

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



## Is Chronic Lymphocytic Leukemia a Mistake of Tolerance Mechanisms?

Ricardo García-Muñoz<sup>1</sup>, Judit Anton-Remirez<sup>2</sup>, Jesus Feliu<sup>3</sup>,  
María Pilar Rabasa<sup>1</sup>, Carlos Panizo<sup>4</sup> and Luis Llorente<sup>5</sup>

<sup>1</sup>Hematology Service, Hospital San Pedro, Logroño, La Rioja,

<sup>2</sup>Rehabilitation Service, Complejo Hospitalario de Navarra, Pamplona, Navarra,

<sup>3</sup>Hematology Service, Complejo Hospitalario de Navarra, Pamplona, Navarra,

<sup>4</sup>Hematology Service, Clínica Universidad de Navarra, Pamplona, Navarra,

<sup>5</sup>Department of Immunology and Rheumatology, Instituto Nacional  
de Ciencias Médicas y Nutrición Salvador Zubirán, Mexico City

<sup>1,2,3,4</sup>Spain

<sup>5</sup>México

### 1. Introduction

Chronic Lymphocytic Leukemia (CLL) is a chronic lymphoproliferative disorder of the B lymphocytes. Small lymphocytic lymphoma (SLL) is considered to be the same disease in a non-leukemic form. CLL remains as an incurable tumour and clinical features have very variable presentation, course, and outcome. The progressive accumulation of monoclonal B lymphocytes leads to leukocytosis, lymphadenopathy, hepatosplenomegaly and marrow failure, and is sometimes associated with autoimmune manifestations.

It has been suggested that CLL cells are defective in apoptosis, which leads to the accumulation of malignant B cells. Furthermore, patients with proliferation rates greater than 0.35% per day have been found to have a more aggressive disease<sup>18,19</sup>. Proliferation of CLL cells is most prominent in proliferative centers that include specific areas in lymph nodes and bone marrow<sup>20,21</sup>. Numerous CD4 T cells and dendritic cells are in close contact with CLL B cells<sup>22</sup>, and micro environmental interactions like BM stromal cells are able to extend the survival of CLL upon direct contact<sup>21</sup>. Thus, the CLL population may originate from a clone with few or no V- domain mutations, or from a more mature clone whose V-domains have undergone the hypermutation process. This creates two separate pools of B cells, both of which originate from antigen-stimulated B lymphocytes. Additionally, IGHV unmutated CLL B cells expressing polyreactive antibodies whereas most IGHV mutated CLL's did not. However, reversion of the IGHV mutated sequences to germline counterparts restored the polyreactivity (Herve et al 2005). Despite these features, the biological etiology of the divergent natural histories of IgVH unmutated vs mutated CLL and the origin of this type of leukemia/lymphoma remains unknown. For this reason we review the immunologic aspects that can help to understand this complex disease based in the findings that suggest that both unmutated and mutated subgroups of patients originally derive from autoreactive clones.

## 2. Diagnosis

The diagnosis of CLL requires the presence of at least 5000 B lymphocytes/ $\mu$ L in the peripheral blood (Hallek, et al 2008). CLL/SLL can be identified by the immunophenotype CD5+, CD10-, CD19+, CD20+, dim expression of surface immunoglobulin, CD23+, CD43 +/-, and cyclin D1- (Matutes, et al 2007). The absence of cyclin D1 is critical in distinguishing CLL/SLL from MCL. Bone marrow involvement is characteristically more than 30% of the nucleated cells in the aspirate are lymphoid.

### Prognostic Markers and Genomic Aberrations

A favorable prognosis in CLL/SLL is associated with the presence of a mutated immunoglobulin heavy chain variable region, and low CD38 and zeta-chain-associated protein kinase 70 protein expression (Damle et al 1999; Kröber et al 2002; Hamblin et al 1999; Tobbin et al 2002; Crespo et al 2003). Chromosomal aberrations in CLL include del 6q, del 11q, del 13q, trisomy 12, and del 17p (Döhner et al 2000). Importantly, specific genomic aberrations have been associated with disease characteristics such better survival for patients with 12q trisomy and 13q deletion, poor survival and massive lymphadenopathy in 11q deletion and resistance to therapy in the group of patients with 17p deletion and p53 abnormalities (Döhner et al 2000; Döhner et al 1995; Döhner et al 1997; Kröber et al 2006). In addition, two miRNA (miR-15a and miR-16-1) were recently identified to be located in the critical region of the 13q14 deletion and their absence in CLL appears to be a major factor in preventing apoptosis and progression through the cell cycle (Aqeilan et al 2010; Cimmino et al 2005; Callin et al 2004; Mertens et al 2006).

### Pathophysiology and cell of origin/normal counterpart of CLL

Different from other types of malignancies derived from mature B cells, the pathogenesis of B-CLL/SLL is much less understood. Notwithstanding extensive searching it is not known whether there is an equivalent normal cell in which the CLL arise. However, several cell types have been suggested as giving rise to chronic lymphocytic leukemia included memory, transitional, B1 and marginal zone B cells (Chiorazzi and Ferrarini 2011; Griffin et al 2011). In addition, it is not certain at what stage in lymphocyte maturation the CLL cell arises, since roughly equal numbers seem to come from pre-germinal center B lymphocytes (unmutated group) and post-germinal center B lymphocytes (mutated group). However, the comparison of CLL gene expression profiles with those of purified normal B cell subpopulations indicates that the common CLL gene expression profile is more related to memory B cells than to those derived from naïve B cells, CD5+ B cells, or germinal center centroblasts and centrocytes (Klein et al 2001; Rosenwald et al 2001, Klein and Dalla-Favera 2005). Interestingly, unmutated and mutated chronic lymphocytic leukemias derive from self reactive B cell precursors despite expressing different antibody reactivity (Herve et al 2005). This similar expression profile also suggest that the consequences or even the mechanism of transformation may be similar, irrespective of IGHV mutations status. This too suggests that rather than having a cellular origin or cellular subtype, CLL is originated by a coordinated normal immunologic tolerance mechanism to destroy self-reactive B cells and to avoid autoimmunity during their process of differentiation. This point of view is supported by the fact that some CLL mutated and unmutated cases derive from self-reactive B cells (Herve et al 2005) had evidence of multiple, related rearranged heavy and light chain immunoglobulin genes (Volkheimer et al 2007; Hadzidimitriou et al 2009, Stamatopoulos et

al 1996); some express more than one functional Ig heavy chain (Rassenti et al 1997), some had been anergized (Mockridge et al 2007; Muzio et al 2008), edited (Hadzidimitriou et al 2009, Stamatopoulos et al 1996), switched (Cerutti et al 2002) and/or had progressive immunoglobulin gene mutation (Volkheimer et al 2007; Roudier et al 1990; Ruzickova et al 2002).

### **Hypothesis: Autoimmunity as origin of CLL**

The basic hypothesis of the origin of autoimmune disease depends of the emergence of a clone or a small number of clones of T and B lymphocytes capable of damaging interaction with normal cells of organ or tissue involved. Each clone is initiated from a cell which has developed an immune receptor adequately reactive with an accessible self antigen as a result of a V/D/J gene recombination in bone marrow (unmutated) or during somatic mutations in germinal centers (mutated). Importantly, this newly self-reactive cell ("forbidden clone") is anomalously resistant to inactivation by central and peripheral tolerance check points (Burnet 1972). Similar to an autoimmune disease, some lymphoproliferative diseases (marginal zone lymphomas and chronic lymphocytic leukemia) depends of the emergence of a clone capable of interact with an (auto) antigen and with other normal cells and an specific microenvironment to proliferate and survive. In a parallel way, newly malignant B cells are anomalously resistant to apoptosis and proliferate as result of acquisition of genetic damage during V/D/J gene recombination, somatic mutations, class switching and receptor edition/revision. Importantly, with the exception of class switching, the other mechanisms to increase the diversity of B cell receptors might induce both self-reactivity and/or DNA damage.

### **B cell development and autoimmunity**

The current model of the pathogenesis of CLL suggest that stimulation by (self) antigens provides a pro-survival and possibly pro-proliferative advantage for CLL (precursor) cells, most likely leading initially to oligoclonal and subsequently monoclonal selection of malignant cells (Mertens et al 2011)

In humans, B cells develop from progenitors within the bone marrow (Fig1). The stages of B cell ontogeny from pro-B to pre-B to early B to mature B cells are marked by phenotypic changes, the most important of which is expression of the BCR for antigen on the cell surface at the early B cell stage of development (van Lochem et al 2004; Fuda et al 2009). During the course of ontogenesis, B cells mature in the bone marrow according to the evolution of the Ig chain synthesis. Starting with the rearrangement of the V/D/J genes for the heavy chain at the pre-B stage, the recombination process continues through the VJ gene rearrangements for kappa light chain or for the lambda light chain at the immature stage. Thus, the resulting receptor (BCR) comprised of randomly selected heavy and light chains have an unpredictable specificity that could include ability to bind "self". However, there are tolerance check points at every stage of B cell activation and maturation (table 1 and 2). This tolerance mechanisms in bone marrow include receptor editing, clonal deletion, clonal anergy and differentiation to B1 cells (Goodnow et al 2005; Radic et al 1993; Tiegs et al 1993; Nemazee et al 2000; Luning Prak et al 2011) Notably, current evidence suggest that anergy, receptor edition and differentiation to B1 B cells could be implicated in the generation of CLL B cells (Herve et al 2005; Chu et al 2010; Mockridge et al 2007; Hadzidimitriou et al 2009, Stamatopoulos et al 1996; Ghia et al 2008a; Rassenti & Kipps 1997; Murray et al 2008;

Griffin et al 2011 ). Additionally , hematopoietic stem cells sorted from a CLL patient's bone marrow produce CLL like disease when transplanted into immunosuppressed mice (Kikushige et al 2011). Importantly, autoreactive B cells may suffer receptor editing and anergy in bone marrow. At the same, recent evidence shows that L chain receptor editing occurs not only in bone marrow with a pre-B/immature B cell phenotype but also in immature/transitional splenic B cells. Nevertheless, editing at the H chain locus appears to occur exclusively in bone marrow cells with pro-B phenotype (Nakajima et al 2009).

Repertoire analyses of antibodies cloned from B cells derived from bone marrow and peripheral blood of healthy donors provide evidence for both a central tolerance check point in the bone marrow and a second peripheral checkpoint, as evidenced by a decrease in the frequency of autoreactive antibodies from 75% in bone marrow to 20% in the circulating naïve compartment (Yurasov, et al. 2005). Other tolerance mechanisms and peripheral check points include memory development check points (Tsuiji et al 2006) CD5+ expression (Morikawa et al 1993; Gary-Gouy et al 2002; Hillion et al 2005; Hippen et al 2000, Gary-Gouy et al 2002b, Dallou et al 2008), germinal centre exclusion (Cappione et al 2005; Pugh-Bernard et al 2001), receptor edition/revision (Luning Prak et al 2011), antibody feedback (Ravetch & Bolland 2001), anti-idiotypic network (Jerne 1974; Jerne 1984; Forni et al 1980) and all contribute to maintain tolerance and avoid autoimmune diseases.

The contribution of this mechanism in the development of CLL remain unknown, however, Ghia et al describe that CLL expressing IGHV3-21/IGVL3-21 most likely were derived from B cells that had experienced somatic mutation and germinal center maturation in an apparent antigen driven immune response previous to undergoing Ig receptor editing and after germinal-center leukemogenic selection (Ghia et al 2008b). This suggest that peripheral tolerance mechanism also contribute to the shape of self reactive CLL B cells generated and selected after somatic hypermutation. Other mechanisms as germinal centre exclusion, defects in antibody feedback and anti-idiotypic network in lymphoproliferative disorders remain unsolved, however some conjectures about their role have been proposed (García-Muñoz 2009a; García-Muñoz et al 2009b).

The fact that unmutated and mutated chronic lymphocytic leukemias derive from self reactive B cell precursors despite expressing different antibody reactivity (Herve et al 2005) suggest that this B cells escape from tolerance mechanisms. Even more Chiorazzi and Ferrarini suggest that CLL derives from competent B lymphocytes selected for clonal expansion and eventual transformation by multiple encounters and responses to (auto)antigen(s) (Chiorazzi and Ferrarini 2003). This two characteristics of CLL B cells guide us to think that CLL is the product of the selective pressure of tolerance check points in an auto-reactive B cell.

### **Development of Unmutated CLL B cells**

Tumors displaying unmutated V genes have a shorter median survival, in one study of 99 months vs 293 months in the mutated cases (Hamblin et al 1999). Here, a cut-off of  $\geq 98\%$  homology to donor germline gene has been used to define unmutated tumor V genes to allow for a low degree of polymorphic allelic variation. There is an association between unfavorable cytogenetic aberrations (del 17p and del 11q) and unmutated CLL, although 13q- is more frequent in mutated CLL. However, there are discrepancies with many cases having some high-risk and other low-risk molecular features and more than 50% of IgVH



unmutated cases have no unfavorable cytogenetics (Krober et al 2006). Prominently unmutated CLL B cells are self reactive or polyreactive (Herve et al 2005) and seem that they are resistant to several tolerance mechanism.

#### **Are unmutated CLL B cells invulnerable to anergy?**

Low BCR signaling induced by weak reactivity to self antigens induce B cells to enter a tolerized but alive state referred to as anergy (Gauld et al 2006; Getahun et al 2009). In most cases, anergic B cells are characterized by chronic low level BCR signaling and exhibit reduced surface IgM levels but can express high levels of IgD (Getahun et al 2009; Goodnow et al 1998; Dolmetsh et al 1997). Interestingly, anergy depends on the degree of BCR occupancy and require constant transduction of a BCR signal (Goodnow et al 1989; Benshop et al 2001; Gauld et al 2005). Although it is clear that stimulation through the BCR occurred during the natural history of all types of CLL, it is quite peculiar that unmutated CLL cells retain the capacity to transmit signals through the BCR via surface IgM (Lanham et al 2003). The low expression of the BCR is the hallmark of CLL cells and anergic B cells, and appears to contribute towards producing poorer responses to BCR stimulation. Despite low levels of surface expressed immunoglobulin, signalling through the B cell receptor is possible. ZAP-70 expression has shown to augment signalling via IgM ligation in CLL cells as measured by phosphorylation of downstream mediators such as Syk, BLNK and PLC and calcium influx (Chen et al 2005). This increased signalling might lead to enhanced proliferation or survival of the leukemic cell (Bernal et al 2001). Significantly, a number of studies have shown a strong association between ZAP-70 expression and unmutated IGHV genes. This findings could imply that if an immature self-reactive B cell recognize an auto-antigen and also express ZAP-70 survival and activating signals prevail over anergy. In this case a self reactive CLL B cell selected by a self-antigen during B cell development in bone marrow might mature despite they undergo an anergy process and likely to progress to transitional and mature B cell.

Unmutated CLL cases are more frequently CD38 (66-77%) and ZAP-70 (93%) positive, exhibit IgM<sup>+</sup> and IgD<sup>+</sup> surface immunoglobulin, express higher amounts of BCR and response better to stimulation compared with mutated CLL's (Wiestner et al 2003, Hamblin et al 2002; Thumberg et al 2001; Döhner et al 2000; Mockridge et al 2007; Guarini et al 2008). This characteristics suggest that this unmutated CLL B cells where resistant to anergy and progress to mature autoreactive naive B cells.

#### **Receptor editing be unsuccessful to avoid self-reactivity and might induce polyreactive BCR in unmutated CLL B cells**

Immature B cells expressing self-reactive IgM antibodies may undergo repeated rounds of light chain rearrangement to lessen the self specificity of the antibody, a process termed receptor editing (Nemazee et al 2000; Luning Prak et al 2011). Evidence of receptor editing in CLL is provided by the fact that a number of CLL's have multiple light chain rearrangements (Hadzidimitriou et al 2009). B cell receptor of CLL B cells react with recurrent self antigens in vitro including IgG, thyroglobulin, DNA, actin, cardiolipin and others as well as microbial antigens and epitopes exposed on cell surface as a result of apoptosis and also could be stimulated by stroma-derived antigens (Sthoeger et al 1989; Dighiero et al 1991; Chiorazzi et al 2005; Lanemo Myhrinder et al 2008). Sustained or repetitive BCR signaling promotes survival in CLL cells (Petlickovsky et al 2005; Bernal et al

2001). Notably, unmutated CLL B cells are self reactive or polyreactive (Herve et al 2005). Interestingly, 79.3% of unmutated CLL antibodies are polyreactive (Herve et al 2005), and reactivity with a particular form of apoptotic cells is a common feature of this subset (Chu et al 2010). Even more, recently Rozcova et al revealed that Toll like receptor 9 (TLR-9) agonists are a potent stimulus from CLL B cells and induce proliferation, expression of CD38 and secretion of cytokines (Rozcova et al 2010). Outstandingly, TLR-9 recognition of self-molecules (nucleic acids in apoptotic cells) of the host, which are not easily distinguishable from those of no-self (infectious organisms) has the potential to provoke autoimmune diseases. Intriguingly, the unmutated CLL subset expresses antibodies with long heavy and light chain CDR3 (Herve 2005) and some cases of unmutated CLL with 100% of IGHV identity have multiple light chain rearrangements (Hadzidimitriou et al 2009), associated with receptor edition. This suggest that receptor editing mechanisms could be not working well in this subset, even more is possible that increase polyreactivity (Luning Prak et al 2011; Binder et al 2010) and promote survival of self-reactive (Sandel et al 1999) CLL B cells. Consequently, BCRs that react with diverse epitopes may be more prone to sustained signaling. As a result, some unmutated CLL B cells expressing multireactive BCR have a more aggressive course than CLLs expressing less reactive BCRs (Binder et al 2010).

#### **Are unmutated CLL B cells insensitive to CD5 action?**

Induction of CD5 by autoantigen might be a mechanism by which the production of autoantibodies is avoided and also maintains tolerance in anergic B cells (Berland et al 2002; Hippen et al 2000 ). Recently, a very interesting observation was made that many CLL leukemia antibodies recognize non-muscle myosin heavy chain IIA exposed apoptotic cells (MEACs) and that natural antibodies from human serum also react with MEACs. In this study 15 of 16 MEAC-reactive CLL mAbs carried unmutated IGVH genes (Chu et al 2010). Several mechanisms are involved in the tolerance associated with expression of CD5. Likewise, CD5 expression prevents B lymphocytes from uncontrolled self reactivity increasing the BCR signalling threshold<sup>51</sup>, and is associated with reexpresion of RAG, receptor edition/revision, and lack of responsiveness to BAFF in some cells outside bone marrow and germinal centres (Lee et al 2009; Hippen et al 2000, Hillion et al 2005). Along this line, the fact that anergic autoreactive B cells may express CD5+ and that immunoglobulin secreted by unmutated B-CLL cells is often autoreactive and react with a variety of autoantigens (including Fc portion of IgG, DNA, histones, cardiolipin, cytoskeletal proteins and insulin) support the notion that unmutated self-reactive B CLL cells are under check to avoid pathogenic autoimmunity (Broker et al 1988; Caligaris-Cappio et al 1996; Morbach et al 2006). We speculate that the expression of ZAP-70 and CD38 could encourage the stimulation of unmutated CLL B cells and overcome the inhibition induced by CD5. In addition, CD5 does not inhibit properly the BCR mediating signalling in leukemic B cells and in some cases provide viability signals or/and promote CLL B cell survival (Perez-Chacon et al 2007; Perez-Chacon 2007b; Gary-Gouy et al 2007; Gary-Gouy et al 2002; Gary-Gouy et al 2002).

#### **Are unmutated CLL B cells transformed human B1 cells?**

Similarities between normal human B1 cells and malignant chronic lymphocytic leukemia (CLL) cells, include that both are CD20+CD27+CD43+CD70-; most normal B1 cells express CD5, as do malignant CLL cells; and, both express relatively nonmutated IGHV. In addition,

normal human B1 cells are ZAP-70+ like unmutated CLL cells. As a final point, in respect to pathophysiology, Griffin et al propose that the chronically activated phenotype of normal B1 cells may predispose to malignant transformation (Griffin et al 2011).

#### **Are unmutated CLL naïve self-reactive B cells efficiently excluded by germinal centres?**

In order to prevent autoimmunity, censoring mechanisms, including anergy and sequestration into the marginal zone, ultimately forbid the participation of mature autoreactive B cells in productive germinal centres reactions, thereby precluding their expansion into the long-lived IgG memory and plasma cell compartments. Importantly, most self reactive and polyreactive IgG antibodies originate from non self-reactive B cells that acquired reactivity by somatic hypermutation (Tiller et al 2007). Significantly, somatic hypermutation does not appear to occur uniformly among CLL IGHV genes (Chiorazzi et al 2005; Fais et al 1998; Tobin et al 2002; Ghia et al 2005) and might suggest the effect of germinal centre exclusion and tolerance mechanisms to maintain the self-reactive BCR in a germ line state and avoid the participation of unmutated CLL cases in germinal centres reactions.

#### **Development of Mutated CLL B cells**

Fifty percent of CLL patients have undergone somatic hypermutation in IGHV, and these patients have a more indolent clinical course and longer survival than those without somatic hypermutation (Hamblin et al 1999; Damle et al 1999). The majority of cases of mutated CLL fail to signal via IgM in vitro (Lanham et al 2003; Chen et al 2002). Interestingly, CLL B cells that express only IgD+ are linked to mutated IGHV genes, negative or low CD38 expression, and 50% of mutated CLL cases unable to signal via IgM were able to signal via IgD (Stevenson et al 2004). Muzio et al, showed that CLL B cells (typically IGH-mutated cases) that do not respond to BCR ligation show activation cellular pathways that suggest anergy (Muzio et al 2008). Essentially, mutated CLL cases derive from B cells with self-reactive receptors that were anergized, edited or regulated to avoid autoimmunity. This is supported by the fact that when mutated non autoreactive immunoglobulin sequences of mutated CLL cases were reverted to their germline counterparts, they encoded polyreactive and autoreactive antibodies (Herve 2005). Despite somatic hypermutation had been proposed as a mechanism to change original BCR self reactivity (germ line) towards some non-self BCR (Murray et al 2008), this is an eccentric mode to loss self reactivity because, self reactive naïve B cells are efficiently excluded from germinal centres (Tsuji et al 2006; Cappione A 3<sup>rd</sup> et al 2005; Pugh-Bernard et al 2001) and if this check point is bypassed B cells progress to plasmatic cells that produce auto-antibodies. Still, a significant fraction of self-reactive BCR fail to be edited or trigger deletion in primary lymphoid tissues, either because the self-antigen are bound with only low avidity or because they are not sufficiently abundant in primary lymphoid organs. For receptors with intermediate avidity for self antigens, the risk they pose for autoimmunity may not overshadow their potential use in fighting infection. B cells with receptors that fall into this zone undergo a conditional type of clonal deletion that is extrinsically regulated through competition with B cells bearing less self reactive BCR (Cyster et al 1994; Lanemo Myhrinder et al 2008). This also can explain that unmutated CLL cases and mutated CLL cases express different antibody repertoires and different VH genes (Fais et al 1988; Johnson et al 1997). Current data support that CLL cells are in active (auto) antigen driven receptor editing, presumably by keeping away from autoreactivity



associated with preferential autoimmune linked IGHV gene utilization in CLL patients like IGHV3-21, IGHV4-34, IGKV1-17 (Foreman et al 2007; Hadzidimitriou et al 2009) and also IGHV5-51 and IGHV1-69 in unmutated IgVH genes (Chapal et al 2000; Vanura et al 2008). Interestingly, highly polyreactive antibodies are expressed frequently by unmutated CLL, but only rarely by mutated cases, supporting the view that the receptor editing mechanism is significantly active to try to elude autoimmunity in CLL.

In mutated CLL cases quite a lot of cellular strategies are used to regulate self-reactive receptors at different points during B cell differentiation.

- |   |
|---|
| <ol style="list-style-type: none"><li>1. The receptor is edited to one that is less self reactive by V(D)J recombination (Hadzidimitriou et al 2009; Rassenti et al 1997 Ghia et al 2008b; Kalinina et al 2011).</li><li>2. Regulation by BCR downregulation and anergy (Muzio et al 2008).</li><li>3. Induction of inhibitory receptors as CD5 by self-reactive BCR (Hippen et al 2000; Morikawa et al 1993; Dallou et al 2008; Hillion et al 2005).</li></ol> |
|---|

Table 1. BCR tolerance mechanisms in central lymphoid organs (bone marrow) include receptor edition, anergy and induction of inhibitory receptors as CD5.

**Regulation of self reactive receptor in follicles**

Each of the checkpoints described above deal with self-reactive receptor generated by V(D)J recombination in the primary lymphoid organs; however, self-reactive BCRs are also generated in a second wave of receptor-gene-diversification through somatic hypermutation in germinal centre follicles of peripheral lymphoid tissues (Shiono et al 2003; Radic et al 1994; Ray et al 1996). Despite somatic hypermutation could produce modifications in BCR to ablate self-reactivity (Murray et al 2008) also might produce new self-reactive BCR. In addition somatic hypermutation poses a particular severe threat of autoimmunity for the reason that increase the affinity of antibodies for self-antigens, the follicular pathway of B cell differentiation generates long lived plasma and memory cells and numerous apoptotic cells be present in germinal centres with self components that are trapped and displayed as immune complexes on follicular dendritic cells. For these reasons the immune system contain a number of mechanisms to elude the maturation of self-reactive B cells that encourage an autoimmune disease. Self-reactivity of mutated CLL cases may derive from immature self-reactive B cells that suffer somatic hypermutation or by non-self reactive B cells that acquire self-reactive BCR during somatic hypermutation in germinal centres. In humans two types of memory B cells have been described: IgM+ memory B cells and class-switched memory B cells (Agematsu et al 1997; Klein et al 1998; Tangye et al 1998). Transition from naive B cells into circulating IgM+ memory B cells is accompanied by efficient counter selection against self reactive naive B cells before the onset of somatic hypermutation and that self reactive IgM+ memory B cells present in the circulation of healthy humans gain self-reactivity as a result of somatic hypermutation (Tsuji et al 2006).

The increase in self-reactivity during transition between mature naive and IgG+ memory B cells might be due to selective advantage for pre-existing self-reactive cells, or selection for cells with self reactive antibodies produced by somatic hypermutation. (Tiller et al 2007) This mechanisms could contribute to generate the IgG+ CLL cases (Ghiotto F, et al 2004).

1. Germinal centre exclusion (Tsuji et al 2006; Cappione A 3 <sup>rd</sup> et al 2005; Pugh-Bernard et al 2001).
2. The receptor is modified to one that is less self reactive by BCR hypermutation (Murray et al 2008, Tiller et al 2007).
3. Receptor edition/revision (Hadzidimitriou et al 2009; Kalinina et al 2011; Rochas et al 2007)
4. CD5 expression (Hillion et al 2005).
6. Absence of T cell help (Shokat et al 1995)
7. Competition for follicular niches (Cyster et al 1994)

Table 2. Tolerance mechanisms in peripheral lymphoid organs.

**Tolerance induced by absence of T-cell help:**

A substantial portion of the activated B cells migrate to germinal centers where they undergo the process of somatic hypermutation. These B cells first remove the BCR from their surface, then undergo several rounds of division, and finally re-express mutated immunoglobulin receptors. The cells then undergo a negative selection process similar to that of transitional B cells. The antigen is provided from antigen-antibody complexes on follicular dendritic cells. Survival requires the receptor to be of high enough affinity to out-compete the already circulating antibody and allow B cell uptake and processing of antigen For display peptides to primed helper T cells, which have also moved into the germinal centers (Kearney et al 1994). If the B cell receives T cell-help it survives and is stimulated to undergo another round of expansion and differentiation. If T cell help is not received, the B cell can become anergized or die by apoptosis (Shokat, et al 1995).

We suggest that in CLL with mutated Ig genes, the proliferating B cells is likely to have traversed a germinal center and acquire “*de novo* self-reactivity” originated in the process of somatic hypermutation mechanism or by receptor editon revision. After this “*de novo* autoreactivity” a normal CD5- B cell can theoretically be transformed into a “*de novo* autoreactive memory B cell” that express CD5+ (increase the threshold for BCR activation), suffer receptor revision (change light chains to evade autoimmunity), down regulate surface Ig (to avoid activation), and remain under check by germinal center exclusion (to diminish the chance to progress in the maturation and become plasma cells that produce autoantibodies). Finally, all this tolerance mechanism converts this B CD5- B cell into an “anergic-edited-CD5+CD27+ memory B cell” excluded from germinal centres. These “*de novo* autoreactive” memory B cells could retain a process of “self-renewal”, a specificity that changes (receptor editing-revision) and/or that can not be activated because this “new malignant cell” is an “anergic cell” excluded from germinal centres. This speculation could

explain why mutated IGVH CLL subsets (“anergic cells”) have an indolent course related to the absence of BCR signalling activation.

IGVH gene usage in CLL is highly selective, and often associated with autoantibody reactivity (Oscier et al 1987). The fact that almost 30 % of CLL patients share BCRs with restricted, quasi-identical immunoglobulins sequences should aid the understanding of the functional interplay between CLL cells and the microenvironment. On the one hand, unmutated IGVH CLL subsets recognizes apoptotic cells in bone marrow and spleen and express a functionally competent BCR, as shown by the fact that most of it can be stimulated following Ig ligation *in vitro*. On the other hand, CLL mutated that has acquired “*de novo*” autoreactivity induced by somatic hypermutation recognizes apoptotic cells in germinal centres; however they become anergic and are unresponsive throughout BCR stimulation. In a CLL mutated subset the “memory-anergic” B cell returns to bone marrow in the same way that normal memory B cells.

#### **Other immunologic alterations that theoretically might predispose the lost of CLL clone control: Impaired immunologic synapses**

CD4 and CD8 T cells of patients with CLL show impaired immunological synapse formation with antigen presenting cells (APC)(Ramsay et al 2008). This dysfunction is in part induced by the CLL B cells. This impaired immunological synapse within T cells and APC could contribute to the failure to mount an effective immune response in patients with CLL. Moreover, it may also add other immunological abnormalities like hipogammaglobulinemia (impaired T cell – B cell interactions), autoimmunity (impaired regulatory T cell control), and second tumours (diminished immunosurveillance mediate by NK and CD8 T cells). Interestingly, lenalidomide, an immunomodulatory drug, could repair this synapses with an enhancement of immune cell function. This effect is clinically observed during treatment of CLL patients with this agent because lenalidomide probably induces a strong activation of the immune system complicated by swelling of involved lymph nodes and fever named tumour flare reaction (Chanan-Khan et al 2006; Aue et al 2009)

#### **Antibody mediated immunoregulation:**

The antigen-antibody complexes are also likely to be responsible for the phenomenon known as original antigenic sin, in which memory B cells, generated during a prior exposure to a cross-reacting antigen, present or down-regulate the response to these unique new determinants on the antigen<sup>70</sup>. Memory B cells seem to have an advantage for rapid activation and this produces antibodies that feed back to inhibit the priming of naïve B cells possessing receptors that are specific to unique determinants of the second immunogen. This feedback mechanism is most likely mediated through antigen-antibody complexes that interact with FcγRIIb on the naïve B cells and inhibit signal transduction through their IgM receptors (Ravetch et al 2001). In patients with hipogammaglobulinemia this feedback mechanism is impaired and might contribute to expansion of autoreactive B cells (García-Muñoz 2009b) , and in patients with CLL it may add an additional risk to uncontrolled proliferation of CLL clones.

**Anti-idiotypic B cell regulation:** In 1974 Jerne proposed that antibody production could be regulated by other antibodies that recognized unique idiotypic determinants in the V regions of the first antibody. He postulated that an increase in the production of the first antibody could negatively regulate the production of anti-idiotypic antibodies, and vice

versa. Because of the interconnected pathways in such a network, perturbation of one segment would be dampened by the presence of others segments and thus the original steady state would be buffered (Jerne 1984; Jerne 1970; Forni et al 1980).

Patients with CLL have an increased proportion of autoimmune haemolytic anemia (AIHA) and idiopathic autoimmune thrombocytopenia (ITP) and infections. It is probable that the idiotypic network is disrupted in CLL patients and that this could lead to an increased risk of autoimmunity on one hand and immunodeficiency on the other. Treatment with intravenous immunoglobulins (IVIg) could in theory restore idiotypic network and antigen-antibody-complexes feedback in CLL B cells. Remarkably, patients with AIHA treated with IVIg experiment a reduction of the size of lymph nodes and spleen (Diehl et al 1998). This suggests that immune-complexes feedback and idiotypic network could contribute indirectly in the control of CLL.

### MYD88 Mutation

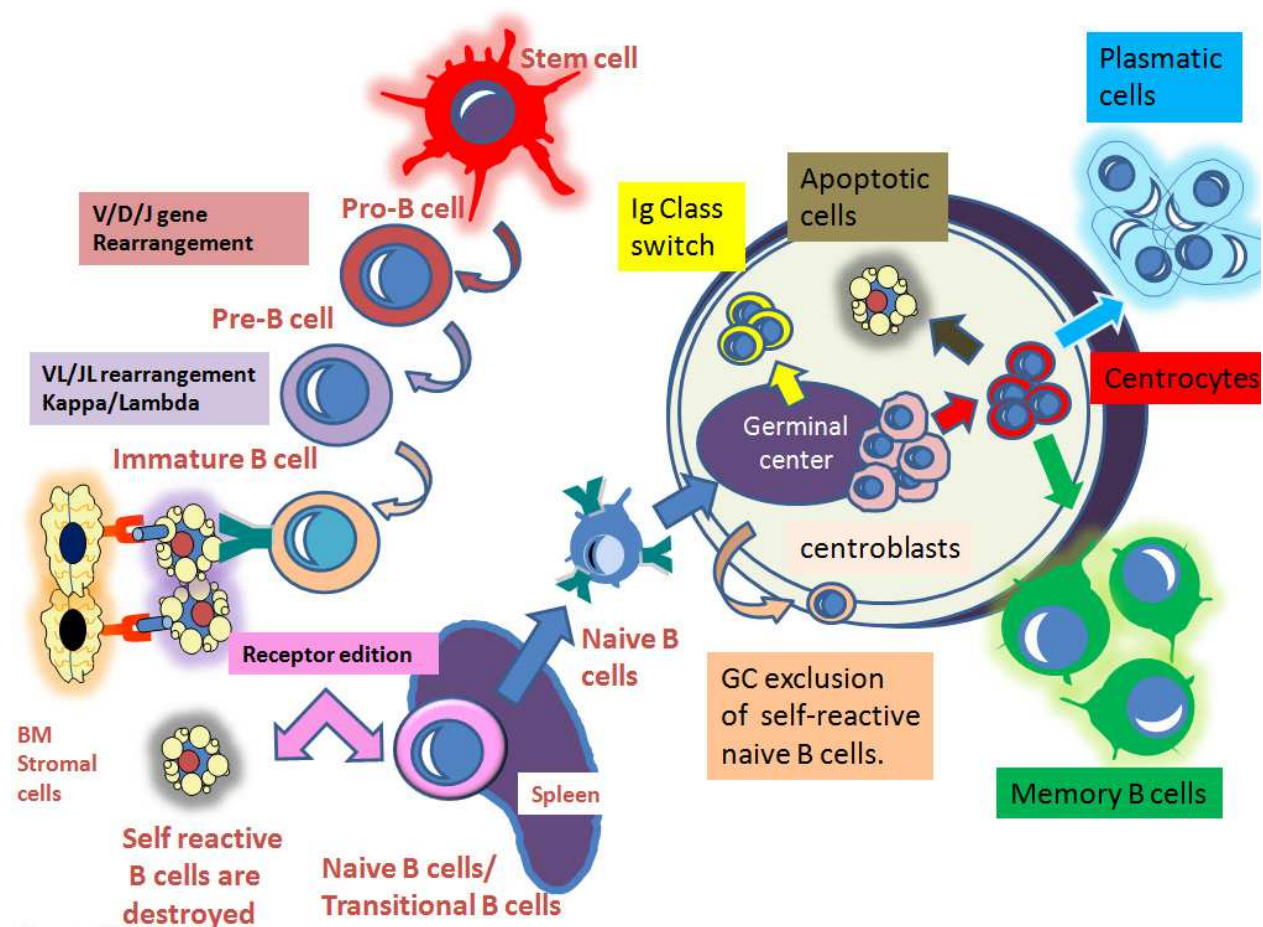
Interestingly, mutations in MYD88 and KLHL6 genes have been reported recently in patients with mutated CLL patients (Puente et al 2011). Significantly, similar to CLL patients, patients with MYD88-deficiency do not secrete autoantibodies (Isnardi et al 2008). We speculate that if mutations in MYD88 gene were acquired during germinal center reaction, is possible that self-reactive B cells cannot progress to plasmatic cells but retain some features or memory B cells. Even more, TLR-9 acts via MYD88 and might induce proliferation of CLL B cells. However, mutations in MYD88 might disturb the function of this TLR-9 and contribute to the biology and better prognosis of mutated CLL cases.

*IGHV* gene usage in CLL is highly selective, and often associated with autoantibody reactivity. The fact that almost 30 % of CLL patients share BCRs with restricted, quasi-identical immunoglobulins sequences should aid to the understanding of the functional interplay between CLL cells and the microenvironment. On the one hand, unmutated *IGHV* CLL subsets recognizes apoptotic cells in bone marrow and spleen and express a functionally competent BCR, as shown by the fact that most of it can be stimulated following Ig ligation *in vitro*. On the other hand, CLL mutated that has acquired “*de novo*” autoreactivity (¿mutations in MYD88?) induced by somatic hypermutation recognizes apoptotic cells in germinal centres; however they become anergic and are unresponsive throughout BCR stimulation. In a CLL mutated subset the “memory-anergic” B cell returns to bone marrow in the same way that normal memory B cells.

### 3. Conclusion

Chronic lymphocytic leukemia can be separated into cases that harbour somatic mutation in their IGVH genes, or cases without somatic mutations. IGVH gene usage in CLL is highly selective, and often associated with autoantibody reactivity. Despite the fact that the cell surface markers and gene expression of CLL cells suggest that both subsets originate from a precursor cell of the same developmental stage, these findings could be only the result of several immunologic mechanisms that try to destroy or avoid the persistence of self-reactive CLL B cells. CLL is characterized by multiple immune deficiencies and autoimmune phenomena associated with persistent tolerance mechanism trying to control self-reactive CLL B cells growth.

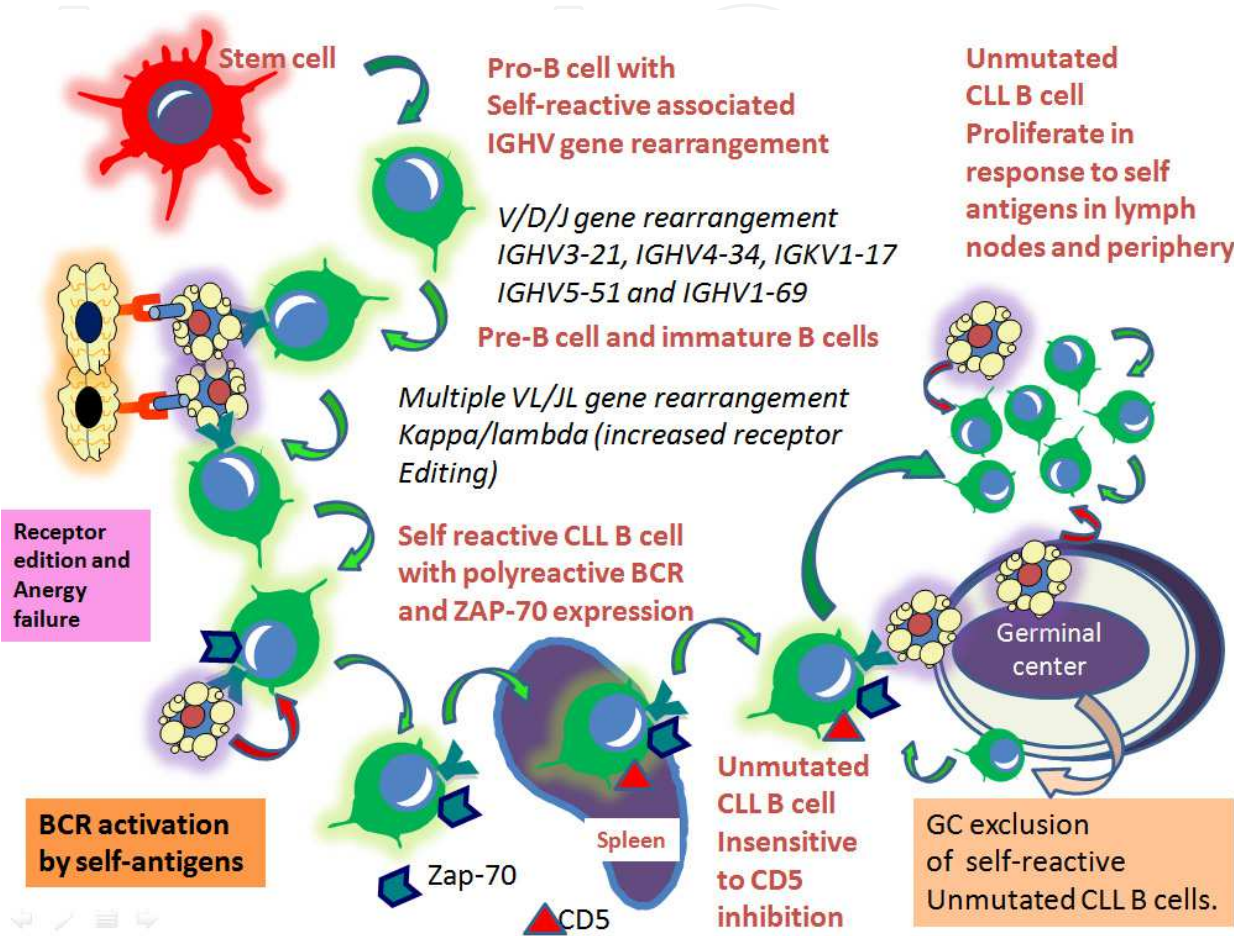




B-cell development occurs initially in the bone marrow and subsequently in lymphoid organs. In bone marrow, hematopoietic progenitor cells (HSC) differentiate into the earliest identifiable cell type committed to the B-cell lineage, the pro-B cell. The pro-B cell undergoes a rearrangement of its immunoglobulin (Ig) heavy chain genes and is called a pre-B cell. Subsequent rearrangement of the light chain enables the cell to express surface IgM and the cell becomes an immature transitional B lymphocyte. These cells leave the bone marrow and are called naïve B cells. They are arrested in the G0 phase of the cell cycle. These naïve B cells enter the lymphoid tissue, where they are exposed to antigen-presenting cells, become activated and differentiate into plasma cells or memory B cells. Through activation by an antigen, B cells differentiate into centroblasts, resulting in Ig isotype switching and somatic mutations in the variable region of the Ig with the generation of high-affinity antibodies. Centroblasts then progress to the centrocyte stage and re-express surface Ig. The centrocytes with high-affinity antibodies differentiate into either memory B cells or plasmablasts, which subsequently move to the bone marrow and terminally differentiate into plasma cells.

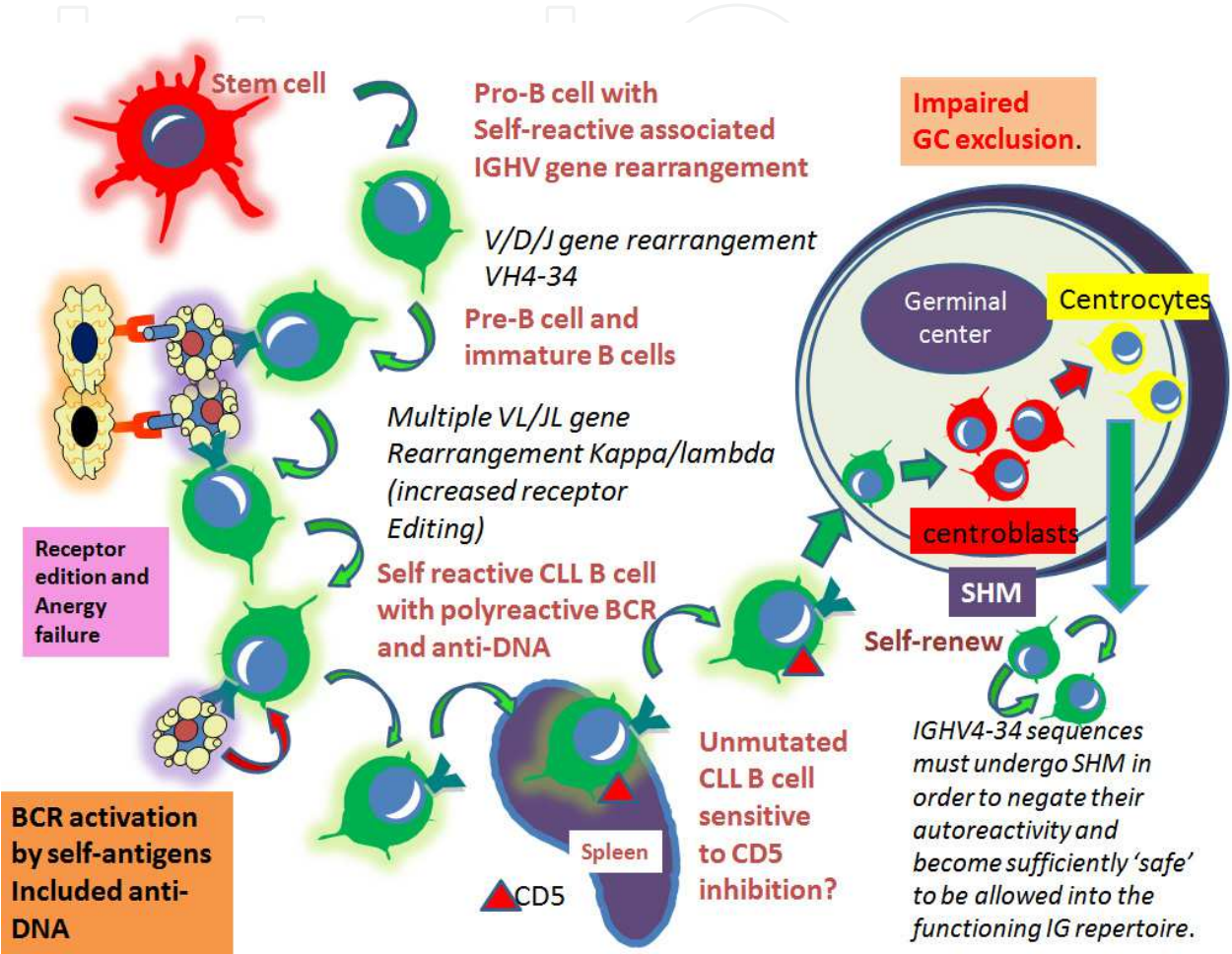
Fig. 1. Normal B cell development.





Unmutate CLL B-cell development occurs initially in the bone marrow and subsequently in lymphoid organs. In bone marrow, hematopoietic progenitor cells (HSC) differentiate into the pro-B cell that use IGHV genes related with autoimmunity. The pro-B cell undergoes a rearrangement of its immunoglobulin (Ig) heavy chain genes and is called a pre-B cell. Subsequent rearrangement of the light chain enables the cell to express surface self reactive BCR that fail to be corrected by several rounds of receptor edition. This self-reactive B cells acquire Zap-70 or other alterations that induce increased BCR activation. This is the way in which this self-reactive CLL B cells pass up tolerance mechanisms as anergy and inhibition exerted by CD5. These cells leave the bone marrow as unmutated polyreactive CLL B cells. These unmutated polyreactive CLL B cells enter in the lymphoid tissue, where they are exposed to antigen-presenting cells and self-antigens, however, they cannot be converted into plasma cells or memory B cells with mutations because they are efficiently excluded by germinal centers.

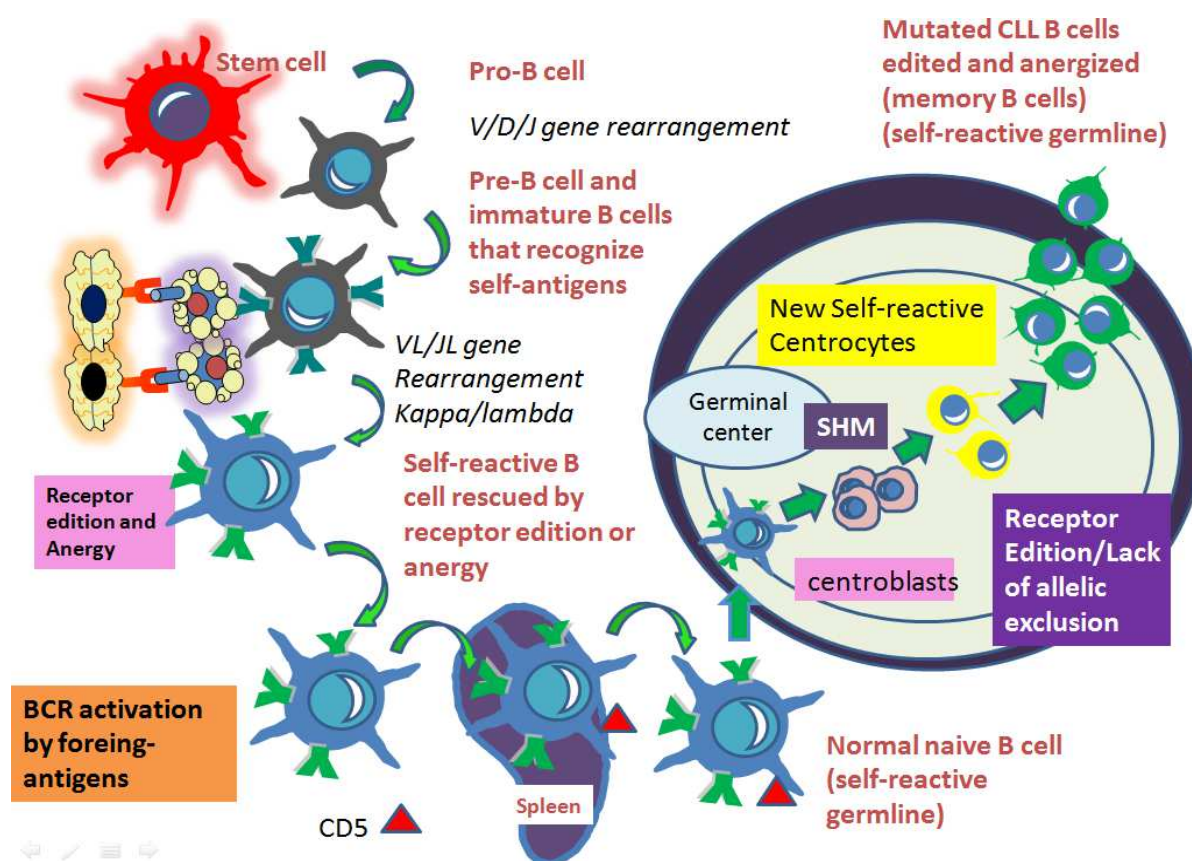
Fig. 2. Hypothesis about generation of unmutated B cells (García-Muñoz et al. Ann Hematol. Accepted).



Mutated CLL B-cell development occurs initially in the bone marrow and subsequently in lymphoid organs. In bone marrow, hematopoietic progenitor cells (HSC) differentiate into the pro-B cell that use IGHV genes related with autoimmunity. The pro-B cell undergoes a rearrangement of its immunoglobulin (Ig) heavy chain genes and is called a pre-B cell. Subsequent rearrangement of the light chain enables the cell to express surface self reactive BCR that fail to be corrected by several rounds of receptor edition. This self-reactive B cells enter in germinal centres and undergo somatic hypermutation in order to negate their autoreactivity. This is the way in which this self-reactive CLL B cells pass up tolerance mechanisms as germinal centre exclusion, however, fortunately they suffer some mutations to reverse their self reactivity and avoid autoimmune diseases as SLE. These cells leave the germinal center as mutated CLL B cells memory like cells.

Fig. 3. "Impaired Germinal Centre exclusion model for development of mutated CLL cases wiht VH4-34.





Mutated CLL B-cell development occurs initially in the bone marrow and subsequently in lymphoid organs. In bone marrow, hematopoietic progenitor cells (HSC) differentiate into the pro-B cell that use IGHV genes related with autoimmunity. The pro-B cell undergoes a rearrangement of its immunoglobulin (Ig) heavy chain genes and is called a pre-B cell. Subsequent rearrangement of the light chain enables the cell to express surface self reactive BCR that succeed to be corrected by several rounds of receptor edition. This ex-self-reactive B cells acquire CD5 or other alterations that induce lesser BCR activation. This is the way in which this ex-self-reactive CLL B cells suffer tolerance mechanisms as receptor edition, anergy and inhibition exerted by CD5. These cells leave the bone marrow as unmutated normal naïve B cells. These naïve ex-self reactive B cells enter in germinal centres and suffer somatic hypermutation (SHM) and acquire a new self-reactive BCR, however, again tolerance mechanisms as receptor edition/revision and CD5 expression make this cells in an anergic memory ex-self-reactive B cells. Importantly, reversion of the IGHV mutated sequences to germline counterparts restored the polyreactivity and self-reactivity.

Fig. 4. Mutated CLL B cells generated by somatic hypermutation (García Muñoz et al. Ann Hematol. Accepted).

#### 4. Acknowledgment

The authors declare that a review paper on immunological aspects in CLL is actually accepted in Ann of Hematology.

#### 5. References

Agematsu K, Nagumo H, Yan FC et al. (1997) B cell subpopulations separated by CD27 and crucial collaboration of CD27<sup>+</sup> B cells and helper T cells in immunoglobulin production. Eur J Immunol 1997;27:2073-2079

- Aqeilan RI, Calin GA, Croce CM, et al. (2010) miR-15 and miR-16-1 in cancer: discovery, function and future perspectives. *Cell Death Differ* 2010;17:215-20.
- Aue G, Njuguna N, Tian X, Soto S, Huges T, Vire B, et al. (2009) Lenalidomide-induced upregulation of CD80 on tumor cells correlates with T cell activation, the rapid onset of a cytokine release syndrome and leukemic cell clearance in chronic lymphocytic leukemia. *Haematologica*. 2009;94:1266-73.
- Bernal A, Pastore RD, Asgary Z, et al. (2001) Survival of leukemic B cells promoted by engagement of the antigen receptor. *Blood* 2001;98:3050-7
- Berland R, Wortis HH. (2002) Origins and functions of B-1 cells with notes on the role of CD5 *Annu Rev Immunol* 2002;20:253-300
- Benschop RJ, Aviszus K, Zhang X et al. (2001) Activation and anergy in bone marrow B cells of a novel immunoglobulin transgenic mouse that is both hapten specific and autorreactive. *Immunity* 2001;14:33-43.
- Binder M, Le'chenne B, Ummanni R, Scharf C, Balabanov S, et al. (2010) Stereotypical Chronic Lymphocytic Leukemia B-Cell Receptors Recognize Survival Promoting Antigens on Stromal Cells. *PLoS ONE* 2010;5: e15992. doi:10.1371
- Binder M, Muller F, Jackst A, Léchenne B, Pantic M, Bacher U, et al. B-cell receptor epitope recognition correlates with the clinical course of chronic lymphocytic leukemia *Cancer* 2011;117:1891-1900
- Broker BM, Klajman A, Youinou P, et al. (1988) Chronic lymphocytic leukemia (CLL) cells secrete multispecific autoantibodies. *J Autoimmune* 1988;1:469-481
- Burnet M. (1972) Pathogenesis of auto-immune disease. In *Auto-immunity and auto-immune disease* (1972). Medical and Technical Publishing CO LTD. 173-290
- Caligaris-Cappio F. B chronic lymphocytic leukemia: a malignancy of anti-self B cells. *Blood* 1996;87:2615-2620
- Callin GA, Liu CG, Shimizu M, et al (2004) MicroRNA profiling reveals distinct signatures in B cell chronic lymphocytic leukemia. *Proc Natl Acad Sci USA* 2004;101:11755-60
- Cappione A 3rd, Anolik JH, Pugh-Bernard A, Barnard J, Dutcher P, Silverman G, Sanz I. Germinal center exclusion of autoreactive B cells is defective in human systemic lupus erythematosus. *J Clin Invest*. 2005;115:3205-3216
- Cerutti A, Zan H, Shan S, Schattner E, et al. Ongoing in vivo class switch DNA recombination in chronic lymphocytic leukemia B cells. *J Immunol* 2002;169:6594-603
- Chanan-Khan A, Miller KC, Musial L, Lawrence D, Padmanabhan S, Tajeshita K, et al. (2006) Clinical efficacy of lenalidomide in patients with relapsed or refractory chronic lymphocytic leukemia: results of a phase II study. *J Clin Oncol*. 2006;24:5343-9
- Chapal N, Peraldi-Roux S, Bresson D, et al. (2000) Human anti-thyroid peroxidase single-chain fragment variable of Ig isolated from a combinatorial library assembled in cell: insights into the In Vivo Situation. *J Immunol* 2000;114:4162-4169.
- Chen L, Apgar J, Huynh L, et al. (2005) Zap-70 directly enhances IgM signaling in chronic lymphocytic leukemia. *Blood* 2005;105:2036-41.
- Chen L, Withopf G, Huynh L, et al. (2002) Expression of ZAP-70 is associated with increased B-cell receptor signaling in chronic lymphocytic leukemia. *Blood*. 2002;100:4609-46-14
- Chiorazzi N, Ferrarini M. (2003) B cell CLL: lessons learned from studies of the B-cell antigen receptor. *Ann Rev Immunol*. 2003;21:841-94.

- Chiorazzi N, Ferrarini M.(2011) Cellular origin(s) of chronic lymphocytic leukemia: cautionary notes and additional considerations and possibilities. *Blood*.2011;117:1781-1791.
- Chiorazzi N, Hatzi K, Albesiano E (2005) B-cell chronic lymphocytic leukemia, a clonal disease of B lymphocytes with receptors that vary in specificity for (auto)antigens. *Ann N Y Acad Sci* 2005;1062: 1-12.
- Chiorazzi N, Rai KR, Ferrarini M. (2005) Chronic lymphocytic leukemia. *N Engl J Med* 2005;352:804-815
- Chu CC, CATERA R, Zhang L, Didier S, Agagnina BM, Damle RN, et al. (2010) Many chronic lymphocytic leukemia antibodies recognize apoptotic cells with exposed nonmuscle myosin heavy chain IIA: implications for patient outcome and cell of origin. *Blood*. 2010;115:3907-3915.
- Cimmino A, Callin GA, Fabbri M, et al. (2005) miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci* 2005;102:13944-9
- Crespo M, Bosch F, Villamor N, et al. (2003) Zap-70 expression as a surrogate for IgV-region mutations in CLL. *N Engl J Med* 2003;348:1764-1775.
- Cyster JG, Hartley SB, Goodnow CC.(1994) Competition for follicular niches excludes self-reactive cells from the recirculating B cell repertoire. *Nature* 1994;371:389-395.
- Dallou A. (2008) CD5: a safeguard against autoimmunity and a shield for cancer cells. *Autoimmun Rev* 2008;8:349-353
- Damle RN, Wasil T, Fais et al,(1999) IGVH gene mutation status and CD38 expression as novel prognostic indicators in CLL. *Blood* 1999;94:1840-1847.
- Diehl LE, Ketchum LH (1998): Autoimmune disease and chronic lymphocytic leukemia: Autoimmune haemolytic anemia, pure red cell aplasia and autoimmune thrombocytopenia. *Semin Hematol* 25:80-97,1998
- Dighiero G, Hart S, Lim A, Borche L, Levy R, et al. (1991) Autoantibody activity of immunoglobulins isolated from B-cell follicular lymphomas. *Blood* 1991; 78:581-585.
- Döhner H, Fischer K, Bentz M, et al.(1995) P53 gene deletion predicts for poor survival and non response to therapy with purine analogs in chronic B-cell leukaemias. *Blood* 1995;85:1580-9
- Döhner H, Stilgenbauer S, Benner A, et al.(2000) Genomic aberrations and survival in CLL. *N Engl J Med*. 2000;343:1910-1916.
- Döhner H, Stilgenbauer S, James MR, et al. (1997) 11q deletions identify a new subset of B-cell chronic lymphocytic leukemia characterized by extensive nodal involvement and inferior prognosis. *Blood* 1997;89:2516-22.
- Dolmetsh RE, Lewis RS, Goodnow CC, et al. (1997) Differential activation of transcription factors induced by Ca<sup>2+</sup> response amplitude and duration. *Nature* 1997;386:855-858
- Fais F, et al (1998) Chronic lymphocytic leukemia B cells express restricted sets of mutated and unmutated antigen receptors. *J Clin Invest* 1998;102:1515-1525.
- Forni L, Coutinho A, Köhler G, Jerne NK.(1980) IgM antibodies induce the production of antibodies of the same specificity. *Proc Natl Acad Sci USA* 1980;77:1125-8
- Foreman AL, van de Water J, Gougeon ML, Gershwin ME. (2007) B cells in autoimmune diseases: insights from analyses of immunoglobulin variable (Ig V) gene usage. *Autoimmun Rev*.2007;6:387-401



- Fuda FS, Karandikar NJ, Chen W. (2009) Significant CD5 expression on normal stage 3 hematogones and mature B lymphocytes in bone marrow. *Am J Clin Pathol* 2009; 132: 733-737
- García-Muñoz R. (2009a) Overall reduction in antibody production could contribute to generate pathogenic autoantibodies and autoimmune manifestations. *Clin Rheumatol* 2009;28:361-3.
- García-Muñoz R, Panizo C, Bendandi M, Llorente L. (2009b). Autoimmunity and lymphoma: is mantle cell lymphoma a mistake of receptor editing mechanism? *Leuk Res* 2009;11:1437-1439.
- García-Muñoz R, Galiacho VR, Llorente L. (2012). Immunological aspects in chronic lymphocytic leukemia (CLL) development. *Ann Hematol.* (in press)
- Gary-Gouy H, Sainz-Perez A, et al. (2007) Natural Phosphorylation of CD5 in chronic lymphocytic leukemia B cells and Analysis of CD5 regulated genes in a B cell line suggest a role of CD5 in malignant phenotype. *J Immunol* 2007;179:4335-4344
- Gary-Gouy H, Harriague J, Bismuth G, Platzer C, Schmitt C, Dallou AH. (2002a) Human CD5+ promotes B cell survival through stimulation of autocrine IL-10 production. *Blood* 2002;100:4537-4543
- Gary-Gouy H, Harriague J, Dallou A, Donnadieu E, Bismuth G. (2002b) CD5-negative regulation of B cell receptor signalling pathways originates from tyrosine residue Y429 outside an immunoreceptor tyrosine-based inhibitory motif. *J Immunol* 2002;168:232-239
- Gauld SB, Benschop RJ, Merrel KT, et al. (2005) Maintenance of B cell anergy requires constant antigen receptor occupancy and signaling. *Nature Immunol* 2005;6:1160-1167).
- Gauld SB, Merrell KT, Cambier JC. (2006) Silencing of autoreactive B cells by anergy; a fresh perspective. *Curr Opin Immunol* 2006;18:292-297.
- Getahun A, O'Neil SK, Cambier JC. (2009) Establishing anergy as a Bona Fide in vivo mechanisms of B cell tolerance. *J Immunol* 2009;5430-5441
- Ghia EM, Jain F, Widhopf II GF, et al. (2008b) Use of IGHV3-21 in chronic lymphocytic leukemia is associated with high risk disease and reflects antigen-driven, post-germinal center leukemogenic selection. *Blood* 2008;111:5101-5108.
- Ghia P, Chiorazzi N, Stamatopoulos K. (2008a) Microenvironmental influences in chronic lymphocytic leukemia: the role of antigen stimulation. *J Internal Med.* 2008;264:549-62.
- Ghia P, Stamatopoulos K, Belessi C, et al. (2005) Geographic patterns and pathogenetic implications of IGHV gene usage in chronic lymphocytic leukemia: the lesson of IGHV3-21 gene. *Blood* 2005;105:1678-1685.
- Ghiotto F, Fais F, Valletto A, et al. (2004) Remarkably similar antigen receptors among a subset of patients with chronic lymphocytic leukemia, *J Clin Invest* 2004;113:1008-1016.
- Goodnow CC. et al. (2005) Self tolerance checkpoints in B cell lymphocyte development. *Adv. Immunol* 2005;58:279-368.
- Goodnow CC, Crosbie J, Adelstein S, et al. (1998) Altered immunoglobulin expression and functional silencing of self reactive B lymphocytes in transgenic mice. *Nature* 1998;334:676-682.
- Goodnow CC, Crosbie J, Jorgensen H, et al. (1989) Induction of self tolerance in mature peripheral B lymphocytes *Nature* 1989;342:385-391

- Griffin DO, Holodick NE, Rothstein TL.(2011) Human B1 cells in umbilical cord and adult peripheral blood express the novel phenotype CD20+CD27+CD43+CD70-. *J Exp Med* 2011;208:67-80.
- Guarini A, Chiaretti S, Tavoraro S, et al. BCR ligation induced by IgM stimulation results in gene expression and functional changes in only in IgVH unmutated chronic lymphocytic leukemia (CLL) cells. *Blood* 2008;112:782-792.
- Hallek M, Cheson D, Catovsky D, Caligaris-Cappio F, Dighiero G, Döner H, Hillmen P, Keating MJ, Montserrat E, Rai KR, Kipps TJ. (2008) Guidelines for the diagnosis and treatment of chronic lymphocytic leukemia: a report from the international Workshop on Chronic Lymphocytic Leukemia updating the National Cancer Institute-Working Group 1996 guidelines. *Blood*. 2008;111:5446-5456.
- Hadzidimitriou A, Darzentas N, Murray F, et al. (2009) Evidence for the significant role of immunoglobulin light chains in antigen recognition and selection in chronic lymphocytic leukemia. *Blood* 2009;113:403-411.
- Hamblin TJ, Davis Z, Gardiner A et al. (1999) Unmutated IGVH genes are associated with a more aggressive form of CLL. *Blood* 1999;94:1848-1854.
- Hamblin TJ, Orchard JA, Ibbotson RE et al. (2002) CD38 expression and immunoglobulin variable region mutations are independent prognostic variables in chronic lymphocytic leukemias, but CD38 may vary during the course of the disease. *Blood* 2002;99:1023-1029.
- Herve M, Xu K, Ng YS, et al. (2005) Unmutated and mutated chronic lymphocytic leukemias derive from self reactive B cell precursors despite expressing different antibody reactivity. *J Clin Invest* 2005;115:1636-1643
- Hillion S, Saraux A, Youinou P, Jamin C. Expression of RAGs in peripheral B cells outside germinal centers is associated with the expression of CD5. *J Immunol* 2005;174:5553-61
- Hippen KL, Tze LE, Behrens T. (2000) CD5 maintains tolerance in anergic B cells. *J Exp Med* 2000;191:883-889
- Isnardi I, Ng YS, Srdanovic I, Motaghedi R, Rudchenko S, von Bernut H, et al. (2008) IRAK-4/MyD88 dependent pathways are essential for the removal of developing of autoreactive B cells in humans. *Immunity* 2008;29:746-757
- Jerne NK. Idiotypic networks and other preconceived ideas. (1984) *Immunol Rev*. 1984;79:5-24
- Jerne NK. (1974) Towards a network theory of the immune system. *Ann Immunol (Paris)* 1974;125C:373-89
- Johnson TA, Rassenti LZ, Kipps TJ (1997) Ig VH1 genes expressed in B cell chronic lymphocytic leukemia exhibit distinctive molecular features. *J Immunol* 1997;158:235-246.
- Kalinina O, Doyle-Cooper, Miksanek J et al. Alternative mechanisms of receptor editing in autoreactive B cells. *PNAS* 2011;108:7125-7130
- Kearney ER, Pape KA, Loh DY, et al. Visualization of peptide-specific T cell immunity and peripheral tolerance induction in vivo. *Immunity*. 1994;1:327-339
- Kikushige Y, Ishikawa F, Miyamoto T, et al (2011) Self-renewing hematopoietic stem cell is the primary target in pathogenesis of human chronic lymphocytic leukemia. *Cancer Cell*. 2011;20:246-59
- Klein U, Dalla-Favera R. (2005) New insights into the phenotype cell derivation of B cells in chronic lymphocytic leukemia. *Curr Top Microbiol Immunol* 2005;294:31-49

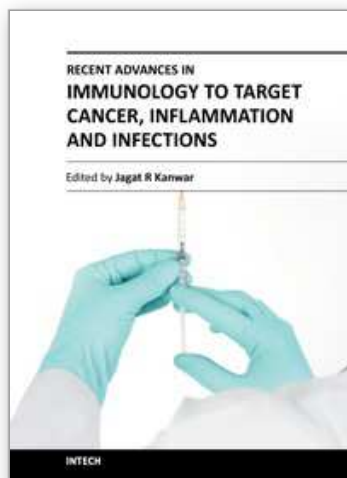
- Klein U, Rajewsky K, Kuppers R. (1998) Human immunoglobulin (Ig)M+IgD+ peripheral blood expressing CD27 cell surface antigen carry somatically mutated variable region genes: CD27 as a general marker for somatically mutated (memory) B cells. *J Exp Med* 1998;188:1679-1689
- Klein U, Tu Y, Stolovitzky GA, et al. (2001) Gene expression profiling of B cell chronic lymphocytic leukemia reveals a homogeneous phenotype related to memory B cells. *J Exp Med* 2001;194:1625-1638.
- Krober A, Seiler T, Benner A et al. (2002) V(H) mutation status, CD38 expression level, genomic aberrations and prognosis in CLL. *Blood* 2002;100:1410-1416.
- Krober A, Bloehdorn J, Hafner S, et al. (2006) Additional genetic high risk features such as 11q deletion, 17p deletion and V3-21 usage characterize discordance of ZAP-70 and VH mutation status in CLL. *J Clin Oncol.* 2006;24:969-975.
- Lanemo Myhrinder A, Hellqvist E, Sidorova E, et al. A new perspective: molecular motifs on oxidized LDL, apoptotic cells, and bacteria are targets for CLL antibodies. *Blood.* 2008;111:3838-3848
- Lanham S, Hamblin T, Oscier D, et al. Differential signaling via surface IgM is associated with VH gene mutational status and CD38 expression in chronic lymphocytic leukemia. *Blood* 2003;101:1087-1093
- Lee J, Kuchen S, Ficher R, et al. (2009) Identification and characterization of circulating human CD5+ pre-naïve B cell population. *J Immunol* 2009;182:4116-4126.
- Luning Prak E, Monestier M, Eisenberg RA. (2011) B cell receptor editing in tolerance and autoimmunity. *Ann NY Acad Sci* 2011;1217:96-121.
- Matutes E, Wrotherspoon A, Catovsky D. (2007) Differential diagnosis in chronic lymphocytic leukemia. *Best Pract Res Clin Haematol* 2007;20:367-84
- Mertens D, Wolf S, Tschuch C et al. (2006) Allelic silencing at the tumor-suppressor locus 13q14.3 suggest an epigenetic tumor-suppressor mechanism. *Proc Natl Acad Sci USA* 2006;103:7741-6
- Mertens D, Bullinger L, Stilgenbauer S. (2011). Chronic lymphocytic leukemia –genomics lead the way. *Haematologica* 2011;96:1402-1405
- Mockridge CI, Potter KN, Wheatley I, Neville LA, Packham G, Stevenson FK. (2007) Reversible anergy of sIgM-mediated signalling in the two subsets of CLL defined by VH-gene mutational status. *Blood* 2007;109:4424-4431.
- Morbach H, Singh S K, Faber C, Lipsky P E, Girschick H J. (2006) Analysis of RAG expression by peripheral blood CD5+ and CD5- B cells in patients with childhood systemic lupus erythematosus *Annals of the Rheumatic Diseases* 2006;65:482-487
- Morikawa K, Oseko F, Morikawa S. (1993) Induction of CD5 antigen on human CD5- B cells by stimulation with staphylococcus Aureus Cowan strain. *Int. Immunol* 1993;5:809-816.
- Murray F, Darzentas N, Hadzidimitriou A, et al.(2008) Stereotyped patterns of somatic hypermutation in subsets of patients with chronic lymphocytic leukemia: implications for the role of antigen selection in leukemogenesis. *Blood* 2008;111:1524-1533
- Muzio M, Apollino B, Scielzo C, et al.(2008) Constitutive activation of distinct BCR-signaling pathways in a subset of CLL patients: a molecular signature of anergy. *Blood* 2008;112:188-195.

- Nakajima PB, Kieffer K, Price A et al. (2009) Two distinct populations of H chain edited B cells show differential surrogate L chain dependence J Immunol. 2009;182:3583-3596.
- Nemazee DA, Weigert MG.(2000) Revising B cell receptors. J Exp Med 2000;191:1813-1818.
- Oscier DG, Thompson A, Zhu D, Stevenson FK. (1987) Differential rates of somatic hypermutation in V(H) genes among subsets of chronic lymphocytic leukemia defined by chromosomal abnormalities. Blood 1987;69:4153-4160
- Perez-Chacon G, Vargas JA, Jorda J, et al.(2007a) CD5 does not regulate the signalling triggered through BCR in B cells from a subset of B-CLL patients. Leuk Lymphoma 2007;48:147-157.
- Perez-Chacon G, Vargas JA, Jorda J, et al.(2007b) CD5 provides viability signals to B cells from a subset of B-CLL patients by a mechanism that involves PKC. Leuk Res 2007;31:183-193
- Petlickovski A, Laurenti L, Li X, et al. Sustained signaling through the B-cell receptor induce Mcl-1 and promotes survival of chronic lymphocytic leukemia B cells. Blood. 2005;105:4820-4827.
- Puente XS, Pinyol M, Quesada V, Conde L, Ordoñez GR, Villamor N, et al. (2011) Whole genome sequencing identifies recurrent mutations in Chronic lymphocytic leukemia. Nature 2011
- Pugh-Bernard, A.E. et al. (2001) Regulation of inherently autoreactive VH4-34 B cells in the maintenance of human B cell tolerance. J Clin Invest. 2001;108:1061-1070.
- Rassenti LZ, Kipps TJ. (1997) Lack of allelic exclusion in B cell Chronic lymphocytic leukemia. J Exp Med 1997;185:1435-1445
- Radic MZ, Erickson J, Litwin S, et al. (1993) Lymphocytes may escape tolerance by revising their antigen receptor. J Exp Med. 1993;177:1165-1163.
- Radic MZ, Wigert M.(1994) Genetic and structural evidence for antigen selection of anti-DNA antibodies. Ann Rev Immunol 1994;12:487-520
- Ramsay AG,Johnson AJ, Lee M, Gorgun G, Le Dieur R, Blum W, et al. (2008) Chronic lymphocytic leukemia T cells show impaired immunological synapse formation that can be reversed with an immunomodulating drug. J Clin Invest 2008;118:2427-37
- Ravetch JV, Bolland S. IgG Fc Receptors. Annu Rev Immunol. 2001;19:275-290
- Ray SK, Putterman C, Diamond B. Pathogenic autoantibodies are routinely generated during response to foreign antigen: a paradigm for autoimmune disease. Proc Natl Acad Sci. USA 1996;93:2019-2024.
- Rochas C, Hillion S, Youinou P et al. (2007) RAG mediated secondary rearrangements of B cell antigen receptors in rheumatoid synovial tissue. Autoimmun Rev 2007;7:155-9
- Rosenwald A, Alizadeh AA, Widhopf G, et al. (2001) Relation of gene expression phenotype to immunoglobulin mutation genotype in B cell chronic lymphocytic leukemia. J Exp Med 2001;194:1639-1647
- Roudier J, Solveman GJ, Chenn PP et al.(1990) Intracloal diversity in the VH genes expressed by CD5-chronic lymphocytic leukemia producing pathologic IgM rheumatoid factor. J Immunol 1990;144:1526-1530
- Rozkova D, Novodna L, Pytlic R, e tal. (2010) Toll like receptors on B cells: Expression and functional consequences of their stimulation. Int J Cancer 2010;126:1132-43



- Ruzickova S, Pruss A, Odendahl M, et al. Chronic lymphocytic leukemia preceded by cold agglutinin disease: intraclonal immunoglobulin light chain diversity in VH4-34 expressing single leukemic B cells. *Blood* 2002;100:3419-3422.
- Sandel PC, Monroe JG. (1999) Negative selection of immature B cells by receptor editing or deletion is determined by site of antigen encounter, *Immunity*. 1999;10:289-299
- Shiono H, et al. (2003) Scenarios of autoimmunization of T and B cells in myasthenia gravis. *Ann NY Acad Sci* 2003;998:237-256.
- Shokat KM, Goodnow CC. Antigen induced B cell death and elimination during germinal centre immune responses. *Nature* 1995;375:334-338.
- Stamatopoulos K, Kosmas C, Stavroyianni N, et al (1996) Evidence of immunoglobulin heavy chain variable region gene replacement in a patient with B cell chronic lymphocytic leukemia. *Leukemia* 1996;10:1551-1556.
- Stevenson FK and Caligaris-Cappio F. Chronic lymphocytic leukemia: revelations from the B cell receptor. *Blood* 2004;103:4389-4395.
- Sthoeger ZM, Wakai M, Tse DB, Vinciguerra VP, Allen SL, et al. (1989) Production of autoantibodies by CD5-expressing B lymphocytes from patients with chronic lymphocytic leukemia. *J Exp Med* 1989;169: 255-268.
- Tangye SG, Liu YJ, Aversa G, et al. (1998) Identification of functional human splenic memory B cells by expression of CD148 and CD27. *J Exp Med* 1998;169:1703
- Thumberg U, Johnson A, Roos G et al.(2001) CD38 is a poor predictor for VH gene mutational status and prognosis in chronic lymphocytic leukemia. *Blood* 2001;97:1892-1894
- Tiegs SL, Russel DM, Nemazee D.(1993) Receptor editing in self-reactive bone marrow B cells. *J Exp Med* 1993;177:1009-1020.
- Tobbin G, Thunberg U, Johnson A, et al. (2002) Somatic mutated Ig V(H)3-21 genes characterize a new subset of chronic lymphocytic leukemia. *Blood* 2002;99:2262-4.
- Tiller T, Tsuiji M, Yurasov S, et al. (2007) Autoreactivity in Human IgG+ Memory B cells. *Immunity* 2007;26:205-213
- Tsuiji M, Yurasov S, Velinzon K, Thomas S, Nussenzweig MC, Wardemann H.(2006) A check point for autoreactivity in human IgM memory B cell development. *J Exp Med* 2006;203:393-400.
- Van Lochem EG, van der Valden VHJ, Wind HK, et al. (2004) Immunophenotypic differentiation patterns of normal hematopoiesis in human bone marrow: Reference patterns for age-related changes and disease induced shifts. *Cytometry B Clin Cytom* 2004;60:1-13
- Vanura K, Le T, Esterbauer H, et al. (2008) Autoimmune conditions and chronic infections in chronic lymphocytic leukemia patients at diagnosis are associated with unmutated IgVH genes. *Haematologica* 2008;93:1912-16.
- Wiestner A, Rosengwald A, Barry TS, et al.(2003) Zap-70 expression identifies a chronic lymphocytic leukemia subtype with unmutated immunoglobulin genes, inferior clinical outcome, and distinct gene expression profile. *Blood* 2003;101:4944-4951
- Yurasov S, Hammersen J, Tiller T, et al. (2005) B cell tolerance check points in healthy humans and patients with systemic lupus erythematosus. *Ann N Y Acad Sci*. 2005;1062:165-174.





## **Recent Advances in Immunology to Target Cancer, Inflammation and Infections**

Edited by Dr. Jagat Kanwar

ISBN 978-953-51-0592-3

Hard cover, 520 pages

**Publisher** InTech

**Published online** 09, May, 2012

**Published in print edition** May, 2012

Immunology is the branch of biomedical sciences to study of the immune system physiology both in healthy and diseased states. Some aspects of autoimmunity draws our attention to the fact that it is not always associated with pathology. For instance, autoimmune reactions are highly useful in clearing off the excess, unwanted or aged tissues from the body. Also, generation of autoimmunity occurs after the exposure to the non-self antigen that is structurally similar to the self, aided by the stimulatory molecules like the cytokines. Thus, a narrow margin differentiates immunity from auto-immunity as already discussed. Hence, finding answers for how the physiologic immunity turns to pathologic autoimmunity always remains a question of intense interest. However, this margin could be cut down only if the physiology of the immune system is better understood. The individual chapters included in this book will cover all the possible aspects of immunology and pathologies associated with it. The authors have taken strenuous effort in elaborating the concepts that are lucid and will be of reader's interest.

### **How to reference**

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Ricardo García-Muñoz, Judit Anton-Remirez, Jesus Feliu, María Pilar Rabasa, Carlos Panizo and Luis Llorente (2012). Is Chronic Lymphocytic Leukemia a Mistake of Tolerance Mechanisms?, Recent Advances in Immunology to Target Cancer, Inflammation and Infections, Dr. Jagat Kanwar (Ed.), ISBN: 978-953-51-0592-3, InTech, Available from: <http://www.intechopen.com/books/recent-advances-in-immunology-to-target-cancer-inflammation-and-infections/is-chronic-lymphocytic-leukemia-a-mistake-of-tolerance-mechanisms>

**INTeCH**  
open science | open minds

### **InTech Europe**

University Campus STeP Ri  
Slavka Krautzeka 83/A  
51000 Rijeka, Croatia  
Phone: +385 (51) 770 447  
Fax: +385 (51) 686 166  
[www.intechopen.com](http://www.intechopen.com)

### **InTech China**

Unit 405, Office Block, Hotel Equatorial Shanghai  
No.65, Yan An Road (West), Shanghai, 200040, China  
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元  
Phone: +86-21-62489820  
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen