We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Trabecular Meshwork and Intraocular Pressure Dynamics: Oxidative Stress-Induced Changes

Bettina Hohberger, Ulrich-Christoph Welge-Lüßen and Alice Yu

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/66138

Abstract

Glaucoma is known as progressive neurodegenerative disease with irreversible loss of vision. Next to perimetric visual field defects, morphological alterations of the optic nerve and an increased intraocular pressure (IOP) occur. IOP is a fine regulated, complex homeostasis of production of aqueous humor (AH) in the non-pigmented ciliary body and its outflow. About 80% of AH is drained throughout the trabecular meshwork (TM). Any outflow resistance, with consecutive increase in IOP, can be generated by a decreased pore size or any other alterations of TM, making it more rigid. Oxidative stress, a disbalance of oxidants and antioxidants, is one mechanism, causing an altered extracellular matrix (ECM), and seems to play a key role in the pathogenesis of glaucomatous nerve atrophy. Damage of DNA, caused by oxidative stress, was shown in TM cells of glaucoma patients. This chapter gives a review about oxidative stress and its pathological alterations in the main outflow pathway—the trabecular meshwork—in glaucoma patients.

Keywords: glaucoma, trabecular meshwork, intraocular pressure, oxidative stress

1. Introduction

Glaucoma is known as a progressive neurodegenerative disease with irreversible loss of vision. Next to perimetric visual field defects, morphological alterations of the optic nerve and an increased intraocular pressure (IOP) occur. Additionally, special examinations were investigated in the last years for glaucoma diagnosis and follow-up [1–4].

 INTECH
 © 20

 open science | open minds
 Attribution

© 2016 The Author(s). Licensee InTech. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. [cc] BY IOP is a fine regulated, complex homeostasis of production of aqueous humor (AH) in the nonpigmented ciliary body and its outflow. About 80% of AH is drained throughout the trabecular meshwork (TM) [5], a sponge-like network of TM cells with glycoproteins, hyaluronic acid, elastic fibers, collagen and extracellular matrix (ECM). Any outflow resistance can be generated by a decreased pore size or any other alterations of TM, making it more rigid, resulting in an increased IOP. TM cells have the ability of different functions. Up to date it is not known, if cellular or intracellular disturbances are the origin of the altered TM in glaucoma. It is also conceivable that both mechanisms are impaired.

Oxidative stress is one mechanism, causing an altered extracellular matrix (ECM) [6] due to a disbalance of oxidants (free radicals, ROS) and antioxidants. If there is any imbalance between both with preponderance of oxidants, an irreversible cell loss is induced. Oxidative stress seems to have a key role in the pathogenesis of glaucomatous nerve atrophy [7, 8]. Damage of DNA, caused by oxidative stress, correlates significantly with IOP as well as with visual field defects [9, 10], and could be shown in TM cells in glaucoma patients [11]. Additionally, an increased lipid peroxidation, being the result of a ROS-induced cell damage, was noticed [12] as well as an impairment of metalloproteinases-2 (MMP-2) expression, resulting in an accumulation of ECM [13]. This accumulation (i.e., 'plaques') in TM could be seen several years before by electron microscopy studies [14, 15]. Additionally, actin expression can be induced by oxidative stress [16, 17], which is responsible for the rigidity of TM—the more amount of actin, the more rigid the TM [18].

In which way is oxidative stress induced, especially in glaucoma patients? Certainly, endogenous and exogenous factors contribute together. For example, daylight (especially ultraviolet light) can induce ROS in AH [19, 20]. Further on a vascular dysregulation [21] is discussed in glaucoma pathogenesis. An impaired blood flow with decreased nutritive supply can result in hypoxia, the main factor of inducing oxidative stress [22]. This hypothesis is supported by an increased concentration of vascular endothelial growth factor (VEGF) and erythropoietin in samples of AH in glaucoma patients [23, 24].

2. Oxidative stress and glaucoma

2.1. Oxidative stress: ROS and RNS

Oxidative stress was first described by Sies [25]. It is the result of an imbalance between oxidant (free radicals, reactive oxygen species and ROS) and antioxidant agents. Commonly, it is assumed that an excess of ROS, which follows from an increased production or decreased depletion, is the main reason for oxidative stress. Furthermore, a reduced antioxidant level can cause elevated ROS levels [26]. This imbalance with a preponderance of ROS results in a cytotoxicity with an irreversible cell loss.

The molecular basic for this damage can be found the reactivity of ROS. Free radicals are very aggressive molecules due to their unpaired electron [27]. They are generated under aerobic conditions. Excessive physical activities, infections or toxicity (e.g. cigarette smoke, UV

radiation), are only some of the ROS causing agents. Even microwave radiation was found to induce oxidative stress [28]. ROS production can be found mainly in mitochondria [29]. Electrons (e⁻) and protons (H⁺), which were released by oxidation of fats and carbohydrates, enter the 'electron transfer chain.' In this chain, several consecutive reactions transfer electrons from NAPH to O₂ (oxygen) via respiratory enzyme complexes [30]. 'NADH dehydrogenase complex' transfers the electrons from NADH via flavin and iron-sulfur centers to ubiquinone. At this point, the second complex 'cytochrome b-c1' passes the electrons to cytochrome c, containing the 'cytochrome oxidase complex'. This step forwards the electrons to react with O₂ [30]. Almost 98% of O₂ is reduced to H₂O. The remaining 2% were reduced only incompletely, resulting in the production of ROS [31]. Considering this fact, it is of interest that not only the biochemical alterations of free radicals, but also the cellular energy level (redox status) is important, when talking about oxidative stress [32].

Several molecules are subsumed in the ROS group, which are all intermediate products, originated from O_2 : hydrogen peroxide (H_2O_2), singlet oxygen ($\frac{1}{2}O_2$), hydroxyl radical (HO) and superoxide (O_2^-). Molecular O_2 reacts with a single electron, followed by the uptake of several protons. This reaction produces the reactive oxygen species [33]:

$$O_2 + e^- \to O_2 \cdot^- \tag{1}$$

$$O_2 \cdot H^+ \to HO_2 \tag{2}$$

$$HO_2 \cdot + e^- + H^+ \to H_2O_2 \tag{3}$$

$$H_2O_2 + e^- + H^+ \rightarrow HO + H_2O \tag{4}$$

$$HO \cdot + e^- + H^+ \rightarrow H_2O$$
(5)

If iron reacts with ROS, the production of further ROS is catalyzed (Fenton reaction, [33]).

$$H_2O_2 + Fe^{2+} \rightarrow OH^- + FeO^{2+} + H^+ \rightarrow HO^- + HO^- + Fe^{3+}$$
(6)

The hydroxyl radical is the most powerful radical, which is known up to now. It can cause membrane peroxidation and protein carbonylation [33], dectract or add electrons and hydrogen to the DNA, resulting in further radicals [34–36].

Organisms show diverse cellular escape mechanisms against ROS, which eliminates superoxide under normal conditions [29, 33, 37–39]:

$$2\mathbf{O}_2 \cdot \overline{\phantom{\mathbf{O}}} \to \frac{1}{2} \mathbf{O}_2 \tag{7}$$

$$2\mathbf{O}_{2}^{-} + \mathrm{SOD} \to \mathrm{H}_{2}\mathrm{O}_{2} + \mathrm{O}_{2} \tag{8}$$

$$H_{2}O_{2} + O_{2}^{-} \rightarrow O_{2} + HO + HO^{-}$$

$$H_{2}O_{2} + Katalase \rightarrow H_{2}O + \frac{1}{2}O_{2}$$
(10)

$$H_2O_2 + 2 GSH + GSH - Peroxidase \rightarrow GSSG + 2 H_2O$$
 (11)

SOD superoxide dismutase

GSH glutathione

GSSG glutathione disulfide

At 'normal' ROS levels, these free radicals are interacting with other cellular molecules in cell regulation, transcription and cell proliferation. Additionally, they are very important as protective mechanism against distinct bacteria or fungi [26, 29, 33]. However with increasing amounts, ROS changes its function from defensive to aggressive.

Next to ROS, N-containing radical molecules were known to have oxidative power [40, 41]. Reactive nitrogen species (RNS) are derived from the nitric oxide (NO). Free radicals react rapidly with NO, forming RNS [42, 43]:

$$O_2^{-} + NO \rightarrow ONOO^{-}$$
 (12)
ONOO^{-} peroxynitrite

Superoxide can be built by diverse reactions (see above) under different conditions. High concentrations of lipoproteins disturb the NOS pathway, which cause an increased amount of superoxide [25]. Peroxynitrite is not a stable molecule. It reacts very fast with almost all classes of biomolecules, thus being a potential cytotoxic agent [42]. It works similar to hydroxyl radicals, thus inducing DNA damages with breaks, nitrations and oxidations [36]. Unlike H_2O_2 peroxynitrite can be degraded without metal ions by inducing radicals [44, 45]:

$$ONOO^{-} + H^{+} \rightarrow OH + NO_{2}$$
(13)

Peroxynitrite can react with several molecules. Next to molecules with aromatic structures (e.g. alpha-tocopherol [46]), thiols [47] can be modified. These reactions generate further radicals, which can induce themselves peroxidation in lipids [48].

In which way does ROS or RNS accumulate? Is there an alteration at the protein or even gene level, which results in this imbalance? Which external factors induce this 'program?' The final answer to all these questions is open up to now. There are evident hints that several transcription factors are involved in this complex network (e.g. p53, [49]).

2.2. Oxidative stress: molecular interactions

ROS and RNS can react with even each biochemical molecule (e.g. lipids, proteins). Cell membranes, containing a high amount of phospholipids and triglycerides, are one of the targets [50, 51]. After peroxidation of the lipids, bond arrangement and nonenzymatic Hock cleavage start. The produced α , β polyunsaturated lipid aldehydes act as radicals in turn [52]. Thus, a reaction chain is triggered. Additionally, the membrane structure and integrity are disturbed [53]. In particular, nerve structures are even more sensitive for oxidative stress as showing a large amount of lipids and high energy requirement [54].

Additionally, free radicals attack the ion channels within the cellular [55] as well as intracellular membranes [56]. The conductance is significantly altered, resulting in an altered excitability of the membranes. After free radical attacks proteins change their backbone and/or side chain structure in that way, that it reacts itself with other amino acid side chains to build carbonyl chains. Unfolding, misfolding and consecutive loss of activity remains [57].

As third target, ROS/RNS attack DNA by purine or pyridine bases structure changes. Yet, protein cross-linking and breaks in the DNA string were detected [39, 58]. These mutations can concern nuclear as well as mitochondrial DNA (mtDNA). However, in particular, mito-chondrial DNA is affected, resulting in an impaired energy production [59, 60].

Why are mutations more dangerous, when affecting mitochondrial than nuclear DNA? Several hypotheses were discussed in this context. It is assumed that because of the local closeness of the enzyme complexes, which are involved in the electron transfer chain, next to the mitochondrial DNA, the produced ROS can attack the mtDNA more severe [61–63]. Additionally, mtDNA can replicate independently of the S-Phase, thus being more sensitive for oxidative agents, when the redox level is more oxidative [64], and free nucleotides are low [65]. Recent reports even suggest that hPrimpol1/CCDC111 (human primase-polymerase 1) is involved in DNA reparation due to its primase and DNA polymerase activity [66].

2.3. Oxidative stress: glaucoma

The influence of oxidative stress is discussed in the pathogenesis of several neurodegenerative diseases, like Morbus Parkinson or Morbus Alzheimer, as well as ophthalmic disorders, for example, glaucoma. The pathogenesis of glaucoma, being the second cause for blindness [67], is unknown up to now.

Glaucoma seems to be a multifactorial disorder. The most important risk factor for converting to glaucoma or glaucoma progression can be seen in the IOP [68, 69]. However, not all patients show a stable glaucoma disease after IOP regulation, and thus, several other factors seem to be involved. 'Vascular dysregulation' [21], glutamate exotoxicity [70], an interrupted retrograde transport of neutrophins [71] as well as ocular ischemia [72–74] and oxidative stress [7, 8, 75–77] are established in this discussion.

Glaucoma is characterized by an altered optic disc, visual field defects and an elevated intraocular pressure (IOP). As consequence of the increased IOP, retinal ganglion cells (RGC) and axons will die and cause the typical optic nerve appearance with glaucomatous excavation. Free radicals are known to induce oxidative stress in retinal ganglion cells, resulting in apoptosis of these cells [78]. Trapping ROS/RNS with NF-E2 related factor 2 (Nrf2), the death of RGC, seems to be prevented [79]. Several studies have been performed to investigate the potential influence of oxidative stress in glaucoma patients. It was shown that the L-arginine/ nitric oxide system is altered in patients with glaucoma [80]. Increased concentrations of superoxide dismutase, malondialdehyde and glutathione peroxidase could be detected in aqueous humor of patients with primary open-angle glaucoma (POAG) [81, 82]. Additionally, antioxidant activity was shown to be decreased in POAG [82]. If systemic antioxidant reserve is low, more severe perimetric defects occurred [83, 84]. Further on the DNA damage, caused by oxidative stress, correlates significantly with glaucomatous visual field defects and IOP [9, 10].

Up to now, there are evident hints for a critical role of oxidative stress in the pathogenesis of glaucoma, yet there are several questions unanswered. However, oxidative stress induces change within different ocular structures, whereas alterations in tissues, influencing IOP, are of special interest.

3. Intraocular pressure

Intraocular pressure is the result of aqueous humor (AH) production and outflow through trabecular meshwork and uveoscleral. This circular flow is regulated until a steady state arises. The production of AH starts at the non-pigmented ciliary body. Plasma of the vascular plexus is filtrated by diffusion, ultrafiltration and active secretion to build the AH. First of all, lipid-soluble molecules diffuse according to a gradient. Second, water and water-soluble molecules are ultrafiltrated because of an osmotic gradient and hydrostatic pressure [85]. Active secretion completes the process of AH production [86]. Electrolytes flow due to an electrochemical gradient [87] into the interstitial space, whereas protein transporters, ion channels, aquaporins and enzymes work together to pass molecules, ions or ascorbic acid through the blood-aqueous barrier [88–91]. Liquid diffusion is a consecutive result of the movement of electric charge [92], which is completed by the work of aquaporins.

The produced AH is similar to plasma, yet proteins are 200× less and ascorbic acid 20× higher than in plasma [93]. Additionally, endothelin-1 [94], several growth factors [95], enzymes

(collagenase) [96] or Immunoglobulin G [97], intrinsic glycoproteins [98] and transferrin [99] can be detected.

After production in the posterior part of the eye, AH flows through the pupil. Convection effectuates the movement of AH in the anterior chamber. AH circulates with an upward trend at the cornea and downstream flow at the iris [100]. The outflow of AH is provided by two pathways: The trabecular meshwork in the anterior chamber angle drains off the AH into the Schlemm's canal. The fluid passes multiple intrascleral collector canals until drainage into episcleral veins, ciliary veins and veins of the extraocular muscles [101, 102]. The second draining pathway is the uveoscleral outflow. Through the uveal meshwork and the anterior parts of the ciliary muscle, AH flows toward suprachoroidal space and sclera, where it is resorbed by several veins [103–105].

To generate the IOP distinct, resistance factors have to be considered: the conventional outflow pathway via trabecular meshwork with resistance of $3-4 \text{ mmHg/}\mu\text{l/min}$ and the episcleral vein pressure of 8-10 mmHg [106–108] results in IOP of about $15.5 \pm 2.57 \text{ mmHg}$ in normals [109–111].

Because the outflow through the trabecular meshwork (TM) is dependent on resistance of the trabecular meshwork, the factors, influencing resistance in TM, are of special interest in IOP.

4. Trabecular meshwork

In patients with POAG, the trabecular meshwork is characterized by an increased outflow resistance leading to an elevated IOP. It is assumed that the increased outflow resistance is attributed to various morphological and biochemical alterations of the TM of glaucomatous eyes. So have electron microscopic and immunohistochemical studies of TM cells demonstrated an increased accumulation of extracellular matrix, the so-called plaque material, in the juxtacanalicular region of the TM [112, 113]. An increased accumulation of fibronectin was found in the TM of glaucomatous donor eyes as compared to the TM of control eyes [114]. In addition, TM cells showed an increased cell loss as compared to age-matched control TM cells [115]. Later, Liton et al. have also demonstrated an accelerated senescence of TM in comparison with age-matched control donors [116].

The reasons for these changes are still not clear. One factor, which may play an important role in the pathogenesis of POAG, is oxidative stress [117]. Patients with POAG showed mutations in the mitochondrial genome and a reduced mitochondrial respiratory activity compared to control groups [118]. Also the antioxidative capacity in the aqueous humor of patients with POAG was found to be reduced by more than 50% compared to non-glaucomatous eyes [119]. Furthermore, an increased level of 8-hydroxy-2'-deoxyguanosine, a molecular biomarker for oxidative damage, was detected in TM samples collected from glaucoma patients undergoing standard filtration surgery [10]. The oxidative damage in these cells correlated with the visual field defects of these operated glaucoma patients [10]. Besides that, oxidative stress can also induce inflammation of the TM cells [8] and alter their mobility [120], which leads to contractile dysfunction and damage of the TM cells. Thus, these studies suggest that oxidative stress may be involved in the pathogenesis of POAG.

The human TM is a highly sensitive tissue to oxidative stress, since it is equipped with few antioxidant defense mechanisms [121, 122]. *In vitro* studies showed that increased levels of hydrogen peroxide stimulated the migration of TM cells, which lead to trabecular thickening and enlargement or collapse [123]. Furthermore, hydrogen peroxide could also cause endothelial cell loss and thus a disruption and collapse of the TM [124]. It is assumed that TM cell loss is caused by oxidative stress-induced apoptosis via inflammation, mitochondrial damage, hypoxia and endothelial dysfunction [125]. We could demonstrate that oxidative stress in form of hydrogen peroxide was able to induce cell death, extracellular matrix production, and accelerated senescence in cultured human TM cells. Based on these studies, it was postulated that oxidative stress may play a role in cell death, release of inflammatory markers, extracellular matrix accumulation, advanced senescence and disarrangement of the cytoskeleton of TM cells. These oxidative stress-induced TM changes may be responsible for a reduced outflow facility and thus an increased IOP.

Author details

Bettina Hohberger^{1*}, Ulrich-Christoph Welge-Lüßen¹ and Alice Yu²

*Address all correspondence to: Bettina.hohberger@uk-erlangen.de

1 Department of Ophthalmology, Friedrich-Alexander-Universität Erlangen-Nürnberg (FAU), Erlangen, Germany

2 Ophthalmology Center-Stachus, Munich, Germany

References

- [1] Brehmer K, Rutrecht P, Mißlinger S, Jünemann A, Horn FK, Kremers J, Hohberger B. 25 Hz adaptation: influence on recovery time in glaucoma. Ophthalmol J 2016;1:1–9.
- [2] Rutrecht P, Brehmer K, Kremers J, Horn FK, Jünemann A, Hohberger B. Temporal contrast sensitivity: a potential parameter for glaucoma progression, especially in advanced stages. Ophthalmol J 2016;1:10–17.
- [3] Kara K, Jünemann A, Redjak R, Hohberger B. Glaucoma: confrontation visual field test using Bagolini striated glasses—a new screening method for detecting visual field defects. Ophthalmol J 2016;1:18–23.

- [4] Horn FK, Scharch V, Mardin CY, Lämmer R, Kremers J. Comparison of frequency doubling and flicker defined form perimetry in early glaucoma. Graefes Arch Clin Exp Ophthalmol. 2016;25:937–946.
- [5] Gabelt BT, Kaufman PL. Changes in aqueous humor dynamics with age and glaucoma. Prog Retin Eye Res 2005;24:612–637.
- [6] Dumont P, Burton M, Chen QM et al. Induction of replicative senescence biomarkers by sublethal oxidative stresses in normal human fibroblast. Free Radic Biol Med 2000;28:361–373.
- [7] Tezel G. Oxidative stress in glaucomatous neurodegeneration: mechanisms and consequences. Prog Retin Eye Res 2006;25:490–513.
- [8] Aslan M, Cort A, Yucel I. Oxidative and nitrative stress markers in glaucoma. Free Radic Biol Med. 2008;45:367–376.
- [9] Gupta N, Yucel YH. Glaucoma as a neurodegenerative disease. Curr Opin Ophthalmol 2007;18:110–114.
- [10] Sacca SC, Pascotto A, Camicione P et al. Oxidative DNA damage in the human trabecular meshwork: clinical correlation in patients with primary open-angle glaucoma. Arch Ophthalmol 2005;123:458–463.
- [11] Izzotti A, Cartiglia C, De Flora S, Sacca S. Methodology for evaluating oxidative DNA damage and metabolic genotypes in human trabecular meshwork. Toxicol Mech Methods 2003;13:161–168.
- [12] Babizhayev MA, Bunin AYa. Lipid peroxidation in open-angle glaucoma. Acta Ophthalmol 1989b;67(4):371–377.
- [13] Welge-Lüssen U, Eichhorn M, Bloemendal H et al. Classification of human scleral spur cells in monolayer culture. Eur J Cell Biol 1998;75:78–84.
- [14] Babizhayev MA, Brodskaya MW. Fibronectin detection in drainage outflow system of human eyes in ageing and progression of open-angle glaucoma. Mech Ageing Dev 1989a;47:145–157.
- [15] Rauhut D, Rohen JW. Electron microscopic study of the trabecular meshwork in alphachymotrypsin glaucoma. Albrecht Von Graefes Arch Klin Exp Ophthalmol 1972;184:29–41.
- [16] Zhu J, Zhou K, Hao JJ et al. Regulation of cortactin/dynamin interaction by actin polymerization during the fission of clathrin-coated pits. J Cell Sci 2005;118:807–817.
- [17] Dalle-Donne I, Rossi R, Giustarini D et al. Actin carbonylation: from a simple marker of protein oxidation to relevant signs of severe functional impairment. Free Radic Biol Med 2001;31:1075–1083.

- [18] Tan JC, Peters DM, Kaufman PL. Recent developments in understanding the pathophysiology of elevated intraocular pressure. Curr Opin Ophthalmol 2006;17:168–174.
- [19] Balansky RM, Izzotti A, D'Agostini F et al. Systemic genotoxic effects produced by light, and synergism with cigarette smoke in the respiratory tract of hairless mice. 2003;24(9): 1525–1532.
- [20] Wei H, Ca Q, Rahn R et al. DNA structural integrity and base composition affect ultraviolet light-induced oxidative DNA damage. Biochemistry 1998;37:6485–6490.
- [21] Flammer J. The vascular concept of glaucoma. Surv Ophthalmol 1994;38:3–6.
- [22] Nakabayashi M. Review of the ischemia hypothesis for ocular hypertension other than congenital glaucoma and closed-angle glaucoma. Ophthalmologica 2004;218:344–349.
- [23] Alge CS, Priglinger SG, Neubauer AS et al. Retinal pigment epithelium is protected against apoptosis by alphaB-crystallin. Invest Ophthalmol Vis Sci 2002;43:3575–3582.
- [24] Cumurcu T, Bulut Y, Demir HD et al. Aqueous humor erythropoietin levels in patients with primary open-angle glaucoma. J Glaucoma 2007;16:645–648.
- [25] Sies H. Oxidative stress: from basic research to clinical application. Am J Med 1991;91:31–38.
- [26] Poljsak B, Suput D, Milisav I. Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants. Oxid Med Cell Longev 2013;2013:Article ID 956792.
- [27] Riley PA. Free radicals in biology: oxidative stress and the effects of ionizing radiation. Int J Radiat Biol 1994;65:27–33.
- [28] Chauhan P, Verma HN, Sisodia R, Kesari KK. Microwave radiation (2.45 GHz)-induced oxidative stress: whole-body exposure effect on histopathology of Wistar rats. Electromagn Biol Med 2017;36(1):20–30.
- [29] Dröge W. Free radicals in the physiological control of cell function. Physiol Rev 2002;82:47–95.
- [30] Alberts B, Johnson A, Lewis J, et al. Molecular Biology of the Cell. 4th edition. New York: Garland Science; 2002. Electron-Transport Chains and Their Proton Pumps. Available from: http://www.ncbi.nlm.nih.gov/books/NBK26904/
- [31] Pinazo-Durán MD, Zanón-Moreno V, Gallego-Pinazo R, García-Medina JJ. Oxidative stress and mitochondrial failure in the pathogenesis of glaucoma neurodegeneration. Prog Brain Res. 2015;220:127–153.
- [32] Lopez-Alarcona C, Denicola A. Evaluating the antioxidant capacity of natural products: a review on chemical and cellular-based assays. Anal Chim Acta 2013;763:1–10.
- [33] Imlay JA. Pathways of oxidative damage. Annu Rev Microbiol 2003;57:395–418.

- [34] Hutchinson F. Chemical changes induced in DNA by ionizing radiation. Prog Nucleic Acid Res. 1985;32:116–154.
- [35] Dizdaroglu M, Rao G, Halliwell B, Gajewski E. Damage to the DNA bases in mammalian chromatin by hydrogen peroxide in the presence of ferric and cupric ions. Arch Biochem Biophys 1991;285:317–324.
- [36] Kohen R, Nyska A. Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. Toxicol Pathol 2002;30:620–650.
- [37] Forth W, Henschler D, Rummel W, Starke K. General and Specific pharmacology and toxicology, 7th version, 1998. Heidelberg: Spektrum Akademischer Verlag.
- [38] Miao L, Clair DKS. Regulation of superoxide dismutase genes: implications in disease. Free Radic Bio Med 2009;47:344–356.
- [39] Gandhi S, Abramov AY. Mechanism of oxidative stress in neurodegeneration. Oxid Med Cell Longev 2012;2012:428010.
- [40] Rubbo H, Darley-Usmar V, Freeman BA. Nitric oxide regulation of tissue free radical injury. Chem Res Toxicol 1996;9:809–820.
- [41] McAndrew J, Patel RP, Jo et al. The interplay of nitric oxide and peroxynitrite with signal transduction pathways: implications for disease. Semin Perinatol 1997;21:351–366.
- [42] Beckman JS, Beckman TW, Chen J et al. Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. Proc Natl Acad Sci USA. 1990;87:1620–1624.
- [43] Crow JP, Beckman JS. The importance of superoxide in nitric oxide-dependent toxicity: evidence for peroxynitrite-mediated injury. Adv Exp Med Bio 1996;387:147–161.
- [44] Green SJ, Mellouk S, Hoffman SL et al. Cellular mechanisms of nonspecific immunity to intracellular infection: cytokine-induced synthesis of toxic nitrogen oxides from Larginine by macrophages and hepatocytes. Immunol Lett 1990;25:15–19.
- [45] Hou YC, Janczuk A, Wang PG. Current trends in the development of nitric oxide donors. Curr Pharm Des 1999;5:417–441.
- [46] Hogg N, Joseph J, Kalyanaraman B. The oxidation of alpha-tocopherol and trolox by peroxynitrite. Arch Biochem Biophys 1994;314:153–158.
- [47] Quijano C, Alvarez B, Gatti RM et al. Pathways of peroxynitrite oxidation of thiol groups. Biochem J 1997;322:167–173.
- [48] Radi R, Beckman JS, Bush KM, Freeman BA. Peroxynitrite-induced membrane lipid peroxidation: the cytotoxic potential of superoxide and nitric oxide. Arch Biochem Biophys 1991;288:481–487.

- [49] Szczepanek K, Lesnefsky EJ, Larner AC. Multi-tasking: nuclear transcription factors with novel roles in the mitochondria. Trends Cell Biol. 2012;22:429–437.
- [50] Bielski BH, Arudi RL, Sutherland MW. A study of the reactivity of HO2/O2- with unsaturated fatty acids. J Biol Chem 1983 Apr 25;258:4759–4761.
- [51] Gardner HW. Oxygen radical chemistry of polyunsaturated fatty acids. Free Radic Biol Med. 1989;7:65–86.
- [52] Benedetti A, Comporti M, Esterbauer H. Identification of 4-hydroxynonenal as a cytotoxic product originating from the peroxidation of liver microsomal lipids. Biochim Biophys Acta 1980;620:281–296.
- [53] Zhong H, Yin H. Role of lipid peroxidation derived 4-hydroxynonenal (4-HNE) in cancer: focusing on mitochondria. Redox Biol 2015;4:193–199.
- [54] Pearson JN, Patel M. The role of oxidative stress in organophosphate and nerve agent toxicity. Ann N Y Acad Sci 2016;1378(1):17–24.
- [55] Evans JR, Bielefeldt K. Regulation of sodium currents through oxidation and reduction of thiol residues. Neuroscience 2000;101:229–236.
- [56] Coronado R, Morrissette J, Sukhareva M, Vaughan DM. Structure and function of ryanodine receptors. Am J Physiol 1994;266:1485–1504.
- [57] Headlamand HA, Davies MJ. Markers of protein oxidation: different oxidants give rise to variable yields of bound and released carbonyl products. Free Radic Biol Med 2004;36:1175–1184.
- [58] Pisoschi AM, Pop A. The role of antioxidants in the chemistry of oxidative stress: a review. Eur J Med Chem 2015;97:55–74.
- [59] Krishnan KJ, Reeve AK, Samuels DC et al. What causes mitochondrial DNA deletions in human cells? Nat genet 2008;40:275–279.
- [60] Schon EA, DiMauro S, Hirano M. Human mitochondrial DNA: roles of inherited and somatic mutations. Nat Rev Genet 2012;13:878–890.
- [61] Wallace DC, Ye JH, Neckelmann SN et al. Sequence analysis of cDNAs for the human and bovine ATP synthase beta subunit: mitochondrial DNA genes sustain seventeen times more mutations. Curr Genet 1987;12:81–90.
- [62] Yakes FM, Van Houten B. Mitochondrial DNA damage is more extensive and persists longer than nuclear DNA damage in human cells following oxidative stress. Proc Natl Acad Sci U S A 1997;94:514–519.
- [63] Rajala N, Gerhold JM, Martinsson P et al. Replication factors transiently associate with mtDNA at the mitochondrial inner membrane to facilitate replication. Nucleic Acids Res 2014;42:952–967.

- [64] Klevecz RR, Bolen J, Forrest G, Murray DB. A genomewide oscillation in transcription gates DNA replication and cell cycle. Proc Natl Acad Sci U S A 2004;101:1200–1205.
- [65] Borst P, Kroon AM. Mitochondrial DNA: physicochemical properties, replication, and genetic function. Int Rev Cytol 1069;26:107–190.
- [66] Wan L, Lou J, Xia Y et al. hPrimpol1/CCDC111 is a human DNA primase-polymerase required for the maintenance of genome integrity. EMBO Rep 2013;14:1104–1112.
- [67] Kass MA, Heuer DK, Higginbotham EJ et al. The ocular hypertensin treatment study: a randomized trial determines that topical ocular hypotensive medication delays or prevents the onset of primary open-angle glaucoma. Arch Ophthalmol 2002;120:701– 713.
- [68] Sommer A. Intraocular pressure and glaucoma. Am J Ophthalmol 1989;107:186–188.
- [69] Garrat S. Primary Open Angle Glaucoma. Preferred Practice Pattern. San Francisco: American Academy of Ophthalmology; 1996.
- [70] Vorwerk CK, Lipton SA, Zurakowski D et al. Chronic low-dose glutamate is toxic to retinal ganglion cells. Toxicity blocked by memantine. Invest Ophthalmol Vis Sci 1996;37:1618–1624.
- [71] Quigley HA. Ganglion cell death in glaucoma: pathology recapitulates ontogeny. Aust N Z J Ophthalmol 1995;23:85–91.
- [72] Begg IS, Drance SM. Progress of the glaucomatous process related to recurrent ischaemic changes at the optic disc. Exp Eye Res 1971;11:141.
- [73] Hayreh SS. Posterior ciliary arterial occlusice disorders. Trans Ophthalmol Soc UK 1971;91:291–303.
- [74] Cioffi GA. Care guidelines and optic nerve assessment. J Glaucoma 1996;5:A12.
- [75] Levin L. Direct and indirect approaches to neuroprotective therapy of glaucomatous optic neuropathy. Surv Ophthalmol 1999;43:98–101.
- [76] Osborne NN. Pathogenesis of ganglion "cell death" in glaucoma and neuroprotection: focus on ganglion cell axonal mitochondria. Prog Brain Res 2008;173:339–352.
- [77] Kong GY, Van Bergen NJ, Trounce IA, Crowston JG. Mitochondrial dysfunction and glaucoma. J Glaucoma 2009;18:93–100.
- [78] Yokoyama Y, Maruyama K, Yamamoto K et al. The role of calpain in an *in vivo* model of oxidative stress-induced retinal ganglion cell damage. Biochem Biophys Res Commun 2014;451:510–515.
- [79] Himori N, Yamamoto K, Maruyama K et al. Critical role of Nrf2 in oxidative stressinduced retinal ganglion cell death. J Neurochem 2013;127:669–680.

- [80] Polak K, Luksch A, Berisha F, Fuchsjaeger-Mayrl G, Dallinger S, Schmetterer L. Altered nitric oxide system in patients with open-angle glaucoma. Arch Ophthalmol 2007;125:494–498.
- ^[81] Ghanem AA, Arafa LF, El-Baz A. Oxidative stress markers in patients with primary open-angle glaucoma. Curr Eye Res 2010;35:295–301.
- [82] Zanon-Moreno V, Marco-Ventura P, Lleo-Perez A et al. Oxidative stress in primary open-angle glaucoma. J Glaucoma 2008;17:263–268.
- [83] Tanito M, Kaidzu S, Takai Y, Ohira A. Correlation between systemic oxidative stress and intraocular pressure level. PLoS One 2015;10:e0133582.
- ^[84] Tanito M, Kaidzu S, Takai Y, Ohira A. Association between systemic oxidative stress and visual field damage in open-angle glaucoma. Sci Rep 2016;6:25792.
- [85] Civan MM, Macknight AD. The ins and outs of aqueous humour secretion. Exp Eye Res 2004;78:625–31.
- [86] Cole DF. Secretion of aqueous humor. Exp Eye Res 1977;25:161–176.
- [87] Gabelt BT, Kaufman PL. Aqueous Humor Hydrodynamics. In: Adler's Physiology of the Eye, 9th ed. 2003a.
- [88] Reddy VN. Dynamics of transport systems in the eye. Friedenwald Lecture. Invest Ophthalmol Vis Sci 1979;18:1000–1018.
- [89] Coca-Prados M, Sanchez-Torres J. Molecular approaches to the study of the Na+-K+-ATPase and chloride channels in the ocular ciliary epithelium. In: Civan MM, editor. The eye's aqueous humor. San Diego: Academic Press 1998.
- [90] Tsukaguchi H, Tokui T, Mackenzie B et al. A family of mammalian Na+-dependent Lascorbic acid transporters. Nature 1999;399:70–75.
- [91] Yamaguchi Y, Watanabe T, Hirakata A, Hida T. Localization and ontogeny of aquaporin-1 and -4 expression in iris and ciliary epithelial cells in rats. Cell Tissue Res 2006;325:101–109.
- [92] Tornquist P, Alm A, Bill A. Permeability of ocular vessels and transport across the bloodretinal-barrier. Eye 1990;4:303–309.
- [93] Gabelt BT, Kaufman PL. Aqueous Humor Hydrodynamics. In: Adlers's Physiology of the Eye. 10th ed. 2003.
- [94] Lepple-Wienhues A, Becker M, Stahl F et al. Endothelin-like immunoreactivity in the aqueous humour and in conditioned medium from cultured ciliary epithelial cells. Curr Eye Res 1992;11:1041–1046.

- [95] Cousins SW, McCabe MM, Danielpour D, Streilein JW. Identification of transforming growth factor-beta as an immunosuppressive factor in aqueous humor. Invest Oph-thalmol Vis Sci 1991;32:2201–2211.
- [96] Vadillo-Ortega F, Gonzalez-Avila G, Chevez P, Abraham CR, Montano M, Selman-Lama M. A latent collagenase in human aqueous humor. Invest Ophthalmol Vis Sci 1989;30:332–335.
- [97] McClellan BH, Whitney CR, Newman LP, Allansmith MR. Immunoglobulins in tears. Am J Ophthalmol 1973;76:89–101.
- [98] Haddad A, Laicine EM, de Almeida JC. Origin and renewal of the intrinsic glycoproteins of the aqueous humor. Graefes Arch Clin Exp Ophthalmol 1991;229:371–379.
- [99] Tripathi RC, Borisuth NS, Tripathi BJ, Gotsis SS. Quantitative and qualitative analyses of transferrin in aqueous humor from patients with primary and secondary glaucomas. Invest Ophthalmol Vis Sci 1992;33:2866–2873.
- [100] Heys JJ, Barocas VH. A boussinesq model of natural convection in the human eye and the formation of Krukenberg's spindle. Ann Biomed Eng 2002;30:392–401.
- [101] Worthen DM. Scanning electron microscopic study of the interior of Schlemm's canal in the human eye. Am J Ophthalmol 1972;74:35–40.
- [102] Tenner A, Jaeger W, Koch W. Demonstration of aqueous veins through injection of fluorescein into the anterior chamber in rabbits. Klin Monatsbl Augenheilkd 1974;164:628–632.
- [103] Bill A, Hellsing K. Production and drainage of aqueous humor in the cynomolgus monkey (*Macaca irus*). Invest Ophthalmol 1965;4:920–926.
- [104] Bill A, Phillips CI. Uveoscleral drainage of aqueous humour in human eyes. Exp Eye Res 1971;12:275–281.
- [105] Johnson M, Erickson K. Mechanisms and routes of aqueous humor drainage. In: Albert DM, Jakobiec FA, Eds. Principles and practice of ophthalmology. Philadelphia: WB Saunders; 2000.
- [106] Brubaker RF. Determination of episcleral venous pressure in the eye. A comparison of three methods. Arch Ophthalmol 1967;77:110–114.
- [107] Brubaker RF. The effect of intraocular pressure on conventional outflow resistance in the enucleated human eye. Invest Ophthalmol 1975;14:286–292.
- [108] Phelps CD, Armaly MF. Measurement of episcleral venous pressure. Am J Ophthalmol 1978;85:35–42.
- [109] Leydhecker W, Akiyama K, Neumann HG. Intraocular pressure of normal human eyes. Klin Monatsbl Augenheilkd 1958;133:662–670.

- [110] Schottenstein EM. Intraocular pressure. In: Ritch R, Shields MB, Krupin T, Eds. The glaucomas. St. Louis: Mosby 1989;301–317.
- [111] Millar C, Kaufman PL. Aqueous humor: secretion and dynamics. In: Tasman W, Jaeger EA, Eds. Duane's foundations of clinical ophthalmology. Philadelphia: Lippincott-Raven; 1995.
- [112] Rohen JW, Witmer R. Electron microscopic studies on the trabecular meshwork in glaucoma simplex. Albrecht Von Graefes Arch Klin Exp Ophthalmol 1972;183:251–266.
- [113] Lütjen-Drecoll E. Morphological changes in glaucomatous eyes and the role of TGFbeta2 for the pathogenesis of the disease. Exp Eye Res 2005;81:1–4.
- [114] Babizhayev MA, Brodskaya MW. Fibronectin detection in drainage outflow system of human eyes in ageing and progression of open-angle glaucoma. Mech Ageing Dev 1989;47:145–157.
- [115] Alvarado J, Murphy C, Juster R. Trabecular meshwork cellularity in primary openangle glaucoma and nonglaucomatous normals. Ophthalmology 1984;91(6):564–79.
- [116] Liton PB, Challa P, Stinnett S, Luna C, Epstein DL, Gonzalez P. Cellular senescence in the glaucomatous outflow pathway. Exp Gerontol 2005;40:745–748.
- [117] Izzotti A, Bagnis A, Sacca SC. The role of oxidative stress in glaucoma. Mutat Res 2006;612:105–114.
- [118] Abu-Amero KK, Morales J, Bosley TM. Mitochondrial abnormalities in patients with primary open-angle glaucoma. Invest Ophthalmol Vis 2006;47:2533–2541.
- [119] Ferreira SM, Lerner SF, Brunizini R, Evelson PA, Liesuy SF. Oxidative stress markers in aqueous humor of glaucoma patients. Am J Ophthalmol 2004;137:62–69.
- [120] Nathanson JA, McKee M. Alterations of ocular nitric oxide synthase in human glaucoma. Invest Ophthalmol Vis Sci 1995;36(9):1774–1784.
- [121] Izzotti A, Saccà SC, Longobardi M, Cartigl C. Sensitivity of ocular anterior chamber tissues to oxidative damage and its relevance to the pathogenesis of glaucoma. Invest Ophthalmol Vis Sci 2009;50(11):5251–5258.
- [122] Nita M, Grzybowski A. The role of the reactive oxygen species and oxidative stress in the pathomechanism of the age-related ocular diseases and other pathologies of the anterior and posterior eye segments in adults. Oxid Med Cell Longev 2016;2016:3164734.
- [123] Hogg P, Calthorpe M, Batterbury M, Grierson I. Aqueous humor stimulates the migration of human trabecular meshwork cells *in vitro*. Invest Ophthalmol Vis Sci 2000;41(5):1091–1098.

- [124] Zhou L, Li Y, Yue BYJT. Oxidative stress affects cytoskeletal structure and cell-matrix interactions in cells from an ocular tissue: the trabecular meshwork. J Cell Physiol 1999;180(2):182–189.
- [125] Saccà SC, Gandolfi S, Bagnis A, Manni G, Damonte G, Traverso CE, Izzotti A. From DNA damage to functional changes of the trabecular meshwork in aging and glaucoma.
 Ageing Res Rev 2016;29:26–41.





IntechOpen