

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Oncogenic Pathway Signatures and Survival Outcome

Xuan Bich Trinh^{1,2}, Peter A. Van Dam¹, Luc Y. Dirix¹,
Steven J. van Laere¹ and Wiebren A. A. Tjalma²

¹*Translational Cancer Research Unit, St Augustinus GZA Hospitals, Antwerp*

²*Department of Gynaecological Oncology, Antwerp University Hospital, Antwerp
Belgium*

1. Introduction

Recent microarray technology and bioinformatics have shown the ability of analysing oncogenic cellular signalling pathways based upon gene signatures in cancers. (Bild et al., 2006; Dressman et al., 2007; Gatz et al., 2010) Epithelial ovarian cancer (EOC) is the most important cause of mortality among gynaecological cancers. Patients with EOC often present in an advanced stage. Treatment modalities consist in general of the sequence of surgical cytoreduction and platinum-taxane based chemotherapy. (Cannistra, 2004) Although the disease is relatively sensitive to cytotoxics, relapses occur in a majority of patients with advanced stage. (Cannistra, 2004) The emergence of resistance to conventional chemotherapeutics is an often-deadly event in the management of ovarian cancer patients. There is an urgent need for additional therapies that increase survival and/or quality of life in these patients.

The objective of our study was to look for cellular pathways that have an effect on survival outcome by a bioinformatical approach. (Trinh et al., 2011) These pathways may guide us to find interesting targets in ovarian cancer. Survival can be used as a measure to quantify the biological relevance in this disease. Ideally, evaluation of survival outcome should be made in a homogenous population with a uniform treatment to avoid treatment-induced biases and uniform histology to find subtler differences independent from histology. Another methodology of estimating prognostic value may be the correlation with documented prognostic gene signatures that have shown to be of prognostic value in breast cancer and other types of cancer. The invasiveness gene signature (IGS) was generated using stem cell-like or tumorigenic breast cancer cells. (Liu et al., 2007) This signature has shown prognostic value in lung cancer, medulloblastoma and prostate cancer. The Wound healing response (WHR) signature, based upon genes induced by wound healing, also has shown its prognostic value in breast cancer, NSLC and bladder cancer. (Chang et al., 2005; Lauss, Ringnér, & Höglund, 2010; Mostertz et al., 2010) The genomic grade index (GGI) is a signature that divides low-grade versus high-grade breast carcinomas. (Sotiriou et al., 2006) Interestingly, using this signature, histological intermediate-grade tumours could be classified as low- or high-grade tumours with the preservation of the gene signatures' prognostic value.

2. Oncogenic pathways

The oncogenic gene signatures were derived from a recent paper by Gatza and colleagues and applied similarly. (Gatza et al., 2010) These pathway signatures were mainly generated by activating or silencing specific genes in cell lines experiments. The signatures were robustly validated afterwards. For each pathway, a pathway activation score was calculated based upon the gene signature to quantify the activation by a score.

Briefly, for each array-sample the pathway-specific informative genes were identified. Next a pathway score was calculated by adding up the products of the gene expression for each gene and its corresponding regression coefficient, which indicates the weight (amplitude of regression coefficient) and the effect (sign of regression coefficient) of the corresponding gene for activation of the corresponding pathway. Finally, the pathway scores were scaled using the intercept values provided in the original manuscript and standardized for comparability by median-centering and setting the standard deviation to 1. Pathways included in the analysis were AKT, β -Catenin, E2F1, EGFR, ER, HER2, INF α , INF γ , MYC, p53, p63, PI3K, PR, RAS, SRC, STAT3, TNF α , and TGF β .

Since PARP inhibitors and VEGF-A inhibitors have shown promising results in ovarian cancer, the BRCA pathway and VEGF-A pathway was also studied. (Audeh et al., 2010; Burger et al., 2010; Fong et al., 2009; 2010) For the BRCA signature, we used one that was published by Konstantinopoulos and colleagues. (Konstantinopoulos et al., 2010) For a VEGF-A signature we have used and validated genes that were reported by Hu et al. (Hu et al., 2009) A BRCA activation score was applied using the same methodology with 60 genes, their weight and sign. (Konstantinopoulos et al., 2010) Prognostic gene signatures (IGS, GGI and WHR) were also applied by previously described methodology. (Chang et al., 2005; Liu et al., 2007; Sotiriou et al., 2006) All gene signature activation scores were handled as a continuous variable. The same standardisation (Median=0; SD=1) was applied for each gene signature.

For the VEGF-A activation signature we used the 13 genes reported by Hu and colleagues. (Hu et al., 2009) To validate and transform this gene signature into a VEGF-A activation probability score we performed subsequent analysis using publicly available gene expression data sets on naïve and VEGF-A treated HUVEC cell lines (GSE18913 (N=21), GSE10778 (N=9; only the HGU133A samples were used) and GSE15464 (N=4)). Each data set was normalised using the GC-RMA algorithm and informative genes (above log 2(100) in at least 25% of the genes) were filtered in. First, we applied a principal component analysis on the GSE18913 data set using the informative VEGF-A signatures genes only (N=10). Only 10 out of 13 genes (*FABP5*, *UCHL1*, *PLOD*, *DDIT4*, *VEGF*, *ADM*, *ANGPTL4*, *NDRG1*, *NP* and *SLC16A3*) were reliably measured (high signal-to-noise ratio). Using these 10 genes in a principal component analysis (PCA) we were able to demonstrate a significant segregation of VEGF-A treated and naïve HUVEC's along the first principal component. Class label permutation analysis revealed that the observed Euclidean distance between the centroids of the VEGF-A treated and naïve HUVEC's on the 2D scatterplot representation of the PCA was significantly different from the expected Euclidean distance (Figure 1A; Observed Euclidean distance=2.185, Expected Euclidean distance=0.682, $P<0.0001$).

Next, we transformed the VEGF-A signature into a VEGF-A activation probability score adopting the methodology described by Gatza and his colleagues. (Gatza et al., 2010). Therefore, we used the regression coefficients that define the first principal component and multiplied these with the gene expression values of their corresponding genes. The products were summed and the resulting score was compared between VEGF-A treated and naïve HUVEC's using a Mann-Whitney U-test (Median VEGF-A treated HUVEC's: 6.416, Median

naïve HUVEC's: 4.276, $P<0.0001$). The boxplot representation is provided in Figure 1B. In addition, we observed a strong correlation between the VEGF-A activation probability scores and the time of VEGF-A incubation of HUVEC's (Correlation coefficient = 0.762; $P=0.038$). (Figure 1C). To validate our procedure, we applied our algorithm on the samples in gene expression data sets GSE10778 and GSE15464.

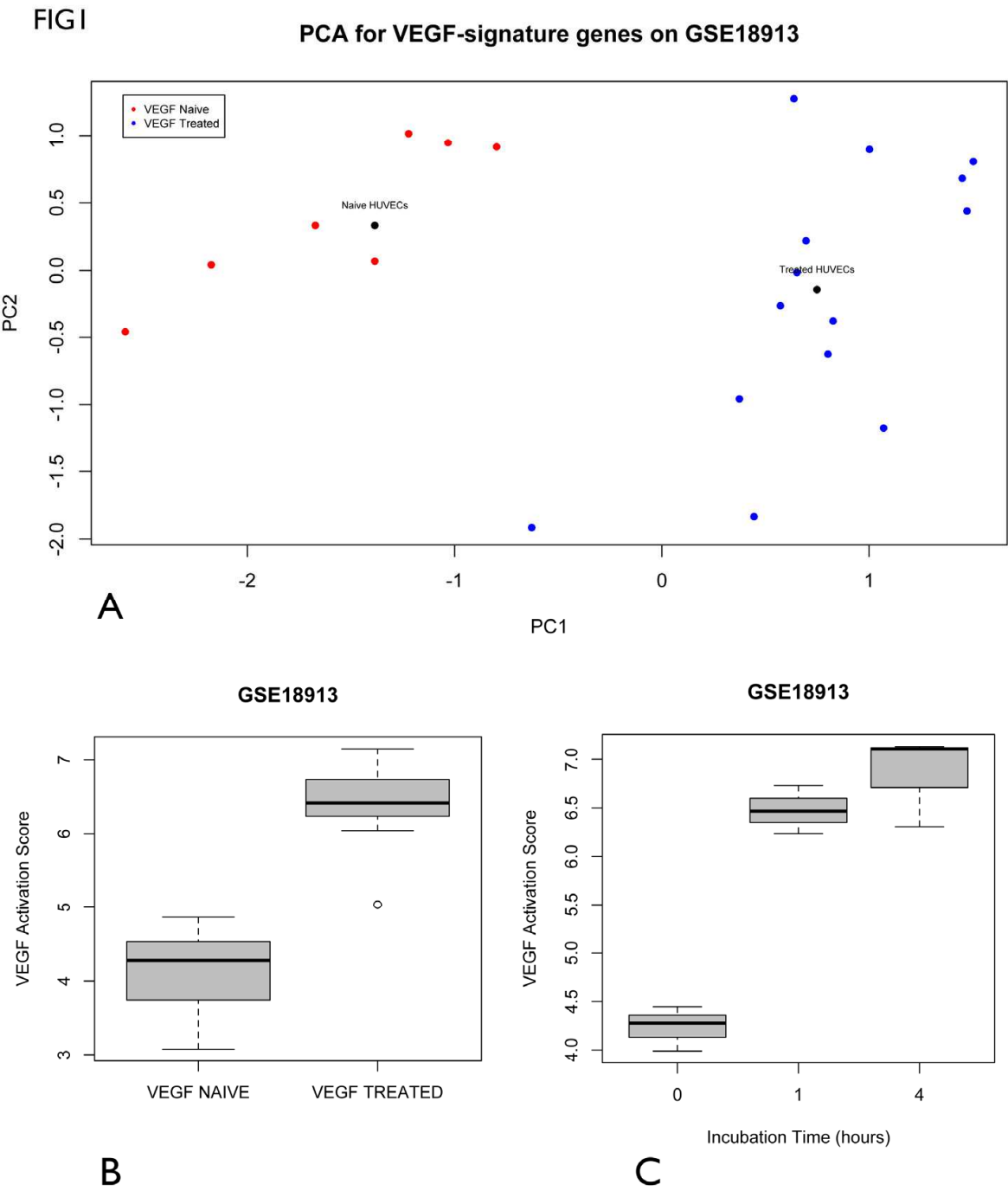


Fig. 1. Principal Component Analysis Plot (A) segregates VEGF-A treated cells versus untreated cells. The calculated activation scores were higher in treated cells versus untreated cells in an apparent time dependent way. (B+C)

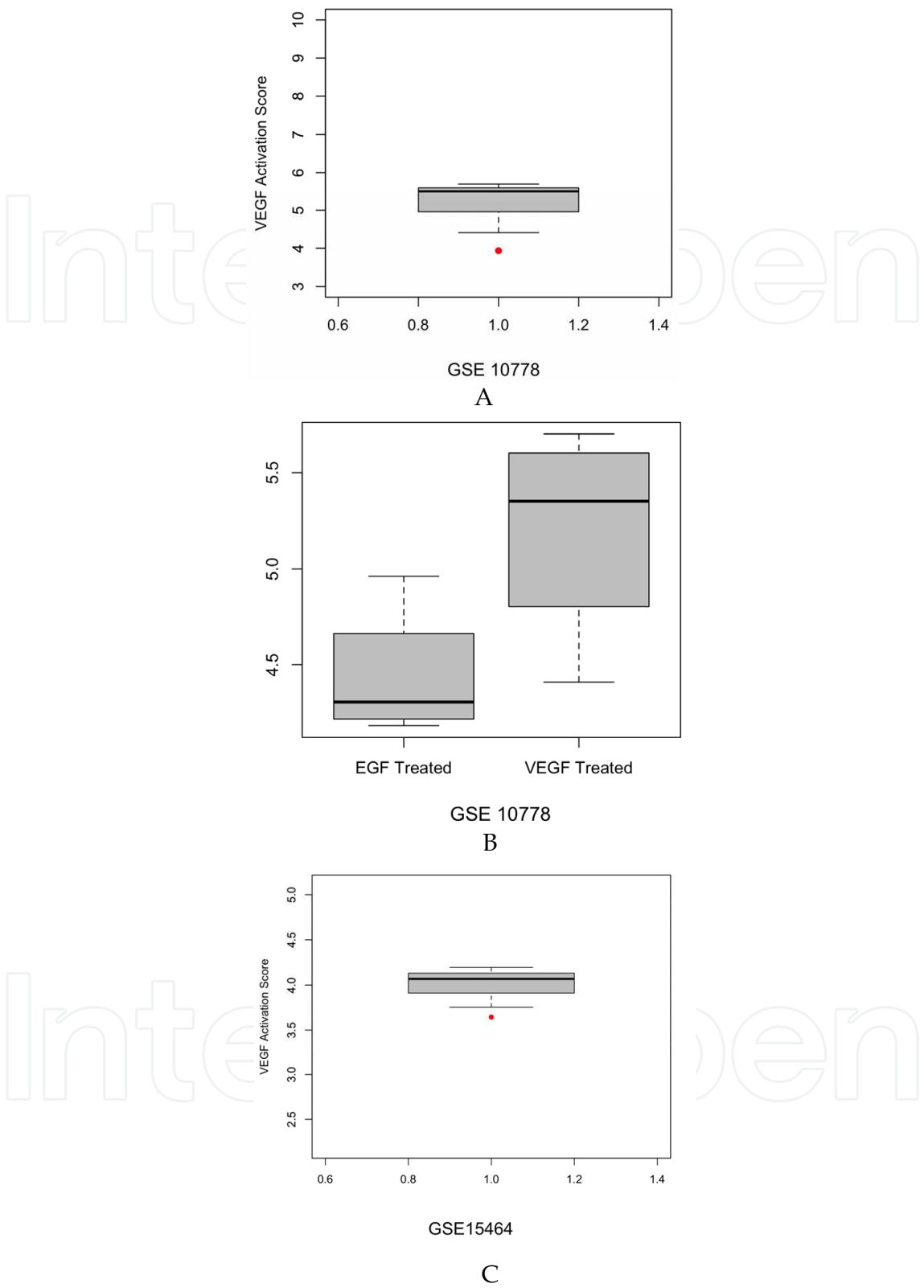


Fig. 2. Validation of the VEGF-A activation score methodology in data derived from two other experiments. The activation scores of VEGF-A treated cells were higher than the untreated condition (red dot). The higher activation scores were observed for VEGF-A treated cells but not for EGF treated cells, suggesting he specificity of the activation score for VEGF-A.

3. Patient datasets

A dataset of 285 patients (Melbourne dataset) was obtained through the Gene Expression Omnibus GEO database (GSE 9891) together with the clinical annotation data file. Only patients that had carcinomas of serous histology in advanced stages (III/IV) were included for analysis. Patients were selected that received platinum and taxane based chemotherapy. Other patients who did not receive chemotherapy or received only one agent, platinum or taxane, were also excluded. After this selection N=165 patients were eligible for further analysis. This dataset contained gene expression data derived from the Affymetrix U133_plus2 platform, which already underwent normalisation using the Robust Multiarray Averaging (RMA) method and subsequent filtering by excluding log expression values of <7 and a variance of <0.5 . After filtering there were 8,732 probe sets left that are considered informative. Progression free survival was used in further analysis. (Tothill et al., 2008)

A second dataset GSE3149 N=153 (North Carolina dataset) with clinical data was also obtained from the GEO website. Here, the same criteria for patient selection were used. After selection N=107 were further analysed. The North Carolina dataset used the same Affymetrix U133_plus2 platform. The raw data were processed in Bioconductor in R software packages. Filtering was done by selecting expressions below a threshold (\log_2 of 100) that are present in at least 25% of the arrayed samples. Normalisation was done using GC-Robust Multiarray Averaging. The number of probe sets that were informative was 7,741. Overall survival data was used, as there was no progression free survival data available. (Bild et al., 2006) A third dataset (Québec dataset) were patients (N=20) that were selected to be either chemoresistant versus chemosensitive. Here, raw microarray data based upon the Agilent platform Human 1A (v2) oligonucleotide microarray were normalised using the Lowess normalisation method. Hereafter, 16,096 genes were eligible for further analysis. Progression free survival data were used. RAW gene expression data is publicly available according to MIAME guidelines through the GEO database (Accession number: GSE 28739). (Bachvarov et al., 2006) A fourth dataset (Niigata Dataset-GSE 17260) contained samples that originated from patients who met the inclusion criteria from present study. Progression free survival data were available. The authors used the Agilent Whole Human Genome Oligo Microarray platform and normalised the data using upper quartile normalisation. 28,446 genes were found to be informative. (Yoshihara et al., 2010) A fifth and sixth dataset (Boston dataset A +B - GSE19829) were derived from a report studying BRCAness in ovarian cancer. (Konstantinopoulos et al., 2010) Progression free survival data was used. After selection, (N=26) and (N=36) patients were eligible. These datasets were RMA-normalised. 35252 and 5626 probe set ID's were informative after filtering. Gene expression data was derived from two platforms: the Affymetrix U133_plus2 platform and the Affymetrix 95UAv2.

4. Correlation of pathway activation scores with prognostic signatures

We applied the oncogenic pathways on the six datasets. These datasets together represent a total of N=464 advanced serous papillary carcinomas. A summary of these 6 datasets is listed in Table 1. Since these are selected oncogenic pathways, it is plausible that many significant correlations were found between pathway activations and the 3 prognostic

| Dataset | N=464 | Platform | Norma- lisation | Clinical outcome | Uniform treat- ment | Advanced stage/ serous papillary histology |
|---------------------------|-------|--|--------------------|---------------------|---------------------------|---|
| Québec 2006 | 20 | Agilent Human 1A (v2) | Lowess | PFS | yes | yes |
| North Carolina 2006 | 107 | Affymetrix U133_plus2 | GC- RMA | OS | yes | yes |
| Melbourne 2008 | 165 | Affymetrix U133_plus2 | RMA | PFS | yes | yes |
| Niigata 2010 | 110 | Agilent Whole Human Genome Oligo Microarray | Upper quartile | PFS | yes | yes |
| Boston A 2010 | 26 | Affymetrix U133_plus2 | RMA | PFS | yes | yes |
| Boston B 2010 | 36 | Affymetrix U95_A2 | RMA | PFS | yes | yes |

Table 1. A summary of datasets that were used in the meta-analysis.

| Pearson Rho | WHR | IGS | GGI |
|---------------|------------|------------|------------|
| Québec | 0.65 | 0.62 | 0.67 |
| | p=3.4 E-4 | p=0.001 | p=2.0 E-4 |
| North Carol | 0.81 | 0.89 | 0.6 |
| | p=7.7 E-40 | p=9.9 E-59 | p=8.0 E-18 |
| Melbourne | 0.73 | 0.54 | 0.79 |
| | p=2.8 E-22 | p=6.9 E-11 | p=5.6 E-28 |
| Niigata | 0.77 | 0.73 | 0.79 |
| | p=2.4 E-19 | 1.0 E-22 | p=4.5 E-25 |
| Boston A | 0.83 | 0.48 | 0.87 |
| | p=1.2 E-7 | p=0.013 | p=5.5 E-9 |
| Boston B | 0.75 | 0.56 | 0.26 |
| | p=1.8 E-7 | p=3.7 E-4 | p=0.13 |
| Meta Analysis | 0.73 | 0.62 | 0.79 |
| | p<0.0001 | p<0.0001 | p<0.0001 |

Table 2. This shows the consistent correlations of the β -Catenin activation scores and WHR/IGS/GGI in each separate dataset (Québec, North Carolina, Melbourne, Niigata, Boston A and Boston B dataset). Overall Rho Coefficients were estimated by a meta-analysis approach using random models effects.

signatures (IGS, WHR and GGI). The β -Catenin pathway showed consistent and strong correlations. (Table 2) Since the six datasets were generated on different platforms with different methodologies, we estimated the overall effect of a pathway activation score by using a meta-analysis approach (Table 2). Similar meta-analysis of correlation coefficients showed that the BRCA, E2F1, EGFR, HER2, MYC, p53, p63 and PI3K showed steady correlations with the WHR, GGI and IGS. The RAS pathway and TGF β pathway showed significant correlations with 2/3 prognostic signatures. Table 3 shows the overall correlation estimates, which were the most significant. While most pathway activation scores showed a positive correlation, the EGFR, HER2, p53 and TGF β pathway showed a negative correlation.

| Rho estimates | WHR | IGS | GGI |
|------------------|----------|----------|----------|
| β -Catenin | 0.73 | 0.62 | 0.79 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| BRCA | 0.43 | 0.36 | 0.36 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| E2F1 | 0.51 | 0.42 | 0.54 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| EGFR | -0.52 | -0.43 | -0.42 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| HER2 | -0.45 | -0.5 | -0.26 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| MYC | 0.69 | 0.53 | 0.4 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| p53 | -0.59 | -0.42 | -0.72 |
| | p<0.0001 | p<0.0001 | p<0.0001 |
| p63 | 0.46 | 0.29 | 0.36 |
| | p<0.0001 | p=0.001 | p<0.0001 |
| PI3K | 0.43 | 0.33 | 0.29 |
| | p<0.0001 | p<0.0001 | p=0.002 |
| RAS | 0.51 | 0.2 | 0.4 |
| | p<0.0001 | p=0.017 | p<0.0001 |
| TGF β | -0.23 | -0.3 | -0.13 |
| | p=0.0001 | p<0.0001 | p=0.004 |

Table 3. Estimates of Pearson rho correlation coefficients after meta-analysis of six datasets between pathway activation scores and prognostic gene signatures: wound healing response signature (WHR)/ Invasiveness gene signature IGS and Genomic grade Index (GGI). Most significant correlations are shown. (Threshold p-value adjusted for multiple testing=0.0025)

5. Association of pathway activation scores with survival outcome

While some pathways were associated with survival outcome in one or more datasets, they showed no or opposite result in another dataset. To estimate the overall survival effect of a given pathway, a similar meta-analysis approach was performed to estimate the overall

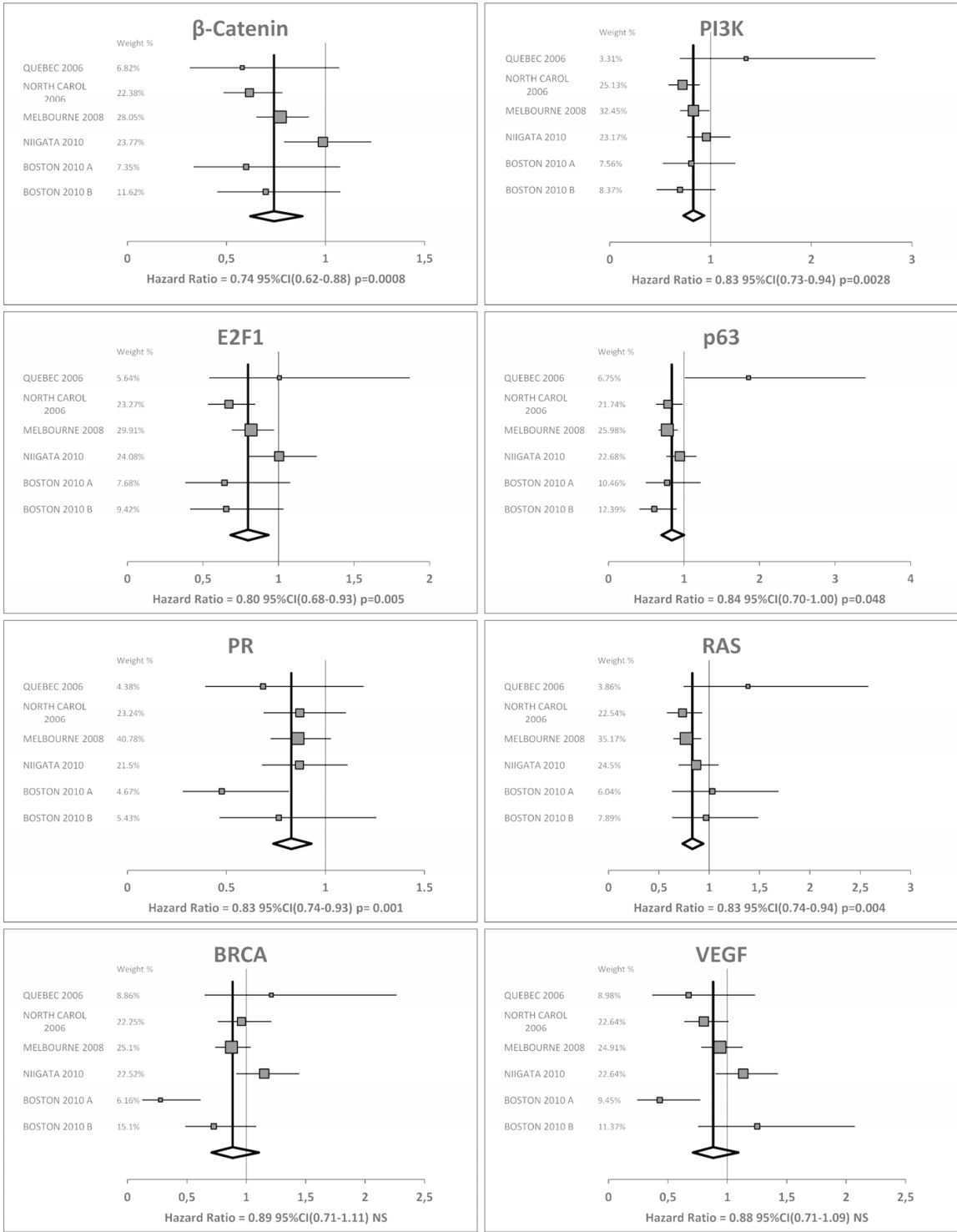


Fig. 3. Forest plots of meta-analysis using a random effects model of the β-Catenin BRCA, E2F1, p63, PR, PI3K, RAS and VEGF pathway.

effect of pathway activation using a random effects model. After this analysis, the β -Catenin, E2F1, PR, p63 PI3K and RAS pathway activation showed a significant association with clinical outcome. Considering the overall effect by means of Hazard Ratios, the β -Catenin pathway showed the most prominent effect after meta-analysis (HR= 0.74; 95%CI [0.62-0.88]). The survival analysis showed that the higher the activation of the β -Catenin pathway, the better the outcome was. Also for PR, E2F1, RAS, PI3K and p63 increased activation of respective pathway was associated with more favourable survival.

Because of these rather unexpected results, we calculated the activation scores of selected discovered pathways in other independent datasets as additional quality control to confirm whether the directions of the activation scores were certainly correct. For β -Catenin the

For the 3 prognostic signatures there was a tendency that a prognostic worse outcome predicted by IGS, WHR and GGI showed an unexpected higher probability of better clinical survival outcome. Further analysis in the Québec dataset showed that chemoresistant patients showed significant lower scores than chemosensitive patients and therefore may explain this finding.

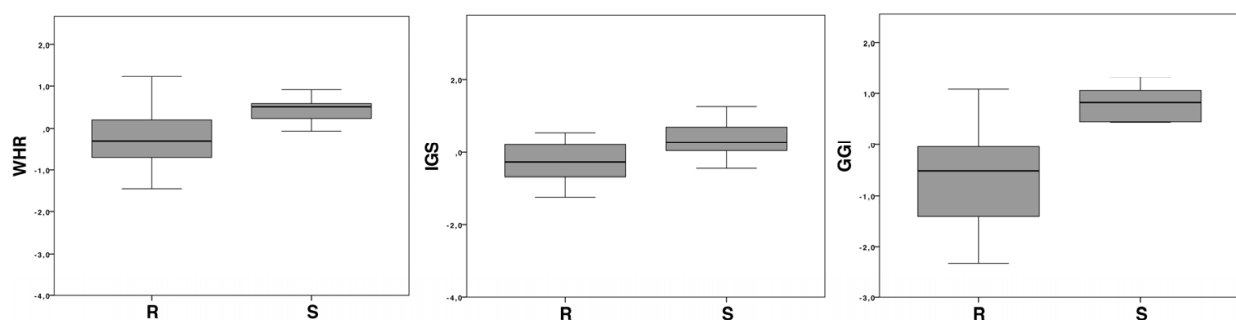


Fig. 4. In the Québec dataset sensitive (S) patients showed a higher genomic grade index (GGI) compared to chemoresistant patients (R) ($p=0.002$). Similarly chemosensitive patients showed a higher wound healing response score ($p=0.02$) and a higher invasiveness gene signature score (IGS) ($p=0.06$).

6. Discussion

Our initial analysis consisted of two datasets. The initial design was to use one dataset, as a discovery dataset while the other one would serve as a validation set. Since bioinformatical mislabelling errors/reproducibility issues have lead to withdrawal of papers of the same research group from which one dataset originated, we sought additional datasets to confirm our findings and render more power and reliability. (Bonnetfoi et al., 2011; Potti et al., 2011) Furthermore, this research group and critical review by another research group have confirmed that the dataset that was used in the present meta-analysis was indeed correctly annotated. (Baggerly, Coombes, & Neeley, 2008; Dressman et al., 2007) With the availability of more datasets, we noticed variation among pathway's association with survival outcome. We therefore used a meta-analysis approach to estimate the overall effect. The advantage is that several studies can be combined despite differences in platforms and methodologies. This overall effect estimation takes into account the number of patients of each separate dataset and confidence interval in the estimation of correlation coefficient of survival hazard ratios. The heterogeneity among datasets (e.g. different patient selection criteria) may partly explain some opposite findings. The Québec dataset is different from others because this

specifically selected patients to study differential expression between chemosensitive versus chemoresistant tumours. (Bachvarov et al., 2006) This dataset therefore may represent the extremities of this disease. Interestingly this dataset showed clearly that chemosensitive patients had tumours that were more likely to be of unfavourable outcome estimated by WHR/IGS/GGI. This contradictory finding may be explained by the finding that these three prognostic signatures are all primarily associated with increased proliferation. (Wirapati et al., 2008) It is known that chemosensitive tumours have higher tumour cell proliferation indexes in serous ovarian cancer. (Itamochi et al., 2002; Têtu et al., 2008) The estimated prognostic values in this survival analysis therefore seems strongly oppositely confounded by the predictive value for platinum/taxane-based chemotherapy. Despite the heterogeneity in datasets and confounding of predictive value versus prognostic value, the E2F1, β -Catenin and the PI3K activation scores showed overall association with survival outcome ($p<0.01$) and consistent significant correlations with three prognostic signatures.

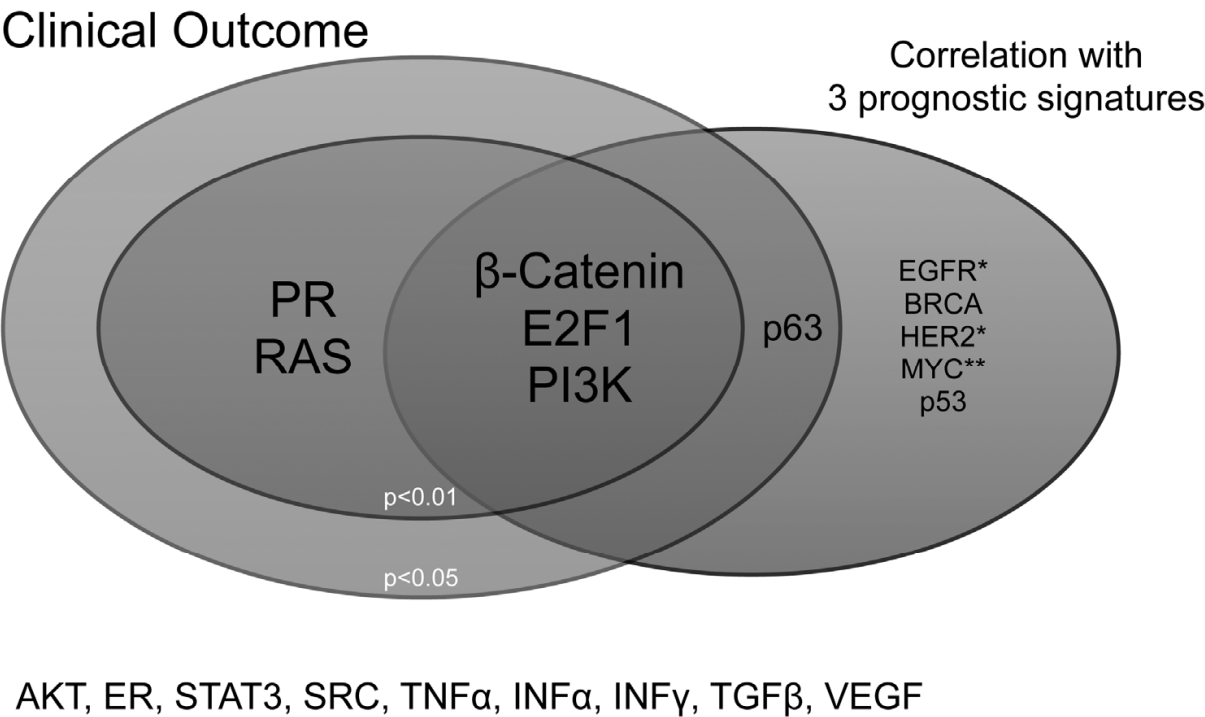


Fig. 5. A Venn diagram is showing combined results of the meta-analysis: β -Catenin, E2F1, p63 and PI3K activation scores showed significant association with survival and were significantly correlated with all three prognostic signatures (WHR/IGS/GGI) after meta-analysis. PR and RAS activation scores were associated with clinical outcome, but did not consistently correlate with prognostic signatures. *Negative correlation coefficient
**borderline significance with clinical outcome

The E2F1 pathway a critical role in proliferation and apoptosis. It has been shown that transcription factor E2F1 interacts with the p53 and PI3K pathway. (Hallstrom, Mori, & Nevins, 2008; Reimer et al., 2006; 2007) Its role in ovarian cancer has been unclear, as other research groups have found similar favourable survival with increased E2F1 pathway activation (Hallstrom et al., 2008), while other findings have shown favourable survival with

decreased *E2F1* gene expression by RT-PCR. (Reimer et al., 2006; 2007) It must be remarked that the latter study included an overrepresentation of patients with clear cell carcinomas (42.9%) and may be less informative here.

The β -Catenin protein is a multifunctional protein. It was originally discovered as a protein that is associated with the cytoplasmatic region of E-cadherin. E-cadherin is a transmembrane protein that is involved in cell-cell contact and cell's adhesive functions. Furthermore, β -Catenin is involved in Wnt signalling as a nuclear transcription factor and is believed to play a role in cancer stem cells. (Nusse, 2008) Loss of its membranous function or a higher nuclear presence has been linked with poor survival in several studies in ovarian cancer based upon immunohistochemical studies. (Faleiro-Rodrigues et al., 2004; Faleiro-Rodrigues, Macedo-Pinto, Pereira, & Lopes, 2004; Irving et al., 2005; 2005; Rosen et al., 2010; 2010; Stawerski et al., 2008; Stawerski, Wagrowska-Danilewicz, Stasikowska, Gottwald, & Danilewicz, 2008; Voutilainen et al., 2006; 2006) In addition, a correlation of β -Catenin protein expression has been described with tumour grade and Ki-67 expression. (Stawerski et al., 2008; Voutilainen et al., 2006) Present results are thus confirmative of earlier findings that β -Catenin is associated with survival outcome. The consideration must be made whether this effect is not attributed to its predictive value to platinum-taxane chemotherapy rather than its prognostic value. In present study, β -Catenin had strong and consistent correlation with IGS/WHR/GGI. Although these signatures were constructed based upon different oncogenic biological processes (wound healing, stem cell phenotype, grade), their major common force has been proven to be cell proliferation. (Wirapati et al., 2008) The observation that chemosensitive patients in present analysis showed significantly higher values of GGI, WHR and IGS renders credibility to this statement.

Similarly, the unexpected findings that increased activation of PI3K-, and RAS- pathways are more favourable for survival may be explained by their predictive value for chemotherapy. This hypothetically may have clinical consequences. Several compounds target the PI3K pathway or downstream effectors (e.g. mTOR) and are under early clinical development in epithelial ovarian cancer. Other compounds have inhibitory effects on the RAS pathway, e.g. lonafarnib (a farnesyltransferase inhibitor). Recent findings of a randomised phase II trial (IGCS meeting 2010, W. Meier et al.) showed that the concomitant addition to standard chemotherapy (first line) and 6-month continuation of lonafarnib in primary epithelial ovarian cancer stage IIB-IV (n=105) resulted in borderline poorer outcome for the experimental- lonafarnib arm (overall survival HR=0.62 95CI%(0.36-1.06) p=0.08) or even resulted in significant unexpected worse outcome (p=0.01) in the experimental stratum of patients with suboptimal debulking. This finding may be relevant in the context of our results. Since increased activation of pathways as RAS and PI3K have been found to be favourable for survival outcome, the question should be asked whether inhibition of one of these pathways concomitant with chemotherapy is desirable. These pathways are driving forces of proliferation, which is an important factor in the efficacy of standard chemotherapeutics. We hypothesize that inhibition of these pathways may therefore also negatively affect the efficacy of these chemotherapies and theoretically induce chemoresistance. This would possibly be an explanation for the recent unexpected findings of lonafarnib in ovarian cancer. Hence, we theorize that these agents may have their potential in ovarian cancer in a sequential adjuvant setting rather than its concomitant combination with chemotherapy.

The PR pathway did not show any relevant association with IGS or GGI. It did show high significant association with survival outcome and WHR. Other immunohistochemical

studies have shown that the PR protein expression has predictive of prognostic value, more than the expression of ER. (Hah et al., 2011; Høgdall et al., 2007; Tangjitgamol, Manusirivithaya, Khunnarong, Jesadapatarakul, & Tanwanich, 2009; X.-Y. Yang, Xi, K.-X. Yang, & Yu, 2009) Since PR expression is a downstream target of the ER pathway, this finding may indicate that an active ER pathway, rather than the expression of ER by itself may be of importance. Anti-hormonal therapies have shown anti-tumoural activity in relapsed/refractory ovarian cancer in phase II studies. (del Carmen et al., 2003; Papadimitriou et al., 2004; Smyth et al., 2007; C. J. Williams, 2001; C. Williams, Simera, & Bryant, 2010) Biomarker studies have shown that increasing ER expression was associated with increasing CA125 response rate. (Smyth et al., 2007) We suggest that further studies are needed to study if PR expression may add value as a suitable biomarker to select patients for anti-hormonal therapy in ovarian cancer.

7. Conclusions

To conclude, oncogenic pathway profiling of advanced serous ovarian tumours revealed that it is difficult to estimate the true prognostic value of a pathway since there seems confounding of predictive factors. Despite these biases, with a meta-analysis approach of 6 independent datasets generated on different micro-array platforms, we found that a PR and RAS activation score was associated with clinical outcome. Activation scores for β -Catenin, p63, E2F1 and PI3K were also associated with survival and were consistently correlated with three prognostic gene signatures.

8. References

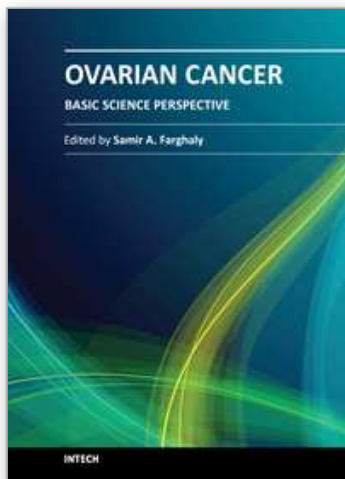
- Audeh, M. W., Carmichael, J., Penson, R. T., Friedlander, M., Powell, B., Bell-McGuinn, K. M., Scott, C., et al. (2010). Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and recurrent ovarian cancer: a proof-of-concept trial *Lancet*, 376(9737), 245–251. doi:10.1016/S0140-6736(10)60893-8
- Bachvarov, D., L'esperance, S., Popa, I., Bachvarova, M., Plante, M., & Têtu, B. (2006). Gene expression patterns of chemoresistant and chemosensitive serous epithelial ovarian tumors with possible predictive value in response to initial chemotherapy *International journal of oncology*, 29(4), 919–933.
- Baggerly, K. A., Coombes, K. R., & Neeley, E. S. (2008). Run batch effects potentially compromise the usefulness of genomic signatures for ovarian cancer *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 26(7), 1186–7; author reply 1187–8. doi:10.1200/JCO.2007.15.1951
- Bild, A. H., Yao, G., Chang, J. T., Wang, Q., Potti, A., Chasse, D., Joshi, M.-B., et al. (2006). Oncogenic pathway signatures in human cancers as a guide to targeted therapies. *Nature*, 439(7074), 353–357. doi:10.1038/nature04296
- Bonnefoi, H., Potti, A., Delorenzi, M., Mauriac, L., Campone, M., Tubiana-Hulin, M., Petit, T., et al. (2011). Retraction--Validation of gene signatures that predict the response of breast cancer to neoadjuvant chemotherapy: a substudy of the EORTC 10994/BIG 00-01 clinical trial *The lancet oncology*, 12(2), 116. doi:10.1016/S1470-2045(11)70011-0
- Burger, R. A., Brady, M. F., Bookman, M. A., Walker, J. L., Homesley, H. D., Fowler, J., Monk, B. J., et al. (2010). Phase III trial of bevacizumab (BEV) in the primary

- treatment of advanced epithelial ovarian cancer (EOC), primary peritoneal cancer (PPC), or fallopian tube cancer (FTC): A Gynecologic Oncology Group study. - ASCO. *Journal of Clinical Oncology*, 28(18s), suppl; abstr LBA1.
- Cannistra, S. A. (2004). Cancer of the ovary. *The New England journal of medicine*, 351(24), 2519–2529. doi:10.1056/NEJMra041842
- Chang, H. Y., Nuyten, D. S. A., Sneddon, J. B., Hastie, T., Tibshirani, R., Sørbye, T., Dai, H., et al. (2005). Robustness, scalability, and integration of a wound-response gene expression signature in predicting breast cancer survival. *Proceedings of the National Academy of Sciences of the United States of America*, 102(10), 3738–3743. doi:10.1073/pnas.0409462102
- del Carmen, M. G., Fuller, A. F., Matulonis, U., Horick, N. K., Goodman, A., Duska, L. R., Penson, R., et al. (2003). Phase II trial of anastrozole in women with asymptomatic müllerian cancer *Gynecologic oncology*, 91(3), 596–602.
- Dressman, H. K., Berchuck, A., Chan, G., Zhai, J., Bild, A., Sayer, R., Cragun, J., et al. (2007). An integrated genomic-based approach to individualized treatment of patients with advanced-stage ovarian cancer. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 25(5), 517–525. doi:10.1200/JCO.2006.06.3743
- Faleiro-Rodrigues, C., Macedo-Pinto, I., Pereira, D., & Lopes, C. S. (2004). Loss of beta-catenin is associated with poor survival in ovarian carcinomas *International journal of gynecological pathology : official journal of the International Society of Gynecological Pathologists*, 23(4), 337–346.
- Fong, P. C., Boss, D. S., Yap, T. A., Tutt, A., Wu, P., Mergui-Roelvink, M., Mortimer, P., et al. (2009). Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers *The New England journal of medicine*, 361(2), 123–134. doi:10.1056/NEJMoa0900212
- Fong, P. C., Yap, T. A., Boss, D. S., Carden, C. P., Mergui-Roelvink, M., Gourley, C., De Greve, J., et al. (2010). Poly(ADP)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 28(15), 2512–2519. doi:10.1200/JCO.2009.26.9589
- Gatza, M. L., Lucas, J. E., Barry, W. T., Kim, J. W., Wang, Q., Crawford, M. D., Datto, M. B., et al. (2010). A pathway-based classification of human breast cancer. *Proceedings of the National Academy of Sciences*, 107(15), 6994–6999. doi:10.1073/pnas.0912708107
- Hah, N., Danko, C. G., Core, L., Waterfall, J. J., Siepel, A., Lis, J. T., & Kraus, W. L. (2011). A Rapid, Extensive, and Transient Transcriptional Response to Estrogen Signaling in Breast Cancer Cells. *Cell*, 145(4), 622–634. Elsevier Inc. doi:10.1016/j.cell.2011.03.042
- Hallstrom, T. C., Mori, S., & Nevins, J. R. (2008). An E2F1-dependent gene expression program that determines the balance between proliferation and cell death *Cancer cell*, 13(1), 11–22. doi:10.1016/j.ccr.2007.11.031
- Hu, Z., Fan, C., Livasy, C., He, X., Oh, D. S., Ewend, M. G., Carey, L. A., et al. (2009). A compact VEGF signature associated with distant metastases and poor outcomes. *BMC medicine*, 7, 9. doi:10.1186/1741-7015-7-9
- Høgdall, E. V. S., Christensen, L., Høgdall, C. K., Blaakaer, J., Gayther, S., Jacobs, I. J., Christensen, I. J., et al. (2007). Prognostic value of estrogen receptor and progesterone receptor tumor expression in Danish ovarian cancer patients: from the “MALOVA” ovarian cancer study *Oncology reports*, 18(5), 1051–1059.

- Irving, J. A., Catasús, L., Gallardo, A., Bussaglia, E., Romero, M., Matias-Guiu, X., & Prat, J. (2005). Synchronous endometrioid carcinomas of the uterine corpus and ovary: alterations in the beta-catenin (CTNNB1) pathway are associated with independent primary tumors and favorable prognosis *Human pathology*, 36(6), 605–619. doi:10.1016/j.humpath.2005.03.005
- Itamochi, H., Kigawa, J., Sugiyama, T., Kikuchi, Y., Suzuki, M., & Terakawa, N. (2002). Low proliferation activity may be associated with chemoresistance in clear cell carcinoma of the ovary *Obstetrics and gynecology*, 100(2), 281–287.
- Konstantinopoulos, P. A., Spentzos, D., Karlan, B. Y., Taniguchi, T., Fountzilas, E., Francoeur, N., Levine, D. A., et al. (2010). Gene expression profile of BRCAness that correlates with responsiveness to chemotherapy and with outcome in patients with epithelial ovarian cancer. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*, 28(22), 3555–3561. doi:10.1200/JCO.2009.27.5719
- Lauss, M., Ringnér, M., & Höglund, M. (2010). Prediction of stage, grade, and survival in bladder cancer using genome-wide expression data: a validation study. *Clinical cancer research : an official journal of the American Association for Cancer Research*, 16(17), 4421–4433. doi:10.1158/1078-0432.CCR-10-0606
- Liu, R., Wang, X., Chen, G. Y., Dalerba, P., Gurney, A., Hoey, T., Sherlock, G., et al. (2007). The prognostic role of a gene signature from tumorigenic breast-cancer cells. *The New England journal of medicine*, 356(3), 217–226. doi:10.1056/NEJMoa063994
- Mostert, W., Stevenson, M., Acharya, C., Chan, I., Walters, K., Lamlertthong, W., Barry, W., et al. (2010). Age- and sex-specific genomic profiles in non-small cell lung cancer. *JAMA : the journal of the American Medical Association*, 303(6), 535–543. doi:10.1001/jama.2010.80
- Nusse, R. (2008). Wnt signaling and stem cell control *Cell research*, 18(5), 523–527. doi:10.1038/cr.2008.47
- Papadimitriou, C. A., Markaki, S., Siapkarakas, J., Vlachos, G., Efstathiou, E., Grimi, I., Hamilos, G., et al. (2004). Hormonal therapy with letrozole for relapsed epithelial ovarian cancer. Long-term results of a phase II study *Oncology*, 66(2), 112–117. doi:10.1159/000077436
- Potti, A., Dressman, H. K., Bild, A., Riedel, R. F., Chan, G., Sayer, R., Cragun, J., et al. (2011). Retraction: Genomic signatures to guide the use of chemotherapeutics *Nature medicine*, 17(1), 135. doi:10.1038/nm0111-135
- Reimer, D., Sadr, S., Wiedemair, A., Goebel, G., Concin, N., Hofstetter, G., Marth, C., et al. (2006). Expression of the E2F family of transcription factors and its clinical relevance in ovarian cancer *Annals of the New York Academy of Sciences*, 1091, 270–281. doi:10.1196/annals.1378.073
- Reimer, D., Sadr, S., Wiedemair, A., Stadlmann, S., Concin, N., Hofstetter, G., Müller-Holzner, E., et al. (2007). Clinical relevance of E2F family members in ovarian cancer--an evaluation in a training set of 77 patients *Clinical cancer research : an official journal of the American Association for Cancer Research*, 13(1), 144–151. doi:10.1158/1078-0432.CCR-06-0780
- Rosen, D. G., Zhang, Z., Chang, B., Wang, X., Lin, E., & Liu, J. (2010). Low membranous expression of beta-catenin and high mitotic count predict poor prognosis in endometrioid carcinoma of the ovary *Modern pathology : an official journal of the*

- United States and Canadian Academy of Pathology, Inc*, 23(1), 113–122. doi:10.1038/modpathol.2009.141
- Smyth, J. F., Gourley, C., Walker, G., MacKean, M. J., Stevenson, A., Williams, A. R. W., Nafussi, A. A., et al. (2007). Antiestrogen therapy is active in selected ovarian cancer cases: the use of letrozole in estrogen receptor-positive patients *Clinical cancer research : an official journal of the American Association for Cancer Research*, 13(12), 3617–3622. doi:10.1158/1078-0432.CCR-06-2878
- Sotiriou, C., Wirapati, P., Loi, S., Harris, A., Fox, S., Smeds, J., Nordgren, H., et al. (2006). Gene expression profiling in breast cancer: understanding the molecular basis of histologic grade to improve prognosis *Journal of the National Cancer Institute*, 98(4), 262–272. doi:10.1093/jnci/djj052
- Stawerski, P., Wagrowska-Danilewicz, M., Stasikowska, O., Gottwald, L., & Danilewicz, M. (2008). Immunoexpression of beta-catenin–E-cadherin complex in primary serous ovarian tumors *Polish journal of pathology : official journal of the Polish Society of Pathologists*, 59(1), 27–32.
- Tangjitgamol, S., Manusirivithaya, S., Khunnarong, J., Jesadapatarakul, S., & Tanwanich, S. (2009). Expressions of estrogen and progesterone receptors in epithelial ovarian cancer: a clinicopathologic study *International journal of gynecological cancer : official journal of the International Gynecological Cancer Society*, 19(4), 620–627. doi:10.1111/IGC.0b013e3181a44b62
- Têtu, B., Popa, I., Bairati, I., L'esperance, S., Bachvarova, M., Plante, M., Harel, F., et al. (2008). Immunohistochemical analysis of possible chemoresistance markers identified by micro-arrays on serous ovarian carcinomas *Modern pathology : an official journal of the United States and Canadian Academy of Pathology, Inc*, 21(8), 1002–1010. doi:10.1038/modpathol.2008.80
- Tothill, R. W., Tinker, A. V., George, J., Brown, R., Fox, S. B., Lade, S., Johnson, D. S., et al. (2008). Novel molecular subtypes of serous and endometrioid ovarian cancer linked to clinical outcome. *Clinical cancer research : an official journal of the American Association for Cancer Research*, 14(16), 5198–5208. doi:10.1158/1078-0432.CCR-08-0196
- Trinh, X. B., Tjalma, W. A. A., Dirix, L. Y., Vermeulen, P. B., Peeters, D. J., Bachvarov, D., Plante, M., et al. (2011). Microarray-Based Oncogenic Pathway Profiling in Advanced Serous Papillary Ovarian Carcinoma. (S. K. Batra, Ed.) *PLoS ONE*, 6(7), e22469. doi:10.1371/journal.pone.0022469.t003
- Voutilainen, K. A., Anttila, M. A., Sillanpää, S. M., Ropponen, K. M., Saarikoski, S. V., Juhola, M. T., & Kosma, V.-M. (2006). Prognostic significance of E-cadherin-catenin complex in epithelial ovarian cancer *Journal of Clinical Pathology*, 59(5), 460–467. doi:10.1136/jcp.2005.029876
- Williams, C. J. (2001). Tamoxifen for relapse of ovarian cancer *Cochrane database of systematic reviews (Online)*, (1), CD001034. doi:10.1002/14651858.CD001034
- Williams, C., Simera, I., & Bryant, A. (2010). Tamoxifen for relapse of ovarian cancer *Cochrane database of systematic reviews (Online)*, (3), CD001034. doi:10.1002/14651858.CD001034.pub2
- Wirapati, P., Sotiriou, C., Kunkel, S., Farmer, P., Pradervand, S., Haibe-Kains, B., Desmedt, C., et al. (2008). Meta-analysis of gene expression profiles in breast cancer: toward a

- unified understanding of breast cancer subtyping and prognosis signatures *Breast cancer research : BCR*, 10(4), R65. doi:10.1186/bcr2124
- Yang, X.-Y., Xi, M.-R., Yang, K.-X., & Yu, H. (2009). Prognostic value of estrogen receptor and progesterone receptor status in young Chinese ovarian carcinoma patients *Gynecologic oncology*, 113(1), 99–104. doi:10.1016/j.ygyno.2008.12.018
- Yoshihara, K., Tajima, A., Yahata, T., Kodama, S., Fujiwara, H., Suzuki, M., Onishi, Y., et al. (2010). Gene expression profile for predicting survival in advanced-stage serous ovarian cancer across two independent datasets. *PLoS ONE*, 5(3), e9615. doi:10.1371/journal.pone.0009615



Ovarian Cancer - Basic Science Perspective

Edited by Dr. Samir Farghaly

ISBN 978-953-307-812-0

Hard cover, 406 pages

Publisher InTech

Published online 17, February, 2012

Published in print edition February, 2012

Worldwide, Ovarian carcinoma continues to be responsible for more deaths than all other gynecologic malignancies combined. International leaders in the field address the critical biologic and basic science issues relevant to the disease. The book details the molecular biological aspects of ovarian cancer. It provides molecular biology techniques of understanding this cancer. The techniques are designed to determine tumor genetics, expression, and protein function, and to elucidate the genetic mechanisms by which gene and immunotherapies may be perfected. It provides an analysis of current research into aspects of malignant transformation, growth control, and metastasis. A comprehensive spectrum of topics is covered providing up to date information on scientific discoveries and management considerations.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Xuan Bich Trinh, Peter A. Van Dam, Luc Y. Dirix, Steven J. van Laere and Wiebren A. A. Tjalma (2012).
Oncogenic Pathway Signatures and Survival Outcome, Ovarian Cancer - Basic Science Perspective, Dr. Samir Farghaly (Ed.), ISBN: 978-953-307-812-0, InTech, Available from: <http://www.intechopen.com/books/ovarian-cancer-basic-science-perspective/oncogenic-pathway-signatures-and-survival-outcome>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen