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Exposure Assessment to Persistent Organic Pollutants in Wildlife: The Case Study of Coatzacoalcos, Veracruz, Mexico

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1. Introduction

Until the early 70s, it was thought that pollution was a phenomenon circumscribed to zones where pollutants were generated. Because of that, in each country concern was limited to regions where pollutant concentration was higher or its danger was greater. However, it has gradually become aware that pollution is a problem that affects everybody and, because of that, everybody is responsible to control it, regardless of the sites distance where pollutants are produced. Therefore, the problem of pollution has become a global phenomenon. Mankind has always depended on natural resources located in the region where they dwell. Nevertheless, the fast population growth coupled with a fast agricultural and industrial development as well as life style changes have increased emissions of pollutants in different ecosystems.

Persistent organic pollutants (POPs) is a group of compounds chemically very stable, able to travel considerable distances and it is resistant to natural degradation processes, most of them were produced to be used as pesticides and certain chemicals to be used as industrial processes, and others are generated as by-products unintentionally from human activities, such as combustion processes or power generation (PNUMA, 2005). Most of these compounds are highly toxic; they bioaccumulate in human and animal tissue, mainly in the fatty tissues, and can damage different organs and systemic targets such as the liver, kidney, hormonal system, nervous system, etc., of both humans and wildlife. According to the Stockholm Convention held in 2001, there are twelve compounds known as POPs: pesticides (DDT, aldrin, chlordane, dieldrin, endrin, mirex, toxaphene and heptachlor), industrial chemicals (hexachlorobenzene and polychlorinated biphenyls -PCB-) and unintentional compounds (dioxins, furans, PCDD-and PCDF-) [Albert, 2004]. In May of 2009 nine new Chemicals were added to the POPs list: alpha hexachlorocyclohexane, beta hexachlorocyclohexane; hexabromodiphenyl heptabromodiphenyl ether and ether tetrabromodiphenyl pentabromodiphenyl ether and ether chlordecone, hexabromobiphenyl, lindane, pentachlorobenzene, perfluorooctane sulfonic acid, its salts and perfluorooctane sulfonyl fluoride.

POPs main route of entry into the organism is food. However, we cannot ignore environmental exposure (inhalation) and dermal exposure (accidents). Because of their properties, POPs are classed as persistent, bioaccumulative and toxic. Therefore, POPs are to be considered as one of the most harmful groups of Toxic environmental pollutants to humans and wildlife. In countries where these compounds have been used are frequently found residuals in food. They are a problem because of their persistence in the environment and characteristics of bioaccumulation and biomagnification along the food chain and because these compounds generate toxic effects in both human population and in biota.

POPs, mainly organochlorine compounds make up a big part of hazardous waste. Mexico annually generates approximately 8 million tons of hazardous waste. Of this amount, only 12 percent is handled properly. The question is: Where does the remaining 88% go? To make matters worse, Mexico's infrastructure for hazardous waste is by far insufficient.

Coatzacoalcos, Veracruz is one of the most commercial and industrialized ports in Mexico. Presently, the Coatzacoalcos River and the areas surrounding it are regarded by many as some of the most heavily polluted sites in Mexico. Several of Mexico's chief petrochemical complexes, such as Cangrejera, Morelos and Pajaritos are based in the region. Furthermore, there have been various toxic substances present in the area which have been stored inside environmental and biological compartments, including persistent organic pollutants (Espinosa-Reyes et al., 2010; Gonzalez-Mille et al., 2010; Stringer et al., 2001), polycyclic aromatic hydrocarbons, volatile organic compounds (Riojas-Rodriguez et al., 2008), polybrominated compounds (Blake 2005), dioxins and metals (Petrlink & DiGangi 2005; Rosales 2005; Vázquez-Botello 2004).

A number of POPs have been registered in Coatzacoalcos. One of these, Hexachlorocyclohexane (HCH), is a manufactured chemical from which there are, theoretically, eight chemical forms or isomers. The three most common isomers are α -HCH, β -HCH and γ -HCH (commonly called lindane). Lindane is used as a pesticide on fruit and vegetable crops as well as on forest plantations; it is also found in medications to treat diseases such as scabies and pediculosis. There are no records indicating that lindane has ever been manufactured in Mexico; however, approximately 20 tons of these compounds are imported and subsequently used in Mexico each year. At present, lindane is authorized for use in Mexico for ectoparasite control in livestock for ticks, fleas, and common fly larvae. It is also registered for use as a seed treatment for oats, barley, beans, corn, sorghum and wheat. Pharmaceutical uses of lindane in Mexico include the formulation of creams and shampoos for scabies and lice treatment (CEC, 2006; ATSDR, 2005). In 1994, the Canadian Environmental Protection Act (CEPA) proclaimed hexachlorobenzene (HCB) as highly toxic. HCB is a manufactured chemical (which was) used as a wood preservative, as a fungicide for treating seeds and as intermediary in organic syntheses. Additionally, hexachlorobenzene can be formed as an unwanted byproduct in preparation processes like in the synthesis of organochlorines from high-temperature sources (Sala et al., 1999; Newhook & Meek 1994). Dichlorodiphenyldichloroethane (DDT) is a synthetic organochloride which is relatively stable with slow degradation rates through sunlight or oxidation, and possesses good absorption capacity and resistance to biodegradation in sediments and soils. It is also insoluble in water (CEC, 2001). In 1945, DDT was used for the first time in Mexico for the control of Malaria, and was widely used in agriculture between the 50s and 70s (CEC, 1997). The use of DDT in the Malaria Control Program was abandoned in the year 2000, when it was replaced by pyrethroids. Polychlorinated

biphenyls (PCBs) are allowed in “totally enclosed uses” such as coolants and lubricants, in transformers and capacitors. Dioxins are produced during the combustion of organic materials containing chlorine as well as during the manufacture of various chlorine-containing chemicals, such as ethylene dichloride. Existing involuntary sources of intake include electric arc furnaces, shredders, sinter plants, cement plants, cremation facilities, and coal-based power plants (Lutharddt et al., 2002).

Biomonitoring wildlife can be used to detect chemical pollution and to evaluate the ecosystem's health, using test species as systematic models in the evaluation of risks associated to paths of real exposure. Wildlife species residing in polluted sites are exposed to complex mixtures of pollutants through multiple pathways which could hardly be evaluated in lab studies. The main purpose of this research was to pinpoint exposure levels to POPs in wildlife from different sets of ecosystems throughout the industrial area of Coatzacoalcos, Veracruz, Mexico to obtain a baseline of the ecological condition of this region.

2. Materials and methods

2.1 Test area, sampling sites and species selection

The Coatzacoalcos region is located in the South eastern State of Veracruz, Mexico, in the municipality under the same name, at 18° 8' 56 " N and 94° 24' 41" W. The average altitude is 14 m.a.s.l. The predominant climate is tropical rain [Am (i) gw"], the average annual temperature is 24.5°C and average annual rainfall is 2780.1 mm (García, 2004). The main inland body of water is the Coatzacoalcos River, which has an area of 322 km. It originates above 2000 m in elevation in the State of Oaxaca and over its course is fed by countless other rivers (Jaltepec, Coachapa, Uxpanapa, Calzada) and streams (Teapa, Tepeyac, San Francisco) which inflows contribute to the discharge of pollutants (Páez-Osuna et al., 1986.; Rosales-Hoz & Carranza-Edwards, 1998). The region is comprised of urban, industrial, livestock, riparian and wetland areas. However, its main activity is chemical, namely petrochemical (Ruelas-Inzunza et al., 2007).

In October 2006, six sampling stations for biological sampling were set up at the lower basin of the Coatzacoalcos River (Fig. 1). The selection of sampling sites was based on wind direction, location of industrial zones and urban areas, the presence of organisms and the influence of riparian systems as well as on previous investigations within the area (Páez-Osuna et al., 1986; Rosales-Hoz & Carranza-Edwards 1998; Stringer et al., 2001; Bahena-Manjarrez et al., 2002.).

In this research we selected earthworms, crabs, fish, toads, turtles, iguanas, and crocodiles to measure levels of POPs in muscle or blood. These groups are critical species because they have an important role in the ecosystems dynamics and/or an importance (economic, cultural and scientific) for man. Species were selected according to the following criteria: The kind of pollutant located in the study area. Based on literature, a revision on the pollutant environmental behavior was conducted, considering their physiochemical characteristics as well as environmental parameters (humidity, temperature, pH, type of soil, etc.) that can influence in the environment pollutant levels. Once pollutant groups to evaluate were determined, potential pathways and exposure routes were established. An important criterion that was also considered when selecting animal groups was their biology, as it should be well documented; finally groups that are relatively easy to capture and handle were selected.

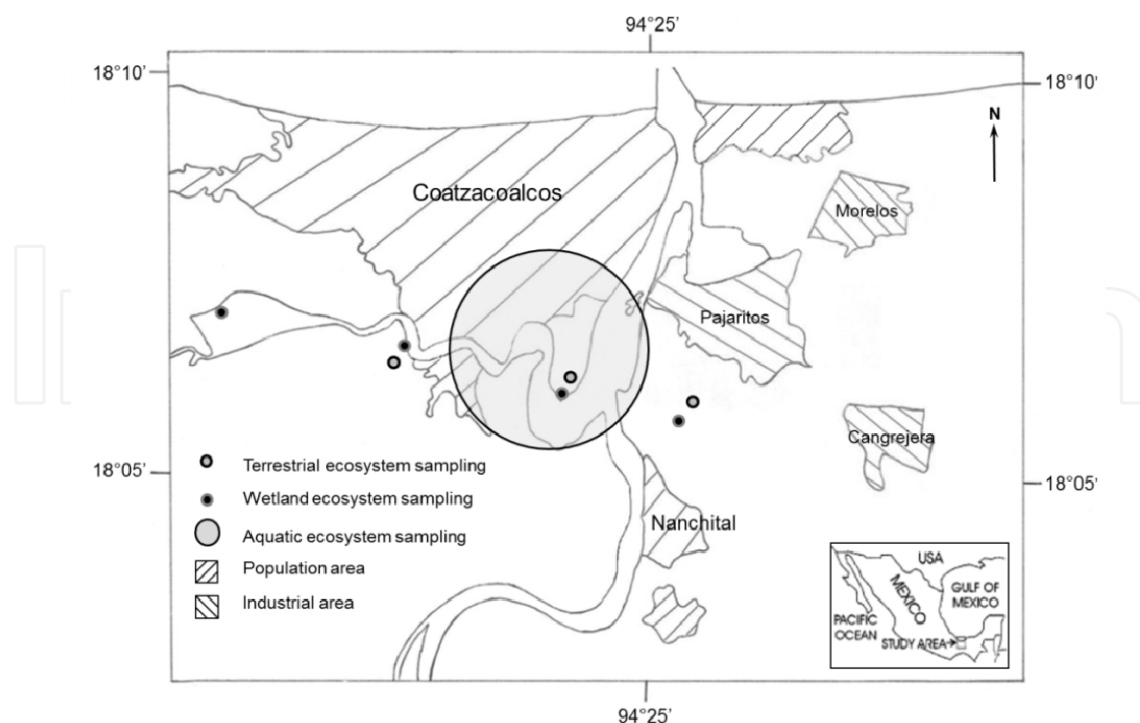


Fig. 1. Study area and location of sampling sites in the region of Coatzacoalcos, Ver.

2.2 Ecological importance of selected species

2.2.1 Earthworms

Earthworms' ecological importance is that they are decomposer organisms (important in biogeochemical cycles) and because of that, they have an important role when adding nutrients to soil (they favor the availability of nitrogen, phosphorus and sulfur) which can be used by vegetable species (Reines et al., 1998; Legall, 2006); they are also an important link in the trophic chain, mainly for some bird species and it has been proven that earthworms can accumulate important metal concentrations (Sánchez-Hernández, 2006). Earthworms can be found in many types of soil and are vulnerable to impacts occurred in soil, their small size represents an advantage to be handled; their distribution is ubiquitous in edaphic horizons with detritus, they are easy to capture, have a close contact with soil, and have a short life-cycle favoring the study of several generations (Ogunseitan, 2002). Persisting organic pollutants have the capacity to bioaccumulate and biomagnify along the trophic chain, as well as animals that belong to decomposers or detritivores levels are very important for the ecosystem functioning, therefore, if animals that are part of soil are affected by pollutants that may show in the ecosystems' health.

2.2.2 Crabs

Crabs are ubiquitous in all temperate and tropical regions in the world. In the wetland ecosystems they are an ecologically important kind, because they play a primary role in the decomposition of organic material and the addition of nutriment to soil. Due to the fact that they build up their galleries by the rivers' basin, lakes or seas, they spend part of their life on land surface and when tides rise they shelter in their burrows (≈ 30 cm. deep) under water. They mainly feed themselves with detritus, so they are excellent filtering organisms, capable to accumulate great pollutant quantities.

2.2.3 Iguanas

There are different kinds of iguanas, however the most common ones in the Coatzacoalcos region are the green iguana (*Iguana iguana*) and the black iguana (*Ctenosaura* spp.); Distribution of both goes from the south of Mexico to South America. According to Lara-Lopez and Gonzalez-Romero, (2002), iguanas are herbivores, the diet of the green iguana is composed mainly as follows: leaves (57.36%), flowers (24.15%) and fruits (3.43%), most of their life is spent on top of the trees, because of the previously said, this specie may be used as a POPs bio-monitor in air. In many coastal communities in Mexico, it is usual to eat iguana as an important source of protein, and it is believed that their blood and eggs contain a lot of energy and help to prevent certain diseases such as anemia. Furthermore people use the skin of this reptile and dissected to sell as ornaments (Alvarez del Toro, 1982). In southern Mexico (Leon & Montiel, 2008), in Central America (FAO, 1997) and in parts of South America, the iguana is one of the most consumed wildlife species. Iguanas have been an important source of protein for humans for over 7000 years FAO (1997). Many of the rural inhabitants of Central America still rely on iguana as a protein source; however, consumption of these species are not uniform during the year because in most cases eating frequency depends on the availability in certain times or seasons that are usually 3 to 4 months per year (Pers. Obs.).

2.2.4 Fish and invertebrates

Fish and aquatic invertebrates are commonly used to monitor pollutants because they bioaccumulate toxic substances and are wide spread, coupled with the diversity and importance of these environments. It has been observed that they are highly sensible to changes in the aquatic environment as well as to low concentrations of environmental pollutants, (Russo et al., 2004; Klobučar et al., 2010). On the other hand, fish have a crucial position in the toxicological field, due to the fact that they have been widely used in studies related to human and ecological health. Fish study includes a wide variety of approaches to detect aquatic pollution impacts from direct measures of mortality, to the analysis of demographical dynamics and the community structure, and to the detection of measures of sub cellular changes (Di Giulio & Hinton, 2008).

At the same time aquatic invertebrates and specifically crustaceans are organisms that have a wide distribution (example: marine, terrestrial and freshwater environment), they are organisms that are in close contact with pollutants in sediment, so they have been used in countless eco-toxicological studies. They have proven to be useful to evaluate effects on different pollutants. They have also served as aquatic pollution indicators. In addition, they may be a source or exposure for local consumers (Nacci et al., 1996; Rinderhagen et al., 2000; Rigonato et al., 2005; Kulköylüolu 2004; Regoli et al., 2006). Otherwise, spatial distribution of pollutants on sediments and biota in aquatic ecosystems have been related to a great variety of biological answers in populations and fish/invertebrates communities, with the purpose to determine a possible relation between pollutants in the environment and health in the organisms (Adams et al., 1999).

2.2.5 Giant toads

The giant toad (*Rhinella marina*, after *Bufo marinus*) is a native and geographically widespread species in Mexico and Central America (Zug & Zug 1979). It is an omnivorous and opportunistic species (Zug & Zug 1979), which indicates that toads would integrate different exposure paths due to the ingestion of a wide variety of food items and amphibious living habits. The giant toad is one of the largest amphibians in Mexico (adult

body length ranges from 10 to 17 cm), with a life expectancy from 10 to 15 years in the wild. The high lipid-somatic index (2 to 10% compared to less than 0.1% in most anuran species after the spawning period) and the elevated hepatosomatic index (Feder & Burggren 1992) along with its breeding biology make this species prone to bioaccumulation of organic and inorganic pollutants and their toxicological effects (Sparling et al., 2010; Linder et al., 2003). Recently, the giant toad has been used as an aquatic ecosystem biomonitor in the evaluation of air pollution (Dohm et al., 2008), infectious diseases (Zupanovic et al., 1998), organochlorine pesticides (Linzey et al., 2003) and endocrine disruptors (McCoy et al., 2008).

2.2.6 Turtles

Slider turtle (*Trachemys scripta*) is geographically widespread across Mexico and Central America (Burger and Gibbons, 1998). They are eligible species because they have several characteristics associated with their metabolism, life history and ecology (Overmann & Krajicek, 1995). These turtles are generally omnivorous and have temperature-dependent of sex determination and studies have investigated contaminant effects on this process (Selcer 2006), this makes them ideal for studies of chronic exposure of local pollutants. This species has been employed for exposure assessment to metals, radiation, organochloride pesticides and polybrominated biphenyls (Bergeron, et al., 1994; Bickham, et al., 1998; Burger and Gibbons, 1998; Lovelette & Wrigth, 1996; Meyers-Schöne & Walton 1994; Willingham et al., 1999; Willingham et al., 2000). In Mexico, slider turtles are considered an endangered species and are protected by Mexican laws.

2.2.7 Crocodiles

Swamp crocodiles (*Crocodylus moreletii*) are aquatic reptiles living in Mexico's tropical regions. They have a life strategy based on late maturation; they are extremely long-lived animals, show parental care and determine sex depending on temperature (Selcer, 2006). Crocodiles reach the highest levels of food chains, so they are useful for the evaluation of persistent and biomagnifying pollutants. Around the world, their populations are endangered (including Mexico), and because of that the concern on effects (mainly reproductive ones) of pollutants in their populations has increased (Guillette et al., 1999); it must be mentioned that the highest DDE levels registered in wild reptile were found in these organisms (De Solla, 2010). Different Crocodylia species have been used to evaluate heavy metals as well as persistent organic compounds.

2.3 Biological sampling techniques

Wild Earthworms (*Eisenia* sp) were collected by excavation. Crabs were harvested using pitfall traps (*Uca* sp) and traditional fishing gear (*Callinectes* sp). fish (*Aplodinotus* sp, *Ariopsis felis*, *Centropomus parallelus*, *Eucinostomus* sp, *Eugerres axillaris*, *Gobiomorus* sp, *Menticirrhus* sp, *Mugil cephalus* and *Oreochromis* sp) were caught using traditional fishing gear (i.e. cast net) with the help of fishermen. Giant toads (*Rhinella marina*) were collected from each site using nets in nocturnal transects within an area of 10,000 m². Crocodiles (*Crocodylus moreletii*) and Iguanas (*Iguana iguana*) were caught using a noose trap. Turtles (*Trachemys scripta*) were captured with a baited piper trap placed near fallen trees and along the edge of the river during the afternoon and checked early the following morning.

Immediately after capture, organisms were measured, weighed and sorted by type of species; type of ecosystem (terrestrial, aquatic and wetland) and by feeding behaviours (carnivores, omnivores, detritivores and herbivores). Blood samples drawn were obtained using

heparinized syringes on endangered animals (turtles and crocodiles). All organisms were subsequently released. Samples were stored at 4°C for transport and subsequent laboratory analysis. Dissection was performed on each of the specimens from the rest of different species to extract the muscle tissue. The tissue was placed in amber glass containers and frozen at -20°C until analysis. All organisms were collected with a Scientific Collector’s Permit (Wild Fauna and Flora Scientific Collector) issued by México’s SEMARNAT (Ministry of Environment) or Secretaría de Medioambiente y Recursos Naturales-No. FAUT-0133.

2.4 Analysis of blood and tissue residues

Concentrations of the following compounds were tested for on biological samples: α-, β-, γ-hexachlorocyclohexane (HCH), hexachlorobenzene (HCB), aldrin, dieldrin, mirex, α-, γ-chlordane, oxychlordane, trans-, cis-nonachlor, heptachlor epoxide, p, p'-DDT, p, p'-DDE, polychlorinated biphenyls (PCBs, IUPAC No 28, 52, 99, 101, 105, 118, 128, 138, 153, 156, 187, 180, 183, and 170) and polybrominated diphenyl ethers (PBDE, only on some species of fish). The method of extraction, separation and cleaning of muscle tissue was carried out according to the method established by Jensen et al., (2003) with slight modifications (Gonzalez-Mille et al., 2010) and Dallaire et al., (2006) for blood samples. The endrin-C13 and PCB 14-C13 were used as internal standards and were added to all samples. The chromatographic method (Gas chromatography-mass spectrometry GC-MS) was carried out according to that reported by Trejo-Acevedo et al., (2009). The detection limit for POPs was approximately 0.3 mg/L.

3. Results and discussion

3.1 Terrestrial ecosystem

In Table 1, it can be observed that earthworms have the highest concentrations of polychlorinated compounds biphenyls (PCBs) y persistent organic pollutants (POPs), followed by iguanas and finally by crabs. PCBs congeners that were analyzed were (PCBs 105, 128, 138, 153, 156, 170, 180 y 183).

ECOSYSTEM	SPECIES	α-HCH	β-HCH	γ-HCH	DDT	DDE	Mirex	ΣPCBs	ΣPOPs
TERRESTRIAL	Eisp	12.8	N.D.	106.3	2.5	13.2	N.D.	13.4	146.2
	(n=6)*	(5.8 - 31.4)		(39.4 - 196.0)	(2.5 - 2.5)	(0.3 - 57.2)		(2.3 - 39.2)	(50.3 - 323.8)
	Igig	4.76	0.59	0.55	N.D.	0.04	0.36	0.11	6.42
	(n=3)	(4.33 - 5.41)	(0.48 - 0.71)	(0.45 - 0.66)		(N.D. - 0.06)	(0.27 - 0.49)	(N.D. - 0.22)	(5.97 - 6.82)
	Ucsp	0.24	0.37	0.06	0.02	0.04	0.73	0.04	1.52
	(n=2)*	(0.17 - 0.31)	(N.D. - 0.74)	(0.05 - 0.08)	(0.020- 0.026)	(0.04 - 0.05)	(0.72 - 0.74)	(N.D. - 0.09)	(1.25 - 1.78)

Values represent the mean and range. Eisp: *Eisenia* sp., Igig: *Iguana iguana*, Ucsp: *Uca* sp., * Pool samples, N.D.: non detected

Table 1. Concentrations of persistent organic pollutants (ng/ g tissue) from terrestrial wildlife collected in Coatzacoalcos, Veracruz.

Registered results of iguanas (*Iguana iguana*) are relevant because until now there are no studies showing POPs exposure background. In addition, because they are a basic part of rural communities' diet, meat intake may be a potential route of exposure to organic pollutants persistent for humans. Furthermore, because it is a tree species, it may be used to indirectly monitor air quality of some volatile organic compounds (VOCs) and semi-volatile ones as DDT and their metabolites, as well as some congeners of PCBs.

With crabs (*Uca* sp.) there are a few exposure studies to POPs. De Sousa et al., (2008) that registered POPs concentrations in crab eggs (*Chasmagnathus granulata*) in different Brasil stereos. Concentrations of DDE, DDT, γ -HCH, PCBs y total POPs are higher (35.95; 0.38; 3.52; 286.27 and 339.68 ng/g) than those reported in the present work. The previously said may be due to the fact that the matrix analyzed by de Sousa et al., contains a greater quantity of lipids, and crabs evaluated in this study (*Uca* sp.) were not in reproductive stage. Bayen et al., (2005) a study was conducted in a Singapore's mangrove swamp where a thropic web was established and the thunder crab (*Myomenippe hardwicki*) was one of the species presenting high concentrations of POPs. Falandysz et al., (2001) also used crabs (*Carcinus means*) to evaluate the exposure to organochlorine pesticides. It is complicated to make a comparison between crab species because they have different etiology and habitants, however, because most crab species are detritivores, we consider them as a good option to do studies related to POPs exposure.

There are several studies on earthworms with ecotoxicological background mainly focused on effects at population levels (lethality), at both laboratory controlled conditions (Heimbach, 1984; Ma & Bodt, 1993; Kula, 1995, Morrison et al., 2000) as in field ones (Thompson, 1970; Tomlin, 1981; Edwards & Brown, 1982; Haque & Ebing, 1983; Potter et al., 1994; Espinosa-Reyes et al., 2010), however, there are a few studies where exposure to POPs are evaluated, Jones and Hart (1998) revised works related to exposure of different earthworms species to various pesticides-Benomyl, Carbaryl, Carbendazim, Carbofuran, Chlordane, Methiocarb, Parathion, Pentachlorophenol, Phorate, Propoxur, Thiophanate-methyl. These same authors mentioned that earthworms of the *Eisenia* type are the most resistant to the pesticides previously mentioned. Morrison et al., (2000) evaluated the bioavailability of DDT, DDE, DDD y Dieldrin in soil samples with different age of use of the mentioned pesticides. They exposed *Eisenia* earthworms. The concentrations registered by Morrison et al., (2000) are high when compared with the registered in this study (DDT 28.3 ± 8.4 & DDE 3.77 ± 0.48 mg/Kg tissue). Based on these backgrounds and results registered in this study, it is possible to postulate the *Eisenia* earthworms as POPs biomonitors in terrestrial ecosystems because earthworms play a major role in facilitating pivotal interactions within ecosystems through the mixing and translocation of soil constituents, or serving as a conduit for contaminants to predators at higher trophic levels (Harris et al., 2000; Langdon et al., 2003). Earthworms have been used extensively in ecotoxicology like biomonitors (Fitzpatrick et al., 1992; Goven et al., 1993; Reinecke & Reinecke, 1998; Espinosa-Reyes et al., 2010) to assess the effects of diffuse contaminants present in soils.

Finally, when doing a POPs biomonitoring in terrestrial ecosystems, it is important to take into account the following factors: a) species susceptibility difference; b) soil type; c) species behavior; d) exposure time; e) pollutant toxicity; f) when using pesticides consider the applying method.

3.2 Aquatic ecosystem

Because fish are organisms with a wide movement range, the sample was taken within an approximate area of 8 Km². Thirty-one fish from five species were caught: *Centropomus parallelus*, *Mugil cephalus*, *Eugerres axillaris*, *Oreochromis* sp, *Ariopsis felis* and thirty organisms of one crustacean species *Callinectes* sp., for quantification of POPs.

From de 29 quantified compounds in the simple, only were detected concentrations of HCB, α -, β -, γ -HCH, DDT, DDE, mirex and 6 congeners of PCBs (52, 101, 105, 118, 138, 153) (Table 2). Most concentrations registered were of β -HCH, α -HCH and mirex. HCB was only detected in 26% of the samples and just on species *Mugil cephalus* y *Callinectes* sp. El γ -HCH and DDT were registered in a 32% and 10% of the samples respectively. Pollutants α -HCH, β -HCH, DDE, mirex and PCBs were detected in a 100% of the samples. Species where a higher number of compounds were found are *Callinectes* sp, *Ariopsis felis* y *Eugerres axillaris* (8, 7 y 6 respectively), however, most concentrations were found in *Callinectes* sp and *Eugerres axillaris*. According to these results, POPs concentrations in muscular tissue by species decrease in this order β -HCH > α -HCH > mirex > DDE > Total PCBs > γ -HCH > DDT > HCB.

ECOSYSTEM	SPECIES	HCB	α -HCH	β -HCH	γ -HCH	DDT	DDE	Mirex	Σ PCBs	Σ POPs
AQUATIC	Cepa	N.D.	1.2	0.1	N.D.	N.D.	0.1	0.3	0.2	1,8
	(n=9)		(0.05-5.3)	(0.05-0.5)			(0.05-0.5)	(0.05-0.6)	(0.05-0.4)	(0.5-5,9)
	Muce	0.1	0.2	0.3	N.D.	N.D.	0.2	0.3	0.1	1.0
	(n=7)	(0.05-0.2)	(0.05-0.3)	(0.05-0.7)			(0.05-0.5)	(0.2-0.4)	(0.05-0.2)	(0.6-1,5)
	Euax	N.D.	0.4	1.7	0.1	N.D.	0.1	0.6	0.1	2.9
	(n=7)		(0.03-1.9)	(0.05-3.8)	(0.05-0.3)		(0.05-0.1)	(0.1-0.9)	(0.1-0.2)	(0.9-4,7)
	Orsp	N.D.	0.3	N.D.	N.D.	N.D.	0.2	0.4	N.D.	0.8
	(n=5)		(0.2-0.4)				(0.1-0.2)	(0.3-0.5)		(0.7-1,0)
	Arfe	N.D.	0.3	N.D.	0.1	0.1	0.1	0.5	0.3	1.3
	(n=3)		(0.2-0.5)		(0.05-0.1)	(0.05-0.1)	(0.05-0.2)	(0.4-0.7)	(0.1-0.6)	(1,1-1,5)
	All fishes	0.1	0.6	0.9	0.1	0.06	0.2	0.4	0.2	1.7
	(n=31)	(0.05-0.2)	(0.03-5.3)	(0.05-3.8)	(0.05-0.3)	(0.05-0.1)	(0.05-0.5)	(0.05-0.9)	(0.05-0.6)	(0.5-5,9)
	Casp	0.2	1.2	1.6	0.2	0.1	0.4	1.3	0.3	14.8
	(n=4)*	(0.05-0.5)	(0.3-2.4)	(0.05-5.2)	(0.05-0.2)	(0.05-0.1)	(0.1-0.9)	(0.9-1,7)	(0.1-0.7)	(13,3-17,6)

Values represent the mean and range. Cepa: *Centropomus parallelus*, Muce: *Mugil cephalus*, Euax: *Eugerres axillaris*, Orsp: *Oreochromis* sp, Arfe: *Ariopsis felis*, Casp: *Callinectes* sp. N.D.: non detected

Table 2. Concentrations of persistent organic pollutants in muscle tissue (ng/g wet weight) from aquatic wildlife collected in Coatzacoalcos, Veracruz.

Out of the *Ariopsis felis* fish species the highest concentrations of PCBs were registered and it is the only one that presented DDT, *Mugil cephalus* is the only species in which HCB was detected and it presented the highest concentrations of DDE, *Eugerres axillaris* showed the highest concentrations of lindane and α , β -HCH's, finally *Oreochromis* sp registered the lowest load of POPs in comparison to the other species.

Concentrations of β -HCH and α -HCH were the most abundant among the evaluated species, which matches with what was registered by Lee et al., (1997) and Yim et al., (2005) in other fish species. This can be explained because β -HCH and α -HCH have a greater bio-concentration factor (log BCF 2.8 and 2.5 respectively) in aquatic animals and they are more persistent than γ -HCH (log BCF 1.2) (Willett et al., 1998). Even though technical grade HCH has a greater constitution of α -HCH (60-70%), concentrations of β -HCH and α -HCH were very similar, this can be due to the fact that α -HCH has a high volatility (Henry' law constant 6.68×10^{-6} atm-3/mol) and environment degradation, which can cause low persistency (Yim et al., 2005). Another possible explanation is that HCH metabolized and excreted faster than β -HCH (Takazawa et al., 2005).

DDT was only detected in *Ariopsis Felis* while DDE was detected in all species. Most concentrations were registered in detritivores fish. As it was mentioned, DDT was widely used in Mexico for malaria control and because of its toxic effects, its use was eliminated. Chemical and biological processes transformed DDT into DDD and DDE, particularly DDE has been the most registered in biota (Takazawa et al., 2005). Registered concentrations in fish from the zone indicate that they are mainly exposed to residual DDT and to its degradation products. However, as it was seen, DDT/DDE relation in sediment suggests current use, reason why it is expected to have greater concentrations in the tissues, which suggests that fish from the zone have the capacity to eliminate DDT fastly.

Furthermore, quantification of polybrominated diphenyl ethers (PBDE) was done in 30 fish that belong to different species (*Aplodinotus* sp, *Centropomus parallelus*, *Mugil cephalus*, *Eucinostomus* sp, *Gobiomorus* sp, *Menticirrhus* sp and Torombolo –common name-), in these organisms, there were concentrations of 6 congeners registered (PBDE 47, PBDE 100, PBDE 99, PBDE 154, PBDE 153, PBDE 209) (Table 3). Considering all species, the order of concentration of congeners found was $47 > 41 > 154 > 209 > 153 > 100$, being *Eucinostomus* sp species where the highest concentrations were found and in Torombolo species the lowest ones.

Within this context, aquatic ecosystems are highly vulnerable because of their tendency to accumulate concentrations relatively greater of pollutants coming from terrestrial ecosystems surrounding them, as well as those from direct entries (downloads), in that, regardless of its source of entry into the environment, aquatic systems are frequently deposits for a great variety of chemicals. Pollution in these environments can have negative effects over aquatic life (example: alteration of reproduction and decreased of species) as well as directly or indirectly affect human health and threat food safety. (Jha, 2008).

3.3 Wetland ecosystem

In Table 4 are presented the levels of persistent organic compounds detected in the different animal species from the Coatzacoalcos wetland. When monitoring them, 6 organochlorine pesticides out of the 14 ones analyzed were detected and 13 polychlorinated compounds biphenyls (PCBs) out of 21. All captured organisms from the different species presented detectable levels of at least 3 persistent organic compounds: DDE, Lindane y PCBs (data not

shown, found in giant toads’ adipose and hepatic tissue). Generally, the pattern of pollutants presence in giant toads’ muscular tissue was $\sum\text{DDT} > \sum\text{HCH} > \text{Mirex} > \text{HCB}$; and for the turtles and crocodiles’ case, the exposure levels of PCBs and DDE in serum were similar. It must be said that lindane and DDT concentrations in crocodiles were higher.

SPECIES	PBDE 47	PBDE 100	PBDE 99	PBDE 154	PBDE 153	PBDE 209	Σ PBDEs
Mesp (n=7)	3.2	1.9	4.5	0.3	0.7	0.7	11.3
	(0.5-6.5)	(0.5-4.3)	(1.0-8.0)	(0.04-0.5)	(0.3-1.2)	(0.3-1.2)	(3.8-19.7)
Eusp (n=2)	12.03	3.7	2.6	18.0	10.2	13.2	59.7
	(7,7-16,4)	(0,8-6,5)	(1,7-3,5)	(9,5-26,5)	(1,5-9,2)	(8,6-17,7)	(43,1-76,2)
Muce (n=7)	5.5	1.6	5.4	1.3	1.1	0.9	15.8
	(0,2-24,8)	(0,1-2,9)	(0,5-10,6)	(0,03-4,2)	(0,1-2,0)	(0,1-2,2)	(1,0-45,1)
Gosp (n=1)	2.9	2.3	8.8	0.8	1.2	1.4	17.3
Apsp (n=3)	1.9	0.7	2.4	0.2	0.3	0.3	5.7
	(1,1-2,7)	(0,5-1,0)	(1,21-3,7)	(0,1-0,2)	(0,1-0,4)	(0,1-0,6)	(3,2-8,7)
Torombolo (n=2)	2.0	0.4	1.3	0.1	0.2	0.2	4.3
	(1,3-2,8)	(0,2-0,5)	(0,8-1,9)	(0,1-0,2)	(0,1-0,3)	(0,1-0,3)	(2,6-6,0)
Cepa (n=8)	2.2	1.0	3.7	0.2	0.5	0.5	8.1
	(1,0-4,0)	(0,5-1,4)	(0,8-5,5)	(0,1-0,5)	(0,2-0,6)	(0,1-0,8)	(3,1-12,6)
All fish (n=30)	4.2	1.7	4.1	3.0	2.0	2.5	17.5
	(0,2-24,8)	(0,1-6,5)	(0,5-10,6)	(0,03-26,5)	(0,1-9,2)	(0,1-17,7)	(1,0-76,2)

Values represent the mean and range. Mesp: *Menticirrhus sp.*, Eusp: *Eucinostomus sp.*, Muce: *Mugil cephalus*, Gosp: *Gobiomorus s.*, Apsp: *Aplodinotus sp.*, Torombolo: -common name-, Cepa: *Centropomus parallelus*.

Table 3. Concentrations of polybrominated diphenyl ethers (ng/g lipid) in fish muscle tissue from species collected in Coatzacoalcos, Veracruz.

ECOSYSTEM	SPECIES	HCB	α -HCH	β -HCH	γ -HCH	DDT	DDE	Mirex	Σ PCBs	Σ POPs
WETLAND	Rhma	0.08	0.75	0.31	0.26	ND	1.51	0.39	ND	3.1
	(n=12)	(N.D.-0.15)	(0.41-2.05)	(N.D.-1.24)	(N.D.-0.53)		(0.05-7.8)	(N.D. 0.67)		(0.84-9.56)
	Crmo†	N.D.	N.D.	N.D.	64.5	59.2	11.5	N.D.	2.7	76.0
	(n=2)				(N.D. - 64.5)	(N.D. - 59.2)	(N.D. - 11.5)		(2.5 - 2.7)	(72.7 - 79.2)
	Trsc‡	N.D.	N.D.	N.D.	3.5	N.D.	8.8	N.D.	3.0	7.0
	(n=4)				(N.D. - 4.58)		(N.D. - 8.78)		(N.D. - 5.83)	(N.D. - 14.61)

Values represent the mean and range. Rhma: *Rhinella marina*, Crmo: *Crocodylus moreletii*, Trsc: *Trachemys scripta*. * Pool samples, ‡Blood samples, N.D.: non detected

Table 4. Concentrations of persistent organic pollutants in muscle tissue (ng/g wet weight) and blood serum (ng/ml) from wildlife collected in Coatzacoalcos, Veracruz.

The DDT/DDE relation obtained in toads' tissue was lower than 1.0 which suggests the organism's ability to metabolize the parental compound to DDE. Studies on amphibians, especially the toads' family (Bufonidae), has proven these organisms capacity to accumulate high concentrations of DDT and its metabolites. It should be said that these studies have also shown that proportions of DDE and DDT were higher than those of the parental compound. In our case, DDD concentrations in tissues were not quantified, so exposure of toads from the Coatzacoalcos region to DDT and its metabolites may be underestimated. Registered DDE concentrations in this research are comparable with those found in adult anuran coming from other polluted sites (Table 4 and 5). There are records of Σ DDT concentrations and their metabolites (mainly DDE) up to 3480 $\mu\text{g/g}$ of fat in samples of *R. clamitans* coming from the wetlands from the south of Ontario (Harris et al 1998); in our study, we found concentrations that go up to 3094.5 $\mu\text{g/g}$ of fat in samples of giant toad's liver (data not shown). POPs exposure works in fresh water turtles' populations are scarce; studies done in Terrapin (*Malaclemys terrapin*), Snapping turtle (*Macrochelys temminckii*) and Common musk (*Sternotherus odoratus*) have determined levels of DDE of 0.73-21.7 ng/g wet mass (Basile, 2010, Moss et al., 2009, De Solla et al., 1998). Several studies on marine turtles have proven levels of DDE of 0.06- 0.73 ng/g wet mass (Stewart et al 2011). Our study shows comparable levels between species of terrestrial and fresh water turtles. Crocodiles present the highest levels of DDT and DDE in plasma; be noted that higher organochlorine pesticides concentrations in the world (mainly DDE) have been found in the Crocodilia species due to their position in the trophic chain. Guillette et al., (1999) Reported DDE levels of 0.9- 17.9 ng/ml and DDT levels of 0.45 to 0.70 in plasma of alligator's residents at Apopka Lake in florida; our study shows comparable levels; even greater to the ones found in Apopka Lake.

The distribution general pattern of Hexachlorociclohexanos (HCH's) observed in toads' tissues was $\alpha\text{-HCH} > \beta\text{-HCH} > \gamma\text{-HCH}$ and for the crocodiles and turtles' case, levels of $\gamma\text{-HCH}$ were detected. In accordance with literature, general distribution patterns of HCH's in mammals, birds and fish are $\beta\text{-HCH} > \alpha\text{-HCH} > \gamma\text{-HCH}$; this distribution pattern is mainly determined by the compound persistency, the exposure path, the species metabolism, and the trophic position (Willet et al., 1998). Our data contrast with the distribution pattern observed in other studies of environmental and biological matrix. Some possible explanations in order to interpret the presence of a higher proportion of $\gamma\text{-HCH}$ is the chronic exposure to isomer of different routes, which can be noted with the found values in crocodile and turtle's plasma where isomer $\gamma\text{-HCH}$ is the only detectable one (Table 4). El $\gamma\text{-HCH}$ is more volatile in comparison with the other isomers, which implies an important transport by air, it is also the isomer with most solubility in water (Walker et al 1999). Coupled to the previously said, it is possible that other important exposure routes exist towards HCH's which have not been explored yet and which may significantly contribute to corporal load and its distribution to these animals. Even though differences in isomers proportions may indicate different sources, routes and times of exposure, for many species it is not clearly understood the influence of processes such as intake, distribution, metabolism and storage in the differences of isomer distribution in tissues (Willet et al., 1998). HCH's concentrations detected in this study are lower than those observed in other studies done on wild amphibians living in agricultural sites (Table 4 and 5). In the case of turtles and crocodiles, there was no useful information found to compare data with.

SPECIE	COMPOUND	CONCENTRATION	TISSUE	REFERENCE
<i>Pseudacris crucifer</i>	DDE	1001	Whole	Russell et al 1995
	DDT	160.6		
	α- HCH	0.37		
	β- HCH	1.37		
	γ- HCH	<DL		
<i>Rana clamitans</i>	DDE	0.58-45.0	Whole	Russell et al 1997
	HCB	0.08-0.49		Guilliland 2001
	ΣDDT	1.24		
	ΣHCH	0.12		
<i>Rana perezi</i>	DDE	<DL-190	Muscle	Rico et al 1987
	ΣDDT	50-550	Whole	Pastor et al 2004
	γ- HCH	<DL-10		
	ΣDDT	35.4		
	HCB	2.7		
	α, γ- HCH	0.5		
<i>Rana pretiosa</i>	DDE	91-173	Whole	Kirk 1988
	DDT	563-1750		
<i>Rana mucosa</i>	DDE	17-100	Whole	Fellers et al 2004
	α- HCH	<DL-4.9		
	γ- HCH	<DL-0.7		
<i>Necurus maculosus</i>	DDE	0.3-90.0	Whole	Bonin et al 1995
	DDT	<DL-8.3		
	ΣHCH	<DL-10.1		
<i>Necturus lewisi</i>	DDE	60	Whole	Hall et al 1985
<i>Chaunus arenarum</i>	DDE	ND-4.5	Whole	Jofre et al 2008
	α- HCH	ND-5.6		
	β- HCH	ND-2.3		
	γ- HCH	ND-2.7		
<i>Hypsiboas cordobae</i>	DDE	1.1-1.7	Whole	
	α- HCH	3.9-5.4		
	β- HCH	3.0-7.3		
	γ- HCH	4.9-7.2		
<i>Leptodactylus mystacinus</i>	DDE	ND-6.0	Whole	
	α- HCH	3.5-6.9		
	β- HCH	0-8.9		
	γ- HCH	ND-5.7		
<i>Melanophryniscus stelzneri</i>	DDE	10.6	Whole	
	α- HCH	6.99		
	β- HCH	ND		
	γ- HCH	ND		
<i>Odontophrynus occidentalis</i>	DDE	0.8-1.8	Whole	
	α- HCH	1.3-4.1		
	β- HCH	1.0-2.7		
	γ- HCH	1.5-3		
<i>Pleurodema tucumanum</i>	α- HCH	4.7	Whole	
	β- HCH	4.1		
	γ- HCH	4.4		

DL-Detection limit, ND-Not detected

Table 5. Persistent Organic Pollutants concentrations (mg/Kg wet weight) measured in various amphibian species from different studies.

Mirex and HCB were pollutants found in less proportion in giant toads’ tissues. There are a few studies related to Mirex and HCB exposure on adult amphibians. Russell y collaborators (2002) reported average concentrations of 0.26-1 ng/g tissue of HCB in cricket frogs (*Acris crepitans*) coming from 5 agricultural locations in Ohio USA; in table (Table 5) other studies are presented with levels of HCB which are higher when compared to the ones obtained in this study (0.14-0.67 ng/g of tissue). Detectable levels of Mirex and HCB in turtles and crocodiles were not found.

PCBs congeners detected in blood and tissue sample were 52,101, 105, 118, 138, 153, 156, 170 and 180, which corresponds with the reported in other studies as the most common for human and biological samples (Table 4). The PCBs congeners presence pattern observed in the giant toad’s tissue is consistent with other studies on amphibians from other regions around the world (Loveridge et al., 2007, Russell et al., 1997); as well as in the found concentrations (Table 6). Studies conducted in diverse species of fresh water and terrestrial turtles have reported levels of PCBs totals of 5-414.8 ng/g wet mass (Basile, 2010, Moss et al., 2009, De Solla et al., 1998) and for alligator populations there have been found levels of 1.54 ± 0.12 ng/ml (Guillette el al 1999); these levels are comparable with the ones obtained in this study. Detected congeners (except 52) correspond in greater proportion (>30%) to aroclor 1254, one of the most sold commercial mixtures in the world, which suggests that the origin of these compounds probably is related to the use of these oils in the region’s industrial areas. Detected congeners are characterized for being some of the most persistent ones in the environment and for being absorbed in greater proportion in the organisms.

SPECIE	TISSUE	CONCENTRATION	REFERENCE
<i>Rana clamitans</i>	Carcass	2.8	Loveridge et al 2007
Various anurans	Carcass	151-4470	DeGarady y Halbrook 2003
<i>Rana clamitans</i>	-	7.51	Rusell et al 1997
<i>Necturus maculosus</i>	Carcass	113-1082	Bonin et al 1995
<i>Rana pipiens</i> and <i>Rana clamitans</i>	Carcass	50-112	Phaneuf et al 1995
<i>Rana perezi</i>	Muscle	50-1080	Rico et al 1987

Table 6. Total PCB concentrations (ng/g wet weight) measured in various amphibian species from different studies.

Amphibian and reptiles populations are declining at an alarming way in the world (Alford, 2020; Todd et al., 2010); some of the determining concomitant causes in this phenomenon is the exposure to toxic agents; let us note that reptile and amphibian toxicological information is growing, however, it is yet limited in comparison with other vertebrate groups (Sparling et al., 2010). Amphibians and reptiles may be exposed to a wide spectrum of toxic substances; pollutant accumulation in these organisms may be influenced by many factors (physiological, trophic, behavioral, etc.) at the same time, exposure may occur by different routes and in different environments during their life-time. It is known that these organisms can accumulate significant pollutant loads in their tissues, mainly of heavy metals and organic compounds. The caused effects by exposure to POPs of greater concern in reptile and amphibian populations are the endocrine disruption, DNA damage, and development abnormalities; some of the studies of greater impact over these effects have been found in these organisms. Ecological importance to maintain viable reptile and amphibian populations is determinant because these organisms are the link between terrestrial and aquatic ecosystems; at the same time they are mainly placed between intermediate links of

trophic chains; they have also diversified and occupied a wide spectrum of ecological niches in different types of ecosystems; the presence of highly persistent, bioaccumulative and biomagnifying pollutants is a potentially dangerous situation worth to be evaluated in the Coatzacoalcos Veracruz ecosystems.

3.4 Trophic levels and bioaccumulation of POPs

In Figure 2, the integration of exposure to POPs in tissue (ng/g lip) of the evaluated species in each one of the ecosystems present in Coatzacoalcos; a trophic hypothetical web was established in the region. Results were classified according to the trophic level that species belong to. It can be observed that herbivores are the ones presenting the lowest POPs concentrations followed by carnivorous, omnivorous, and finally the ones with the highest concentrations are the omnivorous. This is explained because detritivores organisms are found in grater contact with contaminated matrix (as soil and sediment), while omnivorous organisms include a greater number of exposure routes (environment and food) and are, therefore, the ones presenting greater POP's concentrations in their tissues.

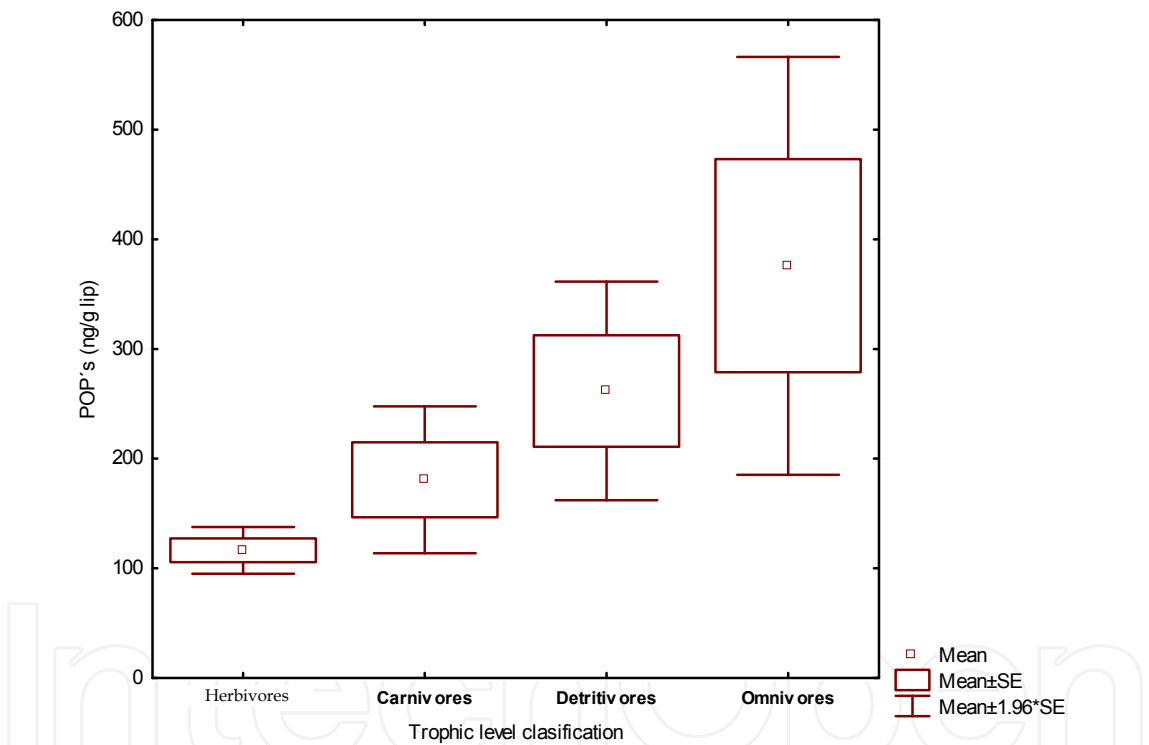


Fig. 2. Concentration Levels of POPs* in tissue taking into account the feeding habits of some species collected at Coatzacoalcos, Ver. *Corresponds to the sum of chloride compounds. Herbivores (iguana); Carnivores (Fish-Cep-); Detritivores (Fish -Arfe, Muce-Crab -Ucsp- and-Casp-); Omnivores (Fish -Tilapia-, Amphibian-Rhma-)

4. Conclusion

With the noted exception of HCB which was not found in the terrestrial ecosystem, traces of all other POPs were identified in species from all three ecosystems. Performing an analysis of feeding habits, it was learned that concentrations of POPs rise as follows: herbivores < carnivores < detritivores < omnivores. With these results it may be established that the

region's biota is in fact exposed to diverse Persistent Organic Pollutants. In similar studies it has been found that exposure to POPs may cause several effects (Espinosa-Reyes et al., 2010; González-Mille et al., 2010). In addition, one must consider that Coatzacoalcos organisms are exposed to other types of pollutants in addition to POPs whose collective action could well increase the magnitude of the effects. On the other hand, as far as brominated compounds go, this is the first ever study on fish in the region. The described scenario leads us to ask ourselves what exactly is the risk to humans and wildlife in the region. The Outlook is not promising if you consider that these organisms are of paramount importance in the food chain both to humans and other species of wildlife.

Available data on concentrations of chemical substances in the environment and human beings as well as over effects of exposure to chemicals complex combinations is still scarce. Chemicals generally pose a risk for the environment; they are a rapidly growing pollution load including chemical compounds increasingly complex from which potential effects on public health and environment are probably known. It is estimated that between 70,000 and 100,000 chemicals are available in the market and this number is growing fast. Around 5,000 of these substances are being produced in high volumes, over one million tons a year. The biggest chemical producers are countries members of the OECD, but countries such as India, China, Brazil, South Africa and Indonesia are rapidly increasing their productions. One of the main reasons for the development and adoption of the REACH Regulation is that a great number of substances have been produced and marketed in Europe during many years, at times in very large quantities yet there is not enough information over the hazards for human health and the environment. The European Commission has estimated that in order to fill in information gaps related to toxic effects of that large number of substances it may be required the use of 9 million of lab animals with an approximate cost of 1.3 billion € to perform the necessary tests. A later estimate suggested that required tests would imply 54 million vertebrate animals and costs would grow up to 9.5 billion €. Within this context, wildlife bio-monitoring may be used to detect chemical pollution and evaluate the health of ecosystems, with species as systematic test models during the evaluation of risks associated to actual exposure routes. Wildlife species residing in polluted zones are exposed to pollutant complex mixtures through multiple paths that could hardly be evaluated in laboratory tests.

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6. References

- Adams, S.M., Bevelhimer, M.S., Greeley, M.S., Levine, D.A., Teh, S.J. 1999. Ecological risk assessment in a large river reservoir: 6. Bioindicators of fish population health. *Environ. Toxicol. Chem.* 18:628–640.
- Alford, R.A, 2010. Declines and the global status of amphibians, D.W. Ecotoxicology of Organic Contaminants in amphibians. In Sparling, D.W. Linder, G., Bishop, C.A., Krest; S.K, editors. *Ecotoxicology of amphibians and reptiles* 2nd ed. Pensacola (FL): SETAC Press p 13-46.

- Álvarez del Toro, M. 1982. *Los reptiles de Chiapas*. Talleres Gráficos del Estado. Tuxtla Gutiérrez, Chiapas.
- ATSDR. 2005. *Toxicological profile for alpha-, beta-, gamma- and delta-hexachlorocyclohexane*. Department of Health and Human Services. Agency for Toxic Substances and Diseases Registry. 377.
- Basile, E.R., Avery, H.W., Bien, W.F., Keller, J.M. 2011. Diamondback terrapins as indicator species of persistent organic pollutants: Using Barnegat Bay, New Jersey as a case study. *Chemosphere* 82:137-44.
- Bayen, S.; Wurl, O.; Karuppiah, S.; Sivasothi, N.; Kee Lee, H.; Philip Obbard, J. 2005. Persistent organic pollutants in mangrove food webs in Singapore. *Chemosphere* 61: 303 -313.
- Bergeron, J.M., Crews, D., Mc Lachlan, J.A. 1994. PCBs as environmental estrogens: turtle sex determination as a biomarker of environmental contamination. *Environ. Health Perspect.* 102: 780-786.
- Bickham, J.W., Hanks, B.G., Smolen, M.J., Lamb, T., Gibbons, J.W. 1988 Flow cytometric analysis of the effects of low-level radiation exposure on natural populations of slider turtles (*Pseudemys scripta*). *Arch. Environ. Contam. Toxicol.* 17:837-841.
- Blake, A. 2005. *The next generation of POP's: PBDE's and lindane*. International POP's Elimination Network (IPEN). Washington D.C. USA. 15.
- Bonin, J., DesGranges, J.L., Bishop, C. A. Rodrigue, J., A. Gendron J. Elliott, E. 1995. Comparative study of contaminants in the mudpuppy (amphibia) and the common snapping turtle (reptilia), St. Lawrence River, Canada. *Arch. Environ. Contam. Toxicol.* 28:184-194.
- Burger, J., Gibbons, J.W. 1998 Trace Elements in Egg Contents and Egg Shells of Slider Turtles (*Trachemys scripta*) from the Savannah River Site. *Arch. Environ. Contam. Toxicol.* 34:382-386.
- CEC. 1997. *Historia del DDT en Norteamérica*. Commission for Environmental Cooperation. Available in:
http://www.cec.org/files/PDF/POLLUTANTS/historiaDDTs_ES.PDF (accessed Jul 2011)
- CEC. 2001. *Diagnostico situacional del uso de DDT y el control de la malaria*. Commission for Environmental Cooperation. Available in:
http://www.cec.org/files/PDF/POLLUTANTS/InfRegDDTb_ES_EN.pdf (accessed Jul 2011)
- CEC. 2006. *The North American regional action plan (NARAP) on lindane and other hexachlorocyclohexane (HCH) isomers*. Commission for Environmental Cooperation. 51p.
- De Solla, S. 2010. Organic Contaminants in reptiles. In Sparling, D.W. Linder, G., Bishop, C.A., Krest; S.K, editors. *Ecotoxicology of amphibians and reptiles*. 2nd ed. Pensacola (FL): SETAC Press p 289-324.
- De Solla, S.R., Bishop, C.A., Van der Kraak, G., Brooks, R.J., 1998. Impact of organochlorine contamination on levels of sex hormones and external morphology of common snapping turtles (*Chelydra serpentina serpentina*) in Ontario, Canada. *Environ. Health. Perspect.* 106:253-260.
- De Souza, A., Machado, J.P., Ornellas, R., Curcio, R., Souto, M., Silveira, C. 2008. Organochlorine pesticides (OCs) and polychlorinated biphenyls (PCBs) in sediments and crabs (*Chasmagnathus granulata*, Dana, 1851) from mangroves of Guanabara Bay, Río de Janeiro state, Brazil. *Chemosphere* 73(1): 186 - 192.

- DeGarady, C. J. and Halbrook, R. S. 2003. Impacts from PCB Accumulation on Amphibians Inhabiting Streams Flowing from the Paducah Gaseous Diffusion Plant. *Arch. Environ. Contam. Toxicol.* 45(4): 525-532.
- Di Giulio, R.T. and Hinton, D.E. 2008. The toxicology of fishes. Ed. CRC Press, Florida, 1071pp.
- Dohm, M.R., Mautz, W.J., Doratt, R.E., Stevens, J.R. 2008 Ozone exposure affects feeding and locomotor behavior of adult *Bufo marinus*. *Environ. Toxicol. Chem.* 27:1209-1216.
- Edwards P.J., Brown S.M. 1982. Use of grassland plots to study the effect pesticides on earthworms. *Pedobiol.* 24: 145-150.
- Espinosa-Reyes, G.; Ilizaliturri, C.; González-Mille, D.; Costilla, R.; Díaz-Barriga, F.; Cuevas, M.C.; Martínez, M.A.; Mejía-Saavedra, J. DNA Damage in earthworms (*Eisenia* spp.) as indicator of environmental Stress in the industrial zone Coatzacoalcos, Veracruz, Mexico. *J Environ. Science Health A* 45: 49-55.
- Falandysz, J.; Strandberg, L.; Zpuzyn, T.; Gucia, M. 2001. Chlorinated cyclodiene pesticide residues in Blue mussel, crab, and fish in the Gulf of Gdansk, Baltic Sea. *Environ. Sci. Technol.* 35: 4163 – 4169.
- FAO. 1997. Lista mundial de vigilancia para la diversidad de los animales domésticos. 2a Ed. Organización de las Naciones Unidas para la Agricultura y la Alimentación. Roma, Italia. <http://www.fao.org/docrep/V8300S/v8300s00.HTM> (última visita diciembre de 2008)
- Feder, M.E., Burggren, W.W. 1992. Environmental physiology of the amphibians. University of Chicago Press, Chicago, p 646.
- Fellers, G.M., McConnell, L.L., Pratt, D., Datta, S. 2004. Pesticides in mountain yellow-legged frogs (*Rana muscosa*) from the Sierra Nevada Mountains of California, USA. *Environ. Toxicol. Chem.* 23: 2170-2177.
- Fitzpatrick, L.C.; Sassani, R.; Venables, B.J.; Goven, A.J. 1992. Comparative toxicity of polychlorinated biphenyls to earthworms *Eisenia fetida* and *Lumbricus terrestris*. *Environ. Pollut.* 77: 65-69.
- Gibbons, J.W. (ed). 1990. The slider turtle. In: *Life history and ecology of the slider turtle*. Smithsonian Institution Press, Washington, DC, pp 3-18
- Gilliland, C.D., Summer, C.L., Gilliland, M.C., Kannan, K., Villeneuve, D.L., Coady, K., Muzzall, P., Mehne, C., Giesy, J.P. 2001 Organochlorine insecticides, polychlorinated biphenyls and metals in water, sediment and green frogs from southwestern Michigan. *Chemosphere* 44:327-339.
- González-Mille, D.J.; Ilizaliturri-Hernández, C.A.; Espinosa-Reyes, G.; Costilla-Salazar, R.; Díaz-Barriga, F.; Ize-Lema, I. and Mejía-Saavedra, J. 2010. Exposure to persistent organic pollutants (POPs) and DNA damage as an indicator of environmental stress in fish of different feeding habits of Coatzacoalcos, Veracruz, Mexico. *Ecotoxicology* 19:1238-1248.
- Goven, A.J.; Eyambe, G.S.; Fitzpatrick, L.C.; Venables, B.J.; Cooper, E.L. 1993. Cellular biomarkers for measuring toxicity of xenobiotics: effects of polychlorinated biphenyls on earthworm *Lumbricus terrestris* coelomocytes. *Environ. Toxicol. Chem.* 12: 863-870.
- Guillette, L.J., Brock, J.W., Rooney, A.A., Woodward, A.R. 1999. Serum concentrations of various environmental contaminants and their relationship to sex steroid concentrations and phallus size in juvenile American alligators *Arch. Environ. Contam. Toxicol.* 36:447-455.

- Hall, R.J, Driscoll, C.T., Likens, G.E., Pratt, J.M.1985. Physical, chemical and biological consequences of episodic aluminium addition to a stream. *Limnol. Oceanogr.* 30, 212-220.
- Haque, A. Ebing, W. 1983. Toxicity determination of pesticides to earthworms in the soil substrate. *J. Plant Dis. Prot.* 90: 395-408.
- Harris, M.L., Bishop, C.A., Struger, J., Van Den Heuvel, M.R., Van Den Kraak, M.R., Dixon, G.J., Ripley, B., Bogart, J.P. 1998. The functional integrity of northern leopard frog (*Rana pipiens*) populations in orchard wetlands. I. Genetics, physiology, and biochemistry of breeding adults and young-of-the-year. *Environ. Toxicol. Chem.* 17:1338-1350.
- Harris, M.L.; Wilson, L.K.; Elliott, J.E.; Bishop, C.A.; Tomlin, A.D.; Henning, K.V. 2000. Transfer of DDT and Metabolites from Fruit Orchard Soils to American Robins (*Turdus migratorius*) Twenty Years after Agricultural Use of DDT in Canada. *Arch. Environ. Contam. Toxicol.* 39, 205-220.
- Heimbach, F. 1984. Correlations between three methods for determining the toxicity of chemicals to earthworms. *Pest. Sci.* 15: 605-611
- Jha, A.N. 2008. Ecotoxicological applications and significance of the comet assay. *Mutagenesis* 23:207-221.
- Jofré, M.B., Antón, I.R. and Caviedes-Vidal, E. 2008. Organochlorine Contamination in Anuran Amphibians of an Artificial Lake in the Semiarid Midwest of Argentina. *Arch. Environ. Contam. Toxicol.* 55 (3): 471-480.
- Jones, A. and Hart, D.M. 1998. Comparison of laboratory toxicity tests for pesticides with field effects on earthworm population: a review. Pp: 247-267. In: Sheppard, S.C; Bembridge J.D.; Holmstrup, M.; Posthuma, L. 1998. *Advances in earthworm ecotoxicology*. Proceedings from the second international Workshop on earthworm ecotoxicology. 2 - 5 april 1997. Amsterdam, The Netherlands. Pensacola FL: Society of environmental Toxicology and chemistry (SETAC) 472 p.
- Kirk, J.J. 1988. Western spotted frog (*Rana pretiosa*) mortality following forest spraying of DDT. *Herp. Review* 19:51-53.
- Klobučar, G. I. V., Štambuk, A., Pavlica, M., Sertić, P.M. Kutuzović, H., Hylland. 2010. Genotoxicity monitoring of freshwater environments using caged carp (*Cyprinus carpio*). *Ecotoxicology* 19:77-84.
- Kula, H. 1995. Comparison of laboratory and field testing for the assessment of pesticide side effects on earthworms. *Acta Zool. Fennica*, 196: 338-341.
- Külköylüolu, O. 2004. On the usage of ostracods (Crustacea) as bioindicator species in different aquatic habitats in the Bolu region, Turkey. *Ecological Indicators* 4: 139-147
- Langdon, C.J., Pearce, T.G., Meharg, A.A., Semple, K.T. 2003. Interactions between earthworms and arsenic in the soil environment: a review. *Environ. Pollut.* 124, 361-373.
- Lara-López, M.S., González-Romero, A. 2002. Alimentación de la iguana verde *Iguana iguana* (Squamata: Iguanidae) en la Mancha, Veracruz, México. *Acta Zool Mex* 85: 139-152.
- Lee R.F., Steinert S. 2003. Use of the single cell gel electrophoresis/comet assay for detecting DNA damage in aquatic (marine and freshwater) animals. *Mutation Res.* 544:43-64.
- Legall, J.R.; L.E., Dicovski, Valenzuela, Z.I. 2006. *Manual Básico de lombricultura para condiciones tropicales*. Escuela de Agricultura y Ganadería de Estelí. Estela, Nicaragua. 16 p.

- León, P., Montiel, S. 2008. Wild Meat Use and Traditional Hunting Practices in a Rural Mayan Community of the Yucatan Peninsula, Mexico. *Hum. Ecol.* 36: 249–257.
- Linzey, D., Burroughs, J., Hudson, L., Marini, M., Robertson, J., Bacon, J., Nagarkatti, M., Nagarkatti, P. 2003. Role of environmental pollutants on immune functions, parasitic infections and limb malformations in marine toads and whistling frogs from Bermuda. *Int. J. Environ. Health Res.* 13:125-148.
- Lutharddt, P.; Mayer, J.; Fuchs, J. 2002. Total TEQ emissions (PCDD/F and PCB) from industrial sources. *Chemosphere.* 46, 1303-1308.
- Ma, W., Bodt, J. 1993. Differences in toxicity of the insecticide chlorpyrifos to six species of earthworms (oligochaeta, lumbricidae) in standardized soils tests. *Bull. Environ. Cont. Toxicol.* 50: 864-870.
- McCoy, K.A., Bortnick, L.J., Campbell, C.M., Hamlin, H.J., Guillette, L.J., St Mary, C.M. 2008. Agriculture alters gonadal form and function in the toad *Bufo marinus*. *Environ. Health Perspect.* 116:1526-1532.
- Morrison, D.E.; Robertson, B.K.; Alexander, M. 2000. Bioavailability to earthworms of aged DDT, DDE, DDD, and Dieldrin in soil. *Environ. Sci. Technol.* 34: 709 – 713.
- Moss, S., Keller, J.M., Richards, S., Wilson, T.P. 2009. Concentrations of persistent organic pollutants in plasma from two species of turtle from the Tennessee River Gorge. *Chemosphere* 76:194-204.
- Nacci, D.E., Cayulab S., Jackim E. 1996. Detection of DNA damage in individual cells from marine organisms using the single cell gel assay. *Aquat. Toxicol.* 35:197-210.
- Newhook, R., Meek, M.E. 1994. Hexachlorobenzene: evaluation of risks to health from environmental exposure in Canada. *Environ. Carcin. Ecotox. Rev.* 12(2), 345-360.
- Ogunseitan, O.A. 2002. *Microbial proteins as biomarkers of ecosystem health.* 217-232 pp. In: Integrated Assessment of Ecosystem Health. Edited por: Scow, K. M.; Fogg, G.E.; Hinton, D.E.; Jonson, M.L. Lewis Publishers. Boca Raton, Florida, U.S.A. 340 p.
- Overman, S.R., Krajicek, J.J. 1995. Snapping turtles (*Chelydra serpentina*) as biomonitors of lead contamination of the Big River in Missouri old lead belt. *Environ. Toxicol. Chem.* 14:689–695.
- Pastor, D., Sanpera, C., González-Solís, J., Ruiz, X., Albaigés, J. 2004. Factors affecting the organochlorine pollutant load in biota of a rice field ecosystem (Ebro Delta, NE Spain). *Chemosphere* 55:567–576.
- Petrlink, J., J. DiGangi. 2005. *The egg report.* International POPs Elimination Network (IPEN). Washington D.C. USA. 52.
- Phaneuf, D., DesGranges, J.L., Plante, N., Rodrigue, J. 1995 Contamination of local wildlife following a fire at a polychlorinated biphenyls warehouse in St. Basile le Grand, Quebec, Canada. *Arch Environ Contam Toxicol* 28:145–153.
- Portter, D.A., Spicer, P.G., Redmond, C.T., Powel, A.L. 1994. Toxicity of pesticides to earthworms in Kentucky bluegrass turf. *Bull. Environ. Cont. Toxicol.* 52: 176-181.
- Regoli, F., Gorbi, S., Fattorini D., Tedesco S., Notti A., et al.,. 2006 Use of the land snail *Helix aspersa* as sentinel organism for monitoring ecotoxicologic effects of urban pollution: an integrated approach. *Environ. Health Perspect.* 114:63-69.
- Reinecke, A.J. and Reinecke S.A. 1998. *The use of earthworms in ecotoxicological evaluation and risk assessment: new approaches* In: Earthworm ecology. Edwards, C.A. (Ed.) St. Lucie Press. Boca Raton. U.S. 273-293

- Reines, M., Rodríguez, C., Sierra, A., Vázquez, M. 1998. *Lombrices de tierra con valor comercial: Biología y técnicas de cultivo*. Universidad de Quintana Roo. Chetumal, Quintana Roo, México. 60 p.
- Rico, M.C., Hernandez, L.M., Gonzalez, M.J., Fernandez, M.A., Montero, M.C. 1987. Organochlorine and metal pollution in aquatic organisms sampled in the Doñana National Park during the period 1983–1986. *Bull. Environ. Contam. Toxicol.* 39:1076–1083.
- Rigonato, J., Mantovani, M.S., Quinzani, J.B. 2005. Comet assay comparasion if different *Corbicula fluminea* (Mollusca) tissues for detection of genotoxicity. *Genet. Mol. Biol.* 28: 464-468.
- Rinderhagen, M., Ritterhoff, J. & Zauke, G.P. 2000. Crustaceans as bioindicators. In *Biomonitoring of polluted water: reviews on actual topics* (A. Gerhardt, ed.). Trans Tech Publications; Environmental Research Forum, Uetikon, p. 161-194.
- Riojas-Rodriguez, H., Baltazar-Reyes, M.C., Meneses, F. 2008. Volatile organic compound presence in environmental samples near a petrochemical complex in Mexico. *Abstracts Epidemiology* 19 (1), S219.
- Rosales, L. Carranza, E. Estudio geoquímico de metales en el estuario del río Coatzacoalcos. In: *Golfo de México contaminación e impacto ambiental: diagnóstico y tendencias*. Vázquez-Botello, A.; Rendón-Von Osten, J.; Gold-Bouchot, G., Agraz-Hernández, C., Eds.; Universidad Autónoma de Campeche, Universidad Autónoma de México, Instituto de Ecología. 2005, 389-406.
- Russell, R., Lipps, G., Hecnar, S., Haffner, D. 2002. Persistent Organic Pollutants in Blanchard's Cricket Frogs (*Acris crepitans blanchardi*) from Ohio. *Ohio J Sci.* 102 (5):119-122.
- Russell, R.W., Gillan, K.A., Haffner, G.D. 1997 Polychlorinated biphenyls and chlorinated pesticides in southern Ontario, Canada, green frogs. *Environ. Toxicol. Chem.* 1:2258–2263.
- Russell, R.W., Hecnar, S.J., Haffner, G.D. 1995. Organochlorine pesticide residues in southern Ontario spring peppers. *Environ. Toxicol. Chem.* 14:815–817.
- Russo, C., Rocco, L., Morescalchi, M.A., Stingo, V. 2004. Assessment of environmental stress by the micronucleus test and the comet assay on the genome of teleost populations from two natural environments. *Ecotoxicol. Environm. Saf.* 57:168-174.
- Sala, M.; Sunyer, O.; Otero, R.; Santiago-Silva, M.; Ozalla, D.; Herrero, C.; To-Figueras, J.; Kogevinas, M.; Anto, J.; Camps, C.; Grimalt, J. 1999. Health effects of chronic high exposure to hexachlorobenzene in a general population sample. *Arch. Environ. Health.* 54(2), 102-109.
- Sánchez-Hernández, J.C. 2006. Earthworm biomarkers in ecological risk assessment. *Rev. Environ. Contam. Toxicol.* 188: 85-126
- Selcer, K.W. 2006. Reptile ecotoxicology: studying the effects of contaminants on populations . In Gardner S.C, Oberdörster, E., editors. *Toxicology of reptiles*. Boca raton (FL): Taylor and Francis Press p 267-297.
- Sparling, D.W. Linder, G., Bishop, C.A., Krest; S.K. 2010. Recent advancements in amphibian and reptile ecotoxicology. In Sparling, D.W. Linder, G., Bishop, C.A., Krest; S.K, editors. *Ecotoxicology of amphibians and reptiles* 2nd ed. Pensacola (FL): SETAC Press p 1-12.

- Stringer, R.; Labunska, I.; Bridgen, K. 2001. *Organochlorine and heavy metals contaminants in the environment around the Complejo Petroquímicos Paharitos, Coatzacoalcas, México*. Technique note Greenpeace. University of Exeter. U.K. 60.
- Takazawa, Y., Kitamura, K., Yoshikane, M., Shibata, Y., Morita, M., Tanaka, A. 2005. Distribution patterns of hexachlorocyclohexanes and other organochlorine compounds in muscles of fish from a Japanese remote lake during 2002–2003. *Bull. Environ. Contam. Toxicol.* 74:652–659.
- Thompson AR. 1970. Effects of nine insecticides on numbers and biomass of earthworms in pasture. *Bull. Environ. Cont. Toxicol.* 5: 577-585.
- Vázquez-Botello, A.; Villanueva-Fragoso, S.; Rosales-Hoz, L. 2004. *Distribución y contaminación por metales en el Golfo de México*. In: Diagnostico ambiental del Golfo de México. Caso, M. Pisanty, I.; Ezcurra, E., Eds.; SEMARNAT-INE. 682-712.
- Venne, L., Anderson, T., Zhang, B., Smith, L., McMurry, S. 2008 Organochlorine pesticide concentrations in sediment and amphibian tissue in playa wetlands in the southern high plains, USA. *Bull. Environ. Toxicol.* 80: 497-501.
- Willett K.L., Ulrich E.M., Hites R.A. 1998. Differential toxicity and environmental fates of hexachlorocyclohexane isomers. *Environ. Sci. Technol.* 32:2197-2207.
- Willett, K.L., Ulrich, E.M., Hites, R.A. 1998 Differential toxicity and environmental fates of hexachlorocyclohexane isomers. *Environ. Sci. Technol.* 32:2197-2207.
- Willingham, E.J., Crews, D. 1999. Organismal effects of environmentally relevant pesticide concentrations on the red-eared slider turtle. *Gen. Comp. Endocrinol.* 113:429–435.
- Willingham, E.J., Crews, D. 2000. The red-eared slider turtle: an animal model for the study of low doses and mixtures. *American Zool.* 40:421–428.
- Yim, U.H., Hong, S.H., Shim, W.J., Oh, J.R. 2005 Levels of persistent organochlorine contaminants in fish from Korea and their potential health risk. *Arch. Environ. Contam. Toxicol.* 48:358–366.
- Zug, G.R., Zug, P.B. 1979. The marine toad, *Bufo marinus*: a natural history resumé of native populations. Smithsonian Institution Press, Washington, D.C. 284 pp. Available in: <http://hdl.handle.net/10088/5188>
- Zupanovic, Z., Musso, C., Lopez, G., Louriero, C.L., Hyatt, A.D., Hengstberger, S., Robinson, A.J. 1998. Isolation and characterization of iridoviruses from the giant toad *Bufo marinus* in Venezuela. *Dis. Aquat. Org.* 33:1-9.



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Ten years after coming into force of the Stockholm Convention on Persistent Organic Pollutants (POPs), a wide range of organic chemicals (industrial formulations, plant protection products, pharmaceuticals and personal care products, etc.) still poses the highest priority environmental hazard. The broadening of knowledge of organic pollutants (OPs) environmental fate and effects, as well as the decontamination techniques, is accompanied by an increase in significance of certain pollution sources (e.g. sewage sludge and dredged sediments application, textile industry), associated with a potential generation of new dangers for humans and natural ecosystems. The present book addresses these aspects, especially in the light of Organic Pollutants risk assessment as well as the practical application of novel analytical methods and techniques for removing OPs from the environment. Providing analytical and environmental update, this contribution can be particularly valuable for engineers and environmental scientists.

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