maintain the normal pH of the body. Elkinton Jr. reported that multiple level of buffering linked different series of ionic exchanges which includes hydrogen, sodium, potassium, and other anions. The buffers absorb excess hydrogen and hydroxyl ions. They help in the maintenance of neutrality during redistribution of the hydrogen ion [16].

A buffer system consists of a weak acid with its conjugate base, or a weak base with its conjugate acid. Blood is a strong solution, and it has many important components that maintain the buffer systems. These include hemoglobin, bicarbonate, carbonic acid, plasma proteins, RBCs and plasma phosphate [17]. $\text{HCO}_3^-/\text{CO}_2$ buffer is the most important buffer system of the body, and plays a major role in regulating pH of the blood. But, the rest of the buffer systems have minimum contribution in pH regulation. In dissolved state, bicarbonate and carbon dioxide ion remains in equilibrium. Bicarbonate reduces strong acid to carbonic acid, whereas carbonic acid neutralizes strong base (Eq. (1)).



When CO_2 and water is converted to HCO_3^- and hydrogen ions, this hydrogen ion is then buffered by hemoglobin [18].

Proteins have a buffering capacity, including hemoglobin. Protein can accept and donate H^+ , if there is H^+ excess or it is reduced. Hemoglobin has a distinct types of buffer action. When blood passes through the capillaries, it loses oxygen and took CO_2 to raise the PaCO_2 and maintain the pH. Hemoglobin plays an important role in transporting both oxygen and carbon dioxide. In 1914, Douglas, Haldane and Christiansen tried to prove that the hemoglobin binds more CO_2 in the reduced form than the oxygenated form [19].

The phosphate buffer system works in the internal environment of all cells. But, in the blood H_2PO_4^- and HPO_4^{2-} are found in a very low concentration. Sodium dihydrogen phosphate neutralizes strong bases and sodium monohydrogen phosphate neutralizes strong acids. The Phosphate buffer system plays an important role in the kidneys.

5. Acid base physiology

Two types of variables, dependent and independent, are important in acid-base balance [20]. Bicarbonate, hydroxyl ion, hydrogen ion or pH, weak acid, anion and carbon trioxide are dependent variables and they are determined by three independent

variables $p\text{CO}_2$, total weak acid and net strong ion charge [21]. Lungs, kidneys, liver and gut regulated this balance. Traditional bicarbonate/carbon-di-oxide approach, base excess approach and Stewart's physicochemical methods are widely discussed for measuring the acid base disorders as well as to explore the physiology of body fluid.

6. Traditional physiological approach

HCO_3/CO_2 buffer system is the basis of this approach. Carbonic acid freely moves in the body fluid and dissociates into bicarbonate automatically when needed. Bicarbonate in the body acts as alkaline reserve. CO_2 , pH and HCO_3 can be calculated by Hasselbalch-Henderson Equation (2) [7, 9].

$$\text{pH} = \text{pK} + \log_{10} \left[\frac{[\text{HCO}_3^-]}{s.\text{PCO}_2} \right] \quad (2)$$

This equation states that not only HCO_3 and CO_2 , but also their ratio determines the pH. In this equation, PCO_2 is the respiratory component and HCO_3^- is the metabolic component of the acid base imbalance. This buffer system is the largest and independent buffer system of the body and whole body acts as an open system for CO_2 . In traditional approach balance is determined by the influx and efflux of H^+ and HCO_3 .

7. Base excess approach

Astrup and Siggaard-Anderson introduced base excess approach, which is close to the traditional approach [22, 23]. Base excess can be calculated from bicarbonate concentration and pH of the body [4]. It can estimate the acid base status of non-respiratory origin. If base excess is too high, then it is metabolic alkalosis. If base excess is too low, then it is metabolic acidosis. When a deviation of normal blood pH is corrected by administering base, then it is called base deficit. Which is a characteristic of metabolic acidosis. Base deficit with increase anion gap suggest the addition of acid in the body fluid. If there is a base deficit with normal anion gap, then there is bicarbonate loss from the body.

8. Stewart approach

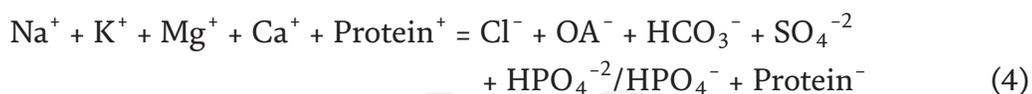
Here H^+ /proton is the preliminary determinant in acid base disturbances, not the CO_2 [21]. The dependent variables are H^+ , OH^- , CO_3^{2-} , HA (weak acid), A^- (weak anions), HCO_3^- and pH. The independent variables are strong ion difference (SID), total non-volatile weak acids (A_{tot}) and PaCO_2 [24]. Among them the strong ion difference has maximum effect on the hydrogen ion concentration. With that, acid base disorder can be divided into three categories: 1. respiratory (increase or decrease PaCO_2), 2. SID changes (excess or deficit of strong ions or water) and 3. inorganic phosphate or albumin deficit or excess (A_{tot} changes). In Stewart approach, a large number of variables are needed to calculate SID. Sodium, potassium, calcium and magnesium are strong positive ions, and chloride and lactate are the negative ions [25]. Bicarbonate and albumin are the balancing ion in strong ion difference. Strong ion difference (mEq/L) = [strong cations] – [strong anions]. Weak acid dissociates in body fluid (Eq. (3)).



A^- Resembles weak anions, that vary with pH. Strong ion difference is filled with this weak A^- , and HCO_3^- , H^+ , OH^- , CO_3^{2-} are also present in minute amount, but are less important. There are many unmeasured anions accounts for ion difference. For electrical neutrality, strong ion difference and the total charge of weak ions must be equal [26]. Normal SID is dominated by sodium and chloride. But other negligible, but measurable ions are present there. Here narrowing of SID from an increase in $[Na^+]$ has alkalizing effect, whereas an increase in $[Cl^-]$ has acidifying effect. From the ionic basis metabolic acid base disturbances are about four major types [25]: (1) The water effect, and it is produced by dilutional effect on SID. Free water intake and intravenous infusion can produce it. (2) The chloride effect is caused by chloride change, and administration of normal saline is the common cause. (3) The protein effect is produced by a change in albumin concentration. (4) There are other factors, and those are influenced by unmeasured anions, that cause a wide anion gap.

9. Anion gap

In vivo, true ion gap cannot exist. There are many anions and cations in the blood. Blood cations and anions must be equal. Sodium, chloride and bicarbonate have the highest concentrations, and they are calculated for anion gap for their largest variability in different pathologic conditions. Anion gap is the difference between serum sodium ion and bicarbonate plus chloride. There are wide variations in the reported anion gap. Widely accepted anion gap is 8–12 mmol/L [15]. Anion gap is clinically important for assessing acidosis. Normal anion gap (hyperchloremic) acidosis and increased anion gap acidosis [27] are two important types of anion gap acidosis. Common serum cation levels are sodium 138.8 ± 4.56 mmol/L, potassium 4.05 ± 0.21 mmol/L, magnesium 0.98 ± 0.05 mmol/L [28] and calcium 2.2–2.7 mmol/L [29]. And normal serum anion levels are chloride 97.7 ± 3.42 mmol/L and acetate 0.23 ± 0.04 mmol/L [28]. The sum of cations and anions should be equal (Eq. (4)).



There are other ions which are not commonly measured, are unmeasured anions and cations [30]. Under normal conditions, albumin and phosphate accounts for this anion gap. There are many clinical conditions, where urate, lactate, ketone bodies, sulfate, salicylates, penicillin's, citrate, pyruvate, and acetates are also responsible for increased anion gap [5]. So, anion gap [31] is Eq. (5)

$$Na - (Cl + HCO_3^-) = UA - UC \quad (5)$$

10. Unmeasured anion

Presence of unmeasured anion in blood is the anion gap and it represents metabolic acidosis [32]. When unmeasured anions like lactate and pyruvate donates proton then that proton is buffered by bicarbonate. And bicarbonate consumption increases the anion gap. The most common causes include lactic acidosis, diabetic ketoacidosis, uremia and acidosis due to drugs and toxins. Methanol, propylene glycol, ethylene glycol, salicylate, and some inborn error of metabolism are

other causes of unmeasured anions [33]. Both lactate and β -hydroxybutyrate are increased in both Gram-positive septicemia [34] and starvation [35]. Krebs cycle intermediate citrate, isocitrate, malate, α -ketoglutarate, succinate and D-lactate are increased in different types of acidosis. Intestinal ischemia and short bowel syndrome cause increase in D-lactate [35]. Plasma proteins are mostly anionic comprising 75% of the unmeasured anion [36–38]. Treatment with Sodium thiosulfate that has no hydrogen can cause severe metabolic acidosis [39].

11. Increased anion gap

It usually indicates acidosis. Increase blood lactate, ketoacidosis, uremia (in advanced renal failure), drugs (salicylate and penicillin), ethylene glycol, methanol are contributor of high anion gap acidosis. But the increase anion gap can be due to laboratory error, hyperphosphatemia [30]. Massive rhabdomyolysis, hippurate, oxalate can also cause increased anion gap acidosis [31]. Diabetes, starvation and alcohol are the most common cause of ketoacidosis. In alcoholic ketoacidosis, primary keto acid is β -hydroxybutyrate. It can be missed in conventional assessment of ketonuria. High anion gap and normal lactate level are characteristics of alcoholic acidosis [40]. Starvation alone can cause high anion gap acidosis [41]. In the third trimester of pregnancy, short period of starvation can cause ketogenesis with a very high anion gap acidosis [42]. Septic shock, hypoxemia, hypovolemic shock, cyanide, mesenteric ischemia, CO poisoning, causes hypoxic type of L-lactic acidosis [43]. Non-hypoxic, L-lactic acidosis develops from seizure, thiamine deficiency, metformin, methanol, ethylene glycol, salicylate, propylene glycol, niacin, isoniazide, iron, propofol, toluene, paraldehyde, non-nucleoside reverse transcriptase inhibitor (NNRTI) drugs [12]. Recurrent 5-oxoprolinuria from inborn errors of metabolism is a rare cause of high anion gap metabolic acidosis [44]. Uremia results from not only reduced ammonia secretion but also reduced filtration of sulfate and phosphate anions, and increases the anion gap [45]. Polyclonal gammopathies are also contributor of increased anion gap [46]. Serum albumin is an important contributor to the anion gap and hypoalbuminemia is a common comorbid condition. That is why, albumin correction is crucial for the anion gap calculation [36, 37]. To explore the cause of the metabolic acidosis anion gap must be corrected for albumin as well as lactate [43]. A high anion gap can be masked by a concomitant low anion gap results from hypoalbuminemia.

12. Reduced anion gap

In anion gap calculation, sodium is the only cation that is measured. But, hypercalcemia, hyperkalemia and hypermagnesemia can produce significant decrements in anion gap. So, clinical correlation and correction of such abnormality is important. Plasma proteins comprise two third of the unmeasured anion, and hypoalbuminemia is a common cause for the low anion gap [31, 36, 37]. The reduced anion gap is usually seen in delusional states, hypernatremia, hypoalbuminemia, hypermagnesemia, hypercalcemia, bromide intoxication, hyperviscosity associated diseases etc. [47]. Sometimes it can be due to laboratory error, paraproteinemia [48, 49], or iodide [30, 50], gastrointestinal bicarbonate loss and diarrhea [31]. It has been reported that Lithium carbonate intoxication can also produce low or absent anion gap [51]. Non-sodium containing paraprotein IgG in multiple myeloma increase the unmeasured cations and reduce the anion gap [48, 52, 53]. Hypercalcemia and hypoalbuminemia in paraproteinemia also contribute to low anion gap [52].

13. Normal anion gap

Measuring anion gap is a routine for evaluating acidosis, and normal anion gap is sometimes misleading. As we know, the increase in anion gap is usual in metabolic acidosis. And acidosis is due to acid retention or ingestion. Normal anion gap acidosis is due to loss of HCO_3^- from the body. Hyperchloremic normal anion gap acidosis is characterized by acidosis with excess chloride ions [54]. Here, the low HCO_3^- level is a characteristic feature. Reduced negatively charged bicarbonate is compensated by the negatively charged chloride movement into the extracellular space, and normal anion gap is maintained. The causes of gastrointestinal and renal loss of bicarbonate are diarrhea, ureteral diversions, pancreatic and biliary fistulas, toluene ingestion, acetazolamide, ifosfamide, topiramite, tenofovir, renal tubular acidosis. These are the causes of normal anion gap acidosis. Rapid infusion of 0.9% normal saline can also cause hyperchloremic metabolic acidosis [55]. If the blood anion gap is normal, but there is acidosis, then the urinary anion gap Eq. (6) is calculated [12].

$$[\text{Na}^+] + [\text{K}^+] - [\text{Cl}^-] \quad (6)$$

The urinary anion gap is negative in diarrhea, sodium infusion and proximal renal tubular acidosis. Whereas, positive urinary anion gap is found in both type 1 and type 4 renal tubular acidosis. Renal tubular acidosis is sometimes the only presenting feature of many chronic diseases and conditions associated with polyclonal gammopathies.

14. Metabolic acidosis and anion gap

Metabolic acidosis results from gain of anions and loss of cations. Potassium chloride, hydrogen chloride, sodium chloride, arginine hydrochloride, calcium chloride, ammonium chloride, lysine hydrochloride can cause hyperchloremia and increase anion gap. Hyperphosphatemia increases the anion gap. But renal tubular acidosis [33], amiloride and triamterene cause a non anion gap hyperchloremic acidosis and hyperkalemia due to impaired bicarbonate production.

Anion gap should be measured for all types of metabolic acidosis. High anion gap metabolic acidosis is a subtype of non-respiratory acidosis. Mnemonics were used for remembering the causes of high gap metabolic acidosis such as KUSMALE (Ketoacidosis, Uraemia, Salicylate poisoning, Methanol, Paraldehyde, Lactate, Ethylene glycol) and MUD PILES (Methanol, Metformin uremia, Diabetic ketoacidosis, Paraldehydes, iron, isoniazid, Lactate, ethylene glycol, Salicylates and starvation). As paraldehyde induced acidosis is extremely rare and recently three anion gap generating organic acid has been recognized. They are Short bowel syndrome producing D-lactic acid, chronic paracetamol use induced 5-oxoproline (or pyroglutamic acid) especially in malnourished woman and high dose propylene glycol (used in lorazepam, phenobarbital) infusions generate acidosis. Also, Iron and Isoniazid can cause lactic acidosis. So, GOLD MARK is a new acronym for metabolic acidosis [Glycols (ethylene and propylene), Oxyproline, L-lactate, D-lactate, Methanol, Aspirin, Renal failure, Ketoacidosis] [56]. Metabolic acidosis also caused by renal bicarbonate loss in type 2 renal tubular acidosis, renal dysfunction in type 4 renal tubular acidosis, type 1 renal tubular acidosis and ingestion of ammonium chloride [31]. Acute rheumatism causes lactate induced acidosis also [57]. Symptomatic correction of acidosis will not eliminate the problem. If the clinical features suggest acidosis, then it should be assessed for anion gap as well.

Following anion gap measurement accordingly history of drug, toxins and diseases need to be evaluated for managing the exact pathology thus acidosis will be properly treated.

15. Albumin, phosphate, lactate and corrected anion gap

At normal blood pH 7.4 plasma proteins are mostly anionic. It has been estimated that anion gap decreases by 2.5 mEq/L for every 10 gm/L drop of serum albumin [36, 37]. Several studies had observed that 2–2.5 times changes in albumin influences in anion gap changes [58]. Albumin contributes a greater part of the normal anion gap [46]. Phosphate and lactate contribute some anion gap as well [59]. Consideration of all of these contributors are important in explaining changes in anion gap. Calculation of anion gap is crucial in critically ill patients. Anion gap should be adjusted for Eq. (7) albumin, phosphate and lactate with the following equation [59].

$$\text{Anion gap} = \{(\text{Na}) + (\text{K}) - (\text{Cl}) - (\text{HCO}_3)\} - \{2 \times \text{albumin g/dl} + 0.5 \times \text{phosphate mg/dl}\} - \{\text{lactate mmol/L}\} \quad (7)$$

Author details

Md. Masudul Hassan^{1,2,3}

1 Bangladesh College of Physicians and Surgeons, Dhaka, Bangladesh

2 Department of Rheumatology, Bangabandhu Sheikh Mujib Medical University, Dhaka, Bangladesh

3 DHCL and IB Hospital, Dhaka, Bangladesh

*Address all correspondence to: masud292@yahoo.com

IntechOpen

© 2020 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Severinghaus JW, Astrup PB. History of blood gas analysis. II. pH and acid base balance measurements. *Journal of Clinical Monitoring and Computing*. 1985;1:259. DOI: 10.1007/BF02832819
- [2] Malkin HM. Historical review: Concept of acid-base balance in medicine. *Annals of Clinical and Laboratory Science*. 2003;33(3):337-344
- [3] Henderson LJ, Black OF. A study of the equilibrium between carbonic acid, sodium bicarbonate, mono-sodium phosphate, and di-sodium phosphate at body temperature. *American Journal of Physiology-Legacy Content*. 1908;21(4):420-426
- [4] Astrup P, Jorgensen K, Andersen OS, et al. Acid-base metabolism: New approach. *Lancet*. 1960;i:1035-1039
- [5] Kimura S, Shabsigh M, Morimatsu H. Traditional approach versus Stewart approach for acid-base disorders: Inconsistent evidence. *SAGE Open Medicine*. 2018;6:2050312118801255. DOI: 10.1177/2050312118801255
- [6] Henderson LJ. Concerning the relationship between the strength of acids and their capacity to preserve neutrality. *The American Journal of Physiology*. 1908;21:173-179
- [7] Hasselbalch KA. Die Berechnung des Wasserstoffzahl des Blutes aus der freien und gebunden Kohlensäure desselben, und die Sauerstoffbindung des Blutes als Funktion der Wasserstoffzahl. *Biochemische Zeitschrift*. 1916;78:112-144
- [8] Myers RJ. One-hundred years of PH. *Journal of Chemical Education*. 2010;87(1):30-32. DOI: 10.1021/ed800002c
- [9] Henderson LJ. The theory of neutrality regulation in the animal organism. *American Journal of Physiology-Legacy Content*. 1908;21:427-448
- [10] Warburg EJ. Studies on carbonic acid compounds and hydrogen ion activities in blood and salt solution—A contribution to the theory of the equation of L. J. Henderson and K. A. Hasselbalch. *The Biochemical Journal*. 1922;16:153-340
- [11] Hamm LL, Nakhoul N, Hering-Smith KS. Acid-base homeostasis. *Clinical Journal of the American Society of Nephrology*. 2015;10(12):2232-2242. DOI: 10.2215/CJN.07400715
- [12] Berend K, de Vries AP, Gans RO. Physiological approach to assessment of acid-base disturbances. *The New England Journal of Medicine*. 2014;371(15):1434-1445. DOI: 10.1056/NEJMra1003327
- [13] Austin JH, Cullen GE. Hydrogen ion concentration of the blood in health and disease. *Medicine*. 1925;4(3):275-343
- [14] Trulock EP III. Chapter 49: Arterial blood gases. In: Walker HK, Hall WD, Hurst JW, editors. *Clinical Methods: The History, Physical, and Laboratory Examinations*. 3rd ed. Boston: Butterworths; 1990. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK37>
- [15] Kellum JA. Determinants of blood pH in health and disease. *Critical Care*. 2000;4:6. DOI: 10.1186/cc644
- [16] Elkinton JR. Whole body buffers in the regulation of acid-base equilibrium. *The Yale Journal of Biology and Medicine*. 1956;29(3):191-210
- [17] Viikari SJ, Harjola P, Maamies T. Clinical studies on the buffer capacity of the blood. *Scandinavian*

Journal of Clinical and Laboratory Investigation. 1954;**6**(2):122-128. DOI: 10.3109/00365515409134852

[18] Ferry RM, Green AA. Studies in the chemistry of hemoglobin. *The Journal of Biological Chemistry*. 1929;**81**:175

[19] Doisy EA, Briggs AP, Eaton EP, Chambers WH. Evaluation of buffers of the blood. *The Journal of Biological Chemistry*. 1922;**54**:305

[20] Stewart PA. Independent and dependent variables of acid-base control. *Respiration Physiology*. 1978;**33**(1):9-26

[21] Stewart PA. Modern quantitative acidbase chemistry. *Canadian Journal of Physiology and Pharmacology*. 1983;**61**:1444-1461

[22] Astrup P. A simple electrometric technique for the determination of carbon dioxide tension in blood and plasma, total content of carbon dioxide in plasma, and bicarbonate content in separated plasma at a fixed carbon dioxide tension (40 mm Hg). *Scandinavian Journal of Clinical and Laboratory Investigation*. 1956;**8**(1):33-43

[23] Andersen OS. The pH-log pCO₂ blood acid-base nomogram revised. *Scandinavian Journal of Clinical and Laboratory Investigation*. 1962;**14**:598-604

[24] Jones NL. A quantitative physicochemical approach to acid-base physiology. *Clinical Biochemistry*. 1990;**23**(3):189-195

[25] Magder S, Emami A. Practical approach to physical-chemical acid-base management. *Stewart at the bedside*. *Annals of the American Thoracic Society*. 2015;**12**:111-117. DOI: 10.1016/j.pbiomolbio.2015.01.00

[26] Morgan TJ. The meaning of acid-base abnormalities in the intensive care

unit – Effects of fluid administration. *Critical Care*. 2004;**9**:204. DOI: 10.1186/cc2946

[27] Kraut J, Madias N. Metabolic acidosis: Pathophysiology, diagnosis and management. *Nature Reviews. Nephrology*. 2010;**6**:274-285. DOI: 10.1038/nrneph.2010.33

[28] Chapp AD, Schum S, Behnke JE, et al. Measurement of cations, anions, and acetate in serum, urine, cerebrospinal fluid, and tissue by ion chromatography. *Physiological Reports*. 2018;**6**(7):e13666. DOI: 10.14814/phy2.13666

[29] Goldstein DA. Chapter 143: Serum Calcium. In: Walker HK, Hall WD, Hurst JW, editors. *Clinical Methods: The History, Physical, and Laboratory Examinations*. 3rd ed. Boston: Butterworths; 1990. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK250>

[30] Kraut JA, Madias NE. Serum anion gap: Its uses and limitations in clinical medicine. *Clinical Journal of the American Society of Nephrology*. 2007;**2**:162-174

[31] Lee S, Kang KP, Kang SK. Clinical usefulness of the serum anion gap. *Electrolyte Blood Press*. 2006;**4**(1): 44-46. DOI: 10.5049/EBP.2006.4.1.44

[32] Selk N, Rodby RA. Unexpectedly severe metabolic acidosis associated with sodium thiosulfate therapy in a patient with calcific uremic arteriolopathy. *Seminars in Dialysis*. 2011;**24**(1):85-88. DOI: 10.1111/j.1525-139X.2011.00848.x

[33] Seifter JL. Integration of acid-base and electrolyte disorders. *The New England Journal of Medicine*. 2014;**371**:1821-1831

[34] Barnardo DE, Cohen RD, Iles RA. “Idiopathic” lactic and

- betahydroxybutyric acidosis. *British Medical Journal*. 1970;**4**(5731):348-349. DOI: 10.1136/bmj.4.5731.348-a
- [35] Forni LG, McKinnon W, Hilton PJ. Unmeasured anions in metabolic acidosis: Unravelling the mystery. *Critical Care*. 2006;**10**(4):220. DOI: 10.1186/cc4954
- [36] Figge J, Jabor A, Kazda A, Fencel V. Anion gap in hypoalbuminemia. *Critical Care Medicine*. 1998;**26**:1807-1810
- [37] Durward A, Mayer A, Skellet S, Taylor D, Hanna S, Tibby SM, et al. Hypoalbuminemia in critically ill children : Incidence, prognosis, and influence on the anion gap. *Archives of Disease in Childhood*. 2003;**88**:419-422
- [38] Rocktaeschel J, Morimatsu H, Uchino S, Bellomo R. Unmeasured anions in critically ill patients: Can they predict mortality? *Critical Care Medicine*. 2003;**31**(8):2131-2136
- [39] Hunt GM, Ryder HF. Metabolic acidosis after sodium thiosulfate infusion and the role of hydrogen sulfide. *Clinical case reports*. 2018;**6**(8):1595-1599. DOI: 10.1002/ccr3.1673
- [40] Noor NM, Basavaraju K, Sharpstone D. Alcoholic ketoacidosis: A case report and review of the literature. *Oxford Medical Case Reports*. 2016;**2016**(3):31-33. DOI: 10.1093/omcr/omw006
- [41] Mubarik A, Jupalli A, Iqbal AM, Muddassir S, Eddib A. Isolated starvation ketoacidosis: A rare cause of severe metabolic acidosis presenting with a pH less than 7. *Cureus*. 2019;**11**(2):e4086. DOI: 10.7759/cureus.4086
- [42] Sinha N, Venkatram S, Diaz-Fuentes G. Starvation ketoacidosis: A cause of severe anion gap metabolic acidosis in pregnancy. *Case Reports in Critical Care*. 2014;**2014**. Article ID: 906283, 4 pages. DOI: 10.1155/2014/906283
- [43] Moviat M, van Haren F, van der Hoeven H. Conventional or physicochemical approach in intensive care unit patients with metabolic acidosis. *Critical Care*. 2003;**7**(3):R41-R45
- [44] Tailor P, Raman T, Garganta CL, Njalsson R, Carlsson K, Ristoff E, et al. Recurrent high anion gap metabolic acidosis secondary to 5-oxoproline (pyroglutamic acid). *American Journal of Kidney Diseases*. 2005;**46**(1):e4-e10
- [45] Ishihara K, Szerlip HM. Anion gap acidosis. *Seminars in Nephrology*. 1998;**18**(1):83-97
- [46] Kirschbaum B. Hyperglobulinemia with an increased anion gap. *The American Journal of the Medical Sciences*. 1998;**316**(6):393-397
- [47] Emmett M, Narins RG. Clinical use of the anion gap. *Medicine (Baltimore)*. 1977;**56**(1):38-54
- [48] Keshgegian AA. Anion gap and immunoglobulin concentration. *American Journal of Clinical Pathology*. 1980;**74**(3):282-284
- [49] Qujeq D, Mohiti J. Decreased anion gap in polyclonal hypergammaglobulinemia. *Clinical Biochemistry*. 2002;**35**(1):73-75
- [50] Fischman RA, Fairclough GF, Cheigh JS. Iodide and negative anion gap. *The New England Journal of Medicine*. 1978;**298**:1035-1036
- [51] Kelleher SP, Raciti A, Arbeit LA. Reduced or absent serum anion gap as a marker of severe lithium carbonate intoxication. *Archives of Internal Medicine*. 1986;**146**:1839-1840
- [52] Murray T, Long W, Narins RG. Multiple myeloma and anion gap.

The New England Journal of Medicine.
1975;**292**:574-575

[53] De Troyer A, Stolarczyk A, De Beyl DZ, Stryckmans P. Value of anion-gap determination in multiple myeloma. The New England Journal of Medicine. 1977;**296**:858-860

[54] Sharma S, Hashmi MF, Aggarwal S. Hyperchloremic Acidosis. [Updated: 13 September 2019]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2019. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK4823>

[55] Prough DS, Bidani A. Hyperchloremic metabolic acidosis is a predictable consequence of intraoperative infusion of 0.9% saline. *Anesthesiology*. 1999;**90**(5):1247-1249

[56] Mehta AN, Emmett JB, Emmett M. GOLD MARK: An anion gap mnemonic for the 21st century. *Lancet*. 2008;**372**(9642):892. DOI: 10.1016/S0140-6736(08)61398-7

[57] Parsons LG, Edgar SH. The Acid-Base equilibrium of the blood in acute rheumatism. *Archives of Disease in Childhood*. 1929;**4**(23):291

[58] Feldman M, Soni N, Dickson B. Influence of hypoalbuminemia or hyperalbuminemia on the serum anion gap. *The Journal of Laboratory and Clinical Medicine*. 2005;**146**(6):317-320

[59] Zampieri FG, Park M, Ranzani OT, et al. Anion gap corrected for albumin, phosphate and lactate is a good predictor of strong ion gap in critically ill patients: A nested cohort study. *Revista Brasileira de Terapia Intensiva*. 2013;**25**(3):205-211. DOI: 10.5935/0103-507X.20130036