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# **Oxidative Stress and Antioxidant Therapy in Chronic Kidney and Cardiovascular Disease**

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David M. Small and Glenda C. Gobe

Additional information is available at the end of the chapter

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## **1. Introduction**

Chronic kidney disease (CKD) and cardiovascular disease (CVD) have major impacts upon the health of populations worldwide, especially in Western societies. The progression of CKD or CVD independently exerts synergistic deleterious effects on the other, for example, patients with CKD are more likely to die of CVD than to develop renal failure. This overlap between CKD and CVD, in part, relates to common etiologies such as diabetes mellitus and hypertension, but important dynamic and bidirectional interactions between the cardiovascular system and kidneys may also explain the occurrence of concurrent organ dysfunction [1]. Cardio-renal syndrome (or reno-cardiac syndrome, the prefix depending on the primary failing organ) is becoming increasingly recognised [2]. Conventional treatment targeted at either syndrome generally reduces the onset or progression of the other [3]. Even though our understanding of various factors and steps involved in the pathogenesis of CKD and CVD and their obvious links has improved, a complete picture of the mechanisms involved is still unclear. Oxidative stress has been identified as one unifying mechanism in the pathogenesis of CKD and CVD [4]. This current chapter gives a brief review of recent literature on the relationship between CKD, CVD and oxidative stress and indicates how, by applying knowledge of the molecular controls of oxidative stress, this information may help improve targeted therapy with antioxidants for these diseases.

## **2. Pathogenesis of chronic kidney and cardiovascular disease – The links**

It is, in fact, very difficult to separate these chronic diseases, because one is a complication of the other in many situations. The development and progression of CKD are closely linked

with hypertension and dyslipidemia, both causes of renal failure. Diabetic nephropathy is arguably the leading cause of renal failure. CKD, hypertension and diabetes mellitus all involve endothelial dysfunction, a change well known in the development of atherosclerosis and CVD that includes coronary artery disease, heart failure, stroke and peripheral arterial disease [5]. Vascular calcification occurs in progressive atherosclerosis and CVD, but it is also an important part of vascular injury in end-stage renal disease (ESRD), where patients need renal replacement therapy to survive. It is paradoxical that approximately 50% of individuals with ESRD die from a cardiovascular cause [6]. Thus, CKD and CVD patients have closely-linked diseases with increasing morbidity and mortality. Prevention and treatment of these diseases are major aims in health systems worldwide.

The initiating causes of CKD are highly variable, with previously-mentioned hypertension and diabetes being two of the key ones [7]. Epidemiological studies reveal other strong risk factors for CKD, such as a previous episode of acute kidney damage, exposure to nephrotoxins, obesity, smoking, and increasing age [8, 9]. However, no matter the cause, the progressive structural changes that occur in the kidney are characteristically unifying [10]. The characteristics of CKD are tubulointerstitial inflammation and fibrosis, tubular atrophy, glomerulosclerosis, renal vasculopathy, and presence of granulation tissue. Alterations in the glomerulus include mesangial cell expansion and contraction of the glomerular tuft, followed by a proliferation of connective tissue which leads to significant damage at this first point of the filtration barrier. Structural changes that occur in the kidney produce a vicious cycle of cause and effect, thereby enhancing kidney damage and giving CKD its progressive nature. Whilst early pathological changes in the kidney can occur without clinical presentations, due to the high adaptability of the kidney [10], once the adaptive threshold is reached, the progression of CKD is rapid and the development of ESRD imminent. Vascular pathology exacerbates development of CKD, and it is perhaps here that the links with CVD are closest. Hypertension induces intimal and medial hypertrophy of the intrarenal arteries, leading to hypertensive nephropathy. This is followed by outer cortical glomerulosclerosis with local tubular atrophy and interstitial fibrosis. Compensatory hypertrophy of the inner-cortical glomeruli results, leading to hyperfiltration injury and global glomerulosclerosis. Note, however, that although glomerulopathy is an important characteristic of CKD, the incidence of tubulointerstitial fibrosis has the best correlation with CKD development [11]. As such, kidney tubular cells and renal fibroblasts may be the founding cell types in the progressive development of CKD.

The main clinical manifestation of CKD is a loss of glomerular filtration rate (GFR), allowing for staging of CKD with progressively decreasing (estimated) GFR. CKD staging was facilitated by the National Kidney Foundation (NKF) Kidney Disease Outcomes Quality Initiative (KDOQI) and the Kidney Disease - Improving Global Outcomes (KDIGO), an outcome that highlighted the condition and facilitated its increased diagnosis [12]. The first two stages have normal, or slightly reduced kidney function but some indication of structural deficit in two samples at least 90 days apart. Stages 3-5 are considered the most concerning, with Stage 3 now being sub-classified into Stages 3a and b because of their diagnostic importance. It is thought that stages 2 and 3 should be targeted with prophylactic therapies, such

as lipid lowering drugs or RAS modifiers [13], to minimize the progression of CKD. Table 1 summarises GFR classification and staging for CKD.

| Stage | GFR*                     | Description  |
|-------|--------------------------|--|
| 1     | 90mL/Min                 | Normal renal function but abnormal urine findings, or structural abnormalities, or a genetic trait indicating kidney disease |
| 2     | 60-89mL/min              | Mildly reduced renal function, and other findings (as for stage 1) indicate kidney disease                                   |
| 3A    | 45-59mL/min              | Moderately reduced kidney function   |
| 3B    | 30-44mL/min              |  |
| 4     | 15-29mL/min              | Severely reduced kidney function   |
| 5     | <15mL/min or on dialysis | Very severe, or end-stage kidney failure (sometimes called established renal failure)  |

\* Measured using the MDRD formula (MDRD= Modification of Diet in Renal Disease). All GFR values are normalized to an average surface area (size) of 1.73m<sup>2</sup>

**Table 1.** Classification and description of the different stages of CKD

Similar to CKD, the initiating causes for CVD are complex. Although exposure to cardiovascular risk factors such as hypertension, dyslipidemia and diabetes mellitus contributes to CVD, obesity, lack of physical exercise, smoking, genetics, and even depression, also play a role [14]. Common themes for causality are oxidative stress and inflammation, be they local or systemic. The prevalence of CVD also has a strong positive correlation with age, with more than 80% of cases of coronary artery disease and 75% of cases of congestive heart failure observed in geriatric patients [14]. Intrinsic cardiac aging, defined as the development of structural and functional alterations during aging, may render the heart more vulnerable to various stressors, and this ultimately favours the development of CVD. In the early stages of CVD, left ventricular hypertrophy and myocardial fibrosis may be seen in many patients [15]. The processes involved in their development, particularly in association with CKD, can be attributed to hypervolaemia, systemic arterial resistance, elevated blood pressure, large vessel compliance, and activation of pathways related to the parathyroid hormone–vitamin D–phosphate axis. Left ventricular hypertrophy and myocardial fibrosis also predispose to an increase in electric excitability and ventricular arrhythmias [16].

Heart failure resulting from CVD may be staged in a system similar to CKD. In its 2001 guidelines, the American College of Cardiology (ACC) and the American Heart Association working groups introduced four stages of heart failure [17]: Stage A with patients at high risk for developing heart failure in the future but no functional or structural heart disorder; Stage B with a structural heart disorder but no symptoms at any stage; Stage C with previous or current symptoms of heart failure in the context of an underlying structural heart problem, but managed with medical treatment; and Stage D with advanced disease requiring hospital-based support, a heart transplant or palliative care. The ACC staging system is

useful in that Stage A may be considered pre-heart failure where intervention with treatment may prevent progression to overt symptoms.

The links between CKD and CVD are so close that it is often difficult to tease out individual causes and mechanisms, given their chronic nature. However, children with CKD present as a particular population without pre-existing symptomatic cardiac disease. This population could also receive significant benefit from preventing and treating CKD and thereby minimising the forthcoming development of CVD which is a major cause of death in children with advanced CKD. Left ventricular hypertrophy and dysfunction, and early markers of atherosclerosis such as increased intimal-medial thickness and stiffness of the carotid artery, and coronary artery calcification, may develop in children with CKD. Early CKD, before needing dialysis, is the optimal time to identify and modify risk factors and intervene in an effort to avert risk of premature cardiac disease and death in these children [18]. These observations have sparked added interest in the mechanisms of the chronic diseases, and in ways to target these mechanisms with additional therapies, such as antioxidants.

### **2.1. Inflammation and chronic kidney and cardiovascular disease**

The circulating nature of many inflammatory mediators such as cytokines, and inflammatory or immune cells, indicates that the immune system can act as a mediator of kidney-heart cross-talk and may be involved in the reciprocal dysfunction that is encountered commonly in the cardio-renal syndromes. Chronic inflammation may follow acute inflammation, but in many chronic diseases like CKD and CVD, it is likely that it begins as a low-grade response with no initial manifestation of an acute reaction. There are many links with visceral obesity and with increased secretion of inflammatory mediators seen in visceral fat [15]. Proinflammatory cytokines are produced by adipocytes, and also cells in the adipose stroma. The links with oxidative stress as an endogenous driver of the chronic diseases become immediately obvious when one admits the close association between oxidative stress and inflammation. The characteristics of dyslipidaemia (elevated serum triglycerides, elevated low-density lipoprotein cholesterol, and/or low high-density lipoprotein cholesterol) are also often seen in obese patients and these are all recognized as risk factors for atherosclerosis. The links between obesity, inflammation, dyslipidaemia, CKD and CVD also occur through yet another syndrome, metabolic syndrome. An improved understanding of the precise molecular mechanisms by which chronic inflammation modifies disease is required before the full implications of its presence, including links with persistent oxidative stress as a cause of chronic disease can be realized.

## **3. Oxidative stress and chronic kidney and cardiovascular disease**

### **3.1. Understanding oxidative stress**

Oxidative stress has been implicated in various pathological systems that are prevalent in both CKD and CVD, most importantly inflammation and fibrosis. Chronic inflammation is induced by biological (eg. infections, autoimmune disease), chemical (eg. drugs, environ-



mental toxins), and physical factors (eg. lack of physical activity) [19]. The inflammatory cells are then a source of free radicals in the forms of reactive oxygen and nitrogen species, although reactive oxygen species (ROS) are considered the most common. The highly reactive ROS are capable of damaging various structures and functional pathways in cells. In consequence, the presence of inflammatory cells is stimulated by cell damage caused by ROS, creating a cycle of chronic damage that is difficult to break. Oxidative stress arises from alterations in the oxidation-reduction balance of cells. Normally, ROS are countered by endogenous natural defences known as antioxidants, and it is the imbalance between ROS and antioxidants which favours greater relative levels of ROS, thereby giving rise to a state of oxidative stress [20-22]. The simple oxidant “imbalance” theory has now grown to incorporate the various crucial pathways and cell metabolism that are also controlled by the interplay between oxidants and antioxidants [23-27]. The rationale for antioxidant therapies lies in restoring imbalances in the redox environment of cells.

The main ROS are superoxide ( $O_2^{\bullet-}$ ), the hydroxyl radical ( $OH^{\bullet}$ ) and hydrogen peroxide ( $H_2O_2$ ). Mitochondria are considered the major source of ROS, however other contributing sites of ROS generation include the endoplasmic reticulum, peroxisomes and lysosomes [28-30]. Estimated levels of ROS within mitochondria are 5-10 fold higher than cytosolic and nuclear compartments in cells [31] due to the presence of the electron transport chain (ETC) within the mitochondrial inner membrane. 1-3% of inspired molecular oxygen ( $O_2$ ) is converted to the most common of the ROS,  $O_2^{\bullet-}$  [32, 33], a powerful precursor of  $H_2O_2$ . Although cellular  $H_2O_2$  is stable in this form, it has the potential to interact with a variety of substrates to cause damage, especially in the presence of the ferrous iron ( $Fe^{2+}$ ), which leads to cleavage and formation of the most reactive and damaging of the ROS, the  $OH^{\bullet}$  [34]. In healthy metabolic cells, the production of the potentially harmful  $H_2O_2$  is countered by the catalyzing actions of mitochondrial or cytosolic catalase (CAT) or thiol peroxidases into water and  $O_2$ . The ETC consists of 5 multi-enzyme complexes responsible for maintaining the mitochondrial membrane potential and ATP generation. Each of these complexes presents a site of ROS generation, however complexes I and III have been identified as primary sites of  $O_2^{\bullet-}$  generation [35-38]. ROS generation from mitochondrial complexes increases with age in mice [39]. In humans, Granata and colleagues [40] have demonstrated that patients with CKD and haemodialysis patients display impaired mitochondrial respiration.

Agreement on the role of oxidative stress in the pathogenesis of chronic disease is, however, not complete. Oxidants are involved in highly conserved basic physiological processes and are effectors of their downstream pathways [41, 42]. The specific mechanisms for “oxidative stress” are difficult to define because of the rapidity of oxidant signalling [31]. For example, protein tyrosine phosphatases are major targets for oxidant signalling since they contain the amino acid residue cysteine that is highly susceptible to oxidative modification [43]. Meng and colleagues [25] demonstrated the oxidation of the SH2 domain of the platelet-derived growth factor (PDGF) receptor, which contains protein tyrosine phosphatases, in response to PDGF binding. This may indicate the induction of free radicals in response to receptor activation by a cognate ligand in a process that is similar to phosphorylation cascades of intracellular signalling.

### 3.2. Endogenous antioxidants – Metabolism or disease modifiers

The production of ROS is usually in balance with the availability and cellular localisation of antioxidant enzymes such as superoxide dismutase (SOD), CAT and glutathione peroxidase (Gpx). *In vivo* studies have found accumulated oxidative damage occurs from decreased levels of these enzymes rather than increased ROS production [44, 45]. However, adequate levels of both are likely to be vital for normal cell function. Mitochondria possess their own pool of antioxidants to counteract their generation of ROS. Mitochondrial manganese-SOD (Mn-SOD) converts  $O_2^{\bullet-}$  to  $H_2O_2$  which is then decomposed to harmless  $H_2O$  and  $O_2$  by CAT and Gpx [46]. Copper/zinc-SOD (Cu/Zn-SOD) has been implicated in stabilizing  $O_2^{\bullet-}$  within other cellular compartments, especially peroxisomes, and must be considered in maintenance of the redox state of the whole cell [47, 48]. Limited antioxidant actions of Cu/Zn-SOD may also occur within the inter-membrane space [49]. There is no evidence to indicate that glutathione synthesis occurs within mitochondria, however the mitochondria have their own distinct pool of glutathione required for the formation of Gpx [50].

Among the various endogenous defences against ROS, glutathione homeostasis is critical for a cellular redox environment. Glutathione-linked enzymatic defences of this family include Gpx, glutathione-S-transferase (GST), glutaredoxins (Grx), thioredoxins (Trx), and peroxiredoxins (Prx) [51]. Many of these proteins are known to interact with each other, forming redox networks that have come under investigation for their contribution to dysfunctional oxidant pathways. Mitochondrial-specific isoforms of these proteins also exist and include Grx2, Grx5, Trx2 and Prx3 [52-54], which may be more critical for cell survival compared to their cystolic counterparts [50]. Mitochondrial dysfunction, resulting in depleted ATP synthesis, has the potential to reduce the redox control of glutathione since the rate of glutathione synthesis is ATP-dependent [55]. Intracellular synthesis of glutathione from amino acid derivatives (glycine, glutamic acid and cysteine) accounts for the majority of cellular glutathione compared with extracellular glutathione uptake [56]. Antioxidant networks in which there is interplay, crosstalk and synergism to efficiently and specifically scavenge ROS, may also exist. If this is the case, these antioxidant networks could be harnessed to develop poly-therapeutic antioxidant supplements to combat oxidant-related pathologies, like CKD and CVD.

### 3.3. Oxidative stress and transcriptional control

The role of oxidative stress in upstream transcriptional gene regulation is becoming increasingly recognised. Not only does this provide insight into the physiological role of oxidative stress, but presents regulatory systems that are possibly prone to deregulation. Furthermore, these sites present targets for pharmacological intervention. Peroxisome proliferator-activated receptors (PPARs) are members of the nuclear hormone receptor superfamily of ligand-dependant transcription factors which have been shown to alter during CKD and CVD [57-59]. They have important roles in the transcriptional regulation of cell differentiation, lipid metabolism, glucose homeostasis, cell cycle progression, and inflammation. There are three PPAR isoforms –  $\alpha$ ,  $\beta/\delta$  and  $\gamma$ . Peroxisome proliferator gamma coactivator (PGC $\alpha$ ), in association with PPAR $\gamma$  activation, leads to a variety of cellular protective responses includ-

ing mitochondrial biogenesis [57]. PPAR $\gamma$  regulation in chronic disease is increasingly recognised, with oxidative stress as the unifying initiating feature. Omega-3 polyunsaturated fatty acids (PUFA) reduce inflammation in kidney tubular epithelial cells by upregulating PPAR $\gamma$  [60]. PPAR $\gamma$  activation by pioglitazone reduced cyclo-oxygenase 2 (COX2) expression in smooth muscle cells from hypertensive rats, and upregulated endogenous antioxidants Mn- and Cu/Zn-SOD [61].

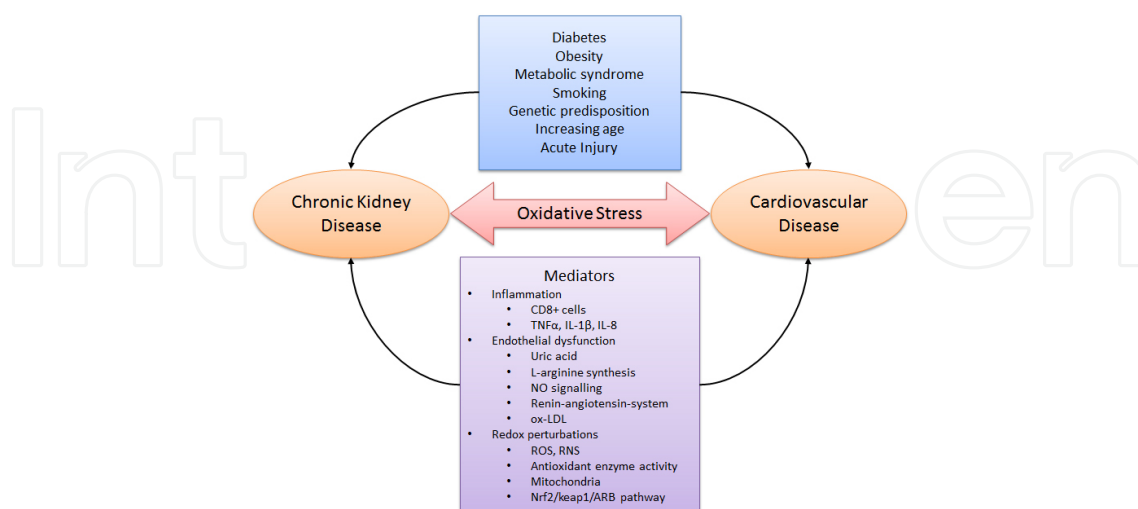
Recently, the protective responses of the nuclear factor E2-related factor 2/Kelch-like ECH-associated protein 1 (Nrf2/Keap1)/antioxidant response element (ARE) were noted [62]. Nrf2 is a nuclear transcription factor that is suppressed in the cytoplasm by the physical binding of Keap1 preventing its translocation into the nucleus. Nrf2 is activated by a loss of Keap1 binding by alterations in cellular redox status, such as increased ROS, by-products of oxidative damage, and reduced antioxidant capacity, thereby promoting its transcriptional response at the ARE [63]. The ARE is a vital component of the promoter regions of genes encoding detoxifying, antioxidant, and glutathione-regulatory enzymes such as quinone-reductase, glutathione-peroxidases, glutathione-reductase, thioredoxins and thioredoxin-reductase, peroxiredoxins, gamma-glutamyl cysteine, heme-oxygenase-1 (HO-1), CAT, SOD metallothionein and ferritin [64-67]. Important to note is that by-products of oxidative damage such as 4-hydroxynoneal and J-isoprostanes act as endogenous activators of Nrf2 [68, 69]. Thus, NRF2/Keap1 and the ARE play a crucial role in cellular defence against ROS. Recent pharmacological protocols have allowed the modulation of this pathway to enhance the capabilities of cells to combat oxidative stress and inflammation [70].

### 3.4. CKD and CVD are unified by oxidative stress

Chronic diseases of the kidney possess various commonalities to chronic disease of the cardiovascular system which can be linked through pathways controlled by oxidative stress, as shown in Figure 1. Vascular, cellular and biochemical factors all contribute. Increased serum uric acid levels (hyperuricaemia) can arise from increased purine metabolism, increasing age and decreased renal excretion, and have harmful systemic effects. Hyperuricaemia is associated with an increased risk for development and progression of CKD. Hyperuricemia is also a risk factor associated with coronary artery disease [71], left ventricular hypertrophy [72], atrial fibrillation [73], myocardial infarction [74] and ischemic stroke [75]. A 20.6% prevalence of hyperuricemia was found in a cross-sectional study of 18,020 CKD patients [76], and a positive correlation was found between serum uric acid and serum creatinine with impaired renal function [77]. Retention of uremic toxins promotes inflammation and oxidative stress, by priming the acute inflammatory polymorphonuclear lymphocytes, activating interleukin (IL)-1 $\beta$  and IL-8 [78] and stimulating the innate immune response through CD8+ cells [79]. Additionally, uric acid synthesis can promote oxidative stress directly through the activity of xanthine oxidoreductase. This enzyme is synthesized as xanthine dehydrogenase, which can be converted to xanthine oxidase by calcium-dependant proteolysis [80] or modification of cysteine residues [81]. In doing so, the enzyme loses its capacity to bind NADH by alterations in its catalytic site and, instead, transfers electrons from O $_2$ , thereby generating O $_2^-$  [82]. However, the role of uric acid in many conditions asso-



ciated with oxidative stress is not clear and there are experimental and clinical data showing that uric acid also has a role *in vivo* as an anti-oxidant [83].



**Figure 1.** Chronic kidney disease and cardiovascular disease are unified by oxidative stress. Mutual risk factors influence the development and progression of CKD and CVD and can either be modifiable (diabetes, obesity, metabolic syndrome, smoking) or non-modifiable (genetic predisposition, increasing age, acute injury). Oxidative stress has been implicated in the majority of initiating factors. The progression of CKD to CVD, or vice versa, is mediated through: (1) inflammation and the release of pro-inflammatory cytokines such as tumor necrosis factor- $\alpha$  (TNF $\alpha$ ), interleukin-1 $\beta$  (IL-1 $\beta$ ) and IL-8 from activated lymphocytes; (2) endothelial dysfunction due to increased retention of uremic toxins, and decreased L-arginine synthesis which causes alterations in nitric oxide (NO) signalling - dyslipidaemia and associated pro-oxidative/inflammatory state lead to increased oxidised-low density lipoproteins (ox-LDL), a major component in the pathogenesis of atherosclerosis; (3) redox perturbations that ultimately underlie oxidative stress due to an imbalance between the production of reactive oxygen species (ROS)/reactive nitrogen species (RNS) and endogenous antioxidants, leading to mitochondrial dysfunction and alterations in redox sensitive pathways such as Nrf2/keap1/ARB.

The kidney is a vital source of L-arginine which is a precursor for nitric oxide (NO). A reduction in renal mass can therefore reduce the production of L-arginine and NO activity. NO is vital for regular vascular endothelial cell function, and decreased amounts have the potential to manifest into CVD [84]. Additionally, oxidized low density lipoprotein (ox-LDL), a by-product of oxidative damage in human blood, plays a pivotal role in the pathogenesis of atherosclerosis [85]. There is also a possible link between CVD and CKD that is regulated by oxidative stress through a functional mitochondrial angiotensin system [86]. Angiotensin type II receptors were co-localised with angiotensin on the inner mitochondrial membrane of human mononuclear cells and mouse renal tubular cells. This system was found to modulate mitochondrial NO production and respiration.

#### 4. Antioxidant therapies in chronic kidney and cardiovascular disease

The current state of antioxidant therapies for CKD and CVD is one of promise, but not without controversy. *In vitro* studies commonly identify agents that are able to detoxify harmful

oxidants. However, these studies are criticised for their isolated, non-holistic, nature [87, 88]. It is largely the positive pre-clinical results from *in vivo* studies, usually in rodents, which drive progress for applicability in chronic human disease, but even these show considerable discrepancies in translation into patients. Despite the well-documented dysregulated endogenous oxidant/antioxidant profile in chronic degenerative disorders such as CVD and CKD, there is still evidence that certain antioxidants have no effect [89-92]. It may first be important to identify patients having an altered oxidative stress profile, since this population provides an ideal “intention to treat” cohort. The following trials of antioxidants need then to be rigorous, identifying not only any positive patient outcomes, but also the underlying mechanism, and of course any deleterious outcome. Various approaches have been taken to reduce oxidative stress in models of CKD and accelerated CVD, ranging from reducing oxidant intake in food stuffs [93, 94] to targeted polypharmaceutical compounds. The benefit of rigorous review of outcome from antioxidant therapies in either CKD or CVD is that the primary and secondary outcomes related to both can be measured. In the following section, some antioxidants used for CKD or CVD are reviewed, as shown in Figure 2.

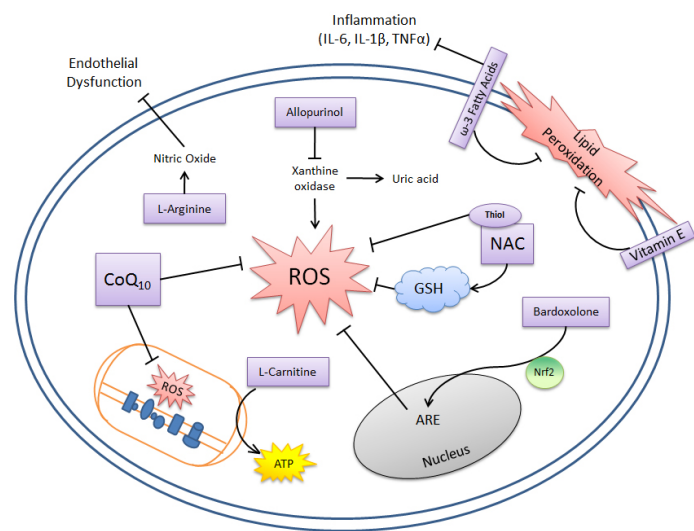
#### 4.1. N-acetylcysteine – An antioxidant with promise

N-acetyl cysteine (NAC) acts as an essential precursor to many endogenous antioxidants involved in the decomposition of peroxides [95]. NAC attenuates oxidative stress from various underlying causes by replenishing intracellular glutathione stores. Glutathione is synthesized in the body by three amino acids by the catalysing of intracellular enzymes gamma-glutamylcysteine synthetase and glutathione synthetase. L-glutamic acid and glycine are two precursors of glutathione that are biologically and readily available. However, the limiting precursor to glutathione biosynthesis and the third amino acid, L-cysteine, is not readily available in a human diet. Although the primary basis for NAC supplementation is to replenish cellular cysteine levels to maintain intracellular glutathione and thus redox control, the sulfhydryl-thiol group of L-cysteine is also able to exert direct antioxidant effects by scavenging free radicals, and NAC may also exert its protective effects against 2,3,5-tris(glutathion-S-yl)-hydroquinone toxicity. This was demonstrated in isolated renal tubular epithelial cells, in part by the activation of extracellular signal regulated protein kinase (ERK) 1/2 [96].

The results of NAC supplementation in kidney disease have been variable and largely dependent on the type and cause of kidney injury and also the timing of treatment. In cultured human proximal tubular epithelial cells, NAC reduced lipid peroxidation and maintained the mitochondrial membrane potential, thereby preventing apoptosis following H<sub>2</sub>O<sub>2</sub> administration [97]. Although NAC had no significant effect on markers of oxidative stress and inflammation in rats following unilateral ureteral obstruction [98], it reduced kidney malondialdehyde (MDA) levels in a diabetic mouse model [99]. The treatment of CKD patients with NAC with the aim of improving renal function and preventing ESKD has been largely disappointing, with no evidence of reduction in proteinuria [100, 101]. However, NAC seems to exert the greatest antioxidant and anti-inflammatory properties when used against the greatest injury, such as in ESKD patients receiving either haemodialysis or peri-

toneal dialysis. In those cases, NAC reduced serum 8-isoprostane and the inflammatory cytokine IL-6 [102, 103]. A recent systemic review on antioxidant therapy in hemodialysis patients highlighted NAC as the most efficacious agent in decreasing oxidative stress [104].

The effect of NAC on cardiovascular pathologies is less well investigated than CKD. Crespo *et al.*, (2011) demonstrated *in vivo* that, although long-term NAC supplementation improved cardiac function, it did not delay progression to cardiomyopathy [105]. Endothelial dysfunction caused by uremic toxins such as indoxyl sulphate induced ROS-dependent expression of the pro-inflammatory and pro-oxidant nuclear factor- $\kappa$ B (NF- $\kappa$ B), which was ameliorated by NAC pre-treatment [106].



**Figure 2.** Cellular sites for antioxidant therapy targets in CKD and CVD. Inflammation, lipid peroxidation and reactive oxygen species (ROS) from mitochondrial, cytoplasmic and extracellular sources contribute to oxidative stress. Vitamin E incorporates into the phospholipid bilayer halting lipid peroxidation chain reactions. Omega ( $\omega$ )-3 fatty acids displace arachadonic acid in the cell membrane and thus reduce arachadonic acid-derived ROS, but also significantly reduce inflammation and subsequent fibrosis. The cysteine residue of N-acetyl-cysteine (NAC) is a precursor for glutathione (GSH) synthesis, and the thiol group is able to scavenge ROS directly. Bardoxolone exerts transcriptional control by promoting nuclear translocation of Nrf2, facilitating antioxidant response element (ARE) binding that upregulates endogenous antioxidant enzyme activity. Allopurinol inhibits xanthine oxidase-derived ROS and the damaging effects of hyperuricemia. Coenzyme Q<sub>10</sub> (CoQ<sub>10</sub>) enhances the efficacy of electron transport in the mitochondria, thereby reducing mitochondrial-derived ROS – it is also able to directly scavenge ROS. L-carnitine enhances mitochondrial fatty acid synthesis and subsequent ATP production and thereby maintains cell health. L-arginine is a precursor for nitric oxide which restores endothelial function.

#### 4.2. Vitamin E – An established antioxidant with controversial outcomes

Vitamin E, or  $\alpha$ -tocopherol, is a lipid-soluble antioxidant that incorporates into the plasma membrane of cells, thereby scavenging free radicals, mainly the peroxy radical, and halting lipid peroxidation chain reactions [107]. A benefit of  $\alpha$ -tocopherol is its ability to restore its antioxidant capacity from its oxidized form following free radical scavenging, and incorporate back into the plasma membrane. Vitamin C (ascorbic acid) is able to directly reduce  $\alpha$ -tocopherol [108-110], and intracellular glutathione and lipoic acid can restore  $\alpha$ -tocopherol

indirectly by restoring vitamin C [111]. This is a prime example of a cellular antioxidant network prone to dysregulation. Administration of  $\alpha$ -tocopherol to kidney proximal tubular cells in culture decreased cisplatin-induced ROS and increased cell viability [112]. The beneficial effects of  $\alpha$ -tocopherol are not limited to its antioxidant properties, and recently attention has focused on its blood oxygenising and endogenous cell signalling functions [113]. Vitamin E foodstuffs primarily consist of  $\alpha$ -tocotrienol, an isoform of  $\alpha$ -tocopherol which has higher antioxidant efficacy in biological membranes. Despite this, the uptake and distribution of  $\alpha$ -tocotrienol is far less than  $\alpha$ -tocopherol. Therefore, the basis of vitamin E supplementation is to enhance  $\alpha$ -tocopherol levels in cell plasma membranes to prevent lipid peroxidation and resultant oxidative stress. One drawback of  $\alpha$ -tocopherol is that it takes several days of pre-treatment to exhibit antioxidant effects [114].

Vitamin E therapy has been extensively researched for renal and cardiovascular benefits in human disease populations. Nevertheless, confounding reports mean there is a lack of consensus as to whether vitamin E therapy induces an overall benefit. It is known that patients with CKD stage 4 display the largest decrease in serum  $\alpha$ -tocopherol levels following a progressive decline from stage 1 indicating an increased need for  $\alpha$ -tocopherol in the CKD population [115]. Interestingly, within the same cohort of patients, a positive correlation of serum  $\alpha$ -tocopherol levels and GFR was found [115]. A large scale trial concluded that vitamin E supplementation to cardiovascular high-risk patients over 4.5 years induced no benefit to cardiovascular outcome [92]. The results from the Selenium and Vitamin E Cancer Prevention Trial (SELECT) are of greater concern. They suggest that vitamin E supplementation significantly increases the risk of prostate cancer for young healthy men [116]. Most studies finding beneficial outcomes of  $\alpha$ -tocopherol supplementation have largely focused on the ESKD dialysis populations compared to healthy controls and found a reduced risk of CVD, decreased oxidative stress and increased erythrocyte antioxidants SOD, Gpx and CAT [117-119]. The use of  $\alpha$ -tocopherol in CKD patients is not without controversy. Miller and colleagues (2005) concluded that high-dose ( $\geq 400$  IU/day) vitamin E supplementation may increase all cause mortality which may be due to  $\alpha$ -tocopherol displacing gamma-( $\gamma$ )-tocopherol and delta-( $\delta$ )-tocopherol in the body [120]. However, this study was highly criticized owing to a bias in data analysis and numerous methodological flaws [121-130]. The apparent lack of clarity surrounding vitamin E supplementation and associated renal and cardiovascular outcomes appears to stem largely from differences in trial design and failure to specify the form of tocopherol used.

#### 4.3. Coenzyme Q<sub>10</sub> - Maintaining mitochondrial health

The heart and kidneys contain the highest endogenous levels of co-enzymes (Co)Q<sub>9</sub> and CoQ<sub>10</sub> compared to all other organs [131, 132]. This is likely due to the respective reliance on aerobic metabolism and high density of mitochondria in the intrinsic functioning cells from these organs. It is imperative that endogenous CoQ<sub>10</sub> levels are maintained to ensure mitochondrial health, and this forms the rationale for CoQ<sub>10</sub> therapy. CoQ<sub>10</sub> is a fundamental lipid-soluble component of all cell membranes including those enclosing subcellular compartments. The physiological roles of CoQ<sub>10</sub> act mostly within the mitochondria where it



has three well-characterised functions: (1) the transfer of electrons from complexes I and II to complex III along the ETC of the inner mitochondrial membrane and subsequent membrane polarisation and ATP generation [133, 134]; (2) the pro-oxidant generation of  $O_2^{\bullet-}$  and  $H_2O_2$  [135, 136]; and (3) the anti-oxidant quenching of free radicals [137]. The continual oxidation-reduction cycle, and existence of CoQ<sub>10</sub> in three different redox states, explains its actions as an important cellular redox modulator through its pro-oxidant and antioxidant actions. The fully oxidised form of CoQ<sub>10</sub>, or ubiquinone, is able to accept electrons, primarily from NADH, to become fully reduced (ubiquinol - CoQ<sub>10</sub>-H<sub>2</sub>). The reduced form of CoQ<sub>10</sub> is able to give up electrons, thereby scavenging free radicals. The intermediate of ubiquinone and ubiquinol is the univalently-reduced ubisemiquinone (CoQ<sub>10</sub>-H<sup>•</sup>) which acts as a pro-oxidant to form  $O_2^{\bullet-}$  and, subsequently,  $H_2O_2$ .

The major antioxidant role of CoQ<sub>10</sub> is in preventing lipid peroxidation directly, and by interactions with  $\alpha$ -tocopherol [138]. Ubiquinol is able to donate a hydrogen atom and thus quench peroxy radicals, preventing lipid peroxidation chain reactions. CoQ<sub>10</sub> and  $\alpha$ -tocopherol co-operate as antioxidants through the actions of CoQ<sub>10</sub>-H<sub>2</sub> restoring  $\alpha$ -tocopheroxyl back to  $\alpha$ -tocopherol [109, 139]. However, the reactivity of  $\alpha$ -tocopherol with peroxy radicals far exceeds that of ubiquinol with peroxy radicals, suggesting that, *in vivo*, ubiquinols do not act as antioxidants but regenerate the antioxidant properties of  $\alpha$ -tocopherols [140]. This is in accordance with *in vivo* studies investigating the effects of CoQ<sub>10</sub> supplementation which have primarily found a limited antioxidant capacity. CoQ<sub>10</sub> acting as a pro-oxidant in all biological membranes including the Golgi, endosome/lysosome systems, as well as mitochondria, has led to much criticism regarding the claimed antioxidant power of CoQ<sub>10</sub> supplementation in humans [141]. Nonetheless, many *in vitro* studies demonstrate antioxidant properties of CoQ<sub>10</sub> in single cells, and benefits of CoQ<sub>10</sub> supplementation in humans are attributed to its ability to maintain efficient mitochondrial energy metabolism and thus prevent mitochondrial dysfunction, rather than act as a direct cellular antioxidant. CoQ<sub>10</sub> supplementation *in vivo* reduced protein oxidation in skeletal muscle of rats but had no effect on mitochondrial  $H_2O_2$  production in the kidney [142]. However, Ishikawa and colleagues (2011) demonstrated a decrease in kidney  $O_2^{\bullet-}$  levels in hemi-nephrectomised rats on a CoQ<sub>10</sub> supplemented diet, and increased renal function compared with rats on a control diet [143]. Recently, CoQ<sub>10</sub> supplementation improved left ventricular diastolic dysfunction and remodelling and reduced oxidative stress in a mouse model of type 2 diabetes [144]. CoQ<sub>10</sub> supplementation in CVD patients also receiving statin therapy is becoming increasingly popular due to the CoQ<sub>10</sub>-inhibitory actions of statins. CoQ<sub>10</sub> levels decrease with age, but there are no studies measuring endogenous CoQ<sub>10</sub> levels in CKD or CVD patients and this could prove vital in the identification of population where CoQ<sub>10</sub> therapy may have beneficial outcomes.

#### 4.4. Omega-3 poly-unsaturated fatty acids – Inflammation and oxidative stress

Inflammation and fibrosis are causes, as well as consequences, of oxidative stress [145, 146]. Direct targeting of inflammatory and fibrotic pathways with more specific modifying compounds presents a way to indirectly decrease oxidative stress in chronic pathologies. Long



chain omega-3 PUFA, including docosahexanoic acid (DHA) and eicosapentanoic acid (EPA), have been investigated in a large range of *in vitro* and *in vivo* models and found to possess anti-inflammatory properties. Recently, omega-3 fatty acid treatment of peripheral blood mononuclear cells from pre-dialysis CKD patients reduced the inflammatory markers IL-6, IL-1 $\beta$ , tumor necrosis factor (TNF)- $\alpha$  and C-reactive protein to levels observed in healthy subjects [147]. Although the beneficial effects of EPA/DHA are attributed to their anti-inflammatory properties, they are also known to enhance endogenous antioxidant defence systems such as  $\gamma$ -glutamyl-cysteinyl ligase and glutathione reductase [148]. DHA and EPA incorporate into the phospholipid bilayer of cells where they displace arachidonic acid. Arachidonic acid can generate ROS through the COX2 and xanthine oxidase inflammatory pathways. DHA/EPA administration to renal epithelial cells and macrophages suppresses this pro-oxidant pathway [149]. Furthermore, chemoattractants derived from EPA are less potent than those derived from arachidonic acid [150, 151]. Recently, *in vitro* studies determined that EPA and DHA attenuated TNF- $\alpha$ -stimulated monocyte chemoattractant protein (MCP)-1 gene expression by interacting with ERK and NF- $\kappa$ B in rat mesangial cells [152]. Earlier evidence had shown that EPA and DHA inhibit NF- $\kappa$ B expression by stimulating PPARs in human kidney-2 cells *in vitro* [60]. *In vivo* studies have now confirmed an improvement in kidney function and structure using EPA/DHA supplementation, with reduced oxidative stress, inflammation and tubulointerstitial fibrosis through the reversal of inflammatory and oxidant pathways [153, 154]. Recently, a highly beneficial outcome of fish oil supplementation was found with heart failure patients with co-morbid diabetes [155]. Clinical studies have found fish oil treatment modulates lipid levels [156, 157], and has anti-thrombotic [158, 159] and anti-hypertensive effects due to its vascular and endothelial actions [160].

#### 4.5. Allopurinol – A xanthine oxidase inhibitor

Allopurinol treatment aims to inhibit xanthine oxidase to decrease serum uric acid and its associated toxic effects. Allopurinol and its metabolite, oxypurinol, act as competitive substrates for xanthine oxidase. They enhance urinary urate excretion and block uric acid reabsorption by urate transporters in the proximal tubule, thereby facilitating enhanced uric acid excretion [161-163]. Allopurinol treatment of diabetic mice attenuated hyperuricaemia, albuminuria, and tubulointerstitial injury [164]. Allopurinol may also have antioxidant activities in addition to its enzyme inhibitory activities, by scavenging OH $\cdot$  as well as chlorine dioxide and HOCl [165, 166]. Although later *in vivo* studies revealed that rat serum obtained after oral administration of allopurinol did not contain allopurinol levels sufficient to scavenge free radicals [167], inhibition of xanthine oxidase-dependent production of NO $\cdot$  and ROS provides allopurinol an indirect mechanism for decreasing oxidative stress in hyperuricaemic CKD patients. Interventional studies of use of allopurinol in renal disease have shown improved uric acid levels, GFR, cardiovascular outcomes and delayed CKD progression. A prospective randomised trial of 113 patients with GFR <60 ml/min/1.73m $^3$  given allopurinol 100mg/d for 2 years found an increase in GFR of 1.31 ml/min/1.73m $^3$  compared to the controls which decreased, and a 71% decreased risk of CVD [168]. Interestingly, Kanbay and colleagues (2007) found that allopurinol at 300mg/d over 3 months improved GFR, uric acid

and C-reactive protein levels but made no change to proteinuria [169]. Allopurinol given to ESKD patients on hemodialysis reduced the risk of CVD by decreasing serum low density lipoproteins, triglycerides and uric acid [170]. Large, long-term interventional studies investigating kidney function in the CKD, and CVD, populations are needed to fully determine if allopurinol is cardio- and reno-protective via anti-oxidant mechanisms.

#### **4.6. Bardoxolone methyl - Targeting the Nrf/Keap1/ARE pathway**

A different approach has been investigated by modulating pathways that respond to oxidative stress, rather than targeting ROS by directly increasing endogenous antioxidants. The Nrf2/keap1/ARE pathway presents an exciting target to enhance the oxidant detoxifying capabilities of cells. Bardoxolone methyl [2-cyano-3,12-dioxooleana-1,9(11)-dien-28-oic acid (CDDO-Me)] is a potent activator of the Nrf2/keap1/ARE pathway and currently shows promise for halting the progressive decline of GFR in type 2 diabetic CKD patients [171, 172]. Bardoxolone methyl is a triterperoid derived from natural plant products that has undergone oleanolic acid-based modification [173]. Its mechanism of action is largely unknown, however, it induces an overall antioxidative protective effect with anti-inflammatory and cytoprotective characteristics [174, 175]. Bardoxolone methyl administered to mice ameliorated ischemia-reperfusion induced acute kidney injury by Nrf2-dependant expression of HO-1 and PPAR $\gamma$  [176]. Its mechanism may also reside in regulating mitochondrial biogenesis given the involvement of PPAR $\gamma$ . A large international study evaluating the full scale of bardoxolone methyl's effects on CKD progression is in progress, the results of which could determine if bardoxolone methyl should become a standard treatment in renal disease patients. Concurrent benefits to CVD will undoubtedly also be measured.

#### **4.7. L-Carnitine – Improving cardiovascular health in dialysis**

Carnitine is an essential cofactor required for the transformation of free fatty acids into acylcarnitine and its subsequent transport into the mitochondria for  $\beta$ -oxidation [177]. This underlies its importance in the production of ATP for cellular energy. Acylcarnitine is also essential for the removal of toxic fat metabolism by-products. Carnitine is obtained primarily from food stuffs, however it can be synthesised endogenously from the amino acid L-lysine and methionine [177]. L-carnitine supplementation primarily benefits ESRD patients on hemodialysis and their associated cardiovascular complications, especially anemia. This is primarily due to the well-described decrease in serum free carnitine in maintenance hemodialysis patients compared to non-dialysis CKD and healthy patients [178]. L-carnitine supplementation offsets renal anemia, lipid abnormalities and cardiac dysfunction in hemodialysis patients [179]. Left ventricular hypertrophy regressed in hemodialysis patients receiving 10mg/kg of L-carnitine immediately following hemodialysis for a 12 month period. [180]. Other measures of cardiac morbidity such as reduced left ventricular ejection fraction and increased left ventricular mass also significantly improved following low dose L-carnitine supplementation [181]. Benefits to the peripheral vasculature have also been demonstrated by L-carnitine through a mechanism thought to involve an associated de-

crease in homocysteine levels [182]. Interestingly, oxidative stress is a major characteristic of hemodialysis patients [183].

As well as the physiological role of L-carnitine in mitochondrial fatty acid synthesis, oxidant reducing capabilities have also been demonstrated and may underlie the health benefits of L-carnitine therapy in CKD and CVD. L-carnitine infusions significantly improved blood urea nitrogen (BUN) and creatinine levels in a 5/6 nephrectomy model of CKD with a concomitant increase in plasma SOD, Gpx, CAT and GSH, and decrease in the oxidative stress marker malondialdehyde [184]. Ye et al., (2010) suggest that L-carnitine attenuates renal tubular cell oxidant injury and subsequent apoptosis by reducing mitochondrial-derived ROS [97]. They suggest that this anti-apoptotic mechanism may also explain the demonstrated reduction in morbidity from cardiomyopathies in L-carnitine supplemented hemodialysis patients.

#### **4.8. L-Arginine - Maintaining endothelial function**

The premise of L-arginine supplementation is to maintain NO signalling and thereby maintain vascular endothelial cell function. L-arginine is a physiological precursor to NO and its availability and transport determine the rate of NO biosynthesis. CKD patients most often present with atherosclerosis, thromboembolic complications, and endothelial dysfunction, primarily due to altered endothelium-dependant relaxation factors [185]. It is believed that the impaired NO synthesis, common in CKD individuals, contributes significantly to their disease pathogenesis [186]. L-arginine synthesis occurs in the liver and kidney, with the kidney functioning to maintain homeostatic plasma levels since the liver processes NO from the diet [187]. The addition of L-aspartic acid or L-glutamic acid with L-citrulline and argininosuccinic acid synthase as the rate determining enzyme forms L-arginine [188]. The proximal tubular cells account for the majority of kidney NO synthesis [189, 190], thus kidney damage and atrophy, a primary corollary of CKD, results in decreased synthesis of L-arginine. The majority of research demonstrates decreased levels of NO production in CKD and CVD patients [191-193]. However, some research suggests NO activity increases [194, 195]. These disparate findings highlight the need to measure L-arginine levels in patients before commencing L-arginine supplementation. Rajapaske *et al.* (2012) demonstrated impaired kidney L-arginine transport and a contributing factor to hypertension in rats, irrespective of an underlying renal disease [196]. During a state of oxidative stress, L-arginine supplementation was shown to decrease MDA, myeloperoxidase and xanthine oxidase and increase glutathione in both heart and kidney tissue from rats [197]. As such, L-arginine supplementation represents an approach to restoring a dysregulation of NO signalling and subsequent endothelial dysfunction in both chronic kidney and heart diseases.

#### **4.9. Combination antioxidants**

Compounds commonly used to alleviate oxidative stress exhibit different antioxidant actions, and so there exists the potential for different antioxidants to work together to improve whole cell and organ function through a targeted polypharmaceutical approach to decrease oxidative stress. However, most clinical studies investigating the effects of combination anti-

oxidants have demonstrated confounding results. Mosca *et al.*, (2002) demonstrated that daily intake of NAC 100mg, L-carnitine 100mg, selenomethionine 0.05mg,  $\alpha$ -tocopherol 10mg, CoQ<sub>10</sub> 100mg and  $\alpha$ -lipoic acid 100mg successfully increased plasma CAT, Gpx and total antioxidant capacity whilst decreasing lipid peroxides and ROS generation by lymphocyte mitochondria [198]. However, this trial only included healthy participants and cannot be extrapolated to the CKD and CVD populations.

In a murine model of diabetic nephropathy, a major cause of CKD with associated CVD, the beneficial effects of NAC, L-ascorbic acid (vitamin C) and  $\alpha$ -tocopherol were demonstrated [199]. Daily supplementation for 8 weeks decreased lipid peroxidation, BUN, serum creatinine and blood glucose, mainly due to a reduction in the inflammatory response induced by hyperglycemia. In comparison, a prospective trial investigating oral supplementation of mixed tocopherols and  $\alpha$ -lipoic acid in stage 3 and 4 CKD patients has revealed disappointing results. Over 2 months, supplementation did not reduce biomarkers of oxidative stress (F<sub>2</sub>-isoprostanes and protein thiol concentration) or inflammation (CRP and IL-6). The short period of time (2 months) of the intervention may explain this result and longer trials need to be carried out. The inclusion of vitamin E in these interventions has polarized discussion on the outcomes, because of its negligible benefits when cardiovascular outcomes were measured [91, 92, 200] and also because of contraindications, discussed previously. Despite this, long-term treatment in with the antioxidants vitamin C, vitamin E, CoQ<sub>10</sub> and selenium has been shown to reduce multiple cardiovascular risk factors [201]. Recently, multiple antioxidants in combination with L-arginine have shown promise in animal models of CKD and associated CVD. Korish (2010) has demonstrated in a 5/6 nephrectomy CKD model that L-arginine improved the effects of L-carnitine, catechin and vitamins E and C on blood pressure, dyslipidemia, inflammation and kidney function [84].

## 5. Conclusion

CKD is a progressive disease with increasing incidence, having very little success in current conventional therapies once CKD reaches stage 4. Stages 2 and 3 are best to target to slow or stop further development of the disease. There is an almost inseparable connection between CKD and CVD, with many patients with CKD dying of the cardiovascular complications before renal failure reaches its fullest extent. Oxidative stress and inflammation are closely interrelated with development of CKD and CVD, and involve a spiralling cycle that leads to progressive patient deterioration. Given the complex nature of oxidative stress and its molecular pathways, antioxidants may need to be given as a polypharmacotherapy to target each aberrant pathway, with the aim of reducing the burden of these chronic diseases. It is vital for the progression of antioxidant therapy research in CKD and CVD that measures of oxidative stress are compared with pathophysiological outcome in the diseases, especially in connection with antioxidant therapies that may be delivered with or without more conventional CKD therapies.



## Author details

David M. Small and Glenda C. Gobe\*

\*Address all correspondence to: [g.gobe@uq.edu.au](mailto:g.gobe@uq.edu.au)

Centre for Kidney Disease Research, School of Medicine, The University of Queensland, Brisbane, Australia

## References

- [1] Rosner MH, Ronco C, Okusa MD. The role of inflammation in the cardio-renal syndrome: a focus on cytokines and inflammatory mediators. *Semin Nephrol.* 2012 Jan; 32(1):70-8.
- [2] Ronco C, McCullough P, Anker SD, Anand I, Aspromonte N, Bagshaw SM, et al. Cardio-renal syndromes: report from the consensus conference of the acute dialysis quality initiative. *Eur Heart J.* 2010 Mar;31(6):703-11.
- [3] Leung FP, Yung LM, Laher I, Yao X, Chen ZY, Huang Y. Exercise, vascular wall and cardiovascular diseases: an update (Part 1). *Sports Med.* 2008;38(12):1009-24.
- [4] Bongartz LG, Cramer MJ, Doevendans PA, Joles JA, Braam B. The severe cardiorenal syndrome: 'Guyton revisited'. *Eur Heart J.* 2005 Jan;26(1):11-7.
- [5] Sallam N, Fisher A, Golbidi S, Laher I. Weight and inflammation are the major determinants of vascular dysfunction in the aortae of db/db mice. *Naunyn Schmiedeberg's Arch Pharmacol.* 2011 May;383(5):483-92.
- [6] Schiffrin EL, Lipman ML, Mann JF. Chronic kidney disease: effects on the cardiovascular system. *Circulation.* 2007 Jul 3;116(1):85-97.
- [7] McDonald SP, Chang S, Excell L, editors. ANZDATA Registry Report. Adelaide 2007.
- [8] Tanner RM, Brown TM, Muntner P. Epidemiology of obesity, the metabolic syndrome, and chronic kidney disease. *Curr Hypertens Rep.* 2012 Apr;14(2):152-9.
- [9] Graf J, Ryan C, Green F. An overview of chronic kidney disease in Australia, 2009. Canberra: Australian Inst Health Welfare 2009.
- [10] Tesch GH. Review: Serum and urine biomarkers of kidney disease: A pathophysiological perspective. *Nephrol (Carlton).* 2010 Sep;15(6):609-16.
- [11] Rodriguez-Iturbe B, Johnson RJ, Herrera-Acosta J. Tubulointerstitial damage and progression of renal failure. *Kidney Int Suppl.* 2005 Dec(99):S82-6.



- [12] Fassett RG, Venuthurupalli SK, Gobe GC, Coombes JS, Cooper MA, Hoy WE. Biomarkers in chronic kidney disease: a review. *Kidney Int.* 2011 Oct;80(8):806-21.
- [13] Choudhury D, Luna-Salazar C. Preventive health care in chronic kidney disease and end-stage renal disease. *Nat Clin Pract Nephrol.* 2008 Apr;4(4):194-206.
- [14] Dutta D, Calvani R, Bernabei R, Leeuwenburgh C, Marzetti E. Contribution of impaired mitochondrial autophagy to cardiac aging: mechanisms and therapeutic opportunities. *Circ Res.* 2012 Apr 13;110(8):1125-38.
- [15] Manabe I. Chronic inflammation links cardiovascular, metabolic and renal diseases. *Circ J.* 2011;75(12):2739-48.
- [16] Glasscock RJ, Pecoits-Filho R, Barberato SH. Left ventricular mass in chronic kidney disease and ESRD. *Clin J Am Soc Nephrol: CJASN.* 2009 Dec;4 Suppl 1:S79-91.
- [17] Hunt SA, Abraham WT, Chin MH, Feldman AM, Francis GS, Ganiats TG, et al. ACC/AHA 2005 Guideline Update for the Diagnosis and Management of Chronic Heart Failure in the Adult: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Writing Committee to Update the 2001 Guidelines for the Evaluation and Management of Heart Failure): developed in collaboration with the American College of Chest Physicians and the International Society for Heart and Lung Transplantation: endorsed by the Heart Rhythm Society. *Circulation.* 2005 Sep 20;112(12):e154-235.
- [18] Mitsnefes MM. Cardiovascular disease in children with chronic kidney disease. *J Am Soc Nephrol: JASN.* 2012 Apr;23(4):578-85.
- [19] Whaley-Connell A, Pavey BS, Chaudhary K, Saab G, Sowers JR. Renin-angiotensin-aldosterone system intervention in the cardiometabolic syndrome and cardio-renal protection. *Ther Adv Cardiovasc Dis.* 2007 Oct;1(1):27-35.
- [20] Gomes P, Simao S, Silva E, Pinto V, Amaral JS, Afonso J, et al. Aging increases oxidative stress and renal expression of oxidant and antioxidant enzymes that are associated with an increased trend in systolic blood pressure. *Oxid Med Cell Longev.* 2009 Jul-Aug;2(3):138-45.
- [21] Pias EK, Aw TY. Apoptosis in mitotic competent undifferentiated cells is induced by cellular redox imbalance independent of reactive oxygen species production. *FASEB J.* 2002 Jun;16(8):781-90.
- [22] Zhuang S, Yan Y, Daubert RA, Han J, Schnellmann RG. ERK promotes hydrogen peroxide-induced apoptosis through caspase-3 activation and inhibition of Akt in renal epithelial cells. *Am J Physiol Renal Physiol.* 2007 Jan;292(1):F440-7.
- [23] Blanchetot C, Tertoolen LG, den Hertog J. Regulation of receptor protein-tyrosine phosphatase alpha by oxidative stress. *EMBO J.* 2002 Feb 15;21(4):493-503.
- [24] Jones DP. Redefining oxidative stress. *Antioxid Redox Signal.* 2006 Sep-Oct;8(9-10):1865-79.

- [25] Meng TC, Fukada T, Tonks NK. Reversible oxidation and inactivation of protein tyrosine phosphatases in vivo. *Mol Cell*. 2002 Feb;9(2):387-99.
- [26] Rao RK, Clayton LW. Regulation of protein phosphatase 2A by hydrogen peroxide and glutathionylation. *Biochem Biophys Res Commun*. 2002 Apr 26;293(1):610-6.
- [27] Tavakoli S, Asmis R. Reactive Oxygen Species and Thiol Redox Signaling in the Macrophage Biology of Atherosclerosis. *Antioxid Redox Signal*. 2012 Jun 11.
- [28] Madesh M, Hajnoczky G. VDAC-dependent permeabilization of the outer mitochondrial membrane by superoxide induces rapid and massive cytochrome c release. *J Cell Biol*. 2001 Dec 10;155(6):1003-15.
- [29] Soubannier V, McBride HM. Positioning mitochondrial plasticity within cellular signaling cascades. *Biochim Biophys Acta*. 2009 Jan;1793(1):154-70.
- [30] Vay L, Hernandez-SanMiguel E, Lobaton CD, Moreno A, Montero M, Alvarez J. Mitochondrial free  $[Ca^{2+}]$  levels and the permeability transition. *Cell Calcium*. 2009 Mar;45(3):243-50.
- [31] Cadenas E, Davies KJ. Mitochondrial free radical generation, oxidative stress, and aging. *Free Radic Biol Med*. 2000 Aug;29(3-4):222-30.
- [32] Boveris A, Chance B. The mitochondrial generation of hydrogen peroxide. General properties and effect of hyperbaric oxygen. *Biochem J*. 1973 Jul;134(3):707-16.
- [33] Nohl H, Hegner D. Do mitochondria produce oxygen radicals in vivo? *Eur J Biochem*. 1978 Jan 16;82(2):563-7.
- [34] Lipinski B. Is it oxidative stress or free radical stress and why does it matter? *Oxid Antioxid Med Sci*. 2012 March;1(1):5-9.
- [35] Cadenas E, Boveris A, Ragan CI, Stoppani AO. Production of superoxide radicals and hydrogen peroxide by NADH-ubiquinone reductase and ubiquinol-cytochrome c reductase from beef-heart mitochondria. *Arch Biochem Biophys*. 1977 Apr 30;180(2):248-57.
- [36] Turrens JF, Boveris A. Generation of superoxide anion by the NADH dehydrogenase of bovine heart mitochondria. *Biochem J*. 1980 Nov 1;191(2):421-7.
- [37] Turrens JF, Alexandre A, Lehninger AL. Ubisemiquinone is the electron donor for superoxide formation by complex III of heart mitochondria. *Arch Biochem Biophys*. 1985 Mar;237(2):408-14.
- [38] Turrens JF. Mitochondrial formation of reactive oxygen species. *J Physiol*. 2003 Oct 15;552(Pt 2):335-44.
- [39] Choksi KB, Nuss JE, Boylston WH, Rabek JP, Papaconstantinou J. Age-related increases in oxidatively damaged proteins of mouse kidney mitochondrial electron transport chain complexes. *Free Radic Biol Med*. 2007 Nov 15;43(10):1423-38.

- [40] Granata S, Zaza G, Simone S, Villani G, Latorre D, Pontrelli P, et al. Mitochondrial dysregulation and oxidative stress in patients with chronic kidney disease. *BMC Genomics*. 2009;10:388.
- [41] Nemoto S, Takeda K, Yu ZX, Ferrans VJ, Finkel T. Role for mitochondrial oxidants as regulators of cellular metabolism. *Mol Cell Biol*. 2000 Oct;20(19):7311-8.
- [42] Werner E, Werb Z. Integrins engage mitochondrial function for signal transduction by a mechanism dependent on Rho GTPases. *J Cell Biol*. 2002 Jul 22;158(2):357-68.
- [43] Cooper CE, Patel RP, Brookes PS, Darley-Usmar VM. Nanotransducers in cellular redox signaling: modification of thiols by reactive oxygen and nitrogen species. *Trends Biochem Sci*. 2002 Oct;27(10):489-92.
- [44] Kokoszka JE, Coskun P, Esposito LA, Wallace DC. Increased mitochondrial oxidative stress in the Sod2 (+/-) mouse results in the age-related decline of mitochondrial function culminating in increased apoptosis. *Proc Natl Acad Sci U S A*. 2001 Feb 27;98(5):2278-83.
- [45] Meng Q, Wong YT, Chen J, Ruan R. Age-related changes in mitochondrial function and antioxidative enzyme activity in fischer 344 rats. *Mech Ageing Dev*. 2007 Mar;128(3):286-92.
- [46] Raha S, McEachern GE, Myint AT, Robinson BH. Superoxides from mitochondrial complex III: the role of manganese superoxide dismutase. *Free Radic Biol Med*. 2000 Jul 15;29(2):170-80.
- [47] Angermuller S, Islinger M, Volkl A. Peroxisomes and reactive oxygen species, a lasting challenge. *Histochem Cell Biol*. 2009 Apr;131(4):459-63.
- [48] Islinger M, Li KW, Seitz J, Volkl A, Luers GH. Hitchhiking of Cu/Zn superoxide dismutase to peroxisomes-evidence for a natural piggyback import mechanism in mammals. *Traffic*. 2009 Nov;10(11):1711-21.
- [49] Sturtz LA, Diekert K, Jensen LT, Lill R, Culotta VC. A fraction of yeast Cu,Zn-superoxide dismutase and its metallochaperone, CCS, localize to the intermembrane space of mitochondria. A physiological role for SOD1 in guarding against mitochondrial oxidative damage. *J Biol Chem*. 2001 Oct 12;276(41):38084-9.
- [50] Soderdahl T, Enoksson M, Lundberg M, Holmgren A, Ottersen OP, Orrenius S, et al. Visualization of the compartmentalization of glutathione and protein-glutathione mixed disulfides in cultured cells. *FASEB J*. 2003 Jan;17(1):124-6.
- [51] Godoy JR, Oesteritz S, Hanschmann EM, Ockenga W, Ackermann W, Lillig CH. Segment-specific overexpression of redoxins after renal ischemia and reperfusion: protective roles of glutaredoxin 2, peroxiredoxin 3, and peroxiredoxin 6. *Free Radic Biol Med*. 2011 Jul 15;51(2):552-61.
- [52] Lillig CH, Holmgren A. Thioredoxin and related molecules--from biology to health and disease. *Antioxid Redox Signal*. 2007 Jan;9(1):25-47.

- [53] Lonn ME, Hudemann C, Berndt C, Cherkasov V, Capani F, Holmgren A, et al. Expression pattern of human glutaredoxin 2 isoforms: identification and characterization of two testis/cancer cell-specific isoforms. *Antioxid Redox Signal*. 2008 Mar; 10(3):547-57.
- [54] Hanschmann EM, Lonn ME, Schutte LD, Funke M, Godoy JR, Eitner S, et al. Both thioredoxin 2 and glutaredoxin 2 contribute to the reduction of the mitochondrial 2-Cys peroxiredoxin Prx3. *J Biol Chem*. 2010 Dec 24;285(52):40699-705.
- [55] Lash LH, Putt DA, Matherly LH. Protection of NRK-52E cells, a rat renal proximal tubular cell line, from chemical-induced apoptosis by overexpression of a mitochondrial glutathione transporter. *J Pharmacol Exp Ther*. 2002 Nov;303(2):476-86.
- [56] Visarius TM, Putt DA, Schare JM, Pegouske DM, Lash LH. Pathways of glutathione metabolism and transport in isolated proximal tubular cells from rat kidney. *Biochem Pharmacol*. 1996 Jul 26;52(2):259-72.
- [57] Funk JA, Odejinmi S, Schnellmann RG. SRT1720 induces mitochondrial biogenesis and rescues mitochondrial function after oxidant injury in renal proximal tubule cells. *J Pharmacol Exp Therapeut*. 2010 May;333(2):593-601.
- [58] Lepenies J, Hewison M, Stewart PM, Quinkler M. Renal PPARgamma mRNA expression increases with impairment of renal function in patients with chronic kidney disease. *Nephrology*. 2010 Oct;15(7):683-91.
- [59] Sakamoto A, Hongo M, Saito K, Nagai R, Ishizaka N. Reduction of renal lipid content and proteinuria by a PPAR-gamma agonist in a rat model of angiotensin II-induced hypertension. *Eur J Pharmacol*. 2012 May 5;682(1-3):131-6.
- [60] Li H, Ruan XZ, Powis SH, Fernando R, Mon WY, Wheeler DC, et al. EPA and DHA reduce LPS-induced inflammation responses in HK-2 cells: evidence for a PPAR-gamma-dependent mechanism. *Kidney Int*. 2005 Mar;67(3):867-74.
- [61] Martin A, Perez-Giron JV, Hernanz R, Palacios R, Briones AM, Fortuno A, et al. Peroxisome proliferator-activated receptor-gamma activation reduces cyclooxygenase-2 expression in vascular smooth muscle cells from hypertensive rats by interfering with oxidative stress. *J Hypertens*. 2012 Feb;30(2):315-26.
- [62] Hybertson BM, Gao B, Bose SK, McCord JM. Oxidative stress in health and disease: the therapeutic potential of Nrf2 activation. *Mol Aspects Med*. 2011 Aug;32(4-6):234-46.
- [63] Wilmes A, Crean D, Aydin S, Pfaller W, Jennings P, Leonard MO. Identification and dissection of the Nrf2 mediated oxidative stress pathway in human renal proximal tubule toxicity. *Toxicol In Vitro*. 2011 Apr;25(3):613-22.
- [64] Nelson SK, Bose SK, Grunwald GK, Myhill P, McCord JM. The induction of human superoxide dismutase and catalase in vivo: a fundamentally new approach to antioxidant therapy. *Free Radic Biol Med*. 2006 Jan 15;40(2):341-7.

- [65] Prestera T, Talalay P, Alam J, Ahn YI, Lee PJ, Choi AM. Parallel induction of heme oxygenase-1 and chemoprotective phase 2 enzymes by electrophiles and antioxidants: regulation by upstream antioxidant-responsive elements (ARE). *Mol Med*. 1995 Nov;1(7):827-37.
- [66] Li Y, Jaiswal AK. Regulation of human NAD(P)H:quinone oxidoreductase gene. Role of AP1 binding site contained within human antioxidant response element. *J Biol Chem*. 1992 Jul 25;267(21):15097-104.
- [67] Okuda A, Imagawa M, Maeda Y, Sakai M, Muramatsu M. Structural and functional analysis of an enhancer GPEI having a phorbol 12-O-tetradecanoate 13-acetate responsive element-like sequence found in the rat glutathione transferase P gene. *J Biol Chem*. 1989 Oct 5;264(28):16919-26.
- [68] Itoh K, Mochizuki M, Ishii Y, Ishii T, Shibata T, Kawamoto Y, et al. Transcription factor Nrf2 regulates inflammation by mediating the effect of 15-deoxy-Delta(12,14)-prostaglandin j(2). *Mol Cell Biol*. 2004 Jan;24(1):36-45.
- [69] Levonen AL, Landar A, Ramachandran A, Ceaser EK, Dickinson DA, Zanoni G, et al. Cellular mechanisms of redox cell signalling: role of cysteine modification in controlling antioxidant defences in response to electrophilic lipid oxidation products. *Biochem J*. 2004 Mar 1;378(Pt 2):373-82.
- [70] Rojas-Rivera J, Ortiz A, Egido J. Antioxidants in kidney diseases: the impact of baradoxolone methyl. *Int J Nephrol*. 2012;2012:321714.
- [71] Brand FN, McGee DL, Kannel WB, Stokes J, 3rd, Castelli WP. Hyperuricemia as a risk factor of coronary heart disease: The Framingham Study. *Am J Epidemiol*. 1985 Jan;121(1):11-8.
- [72] Mitsuhashi H, Tamura K, Yamauchi J, Ozawa M, Yanagi M, Dejima T, et al. Effect of losartan on ambulatory short-term blood pressure variability and cardiovascular remodeling in hypertensive patients on hemodialysis. *Atherosclerosis*. 2009 Nov;207(1):186-90.
- [73] Letsas KP, Korantzopoulos P, Filippatos GS, Mihas CC, Markou V, Gavrielatos G, et al. Uric acid elevation in atrial fibrillation. *Hellenic J Cardiol*. 2010 May-Jun;51(3):209-13.
- [74] Car S, Trkulja V. Higher serum uric acid on admission is associated with higher short-term mortality and poorer long-term survival after myocardial infarction: retrospective prognostic study. *Croat Med J*. 2009 Dec;50(6):559-66.
- [75] Chen JH, Chuang SY, Chen HJ, Yeh WT, Pan WH. Serum uric acid level as an independent risk factor for all-cause, cardiovascular, and ischemic stroke mortality: a Chinese cohort study. *Arthritis Rheum*. 2009 Feb 15;61(2):225-32.
- [76] Shan Y, Zhang Q, Liu Z, Hu X, Liu D. Prevalence and risk factors associated with chronic kidney disease in adults over 40 years: a population study from Central China. *Nephrology (Carlton)*. 2010 Apr;15(3):354-61.



- [77] Chen YC, Su CT, Wang ST, Lee HD, Lin SY. A preliminary investigation of the association between serum uric acid and impaired renal function. *Chang Gung Med J*. 2009 Jan-Feb;32(1):66-71.
- [78] Martinon F, Petrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals activate the NALP3 inflammasome. *Nature*. 2006 Mar 9;440(7081):237-41.
- [79] Sakamaki I, Inai K, Tsutani Y, Ueda T, Tsutani H. Binding of monosodium urate crystals with idiotype protein efficiently promote dendritic cells to induce cytotoxic T cells. *Cancer Sci*. 2008 Nov;99(11):2268-73.
- [80] Amaya Y, Yamazaki K, Sato M, Noda K, Nishino T. Proteolytic conversion of xanthine dehydrogenase from the NAD-dependent type to the O<sub>2</sub>-dependent type. Amino acid sequence of rat liver xanthine dehydrogenase and identification of the cleavage sites of the enzyme protein during irreversible conversion by trypsin. *J Biol Chem*. 1990 Aug 25;265(24):14170-5.
- [81] Nishino T, Okamoto K, Kawaguchi Y, Hori H, Matsumura T, Eger BT, et al. Mechanism of the conversion of xanthine dehydrogenase to xanthine oxidase: identification of the two cysteine disulfide bonds and crystal structure of a non-convertible rat liver xanthine dehydrogenase mutant. *J Biol Chem*. 2005 Jul 1;280(26):24888-94.
- [82] Maia L, Duarte RO, Ponces-Freire A, Moura JJ, Mira L. NADH oxidase activity of rat and human liver xanthine oxidoreductase: potential role in superoxide production. *J Biol Inorg Chem*. 2007 Aug;12(6):777-87.
- [83] Miller NJ, RiceEvans CA. Spectrophotometric determination of antioxidant activity. *Redox Report*. 1996 Jun;2(3):161-71.
- [84] Korish AA. Multiple antioxidants and L-arginine modulate inflammation and dyslipidemia in chronic renal failure rats. *Ren Fail*. 2010 Jan;32(2):203-13.
- [85] Ehara H, Yamamoto-Honda R, Kitazato H, Takahashi Y, Kawazu S, Akanuma Y, et al. ApoE isoforms, treatment of diabetes and the risk of coronary heart disease. *World J Diabetes*. 2012 Mar 15;3(3):54-9.
- [86] Abadir PM, Foster DB, Crow M, Cooke CA, Rucker JJ, Jain A, et al. Identification and characterization of a functional mitochondrial angiotensin system. *Proc Nat Acad Sci USA*. 2011 Sep 6;108(36):14849-54.
- [87] Halliwell B. The wanderings of a free radical. *Free Radic Biol Med*. 2009 Mar 1;46(5):531-42.
- [88] Halliwell B, Whiteman M. Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *Br J Pharmacol*. 2004 May;142(2):231-55.
- [89] Golbidi S, Ebadi SA, Laher I. Antioxidants in the treatment of diabetes. *Curr Diabetes Rev*. 2011 Mar;7(2):106-25.

- [90] Ramos LF, Kane J, McMonagle E, Le P, Wu P, Shintani A, et al. Effects of combination tocopherols and alpha lipoic acid therapy on oxidative stress and inflammatory biomarkers in chronic kidney disease. *J Ren Nutr.* 2011 May;21(3):211-8.
- [91] Yusuf S, Dagenais G, Pogue J, Bosch J, Sleight P. Vitamin E supplementation and cardiovascular events in high-risk patients. The Heart Outcomes Prevention Evaluation Study Investigators. *N Engl J Med.* 2000 Jan 20;342(3):154-60.
- [92] Mann JF, Lonn EM, Yi Q, Gerstein HC, Hoogwerf BJ, Pogue J, et al. Effects of vitamin E on cardiovascular outcomes in people with mild-to-moderate renal insufficiency: results of the HOPE study. *Kidney Int.* 2004 Apr;65(4):1375-80.
- [93] Harcourt BE, Sourris KC, Coughlan MT, Walker KZ, Dougherty SL, Andrikopoulos S, et al. Targeted reduction of advanced glycation improves renal function in obesity. *Kidney Int.* 2011 Jul;80(2):190-8.
- [94] Vlassara H, Torreggiani M, Post JB, Zheng F, Uribarri J, Striker GE. Role of oxidants/inflammation in declining renal function in chronic kidney disease and normal aging. *Kidney Int Suppl.* 2009 Dec(114):S3-11.
- [95] Zafarullah M, Li WQ, Sylvester J, Ahmad M. Molecular mechanisms of N-acetylcysteine actions. *Cell Mol Life Sci.* 2003 Jan;60(1):6-20.
- [96] Zhang F, Lau SS, Monks TJ. The cytoprotective effect of N-acetyl-L-cysteine against ROS-induced cytotoxicity is independent of its ability to enhance glutathione synthesis. *Toxicol Sci.* 2011 Mar;120(1):87-97.
- [97] Ye J, Li J, Yu Y, Wei Q, Deng W, Yu L. L-carnitine attenuates oxidant injury in HK-2 cells via ROS-mitochondria pathway. *Regul Pept.* 2010 Apr 9;161(1-3):58-66.
- [98] Pat B, Yang T, Kong C, Watters D, Johnson DW, Gobe G. Activation of ERK in renal fibrosis after unilateral ureteral obstruction: modulation by antioxidants. *Kidney Int.* 2005 Mar;67(3):931-43.
- [99] Ribeiro G, Roehrs M, Bairros A, Moro A, Charao M, Araujo F, et al. N-acetylcysteine on oxidative damage in diabetic rats. *Drug Chem Toxicol.* 2011 Aug 16.
- [100] Moist L, Sontrop JM, Gallo K, Mainra R, Cutler M, Freeman D, et al. Effect of N-acetylcysteine on serum creatinine and kidney function: results of a randomized controlled trial. *Am J Kidney Dis.* 2010 Oct;56(4):643-50.
- [101] Renke M, Tylicki L, Rutkowski P, Larczynski W, Aleksandrowicz E, Lysiak-Szydlowska W, et al. The effect of N-acetylcysteine on proteinuria and markers of tubular injury in non-diabetic patients with chronic kidney disease. A placebo-controlled, randomized, open, cross-over study. *Kidney Blood Press Res.* 2008;31(6):404-10.
- [102] Hsu SP, Chiang CK, Yang SY, Chien CT. N-acetylcysteine for the management of anemia and oxidative stress in hemodialysis patients. *Nephron Clin Pract.* 2010;116(3):c207-16.

- [103] Nascimento MM, Suliman ME, Silva M, Chinaglia T, Marchioro J, Hayashi SY, et al. Effect of oral N-acetylcysteine treatment on plasma inflammatory and oxidative stress markers in peritoneal dialysis patients: a placebo-controlled study. *Perit Dial Int.* 2010 May-Jun;30(3):336-42.
- [104] Coombes JS, Fassett RG. Antioxidant therapy in hemodialysis patients: a systematic review. *Kidney Int.* 2012 Feb;81(3):233-46.
- [105] Crespo MJ, Cruz N, Altieri PI, Escobales N. Chronic treatment with N-acetylcysteine improves cardiac function but does not prevent progression of cardiomyopathy in Syrian cardiomyopathic hamsters. *J Cardiovasc Pharmacol Ther.* 2011 Jun;16(2):197-204.
- [106] Tumor Z, Shimizu H, Enomoto A, Miyazaki H, Niwa T. Indoxyl sulfate upregulates expression of ICAM-1 and MCP-1 by oxidative stress-induced NF-kappaB activation. *Am J Nephrol.* 2010;31(5):435-41.
- [107] Serbinova E, Kagan V, Han D, Packer L. Free radical recycling and intramembrane mobility in the antioxidant properties of alpha-tocopherol and alpha-tocotrienol. *Free Radic Biol Med.* 1991;10(5):263-75.
- [108] Fujisawa S, Ishihara M, Atsumi T, Kadoma Y. A quantitative approach to the free radical interaction between alpha-tocopherol or ascorbate and flavonoids. *In Vivo.* 2006 Jul-Aug;20(4):445-52.
- [109] Kagan VE, Serbinova EA, Packer L. Recycling and antioxidant activity of tocopherol homologs of differing hydrocarbon chain lengths in liver microsomes. *Arch Biochem Biophys.* 1990 Nov 1;282(2):221-5.
- [110] Kagan VE, Serbinova EA, Forte T, Scita G, Packer L. Recycling of vitamin E in human low density lipoproteins. *J Lipid Res.* 1992 Mar;33(3):385-97.
- [111] Guo Q, Packer L. Ascorbate-dependent recycling of the vitamin E homologue Trolox by dihydrolipoate and glutathione in murine skin homogenates. *Free Radic Biol Med.* 2000 Aug;29(3-4):368-74.
- [112] Schaaf GJ, Maas RF, de Groene EM, Fink-Gremmels J. Management of oxidative stress by heme oxygenase-1 in cisplatin-induced toxicity in renal tubular cells. *Free Radic Res.* 2002 Aug;36(8):835-43.
- [113] Sen CK, Khanna S, Roy S, Packer L. Molecular basis of vitamin E action. Tocotrienol potently inhibits glutamate-induced pp60(c-Src) kinase activation and death of HT4 neuronal cells. *J Biol Chem.* 2000 Apr 28;275(17):13049-55.
- [114] Machlin LJ, Gabriel E. Kinetics of tissue alpha-tocopherol uptake and depletion following administration of high levels of vitamin E. *Ann N Y Acad Sci.* 1982;393:48-60.
- [115] Karamouzis I, Sarafidis PA, Karamouzis M, Iliadis S, Haidich AB, Sioulis A, et al. Increase in oxidative stress but not in antioxidant capacity with advancing stages of chronic kidney disease. *Am J Nephrol.* 2008;28(3):397-404.

- [116] Klein EA, Thompson IM, Jr., Tangen CM, Crowley JJ, Lucia MS, Goodman PJ, et al. Vitamin E and the risk of prostate cancer: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). *JAMA*. 2011 Oct 12;306(14):1549-56.
- [117] Boaz M, Smetana S, Weinstein T, Matas Z, Gafer U, Iaina A, et al. Secondary prevention with antioxidants of cardiovascular disease in endstage renal disease (SPACE): randomised placebo-controlled trial. *Lancet*. 2000 Oct 7;356(9237):1213-8.
- [118] Giray B, Kan E, Bali M, Hincal F, Basaran N. The effect of vitamin E supplementation on antioxidant enzyme activities and lipid peroxidation levels in hemodialysis patients. *Clin Chim Acta*. 2003 Dec;338(1-2):91-8.
- [119] Islam KN, O'Byrne D, Devaraj S, Palmer B, Grundy SM, Jialal I. Alpha-tocopherol supplementation decreases the oxidative susceptibility of LDL in renal failure patients on dialysis therapy. *Atherosclerosis*. 2000 May;150(1):217-24.
- [120] Huang HY, Appel LJ. Supplementation of diets with alpha-tocopherol reduces serum concentrations of gamma- and delta-tocopherol in humans. *J Nutr*. 2003 Oct;133(10):3137-40.
- [121] Baggott JE. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):155-6; author reply 6-8.
- [122] Blatt DH, Pryor WA. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):150-1; author reply 6-8.
- [123] Carter T. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):155; author reply 6-8.
- [124] DeZee KJ, Shimeall W, Douglas K, Jackson JL. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):153-4; author reply 6-8.
- [125] Hemila H. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):151-2; author reply 6-8.
- [126] Krishnan K, Campbell S, Stone WL. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):151; author reply 6-8.
- [127] Lim WS, Liscic R, Xiong C, Morris JC. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):152; author reply 6-8.
- [128] Marras C, Lang AE, Oakes D, McDermott MP, Kieburtz K, Shoulson I, et al. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):152-3; author reply 6-8.
- [129] Meydani SN, Lau J, Dallal GE, Meydani M. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):153; author reply 6-8.
- [130] Possolo AM. High-dosage vitamin E supplementation and all-cause mortality. *Ann Intern Med*. 2005 Jul 19;143(2):154; author reply 6-8.

- [131] Lass A, Forster MJ, Sohal RS. Effects of coenzyme Q10 and alpha-tocopherol administration on their tissue levels in the mouse: elevation of mitochondrial alpha-tocopherol by coenzyme Q10. *Free Radic Biol Med*. 1999 Jun;26(11-12):1375-82.
- [132] Lass A, Sohal RS. Effect of coenzyme Q(10) and alpha-tocopherol content of mitochondria on the production of superoxide anion radicals. *FASEB J*. 2000 Jan;14(1):87-94.
- [133] Merker MP, Audi SH, Lindemer BJ, Krenz GS, Bongard RD. Role of mitochondrial electron transport complex I in coenzyme Q1 reduction by intact pulmonary arterial endothelial cells and the effect of hyperoxia. *Am J Physiol Lung Cell Mol Physiol*. 2007 Sep;293(3):L809-19.
- [134] Ohnishi T, Ohnishi ST, Shinzawa-Ito K, Yoshikawa S. Functional role of coenzyme Q in the energy coupling of NADH-CoQ oxidoreductase (Complex I): stabilization of the semiquinone state with the application of inside-positive membrane potential to proteoliposomes. *Biofactors*. 2008;32(1-4):13-22.
- [135] James AM, Smith RA, Murphy MP. Antioxidant and prooxidant properties of mitochondrial Coenzyme Q. *Arch Biochem Biophys*. 2004 Mar 1;423(1):47-56.
- [136] Linnane AW, Kios M, Vitetta L. Coenzyme Q(10) - its role as a prooxidant in the formation of superoxide anion/hydrogen peroxide and the regulation of the metabolome. *Mitochondrion*. 2007 Jun;7 Suppl:S51-61.
- [137] Nohl H, Gille L, Kozlov AV. Critical aspects of the antioxidant function of coenzyme Q in biomembranes. *Biofactors*. 1999;9(2-4):155-61.
- [138] Frei B, Kim MC, Ames BN. Ubiquinol-10 is an effective lipid-soluble antioxidant at physiological concentrations. *Proc Natl Acad Sci U S A*. 1990 Jun;87(12):4879-83.
- [139] Stoyanovsky DA, Osipov AN, Quinn PJ, Kagan VE. Ubiquinone-dependent recycling of vitamin E radicals by superoxide. *Arch Biochem Biophys*. 1995 Nov 10;323(2):343-51.
- [140] Lass A, Sohal RS. Electron transport-linked ubiquinone-dependent recycling of alpha-tocopherol inhibits autooxidation of mitochondrial membranes. *Arch Biochem Biophys*. 1998 Apr 15;352(2):229-36.
- [141] Linnane AW, Kios M, Vitetta L. Healthy aging: regulation of the metabolome by cellular redox modulation and prooxidant signaling systems: the essential roles of superoxide anion and hydrogen peroxide. *Biogerontology*. 2007 Oct;8(5):445-67.
- [142] Kwong LK, Kamzalov S, Rebrin I, Bayne AC, Jana CK, Morris P, et al. Effects of coenzyme Q(10) administration on its tissue concentrations, mitochondrial oxidant generation, and oxidative stress in the rat. *Free Radic Biol Med*. 2002 Sep 1;33(5):627-38.
- [143] Ishikawa A, Kawarazaki H, Ando K, Fujita M, Fujita T, Homma Y. Renal preservation effect of ubiquinol, the reduced form of coenzyme Q10. *Clin Exp Nephrol*. 2011 Feb;15(1):30-3.



- [144] Huynh K, Kiriazis H, Du XJ, Love JE, Jandeleit-Dahm KA, Forbes JM, et al. Coenzyme Q10 attenuates diastolic dysfunction, cardiomyocyte hypertrophy and cardiac fibrosis in the db/db mouse model of type 2 diabetes. *Diabetologia*. 2012 May;55(5):1544-53.
- [145] Dendooven A, Ishola DA, Jr., Nguyen TQ, Van der Giezen DM, Kok RJ, Goldschmeding R, et al. Oxidative stress in obstructive nephropathy. *Int J Exp Pathol*. 2011 Jun;92(3):202-10.
- [146] Irita J, Okura T, Jotoku M, Nagao T, Enomoto D, Kurata M, et al. Osteopontin deficiency protects against aldosterone-induced inflammation, oxidative stress, and interstitial fibrosis in the kidney. *Am J Physiol Renal Physiol*. 2011 Jul 6.
- [147] Shing CM, Adams MJ, Fassett RG, Coombes JS. Nutritional compounds influence tissue factor expression and inflammation of chronic kidney disease patients in vitro. *Nutrition*. 2011 Sep;27(9):967-72.
- [148] Arab K, Rossary A, Flourie F, Tourneur Y, Steghens JP. Docosahexaenoic acid enhances the antioxidant response of human fibroblasts by upregulating gamma-glutamylcysteinyl ligase and glutathione reductase. *Br J Nutr*. 2006 Jan;95(1):18-26.
- [149] Kim YJ, Chung HY. Antioxidative and anti-inflammatory actions of docosahexaenoic acid and eicosapentaenoic acid in renal epithelial cells and macrophages. *J Med Food*. 2007 Jun;10(2):225-31.
- [150] Mayer K, Meyer S, Reinholz-Muhly M, Maus U, Merfels M, Lohmeyer J, et al. Short-time infusion of fish oil-based lipid emulsions, approved for parenteral nutrition, reduces monocyte proinflammatory cytokine generation and adhesive interaction with endothelium in humans. *J Immunol*. 2003 Nov 1;171(9):4837-43.
- [151] Sperling RI, Benincaso AI, Knoell CT, Larkin JK, Austen KF, Robinson DR. Dietary omega-3 polyunsaturated fatty acids inhibit phosphoinositide formation and chemotaxis in neutrophils. *J Clin Invest*. 1993 Feb;91(2):651-60.
- [152] Diaz Encarnacion MM, Warner GM, Cheng J, Gray CE, Nath KA, Grande JP. n-3 Fatty acids block TNF-alpha-stimulated MCP-1 expression in rat mesangial cells. *Am J Physiol Renal Physiol*. 2011 May;300(5):F1142-51.
- [153] An WS, Kim HJ, Cho KH, Vaziri ND. Omega-3 fatty acid supplementation attenuates oxidative stress, inflammation, and tubulointerstitial fibrosis in the remnant kidney. *Am J Physiol Renal Physiol*. 2009 Oct;297(4):F895-903.
- [154] Peake JM, Gobe GC, Fassett RG, Coombes JS. The effects of dietary fish oil on inflammation, fibrosis and oxidative stress associated with obstructive renal injury in rats. *Mol Nutr Food Res*. 2011 Mar;55(3):400-10.
- [155] Kazemian P, Kazemi-Bajestani SM, Alherbish A, Steed J, Oudit GY. The use of omega-3 poly-unsaturated fatty acids in heart failure: a preferential role in patients with diabetes. *Cardiovasc Drugs Ther*. 2012 May 30.

- [156] Bouzidi N, Mekki K, Boukaddoum A, Dida N, Kaddous A, Bouchenak M. Effects of omega-3 polyunsaturated fatty-acid supplementation on redox status in chronic renal failure patients with dyslipidemia. *J Ren Nutr.* 2010 Sep;20(5):321-8.
- [157] Harris WS. n-3 fatty acids and serum lipoproteins: human studies. *Am J Clin Nutr.* 1997 May;65(5 Suppl):1645S-54S.
- [158] Cohen MG, Rossi JS, Garbarino J, Bowling R, Motsinger-Reif AA, Schuler C, et al. Insights into the inhibition of platelet activation by omega-3 polyunsaturated fatty acids: Beyond aspirin and clopidogrel. *Thromb Res.* 2011 May 26.
- [159] Woodman RJ, Mori TA, Burke V, Puddey IB, Watts GF, Beilin LJ. Effects of purified eicosapentaenoic and docosahexaenoic acids on glycemic control, blood pressure, and serum lipids in type 2 diabetic patients with treated hypertension. *Am J Clin Nutr.* 2002 Nov;76(5):1007-15.
- [160] Matsumoto T, Nakayama N, Ishida K, Kobayashi T, Kamata K. Eicosapentaenoic acid improves imbalance between vasodilator and vasoconstrictor actions of endothelium-derived factors in mesenteric arteries from rats at chronic stage of type 2 diabetes. *J Pharmacol Exp Ther.* 2009 Apr;329(1):324-34.
- [161] El-Sheikh AA, van den Heuvel JJ, Koenderink JB, Russel FG. Effect of hypouricaemic and hyperuricaemic drugs on the renal urate efflux transporter, multidrug resistance protein 4. *Br J Pharmacol.* 2008 Dec;155(7):1066-75.
- [162] Riegersperger M, Covic A, Goldsmith D. Allopurinol, uric acid, and oxidative stress in cardiorenal disease. *Int Urol Nephrol.* 2011 Jun;43(2):441-9.
- [163] Sanders SA, Eisenthal R, Harrison R. NADH oxidase activity of human xanthine oxidoreductase--generation of superoxide anion. *Eur J Biochem.* 1997 May 1;245(3):541-8.
- [164] Kosugi T, Nakayama T, Heinig M, Zhang L, Yuzawa Y, Sanchez-Lozada LG, et al. Effect of lowering uric acid on renal disease in the type 2 diabetic db/db mice. *Am J Physiol Renal Physiol.* 2009 Aug;297(2):F481-8.
- [165] Das DK, Engelman RM, Clement R, Otani H, Prasad MR, Rao PS. Role of xanthine oxidase inhibitor as free radical scavenger: a novel mechanism of action of allopurinol and oxypurinol in myocardial salvage. *Biochem Biophys Res Commun.* 1987 Oct 14;148(1):314-9.
- [166] Moorhouse PC, Grootveld M, Halliwell B, Quinlan JG, Gutteridge JM. Allopurinol and oxypurinol are hydroxyl radical scavengers. *FEBS Lett.* 1987 Mar 9;213(1):23-8.
- [167] Klein AS, Joh JW, Rangan U, Wang D, Bulkley GB. Allopurinol: discrimination of antioxidant from enzyme inhibitory activities. *Free Radic Biol Med.* 1996;21(5):713-7.
- [168] Goicoechea M, de Vinuesa SG, Verdalles U, Ruiz-Caro C, Ampuero J, Rincon A, et al. Effect of allopurinol in chronic kidney disease progression and cardiovascular risk. *Clin J Am Soc Nephrol.* 2010 Aug;5(8):1388-93.

- [169] Kanbay M, Ozkara A, Selcoki Y, Isik B, Turgut F, Bavbek N, et al. Effect of treatment of hyperuricemia with allopurinol on blood pressure, creatinine clearance, and proteinuria in patients with normal renal functions. *Int Urol Nephrol*. 2007;39(4):1227-33.
- [170] Shelmadine B, Bowden RG, Wilson RL, Beavers D, Hartman J. The effects of lowering uric acid levels using allopurinol on markers of metabolic syndrome in end-stage renal disease patients: a pilot study. *Anadolu Kardiyol Derg*. 2009 Oct;9(5):385-9.
- [171] Pergola PE, Krauth M, Huff JW, Ferguson DA, Ruiz S, Meyer CJ, et al. Effect of bardoxolone methyl on kidney function in patients with T2D and Stage 3b-4 CKD. *Am J Nephrol*. 2011;33(5):469-76.
- [172] Pergola PE, Raskin P, Toto RD, Meyer CJ, Huff JW, Grossman EB, et al. Bardoxolone methyl and kidney function in CKD with type 2 diabetes. *N Engl J Med*. 2011 Jul 28;365(4):327-36.
- [173] Honda T, Yoshizawa H, Sundararajan C, David E, Lajoie MJ, Favalaro FG, Jr., et al. Tricyclic compounds containing nonenolizable cyano enones. A novel class of highly potent anti-inflammatory and cytoprotective agents. *J Med Chem*. 2011 Mar 24;54(6):1762-78.
- [174] Eskiocak U, Kim SB, Roig AI, Kitten E, Batten K, Cornelius C, et al. CDDO-Me protects against space radiation-induced transformation of human colon epithelial cells. *Radiat Res*. 2010 Jul;174(1):27-36.
- [175] Nagaraj S, Youn JI, Weber H, Iclozan C, Lu L, Cotter MJ, et al. Anti-inflammatory triterpenoid blocks immune suppressive function of MDSCs and improves immune response in cancer. *Clin Cancer Res*. 2010 Mar 15;16(6):1812-23.
- [176] Wu QQ, Wang Y, Senitko M, Meyer C, Wigley WC, Ferguson DA, et al. Bardoxolone methyl (BARD) ameliorates ischemic AKI and increases expression of protective genes Nrf2, PPARgamma, and HO-1. *Am J Physiol Renal Physiol*. 2011 May;300(5):F1180-92.
- [177] Kelly GS. L-Carnitine: therapeutic applications of a conditionally-essential amino acid. *Altern Med Rev*. 1998 Oct;3(5):345-60.
- [178] Fouque D, Holt S, Guebre-Egziabher F, Nakamura K, Vianey-Saban C, Hadj-Aissa A, et al. Relationship between serum carnitine, acylcarnitines, and renal function in patients with chronic renal disease. *J Ren Nutr*. 2006 Apr;16(2):125-31.
- [179] Eknoyan G, Latos DL, Lindberg J. Practice recommendations for the use of L-carnitine in dialysis-related carnitine disorder. National Kidney Foundation Carnitine Consensus Conference. *Am J Kidney Dis*. 2003 Apr;41(4):868-76.
- [180] Sakurabayashi T, Miyazaki S, Yuasa Y, Sakai S, Suzuki M, Takahashi S, et al. L-carnitine supplementation decreases the left ventricular mass in patients undergoing hemodialysis. *Circ J*. 2008 Jun;72(6):926-31.

- [181] Matsumoto Y, Sato M, Ohashi H, Araki H, Tadokoro M, Osumi Y, et al. Effects of L-carnitine supplementation on cardiac morbidity in hemodialyzed patients. *Am J Nephrol*. 2000 May-Jun;20(3):201-7.
- [182] Signorelli SS, Fatuzzo P, Rapisarda F, Neri S, Ferrante M, Oliveri CG, et al. Propionyl-L-carnitine therapy: effects on endothelin-1 and homocysteine levels in patients with peripheral arterial disease and end-stage renal disease. *Kidney Blood Pressure Res*. 2006;29(2):100-7.
- [183] Zhou Q, Wu S, Jiang J, Tian J, Chen J, Yu X, et al. Accumulation of circulating advanced oxidation protein products is an independent risk factor for ischemic heart disease in maintenance hemodialysis patients. *Nephrology*. 2012 Jun 28.
- [184] Sener G, Paskaloglu K, Satiroglu H, Alican I, Kacmaz A, Sakarcan A. L-carnitine ameliorates oxidative damage due to chronic renal failure in rats. *J Cardiovasc Pharmacol*. 2004 May;43(5):698-705.
- [185] Pecoits-Filho R, Lindholm B, Stenvinkel P. The malnutrition, inflammation, and atherosclerosis (MIA) syndrome -- the heart of the matter. *Nephrol Dial Transplant*. 2002;17 Suppl 11:28-31.
- [186] Brunini TM, da SCD, Siqueira MA, Moss MB, Santos SF, Mendes-Ribeiro AC. Uremia, atherothrombosis and malnutrition: the role of L-arginine-nitric oxide pathway. *Cardiovasc Hematol Disord Drug Targets*. 2006 Jun;6(2):133-40.
- [187] Reyes AA, Karl IE, Klahr S. Role of arginine in health and in renal disease. *Am J Physiol*. 1994 Sep;267(3 Pt 2):F331-46.
- [188] Morris SM, Jr. Enzymes of arginine metabolism. *J Nutr*. 2004 Oct;134(10 Suppl):2743S-7S; discussion 65S-67S.
- [189] Stuehr DJ. Enzymes of the L-arginine to nitric oxide pathway. *J Nutr*. 2004 Oct;134(10 Suppl):2748S-51S; discussion 65S-67S.
- [190] Morel F, Hus-Citharel A, Levillain O. Biochemical heterogeneity of arginine metabolism along kidney proximal tubules. *Kidney Int*. 1996 Jun;49(6):1608-10.
- [191] Mendes RAC, Brunini TM, Ellory JC, Mann GE. Abnormalities in L-arginine transport and nitric oxide biosynthesis in chronic renal and heart failure. *Cardiovasc Res*. 2001 Mar;49(4):697-712.
- [192] Brunini TM, Roberts NB, Yaqoob MM, Ellory JC, Mann GE, Mendes RAC. Activation of L-arginine transport in undialysed chronic renal failure and continuous ambulatory peritoneal dialysis patients. *Clin Exp Pharmacol Physiol*. 2006 Jan-Feb;33(1-2):114-8.
- [193] Mendes RAC, Hanssen H, Kiessling K, Roberts NB, Mann GE, Ellory JC. Transport of L-arginine and the nitric oxide inhibitor NG-monomethyl-L-arginine in human erythrocytes in chronic renal failure. *Clin Sci (Lond)*. 1997 Jul;93(1):57-64.

- [194] Noris M, Benigni A, Boccardo P, Aiello S, Gaspari F, Todeschini M, et al. Enhanced nitric oxide synthesis in uremia: implications for platelet dysfunction and dialysis hypotension. *Kidney Int.* 1993 Aug;44(2):445-50.
- [195] Aiello S, Noris M, Remuzzi G. Nitric oxide synthesis and L-arginine in uremia. *Miner Electrolyte Metab.* 1997;23(3-6):151-6.
- [196] Rajapakse NW, Kuruppu S, Hanchapola I, Venardos K, Mattson DL, Smith AI, et al. Evidence that renal arginine transport is impaired in spontaneously hypertensive rats. *Am J Physiol Renal Physiol.* 2012 Jun;302(12):F1554-62.
- [197] Huang CC, Tsai SC, Lin WT. Potential ergogenic effects of L-arginine against oxidative and inflammatory stress induced by acute exercise in aging rats. *Exp Gerontol.* 2008 Jun;43(6):571-7.
- [198] Mosca L, Marcellini S, Perluigi M, Mastroiacovo P, Moretti S, Famularo G, et al. Modulation of apoptosis and improved redox metabolism with the use of a new antioxidant formula. *Biochem Pharmacol.* 2002 Apr 1;63(7):1305-14.
- [199] Park NY, Park SK, Lim Y. Long-term dietary antioxidant cocktail supplementation effectively reduces renal inflammation in diabetic mice. *Br J Nutr.* 2011 Nov;106(10):1514-21.
- [200] Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: results of the GISSI-Prevenzione trial. Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto miocardico. *Lancet.* 1999 Aug 7;354(9177):447-55.
- [201] Shargorodsky M, Debby O, Matas Z, Zimlichman R. Effect of long-term treatment with antioxidants (vitamin C, vitamin E, coenzyme Q10 and selenium) on arterial compliance, humoral factors and inflammatory markers in patients with multiple cardiovascular risk factors. *Nutr Metab (Lond).* 2010;7:55.

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