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Renal Redox Balance and Na^+ , K^+ -ATPase Regulation: Role in Physiology and Pathophysiology

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1. Introduction

An imbalance between oxidant production and antioxidant defences has been associated with the development of several conditions such as hypertension, obesity-associated hypertension and diabetes, as well as during the ageing process (Bedard & Krause, 2007; Makino et al., 2003; Touyz & Schiffrin, 2004; Valko et al., 2007; Wilcox, 2005). This association is achieved through direct reactive oxygen species damage upon biomolecules or reactive oxygen species-induced alterations in gene and protein regulation (Finkel, 2003; Gill & Wilcox, 2006; McCubrey et al., 2006; Valko et al., 2007).

Being the kidney an organ severely affected in the above-mentioned conditions, most likely tissue redox balance has implications in renal physiology and pathophysiology. In this Na^+ , K^+ -ATPase is especially important since it is a key molecule in renal electrolyte regulation. As such, this chapter concerns the regulation of Na^+ , K^+ -ATPase by reactive products of oxygen metabolism and its physiological and pathophysiological implications. Focus is given to NADPH-oxidase derived reactive oxygen species, since they appear to be important for the redox-signal, and to superoxide dismutases, for being the anti-oxidant molecules that most efficiently scavenge and remove reactive oxygen species.

This chapter begins with a brief introduction to the basic principles of renal function, Na^+ , K^+ -ATPase structure and mechanisms of regulation, followed by a short review on renal reactive oxygen species production and anti-oxidant defence. An exploration of the new findings and ideas on the dynamic interplay between renal redox balance, the molecular effects at the cellular level and Na^+ , K^+ -ATPase function is approached more deeply in the following section. Finally, experimental animal models supporting that loss of redox balance and altered Na^+ , K^+ -ATPase function contribute to the development of renal associated pathologies is addressed. The chapter ends with a broad overview, given in the conclusion section.

2. The kidney function and Na^+ , K^+ -ATPase

In an adult organism the kidney plays an important role in the regulation of blood pressure, nutrient and electrolyte reabsorption and drug and metabolite excretion. This is achieved due

to the presence of specialized proteins that are distributed into specific domains of the apical or basolateral membrane of the distinct nephron segments (Abdolzade-Bavil et al., 2004).

Na^+, K^+ -ATPase is the major transporter of sodium ions in renal basolateral epithelia throughout the nephron and one of the most important renal transporters (Jaitovich & Bertorello, 2010). Na^+, K^+ -ATPase is an oligomeric transmembrane protein composed of two main subunits, α and β (Figure 1). The α -subunit is the catalytic domain of Na^+, K^+ -ATPase and contains the binding site for sodium ions, potassium ions, ATP, steroid hormones and phosphorylation sites for protein kinase A and protein kinase C (Aperia, 2001; Bertorello et al., 1991; Ewart & Klip, 1995; Feraille & Doucet, 2001; Schwartz et al., 1988). The β -subunit is involved in enzyme maturation, localization to the plasma membrane and stabilization of the potassium-occluded intermediate (Geering, 2008). A third subunit has been recently described to bind α and β complex in some tissues, such as heart, kidney and brain. This subunits belongs to the FXYD proteins, a group of structurally similar polypeptides expressed in a tissue-specific manner, and modulates cation binding affinity to Na^+, K^+ -ATPase (Crambert & Geering, 2003; Geering, 2006; Geering et al., 2003).

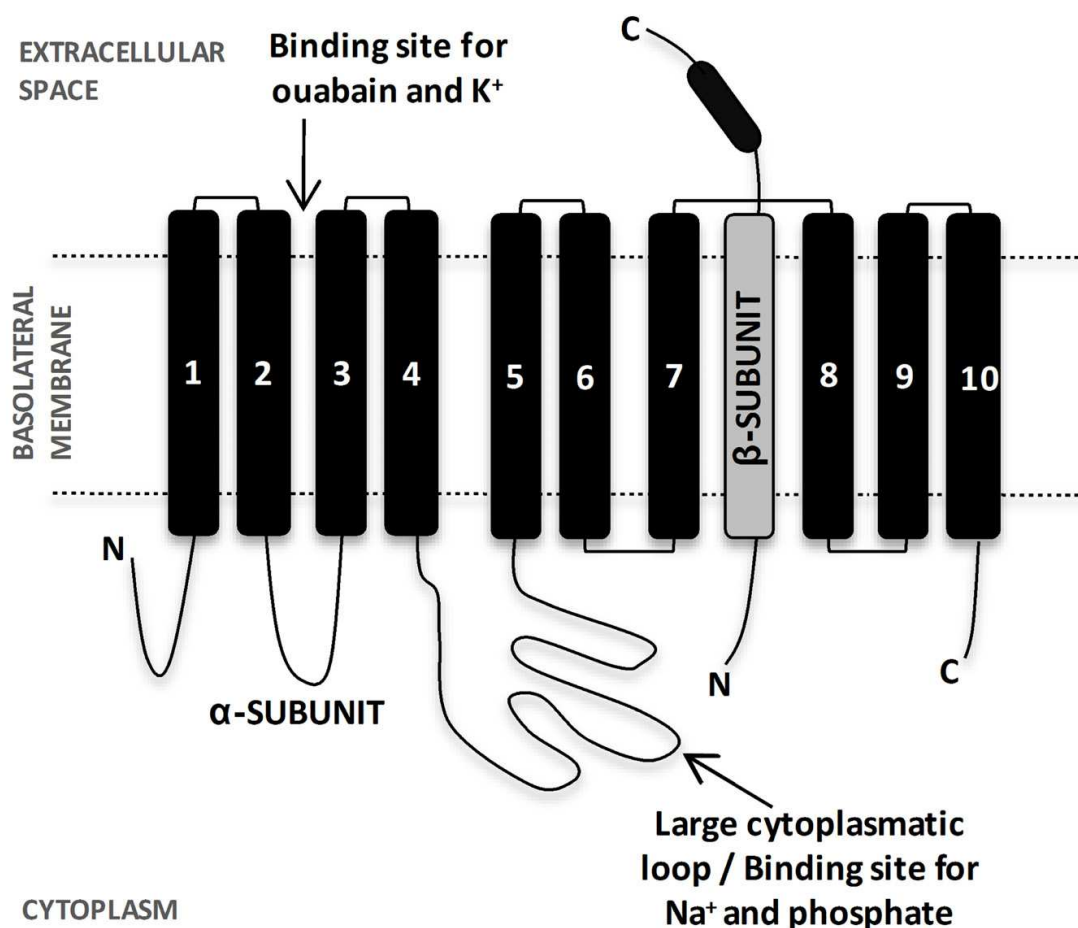


Fig. 1. Schematic representation of Na^+, K^+ -ATPase. Na^+, K^+ -ATPase is composed of two main subunits: a catalytic α -subunit (black) and a glycosylated β -subunit (grey).

There are 4 known isoforms of the α -subunit: α_1 , α_2 , α_3 and α_4 , all with a unique tissue distribution. The α_1 -isoform is expressed ubiquitously (Blanco & Mercer, 1998), and it is the major isoform expressed in the kidney (Kaplan, 2002).

The β -subunit has 3 known isoforms: β_1 , β_2 and β_3 . Detection of the tissue distribution of the β -subunit isoforms has been more difficult due to the lack of specific antibodies. However, antibody sensitivity has been improved by deglycosylation of the β -subunit. Current knowledge is that the β_1 -isoform is expressed in most tissues, including the kidney (Vagin et al., 2007). The tissue specific distribution of α and β subunits indicates that each combination exhibits unique cellular functions.

In the kidney, Na⁺,K⁺-ATPase catalyzes ATP-dependent transport of three sodium ions in exchange for two potassium ions, maintains intracellular ion balance and membrane potential and is also responsible for maintaining sodium gradient. Thus providing the driving force for nutrients, electrolytes and water reabsorption (Aperia, 2001; Feraille et al., 2001; Kaplan, 2002; Skou, 1957). In the renal proximal tubules Na⁺,K⁺-ATPase plays an essential role in the bulk reabsorption of sodium and sodium-dependent reabsorption of nutrients and other electrolytes (Feraille et al., 2001) (Figure 2). Despite approximately 70% of sodium and potassium being reabsorbed in the proximal tubules the final adjustment is made in the distal tubules and the collecting ducts, where Na⁺,K⁺-ATPase also plays a crucial role.

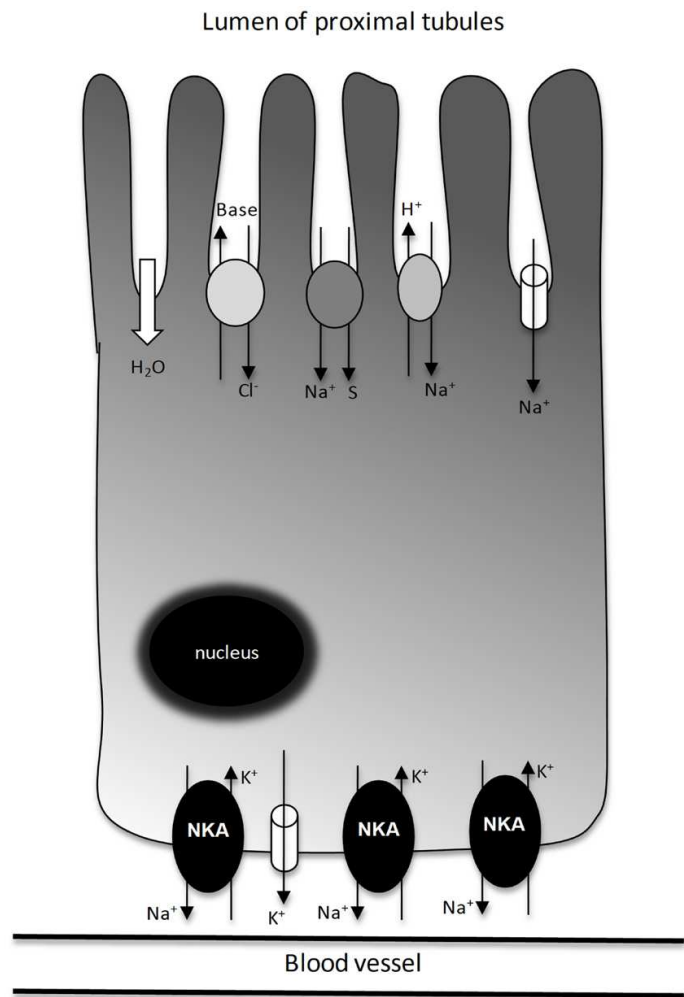


Fig. 2. Schematic representation of solute transport in the proximal tubules. A sodium gradient produced by the Na⁺,K⁺-ATPase allows sodium and organic substrates (S –sugars; amino acids; organic anions) to be transported into the cell. K⁺ is recycled across basolateral membrane through K⁺ channels.

Given the key role of Na^+, K^+ -ATPase for normal renal function changes in Na^+, K^+ -ATPase regulation are associated with the development of several conditions such as hypertension, obesity-associated hypertension and diabetes as well as during the ageing process (Jaitovich et al., 2010; Silva et al., 2010; Vague et al., 2004; Wang et al., 2009). As such understanding the mechanisms involved in regulation of Na^+, K^+ -ATPase throughout the nephron is of major importance.

Renal Na^+, K^+ -ATPase is regulated by several hormones such as dopamine, noradrenaline, aldosterone and ouabain, growth factors, peptides, several cytoskeleton proteins such as ankyrins, spectrins, adducins, actin and moesin (Aperia, 2001; Cantiello, 1997; Devarajan et al., 1994; Kraemer et al., 2003; Nelson & Veshnock, 1987; Therien & Blostein, 2000; Tripodi et al., 1996; Zhang et al., 1998) and more directly by ionic distribution across the membrane (Aperia, 2001; Feraille et al., 2001; Haber et al., 1987; Therien et al., 2000; Xie & Askari, 2002; Xie & Cai, 2003; Zhou et al., 2003). These regulatory factors may alter Na^+, K^+ -ATPase function through interference with protein synthesis, insertion in membrane compartments, enzyme internalization and substrate affinity. Protein kinases, calcium, cAMP and reactive oxygen species are known secondary messengers involved in Na^+, K^+ -ATPase regulation. In comparison to the other secondary messengers, little was known about reactive oxygen species-mediated Na^+, K^+ -ATPase regulation and much information has been gathered in the last decade.

3. Renal reactive oxygen species generation and anti-oxidant defence

Reactive oxygen species are now looked at as normal products of cell metabolism used in various physiological functions and recognised for playing a dual role as both harmful and beneficial to the organism (Valko et al., 2007; Valko et al., 2006). Produced in low/moderate concentrations reactive oxygen species may be important mediators in cellular responses to noxia, in the defence against infectious agents, in cellular signalling pathways and in the induction of a mitogenic response (Finkel & Holbrook, 2000; Gill et al., 2006; McCubrey et al., 2006; Valko et al., 2007).

An imbalance between reactive oxygen species production and anti-oxidant defence leads to the disruption of redox homeostasis and is defined as oxidative stress. Oxidative stress is a deleterious process that can induce damage to cell structures (lipids, membranes, proteins and DNA) and lead to cellular dysfunction and eventually cell death (Harman, 1956).

Reactive oxygen species encompass a series of oxygen intermediates that include the superoxide anion, hydrogen peroxide, the hydroxyl radical and hypochlorous acid. In the organism they can be produced by xanthine oxidase, NADPH-oxidase, mitochondrial oxidative phosphorylation, lipoxygenase, cytochrome P450 mono-oxygenase and heme-oxygenase 1 (Figure 3).

Despite the existence of several sources of reactive oxygen species, NADPH-oxidase appears to be especially important for the redox-signal (Gill et al., 2006; Lassegue et al., 2001). Seven NADPH-oxidase isoforms with tissue specific distribution have been identified: NOX1, NOX2, NOX3, NOX4, NOX5, DUOX1 and DOUX2. NOX1, NOX2, NOX3 and NOX4 form heterodimers with membrane $\text{p}22^{\text{phox}}$, that stabilizes the NOX proteins and docks cytosolic specific regulatory and activator subunits needed for NADPH-oxidase function (Bedard et al., 2007; Gill et al., 2006; Nauseef, 2008). NOX1 activity requires cytosolic NOXO1, in some

cases p47^{phox}, NOXA1, and Rac. NOX2 activity requires cytosolic p47^{phox}, p67^{phox}, and Rac. Moreover, p40^{phox} may also contribute to activation of NOX2. NOX3 activity requires NOXO1. NOX4 is active without cytosolic subunits. NOX5, DUOX1, and DUOX2 are activated by calcium and do not appear to require cytosolic subunits (Bedard et al., 2007).

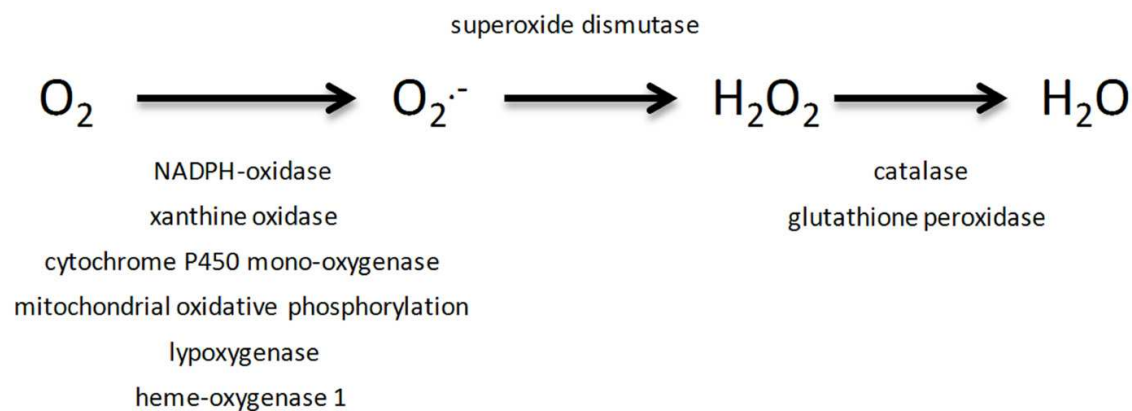


Fig. 3. Schematic representation of the balance oxidant and anti-oxidant enzymes. Multiple enzymes may induce reactive oxygen species generation that in the organism is efficiently detoxified by anti-oxidant enzymes.

Most of NADPH-oxidase isoforms are responsible for the generation of superoxide anion. However, there is still some debate on whether NOX4 generates hydrogen peroxide or superoxide anion that rapidly dismutates into hydrogen peroxide (Bedard et al., 2007; Gill et al., 2006).

The kidney is known to express at least 4 NADPH-oxidase isoforms: NOX1, NOX2, NOX4, NOX5 and NADPH-oxidase regulatory subunits (Bedard et al., 2007; Gill et al., 2006). Despite receiving considerable attention, little is known about NADPH-oxidase function in normal renal physiology (Bedard et al., 2007; Lambeth, 2007). It has been suggested that NADPH-oxidase family may play a role in secretion of erythropoietin, regulation of blood pressure by reaction of superoxide with nitric oxide limiting nitric oxide relaxing effect on afferent arterioles, alteration of cell fate through MAPK pathways activation, induction of apoptosis or cell hypertrophy through ERK 1/2 activation, regulation of gene expression by activation of transcription factors such as NF- κ B or c-jun, and innate immunity (Cui & Douglas, 1997; Dorsam et al., 2000; Gorin et al., 2004; Lodha et al., 2002; Lopez et al., 2003; Rhyu et al., 2005; Wilcox, 2003). Renal NADPH-oxidase activity is influenced by diverse stimuli such as angiotensin II, chemokine receptors and aldosterone (Bedard et al., 2007; Cave et al., 2006; Dworakowski et al., 2006; Gill et al., 2006; Lambeth, 2007; Lambeth et al., 2007).

Under physiological conditions renal reactive oxygen species production is largely contained by a complex and efficient array of antioxidant defence systems. These include antioxidant free radical scavengers such as ascorbate, vitamin E, C and A and antioxidant enzymes such as superoxide dismutase, catalase and glutathione peroxidase. Renal superoxide dismutase can rapidly dismutate cellular superoxide anion into hydrogen peroxide that is converted to water and molecular oxygen by catalase or glutathione peroxidase (Figure 3).

Transduction of the chemical reactive oxygen species signal into biological relevant events can occur through a stable sulfenic acid modification of cysteine residues in selected proteins, resulting in protein function alterations (Cave et al., 2006; Finkel, 2003; McCubrey et al., 2006). Once oxidized, proteins can undergo spontaneous or enzymatic reduction back to the initial conformation. This mechanism represents a form of signal transduction similar to phosphorylation.

A large number of proteins have been identified as specific targets of reversible oxidation, including structural proteins, transcription factors, membrane receptors, ion channels, protein kinases and protein phosphatases (Bedard et al., 2007; Cave et al., 2006) (Figure 4). Protein tyrosine phosphatases are probably the best studied, since they control the phosphorylation status of numerous signal-transducing proteins (Finkel, 2003; Meng et al., 2002). Reactive oxygen species-induced oxidation of protein tyrosine phosphatases decreases phosphatase activity by altering the tyrosine/phosphatase balance and thereby influencing signal transduction (Figure 4). This mechanism constitutes an indirect way of reactive oxygen species-mediated activation of the mitogen activated protein kinases signal pathway. However, a direct mechanism of activation of these pathways is also possible through reactive oxygen species-induced activation of membrane receptors, such as

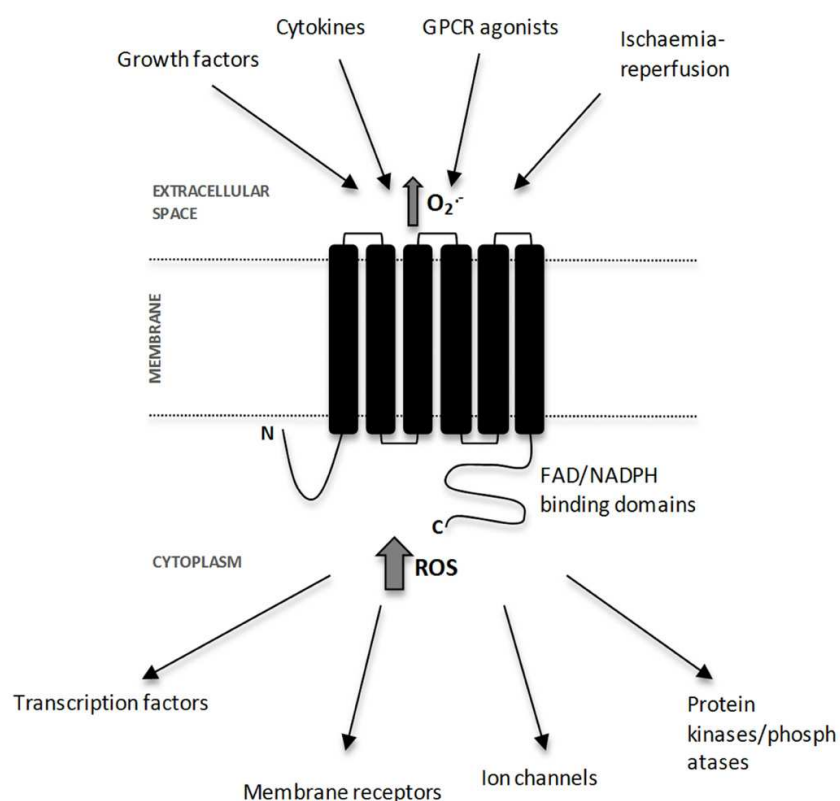


Fig. 4. Schematic representation of known activators of NADPH-oxidase isoforms and downstream effects of NADPH-oxidase derived reactive oxygen species. Diverse stimuli activate NADPH-oxidase isoforms including G-protein coupled receptor (GPCR) agonists, cytokines, growth factors and ischemia-reperfusion. NADPH-oxidase derived reactive oxygen species may influence several signalling pathways through changes in the activity of structural proteins, transcription factors, membrane receptors, ion channels and protein kinases/phosphatases.

endothelial growth factor receptor and platelet-derived growth factor receptor (McCubrey et al., 2006). As such, superoxide anion and hydrogen peroxide can be cell damaging or cell signalling molecules and play an important role in the renal physiology, as well as in the development of renal associated conditions such as diabetes and hypertension (Bedard et al., 2007; Makino et al., 2003; Touyz et al., 2004; Valko et al., 2007; Wilcox, 2005).

4. Redox balance and renal Na⁺, K⁺-ATPase regulation

Reactive oxygen species-mediated molecular mechanisms underlying Na⁺,K⁺-ATPase regulation may rely on an oxidative modification of the enzyme. In opossum kidney cells Na⁺,K⁺-ATPase activity is inhibited by acute incubation with 500 mM of hydrogen peroxide (unpublished results). This finding is in accordance with the works performed by Boldyrev *et al* (Boldyrev & Kurella, 1996) addressed at studying the kinetic parameters of the Na⁺,K⁺-ATPase after its partial inhibition by hydrogen peroxide. They used hydrogen peroxide in the mM concentration range and suggest that oxidized SH-groups of Na⁺,K⁺-ATPase interfered with the capacity of the enzyme to form active oligomers which are essential for higher Na⁺,K⁺-ATPase activity (Figure 5).

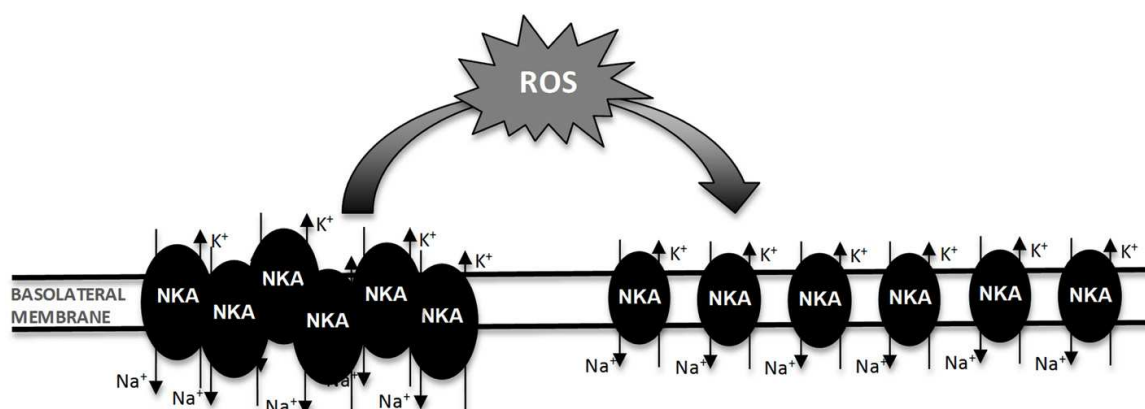


Fig. 5. Schematic representation of direct modulation of Na⁺,K⁺-ATPase by reactive oxygen species (ROS). ROS oxidize Na⁺,K⁺-ATPase interfering with the capacity of the enzyme to form active oligomers and thus decreasing Na⁺,K⁺-ATPase activity.

In cells reactive oxygen species are produced in specific domains, such as membrane microdomains, where NADPH-oxidase isoforms are present, or in the mitochondria. Depending on the cellular location where up-regulation of reactive oxygen species production takes place different modifications in cellular receptors and signalling pathways that alter phosphorylation, gene or protein expression can be achieved. Thus, also different outcomes regarding Na⁺,K⁺-ATPase activity and expression may be observed.

Reactive oxygen species-mediated alteration of cell signalling events that are participate in Na⁺,K⁺-ATPase regulation have been described both *in vivo* and *in vitro*. In Madin-Darby canine kidney cells it has been demonstrated that low potassium-induced an increase in Na⁺,K⁺-ATPase protein content and cell surface Na⁺,K⁺-ATPase expression (Zhou et al., 2003). The signal pathway that transduces the stimulatory effect of low potassium onto up-regulation of Na⁺,K⁺-ATPase is dependent on NADPH-oxidase-derived reactive oxygen

species production, Sp1 up-regulation and enhanced transcription of both α and β -subunits of Na^+, K^+ -ATPase (Yin et al., 2008). Also, in opossum kidney cell line increased levels of reactive oxygen species production in cells with 80 passages in culture were responsible for up-regulation of Na^+, K^+ -ATPase (Silva et al., 2006; Silva & Soares-da-Silva, 2007). Opossum kidney cells with 80 passages in culture were shown to have overexpression of NOX1, superoxide dismutase 1, superoxide dismutase 2 and superoxide dismutase 3 isoforms (Silva et al., 2007) and decreased availability to catalyze hydrogen peroxide degradation (unpublished results). When opossum kidney cells with 80 passages in culture were treated with antioxidants (apocynin or TEMPOL) Na^+, K^+ -ATPase activity was found to be significantly decreased (Silva et al., 2007) (Figure 6).

In the proximal tubules of aged rats age-related increase in reactive oxygen species levels were found to be responsible for decreased basal Na^+, K^+ -ATPase activity (Asghar et al., 2001; Silva et al., 2010). The molecular mechanism responsible for the observed decrease in Na^+, K^+ -ATPase activity was a higher basal Na^+, K^+ -ATPase phosphorylation due to reactive oxygen species-mediated increase in protein kinase C activity (Asghar et al., 2003; Asghar et al., 2001; Asghar & Lokhandwala, 2004) (Figure 6).

In the renal medulla where the final regulation of sodium and potassium in the urine takes place, ageing was accompanied by a significant increase in Na^+, K^+ -ATPase activity and expression of the α_1 -subunit (Silva et al., 2010). Furthermore, not only was Na^+, K^+ -ATPase activity increased in renal medulla of aged Wistar Kyoto rats but also, in this part of the kidney, hydrogen peroxide production was increased with age and in comparison with renal cortex (Silva et al., 2010). Given that Na^+, K^+ -ATPase regulation differs between the proximal and distal nephron segments it is possible that, in the renal medulla, increased reactive oxygen species may activate cell specific signal pathways that up-regulate Na^+, K^+ -ATPase activity. In fact, a study performed by Beltowski and co-workers (Beltowski et al., 2004) investigating whether superoxide anion was involved in the regulation of renal Na^+, K^+ -ATPase support this hypothesis (Figure 6). In this study, infusion of compounds modulating superoxide anion concentration into the abdominal aorta proximally to the renal rat arteries increased medullary Na^+, K^+ -ATPase activity but had no effect on cortical Na^+, K^+ -ATPase activity. Both apocynin and TEMPOL decreased the medullary Na^+, K^+ -ATPase activity. The inhibitory effect of apocynin and TEMPOL was abolished by inhibitors of nitric oxide synthase, soluble guanylate cyclase and protein kinase G. The suggested mechanisms of Na^+, K^+ -ATPase regulation is that NADPH-oxidase-derived superoxide anion increases Na^+, K^+ -ATPase activity in the renal medulla by reducing the availability of nitric oxide. Other mechanism of reactive oxygen species-mediated regulation of Na^+, K^+ -ATPase has been described by the same group. They reported that leptin-induced stimulation of renal Na^+, K^+ -ATPase involves hydrogen peroxide generation, Src kinase, transactivation of the EGF receptor, and stimulation of ERK (Beltowski et al., 2006; Wojcicka et al., 2008) (Figure 6). However, the mechanism of Na^+, K^+ -ATPase regulation in leptin-treated rats shifts from hydrogen peroxide/ERK-dependent to superoxide anion/nitric oxide-dependent after 8 days of treatment, due to a decrease in superoxide dismutase activity and as a consequence higher cellular levels of superoxide anion (Beltowski, 2010; Beltowski et al., 2008).

Finally, reactive oxygen species have also been demonstrated to alter Na^+, K^+ -ATPase activity due to interference with membrane receptor function (Figure 6). Asghar et al (2004)

demonstrated that dopamine was unable to inhibit Na⁺,K⁺-ATPase activity in old rats and treatment with antioxidants restored the coupling of dopamine type 1 receptor to G proteins. Further studies allowed the identification of the mechanism responsible for decoupling of dopamine type 1 receptor from G proteins (Asghar & Lokhandwala, 2006; Fardoun et al., 2007). In renal proximal tubules increased reactive oxygen species production activates NF- κ B and promotes its translocation to the nucleus, where it increases transcription of protein kinase C. This causes an increase in protein kinase C expression and activity, which leads to GRK-2 translocation to the membrane and subsequent dopamine type 1 receptor hyper-phosphorylation and uncoupling from protein G.

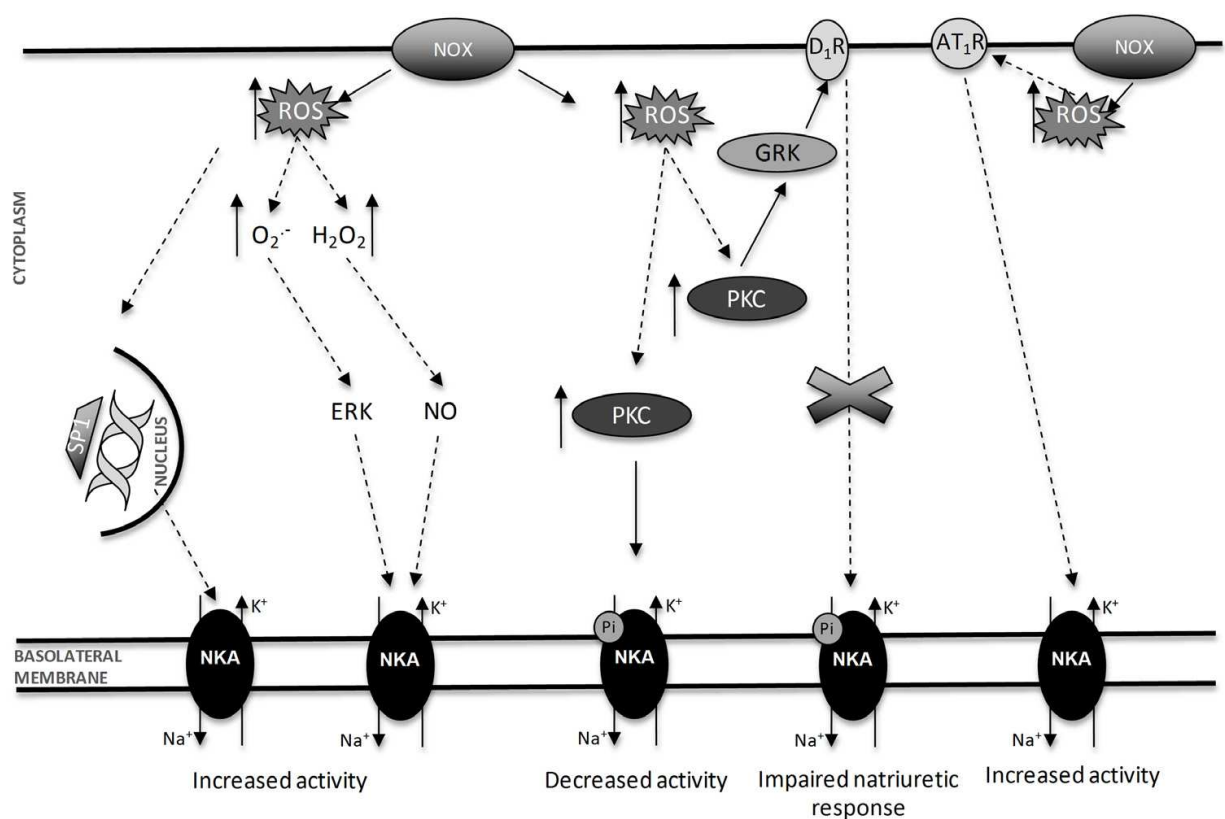


Fig. 6. Schematic representation of reactive oxygen species-mediated modulation of Na⁺,K⁺-ATPase. An increase in cellular levels of reactive oxygen species, mainly due to increase activity of NADPH-oxidase, activates specific cell signal cascades that interfere with Na⁺,K⁺-ATPase function. Filled arrows indicate a direct action. Dashed arrows indicate that the pathway is not known or was simplified.

5. Loss of redox balance: Functional consequences in renal physiology

Na⁺,K⁺-ATPase function in the renal proximal tubules has been a target of extensive research due to the fact that, as mentioned before, it plays a crucial role in the bulk of sodium and nutrient reabsorption. Moreover, as recently reviewed by Wang and co-workers (Wang et al., 2009) it is well known that altered renal proximal tubular sodium reabsorption is implicated in the development of essential hypertension.

In the renal proximal tubules it is well established that dopamine promotes natriuresis while angiotensin II increases sodium retention. Dopamine promotes natriuresis via the activation of dopamine type 1 receptors. The dopamine type 1 receptors couple to G_s -proteins and activate the adenylate cyclase-cAMP-protein kinase A signaling pathway. In the kidney dopamine type 1 receptors can also couple to $G_{q/11}$ and activate the phospholipase C-diacylglycerol-protein kinase C pathway. Both a cross-talk between the protein kinase A and protein kinase C signaling pathways or an activation of the phospholipase C-protein kinase C pathway by protein kinase A have been described (Aperia, 2000; Brismar et al., 2000; Gomes & Soares-da-Silva, 2002; Hussain & Lokhandwala, 1998; Jose et al., 2000). Thus, activation of plasma membrane dopamine type 1 receptors stimulates a tissue specific signal cascade that leads to the activation of the protein kinase C δ -isoform. Protein kinase C δ -isoform phosphorylates the α_1 -subunit of Na^+,K^+ -ATPase producing a conformational change of amino-terminal, which through interaction with other domains of the α_1 -subunit of Na^+,K^+ -ATPase exposes the binding domains for phosphoinositide 3-kinase and adaptor protein-2. Binding of these proteins induces the activation of Na^+,K^+ -ATPase endocytosis in the proximal tubules and promotes natriuresis (Cinelli et al., 2008; Efendiev et al., 2003; Pedemonte et al., 2005). Angiotensin II exerts an anti-natriuretic effect via the activation of angiotensin II type 1 receptors. Angiotensin II type 1 receptors are predominantly coupled to G-proteins and signal through phospholipases, inositol-phosphatases, calcium channels and serine/threonine and tyrosine kinases. Activation of plasma angiotensin II type 1 receptors stimulates a tissue-specific signaling cascade that leads to the activation of the protein kinase C β -isoform. The protein kinase C β -isoform phosphorylates the α_1 -subunit of Na^+,K^+ -ATPase producing a conformational change that increases the interaction between the α_1 -subunit of Na^+,K^+ -ATPase and adaptor protein-1, which results in the recruitment of the enzyme to the plasma membrane (Efendiev et al., 2000; Efendiev et al., 2003).

A reactive oxygen species-associated defect in renal dopamine type 1 receptor function has been observed not only in experimental models of hypertension (Banday et al., 2008; Hussain et al., 1999; Hussain & Lokhandwala, 1997a; Hussain & Lokhandwala, 1997b) but also in diabetes (Banday et al., 2005; Marwaha & Lokhandwala, 2006) and ageing (Fardoun et al., 2006; Hussain et al., 1999; Vieira-Coelho et al., 1999). Failure of dopamine to modulate sodium reabsorption results in diminished natriuresis and blood pressure elevation. More recently it was also demonstrated that reactive oxygen species-mediated angiotensin type 1 receptor up-regulation increases sodium transporters and subsequently contributes to sodium retention and blood pressure elevation (Banday & Lokhandwala, 2008a; Banday & Lokhandwala, 2008b). Angiotensin type 1 receptor up-regulation has been observed in experimental models of hypertension (Reja et al., 2006).

Interest in the regulation of sodium transport in renal medulla is more recent and mainly due to the existence of a possible role in the initiation and development of several forms of experimental hypertension (Cowley & Roman, 1996; Cowley et al., 1992). In renal medulla reactive oxygen species appear to directly alter sodium reabsorption and indirectly alter medullar blood flow, contributing to the development of hypertension (Cowley, 2008; Taylor et al., 2006a; Taylor et al., 2006b). One of the main sources of superoxide anion in this part of the kidney is NADPH-oxidase. NADPH-oxidase-derived superoxide anion was found to contribute to the development of salt-induced hypertension in Dahl salt-sensitive rats (Taylor et al., 2006a). A functional consequence of elevations of superoxide anion within the renal medulla was found to be an immediate reduction of sodium excretion. In this setting, Na^+,K^+ -ATPase may play a role since renal medullary superoxide anion can increase

Na⁺,K⁺-ATPase activity by reducing availability of nitric oxide, as previously described (Beltowski et al., 2004). This mechanism may contribute to an increase in sodium reabsorption and the development of hypertension.

It is now evident that by disrupting several mechanisms responsible for maintenance of sodium homeostasis reactive oxygen species can contribute to the development of renal pathologies and hypertension.

6. Conclusion

Reactive oxygen species-mediated regulation of Na⁺,K⁺-ATPase has been receiving considerable attention. There is now an increasing number of publications addressing multiple mechanisms by which reactive oxygen species may alter Na⁺,K⁺-ATPase function and contribute to the development of several conditions, with special focus being given to hypertension. However, many intermediate events in the mechanisms of reactive oxygen species-mediated regulation of Na⁺,K⁺-ATPase are still unknown and much work needs to be done. Moreover, given that normalization of the redox imbalance in the proximal and distal nephron segments may require the use of specific anti-oxidant molecules and/or pharmacological modulation of different signaling pathways attention should be paid in future therapeutic approaches.

7. References

- Abdolzade-Bavil A, Hayes S, Goretzki L, Kroger M, Anders J & Hendriks R (2004). Convenient and versatile subcellular extraction procedure, that facilitates classical protein expression profiling and functional protein analysis. *Proteomics* 4(5): 1397-1405. ISSN 1615-9853
- Aperia A (2001). Regulation of sodium/potassium ATPase activity: impact on salt balance and vascular contractility. *Current hypertension reports* 3(2): 165-171. ISSN 1522-6417
- Aperia AC (2000). Intrarenal dopamine: a key signal in the interactive regulation of sodium metabolism. *Annual review of physiology* 62: 621-647. ISSN 0066-4278
- Asghar M, Hussain T & Lokhandwala MF (2003). Overexpression of PKC-betaI and -delta contributes to higher PKC activity in the proximal tubules of old Fischer 344 rats. *American journal of physiology. Renal physiology* 285(6): F1100-1107. ISSN 1931-857X
- Asghar M, Kansra V, Hussain T & Lokhandwala MF (2001). Hyperphosphorylation of Na-pump contributes to defective renal dopamine response in old rats. *Journal of the american society of nephrology* 12(2): 226-232. ISSN 1046-6673
- Asghar M & Lokhandwala MF (2004). Antioxidant supplementation normalizes elevated protein kinase C activity in the proximal tubules of old rats. *Experimental biology and medicine (Maywood)* 229(3): 270-275. ISSN 1535-3702
- Asghar M & Lokhandwala MF (2006). Antioxidant tempol lowers age-related increases in insulin resistance in Fischer 344 rats. *Clinical and experimental hypertension* 28(5): 533-541. ISSN 1064-1963
- Banday AA, Lau YS & Lokhandwala MF (2008). Oxidative stress causes renal dopamine D1 receptor dysfunction and salt-sensitive hypertension in Sprague-Dawley rats. *Hypertension* 51(2): 367-375. ISSN 1524-4563
- Banday AA & Lokhandwala MF (2008a). Loss of biphasic effect on Na/K-ATPase activity by angiotensin II involves defective angiotensin type 1 receptor-nitric oxide signaling. *Hypertension* 52(6): 1099-1105. ISSN 1524-4563

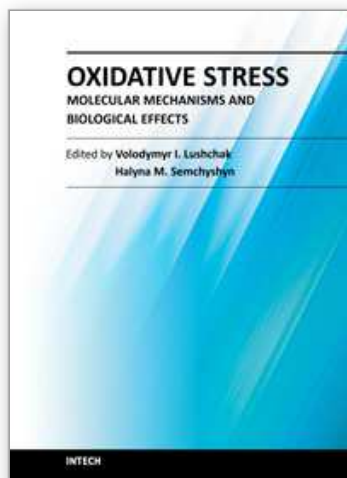
- Banday AA & Lokhandwala MF (2008b). Oxidative stress-induced renal angiotensin AT1 receptor upregulation causes increased stimulation of sodium transporters and hypertension. *American journal of physiology. Renal physiology* 295(3): F698-706. ISSN 1931-857X
- Banday AA, Marwaha A, Tallam LS & Lokhandwala MF (2005). Tempol reduces oxidative stress, improves insulin sensitivity, decreases renal dopamine D1 receptor hyperphosphorylation, and restores D1 receptor-G-protein coupling and function in obese Zucker rats. *Diabetes* 54(7): 2219-2226. ISSN 0012-1797
- Bedard K & Krause KH (2007). The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiological reviews* 87(1): 245-313. ISSN 0031-9333
- Beltowski J (2010). Leptin and the Regulation of Renal Sodium Handling and Renal Na-Transporting ATPases: Role in the Pathogenesis of Arterial Hypertension. *Current cardiology reviews* 6(1): 31-40. ISSN 1875-6557
- Beltowski J, Jamroz-Wisniewska A, Wojcicka G, Lowicka E & Wojtak A (2008). Renal antioxidant enzymes and glutathione redox status in leptin-induced hypertension. *Molecular and cellular biochemistry* 319(1-2): 163-174. ISSN 1573-4919
- Beltowski J, Marciniak A, Jamroz-Wisniewska A & Borkowska E (2004). Nitric oxide -- superoxide cooperation in the regulation of renal Na(+),K(+)-ATPase. *Acta biochimica polonica Pol* 51(4): 933-942. ISSN 0001-527X
- Beltowski J, Wojcicka G, Trzeciak J & Marciniak A (2006). H₂O₂ and Src-dependent transactivation of the EGF receptor mediates the stimulatory effect of leptin on renal ERK and Na⁺,K⁺-ATPase. *Peptides* 27(12): 3234-3244. ISSN 0196-9781
- Bertorello AM, Aperia A, Walaas SI, Nairn AC & Greengard P (1991). Phosphorylation of the catalytic subunit of Na⁺,K⁺-ATPase inhibits the activity of the enzyme. *Proceedings of the national academy of sciences of the United States of America* 88(24): 11359-11362. ISSN 0027-8424
- Blanco G & Mercer RW (1998). Isozymes of the Na⁺,K⁺-ATPase: heterogeneity in structure, diversity in function. *American journal of physiology* 275(5 Pt 2): F633-650. ISSN 0002-9513
- Boldyrev A & Kurella E (1996). Mechanism of oxidative damage of dog kidney Na/K-ATPase. *Biochemical and biophysical research communications* 222(2): 483-487. ISSN 0006-291X
- Brismar H, Holtback U & Aperia A (2000). Mechanisms by which intrarenal dopamine and ANP interact to regulate sodium metabolism. *Clinical and experimental hypertension* 22(3): 303-307. ISSN 1064-1963
- Cantiello HF (1997). Changes in actin filament organization regulate Na⁺,K⁺-ATPase activity. Role of actin phosphorylation. *Annals of the New York academy of sciences* 834: 559-561. ISSN 0077-8923
- Cave AC, Brewer AC, Narayanapanicker A, Ray R, Grieve DJ, Walker S, et al. (2006). NADPH oxidases in cardiovascular health and disease. *Antioxidants & redox signaling* 8(5-6): 691-728. ISSN 1523-0864
- Cinelli AR, Efendiev R & Pedemonte CH (2008). Trafficking of Na⁺,K⁺-ATPase and dopamine receptor molecules induced by changes in intracellular sodium concentration of renal epithelial cells. *American journal of physiology. Renal physiology* 295(4): F1117-1125. ISSN 0363-6127
- Cowley AW, Jr. (2008). Renal medullary oxidative stress, pressure-natriuresis, and hypertension. *Hypertension* 52(5): 777-786. ISSN 1524-4563
- Cowley AW, Jr. & Roman RJ (1996). The role of the kidney in hypertension. *The journal of the american medical association* 275(20): 1581-1589. ISSN 0098-7484

- Cowley AW, Roman RJ, Fenoy FJ & Mattson DL (1992). Effect of renal medullary circulation on arterial pressure. *Journal of hypertension* 10(7): S187-193. ISSN 0952-1178
- Crambert G & Geering K (2003). FXYD proteins: new tissue-specific regulators of the ubiquitous Na⁺,K⁺-ATPase. *Science signaling - The signal transduction knowledge environment* 2003(166): RE1. ISSN 1525-8882
- Cui XL & Douglas JG (1997). Arachidonic acid activates c-jun N-terminal kinase through NADPH oxidase in rabbit proximal tubular epithelial cells. *Proceedings of the National Academy of Sciences of the United States of America* 94(8): 3771-3776. ISSN 0027-8424
- Devarajan P, Scaramuzzino DA & Morrow JS (1994). Ankyrin binds to two distinct cytoplasmic domains of Na⁺,K⁺-ATPase α -subunit. *Proceedings of the National Academy of Sciences of the United States of America* 91(8): 2965-2969. ISSN 0027-8424
- Dorsam G, Taher MM, Valerie KC, Kuemmerle NB, Chan JC & Franson RC (2000). Diphenyleneiodium chloride blocks inflammatory cytokine-induced up-regulation of group IIA phospholipase A(2) in rat mesangial cells. *Journal of pharmacology and experimental therapeutics* 292(1): 271-279. ISSN 0022-3565
- Dworakowski R, Anilkumar N, Zhang M & Shah AM (2006). Redox signalling involving NADPH oxidase-derived reactive oxygen species. *Biochemical Society transactions* 34(Pt 5): 960-964. ISSN 0300-5127
- Efendiev R, Bertorello AM, Pressley TA, Rousselot M, Feraille E & Pedemonte CH (2000). Simultaneous phosphorylation of Ser11 and Ser18 in the α -subunit promotes the recruitment of Na⁺,K⁺-ATPase molecules to the plasma membrane. *Biochemistry* 39(32): 9884-9892. ISSN 0006-2960
- Efendiev R, Budu CE, Cinelli AR, Bertorello AM & Pedemonte CH (2003). Intracellular Na⁺ regulates dopamine and angiotensin II receptors availability at the plasma membrane and their cellular responses in renal epithelia. *The journal of biological chemistry* 278(31): 28719-28726. ISSN 0021-9258
- Ewart HS & Klip A (1995). Hormonal regulation of the Na⁺,K⁺-ATPase: mechanisms underlying rapid and sustained changes in pump activity. *American journal of physiology* 269(2 Pt 1): C295-311. ISSN 0002-9513
- Fardoun RZ, Asghar M & Lokhandwala M (2007). Role of nuclear factor kappa B (NF-kappaB) in oxidative stress-induced defective dopamine D1 receptor signaling in the renal proximal tubules of Sprague-Dawley rats. *Free radical biology & medicine* 42(6): 756-764. ISSN 0891-5849
- Fardoun RZ, Asghar M & Lokhandwala M (2006). Role of oxidative stress in defective renal dopamine D1 receptor-G protein coupling and function in old Fischer 344 rats. *American journal of physiology. Renal physiology* 291(5): F945-951. ISSN 1931-857X
- Feraille E & Doucet A (2001). Sodium-potassium-adenosinetriphosphatase-dependent sodium transport in the kidney: hormonal control. *Physiological reviews* 81(1): 345-418. ISSN 0031-9333
- Finkel T (2003). Oxidant signals and oxidative stress. *Current opinion in cell biology* 15(2): 247-254. ISSN 0955-0674
- Finkel T & Holbrook NJ (2000). Oxidants, oxidative stress and the biology of ageing. *Nature* 408(6809): 239-247. ISSN 0028-0836
- Geering K (2008). Functional roles of Na⁺,K⁺-ATPase subunits. *American journal of physiology. Renal physiology* 17(5): 526-532. ISSN 1062-4821
- Geering K (2006). FXYD proteins: new regulators of Na⁺,K⁺-ATPase. *American journal of physiology. Renal physiology* 290(2): F241-250. ISSN 1931-857X

- Geering K, Beguin P, Garty H, Karlsh S, Fuzesi M, Horisberger JD, *et al.* (2003). FXYP proteins: new tissue- and isoform-specific regulators of Na⁺,K⁺-ATPase. *Annals of the New York academy of sciences* 986: 388-394. ISSN 0077-8923
- Gill PS & Wilcox CS (2006). NADPH oxidases in the kidney. *Antioxidants & redox signaling* 8(9-10): 1597-1607. ISSN 1523-0864
- Gomes P & Soares-da-Silva P (2002). Role of cAMP-PKA-PLC signaling cascade on dopamine-induced PKC-mediated inhibition of renal Na⁺,K⁺-ATPase activity. *American journal of physiology. Renal physiology* 282(6): F1084-1096. ISSN 0363-6127
- Gorin Y, Ricono JM, Wagner B, Kim NH, Bhandari B, Choudhury GG, *et al.* (2004). Angiotensin II-induced ERK1/ERK2 activation and protein synthesis are redox-dependent in glomerular mesangial cells. *Biochemical journal* 381(Pt 1): 231-239. ISSN 1470-8728
- Haber RS, Pressley TA, Loeb JN & Ismail-Beigi F (1987). Ionic dependence of active Na-K transport: "clamping" of cellular Na⁺ with monensin. *American journal of physiology* 253(1 Pt 2): F26-33. ISSN 0002-9513
- Harman D (1956). Aging: a theory based on free radical and radiation chemistry. *Journal of gerontology* 11(3): 298-300. ISSN 0022-1422
- Hussain T, Kansra V & Lokhandwala MF (1999). Renal dopamine receptor signaling mechanisms in spontaneously hypertensive and Fischer 344 old rats. *Clinical and experimental hypertension* 21(1-2): 25-36. ISSN 1064-1963
- Hussain T & Lokhandwala MF (1997a). Dopamine-1 receptor G-protein coupling and the involvement of phospholipase A2 in dopamine-1 receptor mediated cellular signaling mechanisms in the proximal tubules of SHR. *Clinical and experimental hypertension* 19(1-2): 131-140. ISSN 1064-1963
- Hussain T & Lokhandwala MF (1997b). Renal dopamine DA1 receptor coupling with G(S) and G(q/11) proteins in spontaneously hypertensive rats. *The American journal of physiology* 272(3 Pt 2): F339-346. ISSN 0002-9513
- Hussain T & Lokhandwala MF (1998). Renal dopamine receptor function in hypertension. *Hypertension* 32(2): 187-197. ISSN 0194-911X
- Jaitovich A & Bertorello AM (2010). Salt, Na⁺,K⁺-ATPase and hypertension. *Life sciences* 86(3-4): 73-78. ISSN 1879-0631
- Jose PA, Eisner GM & Felder RA (2000). Renal dopamine and sodium homeostasis. *Current hypertension reports* 2(2): 174-183. ISSN 1522-6417
- Kaplan JH (2002). Biochemistry of Na⁺,K⁺-ATPase. *Annual review of biochemistry* 71: 511-535. ISSN 0066-4154
- Kraemer DM, Strizek B, Meyer HE, Marcus K & Drenckhahn D (2003). Kidney Na⁺,K⁺-ATPase is associated with moesin. *European journal of cell biology* 82(2): 87-92. ISSN 0171-9335
- Lambeth JD (2007). Nox enzymes, ROS, and chronic disease: an example of antagonistic pleiotropy. *Free radical biology & medicine* 43(3): 332-347. ISSN 0891-5849
- Lambeth JD, Kawahara T & Diebold B (2007). Regulation of Nox and Duox enzymatic activity and expression. *Free radical biology & medicine* 43(3): 319-331. ISSN 0891-5849
- Lassegue B, Sorescu D, Szocs K, Yin Q, Akers M, Zhang Y, *et al.* (2001). Novel gp91(phox) homologues in vascular smooth muscle cells : nox1 mediates angiotensin II-induced superoxide formation and redox-sensitive signaling pathways. *Circulation research* 88(9): 888-894. ISSN 1524-4571
- Lodha S, Dani D, Mehta R, Bhaskaran M, Reddy K, Ding G, *et al.* (2002). Angiotensin II-induced mesangial cell apoptosis: role of oxidative stress. *Molecular medicine* 8(12): 830-840. ISSN 1076-1551

- Lopez B, Salom MG, Arregui B, Valero F & Fenoy FJ (2003). Role of superoxide in modulating the renal effects of angiotensin II. *Hypertension* 42(6): 1150-1156. ISSN 1524-4563
- Makino A, Skelton MM, Zou AP & Cowley AW, Jr. (2003). Increased renal medullary H₂O₂ leads to hypertension. *Hypertension* 42(1): 25-30. ISSN 1524-4563
- Marwaha A & Lokhandwala MF (2006). Tempol reduces oxidative stress and restores renal dopamine D1-like receptor- G protein coupling and function in hyperglycemic rats. *American journal of physiology. Renal physiology* 291(1): F58-66. ISSN 1931-857X
- McCubrey JA, Lahair MM & Franklin RA (2006). Reactive oxygen species-induced activation of the MAP kinase signaling pathways. *Antioxidants & redox signaling* 8(9-10): 1775-1789. ISSN 1523-0864
- Meng TC, Fukada T & Tonks NK (2002). Reversible oxidation and inactivation of protein tyrosine phosphatases in vivo. *Molecular cell* 9(2): 387-399. ISSN 1097-2765
- Nauseef WM (2008). Biological roles for the NOX family NADPH oxidases. *The Journal of biological chemistry* 283(25): 16961-16965. ISSN 0021-9258
- Nelson WJ & Veshnock PJ (1987). Ankyrin binding to Na⁺,K⁺-ATPase and implications for the organization of membrane domains in polarized cells. *Nature* 328(6130): 533-536. ISSN 0028-0836
- Pedemonte CH, Efendiev R & Bertorello AM (2005). Inhibition of Na⁺,K⁺-ATPase by dopamine in proximal tubule epithelial cells. *Seminars in nephrology* 25(5): 322-327. ISSN 0270-9295
- Reja V, Goodchild AK, Phillips JK & Pilowsky PM (2006). Upregulation of angiotensin AT1 receptor and intracellular kinase gene expression in hypertensive rats. *Clinical and experimental pharmacology & physiology* 33(8): 690-695. ISSN 0305-1870
- Rhyu DY, Yang Y, Ha H, Lee GT, Song JS, Uh ST, et al. (2005). Role of reactive oxygen species in TGF-beta1-induced mitogen-activated protein kinase activation and epithelial-mesenchymal transition in renal tubular epithelial cells. *Journal of the american society of nephrology* 16(3): 667-675. ISSN 1046-6673
- Schwartz A, Grupp G, Wallick E, Grupp IL & Ball WJ, Jr. (1988). Role of the Na⁺,K⁺-ATPase in the cardiotoxic action of cardiac glycosides. *Progress in clinical and biological research* 268B: 321-338. ISSN 0361-7742
- Silva E, Gomes P & Soares-da-Silva P (2006). Overexpression of Na⁺,K⁺-ATPase parallels the increase in sodium transport and potassium recycling in an in vitro model of proximal tubule cellular ageing. *The Journal of membrane biology* 212(3): 163-175. ISSN 0022-2631
- Silva E, Pinto V, Simao S, Serrao MP, Afonso J, Amaral J, et al. (2010). Renal aging in WKY rats: changes in Na⁺,K⁺ -ATPase function and oxidative stress. *Experimental gerontology* 45(12): 977-983. ISSN 1873-6815
- Silva E & Soares-da-Silva P (2007). Reactive oxygen species and the regulation of renal Na⁺-K⁺-ATPase in opossum kidney cells. *American journal of physiology. Regulatory, integrative and comparative physiology* 293(4): R1764-1770. ISSN 0363-6119
- Skou JC (1957). The influence of some cations on an adenosine triphosphatase from peripheral nerves. *Biochimica et biophysica acta* 23(2): 394-401. ISSN 0006-3002
- Taylor NE, Glocka P, Liang M & Cowley AW, Jr. (2006a). NADPH oxidase in the renal medulla causes oxidative stress and contributes to salt-sensitive hypertension in Dahl S rats. *Hypertension* 47(4): 692-698. ISSN 1524-4563
- Taylor NE, Maier KG, Roman RJ & Cowley AW, Jr. (2006b). NO synthase uncoupling in the kidney of Dahl S rats: role of dihydrobiopterin. *Hypertension* 48(6): 1066-1071. ISSN 1524-4563

- Therien AG & Blostein R (2000). Mechanisms of sodium pump regulation. *American journal of physiology. Cell physiology* 279(3): C541-566. ISSN 0363-6143
- Touyz RM & Schiffrin EL (2004). Reactive oxygen species in vascular biology: implications in hypertension. *Histochemistry and cell biology* 122(4): 339-352. ISSN 0948-6143
- Tripodi G, Valtorta F, Torielli L, Chiergatti E, Salardi S, Trusolino L, *et al.* (1996). Hypertension-associated point mutations in the adducin α and β subunits affect actin cytoskeleton and ion transport. *The Journal of clinical investigation* 97(12): 2815-2822. ISSN 0021-9738
- Vagin O, Sachs G & Tokhtaeva E (2007). The roles of the Na^+, K^+ -ATPase β_1 subunit in pump sorting and epithelial integrity. *Journal of bioenergetics and biomembranes* 39(5-6): 367-372. ISSN 0145-479X
- Vague P, Coste TC, Jannot MF, Raccah D & Tsimaratos M (2004). C-peptide, Na^+, K^+ -ATPase, and diabetes. *Experimental diabetes research* 5(1): 37-50. ISSN 1543-8600
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M & Telser J (2007). Free radicals and antioxidants in normal physiological functions and human disease. *The international journal of biochemistry & cell biology* 39(1): 44-84. ISSN 1357-2725
- Valko M, Rhodes CJ, Moncol J, Izakovic M & Mazur M (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-biological interactions* 160(1): 1-40. ISSN 0009-2797
- Vieira-Coelho MA, Hussain T, Kansra V, Serrao MP, Guimaraes JT, Pestana M, *et al.* (1999). Aging, high salt intake, and renal dopaminergic activity in Fischer 344 rats. *Hypertension* 34(4 Pt 1): 666-672. ISSN 0194-911X
- Wang X, Armando I, Upadhyay K, Pascua A & Jose PA (2009). The regulation of proximal tubular salt transport in hypertension: an update. *Current opinion in nephrology and hypertension* 18(5): 412-420. ISSN 1535-3842
- Wilcox CS (2005). Oxidative stress and nitric oxide deficiency in the kidney: a critical link to hypertension? *American journal of physiology* 289(4): R913-935. ISSN 0363-6119
- Wilcox CS (2003). Redox regulation of the afferent arteriole and tubuloglomerular feedback. *Acta physiologica scandinavica* 179(3): 217-223. ISSN 0001-6772
- Wojcicka G, Jamroz-Wisniewska A, Widomska S, Ksiazek M & Beltowski J (2008). Role of extracellular signal-regulated kinases (ERK) in leptin-induced hypertension. *Life sciences* 82(7-8): 402-412. ISSN 0024-3205
- Xie Z & Askari A (2002). Na^+, K^+ -ATPase as a signal transducer. *European journal of biochemistry / FEBS* 269(10): 2434-2439. ISSN 0014-2956
- Xie Z & Cai T (2003). Na^+, K^+ -ATPase-mediated signal transduction: from protein interaction to cellular function. *Molecular interventions* 3(3): 157-168. ISSN 1534-0384
- Yin W, Yin FZ, Shen WX, Cai BC & Hua ZC (2008). Requirement of hydrogen peroxide and Sp1 in the stimulation of Na, K -ATPase by low potassium in MDCK epithelial cells. *The international journal of biochemistry & cell biology* 40(5): 942-953. ISSN 1357-2725
- Zhang Z, Devarajan P, Dorfman AL & Morrow JS (1998). Structure of the ankyrin-binding domain of α - Na^+, K^+ -ATPase. *The journal of biological chemistry* 273(30): 18681-18684. ISSN 0021-9258
- Zhou X, Yin W, Doi SQ, Robinson SW, Takeyasu K & Fan X (2003). Stimulation of Na^+, K^+ -ATPase by low potassium requires reactive oxygen species. *American journal of physiology. Cell physiology* 285(2): C319-326. ISSN 0363-6143



Oxidative Stress - Molecular Mechanisms and Biological Effects

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ISBN 978-953-51-0554-1

Hard cover, 362 pages

Publisher InTech

Published online 25, April, 2012

Published in print edition April, 2012

Since the discovery of free radicals in biological systems researchers have been highly interested in their interaction with biological molecules. Denoted in 1980, and due to fruitful results and ideas, oxidative stress is now appreciated by both basic and applied scientists as an enhanced steady state level of reactive oxygen species with wide range of biological effects. This book covers a wide range of aspects and issues related to the field of oxidative stress. The association between generation and elimination of reactive species and effects of oxidative stress are also addressed, as well as summaries of recent works on the signaling role of reactive species in eukaryotic organisms. The readers will gain an overview of our current understanding of homeostasis of reactive species and cellular processes they are involved in, as well as useful resources for further reading.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Elisabete Silva and Patrício Soares-da-Silva (2012). Renal Redox Balance and Na⁺, K⁺-ATPase Regulation: Role in Physiology and Pathophysiology, *Oxidative Stress - Molecular Mechanisms and Biological Effects*, Dr. Volodymyr Lushchak (Ed.), ISBN: 978-953-51-0554-1, InTech, Available from:
<http://www.intechopen.com/books/oxidative-stress-molecular-mechanisms-and-biological-effects/renal-redox-balance-and-na-k-atpase-regulation-role-in-physiology-and-pathophysiology>

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