

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Osteoarthritis as a Chronic Inflammatory Disease: A Review of the Inflammatory Markers

José Fábio dos Santos Duarte Lana and Bruno Lima Rodrigues

Abstract

Osteoarthritis (OA) is the most prevalent joint disease and a common cause of joint pain, functional loss, and disability. In addition to macroscopic features, such as cartilage degradation with subchondral bone remodeling, hypertrophy of the joint capsule, and osteophytes formation, several cellular and molecular alterations are present in OA, which lead to a chronic low-grade inflammation. Inflammatory mediators observed in OA joints are thought to be the downstream effectors of the pathogenesis of the disease. Although cytokines are among the most extensively studied mediators of inflammation, such as IL-1 β and TNF, there has been an increase in studies showing the contribution of chemokines and adipokines in OA progression. This fact is supported by recent progress, which has considerably improved knowledge of the factors involved in the development of OA and the mechanisms responsible for its progression. Therefore, the aim of this chapter is to discuss the involvement of the inflammatory response in OA maintenance, focusing on the main inflammatory markers observed in studies with OA.

Keywords: osteoarthritis, immune response, inflammation, biomarkers, cytokines

1. Introduction

Osteoarthritis (OA) is a common disease that can affect joints from any part of the body, and it represents a major cause of disability and joint pain worldwide [1, 2]. OA most commonly affects the knee, hip, and shoulder, and it was estimated that more than 25 million people in the USA were affected by some form of OA in the last decade [3]. In addition, OA presents a high susceptibility to affect female gender, elderly people, and obese individuals [4].

The progression of OA leads to cartilage degradation with subchondral bone remodeling, hypertrophy of the joint capsule, and osteophytes formation, causing pain [1, 5, 6]. Although the development of OA is considered a heterogeneous process, which comprises a number of genetic and environmental causes, the presence of local causes, such as trauma and hypermobility of the joint, may worsen OA [2, 7].

The accurate identification of osteoarthritic features has been studied in order to radiographically grade the stages of OA. The Kellgren-Lawrence classification is the most widely used, especially in clinical researches. This classification evaluates the appearance of osteophytes and cysts, joint space loss, and sclerosis, and it grades the severity from 0 to 5 points. The radiological features found in OA joints were

graded as follows: (1) formation of osteophytes on joint margins or on tibia spines for knee OA; (2) periarticular ossicles in relation to distal and proximal interphalangeal joints; (3) narrowing of joint cartilage and sclerosis of subchondral bone; (4) pseudocystic areas with sclerotic walls in the subchondral bone; and (5) altered shape of the bone ends [8]. Some of these criteria were adopted by the World Health Organization (WHO) as the standard for studies on OA.

Current options for the treatment of OA focus on reducing pain (non-steroidal anti-inflammatory drugs—NSAIDs) and joint viscosupplementation (intra-articular injections of hyaluronic acid) [1]. Besides presenting a short-term effect, the chronic use of some of these medications, especially NSAIDs, may cause serious adverse events, including toxicity and risk of thromboembolism [9, 10]. In severe cases, surgical procedures, mostly joint replacement, are suggested [1]. Novel alternative therapies, called orthobiologics, have emerged from the need of tissue regeneration. Clinical trials using orthobiologics, which comprise platelet-rich plasma (PRP), bone marrow aspirate concentrate (BMAC), fat graft (Biofat), and expanded mesenchymal stem cells, have shown promising results for the treatments of OA from any origin [11–14].

Moreover, monoclonal antibody (mAb) therapy represents one of the alternative treatments that aim to control inflammation and slow structural progression [15]. This approach focuses on blocking specific molecules responsible for the maintenance of OA. Preclinical studies with ADAMTS mAbs reported a significant decrease in histological scores after 3 months of treatment [16]. Adalimumab is an anti-TNF- α therapy used in diverse immune-mediated diseases, and it presents a protective role for OA as it reduces the severity of the cartilage lesion and improves the structure of subchondral bone [17]. Since IL-1 family may induce the production of metalloproteinases (MMP), it has also become a target for mAb therapy, and, in a randomized controlled trial with patients who presented knee OA, it was reported great improvement on pain relief [18].

In addition to macroscopic features, several cellular and molecular alterations are present in OA, such as catabolism and anabolism events; hypertrophy and, consequently, death of chondrocytes; and impaired autophagy process [19]. Also, a chronic low-grade inflammation interplayed with immune system has been considered to present a crucial role in the maintenance of OA [1]. This fact is supported by recent progress, which has considerably improved the knowledge regarding factors involved in the OA development and the mechanisms responsible for its progression.

2. Osteoarthritis and immune response

The inflammation observed in OA is believed to involve innate immune response prior to a mild degree of adaptive immunity [20]. During tissue damage, a group of endogenous molecules, called damage-associated molecular pattern (DAMP), signals the immune cells to induce a protective response against the tissue, causing tissue repair. However, a prolonged signaling of DAMP to immune cells leads to an exacerbated cytokine release, which can be destructive to the tissue [21, 22].

Innate immune cells activated by DAMP include macrophages and mast cells, which have shown to present (displayed or demonstrated) a key role in the pathogenesis of OA. Mast cells are considered regulators of vascular permeability, and they may play a crucial role in OA joint inflammation as they facilitate leukocyte infiltration [23].

Macrophages exhibit a functional plasticity based on the environment in which they are located or present [22]. However, their chronic activation can lead to the

production of proinflammatory cytokines, which worsen the osteoarthritic joints [24]. *In vitro* studies of human OA synovium-derived cells showed that macrophage depletion results in diminishing of inflammatory response by decrease of proteolytic enzyme expression, such as metalloproteinases (MMP). This fact is supported by *in vitro* studies with cell culture suspension of human OA synovium, which reported that, after macrophage depletion, there was a decrease in the production of inflammatory response by less activity of proteolytic enzymes, such as metalloproteinases (MMP) [25]. Although macrophages may also present a protective role, as they are known to secrete transforming growth factor β (TGF- β), which would enhance cartilage repair, intra-articular injection of TGF- β in mice knee led to osteophyte formation and fibrosis [26].

In addition, natural killer (NK) infiltrates are commonly found in synovial tissue from patients who underwent joint replacement surgeries, and a subset of NK cells (CD56^{bright}) was found to be greatly expanded in patients with inflammatory arthritis who have not undergone joint replacement surgeries. However, the effect of these cells on the development of OA has not been elucidated yet [27–29]. NK cells secrete protease enzymes called granzyme type A and B, which correlate to cytolytic potency. Granzymes can be released during degranulation of cytotoxic cells and, when delivered intracellularly to the target cells, they induce apoptosis. Granzyme A also stimulates the production of tumor necrosis factor (TNF- α), IL-6, and IL-8, while granzyme B may intensify the degradation of extracellular matrix [30, 31]. Tak et al. identified both types of granzymes in synovia from OA and rheumatoid arthritis. However, another study later showed that NK cells within OA synovia were granzyme negative with potential to produce the interferon- γ (IFN- γ) when expanded with IL-2 and stimulated with cytokines known to trigger IFN- γ production in blood NK cells, such as IL-12 and IL-18 [27, 32].

The presence of IFN- γ has a role in the bone resorption and consequently in the osteoclastogenesis process, but the studies have shown controversial results in this regard: *in vitro* evidence reported that IFN- γ , via TRANCE pathway, strongly suppresses osteoclastogenesis in culture of mononuclear phagocyte cells, which are the osteoclast precursors [33], whereas in culture of peripheral blood it may enhance osteoclast production as IFN- γ increases superoxide generation by neutrophils [34]. In addition, experimental studies in which IFN- γ receptor was silenced suggested a more rapid onset of collagen-induced arthritis [35]. Although IFN- γ plays a key role in angiogenesis, there is no evidence that this cytokine is able to promote angiogenesis in OA.

Proteins from complement system have been found to play a role in OA, especially in early stages, as they were upregulated in both synovial membrane and fluid [23, 36]. Additionally, the deposition of the membrane attack complex (MAC, C5b-9) is correlated with the presence of inflammation on histology of synovial membrane, and it was present in chondrocytes in late OA [36]. MAC can lead to chondrocyte destruction as it stimulates catabolic events through the increase of leukocytes and, consequently, the production of MMP [23]. Also in the studies with experimental knockout models for C5 and C6, the joint damages were attenuated [36].

Cellular infiltrates from adaptive immune response have also been observed in synovial fluid from OA joints. Although the main cell type present in this infiltrate is CD3⁺ T cells, both CD4⁺ and CD8⁺ cells have also been found in OA [37]. Th1 cells, and consequently their secretory cytokines, such as IL-2 and INF- γ , appear to be expressed five times greater than Th2 in most of OA patients [37]. Based on lymphocyte aggregates, there is a suggestion of an active cell-mediated immune response since T-cells in lymphocytic aggregates in OA synovium were shown to bear early (CD69), intermediate (CD25 and CD38), and late (CD45RO) activation markers [38].

3. Inflammatory markers in osteoarthritis

3.1 Cytokines

Inflammatory mediators observed in OA joints are thought to be the downstream effectors of the pathogenesis of the disease. Cytokines are among the most extensively studied mediators of inflammation. Several cytokines have been reported to play a role in the progression of OA, such as TNF, IL-1 β , IL-6, IL-15, IL-17, IL-18, IL-4, and IL-10. Although their precise mechanism of action has not been completely elucidated yet, it has been proposed that their presence influences cartilage homeostasis as they induce catabolic events as well as inhibit anabolic processes [21, 39, 40].

3.1.1 IL-1 β and TNF

Interleukin (IL)-1 β and tumor necrosis factor (TNF) are considered the major mediators in the pathophysiology of OA. They both are secreted not only by immune cells, especially mononuclear cells, but also by chondrocytes and osteoblasts. In OA joints, these cytokines are increased in both synovial fluid and membrane. They are known to drive the inflammatory cascade, and their increased expression induces catabolic events as they enhance MMP [39]. IL-1 β and TNF downregulate the synthesis of major extracellular matrix (ECM) components by inhibiting anabolic activities of chondrocytes [40] and reducing type II collagen production [41].

IL-1 β is activated through the binding of its specific receptor type I (IL-1RI). Overexpression of IL-1RI in cartilage proximal to the macroscopic injury in OA joints resulting in increased binding of IL-1 β was observed [42]. IL-1 β has also been reported to be responsible for the catabolic events present in OA: its expression combined with TNF induces the production of MMP-1, -3, and -13 and stimulates the production of aggrecanases (ADAMTS)-4 and -5 in human and bovine chondrocytes [43, 44]. TNF receptor type I (TNFRI) is overexpressed in OA chondrocytes [45]. High levels of TNF- α in cartilage explants seem to inhibit the synthesis of proteoglycan and stimulate resorption [40].

In OA joint, IL-1 β and TNF amplify the arthritic condition by inducing the production of proinflammatory cytokines, such as IL-6, IL-8, and monocyte chemoattractant protein 1. In addition, chondrocytes treated with IL-1 β and TNF increase the production of nitric oxide (NO), cyclooxygenase 2 (COX-2), and prostaglandin E2 (PGE2), which contribute to articular inflammation and cartilage destruction as they enhance MMP activity, inhibit the production of anabolic products such as collagen and proteoglycan, and induce chondrocyte apoptosis [39].

The catabolic events observed (the catabolic events that occur due to the presence of...) in the presence of IL-1 β and TNF are mediated through the activation of signaling pathways, including nuclear factor- κ B (NF- κ B) signaling. NF- κ B pathway induces the expression of the genes related to the inflammatory mediators cited above and also contributes to the induction of MMP-1 and -13 and ADAMTS-4 [46]. However, some signaling pathways are involved in the downregulation of the IL-1 β and TNF effects in OA, such as peroxisome proliferator-activated receptor- γ (PPAR- γ). The activation of PPAR- γ seems to reduce the progression of cartilage lesion in experimental models of OA as it assists the downregulation of inflammatory and catabolic responses mediated by IL-1 β and TNF [47, 48].

3.1.2 IL-6

IL-6 is a proinflammatory cytokine, whose signaling pathway involves the activation of receptors, such as membrane-bound IL-6 receptor (IL6R), soluble

IL-6R (sIL-6R), and gp130, followed by the activation of STAT1 and STAT3 pathways [39]. In physiological conditions, the production of IL-6 by chondrocytes is considerably low. However, the exact mechanism of IL-6 action in OA is unknown, but its production can be stimulated by the number of cytokines and growth factors present in OA, including IL-1 β , TGF- β , and PGE₂ [25, 49].

Increased levels of IL-6 in synovial fluid and serum have been correlated with the severity of lesions in X-ray imaging [50]. *In vitro* studies have shown that IL-6, in combination with IL-1 β and TNF, upregulates the production of MMP-1 and -13 in human and bovine chondrocytes and induces proteoglycan and type II collagen degradation [51, 52]. The effect of IL-6 in studies with animal models has shown uncertain results. IL-6 knockout mice revealed more advanced degenerative changes compared to wild-type animals [53]. However, when IL-6 was injected in the joint cavity of IL-6-deficient mice, the reduction in the loss of proteoglycans in the acute phase of inflammation was observed [54].

One of the most considered active components in OA is the change in subchondral bone tissue, and IL-6 has been a critical mediator in this regard. Its effect, together with IL-1 β and TNF, is based on promoting osteoclast formation and, consequently, bone resorption [55]. In response to IL-6, osteoblasts stimulate the production of receptor activator of NF- κ B ligand, IL-1 β , and PGE₂, which activate osteoclasts [56]. In addition, osteoblasts activated by these cytokines produce MMPs, which adversely affect the surrounding cartilage [57].

3.1.3 IL-15

Despite a better documented involvement in rheumatoid arthritis [58], the knowledge regarding IL-15 and its action in OA is still poor. It acts based on the stimulation and proliferation of T cells and NK cells, and it may also induce the production of MMP [59]. IL-15 levels are elevated in synovial fluid in early stages of OA, and this concentration correlates with pain and severity of lesions seen on X-ray imaging [60, 61].

3.1.4 IL-17

Due to its inflammatory effects, IL-17 family has been implied to play a role in OA [62]. IL-17 is mainly stimulated by CD4⁺ T cells and mast cells, which are present in the cellular infiltrates observed in OA joints [63]. Within the joints, IL-17 primarily targets chondrocytes and fibroblast-like synoviocytes, which express IL-17 receptor (IL-17R) on their surface [64]. It was reported that IL-17 is able to inhibit proteoglycan synthesis by chondrocytes and increase the production of MMPs [65]. Also, high levels of IL-17 in both serum and synovial fluid were correlated with radiographic lesions in OA [66].

The genetic correlation between IL-17 and OA was suggested: a polymorphism in the gene IL-17A G-197A could be associated with the susceptibility to the development of OA [67]. In addition, IL-17 is produced by a specific T cell lineage called T helper 17, and it is able to cause hypertrophy of synovial membrane as its presence influences the secretion of vascular endothelial growth factor (VEGF), which leads to excessive blood vessel formation [68]. It can also indirectly affect cartilage by inducing the production of cytokines responsible for tissue degradation, such as IL-1 β , TNF, IL-6, NO, and PGE₂ [64].

3.1.5 IL-18

The active form of IL-18 results from the activation of caspase-1, which has been reported to be elevated in articular cartilage and synovium of OA, leading to great

promotion of IL-18 and IL-1 β . The production of IL-18 in joints is mainly determined by chondrocytes, osteoblasts, and macrophages [69]. IL-18 affects cartilage by upregulating the production of IL-18R α on chondrocyte surface and stimulates excessive production of MMP-1, -3, and -13 [70]. Also, IL-18 negatively influences the production of proteoglycans, aggrecan, and type II collagen and may cause morphological changes typically observed in apoptotic processes [71, 72].

The increased concentration of IL-18 observed in synovial fluid, synovium, cartilage, and even blood serum from patients with OA has been correlated with the severity of lesions seen in radiographic imaging [73]. Also, studies have correlated the development of OA and lumbar disc degeneration with polymorphisms in the gene encoding IL-18 and its receptor (IL-18R) [74, 75].

3.1.6 IL-4

Anti-inflammatory cytokines also present a role in the maintenance of OA. IL-4 is associated with chondroprotective effects as it is shown to reduce MMP production and, consequently, inhibit the degradation of proteoglycans in the articular cartilage [76]. However, chondrocytes from OA joints have shown a decreased susceptibility to this IL-4 protective effect, leaving the cartilage unprotected, quickening the degeneration via the action of the proinflammatory cytokines cited above [77]. In addition, a polymorphism in the gene encoding IL-4 and its main receptor (IL-4R α) could predetermine the development of OA in hand and knee joints [78, 79]. It was also further reported that, when compared with healthy patients, OA patients present an elevated level of soluble IL-4R α (sIL-4R α) [80].

The activation of IL-4 depends on intracellular signal transduction by gradual phosphorylation of IL-4R α , which leads to the expression of several proinflammatory genes [81]. IL-4 production is mainly determined by T cells, especially Th2, which are present in the cellular infiltrates observed in OA [37]. It was reported that IL-4 alone or in combination with IL-10 is able to reduce the production of diverse proinflammatory mediators, such as IL-1 β , TNF- α receptors, IL-6, PGE₂, and COX-2 [82–84].

3.1.7 IL-10

Due to its anti-inflammatory features, IL-10 is another cytokine that presents chondroprotective effects, and it is linked to the release of IFN [62]. *In vitro* studies have shown increased proteoglycan and type II collagen syntheses after the administration of IL-10 in chondrocytes [62]. The protective effects that IL-10 exhibits are likely due to a stimulation of the synthesis of IL-1 β antagonist and a tissue inhibition of MMP-1 (TIMP-1) [85]. Also, IL-10, as well as IL-4, reduces apoptotic events in chondrocytes and production of MMP [86, 87].

IL-10 induces the expression of bone morphogenetic protein-2 and -6 (BMP2 and BMP6), which are related to chondrogenesis as they belong to TGF- β family [88]. Together with BMP production, IL-10 activates signaling pathways, such as NKX-3.2/SOX9, that induce the differentiation of mesenchymal stem cells into chondrocytes [89]. Also, by reducing the expression of TNF- α receptors, IL-10 is able to attenuate the effect of TNF- α on synovial fibroblasts. A decrease in COX-2 production was also noted in the same study [90].

The secretion of IL-10 can be influenced by physical exercises. Patients with and without OA had synovial fluid and periarticular tissue harvested from their knee before, during, and after they underwent exercise practice for 3 hours. A significant increase in IL-10 levels was observed in these patients after the exercise. Although it is not clear what exact mechanism led to this result, this observation is

likely attributed to an increase in intra-articular pressure and subsequent effects on cellular secretion [91, 92].

3.2 Chemokines

Chemokines comprise small proteins that act as chemoattractants to assist cells to migrate to injured tissue. Diverse chemokines have gained attention in the development of OA. Some of them including their receptors, such as IL-8, CCL5, CCL19, CCR1, CCR2, CCR3, and CCR5, may induce the production of MMP-3 by chondrocytes and increase the breakdown of cartilage matrix components, which trigger the onset of OA [60, 93]. However, some chemokines might present a protective role in OA, such as stromal cell-derived factor-1 (also called CXCL12), whose main function is to recruit mesenchymal stem cells to the injured area in order to promote tissue repair [94].

Several chemokines were reported to be overexpressed in OA, such as IL-8/CXCL-8, GRO α /CXCL-1, MCP-1/CCL-2, RANTES/CCL-5, MIP-1 α /CCL-3, and MIP-1 β /CCL-4. Some of these chemokines are stimulated by IL-1 β , which is upregulated in OA, and they induce MMP production upon binding to their ligands, causing tissue degradation [93]. Levels of INF- γ -inducible protein 10 (IP-10), also called as CXCL-10, in plasma and synovial fluid have been correlated with radiographic knee OA. CX3CL1, a serum fractalkine, has also been reported to be significantly elevated in severe knee OA in a study that compared OA patients with healthy patients [95].

To support the role of macrophage in the inflammatory response observed in OA, MCP-1, also known as chemokine ligand-2 (CCL2), has been reported to recruit macrophages into adipose tissue and atherosclerotic lesions [96]. Also, MCP-1 levels in both serum and synovial fluid has been associated with self-reported pain and disability in patients who present knee OA [97]. In addition, it was observed that, in severe knee OA, the levels of macrophage-derived chemokine (MDC) and IP-10 in synovial fluid were elevated, while eotaxin levels, an eosinophil chemotactic protein, were lower when compared with healthy patients [98].

3.3 Adipokines

Adipokines have been associated with the incidence and severity of OA [99]. *In vitro* studies reported that the presence of adipokines, such as leptin, adiponectin, visfatin, and resistin, increases the production of inflammatory mediators and also induces chondrolysis [99]. Although the exact mechanism of how these cytokines derived from adipose tissue act on arthritic joints has not yet been elucidated, researchers have studied the role of fat pad as a local inflammation mediator in OA, particularly in knee OA due to the infrapatellar fat pad, which has proven to be infiltrated with macrophages, lymphocytes, and granulocytes [100]. These findings support the thought that obesity supports the development of OA more through biochemical pathways rather than biomechanical overload risks on a weight-bearing joint.

3.4 Lipid mediators

The COX-2 enzyme is responsible for the production of lipid mediators, including PGE₂ and leukotrienes, and it is also upregulated in OA joints. In addition, the overexpression of COX-2 in OA has been associated with the increased production of IL-1 β , TNF, and IL-6 via toll-like receptor-4 (TLR-4) [101]. Besides assisting the production of MMPs and other functions already cited above, PGE₂ is also involved in apoptosis and structural changes that characterize arthritic disease [102].

Leukotrienes have also been investigated for their role in OA. These mediators are converted from arachidonic acid, which also produces PGE₂ via the activity of the enzyme phospholipase A2 [21]. Leukotrienes, mainly leukotriene B4 (LTB₄), are present, to a lesser extent, in OA synovium, bone, and cartilage. Also, LTB₄ has been reported to stimulate the production of IL-1 β and TNF in arthritic synovium [103].

4. Conclusions

The cumulative evidences over the years have shown that increased expression of proinflammatory cytokines, in particular IL-1 β , TNF, and IL-6, in cartilage as well as synovial fluid and membrane, has played a key role in the pathogenesis of OA. Inflammatory processes linked with immune responses have characterized OA as a complex disease and not as a simple age-related cartilage degeneration as it is thought to be. The understanding of individual roles of inflammatory mediators and their compounds is of utmost importance to target new therapies for OA, since the current options are elusive and may be noneffective, invasive, or even capable of presenting serious side effects. Due to advancements in molecular tools, the overall aim would be to dissect the role of each cytokine in the pathophysiology of OA and, together with drug delivery systems, to develop specific anticytokine therapy, given that inflammatory responses contribute substantially to OA maintenance.

Conflict of interest

The authors have no conflict of interest to declare.

Author details

José Fábio dos Santos Duarte Lana* and Bruno Lima Rodrigues
Institute of Bone and Cartilage, Indaiatuba, Sao Paulo, Brazil

*Address all correspondence to: josefabiolana@gmail.com

IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

References

- [1] Robinson WH, Lepus CM, Wang Q, Raghu H, Mao R, Lindstrom TM, et al. Low-grade inflammation as a key mediator of the pathogenesis of osteoarthritis. *Nature Reviews Rheumatology*. 2016;**12**(10):580-592
- [2] Jonsson H, Olafsdottir S, Sigurdardottir S, Aspelund T, Eiriksdottir G, Sigurdsson S, et al. Incidence and prevalence of total joint replacements due to osteoarthritis in the elderly: Risk factors and factors associated with late life prevalence in the AGES-Reykjavik study. *BMC Musculoskeletal Disorders*. 2016;**17**(1):1-9
- [3] Helmick CG, Felson DT, Lawrence RC, Gabriel S, Hirsch R, Kwoh CK, et al. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part I. *Arthritis and Rheumatism*. 2008;**58**(1):15-25
- [4] Blagojevic M, Jinks C, Jeffery A, Jordan KP. Risk factors for onset of osteoarthritis of the knee in older adults: A systematic review and meta-analysis. *Osteoarthritis and Cartilage*. 2010;**18**(1):24-33
- [5] Sellam J, Berenbaum F. The role of synovitis in pathophysiology and clinical symptoms of osteoarthritis. *Nature Reviews Rheumatology*. 2010;**6**(11):625-635
- [6] Mapp PI, Walsh DA, Bowyer J, Maciewicz RA. Effects of a metalloproteinase inhibitor on osteochondral angiogenesis, chondropathy and pain behavior in a rat model of osteoarthritis. *Osteoarthritis and Cartilage*. 2010;**18**(4):593-600
- [7] Sharma L, Cooke TD, Guermazi A, Roemer FW, Nevitt MC. Valgus malalignment is a risk factor for lateral knee osteoarthritis incidence and progression: Findings from the multicenter osteoarthritis study and the osteoarthritis initiative. *Arthritis Rheumatism*. 2014;**65**(2):355-362
- [8] Kellgren JH, Lawrence JS. Radiological assessment of osteoarthrosis. *Annals of the Rheumatic Diseases*. 1957;**16**(4):494-502
- [9] Solomon DH, Husni ME, Libby PA, Yeomans ND, Lincoff AM, Lüscher TF, et al. The risk of major NSAID toxicity with celecoxib, ibuprofen, or naproxen: A secondary analysis of the PRECISION trial. *The American Journal of Medicine*. 2017;**130**(12):1415-1422.e4
- [10] Lee T, Lu N, Felson DT, Choi HK, Dalal DS, Zhang Y, et al. Use of non-steroidal anti-inflammatory drugs correlates with the risk of venous thromboembolism in knee osteoarthritis patients: a UK population-based case-control study. *Rheumatology*. 2016;**55**(6):1099-1105
- [11] Lana J, Weglein A, Sampson S, Vicente F, Huber S, Souza C, et al. Randomized controlled trial comparing hyaluronic acid, platelet-rich plasma and the combination of both in the treatment of mild and moderate osteoarthritis of the knee. *Journal of Stem Cells and Regenerative Medicine*. 2016;**12**(2):69-78
- [12] Dallari D, Stagni C, Rani N, Sabbioni G, Pelotti P, Torricelli P, et al. Ultrasound-guided injection of platelet-rich plasma and hyaluronic acid, separately and in combination, for hip osteoarthritis. *The American Journal of Sports Medicine*. 2016;**44**(3):664-671
- [13] Centeno CJ, Al-Sayegh H, Bashir J, Goodyear SH, D Freeman M. A prospective multi-site registry study of a specific protocol of autologous bone marrow concentrate for the treatment

of shoulder rotator cuff tears and osteoarthritis. *Journal of Pain Research*. 2015;**8**:269-276

[14] Jo C, Lee Y, Shin W, Kim H, Chai J, Jeong E, et al. Intra-Articular Injection of Mesenchymal Stem Cells for the Treatment of Osteoarthritis of the of the Knee: A Proof-of-Concept Clinical Trial CHRIS. *Stem cells*. 2014;**32**(5):1254-1266

[15] Zheng S, Hunter DJ, Xu J, Ding C. Monoclonal antibodies for the treatment of osteoarthritis. *Expert Opinion on Biological Therapy*. 2016;**16**(12):1529-1540

[16] Chiusaroli R, Visentini M, Galimberti C, Casseler C, Mennuni L, Covaceuszach S, et al. Targeting of ADAMTS5's ancillary domain with the recombinant mAb CRB0017 ameliorates disease progression in a spontaneous murine model of osteoarthritis. *Osteoarthritis and Cartilage [Internet]*. 2013;**21**(11):1807-1810. DOI: 10.1016/j.joca.2013.08.015

[17] Ma CH, Lv Q, Yu YX, Zhang Y, Kong D, Niu KR, et al. Protective effects of tumor necrosis factor—A blockade by adalimumab on articular cartilage and subchondral bone in a rat model of osteoarthritis. *Brazilian Journal of Medical and Biological Research*. 2015;**48**:863-870

[18] Cohen SB, Proudman S, Kivitz AJ, Burch FX, Donohue JP, Burstein D, et al. A randomized, double-blind study of AMG 108 (a fully human monoclonal antibody to IL-1R1) in patients with osteoarthritis of the knee. *Arthritis Research & Therapy [Internet]*. 2011;**13**(4):R125. Available from: <http://arthritis-research.biomedcentral.com/articles/10.1186/ar3430>

[19] Loeser RF, Goldring SR, Scanzello CR, Goldring MB. Osteoarthritis: A disease of the joint as an organ. *Arthritis and Rheumatism*. 2013;**64**(6):1697-1707

[20] Haseeb A, Haqqi TM. Immunopathogenesis of osteoarthritis. *Clinical Immunology*. 2009;**6**(3):247-253

[21] Sokolove J, Lepus CM. Role of inflammation in the pathogenesis of osteoarthritis: Latest findings and interpretations. *Therapeutic Advances in Musculoskeletal Disease*. 2013;**5**(2):77-94

[22] Orlowsky EW, Kraus VB. The role of innate immunity in osteoarthritis: When our first line of defense goes on the offensive. *The Journal of Rheumatology*. 2015;**42**(3):363-371

[23] Kandahari AM, Yang X, Dighe AS, Pan D, Cui Q. Recognition of immune response for the early diagnosis and treatment of osteoarthritis. *Journal of Immunology Research*. 2015;**2015**(192415):1-13

[24] Pessler F, Dai L, Diaz-Torne C, Gomez-Vaquero C, Paessler ME, Zheng DH, et al. The synovitis of “non-inflammatory” orthopaedic arthropathies: A quantitative histological and immunohistochemical analysis. *Annals of the Rheumatic Diseases*. 2008;**67**(8):1184-1187

[25] Bondeson J, Wainwright SD, Lauder S, Amos N, Hughes CE. The role of synovial macrophages and macrophage-produced cytokines in driving aggrecanases, matrix metalloproteinases, and other destructive and inflammatory responses in osteoarthritis. *Arthritis Research & Therapy*. 2006;**8**(6):1-12

[26] Van Lent PLEM, Blom AB, Van Der Kraan P, Holthuysen AEM, Vitters E, Van Rooijen N, et al. Crucial role of synovial lining macrophages in the promotion of transforming growth factor?—Mediated osteophyte formation. *Arthritis and Rheumatism*. 2004;**50**(1):103-111

[27] Huss RS, Huddleston JI, Goodman SB, Butcher EC, Zabel BA. Synovial tissue-infiltrating natural killer cells

in osteoarthritis and peri-prosthetic inflammation. *Arthritis and Rheumatism*. 2011;**62**(12):3799-3805

[28] Xiaoqiang E, Yang C, Hongxue M, Yuebin Q, Guangye D, Jun X, Bi Z. Dendritic cells of synovium in experimental model of osteoarthritis of rabbits. *Cellular Physiology and Biochemistry*. 2012;**30**(1):23-32

[29] Dalbeth N, Callan MFC. A subset of natural killer cells is greatly expanded within inflamed joints. *Arthritis and Rheumatism*. 2002;**46**(7):1763-1772

[30] Sower LE, Klimpel GR, Hanna W, Froelich CJ. Extracellular activities of human granzymes. *Cellular Immunology* [Internet]. 1996;**171**(1):159-163. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/8660852>

[31] Froelich CJ, Dixit VM, Yang X. Lymphocyte granule-mediated apoptosis: Matters of viral mimicry and deadly proteases. *Immunology Today*. 1998;**19**(1):30-36

[32] Tak PP, Spaeny-Dekking L, Kraan MC, Breedveld FC, Froelich CJ, Hack CE. The levels of soluble granzyme A and B are elevated in plasma and synovial fluid of patients with rheumatoid arthritis (RA). *Clinical and Experimental Immunology*. 1999;**116**:366-370

[33] Fox SW, Chambers TJ. Interferon- γ directly inhibits TRANCE-induced osteoclastogenesis. *Biochemical and Biophysical Research Communications*. 2000;**276**(3):868-872

[34] Madyastha PR, Yang S, Ries WL, Key LL. IFN- γ enhances osteoclast generation in cultures of peripheral blood from Osteopetrotic patients and normalizes superoxide production. *Journal of Interferon & Cytokine Research* [Internet]. 2000;**20**(7):645-652. Available from:

<http://www.liebertonline.com/doi/abs/10.1089/1079990000414826>

[35] Vermeire K, Heremans H, Vandeputte M, Huang S, Billiau A, Matthysz P. Accelerated collagen-induced arthritis in IFN- γ receptor-deficient mice. *Journal of Immunology*. 1997 Jun 1;**158**(11):5507-5513

[36] Wang Q, Rozelle AL, Lepus CM, Scanzello CR, Song JJ, Larsen M, et al. Identification of a central role for complement in osteoarthritis. *Nature Medicine*. 2012;**17**(12):1674-1679

[37] Ishii H, Tanaka H, Katoh K, Nakamura H, Nagashima M, Yoshino S. Characterization of infiltrating T cells and Th1/Th2-type cytokines in the synovium of patients with osteoarthritis. *Osteoarthritis and Cartilage*. 2002;**10**(4):277-281

[38] Sakkas LI, Scanzello C, Johanson N, Burkholder J, Mitra A, Salgame P, et al. T cells and T-cell cytokine transcripts in the synovial membrane in patients with osteoarthritis. *Clinical and Diagnostic Laboratory Immunology*. 1998;**5**(4):430-437

[39] Kapoor M, Martel-Pelletier J, Lajeunesse D, Pelletier JP, Fahmi H. Role of proinflammatory cytokines in the pathophysiology of osteoarthritis. *Nature Reviews Rheumatology*. 2011;**7**(1):33-42

[40] Saklatvala J. Tumour necrosis factor α stimulates resorption and inhibits synthesis of proteoglycan in cartilage. *Nature*. 1986 Aug 7-13;**322**(6079):547-549

[41] Chadjichristos C, Ghayor C, Kypriotou M, Martin G, Renard E, Ala-Kokko L, et al. Sp1 and Sp3 transcription factors mediate interleukin-1 β down-regulation of human type II collagen gene expression in articular chondrocytes. *The Journal of Biological Chemistry*. 2003;**278**(41):39762-39772

- [42] Shlopov BV, Gumanovskaya ML, Hasty KA. Autocrine regulation of collagenase 3 (matrix metalloproteinase 13) during osteoarthritis. *Arthritis and Rheumatism*. 2000;**43**(1):195-205
- [43] Fan Z, Bau B, Yang H, Soeder S, Aigner T. Freshly isolated osteoarthritic chondrocytes are catabolically more active than normal chondrocytes, but less responsive to catabolic stimulation with interleukin-1? *Arthritis and Rheumatism*. 2005;**52**(1):136-143
- [44] Cortial D, Gouttenoire J, Rousseau CF, Ronzière MC, Piccardi N, Msika P, et al. Activation by IL-1 of bovine articular chondrocytes in culture within a 3D collagen-based scaffold. An in vitro model to address the effect of compounds with therapeutic potential in osteoarthritis. *Osteoarthritis and Cartilage*. 2006;**14**(7):631-640
- [45] Silvestri T, Pulsatelli L, Dolzani P, Frizziero L, Facchini A, Meliconi R. In vivo expression of inflammatory cytokine receptors in the joint compartments of patients with arthritis. *Rheumatology International*. 2006;**26**(4):360-368
- [46] Roman-Blas JA, Jimenez SA. NF- κ B as a potential therapeutic target in osteoarthritis and rheumatoid arthritis. *Osteoarthritis and Cartilage*. 2006;**14**(9):839-848
- [47] Kobayashi T, Notoya K, Naito T, Unno S, Nakamura A, Martel-Pelletier J, et al. Pioglitazone, a peroxisome proliferator-activated receptor γ agonist, reduces the progression of experimental osteoarthritis in Guinea pigs. *Arthritis and Rheumatism*. 2005;**52**(2):479-487
- [48] Boileau C, Martel-Pelletier J, Fahmi H, Mineau F, Boily M, Pelletier JP. The peroxisome proliferator-activated receptor γ agonist pioglitazone reduces the development of cartilage lesions in an experimental dog model of osteoarthritis: In vivo protective effects mediated through the inhibition of key signaling and catabolic pathways. *Arthritis and Rheumatism*. 2007;**56**(7):2288-2298
- [49] Wang P, Zhu F, Konstantopoulos K. Prostaglandin E 2 induces interleukin-6 expression in human chondrocytes via cAMP/protein kinase A- and phosphatidylinositol 3-kinase-dependent NF- κ B activation. *American Journal of Physiology. Cell Physiology*. 2010;**298**:1445-1456
- [50] Stannus O, Jones G, Cicuttini F, Parameswaran V, Quinn S, Burgess J, et al. Circulating levels of IL-6 and TNF- α are associated with knee radiographic osteoarthritis and knee cartilage loss in older adults. *Osteoarthritis and Cartilage*. 2010;**18**(11):1441-1447
- [51] Sui Y, Lee JH, DiMicco MA, Vanderploeg EJ, Blake SM, Hung HH, et al. Mechanical injury potentiates proteoglycan catabolism induced by interleukin-6 with soluble interleukin-6 receptor and tumor necrosis factor α in immature bovine and adult human articular cartilage. *Arthritis and Rheumatism*. 2009;**60**(10):2985-2996
- [52] Rowan AD, Koshy PJT, Shingleton WD, Degnan BA, Heath JK, Vernallis AB, et al. Synergistic effects of glycoprotein 130 binding cytokines in combination with interleukin-1 on cartilage collagen breakdown. *Arthritis and Rheumatism*. 2001;**44**(7):1620-1632
- [53] de Hoo ASK, van de Loo FAJ, Bennink MB, Arntz OJ, de Hoo P, van den Berg WB. Male IL-6 gene knockout mice developed more advanced osteoarthritis upon aging. *Osteoarthritis and Cartilage*. 2005;**13**(1):66-73
- [54] van de Loo FA, Kuiper S, van Enkevort FH, Arntz OJ, van den Berg WB. Interleukin-6 reduces cartilage destruction during experimental

- arthritis. A study in interleukin-6-deficient mice. *The American Journal of Pathology*. 1997;**151**(1):177-191
- [55] Steeve KT, Marc P, Sandrine T, Dominique H, Yannick F. IL-6, RANKL, TNF- α /IL-1: Interrelations in bone resorption pathophysiology. *Cytokine & Growth Factor Reviews*. 2004;**15**(1):49-60
- [56] Massicotte F, Lajeunesse D, Benderdour M, Pelletier JP, Hilal G, Duval N, et al. Can altered production of interleukin-1 β , interleukin-6, transforming growth factor- β and prostaglandin E2 by isolated human subchondral osteoblasts identify two subgroups of osteoarthritic patients. *Osteoarthritis and Cartilage*. 2002;**10**(6):491-500
- [57] Sakao K, Takahashi KA, Arai Y, Saito M, Honjo K, Hiraoka N, et al. Osteoblasts derived from osteophytes produce interleukin-6, interleukin-8, and matrix metalloproteinase-13 in osteoarthritis. *Journal of Bone and Mineral Metabolism*. 2009;**27**(4):412-423
- [58] Baslund B, Tvede N, Danneskiold-Samsøe B, Larsson P, Panayi G, Petersen J, et al. Targeting interleukin-15 in patients with rheumatoid arthritis: A proof-of-concept study. *Arthritis and Rheumatism*. 2005;**52**(9):2686-2692
- [59] Waldmann TA, Tagaya Y. The multifaceted regulation of Interleukin-15 expression and the role of this cytokine in Nk cell differentiation and host response to intracellular pathogens. *Annual Review of Immunology*. 1999;**17**(1):19-49
- [60] Scanzello CR, Goldring SR. The role of synovitis in osteoarthritis pathogenesis. *Bone*. 2013;**51**(2):249-257
- [61] Sun J-M, Sun L-Z, Liu J, Su B, Shi L. Serum Interleukin-15 levels are associated with severity of pain in patients with knee osteoarthritis. *Disease Markers*. 2013;**35**(3):203-206
- [62] Wojdasiewicz P, Poniatowski ŁA, Szukiewicz D. The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis. *Mediators of Inflammation*. 2014;**2014**:561459
- [63] Noordenbos T, Yeremenko N, Gofita I, Van De Sande M, Tak PP, Cañete JD, et al. Interleukin-17-positive mast cells contribute to synovial inflammation in spondylarthritis. *Arthritis and Rheumatism*. 2012;**64**(1):99-109
- [64] Honorati MC, Neri S, Cattini L, Facchini A. Interleukin-17, a regulator of angiogenic factor release by synovial fibroblasts. *Osteoarthritis and Cartilage*. 2006;**14**(4):345-352
- [65] Lubberts E, Joosten LAB, Van De Loo FAJ, Van Den Berselaar LAM, Van Den Berg WB. Reduction of interleukin-17-induced inhibition of chondrocyte proteoglycan synthesis in intact murine articular cartilage by interleukin-4. *Arthritis and Rheumatism*. 2000;**43**(6):1300-1306
- [66] Chen B, Deng Y, Tan Y, Qin J, Bin CL. Association between severity of knee osteoarthritis and serum and synovial fluid interleukin 17 concentrations. *The Journal of International Medical Research*. 2014;**42**(1):138-144
- [67] Han L, Lee HS, Yoon JH, Choi WS, Park YG, Nam SW, et al. Association of IL-17A and IL-17F single nucleotide polymorphisms with susceptibility to osteoarthritis in a Korean population. *Gene*. 2014;**533**(1):119-122
- [68] Honorati MC, Cattini L, Facchini A. VEGF production by osteoarthritic chondrocytes cultured in micromass and stimulated by IL-17 and TNF- α . *Connective Tissue Research*. 2007;**48**(5):239-245

- [69] Olee T, Hashimoto S, Quach J, Lotz M. IL-18 is produced by articular chondrocytes and induces proinflammatory and catabolic responses. *Journal of Immunology*. 1999;**162**(2):1096-1100
- [70] Dai SM, Shan ZZ, Nishioka K, Yudoh K. Implication of interleukin 18 in production of matrix metalloproteinases in articular chondrocytes in arthritis: Direct effect on chondrocytes may not be pivotal. *Annals of the Rheumatic Diseases*. 2005;**64**(5):735-742
- [71] Joosten LAB, Smeets RL, Koenders MI, Van Den Bersselaar LAM, Helsen MMA, Oppers-Walgreen B, et al. Interleukin-18 promotes joint inflammation and induces interleukin-1-driven cartilage destruction. *The American Journal of Pathology*. 2004;**165**(3):959-967
- [72] John T, Kohl B, Mobasheri a EW, Shakibaei M. Interleukin-18 induces apoptosis in human articular chondrocytes. *Histology and Histopathology*. 2007;**22**(5):469-482
- [73] Wang Y, Xu D, Long L, Deng X, Tao R, Huang G. Correlation between plasma, synovial fluid and articular cartilage Interleukin-18 with radiographic severity in 33 patients with osteoarthritis of the knee. *Clinical and Experimental Medicine*. 2014;**14**(3):297-304
- [74] Hulin-Curtis SL, Bidwell JL, Perry MJ. Evaluation of IL18 and IL18R1 polymorphisms: Genetic susceptibility to knee osteoarthritis. *International Journal of Immunogenetics*. 2012;**39**(2):106-109
- [75] Omair A. An association study of interleukin 18 receptor genes (IL18R1 and IL18RAP) in lumbar disc degeneration. *Open Orthopaedics Journal*. 2012;**6**(1):164-171
- [76] Van Lent PLEM, Holthuysen AEM, Slöetjes A, Lubberts E, Van Den Berg WB. Local overexpression of adenoviral IL-4 protects cartilage from metalloproteinase-induced destruction during immune complex-mediated arthritis by preventing activation of pro-MMPs. *Osteoarthritis and Cartilage*. 2002;**10**(3):234-243
- [77] Millward-Sadler SJ, Wright MO, Davies LW, Nuki G, Salter DM. Mechanotransduction via integrins and interleukin-4 results in altered aggrecan and matrix metalloproteinase 3 gene expression in normal, but not osteoarthritic, human articular chondrocytes. *Arthritis and Rheumatism*. 2000;**43**(9):2091-2099
- [78] Vargiolu M, Silvestri T, Bonora E, Dolzani P, Pulsatelli L, Addimanda O, et al. Interleukin-4/interleukin-4 receptor gene polymorphisms in hand osteoarthritis. *Osteoarthritis and Cartilage*. 2010;**18**(6):810-816
- [79] Yigit S, Inanir A, Tekcan A, Tural E, Ozturk GT, Kismali G, et al. Significant association of interleukin-4 gene intron 3 VNTR polymorphism with susceptibility to knee osteoarthritis. *Gene*. 2014;**537**(1):6-9
- [80] Silvestri T, Pulsatelli L, Dolzani P, Facchini A, Meliconi R. Elevated serum levels of soluble interleukin-4 receptor in osteoarthritis. *Osteoarthritis and Cartilage*. 2006;**14**(7):717-719
- [81] Bhattacharjee A, Shukla M, Yakubenko VP, Mulya A, Kundu S, Cathcart MK. IL-4 and IL-13 employ discrete signaling pathways for target gene expression in alternatively activated monocytes/macrophages. *Free Radical Biology & Medicine*. 2013;**54**(216):1-16
- [82] Van Meegeren MER, Roosendaal G, Jansen NWD, Wenting MJG, Van Wesel ACW, Van Roon JAG, et al. IL-4 alone and in combination with IL-10

protects against blood-induced cartilage damage. *Osteoarthritis and Cartilage*. 2012;**20**(7):764-772

[83] Schuerwegh AJ, Dombrecht EJ, Stevens WJ, Van Offel JF, Bridts CH, De Clerck LS. Influence of pro-inflammatory (IL-1 α , IL-6, TNF- α , IFN- γ) and anti-inflammatory (IL-4) cytokines on chondrocyte function. *Osteoarthritis and Cartilage*. 2003;**11**(9):681-687

[84] Yorimitsu M, Nishida K, Shimizu A, Doi H, Miyazawa S, Komiyama T, et al. Intra-articular injection of interleukin-4 decreases nitric oxide production by chondrocytes and ameliorates subsequent destruction of cartilage in instability-induced osteoarthritis in rat knee joints. *Osteoarthritis and Cartilage*. 2008;**16**(7):764-771

[85] Lacraz S, Nicod LP, Chicheportiche R, Welgus HG, Dayer JM. IL-10 inhibits metalloproteinase and stimulates TIMP-1 production in human mononuclear phagocytes. *The Journal of Clinical Investigation*. 1995;**96**(5):2304-2310

[86] Wang Y, Lou S. Direct protective effect of interleukin-10 on articular chondrocytes in vitro. *Chinese Medical Journal*. 2001;**114**(7):723-725

[87] John T, Müller RD, Oberholzer A, Zreiqat H, Kohl B, Ertel W, et al. Interleukin-10 modulates pro-apoptotic effects of TNF- α in human articular chondrocytes in vitro. *Cytokine*. 2007;**40**(3):226-234

[88] Umulis D, O'Connor MB, Blair SS. The extracellular regulation of bone morphogenetic protein signaling. *Development*. 2009;**136**(22):3715-3728

[89] Zeng L, Kempf H, Murtaugh LC, Sato ME, Lassar AB. Shh establishes an Nkx3.2/Sox9 autoregulatory loop that is maintained by BMP signals to induce

somitic chondrogenesis. *Genes & Development*. 2002;**16**(15):1990-2005

[90] Alaaeddine N, Di Battista JA, Pelletier JP, Kiansa K, Cloutier JM, Martel-Pelletier J. Inhibition of tumor necrosis factor alpha-induced prostaglandin E2 production by the anti-inflammatory cytokines interleukin-4, interleukin-10, and interleukin-13 in osteoarthritic synovial fibroblasts: Distinct targeting in the signaling pathways. *Arthritis and Rheumatism*. 1999;**42**(4):710-718

[91] Helmark IC, Mikkelsen UR, Børglum J, Rothe A, Petersen MC, Andersen O, et al. Exercise increases interleukin-10 levels both intraarticularly and peri-synovially in patients with knee osteoarthritis: A randomized controlled trial. *Arthritis Research & Therapy*. 2010;**12**(4):R126

[92] Angele P, Yoo JU, Smith C, Mansour J, Jepsen KJ, Nerlich M, et al. Cyclic hydrostatic pressure enhances the chondrogenic phenotype of human mesenchymal progenitor cells differentiated in vitro. *Journal of Orthopaedic Research*. 2003;**21**(3):451-457

[93] Borzì RM, Mazzetti I, Cattini L, Ugucioni M, Baggiolini M, Facchini A. Human chondrocytes express functional chemokine receptors and release matrix-degrading enzymes in response to C-X-C and C-C chemokines. *Arthritis and Rheumatism*. 2000;**43**(8):1734-1741

[94] Miller RJ, Banisadr G, Bhattacharyya BJ. CXCR4 signaling in the regulation of stem cell migration and development. *Journal of Neuroimmunology*. 2008;**198**(1-2):31-38

[95] Zou Y, Li Y, Lu L, Lin Y, Liang W, Su Z, et al. Correlation of fractalkine concentrations in serum and synovial fluid with the radiographic severity of

knee osteoarthritis. *Annals of Clinical Biochemistry*. 2013;**50**(6):571-575

Seminars in Arthritis and Rheumatism. 2003;**33**(3):155-167

[96] Kanda H, Tateya S, Tamori Y, Kotani K, Hiasa K, Kitazawa R, et al. MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *The Journal of Clinical Investigation*. 2006;**116**(6):1494-1505

[103] Oliveira SHP, Canetti C, Ribeiro RA, Cunha FQ. Neutrophil migration induced by IL-1 β depends upon LTB₄ released by macrophages and upon TNF- α and IL-1 β released by mast cells. *Inflammation*. 2008;**31**(1):36-46

[97] Li L, Jiang BE. Serum and synovial fluid chemokine ligand 2/monocyte chemoattractant protein 1 concentrations correlates with symptomatic severity in patients with knee osteoarthritis. *Annals of Clinical Biochemistry*. 2015;**52**(2):276-282

[98] Beekhuizen M, Gierman LM, van Spil WE, Van Osch GJVM, Huizinga TWJ, Saris DBF, et al. An explorative study comparing levels of soluble mediators in control and osteoarthritic synovial fluid. *Osteoarthritis and Cartilage*. 2013;**21**(7):918-922

[99] Conde J, Scotece M, Gómez R, Lopez V, Gómez-Reino JJ, Gualillo O. Adipokines and osteoarthritis: Novel molecules involved in the pathogenesis and progression of disease. *Arthritis*. 2011;**2011**:1-8

[100] Clockaerts S, Bastiaansen-Jenniskens YM, Runhaar J, Van Osch GJVM, Van Offel JF, Verhaar JAN, et al. The infrapatellar fat pad should be considered as an active osteoarthritic joint tissue: A narrative review. *Osteoarthritis and Cartilage*. 2010;**18**(7):876-882

[101] Geng Y, Blanco FJ, Cornelisson M, Lotz M. Regulation of cyclooxygenase-2 expression in normal human articular chondrocytes. *Journal of Immunology*. 1995;**155**(2):796-801

[102] Martel-Pelletier J, Pelletier JP, Fahmi H. Cyclooxygenase-2 and prostaglandins in articular tissues.