

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Principles and Methodologies for the Determination of Shelf-Life in Foods

Antonio Valero¹, Elena Carrasco^{1,2} and Rosa M^a García-Gimeno¹

¹*University of Cordoba*

²*Centro Tecnológico del Cárnico (TEICA)
Spain*

1. Introduction

The establishment of validated methodologies for the determination of food shelf-life is currently demanded by both food industries and Health Authorities at national and international scale. It is well known that most foods are perishable, since they are subjected to modifications in their structure, composition and properties during storage before consumption. These changes are of physico-chemical origin attributed to food composition together with the action of intrinsic and extrinsic environmental factors, and also microbiological, where spoilage flora play an important role. These modifications are “translated into” sensorial deterioration at a specific time point. In this respect, food-borne bacteria, despite representing a threat for consumers’ health, do not affect sensorial changes.

Product “shelf-life” is defined, according to the American Heritage Dictionary of the English Language (Mifflin, 2006) as “the term or period during which a stored commodity remains effective, useful, or suitable for consumption”. But, which is understood by “unsuitable”? Mifflin (2006) defines “unsuitability” as “the quality of having the wrong properties for a specific purpose”. To know which is wrong and which is fine is not a straightforward question to answer, and often is subjected to individual perceptions. This issue is discussed later. Anyway, whichever the method to detect the unsuitability of a food product, once it is detected and established, its cause should be sought. Among the different elements which constitute and characterize a food product, generally only one is the responsible for the unsuitability of a product, namely, “the specific cause of unsuitability”. With this, going back to the definition of shelf-life, we could redefine “product shelf-life” in the food field as “the term or period a product may be stored before a specific element of the product makes it unsuitable for use or consumption”. This element could be of biological or physico-chemical nature.

In the last years, different procedures have been reported for the establishment of shelf-life, mainly based on the detection of microbial alteration, as well as physico-chemical and sensorial changes. The traditional approach consists of setting a cut-off point along the storage period at the time when any of the measured attributes exceeds a pre-established limit. Experimental work usually includes the storage of food product at different temperatures, performance of microbial analysis and the assessment of spoilage by

sensorial testing. In the case of foods whose shelf-lives might be conditioned by the presence and proliferation of pathogenic microorganisms, experiments also involve challenge testing with the target organism prior to storage. The cut-off point has been traditionally referred as quality limit (if deterioration of food is known to be produced by physico-chemical changes), or safety limit (if deterioration of food is due to the presence of nocive chemical substances and/or pathogenic microorganisms, parasites or virus at levels of concern). This method is usually labour-intensive and expensive.

Regarding microbiological proliferation of spoilage and/or pathogenic microorganisms, predictive microbiology is recognized as a reliable tool for providing an estimation of the course of the bacteria in the foods, and indirectly, provide an estimation of shelf-life of the product in the cases when the cause of food spoilage or unacceptability is known to be microbiological. Indeed, mathematical modelling is a science-based discipline which aims to explain a reality with a few variables, and whose applications have been extended beyond research as a real added-value industrial application (Brul et al., 2007; McMeekin et al., 2002; Peleg, 2006; McMeekin, 2007).

The main concept behind the application of predictive microbiology for the determination of shelf-life based on spoilage is the specific spoilage organisms (SSO), which are associated to sensorial changes and spoilage. As such, the end of shelf-life can be defined as the time needed for SSO to multiply from an initial contamination level to a spoilage level, or the time invested by SSO to produce a certain metabolite causing sensorial rejection (Koutsoumanis & Nychas, 2000). In the case of pathogens, challenge test protocols are available for the determination of kinetic parameters (named maximum growth rate [μ_{\max}] and lag time [lag]) of *Listeria monocytogenes* in ready-to-eat foods (SANCO, 2008). In addition, European Regulation No. 2073/2005, recommends the performance of predictive microbiology studies in order to investigate compliance with the criteria throughout the shelf-life. In particular, this applies to ready-to-eat foods that are able to support the growth of *L. monocytogenes* and that may pose a *L. monocytogenes* risk for public health. In general, proliferation of pathogens for which absence is required although might be present in foods, growth/no growth models are generally accepted as useful tools for the determination of the probability of growth, whereas those pathogens for which hazardous levels (i.e. *Bacillus cereus*) or toxin-producing organisms (i.e. *Staphylococcus aureus*) have been set, growth kinetic models are more appropriate to estimate the time until reaching such levels.

Quality, in a very broad sense, means satisfaction of consumers' expectations; in other words, quality experience delivered by a food should match quality expectations of a consumer (van Boekel, 2008). Quality aspects of foods, such as colour, nutrient content, chemical composition etc., are governed by biochemical reactions (oxidation, Maillard reactions, enzyme activity) together with physical changes (aggregation of proteins, coalescence, sedimentation etc.). As for microbials, there are kinetic models for quality attributes. However, these models only provide a representation of single biochemical reactions within a well-diluted ideal system; thus, they are not easily extrapolated to other more complex matrices like foods.

Sensory testing is designed to validate the length of time that a product will remain with the same "acceptable quality" level or presents "no changes in desired sensory characteristics" over the entire life of a product (IFTS, 1993; Kilcast & Subramaniam, 2000). Some product properties are difficult to measure objectively. Moreover, instrumental measurement alone

cannot indicate consumer acceptability or rejection. It is very important to ensure no changes in sensory properties of foods during their shelf-lives, since consumers pay for a unconsciously established set of desired sensory characteristics.

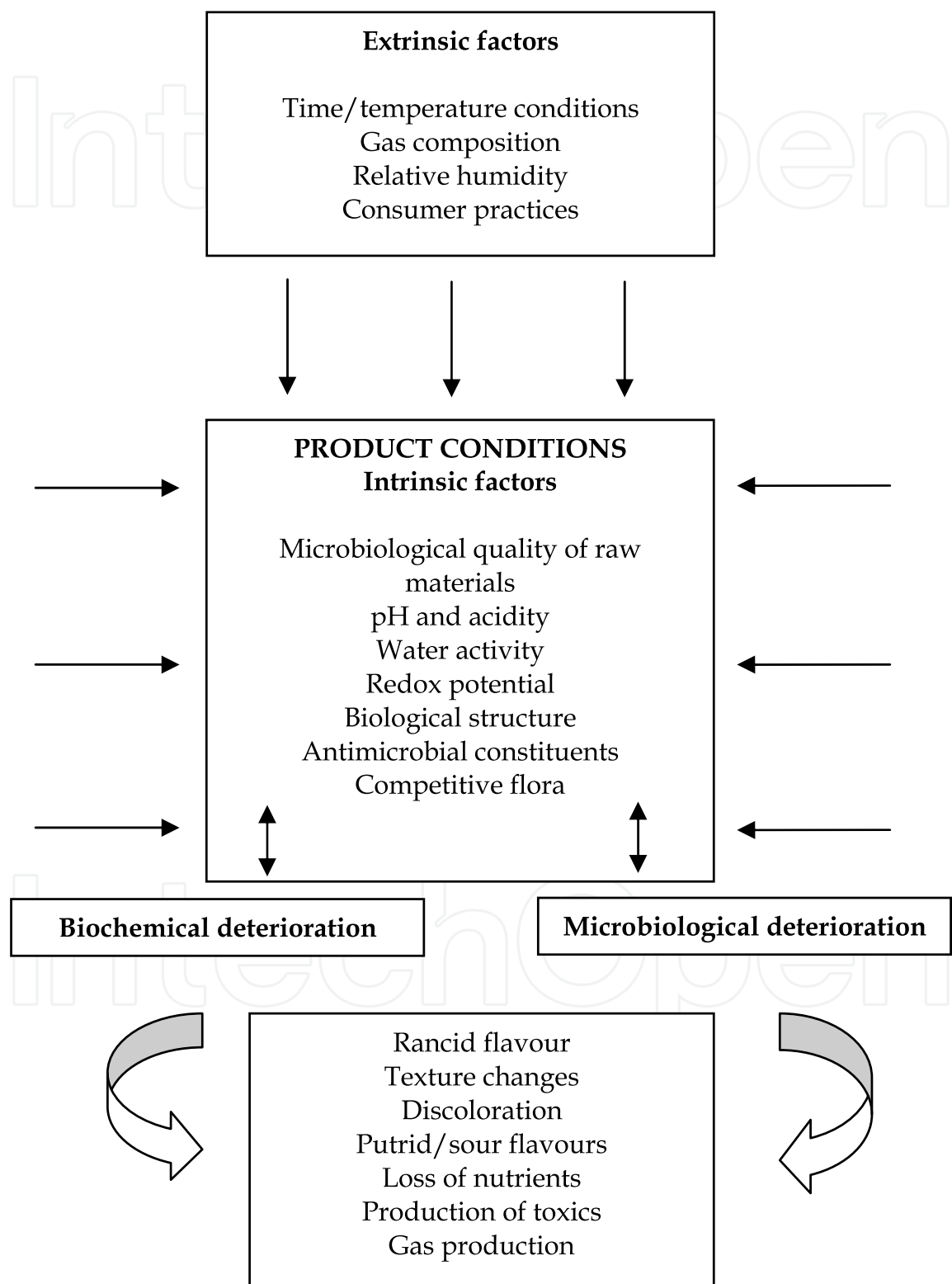


Fig. 1. Deterioration processes during food storage (adapted from Huis in't Veld, 1996)

A unifying description of the interaction between microflora behaviour and physico-chemical changes undertaken in a food, represents a special challenge for food technologists. Such an integrated understanding of the interactions occurring during food storage should motivate the development of alternative preservation systems. A schematic representation of these phenomena is presented in Figure 1.

Throughout this chapter, the principles and methodologies for the establishment of shelf-life of foodstuffs are discussed. Also microbial, sensorial and physico-chemical parameters influencing shelf-life are analyzed. Subsequently, foods testing for data generation as well as procedures for assessment of shelf-life are reviewed. Finally, we will present some comments on future prospects.

2. Environmental factors affecting microbial shelf-life

Food spoilage is greatly influenced by environmental conditions concerning the food matrix, microbial characteristics, temperature, pH, water activity (a_w), processing time, etc. The main objective of studying the influence of environmental factors in food preservation is to inhibit spoilage due to microbial survival and growth and/or occurrence of chemical reactions. For this, a number of factors must be evaluated for different foods which could play an important role in the Hazard Analysis and of Critical Control Points (HACCP) system.

Regarding microbial growth, environmental conditions can affect largely the microbial load along the food chain. They can be classified as:

- Physical factors, such as temperature, food matrix.
- Chemical factors, such as pH, preservatives, etc.
- Biological factors, such as competitive flora, production of metabolites or inhibiting compounds etc.
- Processing conditions affecting foods (slicing, mixing, removing, washing, shredding etc.) as well as influencing transfer of microorganisms (cross-contamination events).

The application of one these factors alone can produce the desired effect on the food in terms of quality and safety, but this is not usual, especially in processed, ready-to-eat or perishable foods. Generally, the establishment of an adequate combination of more than one factor at moderate levels can offer the same result, but an improvement of the sensorial characteristics is achieved. This is the basic idea underlying the hurdle technology concept stated by Leistner (1995). Increasing the severity of one factor alone could produce a negative effect on food quality (e.g. chilling injury), while a moderate combination of several factors can lead to a shelf-life increase maintaining the original sensorial properties. For instance, in meat products, barriers such as the addition of salt, nitrite, modified atmosphere packaging, etc., can reduce the survival of pathogens (if present) and the proliferation of lactic acid bacteria during shelf-life.

2.1 Intrinsic factors

2.1.1 Microbiological quality of raw materials

Raw material entering the food industry represents a potential source of microbial contamination. The potential growth of pathogens and spoilage flora will be affected by the initial level of contamination and the efficacy of processing steps in eliminating bacteria in

the food. Figure 2 represents the effect of various initial contamination levels on shelf-life of foods. At high contamination levels, less time would be needed by SSOs to reach the minimum spoilage level, thus, shelf-life would have to be reduced.

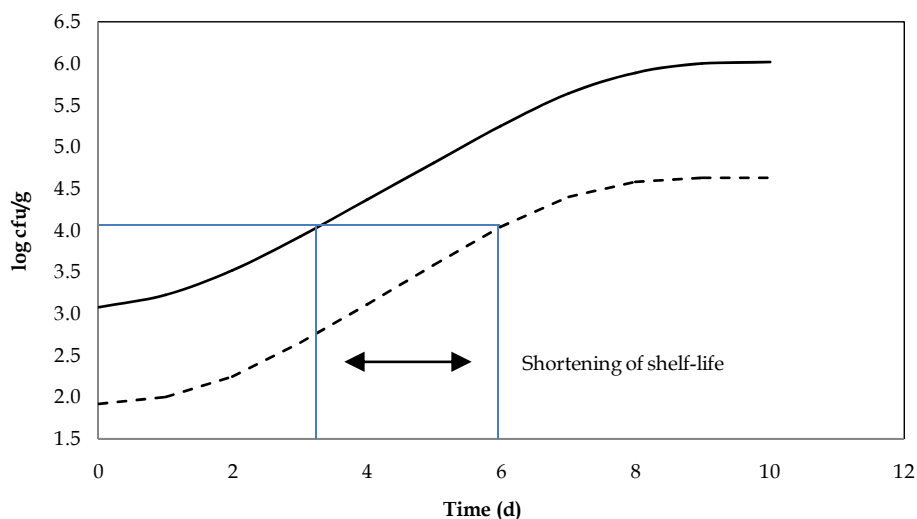


Fig. 2. Effect of the microbial initial contamination on shelf-life in a food.

A better raw material quality in terms of microbial contamination can be achieved by setting out a more strict suppliers control at primary production level, and optimizing sampling schemes. With an initial good quality of raw materials, processing operations, formulation of foods and storage conditions, shelf-life of foods could be extended.

2.1.2 pH and acidity

A widely used preservation method consists of increasing the acidity of foods either through fermentation processes or the addition of weak acids. The pH is a measure of the product acidity and is a function of the hydrogen ion concentration in the food product. It is well known that groups of microorganisms have pH optimum, minimum and maximum for growth in foods. Bacteria normally grow faster between pH ranges of 6.0 - 8.0, yeasts between 4.5 - 6.0 and moulds between 3.5 - 4.0.

An important characteristic of a food is its buffering capacity, i.e. its ability to resist changes in pH. Foods with a low buffering capacity will change pH quickly in response to acidic or alkaline compounds produced by microorganisms, whereas foods with high buffering capacity are more resistant to such changes. In any case, if low pH is a factor included in the preservation system of a food, control of pH and the application of a margin of safety are required for these foods.

2.1.3 Water activity (a_w)

The requirements for moisture by microorganisms are expressed in terms of water activity (a_w). The a_w is therefore one of the most important properties of food governing microbial growth and is defined as the free or available water in a food product (Fennema, 1996). Thus, the a_w of a food describes the fraction of water “not bounded” to the components of a

food, i.e. the fraction of “available” water to participate in chemical/biochemical reactions and promote microbial survival and growth.

As for pH, control of a_w and the application of a margin of safety are required for these food products whose preservation system includes low a_w . Microorganisms respond differently to a_w depending on a number of factors. These factors can modify the minimum and maximum a_w values to grow. Generally, Gram (-) bacteria are more sensitive to changes in a_w than Gram (+) bacteria. The growth of most foodborne pathogens is inhibited at a_w values below 0.86.

2.1.4 Redox potential (E_h)

The oxidation-reduction potential (E_h) of a food is the ease by which it gains or loses electrons (Jay, 1992). The E_h value at which microorganisms will grow determines whether they require oxygen (i.e. aerobic/microaerophilic environment) for growth or not (i.e. anaerobic environment). According to their E_h value, microorganisms can be classified into the three following groups:

Aerobes	+500 to +300 mV;
Anaerobes	+100 to -250 mV; and
Facultative anaerobes	+300 to -100 mV

Although E_h as inhibitory factor is especially important in meat products (Leistner, 2000), product safety control should not solely focus on this factor since its measurement is subjected to limitations. Indeed, the E_h values are highly variable depending on the pH of the food, extent of microbial growth, packaging conditions, oxygen partial pressure in the storage environment, and ingredients. Measurement requirements for E_h in foods are reported by Morris (2000).

2.1.5 Biological structure

The natural surfaces of foods usually provide high protection against the entry and subsequent damage by spoilage organisms. External layers of seeds, the outer covering of fruits, and the eggshell are examples of biological protective structures. Several factors can influence the penetration of organisms through these barriers: (i) the maturity of plant foods enhances the effectiveness of the protective barriers; (ii) physical damage due to handling during harvest, transport, or storage, as well as invasion of insects can allow the penetration of microorganisms (Mossel et al., 1995); (iii) during the preparation of foods, processes such as slicing, chopping, grinding, and shucking destroy the physical barriers, thus favouring contamination inside the food.

Eggs can be seen as a good example of an effective biological structure that, when intact, will prevent external microbial contamination of the perishable yolk. Contamination by *Salmonella*, one of the most prevalent contaminant, is possible through transovarian infection before this shell structure is established. Additional factor is the egg white and its antimicrobial components. When there are cracks through the inner membrane of the egg, microorganisms may further penetrate into the egg. Factors such as storage temperature, relative humidity, age of eggs, and level of surface contamination will influence the internalization of microorganisms.

2.1.6 Antimicrobial constituents

Food products may contain substances (i.e. antimicrobial constituents) which have antimicrobial properties against the growth of specific microorganisms. There is a wide variety of substances with recognized antimicrobial activity in a wide variety of food products. Other antimicrobial constituents in food products are added as preservatives. Some forms of food processing will also result in the formation of antimicrobial substances in food products including:

- Smoking, e.g. fish and meat products;
- Fermentation, e.g. meat and dairy products;
- Condensation reactions between sugars and amino acids (i.e. Maillard reaction) during heating of certain foods.

2.1.7 Competitive flora (biopreservation)

Biopreservation techniques are the result of the development of a microorganism which may have an antagonistic effect on the microbial activity of other undesired microorganisms present in the food product (Mossel et al., 1995). There has been an increasing interest in biopreservation treatments in minimally processed foods in order to guarantee food safety whilst maintaining organoleptical properties.

Antagonistic processes include competition for essential nutrients, changes in pH value or E_h or the formation of antimicrobial substances like bacteriocins, which may negatively affect the survival or growth of other microorganisms (Huis in't Veld et al., 1996). Bacteriocins have been widely recognized as natural food biopreservatives, and latest advances on bacteriocin research have opened new fields to explore their role (García et al., 2010). The use of competitive microorganisms such as lactic acid bacteria against *L. monocytogenes* has been proven as a useful preservation technique in several food products (Brillet et al., 2005).

2.2 Extrinsic factors

2.2.1 Time/temperature conditions

All microorganisms have a defined temperature range within which they can grow. This range is limited by a minimum temperature needed for growth and a maximum temperature which does not support the growth. While the growth speed increases with increasing temperature, it tends to decline rapidly after the optimum temperature has been reached. An understanding of the interplay between time, temperature, and other intrinsic and extrinsic factors is crucial to select appropriate storage conditions for a food product. Temperature has a dramatic impact on the growth of microorganisms. Time plays an important role as a factor facilitating (long time) or avoiding (short time) the growth of microorganisms during food storage.

At low temperatures, two phenomena avoid microbial growth:

- Intracellular reactions, which become much slower; and
- Fluidity of the cytoplasmic membrane, interfering with transport mechanisms (Mossel et al., 1995).

At temperatures above the maximum temperature, structural cell components become denatured and heat-sensitive enzymes are inactivated.

2.2.2 Gas composition

Many scientific studies have demonstrated the antimicrobial activity of gases at ambient and sub-ambient pressures in foods (Loss & Hotchkiss, 2002). The oxygen availability is related to the E_h and plays a role in product packaging, together with CO_2 concentration. A variety of common technologies are used to inhibit the growth of microorganisms, and a majority of these methods have shown high inhibitory effect when applied with low temperatures. Technologies include Modified Atmosphere Packaging (MAP), Controlled Atmosphere Packaging (CAP), Controlled Atmosphere Storage (CAS) or Direct Addition of Carbon Dioxide (DAC) (Loss & Hotchkiss, 2002).

An adequate packaging formulation in food products should extend shelf-life without affecting their sensorial properties. In general, the inhibitory effect of CO_2 increases at low temperatures due to the fact that solubility of CO_2 becomes higher (Jay, 2000). Also, a synergistic effect has been observed when CO_2 has been applied together with low pH. The major concern when extending the shelf-life of a product by MAP is that the inhibition of spoilage bacteria could allow the proliferation of foodborne pathogens as they do not compete for nutrients or physical space anymore, and consequently foods, in spite of presenting an acceptable sensorial quality, may be unsafe.

There are several additional intrinsic and extrinsic factors affecting the efficacy of antimicrobial atmospheres, such as product temperature, product-to-headspace gas volume ratio, initial microbial load, package barrier properties, biochemical composition of the food, etc. By combining antimicrobial atmospheres with other preservation techniques, further enhancement of food quality and safety can be achieved (Leistner, 2000).

2.2.3 Relative humidity (RH)

Relative humidity (RH) is the quantity of moisture in the atmosphere surrounding a food product whether packaged or not. It is calculated as a percentage of the humidity required to completely saturate the atmosphere (i.e. saturation humidity). Typically, there will be an exchange of moisture between a food product and the surrounding atmosphere which continues until the food reaches equilibrium.

The RH is closely related with a_w and may actually alter the a_w of a food. It is important to assure that the product is stored with an environment where the RH prevents a_w changes. For example, if the a_w of a food is set at 0.60 to assure its stability, it is important that this a_w value remains during storage by establishing adequate RH conditions so that the food does not pick up moisture from the ambient air. When foods with low RH values are placed in environments of high RH, they pick up moisture until equilibrium is established. Likewise, foods with high a_w lose moisture when placed in an environment of low RH.

Determining the appropriate storage/packaging conditions for a food product is therefore essential for food safety and quality assurance. It is important to note that RH is related to temperature during storage (Esse & Humidipack, 2004).

2.2.4 Consumer practices

Consumer practices during dispatch, storage and use of food products in the home are obviously outside the control of food industries. However, food industries should take account of poor consumer handling in the establishment of product shelf-life, at least, those known to be popular. For example, many domestic refrigerators do not operate at refrigeration temperatures between 0-5°C, being above this range; this can greatly affect the safety of food products and shorten their shelf-life (Carrasco et al., 2007). Food industries should wisely establish food shelf-lives according to real temperature records of domestic refrigerators, and not to theoretical refrigeration temperatures.

3. Role of spoilage microorganisms in foods

Foods are dynamic systems which experience changes in pH, atmosphere, nutrient composition and microflora over time. Each food product has its own unique flora, determined by the raw materials used, food processing parameters and subsequent storage conditions. The growth of SSOs in perishable foods is largely affected by several environmental factors already explained in the Section 2.

The main purpose of the application of one or a combination of intrinsic and extrinsic factors on a food product is to destroy and/or inhibit SSOs and pathogenic microorganisms, thus providing a more extended shelf-life, or to improve the sensorial characteristics of the food.

The microorganisms dominating a product can be predicted by understanding how the major preservation parameters affect microbial selection. The identification of SSOs are very important for the calculation of the remaining shelf-life of a given food product (Gram et al., 2002). Some examples are *Lactobacillus sakei* in cooked meat products (Devlieghere et al., 2001), *Photobacterium phosphoreum*, *Shewanella* spp and *Pseudomonas* spp in fishery products (Dalgaard et al., 1997; Gram and Dalgaard, 2002), *Brochothrix thermosphacta* in precooked chicken (Patsias et al., 2006), or gram negative bacteria in fresh-cut (Rico et al., 2007).

It is crucial to introduce quantitative considerations (Gram, 1989) since the spoilage activity of an organism is its quantitative ability to produce spoilage metabolites (Dalgaard, 1995; Dalgaard et al., 1993). In other words, it is necessary to evaluate if the levels of a particular organism reached in naturally spoiled foods are capable of producing the amount of metabolites associated with spoilage. In general, it requires a careful combination of microbiology, sensory analyses and chemistry to determine which microorganism(s) are the SSOs of a particular food product.

It is generally known that almost all microbial groups include species that could cause spoilage in a given food under specific conditions. Initially, different populations are present in a contaminated food, and some groups become more predominant after a certain storage period. This selection might depend on several physico-chemical factors (as discussed previously) and processing and storage conditions. For instance, in meat products stored at aerobic conditions, *Pseudomonas* spp. can produce metabolites and slime formation, but in an anaerobic atmosphere, the predominant SSOs are shift to lactic-acid bacteria, producing lactic acid and decreasing pH.

Vegetables are a special case due to the nutrient composition. The high pH (usually close to neutrality) will allow a range of Gram-negative bacteria to grow, but spoilage is specifically caused by organisms capable of degrading the vegetable polymer, pectin (Liao et al., 1997). These organisms, typically *Erwinia* spp. and *Pseudomonas* spp. are the SSO of several ready-to-eat vegetable products (Lund, 1992); also pectin degrading fungi can play a role in vegetable products (Pitt & Hocking, 1997). However, when decreasing pH due to the addition of organic acids in minimally processed fruits and vegetables, growth of fungi, yeasts and lactic-acid bacteria become the SSOs (Edwards et al., 1998).

After identification of the SSOs and the range of environmental conditions under which a particular SSO is responsible for spoilage, the next step in microbial shelf-life establishment is the decision about the microbial level of SSO above which spoilage occurs and shelf-life ends (Dalgaard, 1995; Koutsoumanis & Nychas, 2000). This step often requires a good understanding of the microbial evolution as a function of time. Initially, SSO is present in low quantities and constitutes only a minor part of the natural microflora. During storage, SSO generally produce the metabolites responsible for off-odours, off-flavours slime and finally cause sensory rejection. The cell concentration of SSO at rejection may be called the Minimal Spoilage Level (MSL) and the concentration of metabolites corresponding to spoilage is named Chemical Spoilage Index (CSI) (Dalgaard, 1993).

An example of these concepts is presented in Figure 3. In this case, the evolution of SSO together with histamine content occurs during storage of a seafood product. If MSL is set at 4 log cfu/g (if this limit is proven to cause food spoilage), the established shelf-life would be 4.2 days. However, this limit is largely different if the results are based on the histamine content, because if CSI is set at 200 ppm, the shelf-life would be 8.8 days. Generally, shelf-life may be determined based on the parameter that firstly produces alteration, in this example, microbial growth.

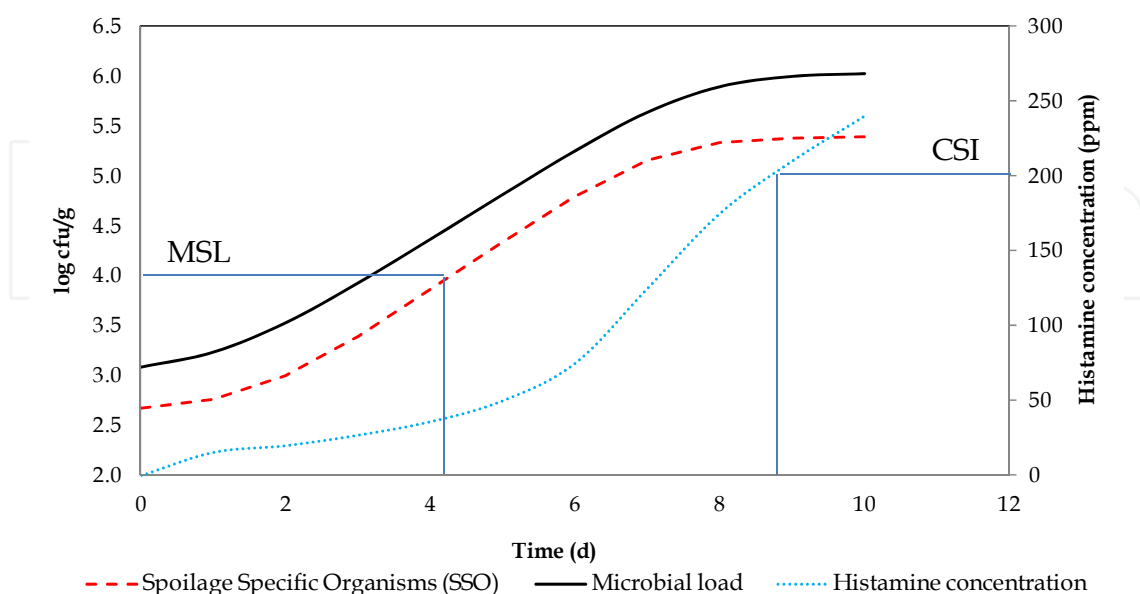


Fig. 3. Theoretical growth of SSO together with evolution of histamine content in a seafood product. MSL (Microbial Spoilage Level), CSI (Chemical Spoilage Index).

The limit of microbial growth that determines shelf-life differs according to the food type and storage conditions. SSO counts from 10^5 to 10^8 cfu/g are commonly considered as convenient quality limits. For microorganisms which produce toxins, 10^5 cfu/g has been established as a limit for risk management in processed foods (Rho & Schaffner, 2007). The time to reach hazardous levels by pathogens should determine the shelf-life limit. In these cases, shelf-life is greatly influenced by the initial contamination level. Thus, hygienic control measures must be taken in order to prevent their presence in raw material and processed food products.

Several studies relate limits for microbial groups defining the end of shelf-life with specific food products. In this sense, 10^7 cfu/g has been chosen for aerobic bacteria in minced chicken meat (Grandinson & Jennings, 1993), freshwater crayfish (Wang & Brown, 1983), or fresh-cut lettuce (Koseki & Ito, 2002). For other microbial groups such as coliforms in cottage cheese, yeasts in shredded chicory endive, or mesophilic bacteria in fresh-cut spinach, limit levels of 10^2 , 10^5 or 10^8 cfu/g, respectively, have been proposed (Babic & Watada, 1996; Jacxsens et al., 2001; Mannheim & Soffer, 1996). To set these limits, a thorough knowledge about the particular microbial ecology and deterioration causes of foods during storage is required.

In relation with the microbiological limits previously mentioned, Regulation (EC) No. 2073/2005 regarding microbiological criteria in foodstuffs sets out two groups of criteria:

- Food safety criteria which define the acceptability of a product or a batch. They are applicable to foodstuffs placed on the market and throughout the shelf-life of the food.
- Process hygiene criteria which define the acceptability of the process. These apply only during the manufacturing process.

Other microbiological criteria for the end of shelf-life have been suggested by International Guidelines in Ready-To-Eat (RTE) foods. Some guidance documents are mainly focused on the study of growth of *L. monocytogenes* (SANCO, 2008) while other more general reports assess the microbiological quality of RTE foods placed in the market (Health Protection Agency, 2009; Refrigerated Food Association, 2009) or recommend specific principles to food industries (Food Safety Authority of Ireland, 2005; New Zealand Food Safety Authority, 2005). All of them establish microbiological criteria by specifying numerical limits, or admissible log increases during shelf-life.

4. Mathematical modelling of bacterial growth

It is a goal of food microbiologists to know in advance the behavior of microorganisms in foods under foreseeable conditions. If it was possible to predict the growth/survival/inactivation of a microorganisms at given fixed conditions, it would be possible to systematically establish a shelf-life based on bacterial behaviour. The development of mathematical models describing the behavior of microorganisms under established conditions satisfactorily fulfill the challenge proposed. Already in 1921, Bigelow (1921) described the logarithm nature of thermal death, but it was not until the 1980s when mathematical models for predicting bacterial behavior experienced a great development. A new field emerged in the food microbiology area: "predictive microbiology", or as redefined recently, "modeling of microbial responses in foods". McKellar & Lu (2004a) presented a detailed review of predictive models published so far. Since primary to tertiary models,

mathematical models have been extensively applied, in part due to the predictive friendly-software developed, where most models are implemented.

The advent of computer technology and associated advances in computational power have made possible to perform complex mathematical calculations that otherwise would be too time-consuming for useful applications in predictive microbiology (Tamplin et al., 2004). Computer software programs provide an interface between the underlying mathematics and the user, allowing model inputs to be entered and estimates to be observed through simplified graphical outputs. Behind predictive software programs are the raw data upon which the models are built. Some of the most popular software are: GInaFiT (Geeraerd, et al., 2005), where different inactivation models are available (<http://cit.kuleuven.be/biotec/downloads.php>); DMFit (Baranyi & Roberts, 1994), with implementation of a dynamic growth primary model (<http://www.combase.cc/index.php/en/downloads/category/11-dmfit>); Pathogen Modeling Program (Buchanan, 1993), which incorporates a variety of models of different pathogens in broth culture and foods (<http://pmp.arserrc.gov/PMPOnline.aspx>); Seafood Spoilage and Safety Predictor (SSSP) (Dalgaard et al., 2002), offering models for specific spoilage microorganisms and also for *L. monocytogenes* in seafood (<http://sssp.dtuaqua.dk/>); ComBase (Baranyi & Tamplin, 2004), a predictive tool for important foodborne pathogenic and spoilage microorganisms (<http://www.combase.cc/index.php/en/predictive-models>); Sym'Previus (Leporq et al., 2005), a tool with a collection of models and data to be applied in the food industry context, e.g. strengthening HACCP plans, developing new products, quantifying microbial behavior, determining shelf-lives and improving safety (<http://www.symprevius.net/>); Microbial Responses Viewer (Koseki, 2009), a new database consisting of microbial growth/no growth data derived from ComBase, and also modelling of the specific growth rate of microorganisms as a function of temperature, pH and a_w (<http://mrv.nfri.affrc.go.jp/Default.aspx#/About>); *Escherichia coli* fermented meat model (Ross & Shadbolt, 2004), describing the rate of inactivation of *Escherichia coli*, due to low a_w or pH or both, in fermented meats (<http://www.foodsafetycentre.com.au/fermenter.php>).

In a further step, a number of software called expert systems have been developed to provide more complex decision support based on a set of rules and algorithms for inference based on the relationships underlying these rules. In these systems, the core knowledge is stored as a series of IF-THEN rules that connect diverse evidence such as user input, data from databases, and the formalized opinions of experts into a web of knowledge. There are various examples of decision-support systems which encapsulate knowledge that is based in predictive microbiology, such as those applied in predicting food safety and shelf-life (Witjes et al., 1998; Zwietering et al., 1992); a step-wise system structured as a standard risk assessment process to assist in decisions regarding microbiological food safety (van Gerwen et al., 2000); or other systems for microbial processes described by Voyer & McKellar (1993) and Schellekens et al. (1994).

Whiting & Buchanan (1994) called the above integrated software models-based "tertiary models". They defined tertiary-level models as personal computer software packages that use the pertinent information from primary- and secondary-level models to generate desired graphs, predictions and comparisons. "Primary-level models" describe the change in microbial numbers over time, and "secondary-level models" indicate how the features of primary models change with respect to one or more environmental factors, such as pH,

temperature and a_w . Below is a description of the most relevant primary and secondary models, as well as their uses and scope.

4.1 Primary models

Primary models aim to describe the kinetics of the process (growth or survival or inactivation) with as few parameters as possible, while still being able to accurately define the distinct stages of the process.

In the context of foods shelf-life, the process observed depends on the group of microorganisms studied (e.g. total aerobic flora, lactic acid bacteria, *Enterobacteriaceae*, etc.) and the characteristics and storage conditions of foods (e.g. presence of antimicrobials like nitrites, a_w , pH, temperature of storage, atmosphere inside the food package, etc.). Often, different groups of microorganisms behave differently in the same food product. For example, in fermented meat products, it is usual to observe growth of lactic acid bacteria against the survival or inactivation of other bacteria groups (González & Díez, 2002; Moretti et al., 2004; Rubio et al., 2007).

The application of primary models to a set of microbiological data proceeds as follows. In a first step, a mathematical model is assumed to explain the data, that is, how microbial counts change over time. In a second step, such model is fitted to microbiological data by means of regression (linear or non-linear). As a consequence of the fitting process, estimates for a number of kinetics parameters embedded in the model is provided, for example, for the rate of growth/inactivation or lag time (in growth processes) or “shoulder” (in inactivation processes). The dataset used to fit the model is assumed to be obtained under specific intrinsic and extrinsic factors. For this reason, the kinetic parameters provided after fitting solely apply for the specific intrinsic and extrinsic factors characterizing the dataset.

Several primary models have been published. Below is described the most important models used for growth and survival/inactivation.

4.1.1 Growth models

The first book devoted exclusively to the field of predictive microbiology (McMeekin et al., 1993) provide an excellent review and discussion of the classical sigmoid growth functions, especially the modified logistic and Gompertz equations. As they point out, these are empirical applications of the original logistic and Gompertz functions. Over the last years, a new generation of bacterial growth curve models have been developed that are purported to have a mechanistic basis: for example, the Baranyi model (Baranyi et al., 1993; Baranyi et al., 1994), the Hills model (Hills & Mackey, 1995; Hills & Wright, 1996), the Buchanan model (Buchanan et al., 1997), and the heterogeneous population model (McKellar, 1997).

- a. Sigmoidal functions such as the modified logistic (Equation 1) and the modified Gompertz (Equation 2) introduced by Gibson et al. (1987), have been the most popular ones used to fit microbial growth data since these functions consist of four phases, similar to the microbial growth curve.

$$\text{Log}x(t) = A + \left[C / \left(1 + e^{(-B(t-M))} \right) \right] \quad (1)$$

$$\text{Log}x(t) = A + C \exp\{-\exp[-B(t - M)]\} \quad (2)$$

where $x(t)$ is the number of cells at time t , A is the lower asymptotic value as t decreases to zero, C is the difference between the upper and lower asymptote, M is the time at which the absolute growth rate is maximum, and B is the relative growth rate at M .

The parameters of the modified Gompertz equation (A , C , B and M) can be used to characterize bacterial growth as follows:

$$\text{lag} = M - 1/B + [(\text{Log}N(0) - A)/(BC/e)] \quad (3)$$

$$\text{specific growth rate} = BC/e \gg BC/2.18 \quad (4)$$

In order to simplify the fitting process, reparameterized versions of the Gompertz equation have been proposed (Zwietering et al., 1990; Willox et al., 1993):

$$\text{Log}_{10}x = A + C \exp\left\{-\exp\left[2.71(R_g/C)(\lambda - t) + 1\right]\right\} \quad (5)$$

where $A = \log_{10} x_0$ (\log_{10} cfu \times ml⁻¹), x_0 is the initial cell number, C the asymptotic increase in population density (\log_{10} cfu \times ml⁻¹), R_g the growth rate (\log_{10} cfu h⁻¹), and λ is the lag-phase duration (h).

Although used extensively, some authors (Whiting & Cygnarowicz-Provost, 1992; Baranyi, 1992; Dalgaard et al., 1994; Membré et al., 1999) reported that the Gompertz equation systematically overestimated growth rate compared with the usual definition of the maximum growth rate. Note that there is also no correspondence between the lower asymptote and the inoculum size x_0 . Baty et al. (2004), while comparing the ability of different primary models to estimate lag time, found that the Gompertz model is perhaps the least consistent. Nevertheless, this model is one of the most widely used to fit bacterial curves, notably by the Pathogen Modeling Program (Buchanan, 1993).

- b. Baranyi et al. (1993) and Baranyi & Roberts (1994, 1995) introduced a mechanistic model for bacterial growth (Equation 6). In this model, it is assumed that during lag phase, bacteria need to synthesize an unknown substrate q critical for growth. Once cells have adjusted to the environment, they grow exponentially until limited by restrictions dictated by the growth medium.

$$\frac{dx}{dt} = \frac{q(t)}{q(t) + 1} \cdot \mu_{\max} \cdot \left(1 - \left(\frac{x(t)}{x_{\max}}\right)^m\right) x(t) \quad (6)$$

where x is the number of cells at time t , x_{\max} the maximum cell density, and $q(t)$ is the concentration of limiting substrate, which changes with time (Equation 7). The parameter m characterizes the curvature before the stationary phase. When $m = 1$ the function reduces to a logistic curve, a simplification of the model that is often assumed.

$$\frac{dq}{dt} = \mu_{\max} \cdot q(t) \quad (7)$$

The initial value of q (q_0) is a measure of the initial physiological state of the cells. A more stable transformation of q_0 may be defined as:

$$h_0 = \ln \left(1 + \frac{1}{q_0} \right) = \mu_{\max} \lambda \quad (8)$$

Thus, the final model has four parameters: x_0 , the initial cell number; h_0 ; x_{\max} ; and μ_{\max} . The parameter h_0 describes an interpretation of the lag first formalized by Robinson *et al.* (1998). Using the terminology of Robinson *et al.* (1998), h_0 may be regarded as the “work to be done” by the bacterial cells to adapt to their new environment before commencing exponential growth at the rate, μ_{\max} , characteristic of the organism and the environment. The duration of the lag, however, also depends on the rate at which this work is done which is often assumed to be μ_{\max} .

- c. Hills & Mackey (1995) and Hills & Wright (1996) developed a theory of spatially dependent bacterial growth in heterogeneous systems, in which the transport of nutrients was described by the combination a structured-cell kinetic model with reaction-diffusion equations. The total biomass in the culture M at a time t is dependent on time and a rate constant A (Equation 9), while the total number of cells in the culture N depends on time and the rate constants A and K_n (Equation 10).

$$M(t) = M(0) \exp(At) \quad (9)$$

$$N(t) = N(0) [K_n \exp(At) + A \exp(-K_n t)] / (A + K_n) \quad (10)$$

The rate constants A and K_n depend on all the environment factors. The lag time and the doubling time have the following relationships:

$$t_{LAG} = A^{-1} - \log[1 + (A / K_n)] \quad (11)$$

$$t_{LAG} / t_D = (\ln 2)^{-1} \log[1 + (A / K_n)] \quad (12)$$

This shows that if the rate constants A and K_n have similar activation energies, the ratio of lag to doubling time (t_D) should be nearly independent of temperature. This model takes no account of possible lag behavior in the total biomass (M).

Hills model can also be generalized to spatially inhomogeneous systems such as food surfaces (Hills & Wright, 1996). If more detailed kinetic information on cell composition is available, more complex multicompartment kinetic schemes can be incorporated.

- d. Buchanan *et al.* (1997) proposed a three-phase linear model. It can be described by three phases: lag phase, exponential growth phase and stationary phase, described as follows:

Lag phase:

$$\text{For } t \leq t_{LAG}, N_t = N_0 \quad (13)$$

Exponential growth phase:

$$\text{For } t_{LAG} < t < t_{MAX}, N_t = N_0 + \mu(t - t_{LAG}) \quad (14)$$

Stationary phase:

$$\text{For } t > t_{MAX}, N_t = N_{MAX} \quad (15)$$

where N_t is the log of the population density at time t (log cfu ml⁻¹); N_0 the log of the initial population density (log cfu ml⁻¹); N_{MAX} the log of the maximum population density supported by the environment (log cfu ml⁻¹); t the elapsed time; t_{LAG} the time when the lag phase ends (h); and μ is the maximum growth rate (log cfu ml⁻¹ h⁻¹).

In this model, the growth rate was always at maximum between the end of the lag phase and the start of the stationary phase. The μ was set to zero during both the lag and stationary phases. The lag was divided into two periods: a period for adaptation to the new environment (t_a) and the time for generation of energy to produce biological components needed for cell replication (t_m).

- e. McKellar model assumes that bacterial population exists in two “compartments” or states: growing or nongrowing. All growth was assumed to originate from a small fraction of the total population of cells that are present in the growing compartment at $t = 0$. Subsequent growth is based on the following logistic equation:

$$\frac{dG}{dt} = G \cdot \mu \cdot \left(1 - \frac{G}{N_{MAX}}\right) \quad (16)$$

where G is the number of growing cells in the growing compartment. The majority of cells were considered not to contribute to growth, and remained in the nongrowing compartment, but were included in the total population. While this is an empirical model, it does account for the observation that growth in liquid culture is dominated by the first cells to begin growth, and that any cells that subsequently adapt to growth are of minimal importance (McKellar, 1997). The model derives from the theory that microbial populations are heterogeneous rather than homogeneous, existing two populations of cells that behave differently; the sum of the two populations effectively describes the transition from lag to exponential phase, and defines a new parameter G_0 , the initial population capable of growing. Reparameterization of the model led to the finding that a relationship existed between μ_{max} and λ as described in Baranyi model. In fact, Baranyi & Pin (2001) stated that the initial physiological state of the whole population could reside in a small subpopulation. Thus, the McKellar model constitutes a simplified version of the Baranyi model, and has the same parameters.

The concept of heterogeneity in cell populations was extended further to the development of a combined discrete-continuous simulation model for microbial growth (McKellar & Knight, 2000). At the start of a growth simulation, all of the cells were assigned to the nongrowing compartment. A distribution of individual cell lag times was used to generate a series of discrete events in which each cell was transferred from nongrowing to the growing compartment at a time corresponding to the lag time for that cell. Once in the growing compartment, cells start growing immediately according to Equation 16. The combination of the discrete step with the continuous growth function accurately described the transition from lag to exponential phase.

At the present time it is not possible to select one growth model as the most appropriate representation of bacterial growth (McKellar & Lu, 2004b). If simple is better, then the three-phase model is probably sufficient to represent fundamental growth parameters accurately (Garthright, 1997). The development of more complex models (and subsequently more mechanistic models) will depend on an improved understanding of cell behavior at the physiological level.

4.1.2 Survival/inactivation models

Models to describe microbial death due to heating have been used since the 1920s, and constitute one of the earliest forms of predictive microbiology. Much of the early work centered on the need to achieve the destruction of *Clostridium botulinum* spores. This type of models has undertaken great advances from the classical linear inactivation model to others more sophisticated nonlinear models.

- a. The classical linear model assumes that inactivation is explained by a simple first-order reaction kinetics under isothermal conditions (Equation 17):

$$\frac{dS_t}{dt} = -k'S_t \quad (17)$$

where S_t is the survival ratio (N_t/N_0) and k' is the rate constant. Thus the number of surviving cells decreases exponentially:

$$S_t = e^{-k't} \quad (18)$$

and when expressed as \log_{10} , gives:

$$\log S_t = -kt \quad (19)$$

where $k = k'/\ln 10$. The well-known *D-value* (time required for a 1-log reduction) is thus equal to $1/k$, where k is the slope. The *D-value* can also be expressed as:

$$D\text{-value} = \frac{t}{\log N_0 - \log N_t} \quad (20)$$

When $\log D\text{-values}$ are plotted against the corresponding temperatures, the reciprocal of the slope is equal to the *z-value*, which is the increase in temperature required for a 1-log decrease in *D-value*. The rate constant can also be related to the temperature by the Arrhenius equation:

$$k = N_0 e^{\left(-\frac{E_a}{RT}\right)} \quad (21)$$

where E_a is the activation energy, R the universal gas constant, and T is the temperature in Kelvin degrees.

Extensive work has been performed to calculate *D-values* and the *z-value* for most pathogens in broth culture and different food matrices (Mazzota, 2001; Murphy et al., 2002; van Asselt & Zwietering, 2006).

- b. Although the concept of logarithmic death is still applied, nonlinear curves have been reported for many years, i.e. curves exhibiting a “shoulder” region at the beginning of the inactivation curve and/or a “tail” region at the end of the inactivation curve. Stringer et al. (2000) summarized the possible explanations for this behavior. Some of them are the variability in heating procedure, the use of mixed cultures, clumping, protective effect of dead cells, multiple hit mechanisms, natural distribution of heat sensitivity or heat adaptation.

We now know that cells do not exist simply as alive or dead, but may also experience various degrees of injury or sublethal damage, which may give rise to apparent nonlinear survival curves (Stringer et al., 2000). Survival modeling should also include a more complete understanding of the molecular events underpinning microbial resistance to the environment.

As heterogeneity is the most plausible reason for observing nonlinearity, the use of distributions to account for this nonlinearity such as gamma (Takumi et al., 2006) or Weibull (Coroller et al., 2006; van Boekel, 2002) have been suggested; from these, Weibull is the favored approach at the moment.

Also, “mirror images” of the primary growth models described above have been used to explain the inactivation process, such as the “mirror image” of the logistic function (Cole et al., 1993; Pruitt et al., 1993; Whiting, 1993), the “mirror image” of the Gompertz function (Linton et al., 1995) and the “mirror image” of the Baranyi model (Koutsoumanis et al., 1999).

4.2 Secondary models

These models predict the changes in the parameters of primary models such as bacterial growth rate and lag time as a function of the intrinsic and extrinsic factors. A description of the most popular secondary models is provided below. Two different approaches can be distinguished: i) the effects of the environmental factors are described simultaneously through a polynomial function; this type of model has probably been the most extensively used within predictive microbiology; and ii) the environmental factors are individually modeled, and a general model describes the combined effects of the factors; this approach is notably applied in the development of the increasingly popular square root and cardinal parameter-type models.

4.2.1 Polynomial models

Polynomial models describe the growth responses (μ_{max} , lag) or their transformations ($\sqrt{\mu_{max}}$, $\ln(\mu_{max})$, ...) through a polynomial function. An example of a polynomial model is that proposed by McClure et al. (1993) for *Brochothrix thermosphacta*:

$$\ln(\mu_{max}) = a_0 + a_1T + a_2pH + a_3(\%NaCl) + a_4T \cdot pH + a_5T(\%NaCl) + a_6pH(\%NaCl) + a_7T^2 + a_8pH^2 + a_9(\%NaCl)^2 \quad (22)$$

Polynomial models allow in almost every case the development of a model describing the effect of any environmental factor including interactions between factors. Moreover, the fit of polynomial models does not require the use of advanced techniques such as the non-

linear regression. The disadvantages of polynomial models lie in the high number of parameters and their lack of biological significance.

4.2.2 Square root-type

Temperature was the first environmental factor taken into account in square root-type models. Based on the observation of a linear relationship between temperature and the square root of the bacterial growth rate in the suboptimal temperature range, Ratkowsky et al. (1982) proposed the following function:

$$\sqrt{\mu_{\max}} = b(T - T_{\min}) \quad (23)$$

where b is a constant, T is the temperature and T_{\min} the theoretical minimum temperature for growth. This equation was extended to describe the effect of temperature in the entire temperature range allowing bacterial growth, the so-called 'biokinetic range' (Ratkowsky et al., 1983):

$$\sqrt{\mu_{\max}} = b(T - T_{\min}) \left(1 - \exp(-c(T - T_{\max}))\right) \quad (24)$$

where T_{\max} is the theoretical maximum temperature for growth and c a model parameter without biological meaning. These models were extended to take into account other environmental factors such as pH and water activity. McMeekin et al. (1987) proposed the following equation to describe the combined effects of temperature (suboptimal range) and water activity on the growth rate of *Staphylococcus xylosum*:

$$\sqrt{\mu_{\max}} = b(T - T_{\min}) \sqrt{(a_w - a_{w\min})} \quad (25)$$

where $a_{w\min}$ is the theoretical minimum water activity for growth. More recently, square root models have also been expanded to include, for example, the effects of CO₂, (Devlieghere et al., 1998) or pH and lactic acid concentration (Presser et al., 1997; Ross et al., 2003).

4.2.3 The gamma concept and the cardinal parameter model

Zwietering et al. (1992) proposed a model called "Gamma model", describing the growth rate relative to its maximum value at optimal conditions for growth:

$$\mu_{\max} = \mu_{opt} \gamma(T) \gamma(pH) \gamma(a_w) \quad (26)$$

where μ_{opt} is the growth rate at optimum conditions, and $\gamma(T)$, $\gamma(pH)$, $\gamma(a_w)$ are the relative effects of temperature, pH and a_w , respectively. The concept underlying this model ("Gamma concept") is based on the following assumptions: i) the effect of any factor on the growth rate can be described, as a fraction of μ_{opt} , using a function (γ) normalized between 0 (no growth) and 1 (optimum condition for growth); and ii) the environmental factors act independently on the bacterial growth rate. Consequently, the combined effects of the environmental factors can be obtained by multiplying the separate effects of each factor (see Equation 26). Ross & Dalgaard (2004) considered that while this apparently holds true for growth rate under conditions where growth is possible, environmental

factors do interact synergistically to govern the biokinetic ranges for each environmental factor.

At optimal conditions for growth, all γ terms are equal to 1 and therefore μ_{max} is equal to μ_{opt} . The γ terms proposed by Zwietering et al. (1992) for the normalized effects of temperature, pH and a_w are given in Equations 16 to 18.

$$\gamma(T) = \left(\frac{T - T_{min}}{T_{opt} - T_{min}} \right)^2 \quad (27)$$

$$\gamma(pH) = \frac{pH - pH_{min}}{pH_{opt} - pH_{min}} \quad (28)$$

$$\gamma(a_w) = \frac{a_w - a_{wmin}}{1 - a_{wmin}} \quad (29)$$

γ -type terms for pH and lactic acid effects on growth rate were also included in square root-type models by Presser et al. (1997) and Ross et al. (2003).

Introduced by Rosso et al. (1993, 1995), the cardinal parameter models (CPMs) were also developed according to the Gamma concept. The relative effects of temperature, pH and a_w on the bacterial growth rate are described by a general model called CPM_n :

$$CM_n(X) = \begin{cases} 0 & , X \leq X_{min} \\ \frac{(X - X_{max})(X - X_{min})}{(X_{opt} - X_{min})^{n-1} \left[(X_{opt} - X_{min})(X - X_{opt}) - (X_{opt} - X_{max})((n-1)X_{opt} + X_{min} - nX) \right]} & , X_{min} < X < X_{max} \\ 0 & , X \geq X_{max} \end{cases} \quad (30)$$

where X is temperature, pH or a_w . X_{min} and X_{max} are respectively the values of X below and above which no growth occur. X_{opt} is the value of X at which bacterial growth is optimum. n is a shape parameter. As for the "Gamma model" of Zwietering et al. (1992), $CM_n(X_{opt})$ is equal to 1, $CM_n(X_{min})$ and $CM_n(X_{max})$ are equal to 0.

For the effects of temperature and pH, n is set to 2 and 1 respectively (Augustin et al., 2000a, 2000b; Le Marc et al., 2002; Pouillot et al., 2003 Rosso et al., 1993, 1995). For the effects of a_w , n is set to 2 (Augustin et al., 2000a, 2000b; Rosso & Robinson, 2001). The combined effects of the environmental factors are also obtained by multiplying the relative effects of each factor. Thus, the Cardinal Parameter Model for the effects of temperature, pH and a_w on μ_{max} can be written as:

$$\mu_{max} = \mu_{opt} CM_2(T) CM_1(pH) CM_2(a_w) \quad (31)$$

or alternatively:

$$\mu_{max} = \mu_{opt} CM_2(T) CM_1(pH) CM_1(a_w) \quad (32)$$

In many ways, CPMs resemble the square root model. Responses predicted by the two types of models can be almost identical (Oscar, 2002; Ross & Dalgaard, 2004; Rosso et al., 1993,

1995). The advantages of the CPMs lie in the lack of structural correlation between parameters and the biological significance of all parameters (Rosso et al., 1995).

Several attempts have been made to include in CPMs the effects of organic acids (Augustin et al., 2000a, 200b; Coroller et al., 2003; Le Marc et al., 2002) or other inhibitory substances (Augustin et al., 2000a, 200b).

4.2.4 Artificial neural networks

“Black boxes” such as artificial neural networks are an alternative to the models above and have been used to develop secondary growth (or lag time) models (Garcia-Gimeno et al., 2002, 2003, Geeraerd et al., 1998). For further details on the principles of this method in the context of predictive microbiology, consult Hajmeer et al. (1997), Hajmeer & Basher (2002, 2003).

4.2.5 Probabilistic models

Probabilistic growth/no growth models can be seen as a logic continuation of the kinetic growth models. If environmental conditions are getting more stressful, growth rate is decreasing until zero and lag phase is increasing until eternity or at least longer than the period of experimental data generation. At these conditions, the boundary between growth and no growth has been reached and the modeling needs to shift from a kinetic model to a probabilistic model. The output of a probabilistic model will be the chance that a certain event will take place within a certain given time under conditions specified by the user. The period of time is an important criterion, as evolving from ‘no growth’ to ‘growth’ is, in many cases, possible as long as the period of analysis is sufficiently extended.

In the case of probabilistic models, responses will be coded as either 0 (response not observed) or 1 (response observed), under a specific set of conditions. If replicate observations are performed at a specific condition, a probability between 0 and 100% will be obtained. To relate this probability to the predictor variables, logistic regression is used (McMeekin et al., 2000; Ratkowsky & Ross, 1995). The logit function is defined as:

$$\text{Logit}P = \log(P / (1 - P)) \quad (33)$$

where P is the probability of the outcome of interest.

Logit P is commonly described as some function Y of the explanatory variables, i.e.:

$$\text{Logit}P = Y \quad (34)$$

Equation 34 can be rearranged to:

$$1 / (1 + e^{-Y}) = P$$

or

$$e^Y / (1 + e^Y) = P$$

where Y is the function describing the effects of the independent variables.

Several works have reported the application of probability models to different pathogens and spoilage microorganisms in foods (Lanciotti et al., 2001; Membré et al., 2001; Stewart et al., 2001).

5. Chemical spoilage

Chemical spoilage is mainly characterized by flavour and colour changes due to oxidation, irradiation, lipolysis (rancid) and heat. These changes may be induced by light, metal ions or excessive heat during processing or storage. Chemical processes may also bring physical changes such as increased viscosity, gelation, sedimentation or color change.

Biochemical reactions during food storage are relevant for the appearance of sensorial defects. Some of them are summarized as follows (Van Boekel, 2008):

- Lipid oxidation can lead to a loss of essential fatty acids and presence of rancidity, mainly due to the presence of free fatty acids. This phenomenon is associated to enzymatic oxidation, lipolysis, and discoloration.
- Non enzymatic and enzymatic browning can affect color, taste and aroma, nutritive value, and can contribute to the formation of toxicologically suspect compounds (acrylamide).
- Hydrolysis, lipolysis and proteolysis can cause changes in flavor, texture, vitamin content and formation of bitter taste.
- Separation and gelation are physical phenomena associated to sedimentation, creaming, gel formation and texture changes.

The first two processes normally characterize the majority of chemical spoilage phenomena in foods, and are explained below.

5.1 Lipid oxidation

Lipid oxidation is one of the most common causes of deterioration of food quality. Unsaturated fats are oxidized by free radical autoxidation, a chain reaction process catalyzed by the products of the reaction. The susceptibility and the rate of oxidation increase as the number of double bonds in the fatty acid increases.

Foods containing fat, e.g. milk, can undergo a number of subtle chemical and physical changes caused by lipolysis. Lipolysis can be defined as the enzymatic hydrolysis of fats by lipases. The accumulation of the reaction products, especially free fatty acids is responsible for the common off-flavour, frequently referred to as rancidity.

Fat stabilization is based on the use of physical methods (controlling temperature and light conditions during storage) and addition of antioxidants. In this sense, many foods or food ingredients contain components which with antioxidant properties (King et al., 1993). Examples of well known natural antioxidants are ascorbic acid, vitamin E (tocopherols), carotenoids and flavonoids. Antioxidants inhibit lipid oxidation by acting as hydrogen or electron donors, and interfere with the radical chain reaction by forming non radical compounds.

5.2 Non enzymatic and enzymatic browning

Non enzymatic browning resides in various chemical reactions occurring during food processing and storage. It is related to the formation of brown compounds and volatile substances that influence the sensorial quality of foods. Non enzymatic browning is in part associated to Maillard reactions. These include a number of complex reactions that are influenced by a large variety of factors (substrates nature, time/temperature combinations, pH, a_w , or presence of inhibitor compounds). Control and prevention measures of non enzymatic browning are based on the elimination of substrates, control of physico-chemical factors (shortening time and temperature, lowering pH) or addition of inhibitors such as sulfites, which delay the presence of pigments by means of fixing intermediate reaction products. Likewise, sulfites act against enzymatic browning and growth of microorganisms. Thus, they are used as preservatives in grape juices, wine, dehydrated fruits and concentrated juices.

Non enzymatic browning is a quality parameter for many dehydrated foods. For other products such as baked goods, coffee, molasses, and some breakfast foods, browning is desirable. However, in other foods browning is undesirable, decreasing the value or quality of the product. The browning include: decreased nutritional value from protein loss, off-flavor development, undesirable color, decreased solubility, textural changes, destruction of vitamins and increased acidity (Franzen et al., 1990). Other effects are destruction of essential amino acids such as lysine, formation of mutagenic compounds and modifications in the antioxidant capacity of foods.

Enzymatic browning is defined as the enzymatic transformation, in presence of oxygen at first stage, from phenolic compounds to colored polymers. Dark pigments formed as result of these reactions are known as melanines. Enzymatic browning is produced in fruits vegetables, which are rich in phenolic compounds, normally after harvesting and during processing conditions (washing, peeling, chopping, etc.). Polyphenoloxidases (PPO) and peroxidases found in fruit tissues can catalyze the oxidation of certain endogenous phenolic compounds to quinones that polymerize to form intense brown pigments (Lewis & Shibamoto, 1986).

Since enzymatic browning can cause important losses in agriculture production, control and prevention of the principal mechanisms of these reactions are crucial. These methods are based on the activity against enzymes, substrates and/or product reactions. Their use might depend on the specific food to be considered, as well as other nutritional, technical, economical, sensorial or ethical considerations.

6. Sensorial evaluation for determining shelf-life

As stated previously, sensorial evaluation is an essential task for establishing shelf-life of food products. Once a food product is designed (composition, packaging, foreseeable conditions of storage and use, target population), next step consists of estimating the period of time during which the food maintains its original sensorial properties. An immediate question arises from the last assertion: "how could I assess that sensorial properties do not change along time?" In other words, "how can these differences be detected?". There are different sensory tests to provide answer to this question; they are named sensory difference tests. However, at the same time, another question may come up to our minds: "Who should

detect these difference (in case they are detectable)?". This is not a straightforward question to answer, as it greatly depends on the food quality management of the company. One could think that trained panellists are the most appropriate to detect possible differences in sensorial characteristics of foods during storage. After all, they are "sensorial instruments". However, actually, they do not represent the target population, and probably, target population in general would not detect differences in such a perfect manner as trained panellists would do. So, it is very important to design an adequate test fitted to the needs, potential and politics of the company.

A sensory difference test allows for the possible distinction between various foods. In the case of the application of these tests to shelf-life estimation, a distinction should be made between the food under storage (at the foreseeable conditions) and the food just produced. The most common sensory difference tests are the paired-comparison test, the duo-trio test, and the triangle test (O'Mahony, 1985).

The paired-comparison difference test presents two samples to a judge who has to determine which one has more of a given attribute (sweeter, spicier, et.). Should he consistently pick the right sample over replicate tests, it would indicate that he can distinguish between the two. Should he pick the right sample more often than the wrong one, a binomial test will determine the probability of picking the right sample this often by chance, so as to be able to make a decision about whether the judge really would tell the difference. The disadvantage of the paired-comparison test is that the experimenter has to specify which attribute the judge has to look for, and it is not easy to explain, for example, what is meant by "having more off-flavor". The duo-trio test does not require the judge to determine which of two samples has more of a given attribute; it is used to determine which of two samples are the same as a standard sample presented before the test. This saves the difficulty of having to describe the attribute under consideration. The test is analyzed statistically in the same way as the paired-comparison test, to see whether a judge demonstrated that he could distinguish the two samples, by consistently picking the sample that matched the standard.

An alternative to the duo-trio test, that also avoids the difficulty of having to define the attribute that is varying between the samples, is the triangle test. Rather than picking one of a pair that is the same as a standard, the judge has to pick out the odd sample from a group of three (two the same, one different). Again, a binomial test is the appropriate statistical analysis.

A summary of the sensory difference tests is presented in Table 1. By setting an adequate sensory analysis plan, it is possible to fix the day from which differences are detected at a pre-established confidence level.

Carpenter et al. (2000) provided a case-study of the application of the sensory analysis in the establishment of shelf-life of chocolate filled and covered tabs, where triangular test was applied. In order to provide with control samples (samples just produced) to be able to make comparisons at every analysis day, a number of samples just produced was frozen at the beginning of the study, and at every analysis day, the necessary control samples were left to defrost. Previously, it was demonstrated that freezing and defrosting of samples did not influence the sensory characteristics of the product.

Paired-comparison		
A vs B		
Which is greater in a given attribute?	A or B?	One-tailed binomial test, $p = q = 1/2$ or two tails, $p = q = 1/2$.
Is there a difference between the two?	Same or different?	Two-tailed binomial test, $p = q = 1/2$.
Duo-trio		
A: A vs B		
Which is the same as the standard?	A or B?	One-tailed binomial test, $p = q = 1/2$
Triangle		
A B B		
Which is the odd sample?	A or B?	One-tailed binomial test, $p = 1/3, q = 2/2$

Table 1. Sensory difference tests and their analyses.

When sensory difference testing aims at detecting differences by consumers, the result of these tests will tell us the day from which differences are detected; however, these tests do not provide any information about the changes in sensorial attributes of a food. For a through knowledge of the food product, it is necessary to carry out sensory studies able to provide sensorial information about the food. These studies are called sensory descriptive tests. Murray et al. (2001) reviewed the state-of-the-art of descriptive sensory analysis. One of the most relevant descriptive tests is the Quantitative Descriptive Analysis (QDA). In the QDA, colour, odour, flavour, texture and other attributes are, in a preliminary stage, examined by an expert panel, which generates descriptive terms for the attributes. Then, the leader of the sensory analysis gathers and organized the terms, and prepares a draft list which is subsequently presented to the expert panel for discussion and fine-tuning. A definition and a scale for each attribute are established. The aim of this preliminary stage is to agree on a lot of standard attributes for a specific food, so that panellists can use it systematically. A definitive questionnaire is made up and, in a second stage, different sessions are organized according to the sensory analysis plan where every panellist fills in the questionnaire individually. QDA data obtained from every session can be statistically processed in different ways. The most common statistical tests are the *t* test (comparison between two products A and B), and the ANOVA test (comparison between more than two products), although other more sophisticated test designed to summarize a high number of sensory data can also be applied such as principal components analysis, correspondence analysis or discriminant analysis. Some guidelines like that published by Donelly & Mitchell, (2009) propose a scale from 1 to 4 to different attributes (e.g. for odour, 1 “normal odour; 2 = slight off-odour; 3 = moderate off-odour, and 4 = strong off odour), stating that a mean of 2.5 or below indicates an acceptable product, and a mean score of 2.5 marks the end of the product shelf-life.

In the case-study published by Carpenter et al. (2000), a QDA was performed in parallel to the triangle test. In this way, it was possible to identify the nature and magnitude of the changes in sensorial characteristics during storage. Apart from the statistical tests to analyze and interpret QDA data, a graphical representation of scores of the different attributes along time provides a “picture” of the evolution of attributes during storage.

Some authors have modelled the relationship between the time when defects appear or when differences between control and studied samples are detected, and temperature of

storage (Labuza & Fu, 1993; Mataragas et al., 2006; Valero et al., 2006), applying the following model:

$$t_s = t_0 \exp(-bxT) \quad (35)$$

where t_s is the time (d) when defects appear at any temperature T within the examined range; t_0 , the time (d) when defects appear at 0°C ; and b , the slope of the regression line of plot $\ln t_s = f(T)$. Labuza & Fu (1993) stated that when the temperature range of concern is relatively narrow (0 to 12°C) then the shelf-life of a product can be determined by using the lot shelf-life model (Equation 35).

For sensory evaluation, it is very important to establish the objective of the study, and depending on the company quality management, resources and duration of the study, an analysis plan should be properly designed.

7. Food testing for determination of shelf-life

When designing a shelf-life study, several questions arise, such as proper food samples storage, testing time, quantity of samples or tests which should be applied. The key steps for designing a shelf-life study are focused on the duration time of the study, sampling frequency and controls which will be carried out until the food presents a significant spoilage. Usually, controls regarding microbiology, sensory and physico-chemical analysis are established. To design a shelf-life study, factors such as temperature, relative humidity, illumination conditions, etc., should be strictly controlled, preferably in a certified laboratory. Additionally, the design must be simple, easy to follow and interpretable by food operators.

Preliminary information is very useful for refining the experimental design. This information can be obtained from multiple sources like historical data (i.e. processing and storage conditions, formulation of the food, historical microbiological data, etc.), or published studies performed in similar products.

Current International legislation does not provide the application of standard protocols aiming at establishing the shelf-life of food products. However, various National and International Guidelines for this purpose are published (Food Safety Authority of Ireland, 2005; Health Protection Agency, 2009; New Zealand Food Safety Authority, 2005; Refrigerated Food Association, 2009). These Guidelines suggest different methods and analyses protocols for shelf-life establishment, being mainly focused on RTE foods, as mentioned in Section 3.

7.1 Steps to be followed for the design of a shelf-life study

There are common steps to follow when designing a shelf-life study. The study can differ according to the previous knowledge one has with regards to the food product.

7.1.1 Identification of the main causes of food spoilage

Each food product includes a series of factors that limit its shelf-life. Some of them were described in Sections 2 and 5 and they can affect food quality and safety during storage and

distribution if they are not adequately controlled. Throughout this first step, identification of all related factors that can potentially influence food shelf-life from production to consumption is crucial. Consequently, the entire food chain must be examined.

7.1.2 Planning the shelf-life study

Once factors are identified, the shelf-life studies must be planned in order to give answer to the following questions:

- a. Which type of analyses must be carried out? A number suitable tests should be performed during a predetermined storage period. These comprise sensorial, microbiological and physico-chemical analyses. Regarding sensorial aspects, different attributes have to be evaluated such as odour, appearance, flavour, texture, as well as other organoleptical characteristics by a trained panel. In section 6, several sensory tests are proposed. Microbial analyses must be performed along the storage time, aiming at monitoring the changes in microbial concentration. The most common microbial groups analyzed are aerobic mesophilic bacteria, coliforms, yeasts, lactic-acid bacteria, and some indicators such as *E. coli*, *Staphylococcus aureus* or sporulated bacteria in thermal treated foods (*Bacillus* spp., *Clostridium* spp.). Physical analyses include texture changes, package type, appearance, etc., and they are usually considered as sensorial attributes. Regarding chemical analyses, changes in concentration of metabolites or chemical parameters (pH, nitrogen content, salt, organic acids, etc.) are monitored during the storage period. These tests are recommended to be performed at three different storage conditions as minimum: one established by the food operator (commercial conditions), one intermediate condition and one abuse condition (in case of breaking of the cold chain). For temperature control during storage, dataloggers should be used in refrigeration cameras.
- b. How many analyses must be carried out? This mainly depends on the pre-established shelf-life or prior knowledge, food type and storage conditions. In principle, for those foods with a long expected shelf-life (i.e. more than 3 months) analyses can be more sporadic. On the contrary, perishable and RTE foods have shorter shelf-lives (between one week and one month); in this case, microbial growth is normally the main reason of food spoilage, so it is necessary to increase the frequency of analyses in order to provide an accurate estimation of shelf-life. In general, for both long and short shelf-lives, it is desirable to set around 7-10 analytical points regularly distributed, although this number will change according to the specific case to study. Sampling points must be separated between 15-20% of the total shelf-life period. Besides, one final point has to be performed once shelf-life is expired (after 15-20% of the shelf-life time). In this way, the total analytical period will comprise 115-120% of the shelf-life period. For instance, if shelf-life is calculated in 30 days, analyses are recommended to be performed in days 0, 6, 12, 18, 24, 30 and 36. Obviously, frequency of analyses is influenced by the storage temperature.
- c. How many samples need to be withdrawn per analytical point? It is recommended to analyze, at least, two samples at each analytical point and storage condition. For instance, 7 analytical points \times 2 temperature conditions \times 2 samples = 28 samples. Normally, the samples destined to microbial or physico-chemical analyses, if not completely destructed, can be used for sensorial analyses.

- d. Which period is the most appropriate to perform the study? In any case, shelf-lives studies have to be repeated to control the variability of foods, but the most suitable period corresponds to summer months.

7.1.3 Shelf-life establishment

Shelf-life ends when the food loses its original quality and/or safety attributes. Based on the information collected in previous steps, the maximum storage time of the food at given conditions assuring the maintenance of appropriate characteristics, has to be decided. Estimations will be different if decisions are based on microbial or sensorial or physico-chemical changes. In any case, the most conservative approach should be chosen as the best one, unless there are other factors needing to be considered.

7.1.4 Shelf-life monitoring

It is advisable to validate the established shelf-life of a food as well as to repeat the analyses in several occasions to include shelf-life variability due to effects of different environmental variables.

7.2 Shelf-life studies based on *Listeria monocytogenes* growth in RTE foods

L. monocytogenes is a pathogen which may cause disease in humans and it is typically transmitted as a food-borne pathogen. *L. monocytogenes* is frequently present in the environment, soil, vegetation and faeces of animals. The organism can be found in raw foods such as fresh meat, raw milk and fish. The ubiquitous occurrence and the increased ability to grow or survive in a chilled environment compared to other microorganisms, makes *L. monocytogenes* a significant challenge in food production. This concern is of particular relevance in Ready-To-Eat (RTE) foods, as they do not received further treatment after production and their intrinsic characteristics support the growth of *L. monocytogenes*.

It is crucial that producers of RTE foods take actions to control the contamination by *L. monocytogenes* as well as its growth throughout the shelf-life. Knowledge on the growth potential of *L. monocytogenes* in RTE foods is needed, and this must be taken into account when setting their shelf-lives.

A guidance document was launched by the European Commission in 2008 regarding shelf-life studies for *L. monocytogenes* in RTE foods (SANCO, 2008). This document addressed the question stated in Annex II of the Regulation (EC) No 2073/2005, where food operators must demonstrate that *L. monocytogenes* cannot exceed 100 cfu/g throughout the shelf-life of RTE foods able to support the growth of *L. monocytogenes* and that may pose a risk for public health. In this document, microbiological procedures for determining the growth of *L. monocytogenes* through challenge tests were described, as well as durability studies in the frame of the application of the Regulation (EC) No. 2073/2005. The microbiological procedures include:

- a. Challenge tests
 - assessing a growth potential (δ)
 - assessing the maximum growth rate (μ_{\max})
- b. Durability studies

8. Assessment of shelf-life scientific-based

Food testing renders a data set of different nature (microbiological, physico-chemical, sensorial) which should be wisely managed to produce an accurate estimation of shelf-life of foods. It is a decision of the quality department of food companies to define the criteria upon which product shelf-life should be established.

At the beginning of the chapter, “product shelf-life” was defined as “the term or period a product may be stored before a specific element of the product makes it unsuitable for use or consumption”. Unsuitability can be evaluated by sensory testing, while the cause for unsuitability could be of biological or physico-chemical nature. Whichever characteristic is analyzed (microbiological, physico-chemical or sensorial) when a batch of product is analyzed at different time intervals during the period of storage, a “survival curve” can be constructed (Figure 4). A “survival curve” is a representation of the percentage of suitable food along time. In other words, a batch of product, even though belonging to the same lot, fails as storage time increases.

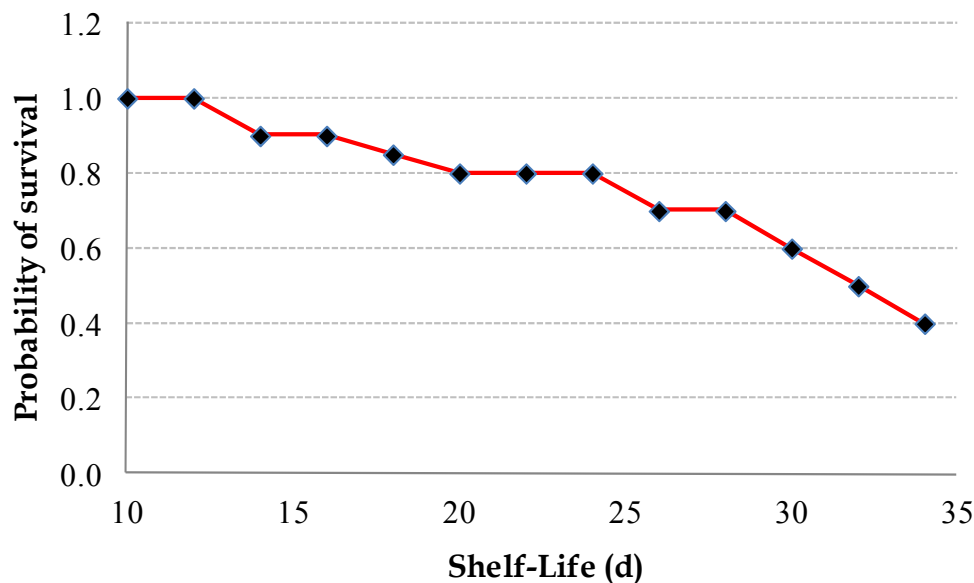


Fig. 4. Survival curve for a Food product.

A number of probability distributions have been proposed to model failure time of a product (Hough et al., 2003). Typically, such a distribution is defined by a small number of parameters and a mathematical equation. Some commonly used statistical distributions are the exponential, Weibull or log-normal distribution. The goodness of fit of these distributions to data can be assessed through the Anderson-Darling statistics; the smaller the value of this test statistics, the better the fit. In the shelf-life context, probability distributions represent storage time *vs* probability. Figure 5 represents the Weibull distribution fitted to a data set, where probability of rejection ($1 - \text{probability of survival}$) is plotted against shelf-life. To use these curves to predict shelf-life, the experimenter needs to define the largest acceptable proportion of defects that can be tolerated. A horizontal line can then be drawn on the plot to determine the corresponding shelf-life. Mataragas et al. (2007) used this approach to estimate the shelf-life of a sliced, cooked, cured meat product.

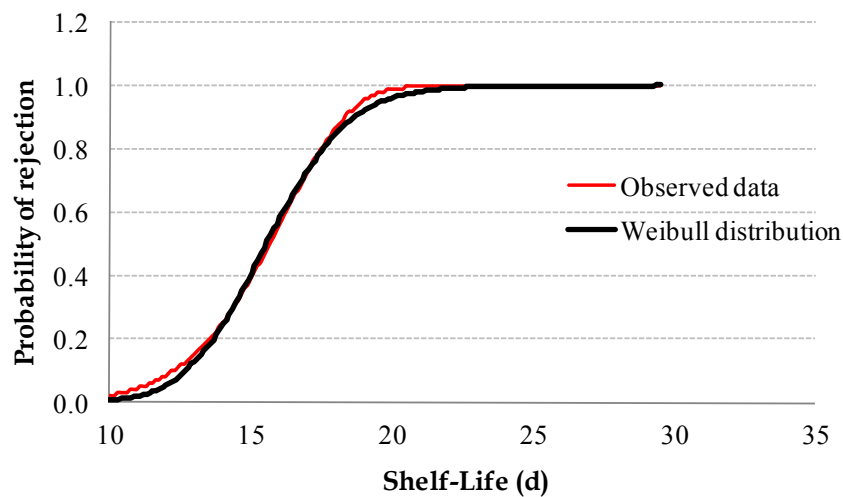


Fig. 5. Weibull distribution to predict shelf-life of a food product.

Statistical tools are of great value for the food industry and to the satisfaction of the different stakeholders, to demonstrate a science-based shelf-life, assuring the safety while maintaining the original sensorial characteristics of the food.

9. Prospects

Shelf-life of foods are being reformulating based on scientific evidence to the satisfaction of Health Authorities, import countries and different stakeholders. For this, a systematic approach for establishing by-product category shelf-lives should be adopted. Research and technological centres, Universities, food industries and sectorial associations should agree on standardized protocols to face this challenge.

Food industries could incorporate informatic tools to systematically produce foods meeting the quality level set up for them. The more homogeneous the food lots, the less deviations from established shelf-lives. A number of possibilities exist for the development of decision-support systems based on rule-based expert systems. Examples (Paoli, 2001, as cited in Tamplin et al., 2004) include:

- Virtual inspection: Simulating interaction with an inspector to allow establishments to self-assess their facility or a food processing operation. This may be of interest to large companies and regulators who find their quality control or inspection resources inadequate for the number of establishments and that they are required to assess.
- Process deviation assessment: Tools which incorporate expert knowledge regarding the best actions to take in case of process deviations, in terms of assessing the seriousness of the deviation and in recommending corrective actions for the implicated product and/or process.
- In-line real-time expert systems: Examples exist (though not in predictive microbiology) of expert systems that received real-time data and provide continuous assessments of the status of systems, based on the combination of these data and embedded knowledge-based rules that interpret the data for display to operators. This could be applied to food production systems where a complex set of variables requires monitoring combined with complex reasoning to assure safety and quality.

10. Acknowledgments

CTS-3620 Project of Excellence from the Andalusia Government, AGL 2008-03298/ALI project from the Spanish government, FP7-KBBE-2007-2A n° 222738 project from the VII Framework Programme and European ERDF funding are greatly acknowledged for providing material and specially human resources, making possible the continuation of Risk Assessment and Management activities at national and European level by our research group AGR170 "HIBRO".

11. References

- Augustin, J.C. & Carlier, V. (2000a). Modelling the growth rate of *Listeria monocytogenes* with a multiplicative type model including interactions between environmental factors. *International Journal of Food Microbiology*, Vol.56, pp. 53-70, ISSN 0168-1605
- Augustin, J.C. & Carlier, V. (2000b). Mathematical modelling of the growth rate and lag time for *Listeria monocytogenes*. *International Journal of Food Microbiology*, Vol.56, pp. 29-51, ISSN 0168-1605
- Babic, I. & Watada, A.E. (1996). Microbial populations of fresh-cut spinach leaves affected by controlled atmospheres. *Postharvest Biology and Technology*, Vol. 9, pp, 187-193. ISSN 0925-5214
- Baranyi, J. & Roberts, T.A. (1994). A dynamic approach to predicting bacterial growth in food. *International Journal of Food Microbiology*, Vol.23, pp. 277-294, ISSN 0168-1605
- Baranyi, J. & Tamplin, M.L. (2004). ComBase: A Common Database on Microbial Responses to Food Environments. *Journal of Food Protection*, Vol.67, pp. 1967-1971, ISSN 0362-028X
- Baranyi, J. (1992). Letters to the Editor: a note on reparameterization of bacterial growth curves. *Food Microbiology*, Vol.9, pp. 169-171, ISSN 0740-0020
- Baranyi, J. Roberts, T.A. & McClure, P. (1993). A non-autonomous differential equation to model bacterial growth. *Food Microbiology*, Vol.10, pp. 43-59, ISSN 0740-0020
- Baranyi, J. Robinson, T.P. & Mackey, B.M. (1995). Predicting growth of *Brochothrix thermosphacta* at changing temperature. *International Journal of Food Microbiology*, Vol.27, pp. 61-75, ISSN 0168-1605
- Baty, F. & Delignette-Müller, M.L. (2004). Estimating the bacterial lag time: which model, which precision? *International Journal of Food Microbiology*, Vol.91, pp. 261-277, ISSN 0168-1605
- Bigelow, W.D. (1921). The logarithmic nature of thermal death time curves. *The Journal of Infectious Diseases*, Vol.29, pp. 528-536, ISSN 0022-1899
- Brillet, A. Pilet, M.F. Prévost, H. Cardinal, M. & Leroi, F. (2005) Effect of inoculation of *Carnobacterium divergens* V41, a biopreservative strain against *Listeria monocytogenes* risk, on the microbiological, and sensory quality of cold-smoked salmon. *International Journal of Food Microbiology*, Vol. 104, pp, 309-324. ISSN 0168-1605
- Brul, S. van Gerwen, S. & Zwietering, M. (2007). Modelling microorganisms in food. Cambridge, Woodhead Publishing Limited.
- Buchanan, R.L. (1993). Developing and Distributing User-Friendly Application Software. *Journal of Industrial Microbiology*, Vol.12, pp. 251-255, ISSN 0169-4146

- Carpenter, R.P. Lyon, D.H. & Hasdell, T.A. (2000). *Guidelines for Sensory Analysis in Food Product Development and Quality Control* (2nd Edition), ISBN 978-0-8342-1642-6, Kluwer Academic/Plenum Publishers, New York, USA
- Carrasco, E. Pérez-Rodríguez, F. Valero, A. García-Gimeno, R.M. & Zurera, G. (2007). Survey of temperature and consumption patterns of fresh-cut leafy green salads: risk factors for listeriosis. *Journal of Food Protection*, Vol. 70, pp, 2407-2412. ISSN 0362-028X
- Cole, M.B. Davies, K.W. Munro, G. Holyoak, C.D. & Kilsby, D.C. (1993). A vitalistic model to describe the thermal inactivation of *Listeria monocytogenes*. *Journal of Industrial Microbiology and Biotechnology*, Vol.12, 232-239, ISSN 1367-5435
- Commission of the European Communities (2008). Technical Guidance Document on *Listeria monocytogenes* shelf-life studies for ready-to-eat foods, under Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. EU Community Reference Laboratory for *Listeria monocytogenes*. SANCO/1628/2008 ver. 9.3 (26112008)
- Commission Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. Official Journal of the European Union, L338/1-L338/26. Available from:
<http://eur-lex.europa.eu /LexUriServ /LexUriServ.do?uri= OJ:L:2005: 338:0001: 0026:EN:PDF>
- Commission Regulation (EC) No 852/2004 of the European Parliament and of the Council of 29 April 2004 on the Hygiene of Foodstuffs. Official Journal of the European Union, L339/1. Available from:
<http://eur-lex.europa.eu /LexUriServ /LexUriServ.do? uri=OJ:L: 2004: 139: 0001: 0054:en:PDF>
- Coroller, L. Guerrot, V. Huchet, V. Le Marc, Y. Sohier, D. & Thuault, D. (2003). Growth kinetics modelling of pathogen bacteria as function of different acids. In: *Predictive Modelling in foods-Conference proceedings*, J.F.M. Van Impe, A.H. Geerard, I. Leguerinel & P. Mafart (Eds.), pp 120-122, Kathoelieke Universiteit Leuven/BioTec, Belgium
- Coroller, L. Leguerinel, I. Mettler, E. Savy, N. & Mafart, P. (2006). General model, based on two mixed Weibull distributions of bacterial resistance, for describing various shapes of inactivation curves. *Applied and Environmental Microbiology*, Vol.72, pp. 6494-6502, ISSN 0099-2240
- Dalgaard, P. (1995). Qualitative and quantitative characterization of spoilage bacteria from packed fish. *International Journal of Food Microbiology*, Vol. 26, pp, 319-333. ISSN 0168-1605
- Dalgaard, P. Mejlholm, O. & Huss, H.H. (1997). Application of an iterative approach for development of a microbial model predicting the shelf-life of packed fish. *International Journal of Food Microbiology*, Vol. 38, pp, 169-179. ISSN 0168-1605
- Dalgaard, P.; Buch, P. & Silberg, S. (2002). Seafood Spoilage Predictor-development and distribution of a product specific application software. *International Journal of Food Microbiology*, Vol.73, pp. 343-349, ISSN 0168-1605
- Dalgaard, P. Ross, T. Kamperman, L. Neumeyer, K. & McMeekin, T.A. (1994). Estimation of bacterial growth rates from turbidimetric and viable count data. *International Journal of Food Microbiology*, Vol.23, pp. 391-404, ISSN 0168-1605

- Dalgaard, P. (1993) Evaluation and prediction of microbial fish spoilage. PhD thesis, Royal Veterinary and Agricultural University. Copenhagen.
- Devlieghere, F. Van Belle, B. & Debevere, J. (2001). Shelf life of modified atmosphere packed cooked meat products: a predictive model. *International Journal of Food Microbiology*, Vol. 46, pp, 57-70. ISSN 0168-1605
- Devlieghere, F. Debevere, J. & Van Impe, J.F.M. (1998). Concentration of carbon dioxide in the water-phase as a parameter to model the effect of a modified atmosphere on microorganisms. *International Journal of Food Microbiology*, Vol.43, pp. 105-113, ISSN 0168-1605
- Donnelly, C.W. & Mitchell, M.M. (2009). Refrigerated Foods Association (RFA) Standardized Protocol for Determining the Shelf-Life of Refrigerated Ready-To-Eat (RTE) Foods, In: *Refrigerated Foods Association*, 10.10.2011, Available from: <http://www.refrigeratedfoods.org/food-safety-resources>
- Edwards, C.G. Haag, K.M. Collins, M.D. Hutson, R.A. & Huang, Y.C. (1998). *Lactobacillus kunkeei* sp. nov.: a spoilage organism associated with grape juice fermentations. *Journal of Applied Microbiology*, Vol. 84, pp, 698-702. ISSN 1364-5072
- Esse, R. & Humidipak, A. (2004). Shelf-life and moisture management. In: *Understanding and Measuring the Shelf-Life of Food*, (ed. Steele, R.). Published by Woodhead Publishing Ltd., Cambridge, England, pp, 24-41.
- Fennema, O.R. (1996). Water and ice. In: Fennema, O.R. (ed.) *Food Chemistry* (3rd Edition), Marcel Dekker Inc., New York. pp. 17-94.
- Food Safety Authority of Ireland (2005). Guidance Document on the determination of product shelf-life. Available from: http://www.fsai.ie/faq/shelf_life.html.
- Franzen, K. Singh, R.K. & Okos, M.R. (1990). Kinetics of nonenzymatic browning in dried skim milk. *Journal of Food Engineering*, Vol. 11, pp, 225-239. ISSN 0260-8774
- García, P. Rodríguez, L. Rodríguez, A. and Martínez, B. (2010). Food biopreservation: promising strategies using bacteriocins, bacteriophages and endolysins. *Trends in Food Science and Technology*, Vol. 21, pp, 373-382. ISSN 0924-2244
- Garcia-Gimeno, R.M. Hervas-Martinez, C. & de Simoniz, M.I. (2002). Improving artificial neural networks with a pruning methodology and genetic algorithms for their application in microbial growth predictions in food. *International Journal of Food Microbiology*, Vol.72, pp. 19-30, ISSN 0168-1605
- Garcia-Gimeno, R.M. Hervas-Martinez, C. Barco-Alcaia, E. Zurena-Cosano, G. & Sanz-Tapia, E. (2003). An artificial neural network approach to *Escherichia coli* O157:H7 growth estimation. *Journal of Food Science*, Vol.68, pp. 639-645, ISSN 1750-3841
- Garthright, W.E. (1997). The three-phase linear model of bacterial growth: a response. *Food Microbiology*, Vol.14, 395-397, ISSN 0740-0020
- Geeraerd, A.H. Herremans, C.H. Cenens, C. & Van Impe, J.F.M. (1998). Application of artificial neural networks as a non linear modular modeling technique to describe bacterial growth in chilled food products. *International Journal of Food Microbiology*, Vol.44, pp. 49-68, ISSN 0168-1605
- Geeraerd, A.H. Valdramidis, V.P. & Van Impe, J.F. (2005.) GInaFiT, a freeware tool to assess non-log-linear microbial survivor curves. *International Journal of Food Microbiology*, Vol.102, pp. 95-105, ISSN 0168-1605
- Gibson, A.M. Bratchell, N. & Roberts, T.A. (1987). The effect of sodium chloride and temperature on the rate and extent of growth of *Clostridium botulinum* tupe A in

- pasterurized pork slurry. *Journal of Applied Bacteriology*, Vol.62, pp. 479-490, ISSN 1364-5072
- González, B. & Díez, V. (2002). The effect of nitrite and starter culture on microbiological quality of "chorizo" – a spanish dry cured sasusage. *Meat Science*, Vol.60, pp. 295-298, ISSN 0309-1740
- Gram, L. (1989). Identification, characterization and inhibition of bacteria isolated from tropical fish. Ph.D. Thesis. Danish Institute for Fisheries Research, Lyngby and The Royal Veterinary and Agricultural University, Copenhagen, Denmark.
- Gram, L., & Dalgaard, P. (2002). Fish spoilage bacteria – problems and solutions. *Current Opinion in Biotechnology*, Vol. 13, pp. 262-266. ISSN 0958-1669
- Gram, L. Ravn, L. Rasch, M. Bruhn, J.B. Christensen, A.B. & Givskov, M. (2002). Food spoilage- interactions between food spoilage bacteria. *International Journal of Food Microbiology*, Vol. 78, pp. 79– 97. ISSN 0168-1605
- Grandinson, A.S. & Jennings, A. (1993). Extension of the shelf-life of fresh minced chicken meat by electron bean irradiation combined with modified atmosphere packaging, *Food Control*, Vol. 4, pp. 83-88. ISSN 0956-7135
- Hajmeer, M.N. & Basher, I. (2003a). A probabilistic neural network approach for probabilistic modelling of bacterial growth/ no growth data. *Journal of Microbiological Methods*, Vol.51, pp. 217-226, ISSN 0167-7012
- Hajmeer, M.N. & Basher, I.A. (2003b). A hybrid Bayesian-neural network approach for probabilistic modelling of bacterial growth/ no growth interface. *International Journal of Food Microbiology*, Vol.82, pp. 233-243, ISSN 0168-1605
- Hajmeer, M.N. Basher, I.A. & Najjar, Y.M. (1997). Computational neural networks for predictive microbiology. 2. Application to microbial growth. *International Journal of Food Microbiology*, 34, pp. 51-66, ISSN 0168-1605
- Health Protection Agency (2009). Guidelines for Assessing the Microbiological Safety of Ready-to-Eat Foods Placed on the market. Available from:
- Hough, G. Langohr, K. Gomez, G. & Curia, A. (2003). Survival analysis applied to sensory shelf-life of foods. *Journal of Food Science*, Vol.68, pp. 359-362, ISSN 0022-1147
http://www.hpa.org.uk/webc/HPAwebFile/HPAweb_C/1259151921557.
- Huis In't Veld, J. (1996). Microbial and biochemical spoilage of foods: an overview. *International Journal of Food Microbiology*, Vol. 33, pp. 1-18. ISSN 0168-1605
- Institute of Food Science and Technology (1993). Shelf life of Foods-Guideline for its Determination and Prediction. IFST, London.
- Jacxsens, L. Devlieghere, F. Van der Steen, C. & Debevere, J. (2001). Effect of high oxygen modified atmosphere packaging on microbial growth and sensorial qualities of fresh-cut produce. *International Journal of Food Microbiology*, Vol. 71, pp. 197-210. ISSN 0168-1605
- Jay, J.M. (1992). Intrinsic and extrinsic parameters of foods that affect microbial growth. In: Jay, J.M. (ed.), *Modern Food Microbiology* 4th Edition, Chapman & Hall, New York. pp. 38-62.
- Jay, J.M. (2000). *Modern food microbiology*. 6th ed. Gaithersburg (MD): Aspen, pp. 679.
- Kilcast, D. & Subramaniam, P. (2000). *Stability and Shelf-life of Foods*. pp. 1-22, 79-105. New York: CRC Press.

- King, D.L. Hahm, T.S. & Min, D.B. (1993) Chemistry of antioxidants in relation to shelf life of foods. In: G. Charalambous (editor), *Shelf-life Studies in Foods and Beverages*. Elsevier, Amsterdam, pp, 629-707.
- Koseki, S. (2009). Microbial Responses Viewer (MRV): a new ComBase-derived database of microbial responses to food environments. *International Journal of Food Microbiology*, Vol.134, pp. 75-82, ISSN 0168-1605
- Koseki, S. & Itoh, K. (2002). Effect of nitrogen gas packaging on the quality and microbial growth of fresh-cut vegetables under low temperatures. *Journal of Food Protection*, Vol. 65, pp, 326-332. ISSN 0362-028X
- Koutsoumanis, K. & Nychas, G.J.E. (2000). Application of a systematic experimental procedure to develop a microbial model for rapid fish shelf-life prediction. *International Journal of Food Microbiology*, Vol. 60, pp, 171-184. ISSN 0168-1605
- Koutsoumanis, K. Lambropoulou, K. & Nychas, G.J.E. (1999). A predictive model for the non-thermal inactivation of *Salmonella enteritidis* in a food model system supplemented with a natural antimicrobial. *International Journal of Food Microbiology*, Vol.49, pp. 63-74, ISSN 0168-1605
- Labuza, T.P. & Fu, B. (1993). Growth kinetics for shelf-life predictions: theory and practice. *Journal of Industrial Microbiology and Biotechnology*, Vol.12, pp. 309-323, ISSN 1367-5435
- Lanciotti, R. Sinigaglia, M. Guardini, F. Vannini, L. & Guerzoni, M.E. (2001). Growth/no growth interface of *Bacillus cereus*, *Staphylococcus aureus* and *Salmonella enteritidis* in model systems based on water activity, pH, temperature and ethanol concentration. *Food Microbiology*, Vol.18, pp. 659-668, ISSN 0740-0020
- Le Marc, Y. Huchet, V. Bourgeois, C.M. Guyonnet, J.P. Mafart P. & Thuault, D. (2002). Modelling the growth kinetics of *Listeria* as a function of temperature, pH and organic acid concentration. *International Journal of Food Microbiology*, Vol.73, pp. 219-237, ISSN 0168-1605
- Leistner, L. (1995). Principles and applications of hurdle technology. In: Gould GW, editor. *New methods of food preservation*. London: Blackie Academic & Professional, pp, 1-21.
- Leistner, L. (2000). Minimally processed, ready-to-eat, and ambient-stable meat. In *Shelf-Life Evaluation of Foods* (ed. Man, C.M. & Jones, A.A.), pp, 242-262. Aspen Publishers, Gaithersburg, USA.
- Leporq, B. Membré, J.M. Dervin, C. Buche, P. & Guyonnet, J.P. (2005). The "Sym'Previus" software, a tool to support decisions to the foodstuff safety. *International Journal of Food Microbiology*, Vol.100, pp. 231-237, ISSN 0168-1605
- Lewis, D.C. & Shibamoto, T. (1986). Shelf-life of fruits, In: G. Charamlabous (editor), *Handbook of food and beverage stability*. Academic Press, Orlando, pp, 353-391.
- Liao, C.H. Sullivan, J. Grady, J., Wong, & L.J.C. (1997). Biochemical characterization of pectate lyases produced by fluorescent pseudomonads associated with spoilage of fresh fruits and vegetables. *Journal of Applied Microbiology*, Vol. 83, pp, 10- 16. ISSN 1365-2672
- Lindqvist, R. & Lindblad, M. (2008). Inactivation of *Escherichia coli*, *Listeria monocytogenes* and *Yersinia enterocolitica* in fermented sausages during maturation/storage. *International Journal of Food Microbiology*, Vol.129, pp. 59-67, ISSN 0168-1605

- Linton, R.H. Carter, W.H. Pierson, M.D. & Hackney, C.R. (1995). Use of a modified Gompertz equation to model nonlinear survival curves for *Listeria monocytogenes* Scott A. *Journal of Food Protection*, Vol.58, pp. 946-954, ISSN 0362-028X
- Loss, C.R. & Hotchkiss, J.H. (2002). Inhibition of microbial growth by low-pressure and ambient pressure gasses. In: Juneja VK, Sofos JN, editors. Control of food-borne microorganisms. New York: Marcel Dekker, pp, 245-79. Forthcoming.
- Lund, B.M. (1992). Ecosystems in vegetable foods. *Journal of Applied Bacteriology*, Vol. 73, pp, 115S- 126S. ISSN 1364-5072
- Mannhein, C.H. & Soffer, T. (1996). Shelf-life extension of cottage cheese by modified atmosphere packaging. *LWT-Food Science and Technology*, Vol. 29, pp, 767-771. ISSN 1365-2672
- Mataragas, M. & Drosinos, E.H. (2007). Shelf life establishment of a sliced, cooked, cured meat product base don quality and safety determinants. *Journal of Food Protection*, Vol.70, pp. 1881-1889, ISSN 0362-028X
- Mataragas, M. Drosinos, E.H. Vaidanis, A. & Metaxopoulos, I. (2006). Development of a predictive model for spoilage of cooked cured meat products and its validation under constant and dynamic temperature storage conditions. *Journal of Food Science*, Vol.71, pp. M157-M167, ISSN 0022-1147
- Mazzota, A.S. (2001). Thermal inactivation of stationary-phase and acid-adaptated *Escherichia coli* O157-H7, *Salmonella*, and *Listeria monocytogenes* in fruit juices. *Journal of Food Protection*, Vol.64, pp. 315-320, ISSN 0362-028X
- McClure, P.J. Baranyi, J. Boogard, E. Kelly, T.M. & Roberts, T.A. (1993). A predictive model for the combined effect of pH, sodium chloride and storage temperature on the growth of *Brochothrix thermosphacta*. *International Journal of Food Microbiology*, Vol.19, pp. 161-178, ISSN 0168-1605
- McKellar, R.B. & Lu, X. (Eds.). (2004a). *Modeling microbial responses in food*. CRC Press, ISBN 0-8493-1237-X, Boca Raton, Florida
- McKellar, R.B. & Lu, X. (2004b). Primary models, In: *Modelling Microbial Responses in Foods*, R.C. McKellar & X. Lu (Eds.), pp. 21-62, CRC Press, ISBN 0-8493-1237-X, Boca Raton, Florida
- McKellar, R.C. & Knight, K.P. (2000). A combined discrete-continuous model describing the lag phase of *Listeria monocytogenes*. *International Journal of Food Microbiology*, Vol.54, pp. 171-180, ISSN 0168-1605
- McKellar, R.C. (1997). A heterogeneous population model for the analysis of bacterial growth kinetics. *International Journal of Food Microbiology*, Vol.36, pp. 179-186, ISSN 0168-1605
- McMeekin, T.A. (2007). Predictive microbiology: quantitative science delivering quantifiable to the meat industry and other food industries. *Meat Science*, Vol. 77, pp, 17-27. ISSN 0309-1740
- McMeekin, T.A. Olley, J. Ratkowsky, D.A. & Ross, T. (2002). Predictive microbiology: towards the interface and beyond. *International Journal of Food Microbiology*, Vol. 73, pp, 395-407. ISSN 0168-1605
- McMeekin, T.A. Chandler, R.E. Doe., P.E. Gardland, C.D. Olley, J. Putro, S. & Ratkowsky, D.A. (1987). Model for the combined effect of temperature and salt/water activity on growth rate of *Staphylococcus xylosus*. *Journal of Applied Bacteriology*, Vol.62, pp. 543-550, ISSN 1364-5072

- McMeekin, T.A. Olley, J.N. Ross, T. & Ratkowsky, D.A. (1993). *Predictive Microbiology: Theory and Application*, Research Studies Press, ISBN 10: 0863801323, Taunton, UK
- McMeekin, T.A. Presser, K. Ratkowsky, D.A. Ross, T. Salter, M. & Tienungoon, S. (2000). Quantifying the furdle concept by modelling the bacterial growth/no growth interface. *International Journal of Food Microbiology*, Vol.55, pp. 93-98, ISSN 0168-1605
- Membré, J.M. Kubaczka, M. & Chene, C. (2001). Growth rate and growth-no-growth interface of *Penicillium brevicompactum* as functions of pH and preservative acids. *Food Microbiology*, Vol.18, pp. 531-538, ISSN 0740-0020
- Membré, J.M. Ross, T. & McMeekin, T.A. (1999). Behaviour of *Listeria monocytogenes* under combined chilling processes. *Letters in Applied Microbiology*, Vol.28, pp. 216-220, ISSN 1472-765X
- Mifflin, H. (Ed.). (2006). *The American Heritage Dictionary of the English Language* (fourth edition), Houghton Mifflin Company, ISBN-10: 0618701729, ISBN-13/EAN: 9780618701728, Florida
- Moretti, V.M. Madonia, G. Diaferia, C. Mentasti, T. Paleary, M.A. Panseri, S. Pirone, G. & Gndini, G. (2004). Chemical and microbiological parameters and sensory attributes of a typical Sicilian salami ripened in different conditions. *Meat Science*, Vol.66, pp. 845-854, ISSN 0309-1740
- Morris, J.G. (2000). The effect of redox potential. In: Lund BL, Baird-Parker TC, Gould GW, editors. *The microbiological safety and quality of food*. Volume 1. Gaithersburg (MD): Aspen. pp, 235-250.
- Mossel, D.A.A. Corry, J.E.L. Struijk, C.B. & Baird, R.M. (1995). *Essentials of the microbiology of foods: a textbook for advanced studies*. Chichester (England): John Wiley and Sons. 699 pp.
- Murphy, R.Y. Duncan, L.K. Johnson, E.R. Davis, M.D. & Smith, J.N. (2002). Thermal inactivation D- and z-values of *Salmonella* serotypes and *Listeria innocua* in chicken patties, chicken tenders, franks, beef patties, and blended beef and turkey patties. *Journal of Food Protection*, Vol.65, pp. 53-60, ISSN 0362-028X
- Murray, J.M. Delahunty, C.M. & Baxter, I.A. (2001). Descriptive sensory analysis: past, present and future. *Food Research International*, Vol.34, pp. 461-471, ISSN 0963-9969
- New Zealand Food Safety Authority (2005). A guide to calculating shelf-life of foods. Information booklet for the food industry. Available from <http://www.nzfsa.govt.nz>.
- O'Mahony, M. (1985). *Sensory evaluation of food. Statistical Methods and Procedures*. Marcel Dekker, Inc., ISBN 0-8247-7337-3, New York
- Oscar, T.P. Development and validation of a tertiary simulation model for predicting the potential growth of *Salmonella typhimurium* on cooked chicken. (2002). *International Journal of Food Microbiology*, Vol.76, pp. 177-190, ISSN 0168-1605
- Paoli, G. (2001). Prospects for expert system support in the application of predictive microbiology, *Proceedings of Decision-Support Tools for Microbial Risk Assessment*, Seattle, WA
- Patsias, A. Chouliara, I. Badeka, A. Savvaidis, I.N. & Kontominas, M.G. (2006). Shelf-life of a chilled precooked chicken product stored in air and under modified atmospheres: microbiological, chemical, sensory attributes. *Food Microbiology*, Vol. 23, pp, 423-429. ISSN 0740-0020

- Peleg, M. (2006). Advanced quantitative microbiology for foods and biosystems: models for predicting growth and inactivation. Boca Ratón, FL, CRC, Taylor & Francis Group.
- Pitt, J.I. & Hocking, A.D. (1997). Fungi and Food Spoilage, 2nd ed. Blackie Academic and Professional, 596 pp
- Pouillot, R. Albert, I. Cornu, M. & Denis, J.B. (2003). Estimation of uncertainty and variability in bacterial growth using Bayesian inference. Application to *Listeria monocytogenes*. *International Journal of Food Microbiology*, Vol.81, pp. 87-104, ISSN 0168-1605
- Presser, K.A. Ratkowsky, D.A. & Ross, T. (1997). Modelling the growth rate of *Escherichia coli* as a function of pH and lactic acid concentration. *Applied and Environmental Microbiology*, Vol.63, pp. 2355-2360. ISSN 0099-2240
- Pruitt, K.M. & Kamau, D.N. (1993). Mathematical models of bacterial growth, inhibition and death under combined stress conditions. *Journal of Industrial Microbiology and Biotechnology*, Vol.12, pp. 221-231, ISSN 1367-5435
- Ratkowsky, D.A. & Ross, T. (1995). Modelling the bacterial growth/no growth interface. *Letters in Applied Microbiology*, Vol.20, pp. 29-33, ISSN 1472-765X
- Ratkowsky, D.A. Lowry, R.K. McMeekin, T.A. Stokes, A.N. & Chandler, R.E. (1983). Model for the bacterial culture growth rate throughout the entire biokinetic temperature range. *Journal of Bacteriology*, Vol.154, pp. 1222-1226, ISSN 0021-9193
- Ratkowsky, D.A. Olley, J. McMeekin, T.A. & Ball, A. (1982). Relationship between temperature and growth rates of bacterial cultures. *Journal of Bacteriology*, Vol.149, pp. 1-5, ISSN 0021-9193
- Refrigerated Foods Association (RFA) (2009). Standardized Protocol for Determining the Shelf Life of Refrigerated Ready-To-Eat (RTE) Foods (revised January 2009). Available from:
https://email.rutgers.edu/pipermail/fen_members/attachments/20100614/7f670f6b/RFA_ShelfLifeProtocol-09-0001.pdf.
- Rho, M.J. & Schaffner, D.W. (2007). Microbial risk assessment of staphylococcal food poisoning in Korean *kimbab*. *International Journal of Food Microbiology*, Vol. 116, pp, 332-338. ISSN 0168-1605
- Rico, D. Martín-Diana, A.B. Barat, J.M. & Barry-Ryan, C. (2007). Extending and measuring the quality of fresh-cut fruit and vegetables: a review. *Trends in Food Science & Technology*, Vol. 18, pp, 373-386. ISSN 0924-2244
- Robinson T.P. Ocio, M.J. Kaloti, A. & Mackey B.M. (1998). The effect of the growth environment on the lag phase of *Listeria monocytogenes*. *International Journal of Food Microbiology*, Vol. 44, pp. 83-92, ISSN 0168-1605
- Ross T. & Dalgaard, P. (2004). Secondary models. In: *Modelling Microbial Responses in Foods*, R.C. McKellar & X. Lu (Eds.), pp. 63-150, CRC Press, ISBN 0-8493-1237-X, Boca Raton, Florida
- Ross, T. & Shadbolt, C.T. (2004). Predicting *Escherichia coli* inactivation in uncooked comminuted fermented meat products, In: *Meat & Livestock Australia*, ISBN 095852548, 20.10.2011, Available from:
<http://www.foodsafetycentre.com.au/docs/Salami%20Final%20report.pdf>
- Ross, T. Ratkowsky, D.A. Mellefont, L.A. & McMeekin, T.A. (2003). Modelling the effects of temperature, water activity, pH and lactic acid concentration on the growth rate of

- Escherichia coli*. *International Journal of Food Microbiology*, Vol.82, pp. 33-43, ISSN 0168-1605
- Rosso, L. & Robinson, T.P. (2001). A cardinal model to describe the effect of water activity on the growth of moulds. *International Journal of Food Microbiology*, Vol.63, pp. 265-273, ISSN 0168-1605
- Rosso, L. Lobry, J.R. & Flandrois, J.P. (1993). An unexpected correlation between cardinal temperatures of microbial growth highlighted by a new model. *Journal of Theoretical Biology*, Vol.162, pp. 447-463, ISSN 0022-5193
- Rosso, L. Lobry, J.R. Bajard, S. & Flandrois, J.P. (1995). A convenient model to describe the combined effects of temperature and pH on microbial growth. *Applied and Environmental Microbiology*, Vol.61, pp. 610-616, ISSN 0099-2240
- Rubio, B. Martínez, B. Sánchez, M.J. García-Cachán, M.D. Rovira, J. & Jaime, I. (2007). Study of the shelf life of a dry fermented sausage “salchichon” made from raw material enriched in monounsaturated and polyunsaturated fatty acids and stored under modified atmospheres. *Meat Science*, Vol.76, pp. 128-137, ISSN 0309-1740
- Schellekens, M. Martens, T. Roberts, T.A. Mackey, B.M. Nicolai, M.B. Van Impe, J.F. & De Baerdemaeker, J. (1994). Computer-aided microbial safety design of food processes. *International Journal of Food Microbiology*, Vol.24, pp. 1-9, ISSN 0168-1605
- Stewart, C.M. Cole, M.B. Legan, J.D. Slade, L. Vandeven, M.H. & Schaffner, D.W. (2001). Modeling the growth boundary of *Staphylococcus aureus* for risk assessment purposes. *Journal of Food Protection*, Vol.64, 51-57, ISSN 0362-028X
- Stringer, S.C. George, S.M. & Peck, M.W. (2000). Thermal inactivation of *Escherichia coli* O157:H7. *Symposium Series (Society for Applied Microbiology)*, Vol.88. 79S-89S, ISSN 1467- 4734
- Takumi, K. de Jonge, R. & Havelaar, A. (2000). Modelling inactivation of *Escherichia coli* by low pH: application to passage through the stomach of young and elderly people. *Journal of Applied Microbiology*, Vol.89, pp. 935-943, ISSN 1364-5072
- Tamplin, M. Baranyi, J. & Paoli, G. (2004). Software programs to increase the utility of predictive microbiology information, In: *Modelling Microbial responses in Foods*, R.C. McKellar & X. Lu (Eds.), pp. 233-242, CRC Press, ISBN 0-8493-1237-X, Boca Raton, Florida
- US Food and Drug Administration (2009). Safe Practices for Food Processes. Chapter 6. Microbiological Challenge Testing: Evaluation and Definition of Potentially Hazardous Foods. Available from: <http://www.fda.gov/Food/ScienceResearch/ResearchAreas/SafePracticesforFoodProcesses/ucm094154.htm>. Accessed 06/18/2009.
- Valero, A. Carrasco, E. Pérez-Rodríguez, F. García-Gimeno, R.M. Blanco, C. & Zurera, G. (2006). Monitoring the sensorial and microbiological quality of pasteurized white asparagus at different storage temperatures. *Journal of the Science of Food and Agriculture*, Vol.86, pp. 1281-1288, ISSN 0022-5142
- Van Asselt, E.D. & Zwietering, M.H. (2006). A systematic approach to determine global thermal inactivation parameters for various food pathogens. *International Journal of Food Microbiology*, Vol.107, pp. 73-82, ISSN 0168-1605
- Van Boekel, M.A.J.S. (2008). Kinetic modeling of food quality: a critical review. *Comprehensive Reviews in Food Science and Food Safety*, Vol. 7, pp, 144-158. ISSN 1541-4337

- Van Boekel, M.A.J.S. (2002). On the use of the Weibull model to describe thermal inactivation of vegetative cells. *International Journal of Food Microbiology*, Vol.74, pp. 139-159, ISSN 0168-1605
- Van Gerwen. S.J.C. te Giffel, M.C. Van't Riet, K. Beumer, R.R. & Zwietering, M.H. (2000). Stepwise quantitative risk assessment as a tool for characterization of microbiological safety. *Journal of Applied Microbiology*, Vol.88, pp. 938-951, ISSN 1364-5072
- Vestergaard, E.M. (2001). Building product confidence with challenge studies. *Dairy, Food and Environmental Sanitation*, Vol. 21, pp, 206-209. ISSN 1043-3546
- Voyer, R. & McKellar, R.C. (1993). MKES tool: a microbial kinetics expert system for developing and assessing food production systems. *Journal of Industrial Microbiology & Biotechnology*, Vol.12, pp. 256-262, ISSN 1367-5435
- Wang, M.Y. & Brown, W.D. (1983). Effects of elevated CO₂ atmosphere on storage of freshwater crayfish (*Pacifastacus leniusculus*). *Journal of Food Science*, Vol. 48, pp, 158-162. ISSN 1750-3841
- Whiting, R.C. & Buchanan, R.L. (1994). Microbial Modeling. *Food Technology*, Vol.48, pp. 113-120, ISSN 0015-6639
- Whiting, R.C. & Cygnarowicz-Provost, M. (1992). A quantitative model for bacterial growth and decline. *Food Microbiology*, Vol.9, pp. 269-277, ISSN 0740-0020
- Whiting, R.C. (1993). Modeling bacterial survival in unfavorable environments. *Journal of Industrial Microbiology and Biotechnology*, Vol.12, 240-246, ISSN 1367-5435
- Willocx, F. Mercier, M. Hendrickx, M. & Tobback, P. (1993). Modelling the influence of temperature and carbon dioxide upon the growth of *Pseudomonas fluorescens*. *Food Microbiology*, Vol.10, 159-173, ISSN 0740-0020
- Witjes, T. Van't Riet, K. in't Veld, J.H.J. & Zwietering, M.H. (1998). A decision support system for the prediction of microbial food safety and food quality. *International Journal of Food Microbiology*, Vol.42, pp. 79-90, ISSN 0168-1605
- Zwietering, M.H. Jongenburger, I. Rombouts, F.M. & van't Riet, D. (1990). Modelling of the bacterial growth curve. *Applied and Environmental Microbiology*, Vol.56, pp. 1876-1881, ISSN 0099-2240
- Zwietering, M.H. Witjes, T. Wit, J.C. de & Van't Riet, K. (1992). A decision support system for prediction of the microbial spoilage in foods. *Journal of Food Protection*, Vol.55, pp. 973-979, ISSN 0362-028X



Trends in Vital Food and Control Engineering

Edited by Prof. Ayman Amer Eissa

ISBN 978-953-51-0449-0

Hard cover, 290 pages

Publisher InTech

Published online 05, April, 2012

Published in print edition April, 2012

This book is an example of a successful addition to the literature of bioengineering and processing control within the scientific world. The book is divided into twelve chapters covering: selected topics in food engineering, advances in food process engineering, food irradiation, food safety and quality, machine vision, control systems and economics processing. All chapters have been written by renowned professionals working in food engineering and related disciplines.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Antonio Valero, Elena Carrasco and Rosa Ma García-Gimeno (2012). Principles and Methodologies for the Determination of Shelf-Life in Foods, Trends in Vital Food and Control Engineering, Prof. Ayman Amer Eissa (Ed.), ISBN: 978-953-51-0449-0, InTech, Available from: <http://www.intechopen.com/books/trends-in-vital-food-and-control-engineering/principles-and-methodologies-for-the-determination-of-shelf-life-in-foods>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen