

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Glycosylation of Integrins in Melanoma Progression

Ewa Pocheć and Anna Lityńska

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/64287>

Abstract

Each stage of melanoma development from transformed melanocytes to metastatic lesions requires the involvement of cell adhesion receptors, among which integrins are of particular importance. Strong N-glycosylation of $\alpha\beta$ integrin heterodimers influences their processing, activation, and functions related to the modulation of cell adhesion to extracellular matrix proteins (ECM) and the basement membrane. A lack of N-glycans on integrin chains significantly reduces their interactions with the ECM. Melanoma progression is accompanied by changes in the composition of N-glycans on integrin subunits. The glycosylation profile of integrins depends on the stage of melanoma development and on the location of the metastasis. Enhanced expression of β 1,6-branched complex-type oligosaccharides and altered sialylation are well-characterized changes in the N-glycosylation of integrins observed in melanoma progression. This chapter summarizes the current state of knowledge about α 3 β 1, α 5 β 1, and α v β 3 integrin glycosylation in melanoma and the functional consequences of changed glycosylation for the development of this cancer.

Keywords: integrin, N-glycosylation, melanoma, β 1,6 branching, migration, extracellular matrix proteins

1. Introduction

Melanoma progression and the acquisition of invasive and metastatic competence by melanoma cells are accompanied not only by changes in integrin expression but also by alterations of the sugar component of these heavily N-glycosylated adhesive proteins [1]. This post-translational modification is critical to integrin functions, mainly its interactions with extracellular matrix proteins (ECM) and the basement membrane [2]. Changes in the expression and glycosylation of integrins contribute to each stage of melanoma progression.

Human cutaneous melanoma develops in a series of definable stages, from the common acquired nevus and dysplastic nevus through the radial growth phase (RGP) and vertical growth phase (VGP) of primary melanoma and finally metastatic melanoma. During these multistep transformations, melanoma cells acquire the ability to invade the dermis and then disseminate throughout the body via blood and lymphatic vessels [3–8]. Adjustment of integrin glycosylation is an important feature of the melanoma cell’s adaptation to the constantly changing conditions of its microenvironment. This chapter reviews the current state of knowledge about integrin glycosylation in the course of melanoma progression.

2. Overall characteristics of integrins

The term “integrins” introduced by Hynes reflects the capacity of these cell surface receptors to integrate ECM proteins with the cytoskeleton and with intracellular signaling pathways by physical connection [9]. The role of integrin-mediated adhesion to the ECM in cell survival is now accepted. Integrins are heterodimeric cell surface adhesion molecules consisting of α and β subunits. By combining 18 α with 8 β subunits, at least 24 integrin dimers can be formed, each with its own characteristic specificity for ligands [10] (Figure 1).

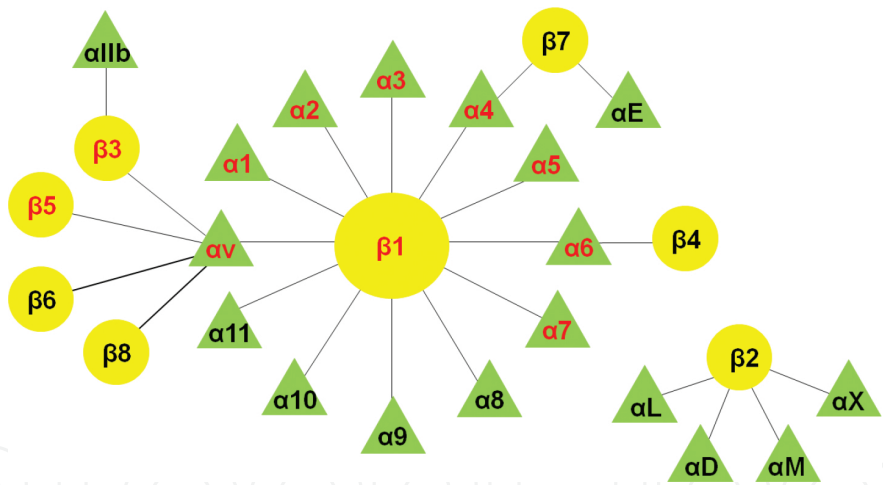


Figure 1. Integrin classification based on β subunits possessed in common. Integrin heterodimers whose expression was observed to increase during melanoma progression are marked in red.

2.1. The structure of α and β subunit ectodomains

Each integrin subunit consists of a large extracellular domain and short transmembrane and cytoplasmic domains. The extracellular domains (ectodomains) of the α and β subunits are constructed of several subdomains joined together by flexible linkers [11, 12]. The crystal structure of the $\alpha v \beta 3$ [13] and $\alpha IIB \beta 3$ [14] ectodomains has been characterized in detail.

The ectodomain of the α -subunit contains four or five elements: a seven-bladed β -propeller, a thigh, and two calfs. There are also nine integrins with an α -subunit containing an additional

α -I domain inserted between blades 2 and 3 of the β -propeller. A structure similar to an α -I domain is also present in the β subunit of integrins. The β -propeller contains Ca^{2+} -binding sites needed for ligand binding. The thigh and calf of the α -subunit have 140–170 residues folded into an immunoglobulin-like domain.

The ectodomain of the β -subunit consists of seven subdomains: a PSI (plexin-semaphorin-integrin), an Ig-like hybrid, a β -I-like domain, and four EGF-like modules (epidermal growth factor-like modules), followed by the β -tail part. The β -I-like domain is inserted into the hybrid modules and shows homology to the α -I domain. The PSI domain is split into two parts. The α -I domain is the primary region of ligand binding in integrins that have this structure, whereas the other integrins form the binding site through the cooperation of both subunit ectodomains (β -propeller/ β -I-like interface) [15]. It has been suggested that the I-domain can exist as an “open” (high-affinity) or “closed” (low-affinity) conformation. The presence of a “metal-ion-dependent-adhesion-site” (MIDAS) motif indicates the role of divalent metal ions in achievement of the high-affinity state by integrins.

The transmembrane segments of each subunit are followed by a short cytoplasmic tail. Although they have no enzymatic activity, cytoplasmic tails play an important role in integrin activity and signal transfer.

2.2. Bidirectional signaling of integrins

Integrins are involved in bidirectional signaling—inside-out and outside-in—through their function as a linker between the ECM and the cytoskeleton [16, 17]. Control of the integrin conformation state is required for their signaling. There is little agreement among the findings from nuclear magnetic resonance (NMR) studies of cytoplasmic tails [12], but other data support the view that transmembrane and cytoplasmic domains play a key role in this signaling. In the inactive state, these domains are closely associated; separation of the chain results in activation of adhesion [11, 16].

Inside-out activation is mediated by talin binding to the β -tail, which interrupts the α/β interaction [18]. In fact, a large number of proteins have been shown to interact with cytoplasmic domains of integrins, among them cytoskeleton proteins (talin, filamin, and kindlins), adaptor proteins, and kinases [11, 19]. Talin and kindlins bound to β -integrin cooperate to regulate integrin affinity [19]. Upon binding of the ligand to the integrins' extracellular domain, signal transduction to the cytoplasm is transmitted in the classical direction: outside-in. Generation of intracellular signals leads to the formation of a focal adhesion complex which involves over 150 intracellular proteins and serves as a center of intracellular signaling [20]. Among these proteins are scaffolding molecules and also kinases such as focal adhesion kinase (FAK) and Src family kinase (SFK). So the function of integrin is related to its ligand affinity, which can be induced either by conformational changes or by clustering on the cell surface [11].

2.3. Classification of integrins and their ligands

The first classification of integrins was based on the presence of a common β subunit having distinct α subfamilies. Recent work has shown that one α subunit may associate with different

β subunits, in particular a αv subunit. However, the largest number of integrins are still assigned to the $\beta 1$ (VLA, very late-activated antigens) subfamily. In this group, are integrins recognizing fibronectin (FN) ($\alpha 5\beta 1$, $\alpha 4\beta 1$), collagen ($\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 10\beta 1$, $\alpha 11\beta 1$) or laminin (LN) ($\alpha 2\beta 1$, $\alpha 3\beta 1$, $\alpha 6\beta 1$, $\alpha 7\beta 1$) [21, 22] (**Figure 1**). The $\alpha 4\beta 1$ integrin present on human lymphocytes has been shown to bind vascular cell adhesion molecule 1 (VCAM-1), the cell surface protein of activated endothelia. The $\beta 2$ subfamily of integrins is limited to white blood cells. Recognition of cell surface receptors of the Ig superfamily by $\beta 2$ integrins is crucial to leukocyte–endothelium interaction [22, 23]. The $\beta 3$ subfamily consists of two members: platelet receptor ($\alpha IIb\beta 3$) and vitronectin - receptor ($\alpha v\beta 3$). Integrin $\alpha IIb\beta 3$ is specific for platelets; it recognizes fibrinogen specifically but upon platelet activation can also bind fibronectin (FN), von Willebrand's factor and thrombospondin. Integrin $\alpha v\beta 3$ binds multiple ligands including vitronectin (VN), fibrinogen, thrombospondin, and von Willebrand's factor [23]. αv subunit can associate with more than one β subunit, such as $\beta 1$, $\beta 5$, $\beta 6$, and $\beta 8$ [22].

Integrins bind to a specific motif in their ligands. The RGD (Arg-Gly-Asp) sequence found within matrix proteins including FN, VN, thrombospondin, and laminin (LN) is usually recognized by integrins [12, 23], but there are integrins that recognize their ligands through motifs other than RGD. Integrins, $\alpha 3\beta 1$, $\alpha 6\beta 1$, and $\alpha 7\beta 1$, being highly specific LN receptors, bind to different regions of this ligand [12]. Fibrinogen contains the binding sequence Lys-Gln-Ala-Gly-Asp-Val, while Asp-Gly-Glu-Ala was found to be the dominant binding motif in type I collagen [24].

3. Integrin expression in melanoma

Changes in integrin expression have been studied extensively in melanoma carcinogenesis [8, 25]. The integrin profile of melanoma cells differs significantly from that of normal melanocytes [26, 27] and is closely related to the stage of melanoma progression [24, 28]. Flow cytometry showed significant differences in the expression of $\alpha 2$, $\alpha 3$, $\beta 1$, and especially $\alpha 5$ integrin subunits between WM35 primary and two metastatic human cell lines (WM9 and A375), indicating that acceleration of melanoma invasion is accompanied by increased integrin subunit synthesis [29]. Significant up-regulation of $\alpha 5$ integrin expression was also shown in highly metastatic B16-F10 murine melanoma cells as compared to weakly metastatic B16-F1 cells [30]. A low level of $\alpha 3\beta 1$ integrin was found in benign lesions of primary melanoma, whereas in malignant cutaneous melanoma, the expression of the heterodimer progressively increased and was connected with the degree of invasion into the dermis [31].

It is well documented in *in vitro* models that melanoma development and acquisition of the metastatic phenotype are also correlated with the expression of $\alpha v\beta 3$ integrin [26, 32, 33]. An early study by Albelda et al. [34] showed that the $\beta 3$ subunit is restricted to the VGP and metastatic melanomas; in the RGP and in nevus cells, this integrin chain was not found. A study of pairs of differing melanoma cells taken from the same patient (primary WM115 and metastatic WM266-4 cell lines) supported previous observations that in primary melanoma the cells survive without αv integrins, while in disease progression, their growth and functions

depend on this receptor's expression [35]. Our group detected $\alpha v \beta 3$ integrin in both primary RGP-derived (WM35) and metastatic melanoma cells (WM9, WM239 and A397 cell lines) [36, 37]. On the other hand, immunohistochemical staining of $\alpha v \beta 3$ in human tumor tissue samples did not confirm a positive correlation of integrin expression with the melanoma metastatic phenotype; melanoma *in situ* with a pre-invasive phenotype showed the highest level of $\alpha v \beta 3$ expression [38].

Most studies have demonstrated up-regulation of integrin expression in melanoma carcinogenesis; only a few integrin receptors have been found to reduce their expression during disease progression. Ziober et al. [39] found that acquisition of a highly metastatic phenotype by melanoma cells was accompanied by loss of $\alpha 7 \beta 1$ expression.

Enhancement of the expression of most integrins promotes conversion of melanoma from the RGP to the VGP and then acquisition of metastatic competence. The switch in expression from LN-binding to FN-binding integrins was shown to contribute to the movement of melanoma cells from the epidermis to the dermis through degraded basement membrane. Apart from induction of $\alpha v \beta 3$ expression, the involvement of $\alpha 3 \beta 1$ [31], $\alpha 5 \beta 1$, and $\alpha v \beta 5$ integrins in this process has been found [40].

4. Functions of integrins: role of glycosylation

Integrins participate in a wide range of biological processes, including growth, proliferation, differentiation, survival/apoptosis, and cell-cycle regulation [41–44]. Apart from the adhesion function, they mediate cell signaling events [45–47].

Tumor progression requires comprehensive alteration of normal cell-cell and cell-ECM interactions [34, 48]. Integrins are the main adhesion proteins responsible for these changes, mainly due to their altered expression. They contribute to regulation of such processes as angiogenesis, tumor growth and metastasis, as well as cell proliferation, survival and motility [49–53]. Abundant glycosylation of the extracellular domains of integrins also significantly affects the function of these receptors [2, 54, 55].

Glycosylation is one of the most frequent post-translational modifications of transmembrane and secreted proteins. Both integrin chains are subject to this modification [56]. Integrin α subunits are more profusely N-glycosylated than their β partners. Subunits $\alpha 3$, $\alpha 5$, and αv in the polypeptide sequences contain 13, 14, and 13 potential N-glycosylation sites, respectively, whereas the $\beta 1$ and $\beta 3$ chains include 12 and 6 N-glycan-linked sequences, respectively [57]. Intensive glycosylation of integrin chains during post-translational processing results in high content of the sugar component of the whole glycoprotein molecule. Peptide N-glycosidase F (PNGase F) digestion showed that ca. 24 and 25% of the glycoprotein's molecular weight (MW) responds to N-glycans in $\alpha 3$ subunits from WM35 primary and A375 metastatic melanoma cells, respectively. N-oligosaccharides on $\beta 1$ subunits account for ca. 24 and 33% of total MW in primary and metastatic cells, respectively. In both subunits, the pool of sialic acids increases in metastatic cells in compared with primary melanoma [58] (**Figure 2**). N-oligosaccharides on

the αv integrin subunit from WM793 primary melanoma cells respond to nearly 30% of glycoprotein MW, and from WM1205Lu metastatic cells 28%. Subunit $\beta 3$ contains 16% of the N-glycans in WM793 cells and 12% of the N-glycans in WM1205Lu cells [59].

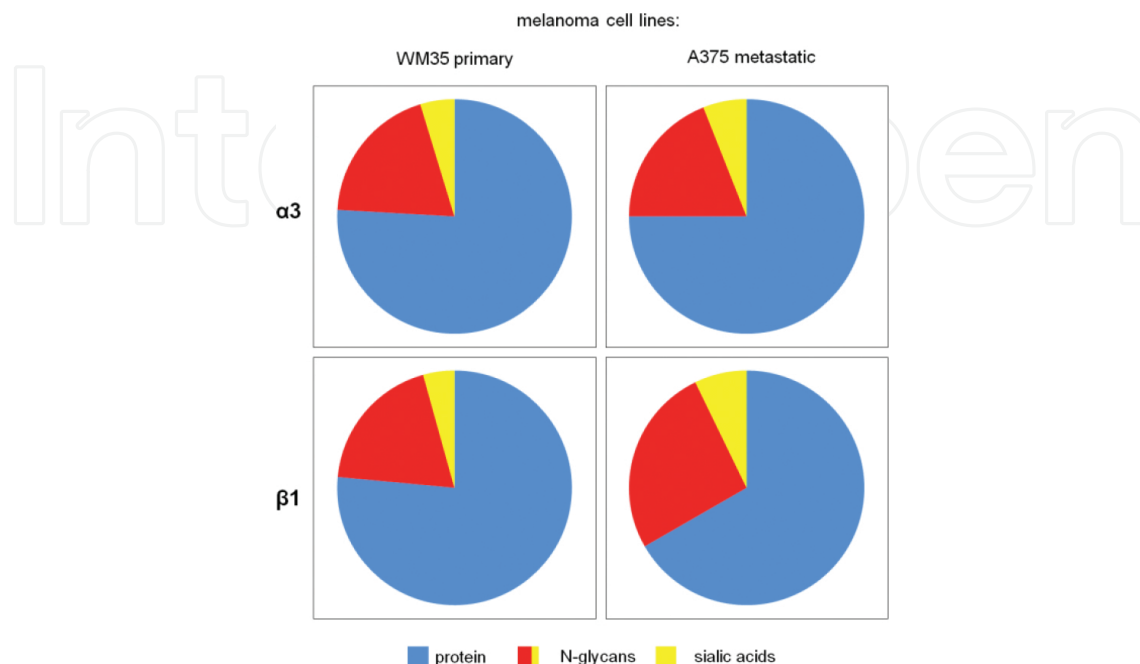


Figure 2. Percentage content of the N-glycan pool and sialic acid in subunits of $\alpha 3\beta 1$ integrin, based on Pocheć et al. [58].

Integrin chains bear all types of N-glycan structures, starting from the evolutionarily oldest structures high-mannose-type, through hybrid glycans, and ending in the most complicated complex-type oligosaccharides [1, 54]. The occurrence of these glycostructures on $\beta 1$ integrins in B16-F10 melanoma cells depends on the stage of integrin maturation. High-mannose glycans recognized by GNA lectin (*Galanthus nivalis* agglutinin) were abundant on the immature form of $\beta 1$ integrins with lower molecular weight. The mature, larger $\beta 1$ chain carried mostly sialylated complex-type structures, identified using DSA (*Datura stramonium* agglutinin) and MAA (*Maackia amurensis* agglutinin) lectins. Only the completely processed form of $\beta 1$ integrin was detected at the cell surface of murine melanoma [60].

Glycosylation is crucial to the processing, activation, and functioning of integrins [56, 61]. The function of integrin glycans has been determined mostly using N-glycan synthesis inhibitors, such as castanospermine and N-methyldeoxynojirimycin, which block glucosidases I and II responsible for trimming glucose from the precursor form of N-linked oligosaccharides; 1-deoxymannojirimycin and swainsonine (SW), inhibitors of mannosidase I and II, respectively, and tunicamycin, which abolishes N-glycosylation by inhibiting the action of N-acetylglucosamine-1-phosphotransferase. Other useful tools for assessing N-glycan functioning are recombinant glycosidases, such as PNGase F, which removes glycans N-linked to the protein backbone, and endo-N-acetylglucosaminidase F (Endo F), which cleaves high-mannose and complex-type N-glycans [62].

Glycosylation of $\alpha v \beta 3$ integrin is necessary to assembly of the heterodimer, proteolytic cleavage of the α chain, and cell surface expression of this VN receptor in human melanoma cells. Application of castanospermine and N-methyldeoxynojirimycin decreased $\alpha v \beta 3$ surface expression as the result of reduced chain assembly and α polypeptide cleavage. On the other hand, 1-deoxymannojirimycin and SW, inhibitors acting on the later stages of glycan synthesis, did not influence $\alpha v \beta 3$ transport to the cell membrane [63]. The importance of N-glycosylation in associating the two subunits was also clearly demonstrated by treating $\alpha 5 \beta 1$ integrin with Endo F and PNGase F. Enzymatic digestion of purified $\alpha 5 \beta 1$ integrin resulted in separate precipitation of the α and β polypeptide chains; undigested integrin subunits underwent co-precipitation [64]. Further research using sequential side-directed mutagenesis showed that N-glycosylation of the I-like domain of the $\beta 1$ subunit is essential for the formation of the $\alpha 5 \beta 1$ heterodimer and for integrin functioning [65].

Cell surface carbohydrates present on adhesion proteins are involved in adhesive and migratory events crucial to each step of the metastatic process. In early studies by Chammas et al., it was found that glycosylation of the $\beta 1$ subunit complexed with $\alpha 6$ integrin is essential for interaction with LN. Binding of B16-F10 melanoma cells to LN via $\alpha 6 \beta 1$ integrin was nearly abolished in tunicamycin-treated cells and after treating LN with Endo F/PNGase F [66]. Similarly, digestion of $\alpha 5 \beta 1$ integrin with a mixture of Endo F and PNGase F led to the loss of FN binding [64]. Lectin analysis showed that both subunits of $\alpha 6 \beta 1$ integrin bear mainly sialylated complex-type N-glycan structures. Exoglycosidase treatment identified galactose residues on the α subunit as the LN-binding determinants involved in cell adhesion to this ECM ligand. The integrin β chain, abundant in complex-type structures, whose synthesis was inhibited by SW (which blocks the formation of complex-type glycans, among them $\beta 1,6$ -branched glycans), was associated with cell spread but not cell adhesion [67]. Also, human metastatic malignant melanoma cell lines LOX and FEMX treated with tunicamycin showed significantly weaker adhesion to LN and to a lesser extent to type IV collagen. Inhibition of N-glycan synthesis by tunicamycin resulted in reduction of LOX and FEMX invasion through Matrigel-coated chambers, as well as diminution of human melanoma aggregation [68].

5. Alterations of integrin glycosylation in melanoma carcinogenesis

The vast majority of studies on integrin glycosylation in melanoma have used mouse melanoma cell line B16-F10 and phenotypic variants of it that show different degrees of invasive potential, mainly the weakly invasive cell lines B16-F1 or B16-Flr, and B16-BL6 cells selected for their higher ability to metastasize to the lungs [60, 67, 69–73], as well as human melanoma cell lines derived from each stage of melanoma progression, most of which were established by Herlyn's group [3].

It has been demonstrated that the glycosylation profile of integrins depends on the stage of melanoma development [37, 58, 59, 74] and the location of the metastasis [75, 76] and that glycosylation is essential to the interaction between integrin and ECM proteins during adhesion and migration processes [58, 75, 76]. These studies have produced ample evidence

for the presence of glycoforms associated with melanoma carcinogenesis on $\alpha 3\beta 1$, $\alpha 5\beta 1$, and $\alpha v\beta 3$ integrins. The changes in the $\beta 1,6$ branching of complex-type N-glycans, and their sialylation, have been observed on these integrins during human melanoma progression.

5.1. Branched complex-type N-glycans

One of the well-characterized changes in N-glycosylation is enhanced expression of $\beta 1,6$ -N-acetylglucosaminyltransferase V (GnT-V) and its products, N-acetylglucosamine (GlcNAc) $\beta 1,6$ -branched N-linked oligosaccharides, observed in the tumorigenesis of many cancers [77–81], including melanoma [74]. $\beta 1,6$ -branched N-glycans are important in invasion of the basement membrane [82] and acquisition of metastatic competence [83]. $\beta 1,6$ branching of glycans on integrin chains has been described in studies of mouse and human melanoma.

The presence of $\beta 1,6$ -branched complex-type oligosaccharides on the integrin receptors that bind LN and FN was first shown by Chammas et al. in mouse melanoma cell line B16-F10 [70] and then confirmed on the $\beta 1$ subunit sharing integrins in this parent cell line and its highly invasive B16-BL6 variant [73, 84, 85]. Significantly enhanced $\beta 1,6$ branching found on highly invasive B16-BL6 cells resulted in their more efficient invasion and migration, as well as impaired adhesion to different ECM proteins (LN, FN, VN, type I and type IV collagen, hyaluronic acid, and Matrigel). Inhibition of $\beta 1,6$ branching on two levels—expression of GnT-V by cell transfection (using antisense cDNA), and oligosaccharide synthesis (using SW)—decreased metastasis and invasion of B16-BL6 cells by half, and reduced the formation of metastatic colonies in lungs [73]. Later it was found that $\alpha 3\beta 1$ and $\alpha 5\beta 1$ integrins on mouse B16-BL6 cells carry $\beta 1,6$ -branched oligosaccharides and that $\beta 1,6$ -glycosylation of integrins has an effect on the spread of melanoma cells on FN and Matrigel. Interestingly, $\beta 1,6$ -branched glycans on $\alpha 3\beta 1$ weakened the association of integrin with CD151 tetraspanin [85]. Earlier the crucial role of glycosylation in the interaction of $\alpha 3\beta 1$ with CD151 had been described in work using MDA-MB-231 human breast cancer cells [86]. For B16-BL6 mouse melanoma cells, it was shown (by co-precipitating $\alpha 3\beta 1$ and CD151 from SW-treated cells) that $\beta 1,6$ -branched N-glycans regulate the association of CD151 with this integrin [85].

In human melanoma cells, we demonstrated $\beta 1,6$ branching of cancer-associated integrin subunits such as $\alpha 2$, $\alpha 3$, $\alpha 4$, $\alpha 5$, αv , $\beta 1$, and $\beta 3$ [37]; integrin heterodimers of special importance in melanoma carcinogenesis are $\alpha 3\beta 1$ [58, 75], $\alpha 5\beta 1$ [74], and $\alpha v\beta 3$ [59, 75, 87].

A number of studies have confirmed the involvement of $\alpha 3\beta 1$ integrin in melanoma development through its participation in cell adhesion, migration, and invasion [88–90]. The ability of $\alpha 3\beta 1$ to promote melanoma metastasis results from its enhanced synthesis [91, 92] and also from altered glycosylation of it, particularly enhanced $\beta 1,6$ branching [58, 74, 75].

Glycosylation of $\alpha 3\beta 1$ integrin was first recognized as a factor promoting tumorigenesis in human colon carcinoma cells. Sialylated $\beta 1,6$ -branched Asn-linked oligosaccharides with short poly-N-acetyllactosamine units were found on both integrin subunits. Due to their role in cancer development, they were suggested to be oncodevelopmental carbohydrate epitopes [93].

Different techniques have been employed to analyze $\alpha 3\beta 1$ integrin glycosylation in detail in melanoma cells derived from primary and metastatic tumors. The use of matrix-assisted laser desorption/ionization mass spectrometry (MALDI-MS) showed the presence of tetra-antennary complex-type glycans on the $\beta 1$ subunit in highly metastatic A375 melanoma cells but not in WM35 cells from the primary site. The reaction of affinity-chromatography-purified $\alpha 3\beta 1$ integrin with *Phaseolus vulgaris* agglutinin (PHA-L) revealed that complex-type glycans are $\beta 1,6$ -branched in the $\alpha 3$ subunit from metastatic but not from primary cells [58]. The presence of GlcNAc $\beta 1,6$ -branched glycans on $\alpha 3\beta 1$ in A375 metastatic cells was confirmed by tandem mass spectrometry (MS/MS) of PHA-L-positive glycoproteins eluted in lectin-affinity chromatography [36]. The absence of this type of branching on the $\alpha 3$ subunit in WM35 primary melanoma was thoroughly documented by MS/MS identification of PHA-L bound proteins and two-sided control of integrin glycosylation: immunoblotting in PHA-L-eluted material and PHA-L blotting in immunoprecipitation [37]. In two other metastatic melanoma cell lines (WM9 and WM239), $\beta 1,6$ branching of $\alpha 3\beta 1$ integrin was shown using MS/MS identification of PHA-L-bound glycoproteins [37] and confirmed using MALDI-MS and PHA-L precipitation [75]. The amount of glycans with $\beta 1,6$ -linked antenna increased in WM1205Lu metastatic melanoma as compared to WM793 primary cells [74]. Using normal-phase high-performance liquid chromatography (NP-HPLC), however, Link-Lenczowski et al. [94] did not observe differences in $\alpha 3\beta 1$ glycosylation profiles between WM115 primary and WM266-4 metastatic human melanoma cell lines originating from the same patient.

The role of $\alpha 5$ integrin in promoting melanoma metastasis has been shown in uveal [95] and cutaneous melanoma [29, 96]. An increase of the metastatic potential of melanoma is accompanied by enhancement of $\alpha 5$ integrin expression [30, 97]. In highly metastatic B16-F10 melanoma cells, the level of $\alpha 5$ integrin was conspicuously elevated as compared to weakly metastatic B16-F1 cells. Pulmonary metastasis in mice as well as the adhesion and spread of B16-F10 cells to FN *in vitro* was significantly reduced after blocking of $\alpha 5$ integrin by a specific antibody. The loss of $\alpha 5$ -mediated melanoma cell-FN anchoring promoted apoptosis of B16-F10 cells [30].

Integrin $\alpha 5\beta 1$ is also a carrier of $\beta 1,6$ -branched glycans in metastatic cells, but on the $\alpha 5$ subunit from primary melanoma, this type of branching was not detected. In each of three analyzed metastatic cell lines (WM9, WM239 and A375), the $\alpha 5$ subunit oligosaccharides were $\beta 1,6$ -branched [37], but not the $\alpha 5$ chain in WM35 melanoma cells [36], as determined using MS/MS analysis of PHA-L-positive glycoproteins. A comparison of $\alpha 5$ integrin chains from early VGP and metastatic lesion cells showed an uptrend of $\beta 1,6$ branching during acquisition of metastatic competence [74]. These findings suggest that GlcNAc $\beta 1,6$ -branched structures appear earlier in melanoma development on the $\beta 1$ subunit than on the $\alpha 3$ and $\alpha 5$ chains and that in melanoma cancerogenesis their content is more stable on the $\beta 1$ subunit than on the $\alpha 3$ and $\alpha 5$ chains [36, 37, 74].

Glycosylation of integrin $\alpha v\beta 3$ is still rather poorly understood [54], although it is well known that this integrin is associated with the metastatic potential of melanoma [33, 35, 98]. Our studies using two genetically related melanoma cell lines showed the presence of $\beta 1,6$ -branched complex-type structures on primary and metastatic cells, but we did not observe

differences in the β 1,6 branching of α v β 3 glycans during the transition from primary VGP melanoma to its metastatic variant. PHA-L precipitation and SW treatment gave similar levels of β 1,6 branching in both α v β 3 subunits in cell lines WM793 and WM1205Lu [59]. This type of glycan was also present on the α v subunit from RGP-derived WM35 melanoma cells, but β 1,6 branching was not found on the β 3 chain from these cells [36]. Integrin α v β 3 from three metastatic cell lines (WM9, WM239 and A375) of varying origin showed expression of these structures [37, 75].

The phenomenon of competition for a substrate between N-acetylglucosaminyltransferase III (GnT-III) and GnT-V is well documented in N-glycan biology. GnT-III activity during N-glycan processing can suppress the biological functions of GnT-V; it results in reduction of N-glycan β 1,6 branching. With respect to integrins, this was first shown on α 3 β 1 in human gastric cancer cell line MKN45 [99]. In B16 melanoma cells, ectopic expression of GnT-III was shown to retard cell metastasis through inhibition of GnT-V activity: the absence of GnT-V products was associated with attenuation of malignant cell motility [83]. Our group showed a significant decrease of bisecting GlcNAc content on α v β 3 integrin subunits during the transition from the VGP to the metastatic stage, but it was not associated with any change in the amount of β 1,6-branched glycans on this integrin [59], although previously in this pair of related cell lines (WM115 vs. WM1205Lu), we observed significant upregulation of GnT-V expression [74].

Integrin-mediated cell migration requires adhesion of cells to ECM substrates and is essential for dissemination of the tumor to distant organs during metastasis [100], so the role of integrin glycosylation is frequently assessed in different adhesion and migration tests. Functional studies have clearly shown that β 1,6 branching on cell surface adhesion receptors, mainly integrins, promotes melanoma cell migration [101], and invasion [90].

The contribution of α 3 β 1 integrin's N-glycans to its binding with its ECM ligands was demonstrated using affinity-chromatography-purified integrin from WM35 primary and A375 metastatic melanoma cells. In direct ligand-binding assays, de-N-glycosylated α 3 β 1 integrin showed enhanced binding of both melanoma cell lines to LN, type IV collagen and FN, except for the binding of α 3 β 1 from WM35 to FN [58]. Enzymatic removal of N-glycans from this integrin in two metastatic melanoma cell lines from metastases of different origin (WM9 and WM239) also resulted in enhanced binding of α 3 β 1 to LN5 [75].

Of the ECM proteins, fibronectin is the major α 5 β 1 ligand [102] and therefore is the one most frequently chosen for assays evaluating the involvement of α 5 β 1 integrin in adhesion and migration processes. β 1,6 branching of FN receptors was shown to contribute to migration of metastatic melanoma on FN, but not to primary cell migration [74].

N-glycan-dependent binding of integrins to the ECM triggers intracellular pathways via phosphorylation of cytoplasmic kinases. FAK is one of the first proteins recruited to integrins aggregated within the cell membrane. Activation of signal pathways leads to the expression of different genes that control cell growth, differentiation, tumor invasion and metastasis [103, 104]. Changes in integrin glycosylation affect intracellular signals triggered by melanoma cell binding to the ECM. Dual immunostaining of melanoma cells growing on VN showed co-localization of α v β 3 integrin and FAK, a downstream target of integrins, in focal adhesion sites

of melanoma cells. Overexpression of GnT-V in human WM266-4 metastatic melanoma cells up-regulated $\alpha v\beta 3$ -integrin-mediated FAK phosphorylation and cell migration on VN, while inhibition of $\beta 1,6$ branching by SW-treatment reduced FAK signaling activation in both A375 and WM266-4 metastatic cells [87].

An interesting aspect of integrin glycans' involvement in melanoma metastasis is their participation in ECM degradation through regulation of the activity of matrix proteases, such as urokinase-type plasminogen activator (uPA) and metalloproteinases (MMPs). Integrins interact with urokinase-type plasminogen activator receptors (uPARs) in the cell membrane [105]. A urokinase-type plasminogen activator (uPA), acting via its receptor (uPAR), catalyzes the activation of plasmin from plasminogen, and the plasmin initiates a proteolytic cascade leading to degradation of the ECM [106, 107]. Our work demonstrated that $\beta 1,6$ -branched oligosaccharides on $\alpha v\beta 3$ and $\alpha 3\beta 1$ integrins are essential for the association of the uPAR with integrins in human melanoma cell lines WM9 and WM239, seen in the failure of co-precipitation of the two integrins with the uPAR in SW-treated cells. Adhesion of the two melanoma cells to VN was dependant on $\beta 1,6$ branching of $\alpha v\beta 3$ and $\alpha 3\beta 1$ integrins in a cell-line-specific manner [76].

N-glycans with $\beta 1,6$ -branched antennae on melanoma integrins also modify the activity of metalloproteinases (MMPs). PHA-L precipitation revealed that $\beta 1$ integrins from B16-BL6 cells are more $\beta 1,6$ -branched than the parent cells with lower invasion ability. $\beta 1,6$ -glycosylation of $\beta 1$ integrin receptors affected the activation of membrane-tethered forms of metalloproteinases (MT1-MMPs). The association of $\beta 1,6$ -glycosylation-suppressed $\beta 1$ integrin with MT1-MMPs was more severely affected in B16-BL6 cells than in the parent cells, suggesting that integrin $\beta 1,6$ branching contributes to melanoma invasion also through activation of MMPs [84].

5.2. Sialylation

Sialic acid-linked $\alpha 2,3$ or $\alpha 2,6$, mostly in terminal positions of the oligosaccharide, gives these molecules a negative charge [108, 109] that significantly influences cell interaction mediated by sialylated adhesion proteins, among them integrins [110]. Hypersialylation of cell surface receptors is important in tumor invasion and metastasis [111]. MAA is the lectin commonly used to analyze a pool of $\alpha 2,3$ -linked sialic acid, while a lectin from *Sambucus nigra* (SNA) is specific for $\alpha 2,6$ -linked sialic acid [112]. The presence of sialic acids on $\alpha 3\beta 1$, $\alpha 5\beta 1$, and $\alpha v\beta 3$ integrins in melanoma cells was confirmed in each stage of melanoma progression [58, 59, 75].

One of the first studies on integrin sialylation in melanoma employed mouse melanoma cell lines differing in their metastatic ability. Analysis of melanoma cell sialylation using HPLC and digestion by *Vibrio cholerae* sialidase did not show changes in the total content of cell surface sialic acids on mouse B16 metastatic melanoma cell variants differing in their invasive potential [69]. Research on specific adhesion proteins provided more detailed information. Integrin $\beta 1$ from both B16-F1 mouse metastatic melanoma and its weakly metastatic wheat germ agglutinin-resistant mutant Wa4-b1 was found to contain high-mannose and bi-, tri-, and tetra-antennary complex-type N-oligosaccharides. Sialylation of the $\beta 1$ subunit was significantly decreased in mutant melanoma cells with low metastatic ability. Alteration of $\beta 1$ integrin

glycosylation resulted in reduction of the mutant's metastatic potential and adhesion to FN and LN, as compared to the parent cells [71]. Higher β 1,6 branching of complex-type glycans on more invasive B16-BL6 melanoma cells versus the parent B16-F10 line was correlated with an increase of α 2,3-linked and α 2,6-linked sialic acid content as determined using MAA and SNA staining in flow cytometry. Hypersialylation of B16-BL6 cells resulted in their higher motility and stronger adhesion to selected ECM proteins [73]. Further results for this pair of murine melanoma cell lines were obtained by lectin blotting: α 2,6-linked sialic acid especially increased on B16-BL6 glycans as a result of enhanced β 1,6 branching. α 2,6-desialylation and down-regulation of the sialyltransferase ST6Gal-I, which transfers sialic acids to oligosaccharides and catalyzes the formation of α 2,6 linkage, negatively affected adhesion and invasion of B16-BL6 cells [113]. In turn, a study by Chang and colleagues showed that α 2,3-linked sialic acid is important in the metastasis of B16-F10 cells. Soyasaponin I (Ssa I), which specifically inhibits the expression of α 2,3-linked sialic acids, reduced the migratory ability of melanoma, up-regulated cell adhesion to ECM proteins, and impaired pulmonary metastasis [114].

Our studies using different human melanoma cell line models indicated reduction of α 2,3 sialylation on the α 3 integrin subunit, and of α 2,6 sialylation on α v β 3 integrin, in melanoma progression [74, 59]. Lectin-probed Western blotting showed that the β 1 subunit from both cell lines and the α 3 subunit from primary melanoma cell line WM35 had both types of sialic acid linkage, while the α 3 subunit from metastatic cell line A375 lost its α 2,3 glycosidic linkage [58]. Using genetically matched cell lines WM793 and WM1205Lu from the last two stages of melanoma progression, we observed a shift in the sialylation of α v β 3 integrin during the transition from VGP to metastatic tumor. Lectin MAA and SNA precipitation as well as digestion by two neuraminidases with narrower (α 2,3) and wider (α 2-3,6,8) specificity showed that α 2,6-linked sialic acid was reduced, whereas α 2,3-linked sialic acid increased on both integrin subunits from metastatic lesion cells. In a wound-healing assay, migration of melanoma cells on VN in the presence of both lectins was affected only in the metastatic cell line [59]. Lectin flow cytometry of another pair of related melanoma cell lines (WM115 derived from RGP/VGP vs. WM266-4 from lymph node metastasis) indicated a more than fourfold increase of cell surface α 2,3 sialylation during the acquisition of metastatic competence. Despite these differences in surface α 2,3 sialylation, the reduction of migration by MAA-treated primary and melanoma cells was comparable, suggesting the involvement of receptor(s) other than α v β 3 integrin and its/their sialylation in metastatic cell migration (data not published).

Digestion of α 3 β 1 glycans with a broad-specificity neuraminidase from *Arthrobacter ureafaciens* led to stronger binding of the integrin to various ECM components (LN, FN, and type IV collagen) in both primary and metastatic melanoma cells. Interestingly, removal of the sialic acids by neuraminidase enhanced integrin binding significantly more than complete de-N-glycosylation did, suggesting an important role of desialylated N-oligosaccharides in integrin-ECM interactions [58]. For efficient cell-ECM adhesion, protein-protein interactions apparently are not enough, and glycosylation is needed to regulate this binding.

Attachment of α 2,8 to underlying glycans by sialic acid is rather rarely detected on integrins. A study using human melanoma cell line G361 is one of the few that have demonstrated the

presence of $\alpha 2,8$ -bound sialic acid on $\alpha 5\beta 1$ integrin—and the role of this type of sialylation in FN binding. Desialylation using an enzyme from *Arthrobacter ureafaciens* specific for $\alpha 2$ -3,6,8-linked sialic acids resulted in reduction of $\alpha 5\beta 1$ -mediated adhesion to FN, an effect not observed for neuraminidase, which cleaves only $\alpha 2$ -3,6 linkages [115].

Undoubtedly, the sialylation state of integrins contributes to the metastatic potential of mouse and human melanoma, but there are blank spots in our understanding of the role of $\alpha 2,3$ -linked and $\alpha 2,6$ -linked sialic acid in melanoma progression. Further studies should establish precisely how sialylation becomes altered, and its contribution to the disease phenotype.

6. Conclusions

The search for glyco-biomarkers on integrins in melanoma progression motivates a host of studies performed by different research groups. Identification of universally present alterations of glycans on adhesion molecules, among them integrins—and elucidation of the molecular mechanisms of these changes—will boost our understanding of how melanoma cells acquire the ability to escape the primary tumor and spread through the body. Enhanced $\beta 1,6$ branching and altered sialylation are the main glyco-features of integrin glycosylation in melanoma progression. The functional consequences of surface glycosylation rearrangements in melanoma progression must be known if we are to find effective ways to stop the process of carcinogenesis. The vast majority of studies on integrin glycosylation in melanoma cells have used cells cultured *in vitro*. A hugely important task for future research is to verify the results obtained from *in vitro* studies of tumor tissue from patients with melanoma, so that those findings can be applied for prevention and treatment of melanoma.

Author details

Ewa Pocheć* and Anna Lityńska

*Address all correspondence to: ewa.pochec@uj.edu.pl

Department of Glycoconjugate Biochemistry, Institute of Zoology, Jagiellonian University, Krakow, Poland

References

- [1] Link-Lenczowski P, Lityńska A. Glycans in melanoma screening. Part 2. Towards the understanding of integrin N-glycosylation in melanoma. *Biochem Soc Trans.* 2011;39(1):374–377. doi:10.1042/BST0390374

- [2] Gu J, Taniguchi N. Potential of N-glycan in cell adhesion and migration as either a positive or negative regulator. *Cell Adh Migr.* 2008;2(4):243–245. doi:10.4161/cam.2.4.67.48
- [3] Herlyn M. Human melanoma: development and progression. *Cancer Metastasis Rev.* 1990;9(2):101–112. doi:10.1007/BF00046337
- [4] Seftor RE, Seftor EA, Hendrix MJ. Molecular role(s) for integrins in human melanoma invasion. *Cancer Metastasis Rev.* 1999;18(3):359–375. doi:10.1023/A:1006317125454
- [5] McGary EC, Lev DC, Bar-Eli M. Cellular adhesion pathways and metastatic potential of human melanoma. *Cancer Biol Ther.* 2002;1(5):459–465. doi:10.4161/cbt.1.5.158
- [6] Satyamoorthy K, Herlyn M. Cellular and molecular biology of human melanoma. *Cancer Biol Ther.* 2002;1(1):14–17. doi:10.4161/cbt.1.1.32
- [7] Chudnovsky Y, Khavari PA, Adams AE. Melanoma genetics and the development of rational therapeutics. *J Clin Invest.* 2005;115(4):813–824. doi:10.1172/JCI24808
- [8] Monteiro AC, Toricelli M, Jasiulionis MG. Signaling pathways altered during the metastatic progression of melanoma. In: Murph M. editor. *Current clinical management and future therapeutics.* InTech; 2015. p. 49–78. doi:10.5772/59747
- [9] Hynes RO. Integrins: a family of cell surface receptors. *Cell.* 1987;48:549–554. doi:10.1016/0092-8674(87)90233-9
- [10] Desgrosellier JS, Cheresch DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer.* 2010;10(1):9–22. doi:10.1038/nrc2748
- [11] Gahmberg CG, Fagerholm SC, Nurmi SM, Chavakis T, Marcheas S, Gronhold M. Regulation of integrin activity and signaling. *Biochim. Biophys. Acta.* 2009;1790(6):431–444. doi:10.1016/j.bbagen.2009.03.007
- [12] Cambell ID, Humphries MJ. Integrin structure, activation, and interactions. *Cold Spring Harb Perspect Biol.* 2011;3(3). pii:a004994. doi:10.1101/cshperspect.a004994
- [13] Xiong JP, Stehle T, Zhang R, Joachimiak A, Frech M, Goodman SL, Arnaout MA. Crystal structure of the extracellular segment of integrin $\alpha v \beta 3$ in complex with an Arg-Gly-Asp ligand. *Science.* 2002;296:151–155. doi:10.1126/science.1069040
- [14] Zhu J, Luo BH, Xiao T, Zhang C, Nishida N, Springer TA. Structure of a complete integrin ectodomain in a physiologic resting state and activation and deactivation by applied forces. *Mol Cell.* 2008;32:849–861. doi:10.1016/j.molcel.2008.11.018
- [15] Xiong JP, Stehle T, Diefenbach B, Zhang R, Dunker R, Scott DL, Joachimiak A, Goodman SL, Arnaout MA. Crystal structure of the extracellular segment of integrin $\alpha v \beta 3$. *Science.* 2001;294(5541):339–345. doi:10.1126/science.1064535

- [16] Wegener KL, Campbell ID. Transmembrane and cytoplasmic domains in integrin activation and protein-protein interactions. *Mol Membr Biol.* 2008;25(5):376–387. doi:10.1080/09687680802269886
- [17] Barczyk M, Carracedo S, Gullberg D. Integrins. *Cell Tissue Res.* 2010;339(1):269–280. doi:10.1007/s00441-009-0834-6
- [18] Wegener KL, Partridge AW, Han J, Pickford AR, Liddington RC, Ginsberg MH, Campbell ID. Structural basis of integrin activation by talin. *Cell.* 2007;128(1):171–182. doi:10.1016/j.cell.2006.10.048
- [19] Montanez E, Ussar S, Schifferer M, Bosl M, Zent R, Moser M, Fassler R. Kindlin-2 controls bidirectional signaling of integrins. *Genes Dev.* 2008;22:1325–1330. doi:10.1101/gad.469408.
- [20] Zaidel-Bar R, Itzkovitz S, Ma'ayan A, Iyengar R, Geiger B. Functional atlas of the integrin adhesome. *Nat Cell Biol.* 2007;9:858–867. doi:10.1038/ncb0807-858
- [21] Takada Y, Ye X, Simon S. The integrins. *Genome Biol.* 2007;8(5):215. doi:10.1186/gb-2007-8-5-215
- [22] Srichai MB, Zent R. Integrin structure and function. In: Zent R, Pozzi A, editors. *Cell-extracellular matrix interactions in cancer*. 1st ed. New York: Springer-Verlag; 2010. p. 19–41. doi:10.1007/978-1-4419-0814-8
- [23] Albelda SM, Buck CA. Integrins and other cell adhesion molecules. *FASEB J.* 1990;4(11):2868–2880. doi:10.1038/346425a0
- [24] Kuphal S, Bauer R, Bosserhoff A-K. Integrin signaling in malignant melanoma. *Cancer Metastasis Rev.* 2005;24(2):195–222. doi:10.1007/s10555-005-1572-1
- [25] Mizejewski GJ. Role of integrins in cancer: survey of expression patterns. *Proc Soc Exp Biol Med.* 1999;222(2):124–138. doi:10.1046/j.1525-1373.1999.d01-122.x
- [26] Cheresch DA. Structure, function and biological properties of integrin alpha v beta 3 on human melanoma cells. *Cancer Metastasis Rev.* 1991;10(1):3–10. doi:10.1007/BF00046839
- [27] Kramer RH, Vu M, Cheng YF, Ramos DM. Integrin expression in malignant melanoma. *Cancer Metastasis Rev.* 1991;10(1):49–59. doi:10.1007/BF00046843
- [28] Felding-Habermann B. Integrin adhesion receptors in tumor metastasis. *Clin Exp Metastasis.* 2003;20(3):203–213. doi:10.1023/A:1022983000355
- [29] Laidler P, Gil D, Pituch-Noworolska A, Ciołczyk D, Ksiazek D, Przybyło M, Lityńska A. Expression of beta1-integrins and N-cadherin in bladder cancer and melanoma cell lines. *Acta Biochim Pol.* 2000;47(4):1159–1170.

- [30] Qian F, Zhang ZC, Wu XF, Li YP, Xu Q. Interaction between integrin $\alpha(5)$ and fibronectin is required for metastasis of B16F10 melanoma cells. *Biochem Biophys Res Commun.* 2005;333(4):1269–1275. doi:10.1016/j.bbrc.2005.06.039
- [31] Natali PG, Nicotra MR, Bartolazzi A, Cavaliere R, Bigotti A. Integrin expression in cutaneous malignant melanoma: association of the $\alpha 3/\beta 1$ heterodimer with tumor progression. *Int J Cancer.* 1993;54(1):68–72. doi:10.1002/ijc.2910540112
- [32] Gehlsen KR, Davis GE, Sriramaraio P. Integrin expression in human melanoma cells with differing invasive and metastatic properties. *Clin Exp Metastasis.* 1992;10(2):111–120. doi:10.1007/BF00114587
- [33] Voura EB, Ramjeesingh RA, Montgomery AM, Siu CH. Involvement of integrin $\alpha(v)\beta(3)$ and cell adhesion molecule L1 in transendothelial migration of melanoma cells. *Mol Biol Cell.* 2001;12(9):2699–2710. doi:10.1091/mbc.12.9.2699
- [34] Albelda SM, Mette SA, Elder DE, Stewart R, Damjanovich L, Herlyn M, Buck CA. Integrin distribution in malignant melanoma: association of the $\beta 3$ subunit with tumor progression. *Cancer Res.* 1990;50(20):6757–6764.
- [35] Koistinen P, Ahonen M, Kähäri VM, Heino J. αV integrin promotes *in vitro* and *in vivo* survival of cells in metastatic melanoma. *Int J Cancer.* 2004;112(1):61–70. doi:10.1002/ijc.20377
- [36] Ochwat D, Hoja-Łukowicz D, Lityńska A. N-glycoproteins bearing $\beta 1$ –6 branched oligosaccharides from the A375 human melanoma cell line analysed by tandem mass spectrometry. *Melanoma Res.* 2004;14(6):479–485.
- [37] Przybyło M, Martuszevska D, Pocheć E, Hoja-Łukowicz D, Lityńska A. Identification of proteins bearing $\beta 1$ –6 branched N-glycans in human melanoma cell lines from different progression stages by tandem mass spectrometry analysis. *Biochim Biophys Acta.* 2007;1770(9):1427–1435. doi:10.1016/j.bbagen.2007.05.006
- [38] Neto DS, Pantaleão L, de Sα BC, Landman G. Alpha-v-beta3 integrin expression in melanocytic nevi and cutaneous melanoma. *J Cutan Pathol.* 2007;34(11):851–856. doi:10.1111/j.1600-0560.2007.00730.x
- [39] Ziober BL, Chen YQ, Ramos DM, Waleh N, Kramer RH. Expression of the $\alpha 7\beta 1$ laminin receptor suppresses melanoma growth and metastatic potential. *Cell Growth Differ.* 1999;10(7):479–490.
- [40] Danen EHJ. Integrin signaling as a cancer drug target. *ISRN Cell Biol.* 2013;Article ID 135164, doi:10.1155/2013/135164
- [41] Frisch SM, Ruoslahti E. Integrins and anoikis. *Curr Opin Cell Biol.* 1997;9(5):701–706. doi:10.1016/S0955-0674(97)80124-X
- [42] van der Flier A, Sonnenberg A. Function and interactions of integrins. *Cell Tissue Res.* 2001;305(3):285–298. doi:10.1007/s004410100417

- [43] Watt FM. Role of integrins in regulating epidermal adhesion, growth and differentiation. *EMBO J.* 2002;21(15):3919–3926. doi:10.1093/emboj/cdf399
- [44] Rathinam R, Alahari SK. Important role of integrins in the cancer biology. *Cancer Metastasis Rev.* 2010;9(1):2232–2237. doi:10.1007/s10555-010-9211-x
- [45] Yamada KM, Even-Ram S. Integrin regulation of growth factor receptors. *Nat Cell Biol.* 2002;4(4):E75–E76. doi:10.1038/ncb0402-e75
- [46] Hehlhans S, Haase M, Cordes N. Signalling via integrins: implications for cell survival and anticancer strategies. *Biochim Biophys Acta.* 2007;1775(1):163–180. doi:10.1016/j.bbcan.2006.09.001
- [47] Harburger DS, Calderwood DA. Integrin signalling at a glance. *J Cell Sci.* 2009;122(Pt 2):159–163. doi:10.1242/jcs.018093
- [48] Christiansen MN, Chik J, Lee L, Anugraham M, Abrahams JL, Packer NH. Cell surface protein glycosylation in cancer. *Proteomics.* 2014;14(4–5):525–546. doi:10.1002/pmic.201300387
- [49] Schadendorf D, Gawlik C, Haney U, Ostmeier H, Suter L, Czarnetzki BM. Tumour progression and metastatic behaviour *in vivo* correlates with integrin expression on melanocytic tumours. *J Pathol.* 1993;170(4):429–434. doi:10.1002/path.1711700405
- [50] Guo W, Giancotti FG. Integrin signalling during tumour progression. *Nat Rev Mol Cell Biol.* 2004;5(10):816–826. doi:10.1038/nrm1490
- [51] Danen EHJ. Integrins: regulators of tissue function and cancer progression. *Curr Pharm Des.* 2005;11(7):881–891. doi:10.2174/1381612053381756
- [52] Ganguly KK, Pal S, Moulik S, Chatterjee A. Integrins and metastasis. *Cell Adh Migr.* 2013;7(3):251–261. doi:10.4161/cam.23840
- [53] Seguin L, Desgrosellier JS, Weis SM, Cheresch DA. Integrins and cancer: regulators of cancer stemness, metastasis, and drug resistance. *Trends Cell Biol.* 2015;25(4):234–240. doi:10.1016/j.tcb.2014.12.006
- [54] Janik ME, Lityńska A, Vereecken P. Cell migration—the role of integrin glycosylation. *Biochim Biophys Acta.* 2010;1800(6):545–555. doi:10.1016/j.bbagen.2010.03.013
- [55] Yuan Y, Wu L, Shen S, Wu S, Burdick MM. Effect of alpha 2,6 sialylation on integrin-mediated adhesion of breast cancer cells to fibronectin and collagen IV. *Life Sci.* 2016;149:138–145. doi:10.1016/j.lfs.2016.02.071
- [56] Gu J, Taniguchi N. Regulation of integrin functions by N-glycans. *Glycoconj J.* 2004;21(1–2):9–15. doi:10.1023/B:GLYC.0000043741.47559.30
- [57] Pigott R, Power C. The adhesion molecules facts book. 1st ed. London: Academic Press; 1993.

- [58] Pocheć E, Lityńska A, Amoresano A, Casbarra A. Glycosylation profile of integrin alpha 3 beta 1 changes with melanoma progression. *Biochim Biophys Acta*. 2003;1643(1–3): 113–123. doi:10.1016/j.bbamcr.2003.10.004
- [59] Pocheć E, Bubka M, Rydlewska M, Janik M, Pokrywka M, Lityńska A. Aberrant glycosylation of $\alpha\text{v}\beta 3$ integrin is associated with melanoma progression. *Anticancer Res*. 2015;5(4):2093–2103.
- [60] Veiga SS, Chammas R, Cella N, Brentani RR. Glycosylation of beta-1 integrins in B16-F10 mouse melanoma cells as determinant of differential binding and acquisition of biological activity. *Int J Cancer*. 1995;61(3):420–424. doi:10.1002/ijc.2910610324
- [61] Fuster MM, Esko JD. The sweet and sour of cancer: glycans as novel therapeutic targets. *Nat Rev Cancer*. 2005;5(7):526–542. doi:10.1038/nrc1649
- [62] Stanley P, Schachter H, Taniguchi N. N-glycans. In: Varki A, Cummings RD, Esko JD, Freeze HH, Stanley P, Bertozzi CR, Hart GW, Etzler ME, editors. *Essentials of glycobiology*. 2nd ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 2009.
- [63] Spiro RC, Laufer DM, Perry SK, Harper JR. Effect of inhibitors of N-linked oligosaccharide processing on the cell surface expression of a melanoma integrin. *J Cell Biochem*. 1989;41(1):37–45. doi:10.1002/jcb.240410105
- [64] Zheng M, Fang H, Hakomori S. Functional role of N-glycosylation in alpha 5 beta 1 integrin receptor. De-N-glycosylation induces dissociation or altered association of alpha 5 and beta 1 subunits and concomitant loss of fibronectin binding activity. *J Biol Chem*. 1994;269(16):12325–12331.
- [65] Isaji T, Sato Y, Fukuda T, Gu J. N-glycosylation of the I-like domain of $\beta 1$ integrin domain is essential for $\beta 1$ integrin expression and biological function. *J Biol Chem*. 2009;284(18):12207–12216. doi:10.1074/jbc.M807920200
- [66] Chammas R, Veiga SS, Line S, Potocnjak P, Brentani RR. Asn-linked oligosaccharide-dependent interaction between laminin and gp120/140. An alpha 6/beta 1 integrin. *J Biol Chem*. 1991;266(5):3349–3355.
- [67] Chammas R, Veiga SS, Travassos LR, Brentani RR. Functionally distinct roles for glycosylation of alpha and beta integrin chains in cell-matrix interactions. *Proc Natl Acad Sci U S A*. 1993;90(5):1795–1799. doi:10.1073/pnas.90.5.1795
- [68] Bironaite D, Nesland JM, Dalen H, Risberg B, Bryne M. N-Glycans influence the *in vitro* adhesive and invasive behaviour of three metastatic cell lines. *Tumour Biol*. 2000;21(3): 165–175. doi:10.1159/000030123
- [69] Passaniti A, Hart GW. Cell surface sialylation and tumor metastasis. Metastatic potential of B16 melanoma variants correlates with their relative numbers of specific penultimate oligosaccharide structures. *J Biol Chem*. 1988;263(16):7591–7603.

- [70] Chammas R, Veiga SS, Brentani RR. Glycobiology of laminin-integrin interaction and the metastatic phenotype. *Mem Inst Oswaldo Cruz*. 1991;86(Suppl 3):29–35. doi: 10.1590/S0074-02761991000700006
- [71] Kawano T, Takasaki S, Tao TW, Kobata A. Altered glycosylation of beta 1 integrins associated with reduced adhesiveness to fibronectin and laminin. *Int J Cancer*. 1993;53(1):91–96. doi:10.1002/ijc.2910530118
- [72] Ogura T, Noguchi T, Murai-Takebe R, Hosooka T, Honma N, Kasuga M. Resistance of B16 melanoma cells to CD47-induced negative regulation of motility as a result of aberrant N-glycosylation of SHPS-1. *J Biol Chem*. 2004;279(14):13711–13720. doi: 10.1074/jbc.M310276200
- [73] Reddy BV, Kalraiya RD. Sialylated beta1,6 branched N-oligosaccharides modulate adhesion, chemotaxis and motility of melanoma cells: effect on invasion and spontaneous metastasis properties. *Biochim Biophys Acta*. 2006;1760(9):1393–1402. doi: 10.1016/j.bbagen.2006.05.003
- [74] Pocheć E, Janik M, Hoja-Łukowicz D, Link-Lenczowski P, Przybyło M, Lityńska A. Expression of integrins $\alpha 3\beta 1$ and $\alpha 5\beta 1$ and GlcNAc $\beta 1,6$ glycan branching influences metastatic melanoma cell migration on fibronectin. *Eur J Cell Biol*. 2013;92(12):355–362. doi:10.1016/j.ejcb.2013.10.007
- [75] Kremser ME, Przybyło M, Hoja-Łukowicz D, Pocheć E, Amoresano A, Carpentieri A, Bubka M, Lityńska A. Characterisation of alpha3beta1 and alpha(v)beta3 integrin N-oligosaccharides in metastatic melanoma WM9 and WM239 cell lines. *Biochim Biophys Acta*. 2008;1780(12):1421–1431. doi:10.1016/j.bbagen.2008.07.011
- [76] Janik ME, Przybyło M, Pocheć E, Pokrywka M, Lityńska A. Effect of $\alpha 3\beta 1$ and $\alpha v\beta 3$ integrin glycosylation on interaction of melanoma cells with vitronectin. *Acta Biochim Pol*. 2010;57(1):55–61.
- [77] Dennis JW, Laferté S, Waghorne C, Breitman ML, Kerbel RS. Beta 1-6 branching of Asn-linked oligosaccharides is directly associated with metastasis. *Science*. 1987;236(4801): 582–585. doi:10.1126/science.2953071
- [78] Taniguchi N, Miyoshi E, Ko JH, Ikeda Y, Ihara Y. Implication of N-acetylglucosaminyltransferases III and V in cancer: gene regulation and signaling mechanism. *Biochim Biophys Acta*. 1999;1455(2–3):287–300. doi:10.1016/S0925-4439(99)00066-6
- [79] Couldrey C, Green JE. Metastases: the glycan connection. *Breast Cancer Res*. 2000;2(5): 321–323. doi:10.1186/bcr75
- [80] Guo HB, Lee I, Kamar M, Akiyama SK, Pierce M. Aberrant N-glycosylation of beta1 integrin causes reduced alpha5beta1 integrin clustering and stimulates cell migration. *Cancer Res*. 2002;62(23):6837–6845.
- [81] Wang L, Liang Y, Li Z, Cai X, Zhang W, Wu G, Jin J, Fang Z, Yang Y, Zha X. Increase in beta1-6 GlcNAc branching caused by N-acetylglucosaminyltransferase V directs

- integrin beta1 stability in human hepatocellular carcinoma cell line SMMC-7721. *J Cell Biochem.* 2007;100(1):230–241. doi:10.1002/jcb.21071
- [82] Yagel S, Feinmesser R, Waghorne C, Lala PK, Breitman ML, Dennis JW. Evidence that beta 1-6 branched Asn-linked oligosaccharides on metastatic tumor cells facilitate invasion of basement membranes. *Int J Cancer.* 1989;44(4):685–690. doi:10.1002/ijc.2910440422
- [83] Dennis JW, Granovsky M, Warren CE. Glycoprotein glycosylation and cancer progression. *Biochim Biophys Acta.* 1999;1473(1):21–34. doi:10.1016/S0304-4165(99)00167-1
- [84] Ranjan A, Kalraiya RD. Invasive potential of melanoma cells correlates with the expression of MT1-MMP and regulated by modulating its association with motility receptors via N-glycosylation on the receptors. *Biomed Res Int.* 2014;2014:804680. doi:10.1155/2014/804680
- [85] Ranjan A, Bane SM, Kalraiya RD. Glycosylation of the laminin receptor ($\alpha 3\beta 1$) regulates its association with tetraspanin CD151: impact on cell spreading, motility, degradation and invasion of basement membrane by tumor cells. *Exp Cell Res.* 2014;322(2):249–264. doi:10.1016/j.yexcr.2014.02.004
- [86] Baldwin G, Novitskaya V, Sadej R, Pochec E, Litynska A, Hartmann C, Williams J, Ashman L, Eble JA, Berditchewski F. Tetraspanin CD151 regulates glycosylation of (α)3(β)1 integrin. *J Biol Chem.* 2008;283(51):35445–35454. doi:10.1074/jbc.M806394200
- [87] Pocheć E, Ząbczyńska M, Bubka M, Homa J, Lityńska A. β 1,6-branched complex-type N-glycans affect FAK signaling in metastatic melanoma cells. *Cancer Invest.* 2016;34(1):45–56. doi:10.3109/07357907.2015.1102928
- [88] Tsuji T, Kawada Y, Kai-Murozono M, Komatsu S, Han SA, Takeuchi K, Mizushima H, Miyazaki K, Irimura T. Regulation of melanoma cell migration and invasion by laminin-5 and α 3 β 1 integrin (VLA-3). *Clin Exp Metastasis.* 2002;19(2):127–134. doi:10.1023/A:1014573204062
- [89] Giannelli G, Astigiano S, Antonaci S, Morini M, Barbieri O, Noonan DM, Albini A. Role of the α 3 β 1 and α 6 β 4 integrins in tumor invasion. *Clin Exp Metastasis.* 2002;19(3):217–223. doi:10.1023/A:1015579204607
- [90] Litynska A, Przybylo M, Pochec E, Kremser E, Hoja-Lukowicz D, Sulowska U. Does glycosylation of melanoma cells influence their interactions with fibronectin? *Biochimie.* 2006;88(5):527–534. doi:10.1016/j.biochi.2005.10.012
- [91] Melchiori A, Mortarini R, Carlone S, Marchisio PC, Anichini A, Noonan DM, Albini A. The α 3 β 1 integrin is involved in melanoma cell migration and invasion. *Exp Cell Res.* 1995;219(1):233–242. doi:10.1006/excr.1995.1223
- [92] Kreidberg JA. Functions of α 3 β 1 integrin. *Curr Opin Cell Biol.* 2000;12(5):548–553. doi:10.1016/S0955-0674(00)00130-7

- [93] Prokopishyn NL, Puzon-McLaughlin W, Takada Y, Laferté S. Integrin $\alpha 3 \beta 1$ expressed by human colon cancer cells is a major carrier of oncodevelopmental carbohydrate epitopes. *J Cell Biochem.* 1999;72(2):189–209. doi:10.1002/(SICI)1097-4644(19990201)72:2<189::AID-JCB4>3.0.CO;2-N
- [94] Link-Lenczowski P, Butters TD, Litynska A. Glycosylation profile of integrin $\alpha 3 \beta 1$ subunits in human melanoma cells at different stages of progression. In: XXI International Symposium on Glycoconjugates (GLYCO 21); 21–26 August. 2011; Vienna. Austria: Glycoconjugate J; 2011. p. 329.
- [95] Béliveau A, Bérubé M, Rousseau A, Pelletier G, Guérin SL. Expression of integrin $\alpha 5 \beta 1$ and MMPs associated with epithelioid morphology and malignancy of uveal melanoma. *Invest Ophthalmol Vis Sci.* 2000;41(8):2363–2372.
- [96] Schaffner F, Ray AM, Dontenwill M. Integrin $\alpha 5 \beta 1$, the fibronectin receptor, as a pertinent therapeutic target in solid tumors. *Cancers.* 2013;5(1):27–47. doi:10.3390/cancers5010027
- [97] Ruoslahti E. The Walter Herbert Lecture. Control of cell motility and tumour invasion by extracellular matrix interactions. *Br J Cancer.* 1992;66(2):239–242. doi:10.1038/bjc.1992.250
- [98] Aznavoorian S, Stracke ML, Parsons J, McClanahan J, Liotta LA. Integrin $\alpha v \beta 3$ mediates chemotactic and haptotactic motility in human melanoma cells through different signaling pathways. *J Biol Chem.* 1996;271(6):3247–3254. doi:10.1074/jbc.271.6.3247
- [99] Zhao Y, Nakagawa T, Itoh S, Inamori K, Isaji T, Kariya Y, Kondo A, Miyoshi E, Miyazaki K, Kawasaki N, Taniguchi N, Gu J. N-acetylglucosaminyltransferase III antagonizes the effect of N-acetylglucosaminyltransferase V on $\alpha 3 \beta 1$ integrin-mediated cell migration. *J Biol Chem.* 2006;281(43):32122–32130. doi:10.1074/jbc.M607274200
- [100] Friedl P, Wolf K. Tumour-cell invasion and migration: diversity and escape mechanisms. *Nat Rev Cancer.* 2003;3(5):362–374. doi:10.1038/nrm2720
- [101] Przybyło M, Pocheć E, Link-Lenczowski P, Lityńska A. Beta1-6 branching of cell surface glycoproteins may contribute to uveal melanoma progression by up-regulating cell motility. *Mol Vis.* 2008;14:625–636.
- [102] Akiyama SK. Integrins in cell adhesion and signaling. *Hum Cell.* 1996;9(3):181–186.
- [103] Zhao Y, Sato Y, Isaji T, Fukuda T, Matsumoto A, Miyoshi E, Gu J, Taniguchi N. Branched N-glycans regulate the biological functions of integrins and cadherins. *FEBS J.* 2008;275(9):1939–1948. doi:10.1111/j.1742-4658.2008.06346.x
- [104] Gu J, Sato Y, Kariya Y, Isaji T, Taniguchi N, Fukuda T. A mutual regulation between cell-cell adhesion and N-glycosylation: implication of the bisecting GlcNAc for biological functions. *J Proteome Res.* 2009;8(2):431–435. doi:10.1021/pr800674g

- [105] Kugler MC, Wei Y, Chapman HA. Urokinase receptor and integrin interactions. *Curr Pharm Des.* 2003;9(19):1565–1574. doi:10.2174/1381612033454658
- [106] Stahl A, Mueller BM. Binding of urokinase to its receptor promotes migration and invasion of human melanoma cells *in vitro*. *Cancer Res.* 1994;54(11):3066–3071.
- [107] Laurenzana A, Biagioni A, D'Alessio S, Bianchini F, Chillí A, Margheri F, Luciani C, Mazzanti B, Pimpinelli N, Torre E, Danese S, Calorini L, Del Rosso M, Fibbi G. Melanoma cell therapy: endothelial progenitor cells as shuttle of the MMP12 uPAR-degrading enzyme. *Oncotarget.* 2014;5(11):3711–3727. doi:10.18632/oncotarget.1987
- [108] Varki A. Sialic acids as ligands in recognition phenomena. *FASEB J.* 1997;11(4):248–255.
- [109] Traving C, Schauer R. Structure, function and metabolism of sialic acids. *Cell Mol Life Sci.* 1998;54(12):1330–1349. doi:10.1007/s000180050258
- [110] Schultz MJ, Swindall AF, Bellis SL. Regulation of the metastatic cell phenotype by sialylated glycans. *Cancer Metastasis Rev.* 2012;31(3–4):501–518. doi:10.1007/s10555-012-9359-7
- [111] Yogeewaran G, Salk PL. Metastatic potential is positively correlated with cell surface sialylation of cultured murine tumor cell lines. *Science.* 1981;212(4502):1514–1516. doi:10.1126/science.7233237
- [112] Sato T. Lectin-probed western blot analysis. *Methods Mol Biol.* 2014;1200:93–100. doi:10.1007/978-1-4939-1292-6_8
- [113] Ranjan A, Kalraiya RD. α 2,6 sialylation associated with increased beta 1,6-branched N-oligosaccharides influences cellular adhesion and invasion. *J Biosci.* 2013;38(5):867–876. doi:10.1007/s12038-013-9382-z
- [114] Chang WW, Yu CY, Lin TW, Wang PH, Tsai YC. Soyasaponin I decreases the expression of α 2,3-linked sialic acid on the cell surface and suppresses the metastatic potential of B16F10 melanoma cells. *Biochem Biophys Res Commun.* 2006;341(2):614–619. doi:10.1016/j.bbrc.2005.12.216
- [115] Nadanaka S, Sato C, Kitajima K, Katagiri K, Irie S, Yamagata T. Occurrence of oligosialic acids on integrin α 5 subunit and their involvement in cell adhesion to fibronectin. *J Biol Chem.* 2001;276(36):33657–33664. doi:10.1074/jbc.M011100200