

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Role of Vegetables in Human Nutrition and Disease Prevention

Taha Gökmen Ülger, Ayşe Nur Songur,
Onur Çırak and Funda Pınar Çakıroğlu

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/intechopen.77038>

Abstract

Vegetables are important for human health because of their vitamins, minerals, phytochemical compounds, and dietary fiber content. Especially antioxidant vitamins (vitamin A, vitamin C, and vitamin E) and dietary fiber content have important roles in human health. Adequate vegetable consumption can be protective some chronic diseases such as diabetes, cancer, obesity, metabolic syndrome, cardiovascular diseases, as well as improve risk factors related with these diseases. In this chapter, basic information will be given about the classification of vegetables, preparation and cooking, and their effects on food content of vegetables and effects on health and diseases (diabetes, obesity, metabolic syndrome, cardiovascular diseases, and cancer).

Keywords: vegetables, diabetes, metabolic syndrome, cardiovascular diseases, cancer, cooking methods, phenolic compounds, antioxidants, fiber

1. Introduction

Vegetables are annual or perennial horticultural crops, with certain sections (roots, stalks, flowers, fruits, leaves, etc.) that can be consumed wholly or partially, cooked or raw [1].

Vegetables are important for human nutrition in terms of bioactive nutrient molecules such as dietary fiber, vitamins and minerals, and non-nutritive phytochemicals (phenolic compounds, flavonoids, bioactive peptides, etc.). These nutrient and non-nutrient molecules reduce the risk of chronic diseases such as cardiovascular diseases, diabetes, certain cancers, and obesity [2, 3].

In recent years, consumers began to change their eating patterns with the growing interest in the effect of foods in staying healthy and maintaining health. “Western” type diets are characterized by increased intake of calories, sugar, saturated fats and animal protein, and reduced consumption of vegetables and fruits. When this type of diet is combined with lack of activity, the prevalence and frequency of diseases such as obesity, diabetes, and cardiovascular pathologies also increases [3]. In healthy diets (Mediterranean diet model), eating plant-based foods such as fruits and vegetables, cereals, legumes and nuts, replacing butter with healthy oils such as olive oil and canola oil, using herbs and spices to add flavor instead of salt, limiting red meat to several times a month and eating fish and poultry at least twice a week are recommended. Evidence from epidemiological studies and clinical trials shows that the Mediterranean diet is associated with many positive health outcomes such as reduced risk of various chronic illnesses, reduced overall mortality, and increased likelihood of healthy aging[4]. One of the most important features of these diets is the high consumption of vegetables, and therefore fiber, vitamins, minerals, flavonoids, phytoestrogens, sulfur compounds, phenolic compounds such as monoterpenes and bioactive peptides, which have positive effects on health [3]. In this chapter, basic information will be presented on the classification of vegetables, their relation to health, and the effects of preparation and cooking on nutrient content of vegetables.

2. Classification of vegetables

There are approximately 10,000 plant species used as vegetables in the world. Classification of these species can be done by considering a common set of features. It is important for food researchers, dietitians, and nutrition educators to subcategorize vegetables by taking into account health and nutrition. This sub-categorization will be more useful if it is based on similarities in food composition [5]. Vegetables can be classified according to the part of the plant used for nutrition and the specific nutritional value [6].

2.1. Green vegetables

2.1.1. Leaf vegetables

This group includes spinach, lettuce, curly lettuce, chard, purslane, chicory, etc. These are important minerals (iron and calcium), vitamins (A, C, and riboflavin) and fiber sources.

Young, fresh leaves contain more vitamin C than mature plants. The green outer leaves of lettuce and cabbage are richer in vitamins, calcium, and iron than white inner leaves. Thinner and greener leaves are more nutritious and usually have lower calories.

2.1.2. Stalk vegetables

The best examples to be given to stalk vegetables are celery and asparagus. They contain minerals and vitamins in proportion to the green color. Asparagus is a particularly rich source of folic acid.

2.1.3. Fruit and flower vegetables

Broccoli, cauliflower, and artichoke are frequently consumed flowering vegetables. Broccoli is a good source of iron, phosphorus, vitamins A and C, and riboflavin. Cauliflower is also a good source of vitamin C. The nutritional value of the outer leaves of cauliflower and broccoli is much higher than the flower buds. They can be consumed raw in salads or cooked. Artichoke is a good source of minerals, especially potassium, calcium, and phosphorus, and has high dietary fiber content. Tomatoes and peppers are the most common fruit vegetables. Both are rich in vitamin C. Other fruit vegetables include cucumber, zucchini, and eggplant. A dark green or yellow color indicates high β -carotene content. The darker the yellow color, the higher the content of β -carotene.

2.2. Root vegetables

2.2.1. Root, bulb, and tuber vegetables

Carrot, beet, turnip, fennel, onion, radish, and potato are examples of this group of vegetables. Yellow and orange varieties are rich in β -carotene, which is the precursor of vitamin A. Onion is an extraordinary example of root vegetables and contains moderate levels of vitamin C.

2.2.2. Legumes

This group includes legumes, peas, and soya beans. This group is rich in saponin and soluble fiber [6].

Subgroups may differ from country to country and classifications in nutritional guidelines are based on nutritional content in different countries. For example, the basic food guidelines used in the United States (Basic 7 and Basic 4 Food Groups and Food Guide Pyramid) are focused on dark green leafy and dark orange/yellow group vegetables for beta-carotene and citrus fruits for vitamin C. Later on, 2010 USDA MyPyramid food guide identified dark green leafy vegetables and broccoli, other leafy vegetables, legumes, unique vegetables (dark orange, tomato, allium vegetables, etc.) and additional vegetables [Table 1]. In the guide prepared by Turkish Ministry of Health (Turkey Nutrition Guide 2015), vegetables have been classified as *Dark green leafy vegetables* (Mediterranean/salad greens such as spinach, chard, quince, blackcurrant, vine leaf, curly, lettuce, spinach, purslane, parsley, cress, arugula, mint, sorrel, radish, dill, radica, and curly-chicory (chopped or in salads)), *other green vegetables* (broccoli, okra, fresh beans, fresh peas, green zucchini, artichokes, asparagus, brussels sprouts, varieties of pointed or stuffed peppers, cucumber, and iceberg lettuce (chopped or in salads)), *Red—orange—blue—purple vegetables* (tomatoes, carrots, red pepper, radish, winter squash, beet, aubergine, and red cabbage), *white vegetables* (onion, celery, cabbage, cauliflower, leek, mushroom, ground apple, turnip), and *starchy vegetables* (potatoes and fresh corn) [7].

Vegetable subgroups	Important sources ^a	Contributes ^b
Dark green leafy vegetables and broccoli	Vitamin C	Iron, copper, manganese
	Vitamin K	Vitamin B6
	Folate	Phytosterol
	Beta-carotene	Alpha-carotene
	Lutein + zeaxanthin, flavones	Flavonols TAC
Other leafy vegetables	Vitamin C	Phytosterol
	Vitamin K	Manganese
	Anthocyanidins	Vitamin B6
		Folate
		Beta-carotene
		Lutein + zeaxanthin
		TAC
Legumes	Copper	Dietary fiber
	Folate	Magnesium, iron, zinc, manganese
	Phytosterol	Vitamin B6
	Flavan-3-ols	
	Flavonols	
	TAC	
Unique vegetables	Vitamin C	Vitamin B6
	Alpha-carotene lycopene	Vitamin K
		Manganese, copper
		Beta-carotene
Additional vegetables	Flavonoids	Vitamin C
		Vitamin K

^aProvides >25% DRI or highest mean concentration of component per 100 g.

^bProvides >10% DRI or second or third highest concentration of component per 100 g.

Table 1. Summary chart for food ingredients in 2010 my pyramid vegetable subgroups [5].

3. The effect of vegetables on some disease

3.1. Effects on diabetes, obesity, and metabolic syndrome

Diabetes mellitus (DM), obesity, and the metabolic syndrome (MS) are increasing health problems in recent years in parallel with the increase in unhealthy eating habits and unhealthy living behaviors. One of the most basic aspects of the control and management of the disease in individuals with these health problems is the regulation of eating habits. In medical nutrition therapy applied to these individuals, it is important to meet the energy and nutritional needs

of individuals, as well as including foods with functional activities against the complications of these diseases in the diet. Phytochemical compounds (carotenoids, alkaloids, terpenoids, and phenolics), which are secondary compounds found in vegetables, are thought to be protective against these diseases.

3.1.1. Root, bulb, and tuber vegetables

Onions and garlic, thanks to the volatile oils, organosulfur compounds, and flavonoids in their content, are among the vegetables thought to be protective against DM, obesity, and MS [8]. Organosulfur compounds such as S-methyl cysteine and flavonoids such as quercetin in these vegetables exert a functional effect by regulating the activities of some enzymes involved in carbohydrate metabolism, increasing insulin secretion and sensitivity, and increasing NADP⁺ and NADPH activities [9]. In addition, these vegetables inhibit the enzymes α -glucosidase and α -amylase, inhibiting the formation of D-glucose from oligosaccharides and disaccharides and delaying the absorption of glucose from the intestines [10]. Onion and garlic are especially protective against dyslipidemia and oxidative stress, which are seen due to DM and MS.

Kumar et al. found that obese patients with Type 2 diabetes who used garlic tablets in addition to metformin had significantly higher fasting blood glucose (FBG), postprandial blood glucose, total cholesterol (TC), triglyceride (TG), low density lipoprotein cholesterol (LDL), C-reactive protein (CRP), and adenosine deaminase levels compared to those of patients using only metformin [11]. In dyslipidemic individuals with Type 2 DM, the use of garlic tablets for 12 weeks significantly decreased TC and LDL levels, while high-density lipoprotein cholesterol (HDL) levels were significantly increased [12]. Although there are similar studies suggesting that garlic has positive effects on blood glucose level and plasma lipid profile in the presence of DM [13], garlic was also found to increase antioxidant enzyme activities in DM and reduce bioactive aldehyde levels [14].

It was found that garlic consumption increased adiponectin levels in MS patients [15]. Considering that adiponectin has antiatherogenic and antiatherosclerotic effects [16], garlic consumption in MS patients is thought to be protective against cardiovascular diseases (CVDs). In addition, it has been determined that garlic has a positive effect on insulin resistance in rats with MS induced by high fructose content feed [17].

In obese rats induced by high-fat diets, garlic supplemented animal feed reduced TG and TC levels, as well as body weight and epididymal fat accumulation [18]. Similarly, in obesity induced rats with a high-fat diet, garlic reduced visceral and epididymal fat accumulation while reducing atherogenic index and cardiac risk factors [19].

It was reported that onion powder added to animal feed in experimental diabetic rats induced by aloxane or streptozotocin had a hypoglycemic effect [9, 20]. In a study comparing the efficacy of glibenclamide, which is an oral antidiabetic drug, with onion application at different doses in DM rats, it was reported that 300 mg/kg of onion extract application reduced fasting glucose levels by 75.4%, whereas 2.5 mg/kg glibenclamide reduced fasting glucose levels by 76.4% [21].

Studies investigating the effects of onion consumption in the presence of DM on antioxidant enzyme levels such as superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GSH-Px) showed that onion consumption increased the levels of these enzymes [22]. In these studies, it was also found that onion reduced bioactive aldehyde levels such as malondialdehyde (MDA) formed by the decomposition of lipid hydroperoxides.

It was reported that onion reduced insulin resistance and improved FBG levels in MS Zucker type rats [23]. It was shown that onion extract reduced weight gain, epididymal fat accumulation, and serum TC levels in BALB/c mice that were made obese with a high-fat diet [24]. It was reported that a daily onion consumption of 100–120 g significantly decreased TC and LDL cholesterol levels of obese women with polycystic ovary syndrome [25].

3.1.2. Leaf vegetables

Purslane and chard are green leafy vegetables thought to have functional activity against DM, MS, and obesity. Purslane shows a functional effect due to free oxalic acids, alkaloids, omega-3 fatty acids, coumarins, flavonoids, cardiac glycosides, anthraquinone, α -linolenic acid, and active compounds in its composition [26], while chard shows its effect via phospholipids, glycolipids, fatty acids (palmitic, stearic, oleic, and linoleic acid), folic acid, ascorbic acid, and pectin in its composition [27]. Purslane, described by WHO as one of the most used medical plants, is also called “global panacea” [28].

In studies investigating the effects of the use of purslane extract on anthropometric and biochemical changes in Type 2 DM patients, it was found that consumption of purslane extract significantly reduced HbA1c levels [29], TG, TC, LDL, FBG, and post-prandial blood glucose, body weight and BMI, whereas it significantly increased HDL levels [30]. In another study, consumption of purslane extract significantly increased glucagon-like peptide-1 concentrations, which has positive effects on beta cell proliferation and insulin secretion, in individuals with Type 2 DM [31].

In rats fed high-fat diets, it was seen that purslane decreased TG, TC, and LDL levels [32] and similarly it decreased TC and TG levels in hypercholesterolemic rats [33]. In rats with DM induced by streptozotocin, it was shown that purslane had hypoglycemic [34] and antioxidant effects [35].

In Type 1 diabetic rats, chard extracts were shown to reduce blood glucose levels and improve beta cell regeneration [27], while significantly decreasing adenosine deaminase levels [36]. It was also found that chard decreased elevated MDA levels due to diabetes and increased antioxidant capacity [36]. In Type 2 diabetic rats, chard extract was also shown to be effective in increasing insulin secretion and lowering blood glucose levels by increasing GLP-1 and acetylcholine levels [37]. It was also found to have a hypolipidemic effect in high fat diet-induced rats [38].

3.1.3. Fruit and flower vegetables

Broccoli and cauliflower are vegetables thought to have protective effects against many diseases thanks to glucosinolates and indole-3-carbinol they contain [39]. Indole-3-carbinol given to obesity induced mice by a high fat diet was shown to reduce epididymal fat accumulation,

body weight, insulin, leptin, and blood glucose levels, increase adiponectin levels, and improve glucose tolerance [39]. Similarly, in Type 2 diabetic rats fed with a high fat diet, indole-3-carbinol reduced blood glucose levels, and HbA1c levels, thereby reducing thio-barbituric acid reactive substances, lipid hydroperoxides and conjugated dienes levels, and increased levels of SOD, CAT, and GSH-Px [40]. Positive effects of broccoli on impaired lipid profile due to high fat diets were detected [41].

3.1.4. Legumes

Leguminosae family peas and vegetables such as peas and soybeans inhibit alpha-amylase enzyme, and show antidyslipidemic and antioxidant effects thanks to phytosterols such as B-sitosterol, campesterol and stigmasterol, and linoleic acid they contain [42]. Studies conducted by Helmstädter revealed that different plant extracts of this family improved glucose tolerance and glycosuria [43]. Consumption of different species of this family such as Pinto beans, Great Northern beans, Navy beans, and Black beans have been reported to reduce the risk of obesity, MS, and DM [44].

3.2. Effects on cardiovascular diseases

CVDs are the primary cause of death and illness in the world. The Global Burden of Disease Study reported that 29.6% of all deaths in the world were due to CVDs [45]. The main factor in these deaths is the increase in unhealthy lifestyle and eating habits. Most of the risk factors associated with CVDs are reversible risk factors and non-pharmacologic measures such as healthy eating habits and healthy lifestyle changes may help control the risk factors for the disease. Increased consumption of vegetables, which are an important part of a healthy diet, has been shown to reduce CVD-related mortality rates [46] and improve risk factors [47]. Vegetables are protective against CVD thanks to low content of saturated fat, trans fat, and cholesterol and being rich in bioactive compounds such as flavonoids, phytoestrogens (lignans, coumestran, isoflavones, resveratrol, and lycopene), organosulfur compounds, soluble dietary fibers (β -glucan, pectin, and psyllium), isothiocyanates, monoterpenes, and sterols (sitostanol, stigmasterol, and campesterol) [48].

3.2.1. Root, bulb, and tuber vegetables

Epidemiological studies indicate that there is an inverse relationship between garlic consumption and CVD development. Studies in the literature reported that garlic and garlic components show cholesterol and lipid lowering effects by inhibiting key enzymes involved in cholesterol and fatty acid synthesis (monooxygenase and HMG-CoA reductase) [49], anti-platelet effect by inhibiting cyclooxygenase enzyme activity [50], and fibrinolytic effect by inhibiting lipid peroxidation and hemolysis in oxidized erythrocytes [51]. It was also reported that onion and garlic had a blood pressure lowering effect by inducing intracellular nitric oxide and hydrogen sulfide production and inhibiting angiotensin-converting enzyme activity [52]. It was also shown that garlic reduced the levels of reactive oxygen species (ROS) that are thought to play a role in the pathogenesis of CVD and increased antioxidant capacity [52].

Although epidemiological studies investigating the relationship of onion consumption and CVD risk and CVD-related mortality rates are limited, a study conducted in Finland found that CVD-induced mortality was lower in individuals with high onion consumption than in those with low onion consumption [53]. Similar to garlic, onion also improves cardiovascular health through the sulfurous compounds, and especially flavonoids such as quercetin in its content. By cutting an onion, *S*-alk(en)yl-L-cysteine sulfoxides are converted into thiosulfinates and copanenes via the enzyme alliinase and these compounds inhibit platelet aggregation [54]. Since platelet aggregation is an important risk factor for the development of coronary thrombosis and atherosclerosis, onion consumption may be beneficial in individuals with risk factors for CVD. In addition, it was reported that onion consumption in hypercholesterolemic rats reduces CVD risk by decreasing the elevated inflammatory biomarkers associated with high cholesterol diet and by increasing the levels of antioxidant enzymes [55]. Onion also eliminates risk factors by correcting the dyslipidemia seen in some chronic diseases such as DM [9].

3.2.2. Leaf vegetables

Green leafy vegetables increase antioxidant capacity through minerals, vitamins, pulp, and phytochemical compounds in their content and protect against oxidative stress which is thought to play an important role in the pathogenesis of CVD [56]. In traditional diets where consumption of green leafy vegetables is high (Mediterranean and Japanese traditional diets), the rate of CVD is lower, and average life span is longer [57]. Moreover, in vegetarian individuals, mortality rates due to ischemic heart diseases and cerebrovascular diseases were also found to be lower than in non-vegetarians [58]. In another study, it was found that paralysis rates were significantly lower in individuals with higher consumption of green leafy vegetables than in individuals with less consumption of green leafy vegetables [59]. The incidence of coronary artery disease was also reported to be lower in individuals with higher green leafy vegetables consumption [53]. Individuals with more than three portions of green leafy vegetable consumption a day were found to have an ischemic heart disease incidence of about 60% less than those consuming less than 1 portion per day [60]. Furthermore, green leafy vegetables such as rocket, spinach, and lettuce also reduce blood pressure, inhibit platelet aggregation and improve endothelial dysfunction due to their rich inorganic nitrate content [57]. Some studies suggest that high nitrate content in the vegetables in this group is transformed into nitrite, nitric oxide, and vasodilator-tissue protective secondary compounds through symbiotic bacteria in the oral cavity, thereby maintaining cardiopulmonary function by lowering blood pressure [61].

3.2.3. Fruit and flower vegetables

The vegetables in this group are rich in sulfur-containing glucosinolates, flavonoids, anthocyanins, coumarins, carotenoids, antioxidant enzymes, and terpenes [62]. However, indole-3-carbinol and sulforaphane, which is a hydrolysis product of glucoraphanin, are thought to be the main bioactive compounds that are protective against CVD [63]. In experimental animals, sulforaphane protects against ischemic damage to the heart through induction of

Nrf2-related phase-II enzymes such as SOD, CAT, and hemoxygenase-1 [64]. Indole-3-carbinol and sulforaphane also protect against inflammation by inhibiting cytokine production [63]. In some epidemiological studies, it has been argued that consumption of vegetables in this group may reduce CVD-related mortality rates [65, 66]. It has been reported that anthocyanins extracted from red cabbage have protective effect on blood platelets [67], while broccoli sprouts decrease TC and LDL levels and increase HDL levels [68].

3.2.4. Legumes

The vegetables in this group are protective against CVD due to their high saponin and soluble fiber content. Another reason why legumes are beneficial for heart health is their low sodium and high potassium, calcium, and magnesium content [69]. The soluble pulp reduces the levels of TC and LDL by inhibiting the absorption of bile acid from the intestines and enabling the formation of short chain fatty acids, particularly propionic acid, that inhibit the synthesis of cholesterol [70]. It also improves heart health by inhibiting platelet aggregation [71]. In long-term observational epidemiologic studies, increased legume consumption has been reported to reduce CVD-related mortality and may protect against these diseases [69, 72, 73].

3.3. Effects on cancer

Cancer occurs as cells grow and proliferative without control [74]. Cancer occurs, progresses, and spreads as a result of abnormal signals in the body due to genetic or epigenetic effects [75]. Cancer is among the main causes of death in the world. On average, 16% of deaths occur each year due to cancer [76]. Lifestyle and many genetic and environmental factors can cause cancer. Smoking, consumed foods, solar radiation, and carcinogens in the environment are among these factors. The most important step in the treatment of cancer is the prevention of cancer. In particular, it is important to use health-related preventive practices in the communities and individuals at risk [75].

Consuming plant-based foods, especially increasing the consumption of vegetables, reduces the risk of cancer [74]. The antioxidants in vegetables help reduce the risk of cancer by preventing oxidative damage to the cells in the body [77]. Vegetables have protective effects against cancer due to the vitamins, minerals, pulp, and phytochemicals they contain [78]. About 14% of deaths worldwide due to inadequate vegetable consumption are caused by gastrointestinal cancers [79].

In a meta-analysis, the effects of vegetable consumption on cancer incidence were examined. Fruit and vegetable consumption were found to decrease cancer risk independently of each other and it was found that an extra portion of vegetables consumed daily resulted in a 3% reduction in cancer incidence [80].

A study investigating the relationship between vegetable and fruit consumption and epithelial ovarian cancer included 500 cancer patients and 500 control subjects. Cancer patients were found to have significantly lower average amounts of vegetables and fruits consumed per day than the control group [81]. However, in a cohort study investigating the relationship between vegetable and fruit consumption and pancreatic cancer, no significant relationship was found [82].

3.3.1. *Root, bulb, and tuber vegetables*

Vegetables in this group exert their protective effect against cancer through inositol, flavonoids, lignans, polyphenols, protease inhibitors, saponins, steroids, triterpenoids, isoflavones, phenolic acids, protein kinase inhibitors, sphingolipids, allicin, aline, and allyl sulfides [78].

Onions prevent tumor formation and cancer cells from spreading in many kinds of cancers such as stomach, ovary, breast, and colon cancer [83]. It has been shown that onion extract has apoptosis-inducing effects in MDA-MB-231 cells that cause breast cancer [84].

Compounds such as thyoallyl found in garlic are effective in preventing cancer. Such compounds in garlic have antioxidant effects that prevent and reduce carcinogens in DNA. They are also effective in reducing free radicals, inducing apoptosis, and stimulating the immune system [85, 86]. In a meta-analysis, the relationship between all cancer types and garlic consumption was investigated and it was concluded that garlic consumption was protective against gastric and intestinal cancers [87].

A controlled study investigating the relationship between onion and garlic consumption and gastric cancer included 759 cancer patients and 750 control subjects. As a result of the study, both onion and garlic consumption were found to have a negative relationship with cancer [88].

Carrot, which is a good source of flavonoids, polyacetylenes, vitamins and minerals, and carotenes, is also effective in protecting against cancer. Carrots have antioxidant, anticarcinogenic, and immune system enhancing properties [89]. In a study, it was determined that carrot consumption was negatively related to prostate cancer [90]. In another study conducted on rats, carrot consumption was shown to have protective effects against cancer due to the high content of carotenoids found in carrots [91].

3.3.2. *Leaf vegetables*

Green leafy vegetables reduce the risk of cancer due to phytochemicals, vitamin C, vitamin E, vitamin K, and vitamin A they contain [92]. The phytochemicals in these vegetables strengthen the immune system, protect against carcinogenic substances, reduce inflammation and oxidative stress that causes cancer, reduce DNA damage, prevent the growth of cancer cells, inhibit angiogenesis that is effective in tumor growth and regulate hormones [78, 93]. It is thought that these effects are exerted especially in cancer types such as breast, skin, lung, and stomach [78]. The main phytochemicals believed to be protective against these cancer types are isothiocyanates [92]. Apart from these phytochemicals, green leafy vegetables are protective against cancer, especially gastrointestinal system carcinomas, due to high pulp content [93].

In this group, spinach shows protective effects against cancer by reducing oxidative stress in the body thanks to vitamins A, C, and E, carotenes such as beta carotene and lutein, flavones and flavonoids it contains [94, 95].

Broccoli is another vegetable that is effective in protecting against cancer. A number of epidemiological studies have associated broccoli to low incidence of cancer. Sulfurous compounds

found in broccoli are cancer preventive agents [96]. In addition to sulfurous compounds, there are carotenes and other antioxidant vitamins in broccoli. But the most effective compounds in preventing cancer are the sulfurous compounds in broccoli. These sulfurous compounds inhibit cancer formation by reducing free radicals and preventing cell damage [97].

3.3.3. *Fruit and flower vegetables*

Tomato, a good source of beta carotene and lycopene, reduces free radical damage in the DNA that causes cancer and prevents the growth and spread of cancer cells just like green leafy vegetables [78]. Lycopene is especially protective against prostate cancer [78, 98].

In an epidemiological study, consuming tomato and tomato products was found to be associated with a lower incidence of prostate cancer [99].

Results of a study investigating the relationship of tomatoes and tomato products with cancer revealed that the consumption of tomatoes and tomato products decreased cancer risk [98].

In another study, lycopene in tomatoes was shown to inhibit the growth and spread of cancer cells in lung cancer by reducing oxidative stress and inducing apoptosis [100].

Another vegetable in this group associated with cancer is pepper because of the capsaicin it contains. Capsaicin is thought to prevent cancer cells from growing, developing, and spreading [101].

4. Effect of preparing and cooking methods on vegetables

Vegetables are one of the most important components of human diet and are rich sources of β -carotene (provitamin A), thiamine (B1), riboflavin (B2), niacin (B3), pantothenic acid (B5), pyridoxine (B6), folic acid, ascorbic acid (vitamin C), vitamins E and K, minerals (such as iron, zinc, calcium, magnesium, and selenium), antioxidants (such as carotenoids, polyphenols, and glucosinolates), and fiber [102].

Preparation and cooking methods can greatly affect the nutritional content and acceptability of vegetables. There is no consensus in the literature as to what is the best way of preserving bioactive compounds while preparing and cooking vegetables [103].

Some vegetables are subjected to peeling in order to remove their shell or skin and make them more digestible. Minerals and other nutrients are affected by peeling. This can also cause severe loss of certain vitamins. It is known that peeling before boiling increases the loss of ascorbic acid, folic acid, or other vitamins of group B. Chopping vegetables can also change the bioavailability of bioactive compounds such as vitamins, carotenoids, polyphenols, and flavonoids [104]. Thawing, cutting, and crashing citrus vegetables can also disrupt antioxidant glucosinolates due to the presence of myrosinase enzyme found in these vegetables [105].

Cooking improves the flavor of vegetables and enables the nutrients in the vegetables to be more easily used by the digestive system. However, cooking results in some physical and

chemical changes in vegetables [106]. The effect of cooking procedure may vary depending on the various factors such as cooking technique, temperature, leakage into the cooking environment, solvent used for extraction, surface area exposed to water and oxygen, and pH [107]. In addition, each food matrix contains different compounds; therefore the same cooking technique may have different effects depending on the type of vegetable [108].

The most commonly used cooking methods are steaming, roasting, boiling, frying, sautéing, sous vide, microwave, and pressure cooking [109]. Cooking techniques affect polyphenol content and antioxidant activity levels in vegetables. Heat treatment can lead to a change in the chemical structures of vegetables, leading to the breakdown of cells and the degradation of some phenolic compounds from biological structures, the release of phenolics from the food matrix, and the conversion of insoluble phenolics to more soluble forms [110, 111]. In addition, the phenolic compounds are soluble in water. Thus, water-based cooking techniques often lead to loss of phenolics by leaking [103].

It has been reported that food processing has negative effects due to oxidation dependent losses in carotenoids and positive effects as it provides increased bioavailability [112]. Among the causes of increased carotenoid concentration in heat treatment may be greater extractability, enzymatic breakdowns, and incalculable moisture losses. Heat treatment also causes inactivation of enzymes and degradation of structures in the food matrix leading to increased bioavailability [113, 114].

The losses of minerals during preparation and cooking stages of vegetables are closely related to their solubility. Minerals are generally stable against a large number of conditions encountered during cooking, such as heat, oxidation, acidity or alkalinity. Potassium is an abundant mineral found in vegetables, and because of its high solubility in water, it is easily lost by leakage during cooking. Calcium and magnesium are usually present in an attached form in plant tissue and are therefore not easily lost by leakage. The loss of vitamin C is due in part to oxidative degradation during preparation and cooking and partly due to the leakage of the vitamin into the water used for cooking. The amount of vitamins degraded during cooking may be quite small compared to the amount lost due to leakage [115]. Due to its solubility and reactivity, folate is susceptible to potentially large losses during food processing and storage. The chemical stability of folates in plant-based foods may be adversely affected by heat, oxygen exposure, and light intensity. Since folate is highly soluble, its losses occur by leakage through the water used for washing, boiling, and cooking [116].

4.1. Root, bulb, and tuber vegetables

Onion is the richest source of quercetin, which is a flavonoid, and it is most widely used source in diet. Gennaro et al. found a 21% reduction in total quercetin uptake after onion peeling [117].

In another study on onions, the effects of boiling, microwaving, frying, and warm holding onion at 60°C for 1–2 hours on flavonoid amounts were investigated and peeling and boiling were found to decrease flavonoid levels in onion by up to 50%. It was found that other cooking methods and warming treatments did not have a significant effect on flavonoid amount [118].

Regarding the effects of cooking on onions, Lombard et al. found a 7% increase in flavonol concentration when sautéed, a 25% increase when oven baking, and an 18% decrease when boiled. They also stated that less than 5 minutes of cooking can result in retention of more than 80% of flavonols [119].

Potato contains various phenolic compounds, mainly chlorogenic acid and caffeic acid. There are several studies showing that cooking reduces [120], does not affect [121] or increases [122–125] phenolic compounds in potato. The reason for the increase in phenolic compounds during cooking is attributed to the increase in the extractability of these compounds from the cellular matrix of potato due to the textural changes in its starch structure during cooking [122].

Carrot is one of the important root vegetables rich in bioactive compounds such as carotenoids and dietary fiber. Bembem and Sadana investigated the effects of different cooking methods (boiling, steaming, pressure cooking, microwaving, and sautéing) on total phenolic content (TPC), total flavonoid content (TFC), total carotenoid and β -carotene content and antioxidant activity, and found that sautéing was the method that increased total carotene, β -carotene, and TPC the most. They reached the conclusion that sautéing and microwaving were the most appropriate ways of cooking carrots [126].

When the effect of boiling and steaming of frozen carrot on phenolic compound content was investigated, it was determined that phenolic content of carrot was significantly decreased at the end of the boiling process, whereas there was an increase in the steaming method [127]. The decrease during boiling may be due to the leakage of phenolic compounds into the boiling water.

4.2. Leaf vegetables

In a study investigating the effect of different cooking methods (boiling, steam cooking, and microwave cooking) on phytochemical content and total antioxidant capacity (TAC) of cabbage and black cabbage, which are part of the *Brassicaceae* family and are rich sources of vitamins and phytochemical compounds such as carotenoids and polyphenols, it was found that the best method in preserving the nutritious quality of vegetables was steam cooking. It was also shown that fresh vegetables preserved phytochemical compounds and TAC better than frozen samples [128].

Chang et al. studied the losses in nutritional value of several green leafy vegetables including Chinese cabbage (*Brassica pekinensis* var. *Cephalata*), swamp cabbage (*Ipomoea aquatica*), spinach (*Spinacia oleracea*), Ceylon spinach (*Basella rubra*), red spinach (*Amaranthus gangeticus*), white spinach (*Amaranthus viridis*), and Tapioca sprouts (*Manihot utilissima*) when they were treated with boiling or deep frying for 4 and 8 minutes, and found that frying reduced lutein content in all vegetables by 8–89%, and boiling reduced lutein content by 0–428%. When 8 minutes boiling procedure was compared with the 4 minutes procedure, β -carotene retention in vegetables other than Chinese cabbage and spinach changed between 18 and 380%, whereas in the frying procedure β -carotene retention increased by 2–3 times except spinach [129]. The difference in cooking conditions (time and temperature), the type of vegetables,

and the interaction between cooking methods and vegetable type may be the cause of differences observed in carotenoid composition.

4.2.1. Fruit and flower vegetables

Alvi et al. investigated the effects of peeling on tomato, which contains vitamins A, C, and E, as well as various phytochemical compounds including lycopene, and found a reduction of 18.3% in fiber, 25.4% in calcium, 32.6% in magnesium, 6.4% in phosphorus, 2.9% in potassium, 28.9% in ascorbic acid, and 17.2% in folic acid after peeling [130].

Dolinsky et al. found that the cooking method that maximized the polyphenol concentration and antioxidant capacity of tomatoes was steaming, and that microwaving significantly reduced the polyphenol content in tomatoes, and recommended microwave cooking less than other cooking methods (boiling, steaming, and pressure cooking) [131].

Pepper (*Capsicum annuum* L.) is considered to be an excellent source of antioxidants and is very rich in ascorbic acid and other phytochemicals. In a study conducted with six species of pepper, three different cooking methods were used (frying, boiling, and microwaving) and antioxidant properties of peppers after cooking procedures were evaluated. Reductions in radical scavenging activity (RSA) and total phenolic contents (TP) were observed after all cooking procedures, but the reductions after frying and microwaving were not statistically significant when compared to the initial RSA and TP levels. After a 5-minute boiling, a 77% reduction compared to initial RSA levels was obtained, and when the boiling time increased to 30 minutes, the RSA totals decreased significantly compared to raw peppers. Significant reductions were also observed in TP after 5 and 30 minutes of boiling [132]. Based on these results, it can be said that the most suitable heat treatment method for peppers are microwave use and frying. If boiling is to be performed, shorter cooking time, less water usage, and consumption of cooking water can also reduce the amount of antioxidants that can be lost.

Artichoke, which is characterized by a complex antioxidant profile, contains many bioactive compounds such as glycosides and phenolic compounds, especially caffeoylquinic acid. Ferracane et al. applied boiling, frying, and steam cooking methods on artichoke and found an increase in overall caffeoylquinic acid concentration due to the formation of different dicaffeoylquinic acid isomers in cooked artichokes compared to raw ones. However, a higher increase in the concentration of dicaffeoylquinic acid was observed in steamed and fried artichokes compared to those boiled. In addition, flavonoid concentrations were reduced in all cooking processes and this reduction was largest in frying [133].

In a study comparing vitamin C content of raw, boiled, and microwaved broccoli and cauliflower, significant reductions in vitamin C contents were found after cooking processes. Boiling process caused more vitamin C loss compared to microwaving. After 6 minutes of boiling, vitamin C levels decreased by 64.5% in broccoli, by 70.7% in white cabbage, and by 66.8% in cauliflower [134]. Based on the results, it can be said that microwaving may be preferred instead of boiling to reduce vitamin C losses.

Yuan et al. investigated the effects of steaming, microwaving, boiling, frying, and boiling followed by frying processes on vitamin C levels in broccoli. At the end of the study, it was found that in all procedures except the steaming method, loss of vitamin C was significant compared to initial levels and the highest loss was obtained in boiling followed by frying (38%) and boiling (33%) [135].

When we look at other studies conducted with broccoli belonging to the *Brassicaceae* family, it is also seen that steaming is the best way to preserve nutritional quality of broccoli [107, 136–138].

It is known for a long time that loss of nutrients in vegetables occurs during the preparation and cooking stages. Knowing the conditions that cause these losses can help limit the losses and increase the nutritional quality of the foods.

5. Conclusion

Numerous preclinical studies carried out in recent years have identified beneficial protective and enhancing effects of vegetables on health, resulting from the nutritional and non-nutritional phytochemical contents of vegetables. These phytochemicals have the ability to modify the cellular function by modulating transcription factors and altering gene expression, cellular metabolism, and cellular signaling. The World Health Organization (WHO) recommends daily intake of 5–8 portions (400–600 g) of fruits and vegetables to reduce the risk of micro nutrient deficiency, cardiovascular diseases, cancer, cognitive impairment, and other nutritional health risks.

In order to make optimum use of the nutritional content of vegetables, choosing the right methods of preparation and cooking is as important as the consumption of adequate amounts of vegetables. To minimize nutritional losses, vegetables should be chopped right before cooking, if possible by hand or by metal tools while making the minimum contact possible, each vegetable should be cooked with the method and time that is most appropriate for that vegetable, and consumed as soon as possible.

Conflict of interest

There is no 'conflict of interest'.

Author details

Taha Gökmen Ülger, Ayşe Nur Songur, Onur Çırak* and Funda Pınar Çakıroğlu

*Address all correspondence to: onrcrk@hotmail.com

Faculty of Health Sciences, Ankara University, Ankara, Turkey

References

- [1] Welbaum GE. Vegetable production and practices; IARC handbooks of cancer prevention: Fruit and vegetables. In: Vegetable History, Nomenclature, and Classification. 2015;**8**: 1-15
- [2] Pennington JAT, Fisher RA. Classification of fruits and vegetables. Journal of Food Composition and Analysis. 2009;**22**(1):23-31
- [3] Septembre-Malaterre A, Remizeb F, Pouchere P. Fruits and vegetables, as a source of nutritional compounds and phytochemicals: Changes in bioactive compounds during lactic fermentation. Food Research International. 2018;**104**:86-99
- [4] Tuttolomondo A, Casuccio A, Butta C, Pecoraro R, Di Raimondo D, Della Corte V, Arnao V, Clemente G, Maida C, Simonetta I, Miceli G, Lucifora B, Cirrincione A, Di Bona D, Corpora F, Maugeri R, Iacopino DG, Pinto A. Mediterranean diet in patients with acute ischemic stroke: Relationships between Mediterranean diet score, diagnostic subtype, and stroke severity index. Atherosclerosis. 2015;**243**:260-267
- [5] Pennington JAT, Fisher RA. Food component profiles for fruit and vegetable subgroups. Journal of Food Composition and Analysis. 2010;**23**:411-418
- [6] Lintas C. Nutritional aspects of fruit and vegetable consumption. In: Lauret F, editor. Les fruits et légumes dans les économies méditerranéennes: actes du colloque de Chania. Montpellier: CIHEAM; 1992. pp. 79-87
- [7] Sağlık, Bakanlığı T.C. Türkiye Beslenme Rehberi 2015. T.C. Sağlık Bakanlığı. TÜRKİYE. Yayın No: 1031. 2016. p. 288
- [8] Belemkar S, Dhameliya K, Pata MK. Comparative study of garlic species (*Allium sativum* and *Allium porrum*) on glucose uptake in diabetic rats. Journal of Taibah University Medical Sciences. 2013;**8**:80-85. DOI: 10.1016/j.jtumed.2013.04.002
- [9] Akash MSH, Rehman K, Chen S. Spice plant *Allium cepa*: Dietary supplement for treatment of type 2 diabetes mellitus. Nutrition. 2014;**30**:1128-1137. DOI: 10.1016/j.nut.2014.02.011
- [10] Nickavar B, Yousefian N. Inhibitory effects of six allium species on-amylase enzyme activity. Iranian Journal of Pharmaceutical Research. 2010;**8**:53-57
- [11] Kumar R, Chhatwal S, Arora S, Sharma S, Singh J, Singh N, Bhandari V, Khurana A. Antihyperglycemic, antihyperlipidemic, anti-inflammatory and adenosine deaminase-lowering effects of garlic in patients with type 2 diabetes mellitus with obesity. Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy. 2013;**6**:49-56. DOI: 10.2147/DMSO.S38888
- [12] Ashraf R, Aamir K, Shaikh AR, Ahmed T. Effects of garlic on dyslipidemia in patients with type 2 diabetes mellitus. Journal of Ayub Medical College, Abbottabad. 2005;**17**:60-64
- [13] Eidi A, Eidi M, Esmaeili E. Antidiabetic effect of garlic (*Allium sativum* L.) in normal and streptozotocin-induced diabetic rats. Phytomedicine. 2006;**13**:624-629. DOI: 10.1016/j.phymed.2005.09.010

- [14] Madkor HR, Mansour SW, Ramadan G. Modulatory effects of garlic, ginger, turmeric and their mixture on hyperglycemia, dyslipidemia and oxidative stress in streptozotocin-nicotinamide diabetic rats. *British Journal of Nutrition*. 2011;**105**:1210-1217. DOI: 10.1017/S0007114510004927
- [15] Gómez-Arbeláez D, Lahera V, Oubiña P, Valero-Muñoz M, De las Heras N, Rodríguez Y, Garcia RG, Camacho PA, Jaramillo PL. Aged garlic extract improves adiponectin levels in subjects with metabolic syndrome: A double-blind, placebo-controlled, randomized, crossover study. *Mediators of Inflammation*. 2013;**2013**:1-6. DOI: 10.1155/2013/285795
- [16] Aktaş G, Şit M, Tekçe H. Yeni adipokinler: Leptin, adiponektin ve omentin. *Abant Medical Journal*. 2013;**2**:56-62. DOI: 10.5505/abantmedj.2013.97269
- [17] Padiya R, Khatua TN, Bagul PK, Kuncha M, Banerjee SK. Garlic improves insulin sensitivity and associated metabolic syndromes in fructose fed rats. *Nutrition & Metabolism*. 2011;**8**:53. DOI: 10.1186/1743-7075-8-53
- [18] Kim M, Kim H. Effect of garlic on high fat induced obesity. *Acta Biologica Hungarica*. 2011;**62**:244-254. DOI: 10.1556/ABiol.62.2011.3.4
- [19] Lee SJ, Hwang CR, Kang JR, Shin JH, Kang MJ, Sung NJ. Anti-obesity effect of red garlic composites in rats fed a high fat-cholesterol diet. *Journal of Life Sciences*. 2012;**22**:671-680. DOI: 10.5352/JLS.2012.22.5.671
- [20] El-Demerdash FM, Yousef MI, El-Naga NA. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. *Food and Chemical Toxicology*. 2005;**43**:57-63. DOI: 10.1016/j.fct.2004.08.012
- [21] Eyo JE, Ozougwu JC, Echi PC. Hypoglycemic effects of *Allium cepa*, *Allium sativum* and *Zingiber officinale* aqueous extracts on alloxan-induced diabetic *Rattus norvegicus*. *Medical Journal of Islamic World Academy of Sciences*. 2011;**19**:121-126. DOI: 10.4314/br.v6i2.28672
- [22] Ogunmodede OS, Saalu LC, Ogunlade B, Akunna GG, Oyewopo AO. An evaluation of the hypoglycemic, antioxidant and hepatoprotective potentials of onion (*Allium cepa* L.) on alloxan-induced diabetic rabbits. *International Journal of Pharmacology*. 2012;**8**:21-29. DOI: 10.3923/ijp.2012.21.29
- [23] Yoshinari O, Shiojima Y, Igarashi K. Anti-obesity effects of onion extract in Zucker diabetic fatty rats. *Nutrients*. 2012;**4**:1518-1526. DOI: 10.3390/nu4101518
- [24] Matsunaga S, Azuma K, Watanabe M, Tsuka T, Imagawa T, Osaki T, Okamoto Y. Onion peel tea ameliorates obesity and affects blood parameters in a mouse model of high-fat-diet-induced obesity. *Experimental and Therapeutic Medicine*. 2014;**7**:379-382. DOI: 10.3892/etm.2013.1433
- [25] Ebrahimi-Mamaghani M, Saghafi-Asl M, Pirouzpanah S, Asghari-Jafarabadi M. Effects of raw red onion consumption on metabolic features in overweight or obese women with polycystic ovary syndrome: A randomized controlled clinical trial. *The Journal of Obstetrics and Gynecology Research*. 2014;**40**:1067-1076. DOI: 10.1111/jog.12311

- [26] Naeem F, Khan SH. Purslane (*Portulaca oleracea* L.) as phyto-genic substance—A review. *Journal of Herbs, Spices & Medicinal Plants*. 2013;**19**:216-232. DOI: 10.1080/10496475.2013.782381
- [27] Bolkent Ş, Yanardağ R, Tabakoğlu-Oğuz A, Özsoy-Saçan Ö. Effects of chard (*Beta vulgaris* L. var. cicla) extract on pancreatic B cells in streptozotocin-diabetic rats: A morphological and biochemical study. *Journal of Ethnopharmacology*. 2000;**73**:251-259. DOI: 10.1016/S0378-8741(00)00328-7
- [28] Sultana A, Rahman K. *Portulaca oleracea* Linn. A global panacea with ethno-medicinal and pharmacological potential. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2013;**5**:33-39
- [29] Wainstein J, Landau Z, Dayan YB, Jakubowicz D, Grothe T, Perrinjaquet-Moccetti T, Boaz M. Purslane extract and glucose homeostasis in adults with type 2 diabetes: A double-blind, placebo-controlled clinical trial of efficacy and safety. *Journal of Medicinal Food*. 2016;**19**:133-140. DOI: 10.1089/jmf.2015.0090
- [30] El-Sayed MIK. Effects of *Portulaca oleracea* L. seeds in treatment of type-2 diabetes mellitus patients as adjunctive and alternative therapy. *Journal of Ethnopharmacology*. 2011;**137**:643-651. DOI: 10.1016/j.jep.2011.06.020
- [31] Heidarzadeh S, Farzanegi P, Azarbayjani MA, Daliri R. Purslane effect on GLP-1 and GLP-1 receptor in type 2 diabetes. *Electronic Physician*. 2013;**5**:582-587. DOI: 10.14661/2013.582-587
- [32] El-Newary SA. The hypolipidemic effect of *Portulaca oleracea* L. stem on hyperlipidemic Wistar albino rats. *Annals of Agricultural Sciences*. 2016;**61**:111-124. DOI: 10.1016/j.aos.2016.01.002
- [33] Zidan Y, Bouderbala S, Djellouli F, Lacaille-Dubois MA, Bouchenak M. *Portulaca oleracea* reduces triglyceridemia, cholesterolemia, and improves lecithin: Cholesterol acyltransferase activity in rats fed enriched-cholesterol diet. *Phytomedicine*. 2014;**21**:1504-1508. DOI: 10.1016/j.phymed.2014.07.010
- [34] Gu JF, Zheng ZY, Yuan JR, Zhao BJ, Wang CF, Zhang L, Xu QY, Yin QW, Feng L, Jia XB. Comparison on hypoglycemic and antioxidant activities of the fresh and dried *Portulaca oleracea* L. in insulin-resistant HepG2 cells and streptozotocin-induced C57BL/6J diabetic mice. *Journal of Ethnopharmacology*. 2015;**161**:214-223. DOI: 10.1016/j.jep.2014.12.002
- [35] Samarghandian S, Borji A, Farkhondeh T. Attenuation of oxidative stress and inflammation by *Portulaca oleracea* in streptozotocin-induced diabetic rats. *Journal of Evidence-Based Complementary & Alternative Medicine*. 2017;**22**:562-566. DOI: 10.1177/2156587217692491
- [36] Gezginci-Oktayoglu S, Sacan O, Bolkent S, Ipci Y, Kabasakal L, Sener G, Yanardag R. Chard (*Beta vulgaris* L. var. cicla) extract ameliorates hyperglycemia by increasing GLUT2 through Akt2 and antioxidant defense in the liver of rats. *Acta Histochemica*. 2014;**116**:32-39. DOI: 10.1016/j.acthis.2013.04.016

- [37] Kabir AU, Samad MB, Ahmed A, Jahan MR, Akhter F, Tasnim J, Hannan JMA. Aqueous fraction of *Beta vulgaris* ameliorates hyperglycemia in diabetic mice due to enhanced glucose stimulated insulin secretion, mediated by acetylcholine and GLP-1, and elevated glucose uptake via increased membrane bound GLUT4 transporters. *PLoS One*. 2015;**10**:e0116546. DOI: 10.1371/journal.pone.0116546
- [38] Hashem AN, Soliman MS, Hamed MA, Swilam NF, Lindequist U, Nawwar MA. *Beta vulgaris* subspecies *cicla* var. *flavescens* (Swiss chard): Flavonoids, hepatoprotective and hypolipidemic activities. *Die Pharmazie - An International Journal of Pharmaceutical Sciences*. 2016;**71**:227-232. DOI: 10.1691/ph.2016.5821
- [39] Chang HP, Wang ML, Chan MH, Chiu YS, Chen YH. Antiobesity activities of indole-3-carbinol in high-fat-diet-induced obese mice. *Nutrition*. 2011;**27**:463-470. DOI: 10.1016/j.nut.2010.09.006
- [40] Jayakumar P, Pugalendi KV, Sankaran M. Attenuation of hyperglycemia-mediated oxidative stress by indole-3-carbinol and its metabolite 3,3'-diindolylmethane in C57BL/6J mice. *Journal of Physiology and Biochemistry*. 2014;**70**:525-534. DOI: 10.1007/s13105-014-0332-5
- [41] Lee JJ, Shin HD, Lee YM, Kim AR, Lee MY. Effect of broccoli sprouts on cholesterol-lowering and anti-obesity effects in rats fed high fat diet. *Journal of the Korean Society of Food Science and Nutrition*. 2009;**38**:309-318. DOI: 10.3746/jkfn.2009.38.3.309
- [42] Mohamed S. Functional foods against metabolic syndrome (obesity, diabetes, hypertension and dyslipidemia) and cardiovascular disease. *Trends in Food Science & Technology*. 2014;**35**:114-128. DOI: 10.1016/j.tifs.2013.11.001
- [43] Helmstädter A. Beans and diabetes: *Phaseolus vulgaris* preparations as antihyperglycemic agents. *Journal of Medicinal Food*. 2010;**13**:251-254. DOI: 10.1089/jmf.2009.0002
- [44] Rebello CJ, Greenway FL, Finley JW. A review of the nutritional value of legumes and their effects on obesity and its related co-morbidities. *Obesity Reviews*. 2014;**15**:392-407. DOI: 10.1111/obr.12144
- [45] Lozano R, Naghavi M, Foreman K, Lim S, Shibuya K, Aboyans V, Abraham J, Adair T, Aggarwal R, Ahn SY, AlMazroa MA, Alvarado M, Anderson R, Anderson L. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: A systematic analysis for the global burden of disease study 2010. *The Lancet*. 2012;**380**:2095-2128. DOI: 10.1016/S0140-6736(12)61728-0
- [46] Roth GA, Forouzanfar MH, Moran AE, Barber R, Nguyen G, Feigin VL, Murray CJ. Demographic and epidemiologic drivers of global cardiovascular mortality. *New England Journal of Medicine*. 2015;**372**:1333-1341. DOI: 10.1056/NEJMoa1406656
- [47] Radhika G, Sudha V, Sathya RM, Ganesan A, Mohan V. Association of fruit and vegetable intake with cardiovascular risk factors in urban south Indians. *British Journal of Nutrition*. 2008;**99**:398-405. DOI: 10.1017/S0007114507803965

- [48] Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE, Hilpert KF, Etherton TD. Bioactive compounds in foods: Their role in the prevention of cardiovascular disease and cancer. *The American Journal of Medicine*. 2002;**113**:71-88. DOI: 10.1016/S0002-9343(01)00995-0
- [49] Yeh YY, Liu L. Cholesterol-lowering effect of garlic extracts and organosulfur compounds: Human and animal studies. *The Journal of Nutrition*. 2001;**131**:989-993. DOI: 10.1093/jn/131.3.989S
- [50] Chang HS, Yamato O, Yamasaki M, Maede Y. Modulatory influence of sodium 2-propenyl thiosulfate from garlic on cyclooxygenase activity in canine platelets: Possible mechanism for the anti-aggregatory effect. *Prostaglandins, Leukotrienes and Essential Fatty Acids*. 2005;**72**:351-355. DOI: 10.1016/j.plefa.2005.01.003
- [51] Moriguchi T, Takasugi N, Itakura Y. The effects of aged garlic extract on lipid peroxidation and the deformability of erythrocytes. *The Journal of Nutrition*. 2001;**131**:1016-1019. DOI: 10.1093/jn/131.3.1016S
- [52] Ried K, Frank OR, Stocks NP. Aged garlic extract reduces blood pressure in hypertensives: A dose-response trial. *European Journal of Clinical Nutrition*. 2013;**67**:64-70. DOI: 10.1038/ejcn.2012.178
- [53] Knekt P, Jarvinen R, Reunanen A, Maatela J. Flavonoid intake and coronary mortality in Finland: A cohort study. *BMJ*. 1996;**312**:478-481. DOI: 10.1136/bmj.312.7029.478
- [54] Briggs WH, Folts JD, Osman HE, Goldman IL. Administration of raw onion inhibits platelet-mediated thrombosis in dogs. *The Journal of Nutrition*. 2001;**131**:2619-2622. DOI: 10.1093/jn/131.10.2619
- [55] Colina-Coca C, González-Peña D, De Ancos B, Sánchez-Moreno C. Dietary onion ameliorates antioxidant defence, inflammatory response, and cardiovascular risk biomarkers in hypercholesterolemic Wistar rats. *Journal of Functional Foods*. 2017;**36**:300-309. DOI: 10.1016/j.jff.2017.07.014
- [56] Joshipura KJ, Hu FB, Manson JE, Stampfer MJ, Rimm EB, Speizer FE, Willett WC. The effect of fruit and vegetable intake on risk for coronary heart disease. *Annals of Internal Medicine*. 2001;**134**:1106-1114. DOI: 10.7326/0003-4819-134-12-200106190-00010
- [57] Lidder S, Webb AJ. Vascular effects of dietary nitrate (as found in green leafy vegetables and beetroot) via the nitrate-nitrite-nitric oxide pathway. *British Journal of Clinical Pharmacology*. 2013;**75**:677-696. DOI: 10.1111/j.1365-2125.2012.04420.x
- [58] Huang T, Yang B, Zheng J, Li G, Wahlqvist ML, Li D. Cardiovascular disease mortality and cancer incidence in vegetarians: A meta-analysis and systematic review. *Annals of Nutrition and Metabolism*. 2012;**60**:233-240. DOI: 10.1159/000337301
- [59] Larsson SC, Virtamo J, Wolk A. Total and specific fruit and vegetable consumption and risk of stroke: A prospective study. *Atherosclerosis*. 2013;**227**:147-152. DOI: 10.1016/j.atherosclerosis.2012.12.022

- [60] Rastogi T, Reddy KS, Vaz M, Spiegelman D, Prabhakaran D, Willett WC, Ascherio A. Diet and risk of ischemic heart disease in India. *The American Journal of Clinical Nutrition*. 2004;**79**:582-592. DOI: 10.1093/ajcn/79.4.582
- [61] Lundberg JO, Feelisch M, Björne H, Jansson EÅ, Weitzberg E. Cardioprotective effects of vegetables: Is nitrate the answer? *Nitric Oxide*. 2006;**15**:359-362. DOI: 10.1016/j.niox.2006.01.013
- [62] Manchali S, Murthy KNC, Patil BS. Crucial facts about health benefits of popular cruciferous vegetables. *Journal of Functional Foods*. 2012;**4**:94-106. DOI: 10.1016/j.jff.2011.08.004
- [63] Jeffery EH, Araya M. Physiological effects of broccoli consumption. *Phytochemistry Reviews*. 2009;**8**:283-298. DOI: 10.1007/s11101-008-9106-4
- [64] Piao CS, Gao S, Lee GH, Park BH, Chae SW, Chae HJ, Kim SH. Sulforaphane protects ischemic injury of hearts through antioxidant pathway and mitochondrial KATP channels. *Pharmacological Research*. 2010;**61**:342-348. DOI: 10.1016/j.phrs.2009.11.009
- [65] Blekkenhorst LC, Bondonno CP, Lewis JR, Devine A, Zhu K, Lim WH, Hodgson JM. Cruciferous and allium vegetable intakes are inversely associated with 15-year atherosclerotic vascular disease deaths in older adult women. *Journal of the American Heart Association*. 2017;**6**:e006558. DOI: 10.1161/JAHA.117.006558
- [66] Zhang X, Shu XO, Xiang YB, Yang G, Li H, Gao J, Zheng W. Cruciferous vegetable consumption is associated with a reduced risk of total and cardiovascular disease mortality. *The American Journal of Clinical Nutrition*. 2011;**94**:240-246. DOI: 10.3945/ajcn.110.009340
- [67] Saluk J, Bijak M, Kołodziejczyk-Czepas J, Posmyk M, Janas K, Wachowicz B. Anthocyanins from red cabbage extract—Evidence of protective effects on blood platelets. *Open Life Sciences*. 2012;**7**:655-663. DOI: 10.2478/s11535-012-0057-9
- [68] Murashima M, Watanabe S, Zhuo XG, Uehara M, Kurashige A. Phase 1 study of multiple biomarkers for metabolism and oxidative stress after one-week intake of broccoli sprouts. *BioFactors*. 2004;**22**:271-275. DOI: 10.1002/biof.5520220154
- [69] Bazzano LA, He J, Ogden LG, Loria C, Vupputuri S, Myers L, Whelton PK. Legume consumption and risk of coronary heart disease in US men and women: NHANES I epidemiologic follow-up study. *Archives of Internal Medicine*. 2001;**161**:2573-2578. DOI: 10.1001/archinte.161.21.2573
- [70] Lattimer JM, Haub MD. Effects of dietary fiber and its components on metabolic health. *Nutrients*. 2010;**2**:1266-1289. DOI: 10.3390/nu2121266
- [71] Shi J, Arunasalam K, Yeung D, Kakuda Y, Mittal G, Jiang Y. Saponins from edible legumes: Chemistry, processing, and health benefits. *Journal of Medicinal Food*. 2004;**7**:67-78. DOI: 10.1089/109662004322984734
- [72] Menotti A, Kromhout D, Blackburn H, Fidanza F, Buzina R, Nissinen A. Food intake patterns and 25-year mortality from coronary heart disease: Cross-cultural correlations in the seven countries study. *European Journal of Epidemiology*. 1999;**15**:507-515

- [73] Nöthlings U, Schulze MB, Weikert C, Boeing H, Van der Schouw YT, Bamia C, Peeters PH. Intake of vegetables, legumes, and fruit, and risk for all-cause, cardiovascular, and cancer mortality in a European diabetic population. *The Journal of Nutrition*. 2008;**138**:775-781. DOI: 10.1093/jn/138.4.775
- [74] Insel P, Turner RE, Ross D. Diet and health. In: Insel P, Turner RE, Ross D, editors. *Nutrition*. 3rd ed. Jones and Barlett Publishers. 2007. Judbury, Massachusetts. pp. 598-640
- [75] Heim KC, Spinalla MJ. In: Bagchi D, Preuss H, Swaropp A, editors. *Nutraceuticals and Functional Foods in Human Health and Disease Prevention*. 1st ed. CRP Press, Taylor Francis Group; 2016. Las Vegas. Pp. 361-390
- [76] WHO (World Health Organization) [Internet]. 2018. Available from: <http://www.who.int/dietphysicalactivity/fruit/en/> [Accessed February 8, 2018]
- [77] Soh Y, Shin M, Lee J, Jang J, Kim OH, Kang H, Surh Y. Oxidative DNA damage and glioma cell death induced by tetrahydropapaveroline. *Mutation Research*. 2003;**544**: 129-142. DOI: 10.1016/j.mrrev.2003.06.023
- [78] AICR (American Institute for Cancer Research) [Internet]. 2015. Available from: <http://www.aicr.org/> [Accessed: March 3, 2018]
- [79] WHO (World Health Organization) [Internet]. 2018. Available from: http://www.who.int/elena/titles/fruit_vegetables_ncds/en/ [Accessed February 10, 2018]
- [80] Wang Y, Li F, Wang Z, Qiu TQ, Shen Y, Wang M. Fruit and vegetable consumption and risk of lung cancer: A dose-response meta-analysis of prospective cohort studies. *Lung Cancer*. 2015;**88**(2):124-130. DOI: 10.1016/j.lungcan.2015.02.015
- [81] Tang L, Lee AH, Su D, Binns CW. Fruit and vegetable consumption associated with reduced risk of epithelial ovarian cancer in southern Chinese women. *Gynecologic Oncology*. 2014;**132**:241-247. DOI: 10.1016/j.ygyno.2013.10.020
- [82] Shigiharaa M, Obara T, Nagai M, Sugawara Y, Watanabe T, Kakizaki M, Nishino Y, Kuriyama S, Tsuji I. Consumption of fruits, vegetables, and seaweeds (sea vegetables) and pancreatic cancer risk: The Ohsaki cohort study. *Cancer Epidemiology*. 2014;**38**(2): 129-136. DOI: 10.1016/j.canep.2014.01.001
- [83] NOA (National Onion Association) [Internet]. 2018. Available from: www.onions-usa.org [Accessed: February 5, 2018]
- [84] Wang Y, Tian W, Ma X. Inhibitory effects of onion (*Allium cepa* L.) extract on proliferation of cancer cells and adipocytes via inhibiting fatty acid synthase. *Asian Pacific Journal of Cancer Prevention*. 2012;**13**:5573-5579. DOI: 10.7314/APJCP.2012.13.11.5573
- [85] Das S. Garlic-A natural source of cancer preventive compounds. *Asian Pacific Journal of Cancer Prevention*. 2002;**3**:305-311
- [86] Nicastro HL, Ross SA, Milner JA. Garlic and onions: Their cancer prevention properties. *Cancer Prevention Research*. 2015;**8**:181-190. DOI: 10.1158/1940-6207.CAPR-14-0172

- [87] Fleischauer AT, Poole C, Arab L. Garlic consumption and cancer prevention: Meta-analyses of colorectal and stomach cancers. *The American Journal of Clinical Nutrition*. 2000;**72**:1047-1052
- [88] Steiawan VW, Yu GP, Lu QY, Lu ML, Yu SZ, Mu L, Zhang JG, Kurtz RC, Cai L, Hsieh CC, Zhang ZF. Allium vegetables and stomach cancer risk in China. *Communication Research*. 2005;**6**:387-395
- [89] Carlos J, Dias S. Nutritional and health benefits of carrots and their seed extracts. *Food and Nutrition Sciences*. 2014;**5**:2147-2156. DOI: 10.4236/fns.2014.522227
- [90] Xu X, Cheng Y, Li S, Zhu Y, Xu X, Zheng X, Mao Q, Xie L. Dietary carrot consumption and the risk of prostate cancer. *European Journal of Nutrition*. 2014;1-9. DOI: 10.1007/s00394-014-0667-2
- [91] Garti H. Effects of carrot consumption on intestinal cancer risk [thesis]. United Kingdom: Newcastle University; 2006
- [92] Pollock RL. Comparing consumption of green leafy vegetables to cruciferous vegetables in relations to incidence of 17 different cancers: A meta-analysis. *The Global Journal of Medical Research*. 2016;**16**:30-39
- [93] Tao J, Li Y, Li S, Li HB. Plant foods for the prevention and management of colon cancer. *Journal of Functional Foods*. 2018;**42**:95-110. DOI: 10.1016/j.jff.2017.12.064
- [94] Tewani R, Sharma JK, Rao SV. Spinach (Palak) natural laxative. *International Journal of Applied Research and Technology*. 2016;**1**:140-148. DOI: 10.4172/2157-7471.1000110
- [95] AND (Academy of Nutrition and Dietetics) [Internet]. 2018. Available from: <https://www.eatright.org/food> [Accessed February 2, 2018]
- [96] Ullah MF. Sulforaphane (SFN): An isothiocyanate in a cancer chemoprevention paradigm. *Medicine*. 2015;**2**:141-156. DOI: 10.3390/medicines2030141
- [97] Palak, Soni K, Thakur A, Kohli K. Broccoli: An insight into formulation and patentability aspects. *Drug design*. 2016;**5**:1-12. DOI: 10.4172/2169-0138.1000139
- [98] Jang J, Surh Y. Potentiation of cellular antioxidant capacity by Bcl-2: Applications for its antiapoptotic function. *Biochemical Pharmacology*. 2003;**(8)**:1371-1379. DOI: 10.1016/S0006-2952(03)00487-8
- [99] Hadley CW, Miller EC, Schwartz SJ, Clinton SK. Tomatoes, lycopene, and prostate cancer. *Progress and Promise*. 2002:869-880. DOI: 1535-3702/02/22710-0869\$15.00
- [100] Palozza P, Simone RE, Catalano A, Mele MC. Tomato lycopene and lung cancer prevention: From experimental to human studies. *Cancer*. 2011;**3**:2333-2357. DOI: 10.3390/cancers3022333
- [101] Clark R, Lee SH. Anticancer properties of capsaicin against human cancer. *Anticancer Research*. 2016;**36**:837-844

- [102] Prodanov M, Sierra I, Vidal-Valverde C. Influence of soaking and cooking on the thiamin, riboflavin and niacin contents of legumes. *Food Chemistry*. 2004;**84**(2):271-277
- [103] Murador D, Braga AR, Da Cunha D, De Rosso V. Alterations in phenolic compound levels and antioxidant activity in response to cooking technique effects: A meta-analytic investigation. *Critical Reviews in Food Science and Nutrition*. 2018;**58**(2):169-177
- [104] Dos Reis LCR, de Oliveira VR, Hagen MEK, Jablonski A, Flôres SH, de Oliveira Rios A. Effect of cooking on the concentration of bioactive compounds in broccoli (*Brassica oleracea* var. avenger) and cauliflower (*Brassica oleracea* var. Alphina F1) grown in an organic system. *Food Chemistry*. 2015;**172**:770-777
- [105] Rodrigues AS, Rosa EAS. Effect of post-harvest treatments on the level of glucosinolates in broccoli. *Journal of the Science of Food and Agriculture*. 1999;**79**:1028-1032
- [106] Rehman ZU, Islam M, Shah WH. Effect of microwave and conventional cooking on insoluble dietary fibre components of vegetables. *Food Chemistry*. 2003;**80**:237-240
- [107] Wachtel-Galor S, Wong KW, Benzie IFF. The effect of cooking on *Brassica* vegetables. *Food Chemistry*. 2008;**110**(3):706-710
- [108] Bernhardt S, Schlich E. Impact of different cooking methods on food quality: Retention of lipophilic vitamins in fresh and frozen vegetables. *Journal of Food Engineering*. 2006;**77**(2):327-333
- [109] Fabbri ADT, Crosby GA. A review of the impact of preparation and cooking on the nutritional quality of vegetables and legumes. *International Journal of Gastronomy and Food Science*. 2016;**3**:2-11
- [110] Dini I, Tenore GC, Dini A. Effect of industrial and domestic processing on antioxidant properties of pumpkin pulp. *LWT - Food Science and Technology*. 2013;**53**(1):382-385
- [111] Gahler S, Otto K, Bohm V. Alterations of vitamin C, total phenolics, and antioxidant capacity as affected by processing tomatoes to different products. *Journal of Agricultural and Food Chemistry*. 2003;**51**(27):7962-7968
- [112] Van den Berg H, Faulks R, Fernando Granado H, Hirschberg J, Olmedilla B, Sandmann G, Southon S, Stahl W. The potential for the improvement of carotenoid levels in foods and the likely systemic effects. *Journal of the Science of Food and Agriculture*. 2000;**80**:880-912
- [113] Porrini M, Riso P, Testolin G. Absorption of lycopene from single or daily portions of raw and processed tomato. *British Journal of Nutrition*. 1998;**80**:353-361
- [114] Stahl W, Sies H. Uptake of lycopene and its geometrical-isomers is greater from heat-processed than from unprocessed tomato juice in humans. *The Journal of Nutrition*. 1992;**122**:2161-2166
- [115] Charlton KE, Patrick P, Dowling L, Khulani K, Jensen E. Ascorbic acid losses in vegetables associated with cook-chill food preparation. *South African Journal of Clinical Nutrition*. 2004;**17**(2):56-63

- [116] Scott J, Rebeille F, Fletcher J. Folic acid and folates: The feasibility for nutritional enhancement in plant foods. *Journal of the Science of Food and Agriculture*. 2000;**80**:795-824
- [117] Gennaro L, Leonardi C, Esposito F, Salucci M, Maiani G, Quaglia G, Fogliano VB. Flavonoid and carbohydrate contents in tropea red onions: Effects of homelike peeling and storage. *Journal of Agricultural and Food Chemistry*. 2002;**50**(7):1904-1910
- [118] Ewald C, Fjelkner-Modig S, Johansson K, Sjöholm I, Akesson B. Effect of processing on major flavonoids in processed onions, green beans, and peas. *Food Chemistry*. 1999;**64**:231-235
- [119] Lombard K, Peffley E, Geoffriau E, Thompson L, Herring A. Quercetin in onion (*Allium cepa* L.) after heat-treatment simulating home preparation. *Journal of Food Composition and Analysis*. 2005;**18**(6):571-581
- [120] Tudela JA, Cantos E, Espin JC, Tomás-Barberán FA, Gil MI. Induction of antioxidant flavonol biosynthesis in fresh-cut potatoes. Effect of domestic cooking. *Journal of Agricultural and Food Chemistry*. 2002;**50**:5925-5931
- [121] Andlauer W, Stumpf C, Hubert M, Rings A, Fürst P. Influence of cooking process on phenolic marker compounds of vegetables. *International Journal for Vitamin and Nutrition Research*. 2003;**73**:152-159
- [122] Blessington T, Nzaramba MN, Scheuring DC, Hale AL, Reddivari L, Miller JC. Cooking methods and storage treatments of potato: Effects on carotenoids, antioxidant activity and phenolics. *American Journal of Potato Research*. 2010;**87**:479-491
- [123] Brown CR, Durst RW, Wrolstad R, De Jong W. Variability of phytonutrient content of potato in relation to growing location and cooking method. *Potato Research*. 2008;**51**: 259-270
- [124] Mattila P, Hellström J. Phenolic acids in potatoes, vegetables, and some of their products. *Journal of Food Composition and Analysis*. 2007;**20**:152-160
- [125] Wu X, Beecher GR, Holden JM, Haytowitz DB, Gebhardt SE, Prior RL. Lipophilic and hydrophilic antioxidant capacities of common foods in the United States. *Journal of Agricultural and Food Chemistry*. 2004;**52**:4026-4037
- [126] Bembem K, Sadana B. Effect of different cooking methods on the antioxidant components of carrot. *Bioscience Discovery*. 2014;**5**(1):112-116
- [127] Mazzeo T, N'Dri D, Chiavaro E, Visconti A, Fogliano V, Pellegrini N. Effect of two cooking procedures on phytochemical compounds, total antioxidant capacity and colour of selected frozen vegetables. *Food Chemistry*. 2011;**128**:627-633
- [128] Pellegrini N, Chiavaro E, Gardana C, Mazzeo T, Contino D, Gallo M, Riso P, Fogliano V, Porrini M. Effect of different cooking methods on color, phytochemical concentration, and antioxidant capacity of raw and frozen *Brassica* vegetables. *Journal of Agricultural and Food Chemistry*. 2010;**58**:4310-4321

- [129] Chang SK, Nagendra Prasad K, Amin I. Carotenoids retention in leafy vegetables based on cooking methods. *International Food Research Journal*. 2013;**20**(1):457-465
- [130] Alvi S, Khan KM, Munir AS, Shahid M. Effect of peeling and cooking on nutrients in vegetables. *Pakistan Journal of Nutrition*. 2003;**2**(3):189-191
- [131] Dolinsky M, Agostinho C, Ribeiro D, De Souza Rocha G, Barroso SG, Ferreira D, Polinati R, Ciarelli G, Fialho E. Effect of different cooking methods on the polyphenol concentration and antioxidant capacity of selected vegetables. *Journal of Culinary Science & Technology*. 2016;**14**(1):1-12
- [132] Chuah AM, Lee Y-C, Yamaguchi T, Takamura H, Yin L-J, Matoba T. Effect of cooking on the antioxidant properties of coloured peppers. *Food Chemistry*. 2008;**111**:20-28
- [133] Ferracane R, Pellegrini N, Visconti A, Graziani G, Chiavaro E, Miglio C, Fogliano V. Effects of different cooking methods on antioxidant profile, antioxidant capacity, and physical characteristics of artichoke. *Journal of Agricultural and Food Chemistry*. 2008;**56**:8601-8608
- [134] Shams El-Din MHA, Abdel-Kader MM, Makhoulouf SK, OSS M. Effect of some cooking methods on natural antioxidants and their activities in some *Brassica* vegetables. *World Applied Sciences Journal*. 2013;**26**(6):697-703
- [135] Yuan G-F, Sun B, Yuan J, Wang Q-M. Effects of different cooking methods on health-promoting compounds of broccoli. *Journal of Zhejiang University Science B*. 2009;**10**(8):580-588
- [136] Bongoni R, Verkerk R, Steenbekkers B, Dekker M, Stiege M. Evaluation of different cooking conditions on broccoli (*Brassica oleracea* var. *italica*) to improve the nutritional value and consumer acceptance. *Plant Foods for Human Nutrition*. 2014;**69**(3):228-234
- [137] Mahn A, Reyes A. An overview of health-promoting compounds of broccoli (*Brassica oleracea* var. *italica*) and the effect of processing. *Journal of Food Science and Technology*. 2012;**18**(6):503-514
- [138] Stea TH, Johansson M, Jägerstad M, Frølich W. Retention of folates in cooked, stored and reheated peas, broccoli and potatoes for use in modern large-scale services systems. *Food Chemistry*. 2007;**101**(3):1095-1107