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Polarisation of Macrophage and Immunotherapy in the Wound Healing

Yu-Sheng Wu, Fan-Hua Nan, Sherwin Chen and Shiu-Nan Chen

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Abstract

Immune cells are involved in virtually every aspect of the wound repair process, from the initial stages where they participate in haemostasis and work to prevent infection to later stages where they drive scar formation. Immunotherapy is being developed offers some advantageous immunomodulation factors that are known in the field of alternative medicine, such as mushroom beta-glucan, anti-microbial peptides and triterpenoid; these factors represent a novel therapeutic approach for anti-inflammation to promote the wound healing.

Keywords: healing, immunotherapy, inflammation, macrophage, polarisation, wound

1. Inflammation

When an organism is injured by a wound injury or infected by a pathogen, inflammation is a crucial response. Inflammation is a complex interaction with molecular mediators; it includes the function of immune cells in a microenvironment through a response that occurs at all levels of biological organisation [1]. Following previous studies, this paper illustrates that the inflammation response involves cooperation between cells and a wide range of mediators, such as cytokines, chemokines and non-enzyme factors involved in the classical immune response. The macrophage is one of the critical inflammatory immune cells involved in the uptake and degradation of infectious agents and senescent cells and also plays critical roles in tissue growth, tissue remodelling and inflammation by producing oxidants, proteinases and anti-microbial peptides [2–4]. Activated inflammatory cells are sources of reactive oxygen

species (ROS) and reactive nitrogen species (RNS) that can initiate changes in cell functions, including cell signalling pathways, transcription factor activation, mediator release and apoptosis. However, whether the ROS and RNS that are produced and released by neutrophils or macrophages are sufficient to diffuse through the extra-cellular matrix, enter epithelial cells and cross the cytoplasm is not clear [5–7]. Even the physiological roles of ROS and RNS in the cellular response are not clear [8–11]. The results obtained from experiments performed on the livers of tilapia showed that extra-cellular hydrogen peroxide (H_2O_2) attracted cell migration. These results suggested that ROS is a crucial factor in initiating the migration of macrophages that trigger cascades of phagocytic activity.

In the microenvironment of inflammation, the platelet-derived growth factor (PDGF), the tumour necrosis factors (TNF)- α and TNF- β , the hepatocyte growth factor, transforming growth factor (TGF)- β_2 , the epidermal growth factor (EGF) and the fibroblast growth factor all play an important role in physiological immune response. The interleukins (IL)-1, IL-6, IL-8, IL-10, and the interferon gamma (INF- γ) also detain key functions in the natural inflammatory response [12–16]. These factors hold a primordial function in fibroblast activation and regulation, also concerning reactive fibrosis that follows their continuing activation. Although these growth factors are also related to fibroblast migration and activation, particular research was recently focused on the PDGF family of growth factors and their relative receptors [17, 18]. Research has documented that PDGF exerts autocrine, mitogenic effects on keratinocytes to support epidermal proliferation and stabilisation of the epidermal junction during wound closure. In addition, it stimulates vessel maturation by recruiting and differentiating pericytes to the immature-endothelial channel [19–22]. According to these references, we investigate whether the produced ROS/RNS is related to the released factors and (if so) what type of relationship exists among ROS/RNS and these factors.

2. Reactive oxygen species production and physical response

The production and scavenging of ROS may be initiated by adverse environmental factors. Research has shown that intra-cellular levels of ROS may rapidly rise and ROS may be generated by the activation of various oxidases and peroxidases in response to certain environmental changes [23]. ROS forms through energy transfer or through electron transfer reactions. ROS formation causes the formation of singlet oxygen, which results in sequential reduction to superoxide, H_2O_2 and hydroxyl radicals [24]. Mitochondria are a crucial source of ROS production in most cells. This ROS production contributes to mitochondrial stress and plays a critical role in redox signalling from the organelles [25]. Mitochondria have a 4-layer structure composed of the outer mitochondrial membrane, intermembrane space, inner mitochondrial membrane and matrix [26]. NADPH oxidase is an enzymatic source in the mitochondrial structure that generates ROS and plays a fundamental role in maintaining normal cell functions. Recent research has focussed on the influence of this enzyme to cellular oxidative stress that may contribute to various pathophysiological conditions and diseases [27, 28]. A crucial function of NADPH oxidase is modulating multiple redox-sensitive intra-cellular signalling pathways; NADPH modulates these pathways by generating ROS molecules,

inhibiting protein tyrosine phosphatases and activating certain redox-sensitive transcription factors. Moreover, the ROS consist of numerous molecular species, including H_2O_2 , oxide ions (O_2^-) and hydroxide (OH^-) [29]. Molecular oxygen is a biradical, containing two unpaired electrons in the outer structure; because these two electrons have the same spin, oxygen can only react with one electron; therefore it is not very reactive when these two electrons have the same spin. Oxygen's unpaired electrons can become excited and can change the spin of one electron. This transforms oxygen into a powerful oxidant because the two electrons with opposing spins can rapidly react with other pairs of electrons [30]. Electrons can be contributed from NADH and FADH₂ enzymes and can pass through the electron transport chain, generating superoxide (O_2^-) at complexes I and III. This generated superoxide can be reduced to H_2O_2 by superoxide dismutase and can be completely reduced into water by glutathione peroxidase, as presented in **Figure 1**.

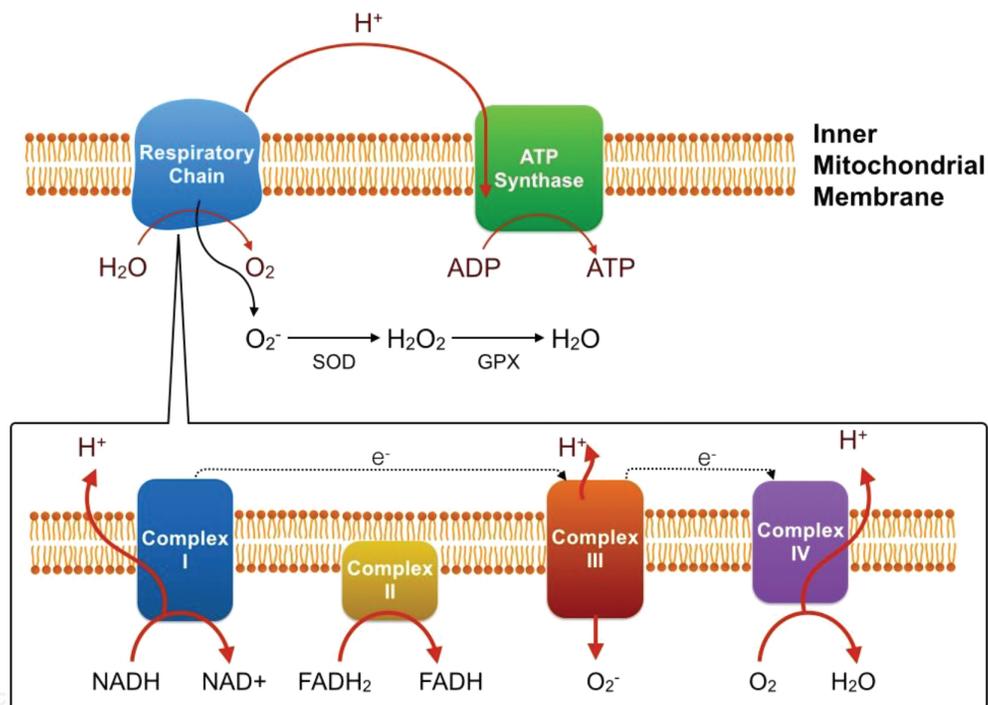


Figure 1. ROS are produced from the electron transport pathway to form superoxide (O_2^-) at complex I and complex III released into the matrix and reduced O_2 to form H_2O at complex IV. Following, the generated O_2^- is transferred to the form of H_2O_2 by superoxide dismutase (SOD) and completely reduced to water (H_2O) by glutathione peroxidase (GPX).

Research has shown that ROS consist of numerous molecular species, including H_2O_2 , oxide ions (O_2^-) and OH^- [29]. These molecular species act as signalling molecules in the migration of profibrogenic cells [31] and peripheral blood monocytes [23, 32]. One of the crucial physiological functions of ROS is the modulation of ion channels. Research has illustrated that ROS may act through Ca^{2+} as an intra-cellular second messenger involved in regulating diverse functions, such as fertilisation, electrical signalling, contraction, secretion, memory, gene transcription and cell death [33, 34]. Furthermore, studies have reported that H_2O_2 may affect

cell energy stores [35], induce DNA strand breaks [36], enhance cell adhesion [37], increase endothelial tissue permeability [38] and stimulate the release of cytokines.

In the research presented in **Figure 2**, the concentration of ROS seems to be considered the concentration of a crucial signalling molecule. Low concentrations of generated ROS are believed to be critical for metabolic adaptation in the organelle. Moderate concentrations of ROS can be produced and released by stress; pathogen-infected and bacterial endotoxin lipopolysaccharide (LPS) are involved in the inflammatory response. The high concentration of ROS in the induced apoptosis/autophagy process can cause cell death [39] and initiate self-healing [40].

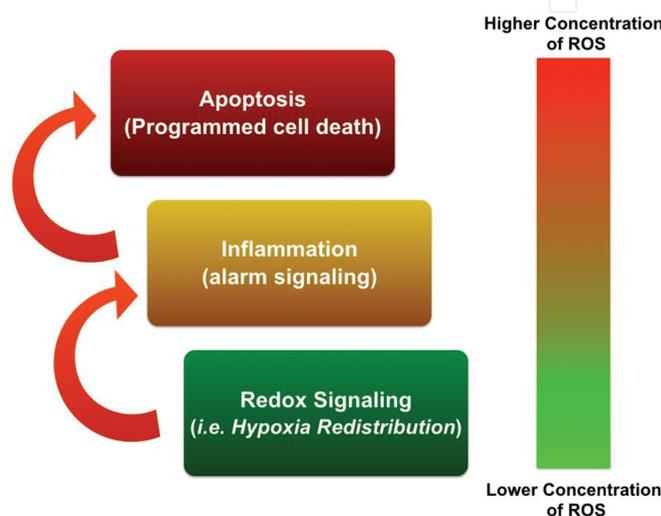


Figure 2. Concentration of generated ROS may involve in the different physiological response. At the low concentration, the ROS regulate in the redox signalling, and at the moderate concentration of induced ROS which participated in the inflammation process. At the high level of ROS concentration increased and was to be involved in the cellular apoptosis.

3. Tissue resident macrophages

Macrophages, which are present in almost all body tissue and display distinct location-specific phenotypes and gene expression profiles, display remarkable functional diversity in innate immune responses, tissue development and tissue homeostasis [41]. In different organs, the resident macrophages are given various appellations: microglia cells have fundamental importance in assessing the pathogenetic significance of perivascular inflammatory phenomena within the brain [42]; Kupffer cells are resident and recruited macrophages that play major roles in the homeostatic function of the liver and in its response to tissue damage [43]; alveolar macrophages are key determinants pulmonary immune responses and in the lung inflammation caused by asthma [44]. Previously, it was hypothesised that tissue macrophages were recruited from circulating blood monocytes. Recent studies have demonstrated that tissue macrophages such as microglia, Kupffer cells and Langerhans cells are established prenatally

and arise independently of the hematopoietic transcription factor Myb [45]. Myb is required for developing hematopoietic stem cells (HSCs) and all CD11b^{high} monocytes and macrophages but is not required for yolk sac (YS) macrophages and for developing YS-derived F4/80^{bright} macrophages. Such macrophages can persist independently of HSCs in several types of tissue in adult mice [46]. Kupffer cells as well as other resident macrophages (e.g., microglia) originate from the YS in a colony-stimulating factor-1/receptor (CSF-1R)-dependent and Myb-independent manner. Researchers have suggested that these macrophages are maintained by local proliferation, which results in extensive mitosis after stress or an exchanged tissue microenvironment [43, 47].

Macrophages are the most crucial and abundant immune cells. They can be categorised into two primary types according to function and differentiation: classically activated macrophages (M1 macrophages) and alternatively activated macrophages (M2 macrophages) [48]. Macrophages are relevant to innate resistance and to the relationship between inflammation and autoimmune disease. In mouse models, macrophages present CD11b, F4/80 and CSF-1R, with F4/80 being the surface proteins for M1 and M2 macrophages [49, 50]. When pathogens enter the organism from the intestinal portal vein, circulating monocytes (surrounding the pathogens and present in the peripheral blood) respond to chemokines (e.g., CCL2) and are exposed to antigens. While interacting with pattern recognition receptors (PRRs), antigens may exert either M1 or M2 polarising activities, depending on the Th1 (IFN- γ) and Th2 (IL-4 and IL-13) cytokines and immune factors [51, 52].

4. Inflammation and macrophages

Inflammation is an important adaptive physiological response of the organism. Inflammation response embodies a complicated interaction among molecular mediators and cells. It globally affects the leukocytes, also the lymphocytes in their micro-environmental function and organisation [48]. Throughout their response, numerous factors are involved in the classical immune response. Macrophages detain a critical role in the uptake and degradation of infectious agents and senescent cells; they also play crucial roles in tissue growth, tissue remodelling and inflammation by producing oxidants, proteinases and anti-microbial peptide [40].

Resident macrophages sense exogenous or endogenous danger signals (e.g., bacterial products or necrotic cell debris) through PRRs. In response to Toll-like receptor (TLR) ligands and interferon-gamma (IFN- γ) or IL-4/IL-13, macrophages undergo M1 (classical) or M2 (alternative) activation. The activation of M1 and M2 macrophages mirrors TH1-TH2 polarisation; M1 and M2 activation span the extremes of a continuum. M1 macrophages, which display a morphology that depends on their tissue location, develop in response to stimulation with IFN- γ and microbial products such as LPS. M1 macrophages can secrete substantial amounts of pro-inflammatory cytokines, such as IL-1 β , IL-15, IL-18, TNF- α and IL-12 [53]. M2 macrophages adapt to similarities and differences between IL-4, TLR ligands with IL-10 and glucocorticoids [54].

The phenotypes of M1 and M2 macrophages exhibit observable differences. The M1 phenotype is characterised by the expression of high levels of pro-inflammatory cytokines, high production of reactive nitrogen and oxygen intermediates, promotion of Th1 response and anti-microbial and tumour-inhibiting activity [43]. The M2 macrophage uses immune inhibitory effects to secrete large amounts of IL-10, TGF- β , and C-C motif chemokine ligands 17 (CCL17) and CCL22. Moreover, the M2 macrophage attracts non-cytotoxic T_{reg} and Type 2 T-helper cells (TH2 cells) to aggregate in tumour tissue, inhibit T-cell differentiation and function, lower cytotoxic T-cell function, induce T-cell apoptosis, secrete CCL18 and attract naive T cells [55].

Macrophage	M1	M2
Transcription factor		
Interferon regulatory factor (IRF)	IRF-3 [61, 62]	IRF-4 [63]
	IRF-5 [64]	
	IRF-8 [65]	
Nuclear factor	NF- κ B [43]	
Signal transducer and activator of transcription (STAT)	STAT-1 [66]	STAT-3 [43]
		STAT-5 [67]
		SATA-6 [68]
Suppressor of cytokine signalling (SOCS)	SOCS-1	
	SOCS-2	
	SOCS-3	
	(controversial) [58]	
Phenotype	iNOS [69, 70]	YM-1 [71]
	IL-6 [72]	Arg-1 [73, 74]
	TNF- α [75]	Fizz-1 [76]
		IL-10 [77]

Table 1. Regulators in the M1 and M2 macrophage.

Macrophage polarisation is highly related to expressions of various TLRs on macrophages [56, 57]. The evidence indicates that TLR signalling (e.g., TLR4), which is activated by LPS and other microbial ligands, drives macrophages to prefer the M1 phenotype. In this reaction, MyD88 and TRIF activate a cascade of kinases, including IRAK4, TRAF6 and IKK β ; this results in the activation of nuclear factor kappa B (NF- κ B), which drives the macrophage forward to the M1 phenotype. By contrast, IL-4 and IL-13 drive the macrophage's phenotype forward to M2. Activation of STAT6 through the IL-4 receptor alpha (IL-4R α) and IL-10 induce activation of STAT3 through receptor IL-10R, which activates JAK1 and JAK3 (38), causing STAT6 activation [58, 59]. IL-10, TGF- β , IL-4 and IL-13 enhance inflammation and cellular immune response with NO, which is generated through IFN- γ -induced iNOS and is reduced in macrophages by Arg1 interactions with mast cells, basophils, eosinophils, NKT cells, IgE and selected subclasses of IgG. This promotes allergies and hypersensitivity [60] (**Table 1**).

5. Inflammation and disease

Accumulating evidence indicates that chronic low-grade inflammation contributes to the systemic metabolic dysfunction that is associated with inflammation disorders [78]. Cytokines and pathogen-associated molecular patterns have been shown to co-stimulate cell surface receptors, including TLRs, to initiate intra-cellular signalling that activates NF- κ B. NF- κ B activation was thought to induce the target gene's expression to promote cellular proliferation and to activate the immune response. However, research has revealed that NF- κ B activation can occur in most cell types; recent reports have demonstrated that high level activation of NF- κ B signalling pathways in the liver, adipose tissue and central nervous system (CNS) is involved in the development of inflammation-associated metabolic diseases [79]. The mutants of the brain-specific serpin, neuroserpin, also form ordered polymers that accumulate within the ER of neurons; these mutations cause an autosomal-dominant type of dementia known as familial encephalopathy with neuroserpin inclusion bodies, which is believed to be an inflammation disorder [80, 81].

Research has shown that, in specific tissue lesions, extra-cellular lipid droplets are forming a core region surrounded by smooth muscle cells and collagen-rich matrix. Lymphocytes as the T cells, monocyte, macrophages and mast cells are infiltrating in the lesion particularly in regions where the atheroma grows. These immune cells also generate important signals in the defence cascade by producing the inflammatory cytokines, largely involved in the atherosclerotic process [82]. A case report indicated that Alzheimer's disease (AD) inflammation appears to arise from within the CNS. Little or no involvement of lymphocytes or monocytes in AD was observed beyond their normal brain surveillance. This observation has placed AD outside the realm of conventional neuroimmunologic studies that largely focus on humoral aspects of such CNS inflammatory disorders as multiple sclerosis [83]. Judging from published reports, we believe that metabolic disorders and even neuronal diseases are highly related to abnormal inflammation.

6. Macrophage and T-cell differentiation

In pathogen infection, dendritic cells (DCs) and macrophages primarily act as phagocytotic antigen-presenting cells (APCs) that degrade infected pathogens into fragments, and then move those fragments to the nearby lymphoid organs. The pathogen fragments combine with cell surface histocompatibility complex (major histocompatibility complex) to activate and differentiate T cells. **Figure 3** displays the cooperation of the antigen-presenting cells, co-stimulatory molecules and cytokines.

The metabolic organs, such as the liver, pancreas and adipose tissue, are composed of parenchymal and stromal cells, which include macrophages to maintain metabolic homeostasis. Bacterial infection innately activates macrophages, causing the secretion of proinflammatory cytokines, such as TNF- α , IL-6 and IL-1 β . This promotes peripheral insulin resistance and reduces nutrient storage during the metabolic reaction. Furthermore, some additional

physiological mechanism can lead to the activation of macrophages. For these latest, the regulatory T cells (T_{reg}), the $Fc\gamma$ receptors, the apoptotic cells and the prostaglandins are increasing the number of macrophages involved in the regulation of inflammation and anti-tumour defences [84]. These inflammatory mediators are involved in activating anti-microbial defence mechanisms, including oxidative processes that contribute to killing pathogens and the secreted IL-12 and IL-23. These direct the differentiation and expansion of anti-microbial T_H1 and T_H17 cells that help to drive inflammatory responses [85]. Recent research shows that intestinal antigen-presenting cells can be divided into $CD11c^+CD11b^-$, $CD11c^+CD11b^+$ and $CD11c^{dull}CD11b^+$ categories. Particularly, the $CD11c^{dull}CD11b^+$ cells are $CD103-F4/80^+$ macrophages, with efficient role in inducing the $Foxp3^+$ regulatory T (T_{reg}) cells [86]. Tumour cells affect the surrounding cellular environment by promoting tumour growth and metastasis by establishing a tumour microenvironment that is conducive to tumour development [87–90]. In the tumour microenvironment, tumour cells secrete inflammatory cytokines, such as TGF- β and IL-10. These cytokines stimulate differentiation of regulatory T and T_{reg} cells [91, 92] as well as differentiation of tumour-associated macrophages (TAMs) into M2 macrophages. This causes the host immune system to locate and attack cancer cells, which generates subsequent tumour cell evasion of this immune surveillance and attack, which enhances tumour growth and metastasis [87, 93–98]. Various cytokines, chemokines and growth factors in the tumour microenvironment are the primary elements that affect the host's anti-tumour ability and evasion of tumour cells [89, 99]. Tumour microenvironments are complicated cellular microcosms [89, 97], and numerous immune cells are located throughout tumour microenvironments. Macrophages are the most crucial and abundant immune cells in the tumour microenvironment. The two most critical types of macrophages, based on function and differentiation, are M1 and M2 macrophages. M1 macrophages are characterised by tumour

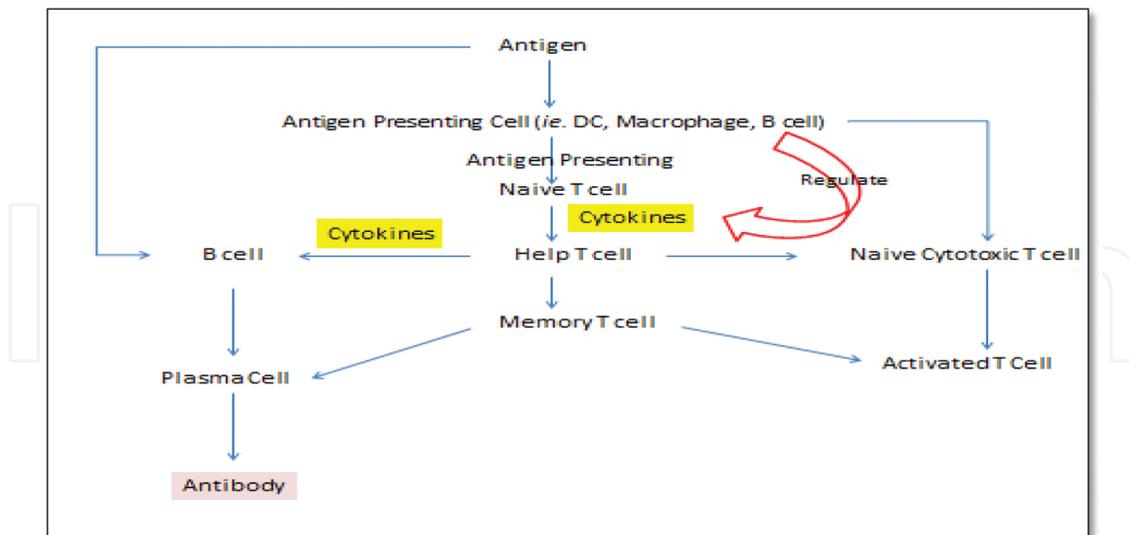


Figure 3. The cooperation of the antigen-presenting cells, costimulatory molecules, and cytokines. Bacterial infection innately activates macrophages, causing the secretion of pro-inflammatory cytokines, such as TNF- α , IL-6, and IL-1 β . This promotes peripheral insulin resistance and reduces nutrient storage during the metabolic reaction. Furthermore, several additional mechanisms can also contribute to the activation of macrophages for immune-regulatory activity.

resistance, whereas M2 macrophages are characterised by tumour promotion [98, 100]. In mouse models, macrophages present CD11b, F4/80, CSF-1R and F4/80 as the surface proteins for M1 and M2 macrophages [93, 101]. Recent studies have noted large quantities of TAMs in tumour tissue. TAMs are the most abundant and critical immune cells in the tumour microenvironment [102–104] and are the main factors that enable the tumour microenvironment to exert immune inhibitory effects [101, 102]. In the tumour microenvironment, tumour cells and the surrounding stroma cells secrete cytokines and growth factors that stimulate TAMs and activate the various expression, function, receptor regulation and secretion types of chemokines [103, 105], including anti-tumour M1 macrophages and pro-tumour M2 macrophages [98, 106–108]. In the tumour microenvironment, the proportions of M1 and M2 macrophages are unequal. Tumour microenvironments contain large amounts of transmitters, such as M-CSF, IL-6, IL-10, TGF- β and COX-2, that induce transformation of TAMs into M2 macrophages that secrete immune inhibitory chemokines and have poor antigen-presenting and cytotoxic abilities, which generates tumour growth and metastasis [49, 98, 102–104, 109–114]. M2 macrophages and TAMs have protumour and immune inhibitory effects, secrete large amounts of IL-10, TGF- β , CCL17 and CCL22, attract non-cytotoxic T_{reg} and TH2 cells to aggregate in tumour tissue, inhibit T-cell differentiation and function, lower cytotoxic T-cell function, induce T-cell apoptosis, secrete CCL18 and attract naïve T cells [49, 98, 115]. NADPH oxidase is a major enzymatic source of cellular ROS. NADPH plays a fundamental role in maintaining normal cell functions. Recent research has focussed on this enzyme's role in cellular oxidative stress, which may eventually contribute to various pathophysiological conditions and diseases [27, 28]. Studies have found that NADPH oxidase modulates multiple redox-sensitive intra-cellular signalling pathways by generating ROS molecules. This modulation includes inhibition of protein tyrosine phosphatases and activation of certain redox-sensitive transcription factors [116, 117]. ROS consist of numerous molecular species, including H₂O₂, oxide ions (O₂⁻) and OH⁻²⁹, that act as signalling molecules involved in the migration of hepatic profibrogenic cells [118] and the functioning of peripheral blood monocytes [119]. ROS and RNS, generated endogenously or in response to environmental stress, have long been implicated in tissue injury for a variety of disease states [120, 121]. Stimulation of the mitochondrial apoptotic pathway through ROS and mitochondrial DNA damage promotes outer membrane permeabilisation, which triggers caspase-dependent or caspase-independent cytosolic signalling events [122]. Activated inflammatory cells serve as sources of ROS and RNS that can initiate the alteration of the cell function, gathering specific cellular signalling, transcription factor activation, physiological factors release, the apoptosis process and compensatory cell proliferation. However, it remains unclear whether the ROS or the RNS production and release through neutrophils or macrophages enhance sufficient diffusion into the intra-cellular cytoplasm as to affect the cellular response [123, 124].

7. Wound healing

Immune cells are involved in virtually every aspect of the wound repair process, from the initial stages, where they participate in haemostasis and work to prevent infection, to later

stages where they drive scar formation [125, 126]. T lymphocytes exercise crucial *in vivo* effects on various parameters of healing [127–129]. Neutrophils help control infection during wound healing, but they also release harmful enzymes that damage healthy tissue surrounding the wound site [130–132]. Recent researchers have noted that several specific proteins produced by wound macrophages at the site of injury are involved: (1) in the recruitment and activation of additional macrophages infiltrating in the wound; (2) in the production of growth factors that promote cellular proliferation and tissue recovery synthesis; (3) in stimulating proteases and extra-cellular matrix growth and (4) in the process of tissue remodelling [133]. β -catenin-dependent Wnt pathways, which are classified according to their ability to promote stabilisation of β -catenin in the cytoplasm, act as cellular signals through cytoplasmic stabilisation and accumulation of β -catenin in the nucleus to activate gene transcription [134]. This could enhance wound healing by lymphocytes [135, 136]. Nicotinamide adenine dinucleotide phosphate (NADPH) oxidase modulates multiple redox-sensitive intra-cellular signalling pathways by generating ROS molecules. This includes inhibiting protein tyrosine phosphatases and activating certain redox-sensitive transcription factors [116, 137, 138]. This shows that ROS regulate the expression of key chemical mediators that further modulate the inflammatory response in animal models; it has also been reported that these redox-sensitive processes may include cytokine action, angiogenesis, cell motility and extra-cellular matrix formation [139–141]; this can enable reliable estimates of wound-healing capacity, which is altered by various conditions, such as inflammation. Furthermore, research on one of the ROS has indicated that H_2O_2 plays a critical role in wound repair, inflammation and anti-inflammation mechanisms [142, 143]. Our published research also showed that the production of ROS (i.e., H_2O_2 after an injury has occurred) may cause healing to generate inflammation through the apoptosis of the cell. Over-inhibition of NADPH oxidase activity may reduce the normal progress of apoptosis under the wound and might delay healing [29].

Inflammation enhances vascular permeability, active migration of blood cells and the passage of plasma constituents into the injured tissue [144]. Blood leukocytes actively participate in the defence and inflammation responses, being activated since the earliest phases of atherosclerosis process. Inflammation and atherosclerosis shelter intricate mechanisms relied to leukocytes recruitment [145]. Neuro-inflammation mediators are described to be closely related to brain cells functioning (such as microglia and astrocytes), to the complement system activation and to cytokines, and chemokines production [146]. Regarding cancer development [147], pro-inflammatory cytokines, including IL-1 α , IL-1 β , IL-6, IL-8, IL-18, chemokines, matrix metalloproteinase-9 and vascular endothelial growth factor, are primarily regulated by the transcription NF- κ B, which is active in most tumours and is induced by carcinogens [148]. Cutaneous wound repair is a tightly regulated and dynamic process involving blood clotting, inflammation, formation of new tissue and tissue remodelling [149]. Thrombin is the protease involved in blood coagulation. Its deregulation can cause haemostatic abnormalities, which range from subtle subclinical problems to serious life-threatening coagulopathies (i.e., during septicæmia) [150]. Inflammation and coagulation are both parts of the natural mechanism that protects the organism against infection. The endothelial cells and the platelets are capable to react in the acute, also in the chronic inflammatory environment. They release pro-inflammatory mediators that produce adhesion of molecules, proteases and clotting factors associated

to leukocytes recruitment [151]. The elements of the PAR family serve as sensors that detect blood-clotting serine proteinases in the inflamed target cells. Activation of PAR-1 by thrombin and of PAR-2 by other factors on the membrane of endothelial cells generates rapid expression and exposure of adhesive proteins that mediate an acute inflammatory reaction and of the tissue factor that initiates the blood coagulation cascade [152] as presented as **Figure 4**.

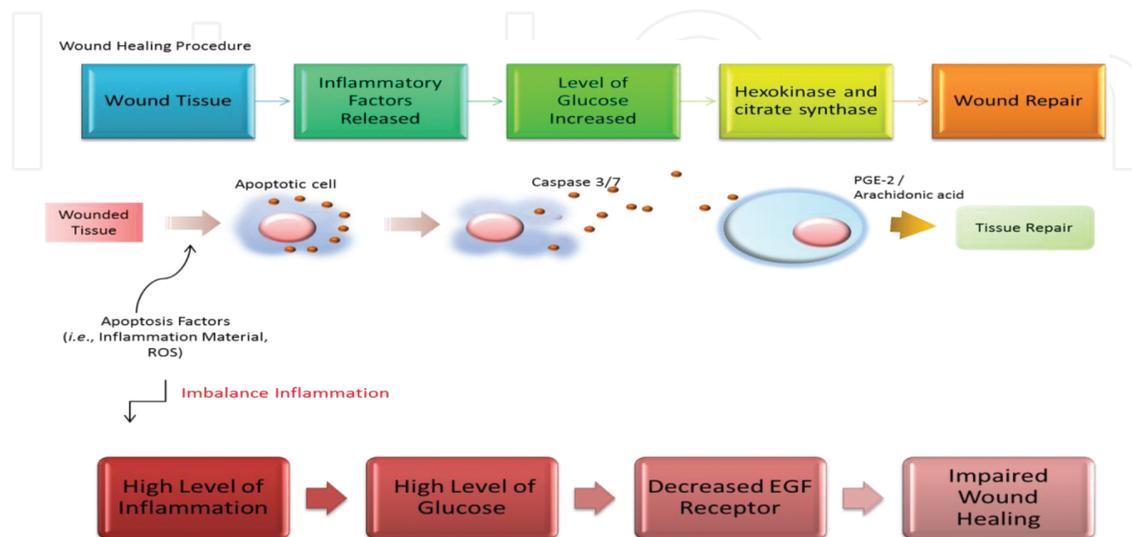


Figure 4. Wound healing was initiated after the injury of the cell, tissue even the organ. In the early stage of the healing, the damaged tissue producing a lot of ROS leading to neighbour cells into the apoptosis, following the apoptotic cells collapsed and released caspases were able to induce the tissue repair. However, the imbalance inflammatory may induce over-production of blood glucose that is leading to decrease the EGF receptor expression further to impair the wound healing.

8. Immunomodulation in anti-inflammation therapy

Nakanishi et al. found that celecoxib can alter the immune inhibitory effects of the tumour microenvironment by promoting transformation of TAMs into M1 macrophages, inhibiting tumour growth [153]. In 1968, Ikekawa et al. found that the fruiting body extracts from *Lentinus edodes*, *Trametes versicolor*, *Ganoderma tsugae*, *Flammulina velutiper* and *Tricholoma matsutake* demonstrated substantial anti-tumour activities towards transplanted tumour cells of Sarcoma 180 [154, 155]. *Autrodia comphorata*-derived beta-glucan inhibited tumour growth for Sarcoma 37, Sarcoma 180, Erlich ascites sarcoma and Yoshida sarcoma as well as inhibited LLC1 transplanted tumour growth [156]. Daily intake of *A. comphorata*-derived beta-glucan for 18 consecutive days was demonstrated to slow tumour growth and reduce the rate of metastasis [157]. Cytotoxic T-cell activity and tumour occurrence rates were observed, and the results illustrated that daily oral intake of *Grifola frondosua*-derived beta-glucan or Lentinan can enhance cytotoxic T-cell activity and reduce tumour occurrence rates [158]. The addition of a conditioned medium along with tumour cells into the progenitor cells of DCs was found to further inhibit maturation of DCs and lower the antigen-presenting capability of the DCs [159]. Tumour cells were found to secrete M-CSF, inhibiting dendritic and T-cell differentiation and

anti-tumour ability [87, 159–161]. In the inflammation environment, the amounts of M1 and M2 macrophages are not equal [162]. The tumour environment contains vast quantities of transmitters such as M-CSF, IL-6, IL-10, TGF- β and COX-2 that induce tumour megakaryocytes to differentiate into M2 macrophages, which, in addition to having inferior antigen-presenting and cytotoxic abilities, also secrete factors that inhibit immune cells, resulting in enhanced immune inhibitory effects in the tumour environment [49, 98, 102–104, 109–114]. M2 macrophages in tumour bearing mice enhance tumour growth and immune inhibitory effects. They also secrete cytokines, such as IL-10 and TGF- β , in high quantities, which attract non-cytotoxic T_{reg} cells and TH2 cells to congregate in tumour tissue; those cells inhibit the differentiation and normal function of T cells, including their cytotoxic ability, and further promote T-cell apoptosis [49, 98, 115, 163, 164]. The polarisation of TH1 and TH2 is built on cytokine patterns; polarisation begins when the antigen-presenting cells interact with naive T cells; they polarise into Type 1 (TH1) and TH2 cells in response to the type of antigen encountered [165]. TH1 and TH2 cells secrete different cytokines; TH1 cells rely on IL-2, IFN- γ and TNF, which are involved in cell-mediated immunity against pathogens, but TH2 cells depend mostly on IL-4 and IL-5, which stimulate the production of IgE antibodies and eosinophil responses, resulting in allergic diseases [166, 167]. Although an imbalanced TH1/TH2 immune response is linked to certain hypersensitivity disorders such as allergies, asthma and hay fever [168], studies have suggested that using a biological response modifier to restore the balance between TH1 and TH2 immune response can be a potential treatment option for IgE-dependent hypersensitivity [169]. *Ganoderma lucidum* is a medicinal mushroom that has been widely used as a folk medicine in Asian countries such as China and Japan for hundreds of years for its immunomodulating and anti-tumour effects. Numerous biologically available substances with immunity enhancement effects, particularly polysaccharides, have been isolated from the extract of *G. lucidum* [170].

Anti-microbial peptides are effective components of innate immunity that exist widely in biological systems. One of the specific anti-microbial peptides, hepcidin, is a 25-amino acid antibiotic peptide synthesised in the liver. Hepcidin is responsible for regulating iron balance and recycling iron in humans and mice. Studies have reported 0–100 $\mu\text{g/ml}$ concentrations of hepcidin incubated with HT1080, Hep-G2 and HeLa for 24 h. The results have indicated higher growth inhibition ratios after 70 $\mu\text{g/ml}$ treatment with hepcidin in HT1080 cells; the treatment has been very effective in inhibiting the growth of fibrosarcoma cells [171, 172]. Tachyplesin is an anti-microbial peptide present in the leukocytes of the horseshoe crab (*Tachypleus tridentatus*); it inhibited the growth of TSU tumour cells on the CAM of chicken embryos as well as the growth of B16 tumour cells in syngenic mice; moreover, it blocked the proliferation of both tumour and endothelial cells in culture in a dose-dependent manner, whereas proliferation was relatively unaffected in non-tumourigenic cell lines Cos-7 and NIH-3T3 [173]. D-K4R2L9 is a peptide comprised of Leu, Lys and Arg residues, totalling 15 amino acid residues that bind to and lyse B16-F10 mouse melanoma cells in culture at concentrations that do not harm normal 3T3 fibroblasts or erythrocytes; this can be conducted to prevent intravenous-injected D122 lung carcinoma cells from forming lung tumours in mice [174, 175]. Bovine lactoferricin (LfcinB), an anti-microbial peptide, is a 25-amino acid long highly basic peptide with a disulfide bridge between two cysteines, thus giving it a cyclic twisted anti-parallel β -sheet solution

structure. LfcinB has been tested on neuroblastoma growth *in vivo*; nude rats carrying SH-SY-5Y xenografts were given injections of 1.0 or 2.0 mg LfcinB; these rats' cancer was significantly inhibited after LfcinB treatment, compared with untreated controls [176]. Anti-microbial peptides can activate specific innate immune responses and immunomodulatory effects in the host, even if the host is at risk or has been damaged. Furthermore, researchers have proposed that anti-microbial peptides can modulate the host's immune system through inflammatory responses and can stimulate beneficial inflammation; anti-microbial peptides might be able to inhibit tumour growth.

9. Conclusion

From the injury to the wound recovery, there are a series of physiological responses that occur in relation to immune cells. Polarisation of the macrophage is an important response in wound healing. A series of inflammatory factors are cited having notable function in the differentiation from novel macrophage into the classical macrophage (M1). The cellular mechanism involved in the regulation of classical macrophage (M1) and alternative macrophage (M2) was documented in the wound healing process. At the present time, the M1/M2 differentiation was studied for selected immune responses. However, future studies may allow possible therapeutic targets considering this process in wound healing.

The immunotherapy that is being developed offers some advantageous immunomodulation factors that are known in the field of alternative medicine, such as mushroom beta-glucan, anti-microbial peptides and triterpenoid; these factors represent a novel therapeutic approach for anti-inflammation. These factors may be a viable alternative approach to the problem of drug resistance. Recent insights into wound healing and anti-inflammation are promising; however, exploiting these insights is complex because it involves chemistry, biology, instrumentation science and formulation science. Discovering new methods that are more effective in targets is difficult. Immunotherapy might be an alternative therapy that can be applied in the early phases of clinical therapy. Similarly, immunomodulation might be applicable in the early phases of immune disease.

Author details

Yu-Sheng Wu¹, Fan-Hua Nan², Sherwin Chen¹ and Shiu-Nan Chen^{1*}

*Address all correspondence to: d97b45004@ntu.edu.tw; snchen@ntu.edu.tw

1 College of Life Science, National Taiwan University, Taipei, Taiwan

2 Department of Aquaculture, National Taiwan Ocean University, Keelung, Taiwan

References

- [1] P. Allavena, A. Sica, G. Solinas *et al.*, "The inflammatory micro-environment in tumor progression: the role of tumor-associated macrophages," *Critical Reviews in Oncology/Hematology*, vol. 66, no. 1, pp. 1–9, 2008.
- [2] S. S. Choe, K. C. Shin, S. Ka *et al.*, "Macrophage HIF-2alpha ameliorates adipose tissue inflammation and insulin resistance in obesity," *Diabetes*, vol. 63, no. 10, pp. 3359–71, 2014.
- [3] Y. Enoki, T. Sato, S. Tanaka *et al.*, "Netrin-4 derived from murine vascular endothelial cells inhibits osteoclast differentiation in vitro and prevents bone loss in vivo," *FEBS Letters*, vol. 588, no. 14, pp. 2262–9, 2014.
- [4] B. Gore, M. Izikki, O. Mercier *et al.*, "Key role of the endothelial TGF-beta/ALK1/ endoglin signaling pathway in humans and rodents pulmonary hypertension," *PLoS One*, vol. 9, no. 6, p. e100310, 2014.
- [5] G. Poschmann, M. Grzendowski, A. Stefanski *et al.*, "Redox proteomics reveal stress responsive proteins linking peroxiredoxin-1 status in glioma to chemosensitivity and oxidative stress," *Biochimica et Biophysica Acta*, 2014.
- [6] L. Wu, H. Chen, C. Curtis *et al.*, "Go in for the kill," *Virulence*, vol. 5, no. 7, pp. 710–21, 2014.
- [7] J. H. Yu, and H. Kim, "Oxidative stress and inflammatory signaling in cerulein pancreatitis," *World Journal of Gastroenterology*, vol. 20, no. 46, pp. 17324–9, 2014.
- [8] E. Y. Choi, H. J. Kim, and J. S. Han, "Anti-inflammatory effects of calcium citrate in RAW 264.7 cells via suppression of NF-kappaB activation," *Environmental Toxicology and Pharmacology*, vol. 39, no. 1, pp. 27–34, 2014.
- [9] C. S. Luo, J. R. Liang, Q. Lin *et al.*, "Cellular responses associated with ROS production and cell fate decision in early stress response to iron limitation in the diatom *Thalassiosira pseudonana*," *Journal of Proteome Research*, vol. 13, no. 12, pp. 5510–23, 2014.
- [10] E. McNeill, M. J. Crabtree, N. Sahgal *et al.*, "Regulation of iNOS function and cellular redox state by macrophage Gch1 reveals specific requirements for tetrahydrobiopterin in NRF2 activation," *Free Radical Biology & Medicine*, vol. 79C, pp. 206–16, 2014.
- [11] K. A. Redgrove, and E. A. McLaughlin, "The role of the immune response in *Chlamydia trachomatis* infection of the male genital tract: a double-edged sword," *Frontiers in Immunology*, vol. 5, p. 534, 2014.
- [12] F. Morescalchi, S. Duse, E. Gambicorti *et al.*, "Proliferative vitreoretinopathy after eye injuries: an overexpression of growth factors and cytokines leading to a retinal keloid," *Mediators Inflammation*, vol. 2013, p. 269787, 2013.

- [13] M. J. Kipanyula, P. F. Seke Etet, L. Vecchio *et al.*, "Signaling pathways bridging microbial-triggered inflammation and cancer," *Cellular Signalling*, vol. 25, no. 2, pp. 403–16, 2013.
- [14] R. Kisielewski, A. Tolwinska, A. Mazurek *et al.*, "Inflammation and ovarian cancer—current views," *Ginekologia Polska*, vol. 84, no. 4, pp. 293–7, 2013.
- [15] E. Przybyt, M. J. van Luyn, and M. C. Harmsen, "Extracellular matrix components of adipose derived stromal cells promote alignment, organization, and maturation of cardiomyocytes in vitro," *Journal of Biomedical Materials Research Part A*, , 2014.
- [16] R. Roshani, F. McCarthy, and T. Hagemann, "Inflammatory cytokines in human pancreatic cancer," *Cancer Letters*, vol. 345, no. 2, pp. 157–63, 2014.
- [17] R. Nemenoff, "Activation of PPARgamma in myeloid cells promotes lung cancer progression and metastasis," *Oncoimmunology*, vol. 1, no. 3, pp. 403–4, 2012.
- [18] R. Nemenoff, "Wound healing: a role for HDACs in inhibition of fibroblast proliferation through repression of PDGF receptor-alpha. Focus on "Repression of PDGF-R-alpha after cellular injury involves TNF-alpha, formation of a c-Fos-YY1 complex, and negative regulation by HDAC", " *American Journal of Physiology – Cell Physiology*, vol. 302, no. 11, p. C1588–9, 2012.
- [19] C. Hellberg, A. Ostman, and C. H. Heldin, "PDGF and vessel maturation," *Recent Results in Cancer Research*, vol. 180, pp. 103–14, 2010.
- [20] C. Liu, W. Zhao, W. Meng *et al.*, "Platelet-derived growth factor blockade on cardiac remodeling following infarction," *Molecular and Cellular Biochemistry*, vol. 397, no. 1–2, pp. 295–304, 2014.
- [21] S. Shimizu, H. Kouzaki, T. Ogawa *et al.*, "Eosinophil-epithelial cell interactions stimulate the production of MUC5AC mucin and profibrotic cytokines involved in airway tissue remodeling," *American Journal of Rhinology & Allergy*, vol. 28, no. 2, pp. 103–9, – 2014.
- [22] K. L. Spiller, R. R. Anfang, K. J. Spiller *et al.*, "The role of macrophage phenotype in vascularization of tissue engineering scaffolds," *Biomaterials*, vol. 35, no. 15, pp. 4477–88, 2014.
- [23] K. Apel, and H. Hirt, "Reactive oxygen species: metabolism, oxidative stress, and signal transduction," *Annual Review of Plant Biology*, vol. 55, pp. 373–99, 2004.
- [24] L. O. Klotz, "Oxidant-induced signaling: effects of peroxynitrite and singlet oxygen," *Biological Chemistry*, vol. 383, no. 3–4, pp. 443–56, 2002.
- [25] A. Y. Andreyev, Y. E. Kushnareva, and A. A. Starkov, "Mitochondrial metabolism of reactive oxygen species," *Biochemistry (Moscow)*, vol. 70, no. 2, pp. 200–14, 2005.

- [26] X. Li, P. Fang, J. Mai *et al.*, "Targeting mitochondrial reactive oxygen species as novel therapy for inflammatory diseases and cancers," *Journal of Hematology & Oncology*, vol. 6, pp. 19, 2013.
- [27] E. C. Chan, F. Jiang, H. M. Peshavariya *et al.*, "Regulation of cell proliferation by NADPH oxidase-mediated signaling: potential roles in tissue repair, regenerative medicine and tissue engineering," *Pharmacology & Therapeutics*, vol. 122, no. 2, pp. 97–108, 2009.
- [28] F. Jiang, Y. Zhang, and G. J. Dusting, "NADPH oxidase-mediated redox signaling: roles in cellular stress response, stress tolerance, and tissue repair," *Pharmacological Reviews*, vol. 63, no. 1, pp. 218–42, 2011.
- [29] Y. S. Wu, S. L. Huang, F. H. Nan *et al.*, "Over-inhibition of NADPH oxidase reduce the wound healing in liver of finfish," *Fish and Shellfish Immunology*, vol. 40, no. 1, pp. 174–81, 2014.
- [30] J. F. Turrens, "Mitochondrial formation of reactive oxygen species," *Journal of Physiology*, vol. 552, no. Pt 2, pp. 335–44, 2003.
- [31] E. Novo, C. Busletta, L. V. Bonzo *et al.*, "Intracellular reactive oxygen species are required for directional migration of resident and bone marrow-derived hepatic pro-fibrogenic cells," *Journal of Hepatology*, vol. 54, no. 5, pp. 964–74, 2011.
- [32] A. Van der Goes, D. Wouters, S. M. Van Der Pol *et al.*, "Reactive oxygen species enhance the migration of monocytes across the blood-brain barrier in vitro," *FASEB Journal*, vol. 15, no. 10, pp. 1852–4, 2001.
- [33] A. Ghosh, and M. E. Greenberg, "Calcium signaling in neurons: molecular mechanisms and cellular consequences," *Science*, vol. 268, no. 5208, pp. 239–47, 1995.
- [34] S. Orrenius, B. Zhivotovsky, and P. Nicotera, "Regulation of cell death: the calcium-apoptosis link," *Nature Reviews Molecular Cell Biology*, vol. 4, no. 7, pp. 552–65, 2003.
- [35] R. G. Spragg, D. B. Hinshaw, P. A. Hyslop *et al.*, "Alterations in adenosine triphosphate and energy charge in cultured endothelial and P388D1 cells after oxidant injury," *Journal of Clinical Investigation*, vol. 76, no. 4, pp. 1471–6, 1985.
- [36] R. G. Spragg, "DNA strand break formation following exposure of bovine pulmonary artery and aortic endothelial cells to reactive oxygen products," *American Journal of Respiratory Cell and Molecular Biology*, vol. 4, no. 1, pp. 4–10, 1991.
- [37] A. C. Gasic, G. McGuire, S. Krater *et al.*, "Hydrogen peroxide pretreatment of perfused canine vessels induces ICAM-1 and CD18-dependent neutrophil adherence," *Circulation*, vol. 84, no. 5, pp. 2154–66, 1991.
- [38] A. Siflinger-Birnboim, H. Lum, P. D. Vecchio *et al.*, "Involvement of Ca²⁺ in the H₂O₂-induced increase in endothelial permeability," *American Journal of Physiology–Lung Cellular and Molecular Physiology*, vol. 14, no. 6, pp. L973, 1996.

- [39] T. Finkel, "Signal transduction by mitochondrial oxidants," *Journal of Biological Chemistry*, vol. 287, no. 7, pp. 4434–40, 2012.
- [40] Y.-S. Wu, and S.-N. Chen, "Apoptotic cell: linkage of inflammation and wound healing," *Frontiers in Pharmacology*, vol. 5, 2014.
- [41] M. Haldar, and K. M. Murphy, "Origin, development, and homeostasis of tissue-resident macrophages," *Immunological Reviews*, vol. 262, no. 1, pp. 25–35, 2014.
- [42] G. J. Guillemin, and B. J. Brew, "Microglia, macrophages, perivascular macrophages, and pericytes: a review of function and identification," *Journal of Leukocyte Biology*, vol. 75, no. 3, pp. 388–97, 2004.
- [43] A. Sica, P. Invernizzi, and A. Mantovani, "Macrophage plasticity and polarization in liver homeostasis and pathology," *Hepatology*, vol. 59, no. 5, pp. 2034–42, 2014.
- [44] S. Przybranowski, C. Wilke, N. Van Rooijen *et al.*, "Resident alveolar macrophages suppress while recruited macrophages promote allergic lung inflammation in murine models of asthma," *American Journal of Respiratory and Critical Care Medicine*, vol. 189, pp. A3685, 2014.
- [45] S. Epelman, K. J. Lavine, A. E. Beaudin *et al.*, "Embryonic and adult-derived resident cardiac macrophages are maintained through distinct mechanisms at steady state and during inflammation," *Immunity*, vol. 40, no. 1, pp. 91–104, 2014.
- [46] C. Schulz, E. G. Perdiguero, L. Chorro *et al.*, "A lineage of myeloid cells independent of Myb and hematopoietic stem cells," *Science*, vol. 336, no. 6077, pp. 86–90, 2012.
- [47] J.-J. Widmann, and H. Fahimi, "Proliferation of mononuclear phagocytes (Kupffer cells) and endothelial cells in regenerating rat liver. A light and electron microscopic cytochemical study," *The American Journal of Pathology*, vol. 80, no. 3, pp. 349, 1975.
- [48] W.-J. Wang, Y.-S. Wu, S. Chen *et al.*, "Mushroom β -glucan may immunomodulate the tumor-associated macrophages in the Lewis lung carcinoma," *BioMed Research International*, 2014.
- [49] F. O. Martinez, L. Helming, and S. Gordon, "Alternative activation of macrophages: an immunologic functional perspective," *Annual Review of Immunology*, vol. 27, pp. 451–83, 2009.
- [50] R. A. Flavell, S. Sanjabi, S. H. Wrzesinski *et al.*, "The polarization of immune cells in the tumour environment by TGF β ," *Nature Reviews Immunology*, vol. 10, no. 8, pp. 554–67, 2010.
- [51] A. Sica, and A. Mantovani, "Macrophage plasticity and polarization: in vivo veritas," *The Journal of Clinical Investigation*, vol. 122, no. 3, pp. 787–95, 2012.
- [52] A. Mantovani, S. K. Biswas, M. R. Galdiero *et al.*, "Macrophage plasticity and polarization in tissue repair and remodelling," *The Journal of Pathology*, vol. 229, no. 2, pp. 176–85, 2013.

- [53] F. O. Martinez, A. Sica, A. Mantovani *et al.*, "Macrophage activation and polarization," *Frontiers in Bioscience: A Journal and Virtual Library*, vol. 13, pp. 453–61, 2007.
- [54] A. Mantovani, S. Sozzani, M. Locati *et al.*, "Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes," *Trends in Immunology*, vol. 23, no. 11, pp. 549–55, 2002.
- [55] F. R. Smiderle, G. Alquini, M. Z. Tadra-Sfeir *et al.*, "Agaricus bisporus and Agaricus brasiliensis (1→6)-β-d-glucans show immunostimulatory activity on human THP-1 derived macrophages," *Carbohydrate Polymers*, vol. 94, no. 1, pp. 91–9, 2013.
- [56] R.-S. Sauer, D. Hackel, L. Morschel *et al.*, "Toll like receptor (TLR)-4 as a regulator of peripheral endogenous opioid-mediated analgesia in inflammation," *Molecular Pain*, vol. 10, no. 1, pp. 10, 2014.
- [57] J. S. Orr, M. J. Puglisi, K. L. Ellacott *et al.*, "Toll-like receptor 4 deficiency promotes the alternative activation of adipose tissue macrophages," *Diabetes*, vol. 61, no. 11, pp. 2718–27, 2012.
- [58] N. Wang, H. Liang, and K. Zen, "Molecular mechanisms that influence the macrophage M1–M2 polarization balance," *Frontiers in Immunology*, vol. 5, 2014.
- [59] C. S. Whyte, E. T. Bishop, D. Rückerl *et al.*, "Suppressor of cytokine signaling (SOCS) 1 is a key determinant of differential macrophage activation and function," *Journal of Leukocyte Biology*, vol. 90, no. 5, pp. 845–54, 2011.
- [60] S. Gordon, and F. O. Martinez, "Alternative activation of macrophages: mechanism and functions," *Immunity*, vol. 32, no. 5, pp. 593–604, 2010.
- [61] H. Zhou, J. Liao, J. Aloor *et al.*, "CD11b/CD18 (Mac-1) is a novel surface receptor for extracellular double-stranded RNA to mediate cellular inflammatory responses," *The Journal of Immunology*, vol. 190, no. 1, pp. 115–25, 2013.
- [62] Y. S. Schwartz, and A. Svistelnik, "Functional phenotypes of macrophages and the M1-M2 polarization concept. Part I. Proinflammatory phenotype," *Biochemistry (Moscow)*, vol. 77, no. 3, pp. 246–60, 2012.
- [63] U. S. Rangaswamy, and S. H. Speck, "Murine gammaherpesvirus M2 protein induction of IRF4 via the NFAT pathway leads to IL-10 expression in B cells," *PLoS Pathogens*, vol. 10, no. 1, pp. e1003858, 2014.
- [64] I. A. Udalova, T. Krausgruber, T. Smallie *et al.*, "IRF5 promotes inflammatory macrophage polarization and Th1/Th17 response," *Nature Immunology*, 2011.
- [65] H. Xu, J. Zhu, S. Smith *et al.*, "Notch-RBP-J signaling regulates the transcription factor IRF8 to promote inflammatory macrophage polarization," *Nature Immunology*, vol. 13, no. 7, pp. 642–50, 2012.

- [66] H. J. Lee, Y. K. Oh, M. Rhee *et al.*, "The role of STAT1/IRF-1 on synergistic ROS production and loss of mitochondrial transmembrane potential during hepatic cell death induced by LPS/d-GalN," *Journal of Molecular Biology*, vol. 369, no. 4, pp. 967–84, 2007.
- [67] W. Xiao, H. Hong, Y. Kawakami *et al.*, "Regulation of myeloproliferation and M2 macrophage programming in mice by Lyn/Hck, SHIP, and Stat5," *The Journal of Clinical Investigation*, vol. 118, no. 3, pp. 924, 2008.
- [68] Y. Ji, S. Sun, A. Xu *et al.*, "Activation of natural killer T cells promotes M2 macrophage polarization in adipose tissue and improves systemic glucose tolerance via interleukin-4 (IL-4)/STAT6 protein signaling axis in obesity," *Journal of Biological Chemistry*, vol. 287, no. 17, pp. 13561–71, 2012.
- [69] J. Wan, M. Benkdane, F. Teixeira-Clerc *et al.*, "M2 Kupffer cells promote M1 Kupffer cell apoptosis: a protective mechanism against alcoholic and nonalcoholic fatty liver disease," *Hepatology*, vol. 59, no. 1, pp. 130–42, 2014.
- [70] M. Heusinkveld, P. J. d. V. van Steenwijk, R. Goedemans *et al.*, "M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to activated M1 macrophages by CD4+ Th1 cells," *The Journal of Immunology*, vol. 187, no. 3, pp. 1157–65, 2011.
- [71] D. Zhou, C. Huang, Z. Lin *et al.*, "Macrophage polarization and function with emphasis on the evolving roles of coordinated regulation of cellular signaling pathways," *Cellular Signalling*, vol. 26, no. 2, pp. 192–7, 2014.
- [72] A. J. Covarrubias, and T. Horng, "IL-6 strikes a balance in metabolic inflammation," *Cell Metabolism*, vol. 19, no. 6, pp. 898–9, 2014.
- [73] P. Kell, T. Ennis, K. Chang *et al.*, "Macrophages mediate the ability of the receptor for advanced glycation end products to prevent formation of abdominal aortic aneurysm in a murine model," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 34, no. Suppl 1, p. A114, 2014.
- [74] A. Gal, T. T. Tapmeier, and R. J. Muschel, "Plasticity of tumor associated macrophages in a metastatic melanoma model in the mouse," *Cancer Research*, vol. 72, no. 8 Suppl, pp. 402, 2012.
- [75] K. L. Spiller, S. Nassiri, C. E. Witherel *et al.*, "Sequential delivery of immunomodulatory cytokines to facilitate the M1-to-M2 transition of macrophages and enhance vascularization of bone scaffolds," *Biomaterials*, vol. 37, pp. 194–207, 2015.
- [76] X. Cai, Y. Yin, N. Li *et al.*, "Re-polarization of tumor-associated macrophages to pro-inflammatory M1 macrophages by microRNA-155," *Journal of Molecular Cell Biology*, vol. 4, no. 5, pp. 341–3, 2012.
- [77] J.-H. Lee, G. T. Lee, S. H. Woo *et al.*, "BMP-6 in renal cell carcinoma promotes tumor proliferation through IL-10–dependent M2 polarization of tumor-associated macrophages," *Cancer Research*, vol. 73, no. 12, pp. 3604–14, 2013.

- [78] N. Ouchi, J. L. Parker, J. J. Lugus *et al.*, "Adipokines in inflammation and metabolic disease," *Nature Reviews Immunology*, vol. 11, no. 2, pp. 85–97, 2011.
- [79] R. G. Baker, M. S. Hayden, and S. Ghosh, "NF- κ B, inflammation, and metabolic disease," *Cell Metabolism*, vol. 13, no. 1, pp. 11–22, 2011.
- [80] B. Gooptu, and D. A. Lomas, "Polymers and inflammation: disease mechanisms of the serpinopathies," *The Journal of Experimental Medicine*, vol. 205, no. 7, pp. 1529–34, 2008.
- [81] R. L. Davis, A. E. Shrimpton, P. D. Holohan *et al.*, "Familial dementia caused by polymerization of mutant neuroserpin," *Nature*, vol. 401, no. 6751, pp. 376–9, 1999.
- [82] G. K. Hansson, "Inflammation, atherosclerosis, and coronary artery disease," *New England Journal of Medicine*, vol. 352, no. 16, pp. 1685–95, 2005.
- [83] H. Akiyama, S. Barger, S. Barnum *et al.*, "Inflammation and Alzheimer's disease," *Neurobiology of Aging*, vol. 21, no. 3, pp. 383–421, 2000.
- [84] T. A. Wynn, A. Chawla, and J. W. Pollard, "Macrophage biology in development, homeostasis and disease," *Nature*, vol. 496, no. 7446, pp. 445–55, 2013.
- [85] A. Sica, and A. Mantovani, "Macrophage plasticity and polarization: in vivo veritas," *Journal of Clinical Investigation*, vol. 122, no. 3, pp. 787–95, 2012.
- [86] T. L. Denning, B. A. Norris, O. Medina-Contreras *et al.*, "Functional specializations of intestinal dendritic cell and macrophage subsets that control Th17 and regulatory T cell responses are dependent on the T cell/APC ratio, source of mouse strain, and regional localization," *The Journal of Immunology*, vol. 187, no. 2, pp. 733–47, 2011.
- [87] J. A. Joyce, and J. W. Pollard, "Microenvironmental regulation of metastasis," *Nature Reviews Cancer*, vol. 9, pp. 239–52, 2009.
- [88] L. M. Coussens, and Z. Werb, "Inflammation and cancer," *Nature*, vol. 420, pp. 860–67, 2002.
- [89] G. Baronzio, G. Fiorentini, and C. R. Cogle, *Cancer microenvironment and therapeutic implications*. Springer, 2009.
- [90] F. Xing, J. Saidou, and K. Watabe, "Cancer associated fibroblasts (CAFs) in tumor microenvironment," *Frontiers in Bioscience (Landmark Ed)*, vol. 15, pp. 166–79, 2010.
- [91] G. Castello, S. Scala, G. Palmieri *et al.*, "HCV-related hepatocellular carcinoma: From chronic inflammation to cancer," *Clinical Immunology*, vol. 134, pp. 237–50, 2010.
- [92] C. A. Janeway, M. Walport, and P. Travers, *Immunobiology: the immune system in health and disease*. Garland Science, 2005.
- [93] D. I. Gabrilovich, and A. A. Hurrwitz, *Tumor-induced immune suppression*. Springer, 2008.
- [94] W. Zou, "Immunosuppressive networks in the tumour environment and their therapeutic relevance," *Nature Reviews Cancer*, vol. 5, pp. 263–74, 2005.

- [95] G. P. Dunn, A. T. Bruce, H. Ikeda *et al.*, "Cancer immunoediting: from immunosurveillance to tumor escape," *Nature Immunology*, vol. 3, pp. 991–8, 2002.
- [96] G. P. Dunn, L. J. Old, and R. D. Schreiber, "The immunobiology of cancer immunosurveillance and immunoediting," *Immunity*, vol. 21, pp. 137–48, 2004.
- [97] B. Z. Qian, and J. W. Pollard, "Macrophage diversity enhances tumor progression and metastasis," *Cell*, vol. 141, pp. 39–51, 2010.
- [98] D. W. Siemann, "Tumor microenvironment. Wiley Online Library, 2011.
- [99] N. Mach, S. Gillessen, S. B. Wilson *et al.*, "Differences in dendritic cells stimulated *in vivo* by tumors engineered to secrete granulocyte-macrophage colony-stimulating factor of flt3-ligand," *Cancer Research*, vol. 60, pp. 3239–46, 2000.
- [100] S. K. Biswas, and A. Mantovani, "Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm," *Nature Immunology*, vol. 11, pp. 889–96, 2010.
- [101] B. Ruffell, N. I. Affara, and L. M. Coussens, "Differential macrophage programming in the tumor microenvironment," *Trends in Immunology*, vol. 33, pp. 119–126, 2012.
- [102] R. A. Flavell, S. Sanjabi, S. H. Wrzesinski *et al.*, "The polarization of immune cells in the tumour environment by TGFbeta," *Nature Reviews Immunology*, vol. 10, pp. 554–67, 2010.
- [103] L. Bingle, N. J. Brown, and C. E. Lewis, "The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies," *The Journal of Pathology*, vol. 196, pp. 254–65, 2002.
- [104] J. W. Pollard, "Tumour-educated macrophages promote tumour progression and metastasis," *Nature Reviews Cancer*, vol. 4, pp. 71–78, 2004.
- [105] A. Mantovani, A. Sica, S. Sozzani *et al.*, "The chemokine system in diverse forms of macrophage activation and polarization," *Trends in Immunology*, vol. 25, pp. 677–86, 2004.
- [106] A. Mantovani, and A. Sica, "Macrophages, innate immunity and cancer: balance, tolerance, and diversity," *Current Opinion in Immunology*, vol. 22, pp. 231–7, 2010.
- [107] A. Mantovani, A. Sica, P. Allavena *et al.*, "Tumor-associated macrophages and the related myeloid-derived suppressor cells as a paradigm of the diversity of macrophage activation," *Human Immunology*, vol. 70, pp. 325–30, 2009.
- [108] C. Steidl, T. Lee, S. P. Shah *et al.*, "Tumor-associated macrophages and survival in classic Hodgkin's Lymphoma," *The New England Journal of Medicine*, vol. 362, pp. 875–85, 2010.
- [109] K. D. Elgert, D. G. Alleva, and D. W. Mullins, "Tumor-induced immune dysfunction: the macrophage connection," *Journal of Leukocyte Biology*, vol. 64, pp. 275–90, 1998.
- [110] C. Sunderkötter, M. Goebeler, K. Schulze-Osthoff *et al.*, "Macrophage-derived angiogenesis factors," *Pharmacology & Therapeutics*, vol. 51, pp. 195–216, 1991.

- [111] E. Giraudo, M. Inoue, and D. Hanahan, "An amino-bisphosphonate targets MMP-9-expressing macrophages and angiogenesis to impair cervical carcinogenesis," *The Journal of Clinical Investigation*, vol. 114, pp. 623–33, 2004.
- [112] A. Ben-Baruch, "Inflammation-associated immune suppression in cancer: the roles played by cytokines, chemokines and additional mediators," *Seminars in Cancer Biology*, vol. 16, pp. 38–52, 2006.
- [113] M. Mitsuhashi, J. Liu, S. Cao *et al.*, "Regulation of interleukin-12 gene expression and its anti-tumor activities by prostaglandin E2 derived from mammary carcinomas," *Journal of Leukocyte Biology*, vol. 76, pp. 322–32, 2004.
- [114] A. Mantovani, S. Sozzani, M. Locati *et al.*, "Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes," *Trends in Immunology* vol. 23, pp. 549–55, 2002.
- [115] G. Solinas, G. Germano, A. Mantovani *et al.*, "Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation," *Journal of Leukocyte Biology*, vol. 86, pp. 1065–73, 2009.
- [116] M. P. Fink, "Role of reactive oxygen and nitrogen species in acute respiratory distress syndrome," *Current Opinion in Critical Care*, vol. 8, no. 1, pp. 6–11, 2002.
- [117] F. Dong, X. C. Zhang, S. Y. Li *et al.*, "Possible involvement of NADPH oxidase and JNK in homocysteine-induced oxidative stress and apoptosis in human umbilical vein endothelial cells," *Cardiovascular Toxicology*, vol. 5, no. 1, pp. 9–20, 2005.
- [118] E. Novo, C. Busletta, L. V. Bonzo *et al.*, "Intracellular reactive oxygen species are required for directional migration of resident and bone marrow-derived hepatic pro-fibrogenic cells," *Journal of Hepatology*, vol. 54, no. 5, pp. 964–74, 2011.
- [119] A. Van der Goes, D. Wouters, S. M. Van Der Pol *et al.*, "Reactive oxygen species enhance the migration of monocytes across the blood-brain barrier in vitro," *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology*, vol. 15, no. 10, pp. 1852–4, 2001.
- [120] J. W. Park, S. W. Ryter, and A. M. Choi, "Functional significance of apoptosis in chronic obstructive pulmonary disease," *COPD*, vol. 4, no. 4, pp. 347–53, 2007.
- [121] S. W. Ryter, H. P. Kim, A. Hoetzel *et al.*, "Mechanisms of cell death in oxidative stress," *Antioxid Redox Signal*, vol. 9, no. 1, pp. 49–89, 2007.
- [122] M. L. Circu, and T. Y. Aw, "Reactive oxygen species, cellular redox systems, and apoptosis," *Free Radical Biology and Medicine*, vol. 48, no. 6, pp. 749–62, 2010.
- [123] B. T. Mossman, "Introduction to serial reviews on the role of reactive oxygen and nitrogen species (ROS/RNS) in lung injury and diseases," *Free Radical Biology and Medicine*, vol. 34, no. 9, pp. 1115–6, 2003.

- [124] S. I. Grivennikov, F. R. Greten, and M. Karin, "Immunity, Inflammation, and Cancer," *Cell*, vol. 140, no. 6, pp. 883–899, 2010.
- [125] T. A. Wilgus, "Immune cells in the healing skin wound: influential players at each stage of repair," *Pharmacological Research*, vol. 58, no. 2, pp. 112–6, 2008.
- [126] R. Alinovi, M. Goldoni, S. Pinelli *et al.*, "Oxidative and pro-inflammatory effects of cobalt and titanium oxide nanoparticles on aortic and venous endothelial cells," *Toxicology In Vitro*, vol. 29, no. 3, pp. 426–37, 2014.
- [127] G. Pellegrini, G. Rasperini, G. Obot *et al.*, "Soft tissue healing in alveolar socket preservation technique: histologic evaluations," *International Journal of Periodontics Restorative Dent*, vol. 34, no. 4, pp. 531–9, 2014.
- [128] V. Kumar, "Innate lymphoid cells: new paradigm in immunology of inflammation," *Immunology Letters*, vol. 157, no. 1–2, pp. 23–37, 2014.
- [129] A. Barbul, and M. C. Regan, "The regulatory role of T lymphocytes in wound healing," *Journal of Trauma*, vol. 30, no. 12 Suppl, pp. S97–100, 1990.
- [130] T. A. Petrie, N. S. Strand, C. Tsung-Yang *et al.*, "Macrophages modulate adult zebrafish tail fin regeneration," *Development*, vol. 141, no. 13, pp. 2581–91, 2014.
- [131] M. Sugaya, "Chemokines and skin diseases," *Archivum Immunologiae et Therapiae Experimentalis (Warsz)*, 2014.
- [132] S. Zhang, S. Dehn, M. DeBerge *et al.*, "Phagocyte-myocyte interactions and consequences during hypoxic wound healing," *Cellular Immunology*, vol. 291, no. 1–2, pp. 65–73, 2014.
- [133] L. A. DiPietro, "Wound healing: the role of the macrophage and other immune cells," *Shock*, vol. 4, no. 4, pp. 233–40, 1995.
- [134] C. Fathke, L. Wilson, K. Shah *et al.*, "Wnt signaling induces epithelial differentiation during cutaneous wound healing," *BMC Cell Biology*, vol. 7, pp. 4, 2006.
- [135] M.-K. Song, Y.-K. Park, and J.-C. Ryu, "Polycyclic aromatic hydrocarbon (PAH)-mediated upregulation of hepatic microRNA-181 family promotes cancer cell migration by targeting MAPK phosphatase-5, regulating the activation of p38 MAPK," *Toxicology and Applied Pharmacology*, vol. 273, no. 1, pp. 130–9, 2013.
- [136] Y. Wang, Y. Zhou, and D. T. Graves, "FOXO transcription factors: their clinical significance and regulation," *BioMed Research International*, vol. 2014, 2014.
- [137] C. W. Chow, M. T. Herrera Abreu, T. Suzuki *et al.*, "Oxidative stress and acute lung injury," *American Journal of Respiratory Cell and Molecular Biology*, vol. 29, no. 4, pp. 427–31, 2003.

- [138] C. Yang, H. Moriuchi, J. Takase *et al.*, "Oxidative stress in early stage of acute lung injury induced with oleic acid in guinea pigs," *Biological and Pharmaceutical Bulletin*, vol. 26, no. 4, pp. 424–8, 2003.
- [139] V. Patel, I. V. Chivukula, S. Roy *et al.*, "Oxygen: from the benefits of inducing VEGF expression to managing the risk of hyperbaric stress," *Antioxidants & Redox Signaling*, vol. 7, no. 9–10, pp. 1377–87, 2005.
- [140] A. C. Bulua, A. Simon, R. Maddipati *et al.*, "Mitochondrial reactive oxygen species promote production of proinflammatory cytokines and are elevated in TNFR1-associated periodic syndrome (TRAPS)," *The Journal of Experimental Medicine*, vol. 208, no. 3, pp. 519–33, 2011.
- [141] R. J. Aitken, K. T. Jones, and S. A. Robertson, "Reactive oxygen species and sperm function—in sickness and in health," *Journal of Andrology*, vol. 33, no. 6, pp. 1096–1106, 2012.
- [142] S. Eligini, I. Arenaz, S. S. Barbieri *et al.*, "Cyclooxygenase-2 mediates hydrogen peroxide-induced wound repair in human endothelial cells," *Free Radical Biology and Medicine*, vol. 46, no. 10, pp. 1428–36, 2009.
- [143] M. Iizuka, and S. Konno, "Wound healing of intestinal epithelial cells," *World Journal of Gastroenterology: WJG*, vol. 17, no. 17, pp. 2161, 2011.
- [144] D. Maslinska, and M. Gajewski, "Some aspects of the inflammatory process," *Folia Neuropathologica*, vol. 36, no. 4, pp. 199–204, 1998.
- [145] P. Libby, P. M. Ridker, and A. Maseri, "Inflammation and atherosclerosis," *Circulation*, vol. 105, no. 9, pp. 1135–43, 2002.
- [146] J. M. Rubio-Perez, and J. M. Morillas-Ruiz, "A review: inflammatory process in Alzheimer's disease, role of cytokines," *Scientific World Journal*, 2012.
- [147] C. D. Gregory, "Inflammation and cancer revisited: an hypothesis on the oncogenic potential of the apoptotic tumor cell," *Autoimmunity*, vol. 46, no. 5, pp. 312–6, 2013.
- [148] B. B. Aggarwal, S. Shishodia, S. K. Sandur *et al.*, "Inflammation and cancer: how hot is the link?," *Biochemical Pharmacology*, vol. 72, no. 11, pp. 1605–21, 2006.
- [149] A. K. Muller, M. Meyer, and S. Werner, "The roles of receptor tyrosine kinases and their ligands in the wound repair process," *Seminars in Cell & Developmental Biology*, vol. 23, no. 9, pp. 963–70, 2012.
- [150] S. Danckwardt, M. W. Hentze, and A. E. Kulozik, "Pathologies at the nexus of blood coagulation and inflammation: thrombin in hemostasis, cancer, and beyond," *Journal of Molecular Medicine: JMM*, vol. 91, no. 11, pp. 1257–71, 2013.
- [151] S. Strukova, "Blood coagulation-dependent inflammation. Coagulation-dependent inflammation and inflammation-dependent thrombosis," *Frontiers in Bioscience*, vol. 11, pp. 59–80, 2006.

- [152] T. N. Dugina, E. V. Kiseleva, I. V. Chistov *et al.*, "Receptors of the PAR-family as a link between blood coagulation and inflammation," *Biochemistry (Moscow)*, vol. 67, no. 1, pp. 65–74, 2002.
- [153] Y. Nakanishi, M. Nakatsuji, H. Seno *et al.*, "COX-2 inhibition alters the phenotype of tumor-associated macrophages from M2 to M1 in ApcMin/+ mouse polyps," *Carcinogenesis*, vol. 32, pp. 1333–9, 2011.
- [154] T. Ikekawa, "Enokitake, *Flammulina velutipes*: host-mediated antitumor polysaccharides," *Food Reviews International*, vol. 11, pp. 203–6, 1995.
- [155] T. Ikekawa, N. Uehara, Y. Maeda *et al.*, "Antitumor activity of aqueous extracts of edible mushrooms," *Cancer Research*, vol. 29, pp. 734–5, 1969.
- [156] S. P. Wasser, "Medicinal mushrooms as a source of antitumor and immunomodulating polysaccharides," *Applied Microbiology and Biotechnology*, vol. 60, pp. 258–74, 2002.
- [157] T. Inomata, G. B. Goodman, C. J. Fryer *et al.*, "Immune reaction induced by X-rays and pions and its stimulation by schizophyllan (SPG)," *The British Journal of Cancer*, vol. 27, pp. 122–5, 1996.
- [158] H. Nanba, and K. Kubo, "Effect of Maitake D-fraction on cancer prevention," *Cancer*, vol. 833, pp. 204–7, 1997.
- [159] C. Menetrier-Caux, G. Montmain, M. C. Dieu *et al.*, "Inhibition of the differentiation of dendritic cells from CD34⁺ progenitors by tumor cells: role of interleukin-6 and macrophage colony-stimulating factor," *Blood*, vol. 92, pp. 4778–91, 1998.
- [160] E. Y. Lin, V. Gouon-Evans, A. V. Nquyen *et al.*, "The macrophage growth factor CSF-1 in mammary gland development and tumor progression," *Journal of Mammary Gland Biology and Neoplasia*, vol. 7, pp. 147–62, 2002.
- [161] C. E. Lewis, and J. W. Pollard, "Distinct role of macrophages in different tumor microenvironment," *Cancer Research*, vol. 66, pp. 605–12, 2006.
- [162] E. S. Ch'ng, H. Jaafar, and S. E. Tuan Sharif, "Breast tumor angiogenesis and tumor-associated macrophages: histopathologist's perspective," *Pathology Research International*, vol. 2011, pp. 1–13, 2011.
- [163] T. J. Standiford, R. Kuick, U. Bhan *et al.*, "TGF- β -induced IRAK-M expression in tumor-associated macrophages regulates lung tumor growth," *Oncogene*, vol. 30, pp. 2475–84, 2011.
- [164] K. Murphy, P. Travers, and M. Walport, *Immunobiology*, 7th ed. Garland Science, 2008.
- [165] T. R. Mosmann, H. Cherwinski, M. W. Bond *et al.*, "Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. 1986," *Journal of Immunology*, vol. 175, no. 1, pp. 5–14, 2005.

- [166] T. R. Mosmann, and R. L. Coffman, "TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties," *Annual Review of Immunology*, vol. 7, pp. 145–73, 1989.
- [167] E. Maggi, "The TH1/TH2 paradigm in allergy," *Immunotechnology*, vol. 3, no. 4, pp. 233–44, 1998.
- [168] P. Kidd, "Th1/Th2 balance: the hypothesis, its limitations, and implications for health and disease," *Alternative Medicine Review*, vol. 8, no. 3, pp. 223–46, 2003.
- [169] M. E. Stern, K. F. Siemasko, and J. Y. Niederkorn, "The Th1/Th2 paradigm in ocular allergy," *Current Opinion in Allergy and Clinical Immunology*, vol. 5, no. 5, pp. 446–50, 2005.
- [170] Z. B. Lin, and H. N. Zhang, "Anti-tumor and immunoregulatory activities of *Ganoderma lucidum* and its possible mechanisms," *Acta Pharmacologica Sinica*, vol. 25, no. 11, pp. 1387–95, 2004.
- [171] J. S. Shi, and A. C. Camus, "Hepcidins in amphibians and fishes: antimicrobial peptides or iron-regulatory hormones?" *Developmental and Comparative Immunology*, vol. 30, no. 9, pp. 746–55, 2006.
- [172] J. Y. Chen, W. J. Lin, and T. L. Lin, "A fish antimicrobial peptide, tilapia hepcidin TH2-3, shows potent antitumor activity against human fibrosarcoma cells," *Peptides*, vol. 30, no. 9, pp. 1636–42, 2009.
- [173] Y. X. Chen, X. M. Xu, S. G. Hong *et al.*, "RGD-tachyplesin inhibits tumor growth," *Cancer Research*, vol. 61, no. 6, pp. 2434–38, 2001.
- [174] D. W. Hoskin, and A. Ramamoorthy, "Studies on anticancer activities of antimicrobial peptides," *Biochimica et Biophysica Acta–Biomembranes*, vol. 1778, no. 2, pp. 357–75, 2008.
- [175] N. Papo, M. Shahar, L. Eisenbach *et al.*, "A novel lytic peptide composed of DL-amino acids selectively kills cancer cells in culture and in mice," *Journal of Biological Chemistry*, vol. 278, no. 23, pp. 21018–23, 2003.
- [176] L. T. Eliassen, G. Berge, A. Leknessund *et al.*, "The antimicrobial peptide, Lactoferricin B, is cytotoxic to neuroblastoma cells in vitro and inhibits xenograft growth in vivo," *International Journal of Cancer*, vol. 119, no. 3, pp. 493–500, 2006.