

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Secondary Prevention of Uterine Cervical Cancer

Seiya Sato and Hiroaki Itamochi

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/intechopen.72144>

Abstract

Secondary prevention by cervical cytology has clearly improved the mortality rate of uterine cervical cancer (CC) by enabling early detection and treatment of high-grade squamous intraepithelial lesion (HSIL) or cervical intraepithelial neoplasia (CIN), which is a precancerous lesion. In the past two decades, HPV-DNA testing, including HPV typing, has clearly brought about positive effects on secondary prevention of CC. However, in practice, CC remains a fatal disease and is the second leading cause of cancer deaths in women aged 20–39 years. Although elucidation of the mechanisms of HPV carcinogenesis and development of a prophylactic vaccine have made CC a preventable disease, eradication of CC is expected to take several decades. Therefore, primary screening to decrease the mortality rate of CC will remain important for a while. In addition, the clinical application of simple biomarkers to stratify HPV-positive women is important for maintenance of medical economy and avoidance of overtreatment in women in the reproductive age. Therefore, the development of an inexpensive therapy or vaccine that can be used worldwide is necessary to overcome cancer deaths due to CC.

Keywords: uterine cervical cancer, cervical intraepithelial neoplasia, secondary prevention, human papillomavirus, carcinogenesis, biomarker, therapeutic vaccine

1. Introduction

Secondary prevention with the use of cervical cytology has clearly improved the mortality rate and early treatment of uterine cervical cancer (CC) by enabling early detection of high-grade squamous intraepithelial lesion (HSIL) or cervical intraepithelial neoplasia (CIN), which is a precancerous lesion [1]. In practice, however, CC was estimated to have 12,820 newly diagnosed cases and 4210 women dying of the disease in 2017 [2]. Moreover, according to the United States data in 2014, CC is the second leading cause of cancer deaths in women aged 20–39 years [2]. Therefore, improvement of screening efficiency remains an important issue.

The etiology of CC is persistent uterine cervical infection with the high-risk human papillomavirus (hrHPV). Therefore, HPV-DNA testing or HPV testing, has become widely used for primary screening of CC. Compared with conventional cytology, HPV testing has higher sensitivity and reproducibility in detecting lesions [3]. However, the specificity of HPV testing is low, with an increase in the number of false-positives, especially in women in their twenties who are highly sexually active [3, 4]. Therefore, HPV testing has been adopted in cancer screening for women over 30 years old. In fact, in the United States (US), the guidelines created by the American Cancer Society, American Society for Clinical Pathology, and American Society for Colposcopy and Cervical Pathology suggested CC screening by cytology starting at the age of 21 and every 3 years until 30 years old; beyond the age of 30, combined HPV testing and cytology for every 5 years was recommended [5]. Based on data from large-scale, longitudinal, randomized-controlled trials in European countries, HPV testing has been adopted as the primary screening tool for CC in women aged 30 years or older [6–9]; in those who are tested positive for HPV, cytology is used as the triage test. In ASC-US cases, HPV testing is performed for triage in the management of CIN, based on the results of available large-scale clinical studies [10–13]. Furthermore, HPV typing has already been used as a biomarker for decisions on therapeutic interventions and subsequent follow-up of CIN [14–19]. Both the US and European guidelines recommended HPV testing to confirm the completion of treatment of CIN.

HPVs are classified according to carcinogenic potential. In general, the frequently reported high-risk types are HPV 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, and 68 [20]. Among these, HPV 16 and 18 are the most common types that are related to carcinogenesis worldwide; both HPV types are controllable by prophylactic vaccines that contain virus-like particles with antigenicity [21, 22]. Bivalent vaccines for HPV 16 and 18 are commercially available, but quadrivalent vaccines are also available for HPV 6, 11, 16, and 18. Although these vaccines have some cross-protective effects [23, 24], these are basically ineffective for infection by all HPV types. To overcome these limitations, a nonavalent vaccine containing HPV 6, 11, 16, 18, 31, 33, 45, 52, and 58 has been launched [25, 26].

As mentioned above, hrHPV testing has clearly brought about positive effects on early detection of CIN and prevention of CC in the past two decades [27–30]. Several researchers all over the world continue to pursue efforts to eradicate CC. **Figure 1** shows the schema of the natural

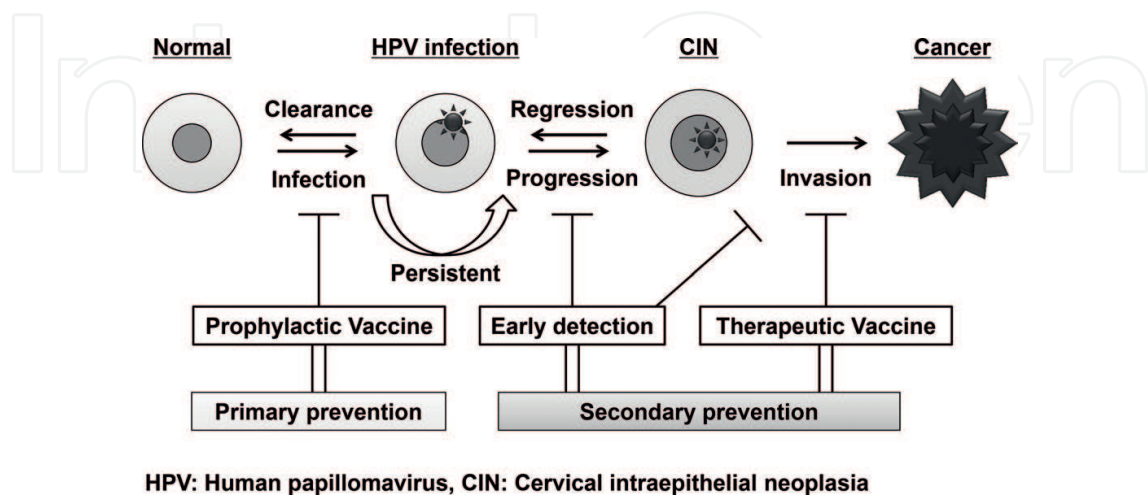


Figure 1. Natural history of HPV and prevention of cervical cancer. Persistent infection of the cervix with high-risk HPV causes cervical cancer (CC), which begins as cervical intraepithelial neoplasia (CIN). Primary prevention of CC can be achieved by prophylactic HPV vaccination. Secondary prevention consists of early detection of CIN and therapeutic vaccination to inhibit progression from CIN to CC.

history of HPV and CC prevention. In this chapter, we will describe the recent developments in secondary prevention of CC.

2. Biology and carcinogenesis of HPV

HPV is a virus with a double-stranded circular DNA in the icosahedral capsid. The genome size is about 8000 bases and contains eight protein-coding genes and a noncoding, regulatory long control region [31]. The early genes (E1, E2, E4, E5, E6, and E7) encode nonstructural proteins involved in replication, transcription, and transformation; whereas the late genes (L1 and L2) encode viral capsid proteins. Among these genes, E6 and E7 play a central role, particularly in carcinogenesis. Notably, a recent whole-genome sequencing study that assessed the risk of viral genetic variation showed that strict preservation of the 98 amino acids of E7, which destroys the function of the retinoblastoma protein (pRB), was critical for HPV 16 carcinogenesis and development of CIN and CC [32].

HPV can infect the epithelial cells of the human mucosa and skin at least once in most women's lives. In other words, HPV infection is a common sexually transmitted disease. Because prophylactic vaccines prevent only initial infection, its value in women is most effective before the first sexual contact [33]. In the early stages of HPV infection, the host is asymptomatic, and in most cases, the virus is eliminated by the immune system within a few years [34]. However, HPV infection can persist in some patients. The reported risk factors for progression of cervical HPV infection to CIN or CC include persistent hrHPV infection, immunosuppression, age over 30 years, and smoking [35].

Persistent hrHPV infection of the cervix is divided into three stages: latent, permissive, and transforming [36–38]. First, HPV invades the epithelial basal cells via minor breaches of the epithelium [39] and become latent as a nuclear episome; the infected cells usually die after virus multiplication. The E6 and E7 genes are rarely integrated into cellular DNA and cause HPV growth in the cells; however, this property also allows continued expression of E6 and E7 proteins at high levels. The expression of E6 and E7 oncogenes in basal cells is tightly controlled; therefore, HPV-infected cells can escape a host's immune defense. In fact, in a small percentage of HPV-infected women, HPV-specific antibodies and T cells are detected at low levels [40, 41]. Recently, it was suggested that the programmed death 1/programmed death 1 ligand (PD-1/PD-L1) pathway might be involved in the mechanism of this immune evasion [42–44].

When infected cells begin to differentiate in the epidermis, the E6 proteins degrade the tumor suppressor protein p53, while the E7 proteins inhibit the function of the pRB; these processes reactivate DNA synthesis and replication of the HPV genome. The cells with integrated E6 and E7 genes will have uncontrolled cell cycles because p53 and pRB are major cell cycle regulators. Furthermore, apoptosis and the tumor suppressor pathway are repressed. During this process, accumulation of genetic mutations and genomic instability ensue [45–50]. As a result, a large number of clones with intratumor heterogeneity are produced, some of which might be able to avoid the host antitumor response [51–54]. Ultimately, with the addition of external factors, these cells will be immortalized and can become cancerous [55].

3. Biomarker for early detection and triage

HPV testing has been introduced for primary screening for CC; it is highly sensitive, but its false-positive rate is high due to the low specificity. Therefore, the need to stratify HPV-positive women with or without abnormal cytology has become a very important issue [56]. At present, HPV-positive women undergo cytology tests and HPV retesting, with colposcopy and tissue biopsy correlations at frequent intervals; however, the precision of this process remains unclear. More objective indicators are required to prevent unnecessary procedures and treatment. As the understanding of the molecular mechanisms of cervical carcinogenesis by HPV has progressed, various biomarkers that predict patient outcomes have been developed not only for early detection but also for triage.

As mentioned earlier, persistent HPV infection of cervical cells leads to tumor formation through several stages. Since HPV infection is often transient, detection of the stage when HPV infection shifts from permissive to transforming is clinically important for cancer screening. Similarly, the histopathologic and molecular diagnostic processes for CIN focus on detection of malignant transformation in HPV-infected cells [57, 58]. The function of HPV-transformed cells is critically dependent on E6 and E7 oncogenes and related molecules such as p16^{INK4a} [59, 60]. Therefore, E6/E7 mRNA and p16^{INK4a} are important targets for early detection and triage. In addition, genetic or epigenetic changes in HPV-transformed cells have been attracting attention as biomarkers for screening of CC, in the triage of HPV-positive women, and as targets of treatment. Because, such new biomarkers can be analyzed from preserved liquid-based cytology (LBC) specimens, their use may be further expanded [61].

3.1. HPV typing

HPV 16 and 18 account for 70% of the causes of CC. The other reported HPV types related to CC are 31, 33, 35, 45, 52, and 58 [62]. Furthermore, the risk of developing CC has been reported to differ according to the type of hrHPV [63]. A cohort study to estimate the risk of disease progression among the HPV genotypes in 570 Japanese women with cytologic low-grade squamous intraepithelial lesion (LSIL) and histologic CIN1/2 showed that the cumulative probability of CIN3 within 5 years was higher in HPV 16, 18, 31, 33, 35, 52, and 58 than in the other hrHPV types [64]. Another Japanese cohort study on cytologic abnormalities, including ASC-US, LSIL, and HSIL (\leq CIN2), reported that infection with HPV types 16, 18, and 33 posed a high risk of developing CIN3 [65]. The Japanese gynecologic guideline 2017 recommended HPV typing to evaluate the risk of disease progression for patients with histologically proven CIN1/2 (**Figure 2**). Taken altogether, HPV typing in CIN patients is useful for risk assessment of disease progression [66].

3.2. p16^{INK4a}

The p16^{INK4a} is a cyclin-dependent kinase inhibitor that blocks the phosphorylation of various cyclins that control the cell cycle. In many human cancers, including colon and breast, the function of the p16^{INK4a} gene is lost by gene deletions, mutations, or epigenetic silencing. In CC,

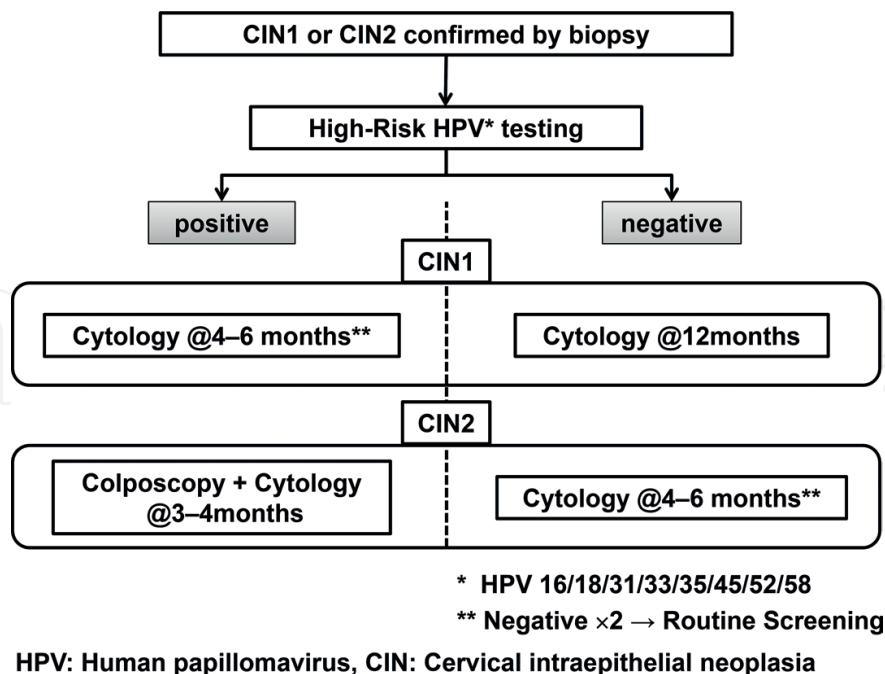


Figure 2. HPV typing for CIN1 and CIN2 in Japan. The Japanese gynecologic guidelines in 2017 recommend HPV typing to evaluate the risk of disease progression in patients with histologically proven CIN1/2. Patients who are positive for high-risk HPV receive more intensive management compared with negative patients.

however, a high level of intracellular E7 expression eliminates the inhibitory methylation mark encoding the CDKN2A gene promoter from p16^{INK4a}, resulting in overexpression of the p16^{INK4a} protein [67]. In addition, since E7 inactivates pRB, there is proliferation of cells that highly express p16^{INK4a}. In other words, high expression of p16^{INK4a} reflects the high expression of E7, which is a good indicator of CIN3 and CC. For this reason, p16^{INK4a} is widely accepted as a valuable surrogate biomarker for the transforming properties of HPV infection [68]. Based on this fact, a therapeutic peptide vaccine using p16^{INK4a} as the antigen has been developed [69]. Moreover, p16^{INK4a} has been used for dual-staining with p16/Ki-67 cytology (p16/Ki-67); this would complement the low sensitivity of cytology and the low specificity of the HPV test for secondary prevention of CC [70–72].

In Europe, p16/Ki-67 was compared with Papanicolaou (Pap) cytology and HPV testing for screening high-grade CIN (CIN2+) in 27,349 women aged 18 years or older; the p16/Ki-67 had high sensitivity and comparable specificity for CIN2 detection, compared with the other tests [73]. This suggested the utility of p16/Ki-67 as a screening method in young women with high HPV infection rates. Other studies showed the effectiveness of p16/Ki-67 as a triage test for CIN2+ detection in Pap-negative and HPV-positive women ≥30 years old [74]. In addition, the usefulness of p16/Ki-67 for follow-up of patients after CIN treatment was suggested [75]. In Germany, a recent study revealed that combined HPV 16/18 testing and p16/Ki-67 resulted in lower cost and clinically efficient CC screening, compared with conventional annual Pap cytology [76]. As described earlier, several evidences on the utility of p16/Ki-67 have accumulated; therefore, p16/Ki-67 will definitely play an important role in the secondary prevention of CC.

3.3. HPV E6/E7 mRNA

The usefulness of HPV E6/E7 mRNA testing in secondary prevention of CC has already been established [77, 78]. Combined testing of LBC cytology and APTIMA® HPV (AHPV) has already been used in the US and Europe. HPV E6/E7 mRNA testing is useful because it can detect HPV infection and the transformation properties of cervical cells. In fact, HPV E6/E7 mRNA testing was reported to detect high-grade CIN with high sensitivity and specificity [79–81].

ASC-US and LSIL on cytology are mainly caused by low-grade cervical lesions that often resolve spontaneously [82]. Currently, the HPV test is used for triage of women with ASC-US and LSIL; however, its low specificity has increased the number of unnecessary examinations and treatment [83]. On the other hand, HPV E6/E7 mRNA testing enables stratification of the risk of developing CIN2+ in women with both hrHPV-positive and hrHPV-negative cytology [84]. A recent meta-analysis to confirm usefulness of HPV E6/E7 mRNA testing for triage revealed that a positive HPV E6/E7 mRNA testing in women with mild cytology findings, such as ASC-US and LSIL, necessitates immediate colposcopy and intensive follow-up because the risk of carcinogenesis is high [85].

3.4. Epigenomic alterations

Epigenetic events in the host and in viral genomic regions and genes are necessary during HPV-mediated cellular transformation and carcinogenesis [86]. DNA methylation is a typical epigenetic change and characterizes the molecular, cellular, and clinical features of HPV-associated neoplasia. Because hypermethylation is a stable and reversible process, detection of methylation marks is used for diagnosis. In addition, new targeted therapies with demethylating compounds have been developed [87].

Combined testing with DNA methylation and hrHPV is one of the promising screening options for CC. The Triage and Risk Assessment of Cervical Precancer by Epigenetic Biomarker (TRACE) study was conducted to examine the usefulness of human epigenetic biomarker testing in the primary prevention of CC. In this study, methylation of the POU4F3 promoter, which is a promising marker for CIN3, showed significantly higher sensitivity and similar specificity for detecting CIN3+, compared with LBC [88]. This finding suggested that detection of POU4F3 methylation is useful for early detection of CIN3. Another study assessed the correlation between CpG methylation of the HPV16 L1 gene and CC in 145 HPV 16-infected Uyghur women who were divided into five groups, as follows: transient infection ($n = 32$), persistent infection ($n = 21$), CIN1 ($n = 21$), CIN2–3 ($n = 33$), and CC ($n = 38$) [89]. After quantifying each CPG methylation by pyrosequencing, results revealed that methylation increased at 13 CpG sites in advanced lesions and that high methylation levels were associated with the risk of developing CIN2+ [89]. These findings may be applied to CC screening.

4. Therapeutic vaccine for CIN

Currently, the standard therapy for CIN is surgical excision such as conization or LEEP. Although these treatments are very effective from the viewpoint of removing HPV-induced

precancerous lesions, these can cause infertility and menstrual disorders secondary to stenosis of the cervix. In addition, the existing CC prophylactic vaccines are ineffective for HPV-infected women and nontargeted HPV types. Therefore, development of a therapeutic vaccine using immunotherapy as a nonsurgical treatment for CIN is an important strategy for the prevention of CC.

The development of therapeutic vaccines has been mainly targeted for HPV E6 and E7 [90], because these proteins are essential for the malignant transformation of HPV-infected cells and are permanently expressed in CIN. In order to induce an E6 or E7 antigen-specific T cell immune response, several kinds of vaccines have been developed; these include adoptive transfer of tumor-specific T cells, chimeric virus-like particle vaccines, dendritic cell, DNA vaccines, peptide vaccines, protein vaccines, and viral or bacterial vector vaccines. Among these, protein vaccines are the most common therapeutic vaccines for HPV 16 because of the simplicity of the method and the lack of HLA restriction [91]. However, there are currently no available therapeutic HPV vaccines against CIN.

Recently, a randomized, double-blind, placebo-controlled phase 2b trial on CIN2/3 patients showed promising results on the efficacy and safety of VGX-3100, which is a synthetic plasmid targeting human HPV 16 and HPV 18 E6 and E7 proteins (ClinicalTrials.gov Identifier: NCT01304524). In the study, the primary endpoint for efficacy was regression to CIN1 or normal pathology at 36 weeks after the first dose. This study enrolled 167 people who were randomized (3:1) to the VGX-3100 group ($n = 125$) and the placebo group ($n = 42$); the rate of histopathologic regression was significantly higher by 18.2% [95% CI 1.3–34.4] in the VGX-3100 group, compared with the placebo group (48.2% vs. 30.0%; $p = 0.034$). The incidence of erythema at the injection site was significantly higher in the VGX-3100 group than in the placebo group (78.4% vs. 57.1%, $p = 0.007$). On the other hand, there was no significant increase in the number of severe side effects that could interfere with the performance of vaccine therapy [92]. Therefore, this vaccine might be a nonsurgical therapeutic option for CIN2/3, but further research and development are needed in this field. A clinical trial on HPV therapeutic vaccines was detailed in a recent review [93].

5. Discussion

The mathematical model by a German group estimated that the incidence and mortality of CC will drastically decrease in the next 30 years due to the increasing number of screening participants since the 1990s [94]. Furthermore, even at a vaccination rate of only 50%, more than 40% of CC is considered to be preventable in the next 100 years [94]. Nevertheless, more effective primary prevention is necessary to eradicate CC. Currently, an effective vaccine can be used to inhibit a part of the hrHPV infection process that leads to cancer. However, several individuals cannot receive vaccination due to economic or geographical problems. In order to solve this problem, international cooperation and national policy are necessary to construct a CC prevention system. One possible problem in the future would be the changes in the distribution of hrHPV types due to an increase in the number of vaccinated cohorts; this would

likely decrease the efficiency of the current screening system. Therefore, it may be necessary to monitor the distribution of hrHPV in each country and region, and to develop screening methods that are suitable for each situation.

Secondary prevention remains important because vaccines only prevent infection with a limited number of HPV types. In order to reduce the mortality rate of CC, the coverage of a screening program needs to be increased and include patients with advanced CC. To address this issue, the usefulness of self-sampling for HPV testing has been studied [95, 96]. The US National Health and Nutrition Examination Survey in 2007–2010 on women aged 18–59 years revealed a 41.9% prevalence of genital HPV infection [97]. Multivariate analysis in this cohort revealed that HPV infection was related to age, number of sexual partners, smoking, educational level, income, and insurance status [97]. Similar results on the risk of persistent HPV infection have been confirmed in other studies [98, 99]. Therefore, populations with these risk factors require more rigorous and continuous monitoring for effective prevention; in these cases, self-sampling may be particularly useful. Importantly, the hrHPV detection rate by continuous self-sampling of vaginal fluid for 28 days was reported to be consistent regardless of the hormonal cycle [100]. In other words, hrHPV detection by self-sampling can be adapted to all women, even those in the nonmenstrual period, including menopause. A recent meta-analysis of 37 studies including 18,516 women revealed that HPV-DNA sampling screening was highly accepted compared with clinician’s sampling. In the future, the importance of self-collection method will increase, especially from the viewpoint

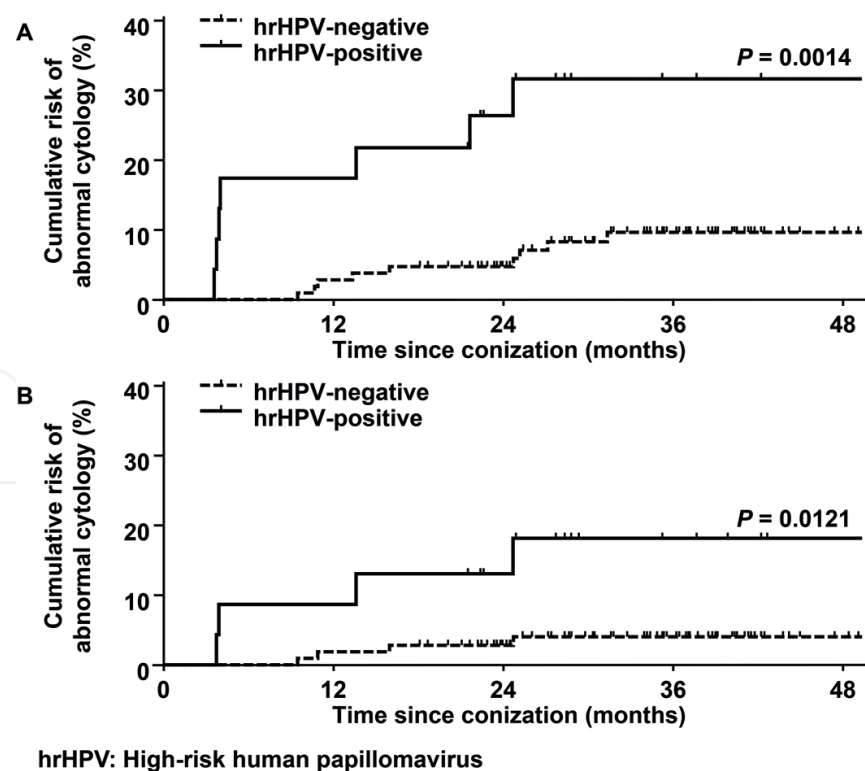


Figure 3. Postoperative infection with high risk (hr) HPV and risk of abnormal cytology. The cumulative risk curves for (a) atypical squamous cells of undetermined significance (ASC-US) or higher and (b) low-grade squamous intraepithelial lesion (LSIL) or higher show that the cumulative risks for recurrence of abnormal cytology and LSIL or higher were significantly increased in postoperative hrHPV-positive patients than in hrHPV-negative patients.

of cost-effectiveness and expansion of screening services [101]. Therefore, HPV-DNA testing by self-sampling has the potential to become the mainstream in cancer prevention.

CIN frequently regresses spontaneously within months or a few years [102, 103]. However, there is no biomarker to predict spontaneous regression of CIN. The standard treatment for CIN is still surgical resection such as conization; for a long time, there had been no other options for treatment. Although surgical excision is successful for CIN treatment most of the time, HPV infection cannot be completely eliminated. We reported that postsurgical hrHPV infection was a positive predictor of the recurrence of abnormal cytology (**Figure 3**). Furthermore, surgical procedure can lead to complications such as pregnancy problems, infertility, incontinence, and sexual dysfunction [104–106]. At the very least, overtreatment of women with fertility must be avoided. With the progression of CC screening, the importance of these problems has increased. In order to overcome this problem, development of a therapeutic vaccine as a new treatment option without surgery is urgently needed. The availability of low-cost therapeutic vaccines for patients with CIN or stage IA CC in the future will lead to a long-term reduction in medical costs [107].

6. Conclusions

Although elucidation of the mechanisms of HPV carcinogenesis and development of a prophylactic vaccine have made CC a preventable disease, eradication of CC is expected to take several decades. To decrease the mortality rate of CC, early detection by screening will remain important for a while. The clinical application of simple biomarkers to stratify HPV-positive women is important for maintenance of medical economy and avoidance of overtreatment of women in the reproductive age. To overcome cancer deaths due to CC, the development of inexpensive treatment options or therapeutic vaccines that can be readily used worldwide is necessary.

Author details

Seiya Sato and Hiroaki Itamochi*

*Address all correspondence to: itamochi@iwate-med.ac.jp

Department of Obstetrics and Gynecology, Iwate Medical University School of Medicine, Iwate, Morioka, Japan

References

- [1] Saslow D. American Cancer Society, American Society for Colposcopy and Cervical Pathology, and American Society for Clinical Pathology screening guidelines for the prevention and early detection of cervical cancer. *Journal of Lower Genital Tract Disease*. 2012; 16(3):175-204

- [2] Siegel RL. Cancer statistics, 2017. *CA: A Cancer Journal for Clinicians*. 2017;**67**(1):7-30
- [3] Wright TC Jr. Interim guidance for the use of human papillomavirus DNA testing as an adjunct to cervical cytology for screening. *Obstetrics and Gynecology*. 2004;**103**(2): 304-309
- [4] Massad LS. 2012 updated consensus guidelines for the management of abnormal cervical cancer screening tests and cancer precursors. *Obstetrics and Gynecology*. 2013; **121**(4):829-846
- [5] Saslow D. American Cancer Society, American Society for Colposcopy and Cervical Pathology, and American Society for Clinical Pathology screening guidelines for the prevention and early detection of cervical cancer. *CA: A Cancer Journal for Clinicians*. 2012;**62**(3):147-172
- [6] Ronco G. Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: A randomised controlled trial. *The Lancet Oncology*. 2010;**11**(3):249-257
- [7] Bulkman NW. Human papillomavirus DNA testing for the detection of cervical intraepithelial neoplasia grade 3 and cancer: 5-year follow-up of a randomised controlled implementation trial. *Lancet*. 2007;**370**(9601):1764-1772
- [8] Rijkaart DC. Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: Final results of the POBASCAM randomised controlled trial. *The Lancet Oncology*. 2012;**13**(1):78-88
- [9] Kitchener HC. HPV testing in combination with liquid-based cytology in primary cervical screening (ARTISTIC): A randomised controlled trial. *The Lancet Oncology*. 2009; **10**(7):672-682
- [10] Results of a randomized trial on the management of cytology interpretations of atypical squamous cells of undetermined significance. *American Journal of Obstetrics and Gynecology*. 2003;**188**(6):1383-1392
- [11] A randomized trial on the management of low-grade squamous intraepithelial lesion cytology interpretations. *American Journal of Obstetrics and Gynecology*. 2003;**188**(6): 1393-1400
- [12] Katki HA. Benchmarking CIN 3+ risk as the basis for incorporating HPV and Pap cotesting into cervical screening and management guidelines. *Journal of Lower Genital Tract Disease*. 2013;**17**(5 Suppl 1):S28-S35
- [13] Wright TC Jr. Evaluation of HPV-16 and HPV-18 genotyping for the triage of women with high-risk HPV+ cytology-negative results. *American Journal of Clinical Pathology*. 2011;**136**(4):578-586
- [14] Hoffman SR. Patterns of persistent HPV infection after treatment for cervical intraepithelial neoplasia (CIN): A systematic review. *International Journal of Cancer*. 2017; **141**(1):8-23

- [15] Kocken M. Risk of recurrent high-grade cervical intraepithelial neoplasia after successful treatment: A long-term multi-cohort study. *The Lancet Oncology*. 2011;**12**(5):441-450
- [16] van der Heijden E. Follow-up strategies after treatment (large loop excision of the transformation zone (LLETZ)) for cervical intraepithelial neoplasia (CIN): Impact of human papillomavirus (HPV) test. *The Cochrane Database of Systematic Reviews*. 2015;**1**:CD010757
- [17] Katki HA. Five-year risk of recurrence after treatment of CIN 2, CIN 3, or AIS: Performance of HPV and Pap cotesting in posttreatment management. *Journal of Lower Genital Tract Disease*. 2013;**17**(5 Suppl 1):S78-S84
- [18] Cubie HA. Evaluation of commercial HPV assays in the context of post-treatment follow-up: Scottish Test of Cure Study (STOCS-H). *Journal of Clinical Pathology*. 2014;**67**(6):458-463
- [19] Gosvig CF. Long-term follow-up of the risk for cervical intraepithelial neoplasia grade 2 or worse in HPV-negative women after conization. *International Journal of Cancer*. 2015;**137**(12):2927-2933
- [20] Brianti P. Review of HPV-related diseases and cancers. *The New Microbiologica*. 2017;**40**(2):80-85
- [21] Quadrivalent vaccine against human papillomavirus to prevent high-grade cervical lesions. *The New England journal of medicine*. 2007;**356**(19):1915-1927
- [22] Lehtinen M. Overall efficacy of HPV-16/18 AS04-adjuvanted vaccine against grade 3 or greater cervical intraepithelial neoplasia: 4-year end-of-study analysis of the randomised, double-blind PATRICIA trial. *The Lancet Oncology*. 2012;**13**(1):89-99
- [23] Wheeler CM. Cross-protective efficacy of HPV-16/18 AS04-adjuvanted vaccine against cervical infection and precancer caused by non-vaccine oncogenic HPV types: 4-year end-of-study analysis of the randomised, double-blind PATRICIA trial. *The Lancet Oncology*. 2012;**13**(1):100-110
- [24] Brown DR. The impact of quadrivalent human papillomavirus (HPV; types 6, 11, 16, and 18) L1 virus-like particle vaccine on infection and disease due to oncogenic nonvaccine HPV types in generally HPV-naïve women aged 16-26 years. *The Journal of Infectious Diseases*. 2009;**199**(7):926-935
- [25] Herrero R. Present status of human papillomavirus vaccine development and implementation. *The Lancet Oncology*. 2015;**16**(5):e206-e216
- [26] Pista A. Potential impact of nonavalent HPV vaccine in the prevention of high-grade cervical lesions and cervical cancer in Portugal. *International Journal of Gynaecology and Obstetrics: the Official Organ of the International Federation of Gynaecology and Obstetrics*. 2017;**139**(1):90-94
- [27] Castle PE. Performance of carcinogenic human papillomavirus (HPV) testing and HPV16 or HPV18 genotyping for cervical cancer screening of women aged 25 years and older: A subanalysis of the ATHENA study. *The Lancet Oncology*. 2011;**12**(9):880-890

- [28] Rijkaart DC. Evaluation of 14 triage strategies for HPV DNA-positive women in population-based cervical screening. *International Journal of Cancer*. 2012;**130**(3):602-610
- [29] Veldhuijzen NJ. Stratifying HPV-positive women for CIN3+ risk after one and two rounds of HPV-based screening. *International Journal of Cancer*. 2017;**141**(8):1551-1560
- [30] Massad LS. 2012 updated consensus guidelines for the management of abnormal cervical cancer screening tests and cancer precursors. *Journal of Lower Genital Tract Disease*. 2013;**17**(5 Suppl 1):S1-S27
- [31] Bernard HU. Genome variation of human papillomavirus types: Phylogenetic and medical implications. *International Journal of Cancer*. 2006;**118**(5):1071-1076
- [32] Mirabello L. HPV16 E7 genetic conservation is critical to carcinogenesis. *Cell*. 2017;**170**(6):1164-1174 e1166
- [33] Hildesheim A. Human papillomavirus vaccine should be given before sexual debut for maximum benefit. *The Journal of Infectious Diseases*. 2007;**196**(10):1431-1432
- [34] Franco EL. Epidemiology of acquisition and clearance of cervical human papillomavirus infection in women from a high-risk area for cervical cancer. *The Journal of Infectious Diseases*. 1999;**180**(5):1415-1423
- [35] Forcier M. An overview of human papillomavirus infection for the dermatologist: Disease, diagnosis, management, and prevention. *Dermatologic Therapy*. 2010;**23**(5):458-476
- [36] Doeberitz M. Host factors in HPV-related carcinogenesis: Cellular mechanisms controlling HPV infections. *Archives of Medical Research*. 2009;**40**(6):435-442
- [37] Doorbar J. Molecular biology of human papillomavirus infection and cervical cancer. *Clinical Science (London, England)*. 2006;**110**(5):525-541
- [38] Doorbar J. The biology and life-cycle of human papillomaviruses. *Vaccine*. 2012;**30**(Suppl 5):F55-F70
- [39] Kines RC. The initial steps leading to papillomavirus infection occur on the basement membrane prior to cell surface binding. *Proceedings of the National Academy of Sciences of the United States of America*. 2009;**106**(48):20458-20463
- [40] Reuschenbach M. Characterization of humoral immune responses against p16, p53, HPV16 E6 and HPV16 E7 in patients with HPV-associated cancers. *International Journal of Cancer*. 2008;**123**(11):2626-2631
- [41] de Jong A. Human papillomavirus type 16-positive cervical cancer is associated with impaired CD4+ T-cell immunity against early antigens E2 and E6. *Cancer Research*. 2004;**64**(15):5449-5455
- [42] Mezache L. Enhanced expression of PD L1 in cervical intraepithelial neoplasia and cervical cancers. *Modern Pathology: An Official Journal of the United States and Canadian Academy of Pathology, Inc*. 2015;**28**(12):1594-1602

- [43] Yang W. Increased expression of programmed death (PD)-1 and its ligand PD-L1 correlates with impaired cell-mediated immunity in high-risk human papillomavirus-related cervical intraepithelial neoplasia. *Immunology*. 2013;**139**(4):513-522
- [44] Yang W. Expressions of programmed death (PD)-1 and PD-1 ligand (PD-L1) in cervical intraepithelial neoplasia and cervical squamous cell carcinomas are of prognostic value and associated with human papillomavirus status. *The Journal of Obstetrics and Gynaecology Research*. 2017;**43**(10):1602-1612
- [45] Kuner R. Identification of cellular targets for the human papillomavirus E6 and E7 onco-genes by RNA interference and transcriptome analyses. *Journal of Molecular Medicine*. 2007;**85**(11):1253-1262
- [46] McLaughlin-Drubin ME. Biochemical and functional interactions of human papilloma-virus proteins with polycomb group proteins. *Virus*. 2013;**5**(5):1231-1249
- [47] Munger K. Complex formation of human papillomavirus E7 proteins with the retino-blastoma tumor suppressor gene product. *The EMBO Journal*. 1989;**8**(13):4099-4105
- [48] Scheffner M. The E6 oncoprotein encoded by human papillomavirus types 16 and 18 promotes the degradation of p53. *Cell*. 1990;**63**(6):1129-1136
- [49] Duensing S. The human papillomavirus type 16 E6 and E7 oncoproteins cooperate to induce mitotic defects and genomic instability by uncoupling centrosome duplication from the cell division cycle. *Proceedings of the National Academy of Sciences of the United States of America*. 2000;**97**(18):10002-10007
- [50] White AE. Differential disruption of genomic integrity and cell cycle regulation in normal human fibroblasts by the HPV oncoproteins. *Genes & Development*. 1994;**8**(6):666-677
- [51] Akagi K. Genome-wide analysis of HPV integration in human cancers reveals recurrent, focal genomic instability. *Genome Research*. 2014;**24**(2):185-199
- [52] Heselmeyer K. Gain of chromosome 3q defines the transition from severe dysplasia to invasive carcinoma of the uterine cervix. *Proceedings of the National Academy of Sciences of the United States of America*. 1996;**93**(1):479-484
- [53] Cahill DP. Genetic instability and darwinian selection in tumours. *Trends in Cell Biology*. 1999;**9**(12):M57-M60
- [54] Thomas LK. Chromosomal gains and losses in human papillomavirus-associated neo-plasia of the lower genital tract—A systematic review and meta-analysis. *European Journal of Cancer*. 2014;**50**(1):85-98
- [55] Moody CA. Human papillomavirus oncoproteins: Pathways to transformation. *Nature Reviews Cancer*. 2010;**10**(8):550-560
- [56] Wentzensen N. Triage of HPV positive women in cervical cancer screening. *Journal of Clinical Virology: The Official Publication of the Pan American Society for Clinical Virology*. 2016;**76**(Suppl 1):S49-S55

- [57] Steenbergen RD. Clinical implications of (epi)genetic changes in HPV-induced cervical precancerous lesions. *Nature Reviews Cancer*. 2014;**14**(6):395-405
- [58] Reuschenbach M. Diagnostic tests for the detection of human papillomavirus-associated cervical lesions. *Current Pharmaceutical Design*. 2013;**19**(8):1358-1370
- [59] von Knebel Doeberitz M. Correlation of modified human papilloma virus early gene expression with altered growth properties in C4-1 cervical carcinoma cells. *Cancer Research*. 1988;**48**(13):3780-3786
- [60] Zur Hausen H. Papillomaviruses in anogenital cancer as a model to understand the role of viruses in human cancers. *Cancer Research*. 1989;**49**(17):4677-4681
- [61] Tota JE. Approaches for triaging women who test positive for human papillomavirus in cervical cancer screening. *Preventive Medicine*. 2017;**98**:15-20
- [62] de Sanjose S. Human papillomavirus genotype attribution in invasive cervical cancer: A retrospective cross-sectional worldwide study. *The Lancet Oncology*. 2010;**11**(11):1048-1056
- [63] Skinner SR. Progression of HPV infection to detectable cervical lesions or clearance in adult women: Analysis of the control arm of the VIVIANE study. *International Journal of Cancer*. 2016;**138**(10):2428-2438
- [64] Matsumoto K. Predicting the progression of cervical precursor lesions by human papillomavirus genotyping: A prospective cohort study. *International Journal of Cancer*. 2011;**128**(12):2898-2910
- [65] Hosaka M. Incidence risk of cervical intraepithelial neoplasia 3 or more severe lesions is a function of human papillomavirus genotypes and severity of cytological and histological abnormalities in adult Japanese women. *International Journal of Cancer*. 2013;**132**(2):327-334
- [66] Kudoh A. Human papillomavirus type-specific persistence and reappearance after successful conization in patients with cervical intraepithelial neoplasia. *International Journal of Clinical Oncology*. 2016;**21**(3):580-587
- [67] McLaughlin-Drubin ME. Human papillomavirus E7 oncoprotein induces KDM6A and KDM6B histone demethylase expression and causes epigenetic reprogramming. *Proceedings of the National Academy of Sciences of the United States of America*. 2011;**108**(5):2130-2135
- [68] Bergeron C. The clinical impact of using p16(INK4a) immunochemistry in cervical histopathology and cytology: An update of recent developments. *International Journal of Cancer*. 2015;**136**(12):2741-2751
- [69] Reuschenbach M. A phase 1/2a study to test the safety and immunogenicity of a p16(INK4a) peptide vaccine in patients with advanced human papillomavirus-associated cancers. *Cancer*. 2016;**122**(9):1425-1433

- [70] Tjalma WAA. Diagnostic performance of dual-staining cytology for cervical cancer screening: A systematic literature review. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2017;**210**:275-280
- [71] Ebisch RM. Evaluation of p16/Ki-67 dual-stained cytology as triage test for high-risk human papillomavirus-positive women. *Modern Pathology: An Official Journal of the United States and Canadian Academy of Pathology, Inc*. 2017;**30**(7):1021-1031
- [72] Wright TC Jr. Triage HPV-positive women with p16/Ki-67 dual-stained cytology: Results from a sub-study nested into the ATHENA trial. *Gynecologic Oncology*. 2017;**144**(1):51-56
- [73] Ikenberg H. Screening for cervical cancer precursors with p16/Ki-67 dual-stained cytology: Results of the PALMS study. *Journal of the National Cancer Institute*. 2013;**105**(20):1550-1557
- [74] Petry KU. Triage Pap cytology negative, HPV positive cervical cancer screening results with p16/Ki-67 dual-stained cytology. *Gynecologic Oncology*. 2011;**121**(3):505-509
- [75] Polman NJ. Good performance of p16/ki-67 dual-stained cytology for surveillance of women treated for high-grade CIN. *International Journal of Cancer*. 2017;**140**(2):423-430
- [76] Petry KU. A model to evaluate the costs and clinical effectiveness of human papilloma virus screening compared with annual papanicolaou cytology in Germany. *European Journal of Obstetrics, Gynecology, and Reproductive Biology*. 2017;**212**:132-139
- [77] Monsonego J. Evaluation of oncogenic human papillomavirus RNA and DNA tests with liquid-based cytology in primary cervical cancer screening: The FASE study. *International Journal of Cancer*. 2011;**129**(3):691-701
- [78] Arbyn M. Evidence regarding human papillomavirus testing in secondary prevention of cervical cancer. *Vaccine*. 2012;**30**(Suppl 5):F88-F99
- [79] Dockter J. Clinical performance of the APTIMA HPV Assay for the detection of high-risk HPV and high-grade cervical lesions. *Journal of Clinical Virology: The Official Publication of the Pan American Society for Clinical Virology*. 2009;**45**(Suppl 1):S55-S61
- [80] Monsonego J. Risk assessment and clinical impact of liquid-based cytology, oncogenic human papillomavirus (HPV) DNA and mRNA testing in primary cervical cancer screening (the FASE study). *Gynecologic Oncology*. 2012;**125**(1):175-180
- [81] Origoni M. E6/E7 mRNA testing for human papilloma virus-induced high-grade cervical intraepithelial disease (CIN2/CIN3): A promising perspective. *Ecancermedicalscience*. 2015;**9**(533). DOI: 10.3332/ecancer.2015.533
- [82] Alanen KW. Assessment of cytologic follow-up as the recommended management for patients with atypical squamous cells of undetermined significance or low grade squamous intraepithelial lesions. *Cancer*. 1998;**84**(1):5-10

- [83] Szarewski A. Comparison of seven tests for high-grade cervical intraepithelial neoplasia in women with abnormal smears: The predictors 2 study. *Journal of Clinical Microbiology*. 2012;**50**(6):1867-1873
- [84] Rijkaart DC. High-risk human papillomavirus (hrHPV) E6/E7 mRNA testing by PreTect HPV-Proofer for detection of cervical high-grade intraepithelial neoplasia and cancer among hrHPV DNA-positive women with normal cytology. *Journal of Clinical Microbiology*. 2012;**50**(7):2390-2396
- [85] Yang L. The clinical application of HPV E6/E7 mRNA testing in triaging women with atypical squamous cells of undetermined significance or low-grade squamous intra-epithelial lesion Pap smear: A meta-analysis. *Journal of Cancer Research and Therapeutics*. 2017;**13**(4):613-620
- [86] Clarke MA. Human papillomavirus DNA methylation as a potential biomarker for cervical cancer. *Cancer Epidemiology, Biomarkers & Prevention: A Publication of the American Association for Cancer Research*, cosponsored by the American Society of Preventive Oncology. 2012;**21**(12):2125-2137
- [87] Prigge ES. Clinical relevance and implications of HPV-induced neoplasia in different anatomical locations. *Mutation Research Reviews in Mutation Research*. 2017;**772**:51-66
- [88] Kocsis A. Performance of a new HPV and biomarker assay in the management of hrHPV positive women: Subanalysis of the ongoing multicenter TRACE clinical trial (n > 6,000) to evaluate POU4F3 methylation as a potential biomarker of cervical precancer and cancer. *International Journal of Cancer*. 2017;**140**(5):1119-1133
- [89] Niyazi M. Correlation between methylation of human Papillomavirus-16 L1 gene and cervical carcinoma in Uyghur women. *Gynecologic and Obstetric Investigation*. 2017;**82**(1):22-29
- [90] Rosales R. Immune therapy for human papillomaviruses-related cancers. *World Journal of Clinical Oncology*. 2014;**5**(5):1002-1019
- [91] Li J. A novel therapeutic vaccine composed of a rearranged human papillomavirus type 16 E6/E7 fusion protein and Fms-like tyrosine kinase-3 ligand induces CD8+ T cell responses and antitumor effect. *Vaccine*. 2017;**35**(47):6459-6467
- [92] Trimble CL. Safety, efficacy, and immunogenicity of VGX-3100, a therapeutic synthetic DNA vaccine targeting human papillomavirus 16 and 18 E6 and E7 proteins for cervical intraepithelial neoplasia 2/3: A randomised, double-blind, placebo-controlled phase 2b trial. *Lancet*. 2015;**386**(10008):2078-2088
- [93] Vici P. Targeting immune response with therapeutic vaccines in premalignant lesions and cervical cancer: Hope or reality from clinical studies. *Expert Review of Vaccines*. 2016;**15**(10):1327-1336
- [94] Horn J. Estimating the long-term effects of HPV vaccination in Germany. *Vaccine*. 2013;**31**(19):2372-2380

- [95] Belinson JL. Improved sensitivity of vaginal self-collection and high-risk human papillomavirus testing. *International Journal of Cancer*. 2012;**130**(8):1855-1860
- [96] Castle PE. Comparative community outreach to increase cervical cancer screening in the Mississippi Delta. *Preventive Medicine*. 2011;**52**(6):452-455
- [97] Shi R. Factors associated with genital human papillomavirus infection among adult females in the United States, NHANES 2007-2010. *BMC Research Notes*. 2014;**7**:544
- [98] Moscicki AB. Natural history of anal human papillomavirus infection in heterosexual women and risks associated with persistence. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America*. 2014;**58**(6):804-811
- [99] Rositch AF. Patterns of persistent genital human papillomavirus infection among women worldwide: A literature review and meta-analysis. *International Journal of Cancer*. 2013;**133**(6):1271-1285
- [100] Sanner K. Daily self-sampling for high-risk human papillomavirus (HR-HPV) testing. *Journal of Clinical Virology: The Official Publication of the Pan American Society for Clinical Virology*. 2015;**73**:1-7
- [101] Nelson EJ. The acceptability of self-sampled screening for HPV DNA: A systematic review and meta-analysis. *Sexually Transmitted Infections*. 2017;**93**(1):56-61
- [102] Moscicki AB. Rate of and risks for regression of cervical intraepithelial neoplasia 2 in adolescents and young women. *Obstetrics and Gynecology*. 2010;**116**(6):1373-1380
- [103] Trimble CL. Spontaneous regression of high-grade cervical dysplasia: Effects of human papillomavirus type and HLA phenotype. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*. 2005;**11**(13):4717-4723
- [104] Kyrgiou M. Obstetric outcomes after conservative treatment for intraepithelial or early invasive cervical lesions: Systematic review and meta-analysis. *Lancet*. 2006;**367**(9509):489-498
- [105] Vrzackova P. Sexual morbidity following radical hysterectomy for cervical cancer. *Expert Review of Anticancer Therapy*. 2010;**10**(7):1037-1042
- [106] Wit EM. Urological complications after treatment of cervical cancer. *Nature Reviews Urology*. 2014;**11**(2):110-117
- [107] Luttjeboer J. Threshold cost-effectiveness analysis for a therapeutic vaccine against HPV-16/18-positive cervical intraepithelial neoplasia in the Netherlands. *Vaccine*. 2016;**34**(50):6381-6387

