

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



# miRNAs in Acute Lymphoblastic Leukemia: Diagnosis, Prognosis and Target Therapeutic

*Yazmín Gómez-Gómez, Jorge Organista-Nava,  
Berenice Illades-Aguilar and Marco Antonio Leyva-Vázquez*

## Abstract

Acute lymphoblastic leukemia (ALL) is more frequent in children than in adults. The ALL is a hematological neoplasia, which is characterized by the hyperproliferation of lymphoid precursors in bone marrow. MicroRNAs (miRNAs) are a class of noncoding RNAs that regulate mRNA expression at posttranscriptional level. miRNAs regulate different biological processes such as development, proliferation, apoptosis, hematopoiesis, drug resistance, and tumorigenesis. It has also been observed that the expression of miRNAs can be used to the classification of the different subtypes of ALL. Likewise, miRNAs can also be used to determine the prognostic value and may represent potential therapeutic target molecules in the treatment of ALL.

**Keywords:** miRNAs, acute lymphoblastic leukemia, diagnosis, prognosis, therapy, biomarkers

## 1. Introduction

The hematopoiesis is primarily regulated at the transcriptional level by transcription factors that act as master regulators of genes expression. However, the transcriptional process alone does not appear to control all aspects of cellular functioning (cell fate, lineage, etc.), suggesting the participation of other mechanisms. The miRNAs constitute another critical way of hematopoietic regulation. The B- and T-lymphocytes develop from progenitor cells that occur in different organs; B-cell lymphopoiesis is completed in the bone marrow, whereas T-cell lymphopoiesis occurs in the thymus. However, their development and activation are controlled by signaling pathways, which are also regulated by the microRNAs (miRNAs) [1]. miRNA expression profile during the normal and malignant hematopoiesis suggests that miRNAs are regulators of hematopoiesis implicated in regulating and maintenance of the “stemness” of the early progenitors, various stages of cell differentiation, and malignance [2].

Nowadays, there is evidence that miRNAs do not just regulate hematopoietic differentiation and proliferation but also their activity. Deregulation of the expression of miRNAs has been observed in leukemias, and mechanistic studies reveal a role for miRNAs in the pathogenesis of this disorder [3].

Leukemia is a clonal disorder in which the normal hematopoiesis is replaced by a malignant clonal expansion of immature hematopoietic cells (blasts) in the bone marrow or peripheral blood [4]. The first approach between miRNAs and leukemia was carried out by Calin et al. [5]. The author showed that the 13q14 deletion in B-cell chronic lymphocytic leukemias (B-CLLs) causes the loss of the precursor gene of miR-16-1 and miR-15a; therefore, the loss of these miRNAs is observed in approximately 70% of the CLLs [5]. Interestingly, it has been reported that at fragile sites, minimal regions of amplification (minimal amplicons), or common break-point regions fragile sites, minimal regions of loss of heterozygosity, and genomic regions related with cancer code for approximately 50% of the miRNAs, hence the aberrant expression of different miRNAs in cancer [6].

The participation of miRNAs in different biological and cellular processes under pathological and normal conditions makes them good candidates in the investigation of functional markers for differential diagnosis, prognosis, and development of new therapeutic regimens, through the investigation of their molecular targets. In this chapter, the role of miRNAs expression profiles in ALL that could be used for classification of the disease establishing specific diagnoses and prognostic values is summarized. Likewise, the relation between the miRNA dysregulation and ALL may be a potential therapeutic target.

2. MiRNA biogenesis

The miRNA genes are transcribed by RNA polymerase II (Pol II) in the nucleus, and the primary miRNAs transcripts (pri-miRNAs) contain cap structures as well as poly(A) tails [7, 8]. The pri-miRNA transcript is processed by the microprocessor complex (Drosha/DGCR8), which crops the pri-miRNAs, producing a pre-miRNA

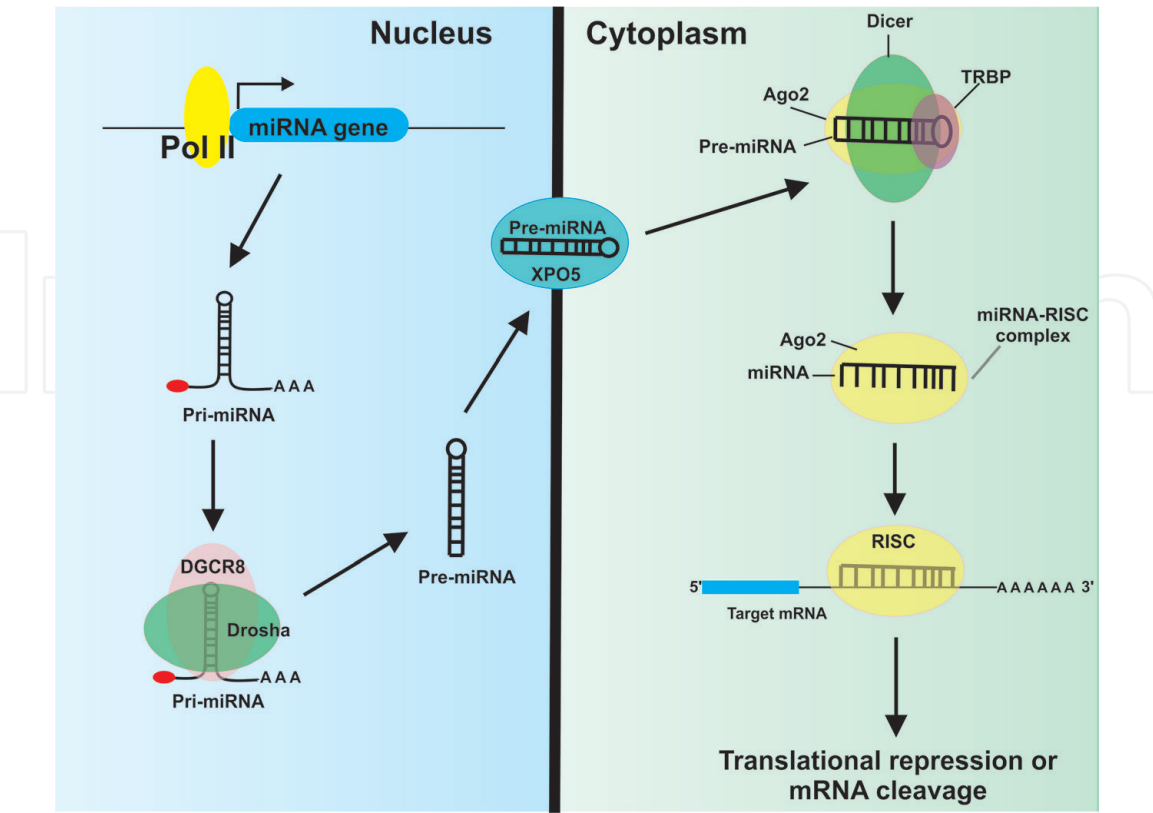


Figure 1.  
miRNA biogenesis.

(transcript of about 70 kb) [9–11]. The exportin 5 (XPO5) mediates the export of the pre-miRNAs from the nucleus to the cytoplasm [12–14]. In the cytosol, the pre-miRNA is recognized by Dicer enzyme (RNase type III), producing a mature **miRNA duplexes** (miRNA:miRNA\*) about 22 nucleotides [10]. The miRNA duplex binds to the RNA-induced silencing complex (RISC) [which is composed by of the transactivation-responsive RNA-binding protein (TRBP) and Argonaute2 (Ago2)] [8, 15]. The mature strand is retained by the Ago2 protein in the RISC complex, who directs the mature mRNA to its mRNA target for posttranscriptional gene silencing, while the complementary strand is degraded [16, 17] (**Figure 1**).

### 3. Functions of the miRNAs in lymphopoiesis

Lymphopoiesis is a process by which the hematopoietic stem cells (HSCs) differentiate into lymphoid progenitors and finally into B- or T-lymphocytes [18]. In the process of differentiation, the miRNAs play an important role. miR-29a and miR-196b are highly expressed by HSCs, and their downregulation is associated with differentiation into lymphoid progenitors [19, 20]. It has been reported that miR-17, miR-24, miR-155, miR-128, and miR-181 act to prevent the differentiation of early-stage progenitors [21].

miRNA-150 is expressed in both mature B- and T-cells. The lymphoid progenitors express the miRNA-150 to give rise to the mature B-cells and assist in the transition from progenitor B-cell (pro-B) to the precursor B-cell (pre-B) stage [18]. And premature expression of miRNA-150 results in blocked transition from the pro-B-cell stage to the pre-B-cell stage [22, 23].

B-cell differentiation is regulated by the miR-155, and it has been observed that miR-155 levels are upregulated rapidly in both activated mature T- and B-cells [24]. Also, miRNA-155 regulates the differentiation of T-cells into Th type 1 cells [24, 25].

miR-181 is specifically expressed in hematopoietic cell, and its expression is dynamically regulated during early hematopoiesis and lineage commitment. miR-181 expression is high in the early B-cell differentiation stage and progressively decreases subsequently, and its ectopic expression in hematopoietic stem/progenitor cells led to an increased fraction of B-lineage cells in both tissue culture differentiation assays and adult mice [26]. Additionally, miR-181 also plays an important role in T-cell development [27].

The miRNA-15 family is an element required to promote the switch from pre-B-cell proliferation to a more differentiated stage. [28]. So, pre-B-cells lacking miRNA-15 family functions exhibit prolonged proliferation because of aberrant expression of the target genes cyclin E1 and D3, and they additionally fail to trigger the transcriptional reprogramming normal to their differentiation, resulting in a developmental block at the pre-B-cell stage [28].

Six miRNAs, miR-17, miR-18a, miR-19a, miR-20a, miR-19b-1, and miR-92a-1 are part of the miR-17-92 cluster; these small molecules are important for mature B-cell development. Absence of the cluster leads to the development of disorders in the maturation from pro-B to pre-B stage [29]. Ventura et al. using miR-17-92-deficient mice found that B-cell development is inhibited at the pro-B to pre-B stage differentiation [30]. The above shows that if the miR-17-92 family miRNAs control the pro- to pre-B transition during B-cell development [31]. Likewise, it has been showed that in helper T cells, the miR-17–92a cluster is critical for the differentiation from Th1 cells [32].

miR-29b is increased in Th1 cells, and the levels from this miRNA decrease significantly upon T cell activation. So, the miR-29 expression can serve as a regulator



of Th1 differentiation [33]. Expression of miR-21 promotes Th2 differentiation in nonpolarized T cells [34]. miR-126 is another miRNA that also regulates the differentiation of the Th2 cells [35].

#### **4. miRNA expression and its role in the differential diagnosis of acute lymphoblastic leukemia subtypes**

Acute lymphoblastic leukemia (ALL) is characterized by clonal proliferation of early B- and T-lymphocyte progenitors that result in the accumulation of lymphoblasts in the bone marrow and various extramedullary sites. ALL is also the hematology neoplasia most commonly observed in the pediatric population, while it is relatively **less common than AML** in adults [36]. Around 75% of childhood ALL cases contain at least one alteration chromosomic, have lymphoid maturation arrest in distinct stages, and involve B- or T-lineages to leaving different immunophenotypes with different miRNA signatures [37].

microRNAs participate in different physiological and cellular processes, such as development and tissue differentiation, cell identity, cell cycle progression, and programmed cell death [38]. Nowadays, it is known that the distinct stages of lymphopoiesis and the direction of lymphoid precursor maturation are influenced by miRNA expression differentially. However, an aberrant expression of miRNAs is related with malignant lymphopoiesis, characteristic that can be utilized as signature to diagnosis and classification diagnosis of acute lymphoblastic leukemia [18]. Interestingly, miRNA groups that can clearly differentiate ALL of its normal counterpart, B-ALL versus T-ALL and ALL subtypes with specific genetic abnormalities have been reported. De Oliveira and collaborators reported miRNA-128a and miRNA-181b overexpressed and miRNA-100, miRNA-196b, and let-7e with lower level when compared the miRNAs expression in normal pediatric bone marrow (BM) samples and BM samples of pediatric ALL. The authors point out miR-196b as a miRNA highly expressed in T-ALL, while miR-100 was related with the presence of t(12;21) [39].

A study in Brazilian children with T- or B-cell acute lymphoblastic leukemia (T-ALL or B-ALL) evaluated a bone marrow miRNAs profile that may be used for distinguishing childhood lymphoblastic leukemia subtypes [40]. The authors mention that miR-708-5p, miR-497-5p, miR-151a-5p, miR-151b, miR-371b-5p, miR-455-5p, miR-195-5p, miR-1266-5p, miR-574-5p, miR-425-5p downregulated and miR-450b-5p, miR-450a-5p, miR-542-5p, miR-424-5p, miR-629-5p, miR-29c-5p upregulated in childhood T-ALL may be used for distinguishing childhood T- and B-ALL subtypes. However, a machine learning analysis showed that miR-29c-5p, which is involved in calcium signaling, is critical for B-cell lymphocyte fate. So, it is the best discriminator between childhood T- and B-ALL [40].

In a series of adult ALL cases, the expression profile of 470 miRNAs was measured by microarray analysis; 3 miRNAs (miR-148, miR-151, and miR-424) were identified as discriminative of T-lineage versus B-lineage ALL; and miR-151 dramatically downmodulated an miR-148a and miR-424 with higher expression in patients with T-ALL [41]. Furthermore, in the B-lineage ALL cases with special molecular lesions, those with BCR/ABL, E2A/PBX1, MLL/AF4 rearrangements and cases lacking known genetic abnormalities can be differentiated by a set of six miRNA, which was highlighted by one-way analysis of variance [41]. These miRNAs were preferentially expressed in each chromosomic rearrangement; miR-425-5p, miR-191, and miR-128 were expressed in the E2A/PBX1-positive case, miR-629 was highly expressed in cases harboring MLL/AF4 rearrangement, while high levels of miR-146b and miR-126 were observed in the BCR/ABL-positive cases [41]. Other study in pediatric ALL showed

ALL subtype	Upregulated expression	Downregulated expression	References
<b>Children</b>			
T-ALL	miR-450b-5p, miR-450a-5p, miR-542-5p, miR-424-5p, miR-629-5p, miR-29c-5p	miR-708-5p, miR-497-5p, miR-151a-5p, miR-151b, miR-371b-5p, miR-455-5p, miR-195-5p, miR-1266-5p, miR-574-5p, miR-425-5p	[39]
MLL-rearranged, T-ALL	miR-196b		[41]
TEL-AML1 BCR-ABL, E2A-PBX1, hyperdiploid, and B-other	miRNA-708		
<b>Adults</b>			
T-ALL	miR-148a, miR-424	miR-151	[40]
E2A/PBX1-positive B-ALL	miR-425-5p, miR-191, miR-128		
MLL/AF4-positive B-ALL	miR-629		
BCR/ABL-positive B-ALL	miR-146b, miR-126		

**Table 1.**  
*Expression of miRNAs in children and adults to differentiate acute lymphoblastic leukemia subtypes.*

in seven major subtypes of pediatric ALL, which included: T-cell, MLL-rearranged, TEL-AML1-positive, E2A-PBX1-positive, hyperdiploid ALL, BCR-ABL-positive, and B-other ALLs, the differential miRNA expression. miRNA-708 was highly expressed in TEL-AML1, BCR-ABL, E2A-PBX1, hyperdiploid, and B-other cases than in the MLL-rearranged and T-ALL cases. On the other hand, the expression of miR-196b was higher in MLL-rearranged and T-ALL cases as compared with the expression level in the precursor B-ALL cases [42]. This information suggests that upregulated expression of miR-424 and downregulated expression of miR-151 might be good diagnostic markers to differentiate T-ALL regardless of age (**Table 1**).

Malik and collaborators propose a novel miR-2909-KLF4 molecular axis to differentiate the pathogeneses of pediatric B- and T-cell ALLs that may represent a new diagnostic marker, through alterations in miRNA expression patterns and their respective targets. The authors demonstrate the ability of miR-2909 to repress KLF4 expression in pediatric B-ALL, but not T-ALL [43]. Another interesting work shows that miR-19b, miR-20a, miR-26a, miR-92, and miR-223 have cooperative effects on tumor suppressor genes implicated in the pathogenesis of T-ALL, including *IKAROS*, *PTEN*, *BIM*, *PHF6*, *NF1*, and *FBXW7*. Interestingly, these miRNAs are capable of promoting T-ALL development in a mouse model [44].

**5. MicroRNAs as prognostic markers in ALL**

MiRNAs are suggested as promising biomarkers not only in the diagnosis but also in the prognosis of ALL patients. Since they have been promising in identifying subgroups of patients with different clinical outcomes [45]. It has been observed that ectopic expression of miRNAs leads to the development of leukemia, such is the case of miR-125b, which has been reported in mice transplanted with fetal liver

cells ectopically expressing miR-125b that showed an increase in white blood cell count, in particular in neutrophils and monocytes, associated with a macrocytic anemia. These mice developed B-cell acute lymphoblastic leukemia, T-cell acute lymphoblastic leukemia, or a myeloproliferative neoplasm, suggesting an important role for miR-125b in early hematopoiesis [46].

Patients group with high miR-21 expression was significantly associated with those aged <2 and > 10 years, lower platelets count, more incidence of central nervous system (CNS) infiltration, and poorer treatment outcome also; patients with high miR-21 showed a significantly poorer disease-free survival (DFS) and overall survival (OS) compared with those with low miR-21 expression group [47]. Also, miR-92a expression is significantly higher in ALL compared with peripheral blood mononuclear cells (PBMNCs) from healthy volunteers. Likewise, the expression levels of miR-99a, miR-100, and miR-128b correlated high-risk prognostic factors, including white blood cell (WBC) count, ALL subclassification (T-cell and B-cell ALL), the MLL-rearranged gene, and the BCR-ABL fusion gene, suggesting possible relation of miR-99a, miR-100, and miR-218b with prognosis [48, 49]. It has also been reported that mir-125b-2 is highly expressed in childhood ETV6/RUNX1 (TEL/AML1) leukemias and confers survival advantage to growth inhibitory signals independent of p53 [50].

More specifically, miR-9, miR-24, and miR-92a expression was significantly increased in a subset of ALL cells, and ALL patients with overexpressed miR-24 and miR-92a had poor prognoses [51–53]. Wang et al. (2010) observed that miR-146a, miR-181a/c, and miR-221 were significantly associated with overall survival of the ALL patients. Expression level of miR-146a and miR-181a/c was associated with a poor outcome (i.e., poor prognosis/short-term survival), whereas that of miR-221 was associated with a good outcome (i.e., good prognosis/long-term survival) [54], while that of miR-423-5p is associated with a poorer survival in patients with ALL [55]. Otherwise, the reduced expression of miR-155, miR-181b, miR-182, miR-143, miR-210, and miR-335 is associated with poor outcome of pediatric ALL [56–60]. Also, the expression of miRNAs miR-18a, miR-532, miR-218, miR-625, miR-193a, miR-638, miR-550, and miR-633 is associated with early relapse in childhood ALL, suggesting possible relation of these miRNAs with prognosis [61].

The high miR-16 expression is associated with hyperleukocytosis and poor cytogenetic groups. In B-cell ALL patients, the DFS was significantly shorter in patients with high miR-16 levels. While in T-cell ALL patients, for both DFS and overall survival, a significant trend was found with a survival shortening from the lowest to the highest miR-16 levels [62, 63]. Likewise, it was reported that the expression of miR-16 was upregulated in cases of T lymphoblastic lymphoma/leukemia (T-LBL/ALL), and the high expression group of miR-16 was significantly correlated with longer over survival [64].

For instance, Gimenes-Teixeira et al. reported that T-ALL patients with high miR-221 expression had significantly lower 5-year overall survival (OS) rates compared with those with low miR-221 expression [65]. Oliveira et al. observed that lower levels of miR-29a were significantly associated with higher blast counts in the bone marrow and with increased disease-free survival in T-ALL patients [66].

## **6. miRNAs in response to commonly used chemotherapy agents in pediatric acute lymphoblastic leukemia**

Despite the great effort of current treatment strategies, drug resistance still remains a major cause of chemotherapy failure and relapse in pediatric patients.



miRNAs have not only become tools for classifying subtypes of ALL and in support of the prognosis of this disease, but also studies have reported the classification of patients sensitive or resistant to drugs based on the expression of miRNAs.

Glucocorticoids (GCs) regulate proliferation, differentiation, metabolism, and cell survival in many tissues. In lymphocytes, they affect cell cycle progression, influence immunoglobulin and lymphokine production, and induce apoptosis in immature lymphoblasts [67]. Actually, these drugs are used clinically in the treatment of childhood acute lymphoblastic leukemia (ALL) and other lymphoid malignancies. In the group of glucocorticoids that is administered to patients with ALL is the prednisone; unfortunately, a proportion of patients are insensitive to this drug. A study in 49 ALL patients showed that miR-18a, miR-532, miR-218, miR-625, miR-193a, miR-638, miR-550, and miR-633 could distinguish prednisone-sensitive patients from prednisone-insensitive patients [68]. In contrast, other authors in a group of 81 children with newly diagnosed ALL, no discriminative microRNAs were found for prednisolone response [69].

It is well known that the presence of translocations in ALL is a frequent and prognostic influence event. In leukemia, MLL rearrangements are a common genetic alteration; MLL-AF4 acute lymphocytic leukemia (ALL), resulting from a balanced translocation between *MLL* and *AF4*, occurs in approximately 50% of ALL cases in infants, 2% in children, and 5–6% in adults. The poor prognosis of MLL-AF4 ALL to glucocorticoid-induced apoptosis is associated with its resistance to this drug [70]. miR-128b and miR-221 are commonly downregulated in MLL-rearranged ALL compared with other types of ALL; also these miRNAs downregulate mRNAs encoding CDKN1B, MLL, AF4, and both MLL-AF4 and AF4-MLL fusion genes that are thought to contribute to leukemia development [71]. Interestingly, the restoration of miRNA-128b downregulates target genes including *MLL*, *AF4*, and both *MLL-AF4* and *AF4-MLL* fusion oncogenes, and the restoration of miRNA-221 downregulates CDKN1B cooperatively. Thus, the sensitivity of MLL-AF4 ALL cells to GCs is strengthened [71]. Study developed by Kotani et al. supports the idea that restoration of miRNA-128b improves the sensitivity of MLL-AF4 ALL cells to GCs. This author mentioned that one novel mutation of miRNA-128b significantly reduced its processing, and the resultant downregulation of mature miRNA-128b gave rise to GCs resistance due to the failure to downregulate the fusion oncogenes [72]. This suggests that miRNA-128b and miRNA-221 could be GC (dexamethasone) sensitizers potential.

Other microRNAs related with drug resistance in pediatric acute lymphoblastic leukemia are miR-454, which present a low expression in L-asparaginase-resistant cases, whereas miR-125b, miR-99a, and miR-100 show an upregulation of their expression in patients resistant to vincristine and daunorubicin [69].

## **7. miRNAs as therapeutic targets in acute lymphoblastic leukemia**

Nowadays, advances in our understanding of the molecular carcinogenesis of the human cancers and the extensive research on generate and implement new combined and targeted therapies, and have allowed to know specific molecular therapeutic targets. However, there is still a continuous need for development of new therapeutic tools for applicability.

RNA molecules actually are the therapeutic targets promising in the molecular oncology. The ability of miRNAs to regulate important cellular processes, by concurrently regulating multiple targets, their inherent role in carcinogenesis as oncogenes or tumor suppressor genes, and the aberrant dysregulation of their



expression levels in cancer, can represent a viable therapeutic strategy and a powerful intervention tool in **leukemia** [73]. For example, in leukemia cells isolated from individuals with BCR/ABL, TKI-resistant Philadelphia-chromosome-positive acute lymphoblastic leukemia (Ph + ALL) was observed an increase in levels of DNMT3A in association with downregulation of miR-217; these observations are clinically relevant; and inhibition of DNMT3A by forced expression of miRNA-217 may benefit in preventing drug resistance to TKI treatment in Philadelphia-chromosome-positive ALL patients [74]. Another therapeutic strategy for BCR-ABL-positive ALL is miRNA-203, which has as direct target to BCR-ABL1 and ABL1, proteins with activity tyrosine kinase. This miRNA is silenced by genetic and epigenetic mechanisms in hematopoietic malignancies expressing either ABL1 or BCR-ABL1. However, the restoration of the miRNA-203 expression reduces ABL1 and BCR-ABL1 levels and inhibits cell proliferation [75]. miRNA-143 was identified as a regulator of MLL-AF4 expression and is epigenetically repressed by promoter hypermethylation in MLL-AF4-positive primary blasts and cell lines; upregulation of miRNA-143 expression by demethylation has therapeutic promise for MLL-AF4 B-cell ALL [76].

It is also important to consider that some miRNAs can behave as oncogenes in one cancer type and as tumor suppressive genes in others. It has been reported that miR-221 maintains a high expression in hepatic cancer and exerts an oncogenic function by targeting tumor suppressor PTEN, but this miRNA acts as a tumor suppressor in erythroblastic leukemia by inhibiting the KIT oncogene expression [77, 78]. Thus, identification of specific biological functions, type of cancer, and targets of miRNAs is a basic aspect when considering miRNA therapeutics.

## **8. Summary and future directions**

Various studies have demonstrated that the oncomiRs or tumor suppressor miRNAs expression may significantly have potential how diagnostic and/or prognostic biomarkers, as well as monitoring the disease progression and in the response to treatment, and it may be a therapeutic target for treatment in ALL. Also, miRNAs expression levels may play an important role in the genesis and evolution of the ALL. Nevertheless, the biological effects and relevant target genes of many miRNAs that are deregulated and/or prognostically relevant in ALL need to be identified and characterized. Therefore, novel anti-ALL agents are needed to overcome chemotherapy resistance and reduce cytotoxicity. The mimics- and/or anti-miRNAs may be a good alternative. However, more experiments are required to evaluate the feasibility and safety of mimics- and/or anti-miRNAs in the clinical treatment.

## **Acknowledgements**

This work was supported by the Universidad Autónoma de Guerrero. Yazmín Gómez-Gómez (CVU: 236728) and Jorge Organista-Nava (CVU: 236745) were recipient of postdoctoral fellowships from CONACYT.

## **Conflict of interest**

The authors declare that there are no conflicts of interest.

IntechOpen

### Author details

Yazmín Gómez-Gómez<sup>†</sup>, Jorge Organista-Nava<sup>\*†</sup>, Berenice Illades-Aguilar and Marco Antonio Leyva-Vázquez\*  
Molecular Biomedicine Laboratory, School of Chemical-Biological Sciences,  
Autonomous University of Guerrero, Chilpancingo, Guerrero, Mexico

\*Address all correspondence to: [leyvamarco13@gmail.com](mailto:leyvamarco13@gmail.com) and [joorna@gmail.com](mailto:joorna@gmail.com)

<sup>†</sup> These authors contributed equally to this work

### IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

## References

- [1] Johanson TM, Skinner JPJ, Kumar A, Zhan Y, Lew AM, Chong MMW. The role of microRNAs in lymphopoiesis. *International Journal of Hematology*. 2014;**100**(3):246-253. DOI: 10.1007/s12185-014-1606-y
- [2] Ultimo S, Martelli AM, Zauli G, Vitale M, Calin GA, Neri LM. Roles and clinical implications of MicroRNAs in acute lymphoblastic leukemia. *Journal of Cellular Physiology*. 2018;**233**(8):5642-5654. DOI: 10.1002/jcp.26290
- [3] Garzon R, Croce CM. MicroRNAs in normal and malignant hematopoiesis. *Current Opinion in Hematology*. 2008;**15**(4):352-358. DOI: 10.1097/MOH.0b013e328303e15d
- [4] Zhao H, Wang D, Du W, Gu D, Yang R. MicroRNA and leukemia: Tiny molecule, great function. *Critical Reviews in Oncology/Hematology*. 2010;**74**(3):149-155. DOI: 10.1016/j.critrevonc.2009.05.001
- [5] Calin GA, Dumitru CD, Shimizu M, Bichi R, Zupo S, Noch E, et al. Frequent deletions and down-regulation of micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proceedings of the National Academy of Sciences of the United States of America*. 2002;**99**(24):15524-15529. DOI: 10.1073/pnas.242606799
- [6] Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proceedings of the National Academy of Sciences of the United States of America*. 2004;**101**(9):2999-3004. DOI: 10.1073/pnas.0307323101
- [7] Lee Y, Kim M, Han J, Yeom K-H, Lee S, Baek SH, et al. MicroRNA genes are transcribed by RNA polymerase II. *The EMBO Journal*. 2004;**23**(20):4051-4060. DOI: 10.1038/sj.emboj.7600385
- [8] Lin S, Gregory RI. MicroRNA biogenesis pathways in cancer. *Nature Reviews Cancer*. 2015;**15**(6):321-333. DOI: 10.1038/nrc3932
- [9] Han J, Lee Y, Yeom K-H, Kim Y-K, Jin H, Kim VN. The Drosha-DGCR8 complex in primary microRNA processing. *Genes & Development*. 2004;**18**(24):3016-3027. DOI: 10.1101/gad.1262504
- [10] Acunzo M, Romano G, Wernicke D, Croce CM. MicroRNA and cancer—A brief overview. *Advances in Biological Regulation*. 2015;**57**:1-9. DOI: 10.1016/j.jbior.2014.09.013
- [11] Wu Q, Song R, Ortogero N, Zheng H, Evanoff R, Small CL, et al. The RNase III enzyme DROSHA is essential for microRNA production and spermatogenesis. *The Journal of Biological Chemistry*. 2012;**287**(30):25173-25190. DOI: 10.1074/jbc.M112.362053
- [12] Kim VN. MicroRNA precursors in motion: Exportin-5 mediates their nuclear export. *Trends in Cell Biology*. 2004;**14**(4):156-159. DOI: 10.1016/j.tcb.2004.02.006
- [13] Yi R, Qin Y, Macara IG, Cullen BR. Exportin-5 mediates the nuclear export of pre-microRNAs and short hairpin RNAs. *Genes & Development*. 2003;**17**(24):3011-3016. DOI: 10.1101/gad.1158803
- [14] Bohnsack MT, Czaplinski K, Gorlich D. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of pre-miRNAs. *RNA*. 2004;**10**(2):185-191. DOI: 10.1261/rna.5167604

- [15] Chendrimada TP, Gregory RI, Kumaraswamy E, Norman J, Cooch N, Nishikura K, et al. TRBP recruits the Dicer complex to Ago2 for microRNA processing and gene silencing. *Nature*. 2005;**436**(7051):740-744. DOI: 10.1038/nature03868
- [16] Bartel DP. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell*. 2004;**116**(2):281-297. DOI: 10.1016/S0092-8674(04)00045-5
- [17] Diederichs S, Haber DA. Dual role for argonautes in microRNA processing and posttranscriptional regulation of microRNA expression. *Cell*. 2007;**131**(6):1097-1108. DOI: 10.1016/j.cell.2007.10.032
- [18] Luan C, Yang Z, Chen B. The functional role of microRNA in acute lymphoblastic leukemia: Relevance for diagnosis, differential diagnosis, prognosis, and therapy. *OncoTargets and Therapy*. 2015;**8**:2903-2914. DOI: 10.2147/OTT.S92470
- [19] Han Y-C, Park CY, Bhagat G, Zhang J, Wang Y, Fan J-B, et al. microRNA-29a induces aberrant self-renewal capacity in hematopoietic progenitors, biased myeloid development, and acute myeloid leukemia. *Journal of Experimental Medicine*. 2010;**207**(3):475-489. DOI: 10.1084/jem.20090831
- [20] Popovic R, Riesbeck LE, Velu CS, Chaubey A, Zhang J, Achille NJ, et al. Regulation of mir-196b by MLL and its overexpression by MLL fusions contributes to immortalization. *Blood*. 2009;**113**(14):3314-3322. DOI: 10.1182/blood-2008-04-154310
- [21] Manterola L, Fernandez-Mercado M, Larrea E, Goicoechea I, Arestin M, Armesto M, et al. MicroRNAs as B-cell lymphoma biomarkers. *Blood and Lymphatic Cancer: Targets and Therapy*. 2015;**5**:25-34. DOI: 10.2147/BLCTT.S60481
- [22] Zhou B, Wang S, Mayr C, Bartel DP, Lodish HF. miR-150, a microRNA expressed in mature B and T cells, blocks early B cell development when expressed prematurely. *Proceedings of the National Academy of Sciences of the United States of America*. 2007;**104**(17):7080-7085. DOI: 10.1073/pnas.0702409104
- [23] He Y, Jiang X, Chen J. The role of miR-150 in normal and malignant hematopoiesis. *Oncogene*. 2014;**33**(30):3887-3893. DOI: 10.1038/onc.2013.346
- [24] Seddiki N, Brezar V, Ruffin N, Lévy Y, Swaminathan S. Role of mi R-155 in the regulation of lymphocyte immune function and disease. *Immunology*. 2014;**142**(1):32-38. DOI: 10.1111/imm.12227
- [25] Banerjee A, Schambach F, DeJong CS, Hammond SM, Reiner SL. MicroRNA-155 inhibits IFN- $\gamma$  signaling in CD4<sup>+</sup> T cells. *European Journal of Immunology*. 2010;**40**(1):225-231. DOI: 10.1002/eji.200939381
- [26] Chen C-Z, Li L, Lodish HF, Bartel DP. MicroRNAs modulate hematopoietic lineage differentiation. *Science*. 2004;**303**(5654):83-86. DOI: 10.1126/science.1091903
- [27] Verduci L, Azzalin G, Gioiosa S, Carissimi C, Laudadio I, Fulci V, et al. microRNA-181a enhances cell proliferation in acute lymphoblastic leukemia by targeting EGR1. *Leukemia Research*. 2015;**39**(4):479-485. DOI: 10.1016/j.leukres.2015.01.010
- [28] Lindner SE, Lohmüller M, Kotkamp B, Schuler F, Knust Z, Villunger A, et al. The miR-15 family reinforces the transition from proliferation to differentiation in pre-B cells. *EMBO Reports*. 2017;**18**(9):1604-1617. DOI: 10.15252/embr.201643735



- [29] Violaine H, Ramiro G. Micronas: Emerging key regulators of hematopoiesis. *American Journal of Hematology*. 2010;**85**(12):935-942. DOI: 10.1002/ajh.21863
- [30] Ventura A, Young AG, Winslow MM, Lintault L, Meissner A, Erkland SJ, et al. Targeted deletion reveals essential and overlapping functions of the miR-17~92 family of miRNA clusters. *Cell*. 2008;**132**(5):875-886. DOI: 10.1016/j.cell.2008.02.019
- [31] Lai M, Gonzalez-Martin A, Cooper AB, Oda H, Jin HY, Shepherd J, et al. Regulation of B-cell development and tolerance by different members of the miR-17~92 family microRNAs. *Nature Communications*. 2016;**7**:12207. DOI: 10.1038/ncomms12207
- [32] Jiang S, Li C, Olive V, Lykken E, Feng F, Sevilla J, et al. Molecular dissection of the miR-17-92 cluster's critical dual roles in promoting Th1 responses and preventing inducible Treg differentiation. *Blood*. 2011;**118**(20):5487-5497. DOI: 10.1182/blood-2011-05-355644
- [33] Smith KM, Guerau-de-Arellano M, Costinean S, Williams JL, Bottoni A, Mavrikis Cox G, et al. miR-29ab1 deficiency identifies a negative feedback loop controlling Th1 bias that is dysregulated in multiple sclerosis. *Journal of Immunology*. 2012;**189**(4):1567-1576. DOI: 10.4049/jimmunol.1103171
- [34] Sawant DV, Wu H, Kaplan MH, Dent AL. The Bcl6 target gene microRNA-21 promotes Th2 differentiation by a T cell intrinsic pathway. *Molecular Immunology*. 2013;**54**(3-4):435-442. DOI: 10.1016/j.molimm.2013.01.006
- [35] Mattes J, Collison A, Plank M, Phipps S, Foster PS. Antagonism of microRNA-126 suppresses the effector function of TH2 cells and the development of allergic airways disease. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;**106**(44):18704-18709. DOI: 10.1073/pnas.0905063106
- [36] Crazzolara R, Bendall L. Emerging treatments in acute lymphoblastic leukemia. *Current Cancer Drug Targets*. 2009;**9**(1):19-31. DOI: 10.2174/156800909787314057
- [37] Li W-y, X-m C, Xiong W, D-m G, Lu L, Li H-y. Detection of microvesicle miRNA expression in ALL subtypes and analysis of their functional roles. *Journal of Huazhong University of Science and Technology [Medical Sciences]*. 2014;**34**(5):640-645. DOI: 10.1007/s11596-014-1330-0
- [38] Pradillo M, Santos JL. Genes involved in miRNA biogenesis affect meiosis and fertility. *Chromosome Research*. 2018;**26**:1-9. DOI: 10.1007/s10577-018-9588-x
- [39] Oliveira JC, Scrideli CA, Brassesco MS, Morales AG, Pezuk JA, Queiroz RP, et al. Differential MiRNA expression in childhood acute lymphoblastic leukemia and association with clinical and biological features. *Leukemia Research*. 2012;**36**(3):293-298. DOI: 10.1016/j.leukres.2011.10.005
- [40] Santos Almeida R, Costa e Silva M, Coutinho LL, Gomes RG, Pedrosa F, Massaro JD, et al. MicroRNA expression profiles discriminate childhood T-from B-acute lymphoblastic leukemia. *Hematological Oncology*. 2018;**0**(0):2567. DOI: 10.1002/hon.2567
- [41] Fulci V, Colombo T, Chiaretti S, Messina M, Citarella F, Tavolaro S, et al. Characterization of B- and T-lineage acute lymphoblastic leukemia by integrated analysis of MicroRNA and mRNA expression profiles. *Genes, Chromosomes and Cancer*. 2009;**48**(12):1069-1082. DOI: 10.1002/gcc.20709

- [42] Schotte D, Chau JCK, Sylvester G, Liu G, Chen C, van der Velden VHJ, et al. Identification of new microRNA genes and aberrant microRNA profiles in childhood acute lymphoblastic leukemia. *Leukemia*. 2008;**23**(2): 313-322. DOI: 10.1038/leu.2008.28
- [43] Malik D, Kaul D, Chauhan N, Marwaha RK. miR-2909-mediated regulation of KLF4: A novel molecular mechanism for differentiating between B-cell and T-cell pediatric acute lymphoblastic leukemias. *Molecular Cancer*. 2014;**13**:175-190. DOI: 10.1186/1476-4598-13-175
- [44] Mavrakis KJ, Van Der Meulen J, Wolfe AL, Liu X, Mets E, Taghon T, et al. A cooperative microRNA-tumor suppressor gene network in acute T-cell lymphoblastic leukemia (T-ALL). *Nature Genetics*. 2011;**43**(7):673-678. DOI: 10.1038/ng.858
- [45] Cao J, Cai J, Huang D, Han Q, Chen Y, Yang Q, et al. miR-335 represents an independent prognostic marker in epithelial ovarian cancer. *American Journal of Clinical Pathology*. 2014;**141**(3):437-442. DOI: 10.1309/AJCPLYTZGB54ISZC
- [46] Bousquet M, Harris MH, Zhou B, Lodish HF. MicroRNA miR-125b causes leukemia. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;**107**(50):21558-21563. DOI: 10.1073/pnas.1016611107
- [47] Labib HA, Elantouny NG, Ibrahim NF, Alnagar AA. Upregulation of microRNA-21 is a poor prognostic marker in patients with childhood B cell acute lymphoblastic leukemia. *Hematology*. 2017;**22**(7):392-397. DOI: 10.1080/10245332.2017.1292204
- [48] Li XJ, Luo XQ, Han BW, Duan FT, Wei PP, Chen YQ. MicroRNA-100/99a, deregulated in acute lymphoblastic leukaemia, suppress proliferation and promote apoptosis by regulating the FKBP51 and IGF1R/mTOR signalling pathways. *British Journal of Cancer*. 2013;**109**(8):2189-2198. DOI: 10.1038/bjc.2013.562
- [49] Nemes K, Csóka M, Nagy N, Márk Á, Váradi Z, Dankó T, et al. Expression of certain leukemia/lymphoma related microRNAs and its correlation with prognosis in childhood acute lymphoblastic leukemia. *Pathology & Oncology Research*. 2015;**21**(3):597-604. DOI: 10.1007/s12253-014-9861-z
- [50] Gefen N, Binder V, Zaliova M, Linka Y, Morrow M, Novosel A, et al. Hsa-mir-125b-2 is highly expressed in childhood ETV6/RUNX1 (TEL/AML1) leukemias and confers survival advantage to growth inhibitory signals independent of p53. *Leukemia*. 2010;**24**(1):89-96. DOI: 10.1038/leu.2009.208
- [51] Ohyashiki JH, Umezumi T, Kobayashi C, Hamamura RS, Tanaka M, Kuroda M, et al. Impact on cell to plasma ratio of miR-92a in patients with acute leukemia: in vivo assessment of cell to plasma ratio of miR-92a. *BMC Research Notes*. 2010;**3**:347. DOI: 10.1186/1756-0500-3-347
- [52] Organista-Nava J, Gómez-Gómez Y, Illades-Aguilar B, Del Carmen Alarcón-Romero L, Saavedra-Herrera MV, Rivera-Ramírez AB, et al. High miR-24 expression is associated with risk of relapse and poor survival in acute leukemia. *Oncology Reports*. 2015;**33**(4):1639-1649. DOI: 10.3892/or.2015.3787
- [53] Sugita F, Maki K, Nakamura Y, Sasaki K, Mitani K. Overexpression of MIR9 indicates poor prognosis in acute lymphoblastic leukemia. *Leukemia & Lymphoma*. 2014;**55**(1):78-86. DOI: 10.3109/10428194.2013.790023
- [54] Wang Y, Li Z, He C, Wang D, Yuan X, Chen J, et al. MicroRNAs expression signatures are associated with lineage and survival in acute leukemias.

Blood Cells, Molecules & Diseases. 2010;**44**(3):191-197. DOI: 10.1016/j.bcmd.2009.12.010

[55] Mosakhani N, Sarhadi VK, Usvasalo A, Karjalainen-Lindsberg M-L, Lahti L, Tuononen K, et al. MicroRNA profiling in pediatric acute lymphoblastic leukemia: Novel prognostic tools. *Leukemia & Lymphoma*. 2012;**53**(12):2517-2520. DOI: 10.3109/10428194.2012.685731

[56] Yan J, Jiang N, Huang G, Tay JLS, Lin B, Bi C, et al. Deregulated MIR 335 that targets MAPK 1 is implicated in poor outcome of paediatric acute lymphoblastic leukaemia. *British Journal of Haematology*. 2013;**163**(1): 93-103. DOI: 10.1111/bjh.12489

[57] Piatopoulou D, Avgeris M, Drakaki I, Marmarinos A, Xagorari M, Baka M, et al. Clinical utility of miR-143/ miR-182 levels in prognosis and risk stratification specificity of BFM-treated childhood acute lymphoblastic leukemia. *Annals of Hematology*. 2018;**97**(7):1169-1182. DOI: 10.1007/s00277-018-3292-y

[58] Mei Y-Y, Li Z-G, Zhang Y, Zhang W-L, Zhang P-W, Wang N, et al. Prognostic significance of joint detection of miR-210 and minimal residual disease in pediatric acute lymphoblastic leukemia. *Zhongguo Shi Yan Xue Ye Xue Za Zhi*. 2017;**25**(1):66-71. DOI: 10.7534/j.issn.1009-2137.2017.01.011

[59] Mei Y, Gao C, Wang K, Cui L, Li W, Zhao X, et al. Effect of microRNA-210 on prognosis and response to chemotherapeutic drugs in pediatric acute lymphoblastic leukemia. *Cancer Science*. 2014;**105**(4):463-472. DOI: 10.1111/cas.12370

[60] Zhou G, Cao Y, Dong W, Lin Y, Wang Q, Wu W, et al. The clinical characteristics and prognostic significance of AID, miR-181b,

and miR-155 expression in adult patients with de novo B-cell acute lymphoblastic leukemia. *Leukemia & Lymphoma*. 2017;**58**(9):2118-2126. DOI: 10.1080/10428194.2017.1283028

[61] Xu L, Y-n L, X-q L, X-d L, H-x G. Association of miRNAs expression profiles with prognosis and relapse in childhood acute lymphoblastic leukemia. *Zhonghua Xue Ye Xue Za Zhi*. 2011;**32**(3):178-181

[62] Kaddar T, Chien WW, Bertrand Y, Pages MP, Rouault JP, Salles G, et al. Prognostic value of miR-16 expression in childhood acute lymphoblastic leukemia relationships to normal and malignant lymphocyte proliferation. *Leukemia Research*. 2009;**33**(9):1217-1223. DOI: 10.1016/j.leukres.2008.12.015

[63] Tong L-G, Wu W-Z, Zhang Y-P, Zhou Z-G, Chen Y-F, Huang W-J, et al. Expression of miR-16 in patients with T lymphoblastic lymphoma/ acute lymphoblastic leukemia. *Zhongguo Shi Yan Xue Ye Xue Za Zhi*. 2014;**22**(1):99-103. DOI: 10.7534/j.issn.1009-2137.2014.01.020

[64] Li J, Li P, Wang J, Xi Y. Significance of microRNA-16 and bcl-2 expression in T lymphoblastic lymphoma/leukemia and its relation with prognosis. *Zhonghua Bing Li Xue Za Zhi*. 2013;**42**(11):748-752. DOI: 10.3760/cma.j.issn.0529-5807.2013.11.007

[65] Gimenes-Teixeira HL, Lucena-Araujo AR, Dos Santos GA, Zanette DL, Scheucher PS, Oliveira LC, et al. Increased expression of miR-221 is associated with shorter overall survival in T-cell acute lymphoid leukemia. *Experimental Hematology & Oncology*. 2013;**2**(1):10. DOI: 10.1186/2162-3619-2-10

[66] Oliveira LH, Schiavinato JL, Fráguas MS, Lucena-Araujo AR, Haddad R, Araújo AG, et al. Potential roles of microRNA-29a in the molecular



pathophysiology of T-cell acute lymphoblastic leukemia. *Cancer Science*. 2015;**106**(10):1264-1277. DOI: 10.1111/cas.12766

[67] Schaaf MJM, Cidlowski JA. Molecular mechanisms of glucocorticoid action and resistance. *The Journal of Steroid Biochemistry and Molecular Biology*. 2002;**83**(1):37-48. DOI: 10.1016/S0960-0760(02)00263-7

[68] Xu L, Liang Y, Luo X, Liu X, Guo H. Association of miRNAs expression profiles with prognosis and relapse in childhood acute lymphoblastic leukemia. *Zhonghua xue ye xue za zhi= Zhonghua xueyexue zazhi*. 2011;**32**(3):178-181

[69] Schotte D, De Menezes RX, Akbari Moqadam F, Khankahdani LM, Lange-Turenhout E, Chen C, et al. MicroRNA characterize genetic diversity and drug resistance in pediatric acute lymphoblastic leukemia. *Haematologica*. 2011;**96**(5):703-711. DOI: 10.3324/haematol.2010.026138

[70] Pui CH, Chessells JM, Camitta B, Baruchel A, Biondi A, Boyett JM, et al. Clinical heterogeneity in childhood acute lymphoblastic leukemia with 11q23 rearrangements. *Leukemia*. 2003;**17**(4):700-706. DOI: 10.1038/sj.leu.2402883

[71] Kotani A, Ha D, Hsieh J, Rao PK, Schotte D, den Boer ML, et al. miR-128b is a potent glucocorticoid sensitizer in MLL-AF4 acute lymphocytic leukemia cells and exerts cooperative effects with miR-221. *Blood*. 2009;**114**(19):4169-4178. DOI: 10.1182/blood-2008-12-191619

[72] Kotani A, Ha D, Schotte D, den Boer ML, Armstrong SA, Lodish HF. A novel mutation in the miR-128b gene reduces miRNA processing and leads to glucocorticoid resistance of MLL-AF4 acute lymphocytic leukemia cells. *Cell cycle (Georgetown, Tex)*.

2010;**9**(6):1037-1042. DOI: 10.4161/cc.9.6.11011

[73] Shah MY, Ferrajoli A, Sood AK, Lopez-Berestein G, Calin GA. microRNA therapeutics in cancer—An emerging concept. *eBioMedicine*. 2016;**12**:34-42. DOI: 10.1016/j.ebiom.2016.09.017

[74] Nishioka C, Ikezoe T, Yang J, Nobumoto A, Tsuda M, Yokoyama A. Downregulation of miR-217 correlates with resistance of Ph(+) leukemia cells to ABL tyrosine kinase inhibitors. *Cancer Science*. 2014;**105**(3):297-307. DOI: 10.1111/cas.12339

[75] Bueno MJ, de Castro IP, de Cedrón MG, Santos J, Calin GA, Cigudosa JC, et al. Genetic and epigenetic silencing of microRNA-203 enhances ABL1 and BCR-ABL1 oncogene expression. *Cancer Cell*. 2016;**29**(4):607-608. DOI: 10.1016/j.ccell.2016.03.013

[76] Dou L, Zheng D, Li J, Li Y, Gao L, Wang L, et al. Methylation-mediated repression of microRNA-143 enhances MLL-AF4 oncogene expression. *Oncogene*. 2011;**31**(4):507-517. DOI: 10.1038/onc.2011.248

[77] Pineau P, Volinia S, McJunkin K, Marchio A, Battiston C, Terris B, et al. miR-221 overexpression contributes to liver tumorigenesis. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;**107**(1):264-269. DOI: 10.1073/pnas.0907904107

[78] Felli N, Fontana L, Pelosi E, Botta R, Bonci D, Facchiano F, et al. MicroRNAs 221 and 222 inhibit normal erythropoiesis and erythroleukemic cell growth via kit receptor down-modulation. *Proceedings of the National Academy of Sciences of the United States of America*. 2005;**102**(50):18081-18086. DOI: 10.1073/pnas.0506216102