

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Short-Term Response of Plants Grown under Heavy Metal Toxicity

Prasann Kumar and Shweta Pathak

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/intechopen.75722>

Abstract

Sorghum vulgare L. plants when exposed to cadmium nitrate with the concentrations of 70 and 150 ppm per kg of soil for 90 days exhibited phytotoxic responses. The observations of specific responses were dependent on treatment combinations. The significant hazardous effects and oxidative damage of cadmium nitrate (70 and 150 ppm) were evident by increased MDA content and hydrogen peroxide content. However, these responses were reversed by exogenous application of putrescine (2.5 and 5.0 mM) and mycorrhiza (*Glomus*; 150 inoculants per kg of soil), more so, in their combined treatment, at different DAS. But combined treatment of putrescine and mycorrhiza enhanced the stability of sorghum by reducing the ROS production in plant cells. On the basis of the data obtained, it is concluded that plants responded up to 70 ppm cadmium nitrate with stress-induced responses, which were ameliorated by combined application of putrescine and *Glomus* mycorrhiza.

Keywords: agriculture, biotic, cadmium, density, economic, forage

1. Introduction

Cadmium (Cd) is one of the components of the earth's crust and present in several places or in different ecosystems on earth i.e., terrestrial, aquatic and others. Benavides et al. [1] reported that cadmium is one of the most hazardous heavy metals in the atmosphere, soil and aquatic system which is finally going into our food chain and responsible for serious environmental problem leading to the health hazards in the living organisms, for instance, mutagenesis, lung cancer, convulsion and brain damage. The alleviation or inhibition of cadmium in plants has, therefore, caused extensive attention of the whole society [2, 3]. It is must to know about

cadmium, its physical and chemical properties, isotopic studies In the atmosphere, Cd can enter by burning of coal, mining of metals as well as refining process which may lead to rise in Cd level in the soil by atmospheric fallout also. If we see the atmospheric fallout of Cd from atmospheric air, it follows in the order Remote area < Rural area < Urban areas. Due to long term effects of cadmium, the countries fixed its tolerance limit. The European Economic Committee proposed the concept of PTE i.e. Potential Toxic Elements. PTE for the Cadmium in soil is 1.0–3.0 mg/kg of dry soil [4]. If we think about the aging of the metal in the soil, then a distinction should be made between persistence of total metals in the soil and the persistence of bioavailable forms of metals. This aging of the metals depends on soil acidity. Evidence of the aging process is provided by studies of metal extractability and liability. There are many terms used to describe and categorize metals, including traces metals, transition metals, micronutrients, toxic metals, heavy metals. Bjerrum's [5] meaning of "heavy metals" depends on the density of the natural type of the metal, and he arranges "heavy metals" as those metals with basic densities over 7 g cm^{-3} . In 1964, the editorial manager of Van Nostrand's Worldwide Reference book of Concoction Science and in 1987, the editors of Allow and Hackh's Compound Lexicon included metals with a thickness more noteworthy than 4 g cm^{-3} . The fortune of various metals viz. Cr, Ni, Cu, Mn, Hg, Compact disc, Pb and metalloids, including As, Sb, and Se, in the encompassing outskirts is of huge misgiving [6], especially close previous mine locales, dumps, following heaps, and impoundments, yet in addition in urban and mechanical focuses. Cause of cadmium in soil is portrayed as agrarian squanders (20%), ooze (38%), manures (2%) and barometrical aftermaths (40%) [7]. Cadmium is one of the most toxic elements with reported carcinogenic effects in humans [8]. Cadmium and cadmium compounds are, compared to other heavy metals, relatively water soluble and mobile compound in most soils, generally more bio- available and tends to bio- accumulate. It induces cell injury and death by interfering with calcium (Ca) regulation in biological system. Cadmium is not essential for plant or animal life (IPCS monographs). Terrestrial plants may accumulate cadmium in the roots where it is found bound to the cell walls [9]. The pH level is one of the most important factors controlling cadmium absorption. The translocation of cadmium is significant in above ground parts with respect to copper and lead. Concentration in roots represents only 2–5 times that in the above ground parts, but cadmium is transferred only with difficulty to reproductive or storage organs of the plant [10]. The polyamine (PA) putrescine (Put) is an important modulator for the mitigation of diverse of stress in the plants [11–18], and it also plays a significant role in the apoptosis and programmed death in both animals and plants [19]. Chemically the PAs are undersized, +ve charged aliphatic amines at cellular pH values and, consequently, bind opposite charged molecules, viz. nucleic acids, acid phospholipids and proteins, consequently [14, 20]. The cellular level of free amino acids in plants depends on the synthesis and degradation of PAs, their bounding with phenolic acids and intracellular transport [14, 15, 21–23]. The inhibitory effects of Cd are manifestations of oxidative stress, which finally reduces crop productivity. Polyamines are involved in abiotic stress tolerance in plants. Increased polyamines level in stressed plants has adaptive significance because of their involvement in the regulation of cellular ionic environment, maintenance of membrane integrity, prevention of chlorophyll loss and stimulation of protein, nucleic acid and protective alkaloids. Interaction of polyamines with membrane phospholipids implicates membrane stability under stress conditions. Polyamines additionally

shield the layer from oxidative harm as they go about as free radical foragers. Reaction to abiotic damage and mineral supplement insufficiency is related with the generation of conjugated PAs in plants. Polyamine substances are adjusted in light of the introduction to overwhelming metals. These viably balance out and secure the film frameworks against the dangerous impacts of metal particles. The mycorrhiza *Glomus* is the significant inhibitor of mobility of cadmium ion into the soil solution and defends the plants from cadmium toxicity. Colonization of AM fungal was observed in highly contaminated soil [24, 25]. *Glomus mosseae* was reported in heavy metal contaminated sites [26]. External mycelium of *Glomus mosseae* produce a type of protein called Glycoprotein (Glomalin), which has heavy metal binding sites [27]. Cadmium metals accumulate at these binding sites. The antioxidant level is also increased as a result of association of AM fungi with plants [28–30]. Some fungal strains were isolated in the past which were resistant to heavy metal contamination. Mycorrhizal fungi overcome the stress of heavy metal contamination [31, 32].

We are interested in the study of the sorghum plant because of several reasons, which we find more relevant:

[A] The research of the poisoning effect of the heavy metal Cd on plant mainly focuses on food crops such as rice, wheat and maize, but less on sorghum.

[B] Sorghum frequently used as animal feed so quality assurance related to heavy metals is not understood very well, hence studies on contamination of heavy metal in fodder crop is required.

[C] The sensitivity level of sorghum for cadmium varies. By deciding the sensitivity one can find genes which are responsible for the same. Literature indicates that tolerance level of a heavy metal for sorghum is 1000 ppm onwards. It indicates that it has a wide range of tolerance. The genomic size of the sorghum ranges from 700 to 772 Mbp, and it has been well sequenced. So, search for genes responsible for tolerance is not that difficult now. After finding the gene, it can be transferred into the sensitive crops such as cow pea and other leafy vegetables. Finally, we can get transgenic of the heavy metal tolerance. Da-Lin et al. [33] reported some important changes in growth and physiological characteristics of the sorghum plant under cadmium stress such as height, chlorophyll content, root activities and MDA content. They concluded that Cd effect was the manifestation of oxidative stress. Low concentration of cadmium can promote growth of hard wheat, while under a relatively high concentration, the growth of wheat and tillering were both inhibited and the degree varied among the various species. Liu's [34] research also showed that corn seedling's height under cadmium treatment was reduced significantly as the concentration of cadmium increased with prolonged growth period. These studies showed that lower concentrations of Cd stimulated increase of sorghum height, which may be related to the certain resistance of the sorghum genus plant. The higher concentrations hindered their development; explanations behind the restraining impact of substantial metals to plant development were presumably due to: [A] A progression of physical and chemical responses between the overabundance substantial metal and soil parts which changes soil properties, along these lines influencing soil fruitfulness [35, 36]. For instance, substantial metal contamination can improve the obsession of soil phosphorus,

which influenced the plant engrossing phosphorus, consequently influencing the development of plants [37, 38].

[B] Substantial metal caused a lessening in plant photosynthesis, in this way decreasing the plant water and supplement adsorption, which influenced the ordinary development and improvement of sorghum [39].

There are large reports about substantial metal contamination influencing root exercises of Gramineous plants. For instance, through the hydroponic way, Yang et al. [40] found the impact of sewage straightforwardly flooded on root and seedlings of the wheat. The outcomes demonstrated that the worry of sewage flooded quickened the decrease of wheat seedlings and root, diminishing the root number and the root exercises essentially. Jiang et al. [41] additionally demonstrated that tainted soil made the underlying foundations of the rice seedlings yellow and red, extended the rhizome, root shading was dark colored and yellow, while Huang et al. [42] demonstrated that under framework or soil with cadmium the root exercises altogether diminished.

So, our objective in the present study was to find out the oxidative damage induced by cadmium toxicity in sorghum and to assess the role of putrescine and mycorrhiza in the mitigation of cadmium induced stress responses in sorghum.

2. Material and methods

2.1. Selection of crop sorghum (*Sorghum vulgare* L.)

Sorghum is one of the main staple foods for the world's poorest and most food-unsecured people across the semi-arid tropics. Sorghum belongs to the grass family Graminae. It is a short-day C4 plant. The optimum photoperiod which induces flower formation is between 10 and 11 h. Sorghum is one of the main staple foods for the world's poorest and most food-unsecured people across the semi-arid tropics. Globally, sorghum is cultivated on 41 million hectares to produce 64.20 million tons, with productivity having around 1.60 tonnes per hectare ([96], Directorate of Sorghum Research, Hyderabad). With exception in some regions, it is mainly produced and consumed by poor farmers. India contributes about 16% of the world's sorghum production ([96], DSR Hyderabad). It is the fifth most important cereal crop in the country. In India, this crop was one of the major staple cereals during 1950s and occupied an area of more than 18 million hectares, but has come down to 7.69 million hectares ([96], Directorate of Sorghum Research, Hyderabad). Sorghum requires warm conditions, but it can be grown under a wide range of conditions. It is grown from sea level to as high as 1500 m. It can tolerate high temperature throughout its life cycle better than any crop. It can be grown in the areas having an average annual rainfall 60–100 cm. The minimum temperature for the germination of the seed is 7–10°C. It needs about 26–30°C for optimal growth. Sorghum is characterized by a vastly diverse germplasm in terms of phenotypic and morphological traits. Sorghum can be classified into four main groups depending on their production characteristic: grain sorghum, forage sorghum (FS), high-tonnage sorghum (energy) and sweet sorghum.

Sorghum cultivars are now being considered as candidates in the search for efficient energy crops due to an increased interest in the conversion of biomass to energy.

2.2. Experimental site

The present investigation was carried out in the polyhouse and the laboratory of the Department of Plant Physiology, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, U.P., India. Its geographical location lies between 25°18' N latitude to 83°03' E longitudinal and the elevation of the experimental site from the sea level is approximately 75.7 m above the mean sea level.

2.3. Climate condition

Varanasi falls in the Northern India belt of semi-arid to sub humid climate. The normal period of onset of monsoon is the third week of June in this region, which lasts up to the end of September or sometimes third week of October. The normal annual rainfall is about 1100 mm, of which 88% are received from June to September as monsoon season rain, 5–7% in October to December as post monsoon and about 3.3% from January to February as winter season or pre –monsoon rain. The temperature fluctuated in the range of 45–19°C as maximum and 28–7°C as a minimum. The mean relative humidity of the area is about 66%, which rises up to 92% during July to September and falls down to 39% during the end of April to early June.

2.4. Plant and mycorrhizal source

Disease free and healthy, bold seeds of sorghum cv. CSV15 were obtained from the Directorate of Sorghum Research, Hyderabad. The endomycorrhiza *Glomus mosseae* was obtained from the Tata Energetical Research Institute, New Delhi.

2.5. Treatments detail

The pot experiment was conducted in the poly house of the Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, with one genotype of sorghum CSV 15. The pot size for the experiment was in the diameter of 30 and 25 cm in height and each of capacity with 10 kg of soil, with a small hole at the bottom. Pots containing soil mix (Soil + FYM in 3:1) are inoculated with seeds of sorghum. Targeted pots were inoculated with Endomycorrhiza *Glomus sp.* (150 inoculants per kg of soil), after that heavy metal stress was created in the plant by the exogenous application of cadmium nitrate in soil. Two best concentrations of heavy metals on the basis of initial screening were selected i.e., 70 and 150 ppm per kg of soil. Putrescine was applied at the rate of 2.5 and 5.0 mM through foliar spray in 7 day interval, starting from seven DAS up to a week before 90 DAS. The various observations were taken in three stages such as 30, 60 and 90 days after sowing in the concerned pots. The detailed plan of treatments were: Control (T0), Control + Mycorrhiza (T1), Control + 2.5 mM Putrescine (T2), Control +5 mM Putrescine (T3), Control +2.5 mM Putrescine + Mycorrhiza (T4), Control +5 mM Putrescine + Mycorrhiza (T5), 70 ppm Cd(NO₃)₂ (T6), 70 ppm Cd(NO₃)₂ + Mycorrhiza (T7),

70 ppm $\text{Cd}(\text{NO}_3)_2$ + 2.5 mM Putrescine (T8), 70 ppm $\text{Cd}(\text{NO}_3)_2$ + 5 mM Putrescine (T9), 70 ppm $\text{Cd}(\text{NO}_3)_2$ + 2.5 mM Putrescine + Mycorrhiza (T10), 70 ppm $\text{Cd}(\text{NO}_3)_2$ + 5 mM Putrescine + Mycorrhiza (T11), 150 ppm $\text{Cd}(\text{NO}_3)_2$ (T12), 150 ppm $\text{Cd}(\text{NO}_3)_2$ + Mycorrhiza (T13), 150 ppm $\text{Cd}(\text{NO}_3)_2$ + 2.5 mM Putrescine (T14), 150 ppm $\text{Cd}(\text{NO}_3)_2$ + 5 mM Putrescine (T15), 150 ppm $\text{Cd}(\text{NO}_3)_2$ + 2.5 mM Putrescine + Mycorrhiza (T16), 150 ppm $\text{Cd}(\text{NO}_3)_2$ + 5 mM Putrescine + Mycorrhiza (T17).

2.6. Design and layout of experiment

The experiment was laid out in completely randomized design (CRD). There were 18 treatments including control. Each treatment was replicated five times.

2.7. Measurement of oxidative damage

Estimation of Lipid Peroxidation (malondialdehyde (MDA) content) in terms of Thiobarbituric Acid Reducing Substances (TBARS) content was estimated by the method given by Heath and Packer [43]. About 0.5 g of leaf tissues from control and treated groups were cut into small pieces and homogenized by the addition of 5 ml of 5% trichloroacetic acid (TCA) solution. The homogenates were then transferred into fresh tubes and centrifuged at 12,000 rpm for 15 min at room temperature. Equal volumes of supernatant and 0.5% thiobarbituric acid (TBA) in 20% TCA solution were added into a new tube and boiled at 96°C for 25 min. The tubes were transferred into an ice bath and then centrifuged at 10,000 rpm for 5 min. The absorbance of the supernatant was measured at 532 nm and corrected for non-specific turbidity by subtracting the absorbance at 600 nm, 0.5% TBA in 20% TCA solution was used as the blank. The amount of MDA-TBA complex (red pigment) was calculated from the extinction coefficient as $155 \text{ M}^{-1} \text{ cm}^{-1}$. Values of MDA contents were taken from measurements. Results were presented as $\mu\text{moles MDA g}^{-1} \text{ FW}$.

The H_2O_2 content was measured by the method given by Jana and Choudhuri [44]. One hundred mg root samples were extracted using 3 ml of 50 mM sodium phosphate buffer (pH 7.4). The homogenate was centrifuged at 6000 g for 15 min. To determine hydrogen peroxide levels, 3 ml of the extracted solution was mixed with 1 ml of 0.1% titanium sulfate in 20% (w/v) sulfuric acid and then centrifuged at 6000 g for 15 min. The intensity of the yellow color in the supernatant was measured at 410 nm. The hydrogen peroxide level was calculated using extinction coefficient $0.28 \mu\text{M}^{-1} \text{ cm}^{-1}$ and was expressed as $\text{nmol H}_2\text{O}_2 \text{ g}^{-1} \text{ tissue FW}$.

2.8. Statistical analysis

All the numerical data obtained were analyzed through Statistical package of Origin 6.1-advance scientific graphing and data analysis [OriginLab Corporation, One RoundHouse Plaza, Northhampton, MA 01060]. Two-way ANOVA was performed for interaction between mycorrhiza and cadmium treatments. One Way ANOVA was performed for comparing the significance difference among individual means.

3. Results

3.1. Malondialdehyde (MDA) [nmole/ml FW] content

Effect of polyamine (putrescine), mycorrhiza and their combination on MDA [nmole/ml FW] was studied in sorghum variety CSV15 under the cadmium stress. Data were recorded at 30, 60 and 90 days after sowing (DAS) (Figure 1). It is evident that the average MDA was significantly increased by 35.64, 41.39 and 64.02% when exposed to heavy metal stress (T6) as compared to control (T0) at 30, 60 and 90 DAS of the interval. Similarly, when plants were exposed to higher doses of heavy metal (T12) then MDA was significantly increased by 62.28, 59.44 and 73.67% as compared to control (T0) on the dates of proposed interval. Exogenous application of endomycorrhiza in the soil (T7) showed the mitigation effect by reducing the MDA by 6.05, 2.87 and 7.87% as compared to T6 at 30, 60 and 90 DAS. During treatment, T13 was compared to T12, the MDA reduced significantly by 12.92, 9.85 and 9.63% at proposed DAS. In comparison to T6, the exogenous application of putrescine (T8) showed the mitigating effect by decreasing MDA by 1.84, 6.64 and 14.90% on proposed DAS. The average MDA was significantly reduced as compared to T6 by 11.15, 7.24 and 29.68% when treated with high dose of putrescine (T9) with respect to T8. Similarly, when treatment, T14 was compared with T12, the MDA reduced significantly by 21.47, 17.32 and 18.45% at proposed DAS. The average MDA was significantly reduced as compared to T12 by 33.17, 23.72 and 20.39% when treated

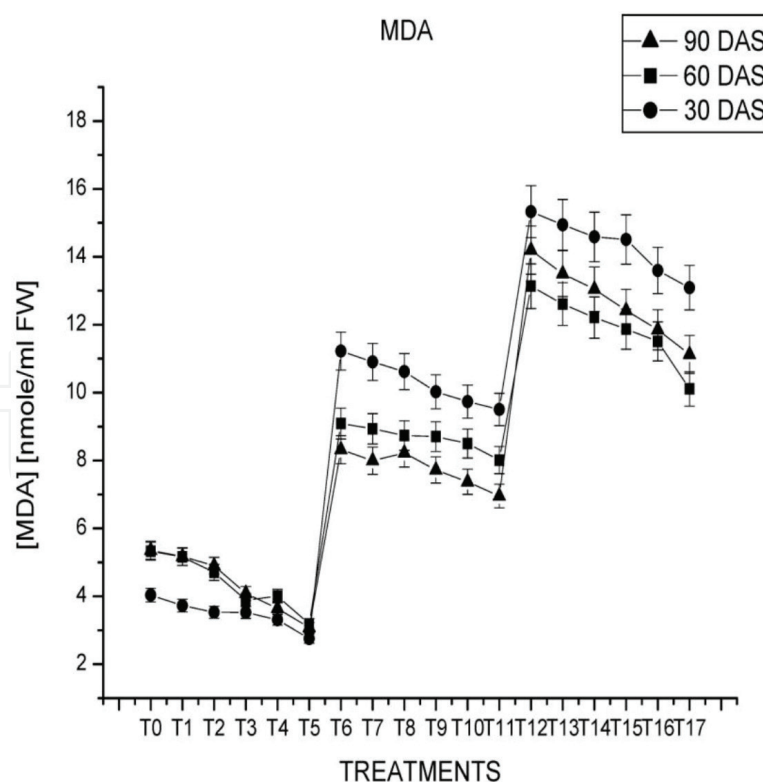


Figure 1. Effects of putrescine and *Glomus* on MDA content in sorghum grown in cadmium contaminated soil.

with high dose of putrescine (T15) with respect to T14. The combination of putrescine and mycorrhiza showed the best mitigation effect in treatment T10 by reducing MDA by 17.65, 11.03 and 36.89% with respect to treatment T6 at proposed DAS. When treatment T11 was compared with treatment T6 then MDA was reduced by 25.51, 20.21 and 42.59%, respectively. A similar effect was seen in the treatment (T16) with respect to treatment T12. In this treatment (T16), the MDA was found to decrease significantly by 43.79, 30.52 and 42.96%, respectively at proposed DAS. The treatment T17 was found to show better results; significant decrease in MDA by 57.32, 56.76 and 55.50% with respect to T12 was observed. So, the combination of putrescine and mycorrhiza showed the best combination for the mitigation of cadmium toxicity for the malondialdehyde.

3.2. H₂O₂ [μ mole/g FW] content

H₂O₂ [μ mole/g FW] content was studied in sorghum variety CSV15 under the cadmium stress. Data were recorded at 30, 60 and 90 days after sowing (DAS) (**Figure 2**). It is evident that the average H₂O₂ was significantly increased by 34.09, 32.02 and 19.37% when exposed to heavy metal stress (T6) as compared to control (T0) at 30, 60 and 90 DAS of the interval. Similarly, when plants were exposed to high doses of heavy metal (T12) then its H₂O₂ was significantly increased by 30.9, 37.55 and 34.66% as compared to control (T0) on the dates of proposed interval. Exogenous application of endomycorrhiza in the soil (T7) showed the mitigation effect by reducing the H₂O₂ by 32.47, 14.21 and 0.54% as compared to T6 at 30, 60 and 90 DAS. During treatment, T13 was compared to T12, the H₂O₂ reduced significantly by 1.13, 2.00 and 1.25% at proposed DAS. In comparison to T6, the exogenous application of

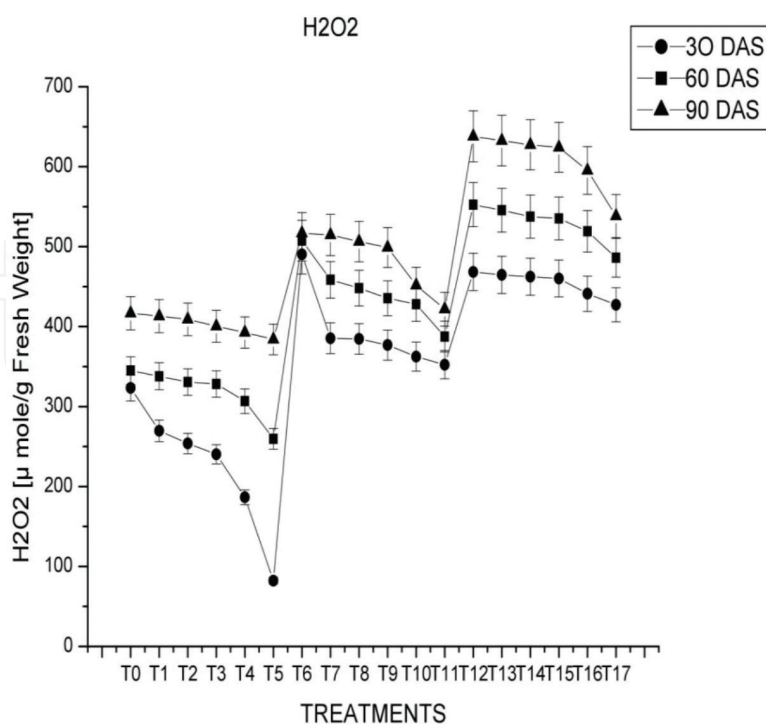


Figure 2. Effects of putrescine and *Glomus* on H₂O₂ content in sorghum grown in cadmium contaminated soil.

putrescine (T8) showed the mitigating effect by decreasing H_2O_2 by 32.74, 17.24 and 2.52% on proposed DAS. The average H_2O_2 was significantly reduced as compared to T6 by 35.10, 20.87 and 4.30% when treated with high dose of putrescine (T9) with respect to T8. Similarly, when treatment, T14 was compared with T12, the H_2O_2 reduced significantly by 1.82, 4.33 and 2.50% at proposed DAS. The average H_2O_2 was significantly reduced as compared to T12 by 2.5, 4.98 and 3.28% when treated with high dose of putrescine (T15) with respect to T14. The combination of putrescine and mycorrhiza showed the best mitigation effect in treatment T10 by reducing H_2O_2 by 17.65, 11.03 and 36.89% with respect to treatment T6 at proposed DAS. When treatment T11 was compared with treatment T6 then significant H_2O_2 was reduced by 42.70, 34.80 and 22.80%, respectively. A similar effect was seen in the treatment (T16) with respect to treatment T12 where H_2O_2 was found to decrease significantly by 8.41, 9.66 and 10.21%, respectively at proposed DAS. The treatment T17 was found to show better results; significant decrease in H_2O_2 by 12.70, 19.24 and 23.89% with respect to T12 was observed. So, the combination of putrescine and mycorrhiza showed the best combination for the mitigation of cadmium toxicity for the hydrogen peroxide.

4. Discussion

The ROS is conceivably unsafe to cell layers, bringing about oxidative corruption of film lipids (lipid peroxidation). Malondialdehyde (MDA) is one of the end breakdown results of lipid peroxidation and can be utilized as a marker of in vivo lipid peroxidation. Cadmium uptake triggered oxidative stress resulting in increased generation of Hydrogen peroxide and lipid peroxidation as reported in earlier studies [45, 46]. To cope with enhanced levels of oxidative stress, plants are equipped with antioxidant system that gets activated under cadmium stress [46, 47]. Ascorbate peroxidase, Peroxidase and Catalase play a crucial role during the degradation of the plant sample grown under cadmium stress. The statistical analysis in the treatment T17 showed significant reduction in MDA content because the most important property of PA conjugates with phenolic acids were their antioxidant characteristic, required by the plants to adapt under stress condition. Bors et al. [48] showed that conjugates with caffeic, cinnamic and ferulic acids displayed higher constraint of binding to reactive oxygen species (ROS) i.e., free polyamines are less efficient radical scavengers than their conjugates. Polyamines conjugation by transglutaminases, especially to Rubisco seems to have a significant role in protecting this protein from protease action, thus protecting its photosynthetic efficiency [49]. Therefore, PAs are likely to play a role in photosynthesis since they are capable of reversing stress-induced damage in the photosynthetic apparatus. Cross-linking of PAs mediated by transglutaminases might play a significant role in polyamine bioactivity for flower development and compatibility in reproduction [50]. Thus, conjugates involved in defense mechanisms against cadmium toxicity and also mediate the regulation of certain growth and developmental events [51, 52]. Similarly, the mycorrhiza *Glomus* acted as the potential inhibitor of mobility of cadmium ion into the soil solution and protected the plants from cadmium toxicity. AM fungal colonization was observed in highly contaminated soil [24, 25]. The presence of *Glomus mosseae* was also reported in heavy metal contaminated sites by some researchers [26]. The external mycelium of certain AM fungi produce a type

of protein called Glycoprotein (Glomalin), which has heavy metal binding sites [53]. Heavy metals accumulate at these binding sites. The antioxidant level is also increased as a result of association of AM fungi with plants [28–30]. Some fungal strains were isolated in the past which were resistant to heavy metal contamination. Many mycorrhizal fungi overcome the stress of heavy metal contamination [31, 32].

In the present study, when plants were subjected to cadmium stress, lipid peroxidation was lesser in mycorrhizal treated plant than the non mycorrhizal plants, in severity of cadmium stress. The similar trend was shown when the plant was treated with putrescence. The best results were obtained when mycorrhiza and putrescine were treated together for alleviation of Cd toxicity in sorghum. Mycorrhizal plants along with putrescine displayed lower lipid peroxidation and low hydrogen peroxide production [40, 54–56]. It is widely accepted that diminishing levels are one of the mechanisms by which AM protects plants against diverse stresses [57–59]. It has been suggested that peroxidase could act as an efficient H_2O_2 scavenging system in plant vacuoles in the presence of phenolics and reduced ascorbate [60]. Direct chelation, or binding to polyphenols, was observed with methanol extracts of rhizome polyphenols from *Nymphaea* for Cr, Pb and Hg [61–96]. Phenolic antioxidants inhibit lipid peroxidation by trapping the lipid alkoxyl radical. This activity depends on the structure of the molecules, and the number and position of the hydroxyl group in the molecules. Santiago et al. [62] hypothesize a cycle where H_2O_2 is scavenged by phenolics through a peroxidase, and phenolics are oxidized to phenoxyl radicals, which can be reduced by amino acids [97–118].

The present perceptions demonstrating a positive connection between metal danger and proline aggregation recommend a defensive part of this amino corrosive against substantial metal poisonous quality. In this specific situation, recommendations have been made that proline gives assurance by keeping up the water adjust, which is frequently exasperates by substantial metals [63, 64], searching hydroxyl radicals [65], chelating overwhelming metals in the cytoplasm [66–68] and lessening metal take-up [69]. Numerous scientists, all things considered, feel that proline gathering is just a side effect of assorted anxieties, and isn't engaged with assurance against metal and different burdens [70–72]. Wu et al. [73] demonstrated that Cu-prompted efflux of K^+ in *Anacystis nidulans* was limited within the sight of proline, proposing that proline conceivably shielded the plasma layer from Cu lethality, or it maybe complexed Cu + in this manner diminishing the centralization of free Cu + in the outer condition. In a consequent paper, Wu et al. [69] exhibited decreased Cu take-up by cells containing high convergences of proline, proposing that proline balanced out the plasma film and furthermore diminished the take-up of Cu by algal cells. The impacts of lead, copper, cadmium and mercury on the substance of proline, were researched in 17-day-old bean seedlings (*Phaseolus vulgaris* L.) developed in Hoagland arrangement spiked with different centralizations of Pb, Cu, Album and Hg. Control and substantial metal-treated plants were developed for 10 days in Hoagland arrangement. A noteworthy increment of proline was identified in essential leaves following ten-day presentation to substantial metals. The most grounded impact on proline, content was found in plants presented to mercury, trailed by the grouping $Cd^{++} > Cu^{++} > Pb^{++}$ [74].

Pioneer studies suggested that peroxidase is not only one of the defense proteins, but an important antioxidant enzyme involved in response to environmental stresses [75]. It constitutes a wide variety of heme containing enzymes involved in many physiological processes in plants. It was reported to be involved in response to biotic and abiotic stresses, auxin catabolism, cell wall lignification and degradation [76]. Peroxidase is involved in the scavenging of Reactive Oxygen Species (ROS). High concentration of cadmium in sorghum resulted in inhibition of several enzymes and an increase in activity of others. Cadmium accumulation in the cellular compartment of the enzyme is a pre-requisite for enzyme inhibition *in vivo* [77, 78]. Peroxidase induction was likely to be related to oxidative reaction at the biomembrane. Metal induced enzymatic changes in plants such as isoperoxidase pattern was used to evaluate the phytotoxicity of metal polluted soils. There is extensive literature indicating the role of peroxidase in developmental processes, possibly due to the involvement of this enzyme in auxin metabolism (auxin oxidation) and in the formation of cross-links between cell wall components. Peroxidase activity was more pronounced in cadmium treated sorghum [79–81]. This high peroxidase activity enables the plants to protect themselves against the oxidative stresses as suggested by Scalet et al. [82]. In fact, in plants, peroxidase protects cell against harmful concentration of hydroperoxides by induction of peroxidases especially anionic peroxidase, which have been found to be involved in response to both abiotic and biotic stresses. Mycorrhiza showed best mitigation against cadmium toxicity with respect to peroxidase activity. In the present study, the activity of peroxidase was significantly reduced due to the presence of *Glomus*, because in soil solution the cadmium ion was trapped by the fungus and its mobility and translocation in the plants was inhibited [83]. The translocation of cadmium depends on several factors. The bioavailability of cadmium to sorghum roots from the soil solution depends on its concentration in soil and it modulated by the presence of organic matter, pH and temperature. It is also affected by the presence of chelating organisms like mycorrhiza. Despite different mobility of metal ions in plants, the cadmium content was generally greater in roots than in the above ground tissue [84–86]. In most environmental conditions, Cd enters first the roots, and consequently they are likely to experience Cd damage first [87]. Similarly, the putrescine is another potential ameliorative agent of heavy metal toxicity in sorghum because of its unique nature. Several studies have shown that PA accumulation occurs under abiotic stresses including drought, salinity, low temperature, heavy metals, and PQ [21, 84, 88–90].

5. Conclusions

Finally, it is concluded that the polyamines like putrescine and mycorrhiza *Glomus* impart significant mitigation of cadmium induced toxicity in sorghum mediated through the defensive role of enzymatic and non enzymatic antioxidants in plants. The MDA and hydrogen peroxide content was found significant increased with cadmium treated plants with respect to plants treated with putrescine and mycorrhiza.

Acknowledgements

Authors are thankful to University Grants Commission for providing Senior Research Fellowship to the first author.

Author details

Prasann Kumar^{1*} and Shweta Pathak²

*Address all correspondence to: prasann0659@gmail.com

1 Department of Agronomy, School of Agriculture, Lovely Professional University, Jalandhar, India

2 Department of Plant Physiology, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, UP, India

References

- [1] Benavides MP, Gallego SM, Tomaro ML. Cadmium toxicity in plants. *Brazilian Journal of Plant Physiology*. 2005;**17**:21-34. DOI: 10.1590/S1677-04202005000100003
- [2] Uraaguchi S, Mori S, Kuramata M, Kawasaki A, Arora T, Ishikawa S. Root-to shoot Cd translocation via the xylem is the major process determining shoot and grain cadmium accumulation in rice. *Journal of Experimental Botany*. 2009;**60**:2677-2268. DOI: 10.1093/jxb/erp119
- [3] Wang L, Zhou QX, Ding LL, Sun Y. Effect of cadmium toxicity on nitrogen metabolism in leaves of *Solanum nigrum* L. as a newly found cadmium hyperaccumulator. *Hazardous Materials*. 2008;**154**(1-3):818-825. DOI: 10.1016/j.jhazmat.2007.10.097
- [4] Smith CJ, Hopmans P, Cook FJ. Accumulation of Cr, Pb, Cu, Ni, Zn and Cd in soil following irrigation with untreated urban effluents in Australia. *Environmental Pollution*. 1996;**94**(3):317-323. PMID:15093492
- [5] Bjerrum J. Metal amine formation in aqueous solution. In: Elving J, editor. *Treatise on Analytical Chemistry*. Vol. I. New York: The Interscience Encyclopedia; 1959
- [6] Adriano DC. Chromium. In: *Trace Elements in the Terrestrial Environment*. New York: Springer; 1986. pp. 58-76
- [7] Juste C, Mench M. Long term application of sewage sludge and its effect on metal uptake by crops. In: Adriano DC, editor. *Biogeochemistry of Trace Metals*. Ann. Arbor, London, Tokyo: Leuwis publishers; 1992. pp. 159-193

- [8] Goering PL, Waalkes MP, Klaassen CD. Toxicity of cadmium. In: Goyer RAC, Herian MG, editors. Handbook of Experimental Pharmacology: Toxicity of Metals, Biochemical Effects. New York: Springer Verlag; 1994. pp. 189-213
- [9] AMAP. AMAP Assessment. Heavy Metals in the Arctic-Pre-Print Files. Oslo, Norway: Arctic Monitoring and Assessment Programme; 2002. p. 870
- [10] Mench M, Amans V, Sappin-Didier V, Fargues S, Gomez A, Loffler M, Masson P, Arrouays D. A study of additives to reduce availability of Pb in soils to plants. In: Iskandar A, Adriano DC, editors. Remediation of Soils Contaminated with Metals. Northwood, UK: Science Reviews; 1997. pp. 202-185
- [11] Baron K, Stasolla C. The role of polyamines during in vivo and in vitro development. In Vitro Cellular & Developmental Biology. Plant. 2008;**44**:384-395. DOI: 10.1007/s11627-008-9176-4
- [12] Dossantos RW, Schmidt EC, Depmartins R, Latini A, Maraschin M, Horta PA, Bouzon ZL. Effects of cadmium on growth, photosynthetic pigments, photosynthetic performance, biochemical parameters and structure of chloroplasts in the agarophyte *Gracilaria domingensis* (Rhodophyta, Gracilariales). American Journal of Plant Sciences. 2012;**3**:1077-1084. DOI: 10.4236/ajps.2012.38129
- [13] Kakkar RK, Sawhney VK. Polyamine research in plants—a changing perspective. Physiologia Plantarum. 2002;**116**:281-292. DOI: 10.1034/j.1399-3054.2002.1160302.x
- [14] Kuznetsov V, Radyukina NL, Shevyakova NI. Polyamines and stress: Biological role, metabolism and regulation. Russian Journal of Plant Physiology. 2006;**53**:583-604. DOI: 10.1134/S1021443706050025
- [15] Kusano T, Berberich T, Tateda C, Takahashi Y. Polyamines: Essential factors for growth and survival. Planta. 2008;**228**:367-381. DOI: 10.1007/s00425-008-0772-7
- [16] Steiner N, Santa-Catarina C, Silveira V, Floh EIS, Guerra MP. Polyamine effects on growth and endogenous hormones levels in *Araucaria angustifolia* embryogenic cultures. Plant Cell, Tissue and Organ Culture. 2007;**89**:55-62. DOI: 10.1007/s11240-007-9216-5
- [17] Santa-Catarina C, Silveira V, Scherer GFE, Floh EIS. Polyamine and nitric oxide levels correlate with morphogenetic evolution in somatic embryogenesis of *Ocotea catharinensis*. Plant Cell, Tissue and Organ Culture. 2007;**90**:93-101. DOI: 10.1007/s11240-007-9259-7
- [18] Tun NN, Santa-Catarina C, Beghum T, Silveira V, Handro W, Floh EIS, Scherer GFE. Polyamines induce the rapid biosynthesis of nitric oxide (NO) in *Arabidopsis thaliana* seedlings. Plant & Cell Physiology. 2006;**47**:346-354. DOI: 10.1093/pcp/pci252
- [19] Kuhen GD, Phillips GC. Role of polyamines in apoptosis and other recent advances in plant polyamines. Critical Reviews in Plant Sciences. 2005;**24**:123-130. DOI: 10.1080/07352680590953161

- [20] Yoda H, Hamaguchi R, Sano H. Induction of hypersensitive cell death by hydrogen peroxide produced through polyamine degradation in tobacco plants. *Plant Physiology*. 2003;**132**:1973-1981. DOI: 10.1104/pp.103.024737
- [21] Bouchereau A, Aziz A, Larher F, Martin-Tanguy J. Polyamines and environmental challenges: Recent development. *Plant Science*. 1999;**140**:103-125. PMCID: PMC2835953
- [22] Bhatnagar P, Minocha R, Minocha SC. Genetic manipulation of the metabolism of polyamines in poplar cells: The regulation of putrescine catabolism. *Plant Physiology*. 2002;**128**:1455-1469. DOI: 10.1104/pp.010792
- [23] Minocha SC, Minocha R. Role of polyamines in somatic embryogenesis. In: Bajaj YPS, editor. *Biotechnology in Agriculture and Forestry*. Vol. 30. Berlin: Springer-Verlag; 1995. pp. 53-70. DOI: 10.12691/plant-1-2-1
- [24] Leung HM, Ye ZH, Wong MH. Survival strategies of plants associated with arbuscular mycorrhizal fungi on toxic mine tailings. *Chemosphere*. 2007;**66**:905-915. DOI: 10.1016/j.chemosphere.2006.06.037
- [25] Pawlowska TE, Blaszkowski J, Ruhling A. The mycorrhizal status of plants colonizing a calamine spoil mound in southern Poland. *Mycorrhiza*. 1996;**6**:499-505. DOI: 10.1007/s005720050154
- [26] Debiane D, Garcon G, Verdin A, Fontaine J, Durand R, Grandmougin-Ferjani A, Shirali P, Lounces- Hadj Sahraoui A. In vitro evaluation of the oxidative stress and genotoxic potentials of anthracene on mycorrhizal chicory roots. *Environmental and Experimental Botany*. 2008;**64**:120-127. DOI: 10.1016/j.envexpbot.2008.04.003
- [27] Vivas A, Azcon R, Biro B, Barea JM, Ruiz Lozano JM. Influence of bacterial strains isolated from lead polluted soil and their interactions with arbuscular mycorrhizae on the growth of *Trifolium pertense* L. under lead toxicity. *Canadian Journal of Microbiology*. 2003a;**49**:577-588. DOI: 10.1139/w03-073
- [28] Gopi R, Jaleel CA, Sairam R, Lakshmanam GMA, Gomathinayagam M, Paneerselvam R. Differential effects of hexaconazole and paclobutrazol on biomass, electrolyte leakage, lipid peroxidation and antioxidant potential of *Daucus carota* L. *Colloids and Surfaces Biointerfaces*. 2007;**60**:180-186. DOI: 10.1016/j.colsurfb.2007.06.003
- [29] Zhang FQ, Wang YS, Lou ZP, Dong JD. Effect of heavy metal stress on antioxidative enzymes and lipid peroxidation in leaves and roots of two mangrove plant seedlings (*Kandelia candel* and *Bruguiera gymnorrhiza*). *Chemosphere*. 2007a;**67**:44-50. DOI: 10.1016/j.chemosphere.2006.10.007
- [30] Zhang LZ, Wei N, Wu QX, Ping ML. Antioxidant response of *Cucumis sativus* L. to fungicide carbendazim. *Pesticide Biochemistry and Physiology*. 2007b;**89**:54-59. DOI: 10.1016/j.pestbp.2007.02.007
- [31] Hildebrandt U, Regvar M, Bothe H. Arbuscular mycorrhiza and heavy metal tolerance. *Phytochemistry*. 2007;**68**:139-146. DOI: 10.1016/j.phytochem.2006.09.023

- [32] Joschim HJ, Makoi R, Ndakidemi PA. The agronomic potential of vesicular-arbuscular mycorrhiza (AM) in cereals-legume mixtures in Africa. *African Journal of Microbiology Research*. 2009;**11**:664-675
- [33] Da-Lin L, Shu-pan Z, Zheng C, Wei-wei Q. Soil cadmium regulates antioxidases in sorghum. *Agricultural Sciences in China*. 2011;**9**(10):1475-1480
- [34] Liu JX. Effects of cadmium and zinc interaction on corn seedlings physiological and biochemical characteristics. *Journal of Yichun College*. 2004;**26**(6):55-57. DOI: 10.5897/AJB11.848
- [35] Cieslinski G, Neilser GH, Hogue EJ. Effect of soil cadmium application and pH on growth and cadmium accumulation in roots, leaves and fruit of strawberry plants. *Plant and Soil*. 1996;**18**:267-271. DOI: 10.1080/01904169409364791.
- [36] Chang ZM, Wu XH. Difference comparison of three alfalfa varieties resistant to cadmium pollution. *Pratac. Sci.* 2005;**22**(2):20-23
- [37] Li J, Gao XH, Guo SR, Zhang RH, Wang X. Effects of exogenous spermidine on photosynthesis of salt-stressed *Cuellaria sativa* seedlings. *Chinese Journal of Ecology*. 2007;**26**(10):1595-1599. DOI: 10.1007/s10265-014-0653-z
- [38] Zhang EH, Zhang XH, Wang HZ. Adaptation effects of phosphorus stress on different genotypes of faba-bean. *Acta Ecologica Sinica*. 2004;**24**(8):1589-1593
- [39] Qin TC, Ruan J, Wang LJ. Effects of cadmium on plant photosynthesis. *Environmental Science and Technology*. 2000;**13**:33-35. DOI: 10.5897/AJB11.848
- [40] Yang Y, Han X, Liang Y, Ghosh A, Chen J, Tang M. The combined effects of arbuscular mycorrhizal fungi (AMF) and lead (Pb) stress on Pb accumulation, plant growth parameters, photosynthesis, and antioxidant enzymes in *Robinia pseudoacacia* L. *PLoS One*. 2015;**10**:e0145726. DOI: 10.1371/journal.pone.0145726
- [41] Jiang Y, Liang WJ, Zhang YG, Xu YF. Research on effect of sewage irrigation on soil heavy metal environmental capacity and rice growth. *Chinese Journal of Eco-Agriculture*. 2004;**12**(3):124-127
- [42] Huang CC, Chen MW, Hsieh JL, Lin WH, Chen PC, Chien LF. Expression of mercuric reductase from *Bacillus megaterium* MB1 in eukaryotic microalga *Chlorella* sp. *Applied Microbiology and Biotechnology*. 2006;**72**:197-205. DOI: 10.1007/s00253-005-0250-0
- [43] Heath RL, Packer L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Archives of Biochemistry and Biophysics*. 1968;**125**:189-198. DOI: 10.1016/0003-9861(68)90654-1
- [44] Jana S, Choudhuri MA. Glycolate metabolism of three submerged aquatic angiosperms during aging. *Aquatic Botany*. 1981;**12**:345-354
- [45] Liu J, Macarasin D, Wisniewski M, Sui Y, Droby S, Norelli J, et al. Production of hydrogen peroxide and expression of ROS-generating genes in peach flower petals in

- response to host and non-host fungal pathogens. *Plant Pathology*. 2013;**62**:820-828. DOI: 10.1111/j.1365-3059.2012.02683
- [46] Singh HP, Batish DR, Kohli RK, Arora K. Arsenic-induced root growth inhibition in mung bean (*Phaseolus aureus* Roxb.) is due to oxidative stress resulting from enhanced lipid peroxidation. *Plant Growth Regulation*. 2007;**53**:65-73. DOI: 10.1007/s10725-007-9205
- [47] Sobrino-Plata J, Meyssen D, Cuypers A, Escobar C, Hernández LE. Glutathione is a key antioxidant metabolite to cope with mercury and cadmium stress. *Plant and Soil*. 2014;**377**:369-381. DOI: 10.1007/s11104-013-2006-4
- [48] Bors W, Langebartels C, Michel C, Sandermann H. Polyamines as radical scavengers and protectants against ozone damage. *Phytochemistry*. 1989;**28**:1589-1595
- [49] Serafini-Fracassini D, Del Duca S, Beninati S. Plant transglutaminases. *Phytochemistry*. 1995;**40**:355-365
- [50] Serafini-Fracassini D, Del Duca S. Transglutaminases: Widespread cross-linking enzymes in plants. *Annals of Botany*. 2008;**102**:145-152. DOI: 10.1093/aob/mcn075
- [51] Bagni N, Tassoni A. Biosynthesis, oxidation and conjugation of aliphatic polyamines in higher plants. *Amino Acids*. 2001;**20**:301-317. PMID:11354606
- [52] Martin-Tanguy J. Metabolism and function of polyamines in plants: Recent development (new approaches). *Plant Growth Regulation*. 2001;**34**:135-148
- [53] Vivas A, Voros I, Biro B, Campos E, Barea JM, Azcon R. Symbiotic efficiency of autochthonous arbuscular *Mycorrhizal fungus* (*G. mosseae*) and *Brevibacillus brevis* isolated from cadmium polluted soil under increasing cadmium levels. *Environmental Pollution*. 2003b;**126**:179-189. DOI: 10.1016/S0269-7491(03)00195-7
- [54] Evelin H, Kapoor R. Arbuscular mycorrhizal symbiosis modulates antioxidant response in salt-stressed *Trigonella foenum-graecum* plants. *Mycorrhiza*. 2014;**24**:197-208. DOI: 10.1007/s00572-0130529-4
- [55] Jiang QY, Zhuo F, Long SH, Zhao HD, Yang DJ, Ye ZH, et al. Can arbuscular mycorrhizal fungi reduce Cd uptake and alleviate cd toxicity of *Lonicera japonica* grown in Cd-added soils? *Scientific Reports*. 2016;**6**:21805. DOI: 10.1038/srep21805
- [56] Tan SY, Jiang QY, Zhuo F, Liu H, Wang YT, Li SS, et al. Effect of inoculation with *Glomus versiforme* on cadmium accumulation, antioxidant activities and phytochelatin of *Solanum 977 photeinocarpum*. *PLoS One*. 2015;**10**:e0132347. DOI: 10.1371/journal.pone.0132347
- [57] Garg N, Singla P. The role of *Glomus mosseae* on key physiological and biochemical parameters of pea plants grown in arsenic contaminated soil. *Scientia Horticulturae*. 2012;**143**:92-101. DOI: 10.1016/j.scienta.2012.06.010
- [58] Hajiboland R, Aliasgharzadeh N, Laiegh SF, Poschenrieder C. Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum lycopersicum* L.) plants. *Plant and Soil*. 2010;**331**:313-327. DOI: 10.1007/s11104-009-0255-z

- [59] Ruiz-Sánchez M, Aroca R, Muñoz Y, Polón R, Ruiz-Lozano JM. The arbuscular mycorrhizal symbiosis enhances the photosynthetic efficiency and the antioxidative response of rice plants subjected to drought stress. *Journal of Plant Physiology*. 2010;**167**:862-869. DOI: 10.1016/j.jplph.2010.01.018
- [60] Zancani M, Nagy G. Phenol-dependent H₂O₂ breakdown by soybean root plasma membrane-bound peroxidase is regulated by ascorbate and thiols. *Journal of Plant Physiology*. 2000;**156**:295-299. DOI: 10.1016/S0176-1617(00)80064-4
- [61] Lavid N, Schwartz A, Yarden O, TelOr E. The involvement of phenylpropanoids and peroxidase activities in heavy metal accumulation by epidermal glands of the waterlily (Nymphaeaceae). *Planta*. 2001;**212**:323-331
- [62] Santiago LJM, Louro RP, De Oliveira DE. Compartmentation of phenolic compounds and phenylalanine ammonia-lyase in leaves of *Phyllanthus tenellus* Roxb. And their induction by copper sulphate. *Annals of Botany*. 2000;**86**:1023-1032. DOI: 10.1006/anbo.2000.1271
- [63] Costa G, Morel JL. Water relations gas exchange and amino acid content in cd treated lettuce. *Plant Physiology and Biochemistry*. 1994;**32**:561-570
- [64] Schat H, Sharma SS, Vooijst R. Heavy metal-induced accumulation of free proline in a metal-tolerant and nontolerant ecotype of *Silene vulgaris*. *Physiologia Plantarum*. 1997;**101**:477-482. DOI: 10.1034/j. 1399-3054.1997.1010304. x
- [65] Smirnoff N, Cumber QJ. Hydroxyl radical scavenging activity of compatible solutes. *Phytochemistry*. 1989;**28**:1057-1060. DOI: 10.1016/0031-9422(89)80182-7
- [66] Farago ME, Mullen WA. Plants which accumulate metals. A possible copper-proline complex from the roots of *Armeria maritima*. *Inorganica Chimica Acta*. 1979;**32**:L93-L94. DOI: 10.1016/0031-9422(89)80182-7
- [67] Hemalatha S, Anburaj A, Francis K. Effect of heavy metals on certain biochemical constituents and nitrate reductase activity in *Orzya sativa* L. seedlings. *Journal of Environmental Biology*. 1997;**18**:313-319. DOI: 10.2478/eko-2014-0012
- [68] Pandey P, Tripathi AK. Effect of heavy metals on morphological and biochemical characteristics of *Albizia procera* (Roxb. Benth.) seedling. *International Journal of Environmental Sciences*. 2011;**1**(5):1009-1018. DOI: 10.2478/eko-2014-0012
- [69] Wu JT, Hsieh MT, Kow LC. Role of proline accumulation in response to toxic copper in *Chlorella* sp. (Chlorophyceae) cells. *The Journal of Physiology*. 1998;**34**:113-117. DOI: 10.1046/j.1529-8817.1998.340113.x
- [70] Bhaskaran S, Smith RH, Newton RJ. Physiological changes in cultured sorghum cells in response to induced water stress. I. Free proline. *Plant Physiology*. 1985;**79**:266-269. PMID: PMC1075388
- [71] Lutts S, Kinet JM, Bouharmont J. Effects of various salts and of mannitol on ion and proline accumulation in relation to osmotic adjustment in rice (*Oryza sativa* L.) callus cultures. *Journal of Plant Physiology*. 1996;**149**:186-195. DOI: 10.1016/S0176-1617(96)80193-3

- [72] Perez-Alfocea F, Santa-Cruz A, Guerrier G, Bolarin MC. NaCl stress-induced organic solute changes on leaves and calli of *Lycopersicon esculentum*, *L. pinnellii* and their inter-specific hybrid. *Journal of Plant Physiology*. 1994;**143**:106-111
- [73] Wu JT, Chang SC, Chen KS. Enhancement of intracellular proline level in cells of *Anacystis nidulans* (cyanobacteria) exposed to deleterious concentrations of copper. *Journal of Phycology*. 1995;**31**:376-379. DOI: 10.1111/j.0022-3646.1995.00376.x
- [74] Rodriguez-Serrano M, RomeroPuertas MC, Pazmino DM, Testillano PS, Risueno MC, Del Rio LA, et al. Cellular response of pea plants to cadmium toxicity: Cross talk between reactive oxygen species, nitric oxide, and calcium. *Plant Physiology*. 2009;**150**:229-243. DOI: 10.1104/pp.108.131524
- [75] Flohé L, Ursini F. Peroxidase: A term of many meanings. *Antioxidants & Redox Signaling*. 2008;**10**:1485-1490. DOI: 10.1089/ars.2008.2059
- [76] Wang LK, Hung YT, Shammas NK. *Physicochemical Treatment Processes*. Vol. 3. New Jersey: Humana Press; 2004, 2004. pp. 141-198
- [77] Rabie MH, Eleiwa ME, Aboseoud MA, Khalil KM. Effect of nickel on the content of carbohydrate and some mineral in corn and broad bean plant. *Journal of King Saud University – Science*. 1992;**4**:37
- [78] Sharma SS, Dietz KJ. The significance of amino acids and amino acid-derived molecules in plant responses and adaptation to heavy metal stress. *Journal of Experimental Botany*. 2006;**57**(4):711-726. DOI: 10.1093/jxb/erj073
- [79] Ahmad P, Sharma S, Srivastava PS. In vitro selection of NaHCO₃ tolerant cultivars of *Morus alba* (Local and Sujanpuri) in response to morphological and biochemical parameters. *Horticultural Science (Prague)*. 2007;**34**:114-122
- [80] Chugh LK, Sawhney SK. Photosynthetic activities of *Pisum sativum* seedlings grown in the presence of cadmium. *Plant Physiology and Biochemistry*. 1999;**37**:297-303. DOI: 10.1016/S0981-9428(99)80028-X
- [81] Deniz B, Merve H, Sermin E. Evaluation of Lead Removal onto Black Cumin by Using Multi Linear Regression. *Ohrid, Republic of Macedonia: BALWOIS 2012*; 2012. pp. 1-6
- [82] Scalet M, Federico R, Guido MC, Manes F. Peroxidase activity and polyamines changes in response to ozone and simulated acidrain in Aleppo pine needles. *Environmental and Experimental Botany*. 1995;**35**:417-425. DOI: 10.1016/0098-8472(95)00001-3
- [83] Ernst WHO, Verkleij JAC, Schat H. Metal tolerance in plants. *Acta Botanica Neerlandica*. 1992;**41**:229-248
- [84] Jentschke G, Godbold DL. Metal toxicity and ectomycorrhizas. *Physiologia Plantarum*. 2000;**109**:107-116
- [85] Leyval C, Turnau K, Haselwandter K. Effect of heavy metal pollution on mycorrhizal colonization and function: Physiological, ecological and applied aspects. *Mycorrhiza*. 1997;**7**:139-153. DOI: 10.1007/s005720050174

- [86] Schützendübel A, Polle A. Plant responses to abiotic stresses: Heavy metal-induced oxidative stress and protection by mycorrhization. *Journal of Experimental Botany*. 2002;**53**:1351-1365. DOI: 10.1093/jexbot/53.372.1351
- [87] Santia di Toppi L, Gabbrielli R. Response to cadmium in higher plants. *Environmental and Experimental Botany*. 1999;**41**:105-130 PII: S0098-8472(98)00058-6
- [88] Groppa MD, Tomaro ML, Benavides MP. Polyamines as protectors against cadmium or copper induced oxidative damage in sunflower leaf discs. *Plant Science*. 2001;**161**:481-488. DOI: 10.1007/s00726-006-0343-9
- [89] Marschner H. *Mineral Nutrition of Higher Plants*. 2nd ed. London: Academic Press; 1995. 889 pp
- [90] Pang XM, Zhang ZY, Wen XP, Ban Y, Moriguchi T. Polyamine, all-purpose players in response to environment stresses in plants. *Plant Stress*. 2007;**1**:173-188
- [91] Aebi HE. Catalase. In: Bergmeyer HU, editor. *Methods of Enzymatic Analysis*. 3rd ed. Weinheim, Florida: Verlag Chemie; 1983. pp. 273-286
- [92] Asada K. The water –water cycle in chloroplast: Scavenging of active oxygen and dissipation of excess photons. *Annual Review of Plant Physiology and Plant Molecular Biology*. 1999;**50**:601-639. DOI: 10.1146/annurev.arplant.50.1.601
- [93] Abdul Jaleel C, Jayakumar K, Chang-Xing Z, Iqbal M. Low concentration of cobalt increases growth, biochemical constituents, mineral status and yield in *Zea mays*. *Journal of Scientific Research*. 2009;**1**:128-137. DOI: 10.3329/jsr.v1i1.1226
- [94] Bates L, Waldren RP, Teare ID. Rapid determination of free proline for water-stress studies. *Plant and Soil*. 1973;**39**:205-207. DOI: 10.1007/BF00018060
- [95] Cho UH, Seo NH. Oxidative stress in *Arabidopsis thaliana* exposed to cadmium is due to hydrogen peroxide accumulation. *Plant Science*. 2005;**168**:113-120. DOI: 10.1016/j.plantsci.2004.07.021
- [96] DSR Annual Report (2011). Directorate of Sorghum Research, Hyderabad; 2010-11
- [97] El-Sayed E, Omran, Afaf A, Abd ER. Mapping and screening risk assessment of heavy metals concentrations in soils of the Bahr El-Baker region, Egypt. *Journal of Soil Science and Environmental Management*. 2010;**6**(7):182-195. DOI: 10.5897/JSSEM12.010.
- [98] Gonzalez-Chavez MC, Carrillo-Gonzalez R, Wright SF, Nichols KA. The role of glomalin, a protein produced by arbuscular mycorrhizal fungi, in sequestering potentially toxic elements. *Environmental Pollution*. 2004;**130**:317-323. DOI: 10.1016/j.envpol.2004.01.004
- [99] Gohre V, Paszkowski U. Contribution of arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. *Planta*. 2006;**223**:1115-1122. DOI: 10.1007/s00425-006-0225-0
- [100] Garg N, Manchanda G. ROS generation in plants: Boon or bane? *Plant Biosystems*. 2009;**143**(1):81-96. DOI: 10.1080/11263500802633626

- [101] Ivanova J, Toncheva-Panova T, Chernev G, Samuneva B. Effect of Ag⁺, Cu²⁺ and Zn²⁺ containing hybrid nano matrixes on the green algae *Chlorella keissleri*. General and Applied Plant Physiology. 2008;**34**:339-348
- [102] Joner EJ, Briones R, Leyval C. Metal-binding capacity of arbuscular mycorrhizal mycelium. Plant and Soil. 2000;**226**:227-234. DOI: 10.1023/A:1026565701391.
- [103] Kar M, Mishra D. Catalase, peroxidase, and polyphenoloxidase activities during rice leaf senescence. Plant Physiology. 1976;**57**:315-319. PMID: 16659474
- [104] Kaldorf M, Kuhn AJ, Schroder WH, Hildebrandt U, Bothe H. Selective element deposits in maize colonized by a heavy metal tolerance conferring arbuscular mycorrhizal fungus. Journal of Plant Physiology. 1999;**154**:718-728. DOI: 10.1016/S0176-1617(99)80250-8
- [105] Kukreja S, Nandval AS, Kumar N, Sharma SK, Unvi V, Sharma PK. Plant water status, H₂O₂ scavenging enzymes, ethylene evolution and membrane integrity of *Cicer arietinum* roots as affected by salinity. Biologia Plantarum. 2005;**49**:305-308
- [106] Mehlhorn H, Lelandais M, Korth HG, Foyer CH. Ascorbate is the natural substrate for plant peroxidase. FEBS Letters. 1996;**378**:203-206. SSDI: 0014-5793(95)01448-9
- [107] Mozgawa W, Król M, Bajda T. Application of IR spectra in the studies of heavy metal cations immobilization on natural sorbents. Journal of Molecular Structure. 2009;**924-926**:427-433. DOI: 10.1016/j.molstruc.2008.12.028
- [108] Nakano Y, Asada K. Hydrogen peroxide is scavenged by ascorbate specific peroxidase in spinach chloroplasts. Plant & Cell Physiology. 1981;**22**:867-880. DOI: 10.1093/oxford-journals.pcp.a076232
- [109] Pooria G, Tahereh TM, Bijan R. Differential scanning calorimetry techniques: Applications in biology and nanoscience. Journal of Biomolecular Techniques. 2010;**21**(4):167-193. PMCID: PMC2977967
- [110] Stoeppler M. Cadmium. In: Merian E, editor. Metals and their Compounds in the Environment: Occurrence, Analyses and Biological Relevance. New York: VCH; 1991. pp. 803-851
- [111] Sadasivam S, Manickam A. Biochemical methods for agricultural sciences; 1992. pp. 12-13. Sumner JB, Gjessing EC. Arch. Biochem. 1943;**2**:291
- [112] Shah K, Kumar RG, Verma S, Dubey RS. Effect of cadmium on lipid peroxidation, superoxide anion generation and activities of antioxidant enzymes in growing rice seedlings. Plant Science. 2001;**161**:1135-1144. DOI: 10.1016/S0168-9452(01)00517-9
- [113] Smeets K, Cuypers A, Lambrechts A, et al. Induction of oxidative stress and antioxidant mechanisms in *Phaseolus vulgaris* after Cd application. Plant Physiology and Biochemistry. 2005;**43**(5):437-444. DOI: 10.1016/j.plaphy.2005.03.007
- [114] Sbartaï H, Rouabhi R, Sbartaï I, Berrebbah H, Djebbar RM. Induction of anti-oxidative enzymes by cadmium stress in tomato (*Lycopersicon esculentum*). African Journal of Plant Science. 2008;**2**(8):72-76. Article Number - E4FC5413005

- [115] Thomet M, Vogel E, Krahenbuhl U. The uptake of cadmium and zinc by mycelia and their accumulation in mycelia and fruiting bodies of edible mushrooms. *European Food Research and Technology*. 1999;**209**(5):317-324. DOI: 10.1155/2013/1499120
- [116] WHO. Lead Environmental Health Criteria. Geneva: WHO; 1995
- [117] Yang X, Baligar VC, Martens DC, Clark RB. Cadmium effects on influx and transport of mineral nutrients in plant species (*Sedum alfredii* Hance). *Plant and Soil*. 1996;**259**: 181-189. DOI: 10.1080/01904169609365148
- [118] Zhang H. Chromium contamination in the soil from an alloy steel factory in Nanjing. *China Environmental Science*. 1997;**17**(2):80-82

