

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Biomarkers and Physiological Agents in Severe Sepsis and Septic Shock

D. Paramythiotis, E. Digkas and G. Dryllis
*1st Propedeutic Surgical Department
Aristotle University of Thessaloniki
A.H.E.P.A. University Hospital
Thessaloniki
Greece*

1. Introduction

The term sepsis implies the presence of infection with signs of systemic body response (tachycardia, tachypnea, fever, etc.) and conditions of severe sepsis and septic shock, further disturbance in organ perfusion (impaired consciousness, hypoxia, oliguria) required fluid administration and inotropic and / or vasopressor drugs, respectively (Levy et al. 2003, Bone et al.1992).

Sepsis is an enormously complex clinical syndrome that arises from the activation of an innate host response to danger. Sepsis is associated with infections of bacteria, viruses, fungi and endotoxins, whether or not being evidenced by culture. The above systematic inflammatory response in humans occurs not only in sepsis but also in other non-infectious conditions such as pancreatitis, ischemia, severe trauma, etc. The host responds in the same way in infectious as in non-infectious events, which is described by the term systemic inflammatory response syndrome (SIRS). Insights into the biology of innate immunity suggest that in terms of impact on outcomes, the specific insult evoking the response is less important than the nature of the response itself. The terms sepsis and septicemia were-until relatively recently- used indiscriminately. However, researchers tend to use those two terms with different and distinct meanings according to several conditions under which each of this condition happens (Pezilli et al, 2010, Dhar et al, 2008, Hoover et al, 2006, Matzinger et al, 2002, Bone et al, 1992). Sepsis is a very complex chain of events including inflammatory and anti-inflammatory processes, circulatory abnormalities and humoral and cellular reactions. Its complexity, in addition to high variability of its non-specific signs and symptoms, make early diagnosis and determination of its severity very important, as sepsis can be lethal for patients. Early diagnosis increases the possibility of starting a specific therapy in time (Lever et al, 2007, Hotchkiss and Karl, 2003, Zambon et al., 2008).

As mentioned above, sepsis may be initialized by local inflammation in order to inactivate invading factors. At first the host recruits and activates leukocytes at infectious foci. During that phase, innate immune cells recognize invading factors through pattern recognition receptors (PRRs) on the cells. Toll-like receptors (TLRs), which belong to PRRs, activate

immune cells. This activation is caused by the production of pro-inflammatory cytokines and chemokines. Bacterial products and viral proteins are linked to various and distinct TLRs and are the main cause of cytokine and chemokine production. Following the complement is activated which activates leukocytes. Cells activated through all this process produce and secrete many agents such as leukotriene, nitric oxide and inducible nitric oxide synthase (iNOS) and free radicals. All this process is initiated by many proinflammatory mediators (cytokines, adhesion molecules, vasodilating mediators and reactive oxygen species). If the above body reaction fails to achieve its goals, then some of the invading factors may invade into the bloodstream and trigger a systemic inflammatory reaction. This happens mainly through overproduction of inflammatory cytokines (eg TNF, IFNs and IL-1,6). In all the inflammatory process many agents are overproduced and are of high importance for patients health. Apoptosis has a key role in sepsis and it is influenced by many factors such as caspases, pro-inflammatory cytokines release –such as TNF- α , IFN- γ , IL-1 β - and anti-inflammatory cytokines e.g. TGF β , IL-10 and IL-13, many different pathogenic toxins, such as LPS, CLP and CASP, while it happens in many organs such as heart, glomeruli, liver, bone marrow, thymus, spleen, lungs and gut. The TNFR- CD40- TRADD- FADD pathway –activated by TNF- α leads to inhibition of cas-8, which is an initiator factor of apoptosis, and therefore that pathway inhibits apoptosis. Additionally, neutrophil and monocyte activation leads to inhibition of Bcl-1, which belongs to Bcl-2 family and inhibits apoptosis. On the contrary, activation of Cardinal and Fas death receptor by Bim (of the Bcl-2 family), which leads to activation of lymphocyte apoptosis, causes a decline of inflammatory response. Apoptosis inhibition leads to SIRS, while its activation leads to immunoparalysis and MODS. However, apoptosis inhibition may have a beneficial role for septic patient. Immunodeficiency is another characteristic of sepsis, which happens due to increased apoptosis of CD4 T cells, dendritic cells and B cells and leads to immunoparalysis. The coagulation system is activated by cytokines and oxidants through tissue factor induction. Antithrombin III plays a dominant role among antithrombotic and fibrinolytic agents. Protein C or autoprothrombin IIA, protect cells from sepsis and it has strong anticoagulative effects through inactivating Va and VIIIa factors of the coagulation system (Remick, 2007, Okazaki and Matsukawa, 2009, Hotchkiss et al, 2003, McDonald, et al 2000, Sohn et al 2003, Hildeman et al 2002, O'Neill et al 2000). **(Figure 1)**

Physiological agents participating in sepsis pathophysiology cascades are used as biomarkers in sepsis diagnosis. Biomarker is “a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic process or pharmacologic responses to a therapeutic intervention (BDWG, 2001). The exact role of biomarkers in septic patient’s management is not yet defined. Nevertheless, biomarkers can play an important role in early diagnosis and severity determination of sepsis. Moreover they can differentiate bacterial from viral and fungal infection and systemic from local infection. Differentiation of Gram-positive from Gram-negative microorganisms as cause of sepsis is a potential use of biomarkers. Biomarkers may also be used in guiding antibiotic therapy, guiding therapy, and evaluating the response to therapy. Other possible biomarker applications are prediction of sepsis complications and development of organ dysfunction (BDWG, 2001, Marshall and Reinhart, 2009, Dellinger et al, 2008). It has been shown that various biomarkers are very useful in clinical practice in sepsis diagnosis and some of them are thought superior to clinical signs as far as diagnosis.

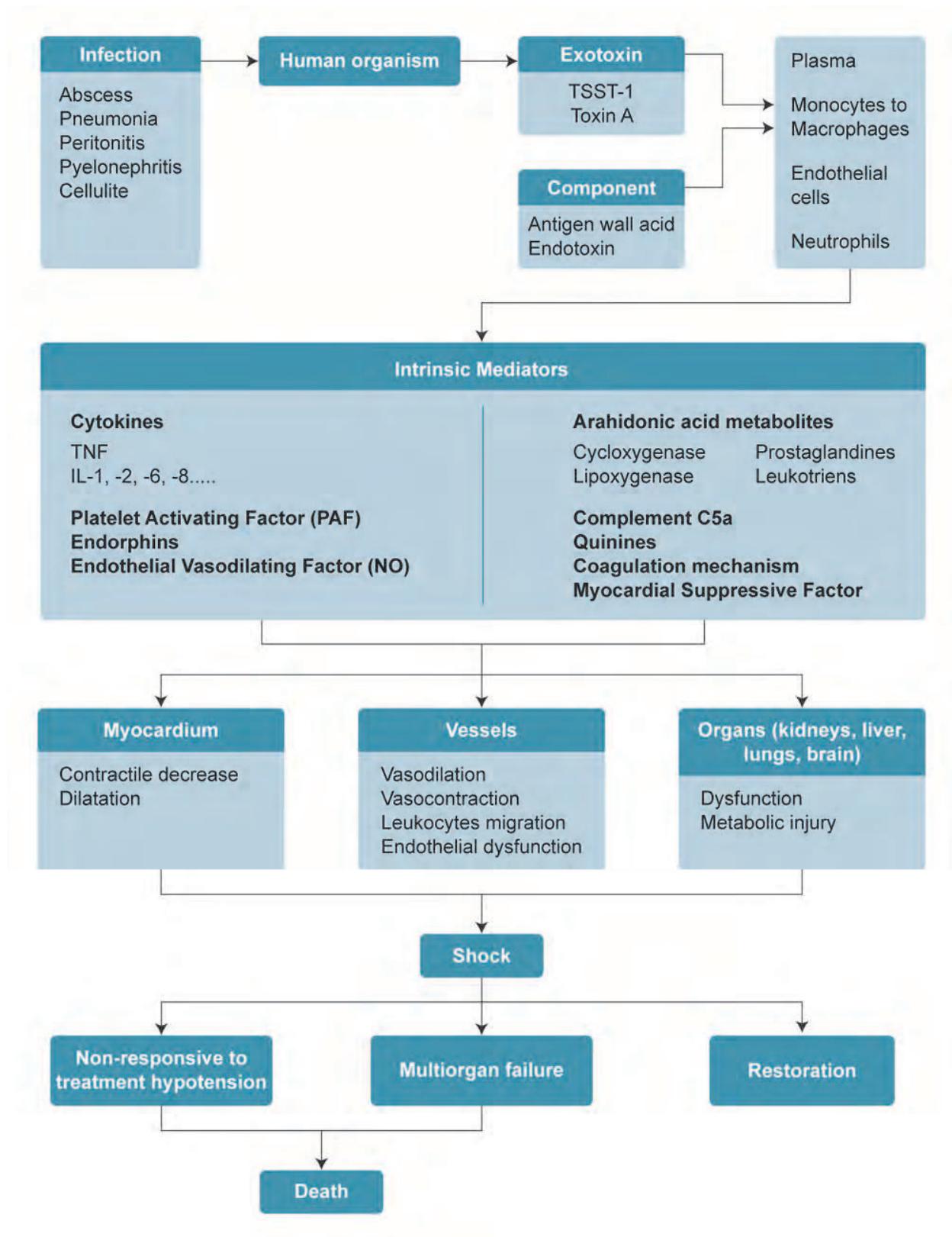


Fig. 1. Pathogenetic mechanism of sepsis (improved figure by Parillo et al, 1993)

Biomarkers of sepsis can be divided in nine categories, which are: 1. Cytokine biomarkers, 2. Coagulation biomarkers, 3. Acute phase protein biomarkers, 4. Receptor biomarkers, 5. Cell marker related biomarkers, 6. Vascular and endothelial damage biomarkers, 7. Vasodilation related biomarkers, 8. Organ dysfunction biomarkers, 9. Genetic biomarkers.

2. Cytokine biomarkers

Cytokines participate in the pro-inflammatory as in the inflammatory cascade of sepsis and septic shock. The release of pro-inflammatory mediators characterizes the initial phase of sepsis. Persistence of the latter provokes acquired immunodepression, related to an anti-inflammatory profile, and hence to a delayed decrease in hypersensitivity, an incapacity to cope with the infection and the onset of inflammation. The first human defence mechanism which is activated is the intrinsic immune system (innate immune system). It is the first line of defence and it is motivated by both exogenous (e.g. Gram-, Gram + bacteria) and endogenous factors (e.g. Heat shock protein, direct cellular injury, ischemia-reperfusion). Both in SIRS and sepsis, there is an excessive production of pro-inflammatory cytokines, adhesion molecules, vasodilating mediators and reactive oxygen species, therefore they are detectable in blood, where they are normally absent. However, the circulating cytokines are merely the tip of the iceberg and leukocyte-associated cytokines can be identified even when amounts in plasma are undetectable. Pro-inflammatory responses lead to the activation of successive biological reactions which include: Inflammatory mediators (cytokines, chemokines and lipid mediators) and coagulation / fibrinolysis system.

Cytokines act as inflammatory mediators and regulators of both the immune and inflammatory reactions. Pro-inflammatory cytokines are released in a cascade, with TNF- α and IL-1 β being the initial cytokines. These cytokines stimulate the production of other cytokines such as IL-6, IL-8, IL-10 and MIP-1 α (macrophage inflammatory protein-1 α). TNF α , IL-1 β , IL-6, IL-8 and IL-10 are linked to morbidity and mortality in septic patients. IL-6 levels in plasma rise according to sepsis severity, but that does not always happen. The median levels of IL-6 in patients with SIRS are approximately 10-fold higher than that of healthy people, in sepsis are approximately 20-fold higher, in severe sepsis 60-fold higher and in septic shock are 100-fold higher. But, in some patients IL-6 levels are very similar to those of healthy people (3pg/ml). (Socha et al, 2006, Halter et al, 2005, Akira et al, 2003, Mokart et al, 2002, Paterson et al, 2000, Hack et al, 1997, Casey et al, 1993, Pinsky et al, 1993). IL-6 is induced by TNF- α and has a longer half-life, reliably measured in blood after insult to the host. IL-6, as a marker of infection, is relatively nonspecific as it is elevated in a variety of inflammatory states. IL-6 is one of the initial cytokines released in inflammation, as mentioned above, and may be an early predictor of more downstream effects, such as organ dysfunction. IL-6 may perform well as a diagnostic and prognostic tool of sepsis. IL-6 levels of more than 1000 ng/mL are highly predictive of sepsis related death. The diagnostic and prognostic accuracy of IL-6 may depend on time and frequency of measurement and underlying illness severity, perhaps suggesting the importance of trending cytokine levels with clinical course and therapy As IL-6, IL-8 is not so a good indicator of systemic infection, although elevated IL-8 levels are indicative of multiple organ failure and an increased likelihood of death. Measurement of IL-6 and IL-8 is not optimal for discriminating patients with infectious from those with noninfectious conditions (Chiesa et al, 2003, Iglesias et al, 2003, Harbarth, 2001, Muller et al, 2000, Panacek, 1999, Martin, 1997, De werra, 1997).

Tumor Necrosis Factor- α (TNF- α), which is mainly produced by macrophages, lymphocytes and fibroblasts, causes fever and elevated liver enzymes among others and activates three

main cell signaling pathways, which lead to inflammation and cell death, by binding to its receptor. Plasma levels of TNF- α are increased rapidly after infection and that's how TNF- α activates the inflammation cascade. Additionally, TNF- α levels in plasma are increased due to the elevated production of Haemoxygenase-1 (HO-1), which is implicated by lung injury and organ perfusion (Socha et al, 2006, Kalil et al, 2004, Kan et al, 1999). There are two types of TNF α receptors: TNF-R type 1 (CD120a) - which binds only to TNF α and TNF-R type 2 (CD120b) - which binds both to TNF α and TNF β . The three cell signaling pathways, leading to inflammation, activated by TNF- α are the following:

1. TRADD- RIP- TRAF2 pathway stimulates NF- κ B pathway and production of antiapoptotic factors, such as Bcl-2.
2. TRADD- RIP-TRAF2-ASK1-MAPK pathway leads mainly to AP-1 production and downstream to the production of pro-apoptotic factors and cell proliferation and cell maturation.

TNFR- CD40 TRADD- FADD pathway leads to cell apoptosis by triggering caspase family cascade (Jaffer et al, 2010) (**Figure 2**).

Cytokines also participate in anti-inflammatory cascade. When the inflammatory response is triggered, anti-inflammatory mechanisms are simultaneously activated. Elevated production of pro-inflammatory cytokines causes the simultaneous production of specific cytokines such as IL-4, IL-10, IL-13, soluble TNF receptor and reduction of lymphocyte cells production. Th1 lymphocytes secrete TNF- α , IL-2 and IFN- γ , while T17 lymphocytes secrete IL-17 leading neutrophils to the infection field, unlike Th2 lymphocytes secrete IL-4, IL-6 and IL-10, while regulatory T-cells (Treg) mediate cytokine secretion amplifying the anti-inflammatory response. Especially IL-10 -produced also by monocytes- is suspensory to Th1-cytokines secretion (and mainly to TNF- α) and inhibits the NF- κ B JAK-STAT pathway. IL-13 is secreted by Th-cells and leads to elevated production of IgE, matrix-metalloproteinases (MMPs) and neutrophils while it regulates TNF- α and leucocyte response to SIRS. TNF- α and IL-10 levels are found increased in patients with bad prognosis and the increase is higher in patients with positive blood culture. IL-10 levels can be used to discriminate sepsis from sever sepsis (Jaffer et al 2010, Giamarellos- Bourboulis 2010, Freitas et al 2009, Sobieski et al 2008, Van der Poll et al 2008, Munford et al 2001, Rodriguez-Gaspar et al 2001, Socha et al 2006, Whitlock et al 2006).

3. Coagulation biomarkers

Macrophage Migration Inhibitory Factor (MIF) has a critical role in activating the immune system and accelerating the pro-inflammatory response in sepsis (that is why it is thought that in the near future MIF will be a significant target in the treatment of sepsis). Tissue factor activates coagulation cascade (**figure 3**) and fibrinolysis system. Antithrombin III plays a dominant role among antithrombotic and fibrinolytic agents. Antithrombin III is involved in the formation of a complex constituted by activated FVIIa, FIXa, Fxa and FXIa factors and kallikrein. When antithrombin III is inhibited e.g by prostacyclin, Disseminated Intravascular Coagulation (DIC) is caused -which is considered as an important factor in Multi Organ Dysfunction (MODS) development because its normal function is to lower platelet adhesion and activate monocytes. Finally, we must refer to protein C or autoproteolytic IIa, which protects cells from sepsis and it has strong anticoagulative effects through inactivating Va and VIIIa factors of the coagulation system (Mosnier et al, 2007, Hotchkiss et al, 2003, Nicolaes et al, 2003, Mather et al, 1996).

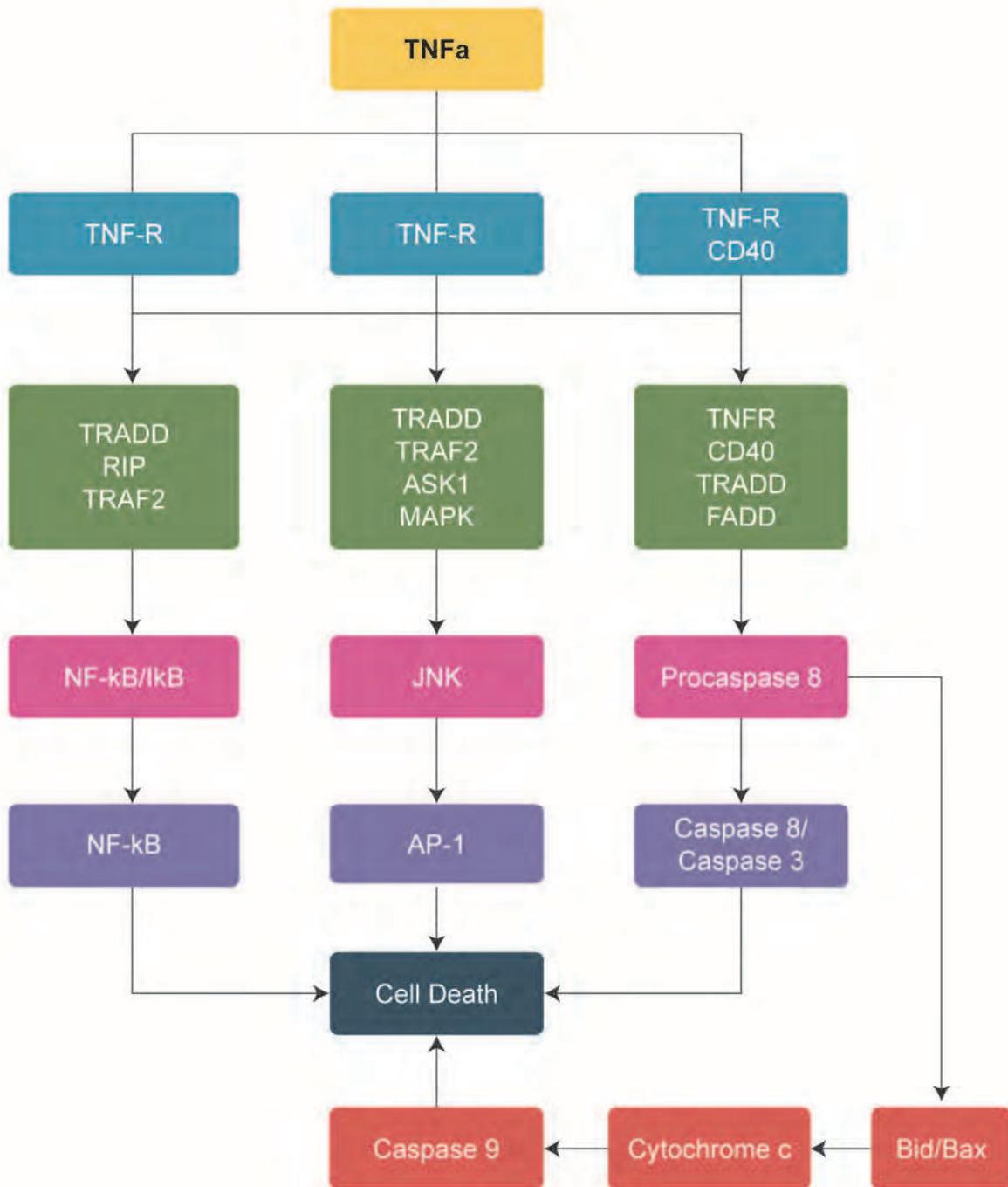


Fig. 2. TNF α activated cellular pathways

The more interesting coagulation factors for sepsis, whose levels have been proposed to be in accordance to sepsis severity and have some prognostic use for septic patient, are easily counted in clinical practice. Activated partial thromboplastin time (aPTT) measurement waveform has been correlated to sepsis. An atypical biphasic transmittance waveform due to decreased light transmission has been observed and it is more common in patients with sepsis. This phenomenon may be due to the formation and precipitation of a calcium dependent complex among C-reactive protein (CRP) and very low density lipoprotein

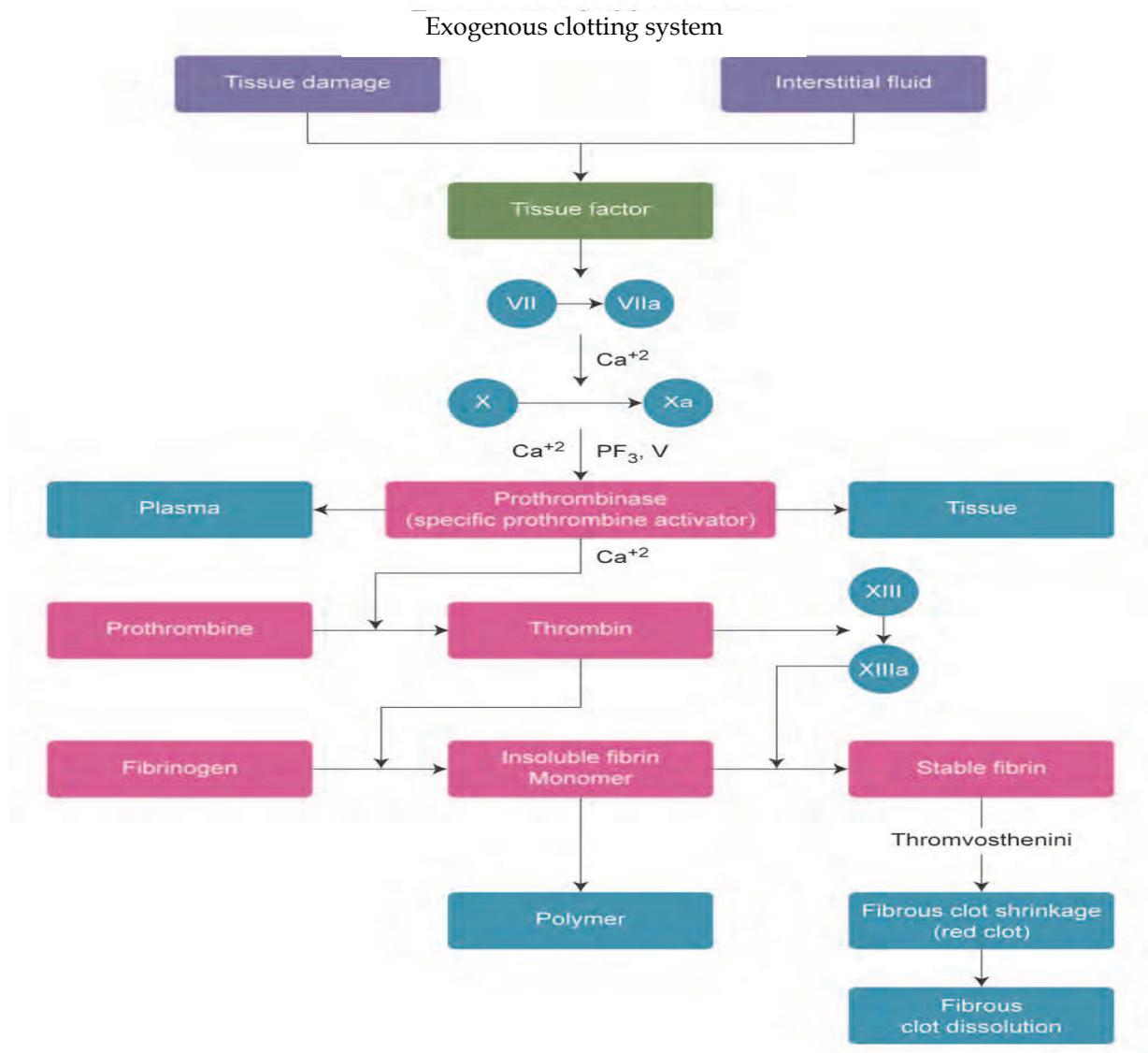


Fig. 3. The exogenous clotting system

(VLDL) in serum of septic patients. That waveform is correlated with negative prediction, sepsis induced disseminated intravascular coagulation and multiorgan failure. Plasminogen activator inhibitor-1 high levels have predictive value for the development of multiorgan failure in septic patients and they are strong correlated with bad prognosis of septic patients with sepsis induced disseminated intravascular coagulation especially when plasminogen activator inhibitor levels are higher than 90 ng/ml. Antithrombin III concentration has been proposed as a prognostic factor, as its levels are lower in septic patients who are not possible to survive. Low protein C levels in septic patient's blood are indicative of increased risk of morbidity and mortality (Zakariah et al, 2008, Madoiwa et al, 2006, Petilla et al, 2002, Fisher et al, 2000).

4. Acute phase protein biomarkers

Acute phase protein biomarkers have been more extensively correlated with prognosis of sepsis, compared to other biomarkers. The two main acute phase protein biomarkers which

are related to sepsis are procalcitonin (PCT) and C-reactive protein (CRP). PCT has a bigger sensitivity and specificity in prognosis of sepsis compared to CRP, but CRP is more available and most widely used in clinical practice. Also, PCT is generally elevated in various conditions correlated with inflammatory response, whether CRP has a better prognostic value in the response of therapy in septic patients. However, there are contradictory views for the utility of both biomarkers in prognosis of sepsis and further clinical studies must be demonstrated in this direction. Serum CRP may be used as a biomarker as its concentrations increase in response to inflammation, its half-life is short but its kinetics are not as good as those of PCT. CRP levels are increased in sepsis, but its use as a diagnostic biomarker is not yet established. CRP is proposed to be used as a discriminating factor between septic from non septic shock while it is less accurate than PCT in SIRS from sepsis differentiation. CRP is a poor predictive biomarker of sepsis outcome. Many experimental and clinical trials suggest that PCT is a specific marker for severe bacterial infection and it may be used to distinguish patients who have sepsis from patients who have SIRS in clinical routine. PCT has been shown to be a better biomarker than IL-6 and CRP in characterizing noninfectious from infectious acute respiratory distress syndrome. PCT remains among the most promising biomarkers in sepsis (Jensen, 2006, Castelli, 2004, Clec'h, 2004, Povoas, 2004, Luzzani, 2003, Claeys, 2002, Harbarth, 2001, Muller, 2000, Assicot, 1993).

Other acute phase protein biomarkers which are widely used in clinical practice, are ceruloplasmin - which is correlated with great liver dysfunction in septic patients- and hepcidin which is shown significantly elevated in septic patients and patients with chronic renal failure. Also, another critical biomarker of this category is serum amyloid A (SAA) which is associated with the elevated levels of CRP in septic patients. Finally, there must be a reference on lipopolysaccharide binding protein (LBP), which is generally elevated in septic patients compared to the healthy population, but -on the other hand- cannot be used as a prognostic marker in the development of sepsis. (Li et al 2009, Cicarrelì et al 2008, Schmit et al 2008, Couto et al 2007, Seller-Perez et al 2005, Oude Nijhuis et al 2003, Becker et al 2008).

5. Receptor biomarkers

Many cell receptors have been proposed to have prognostic value for septic patients. Their reliability for clinical routine use is under investigation and only some experimental findings are indicative of receptor biomarkers quality for usage in septic patient diagnosis. Receptor biomarkers that will be further discussed are IL-2 receptor, Toll-like receptor (TLR2 and TLR 4), TNF receptor and TREM-1 receptor. However, many other receptors have been investigated and proposed as possible biomarkers of sepsis, such as CSL2, Fas Receptor and group II phospholipase A2 receptor. Increased serum levels of soluble IL-2 receptor in patients with gram-negative sepsis may behave as predictive indices of shock. Soluble IL-2 receptor is released in biological fluids mostly from T and B lymphocytes that seem to participate in pathogenesis of sepsis. Soluble IL-2 receptor measurement can be easily performed by the use of diagnostic kits and it is suggested to be routinely measured in sera from patients with gram-negative sepsis in order to identify those, who have the strongest risk to develop septic shock. Microorganisms consist of conserved sequences called Pathogen-Associated Molecular Patterns (PAMPs) such as endotoxin/lipopolysaccharide (LPS) which are located in Gram- negative bacteria cell wall. These PAMPs bind to specific Pattern Recognition Receptors (PRRs) (there are 15 known subtypes), named Toll-like receptors (TLRs) and especially TLR2 and TLR4. TLRs have an

intracellular domain, homologous with IL-1 and the IL-18 receptors. Adaptor proteins facilitate binding to IL-1 receptor-associated kinase, which in turn induces TNF receptor-associated factor-6, leading to nuclear translocation of nuclear factor- κ B (NF- κ B) and ultimately to activation of cytokine gene promoters, resulting in advanced proinflammatory cytokine production (Rittisch et al, 2008, Brunn et al, 2006, Cornell et al, 2005, Delogu et al, 1995, Willatts et al, 1994). Initially in inflammatory response, LPS is bound by a lipopolysaccharide binding protein which is located to CD14 and TLR4. TLR4 and CD14 are receptors of macrophages and circulating monocytes and they are activated by LPS binding. Their activation causes signal transduction through Toll Inerleukin-1 Receptor (TIR) domain which is referred as myeloid differentiation protein 88. Afterwards, the IL-1 receptor-associated kinase (IRAK) is activated and in turn activates the TNFa-receptor associated factor (TRAF) and the TRAF-associated kinase (TAK). That is the role of TLRs in sepsis. But what is their diagnostic use as biomarkers, if there is any? TLR-2 and TLR-4 expressions are relatively increased in septic critically ill patients, as it was found in experiments using mice. Moreover, the increased levels of those two TLRs were related to advanced possibility to death.

In septic patients, soluble TNFRs plasma levels have been found increased. In the same study it was suggested that the increased levels of the receptor correlate in the same way with multiple organ failure, as with mortality. Elevated soluble TNFR levels may be used as a biomarker for severity of sepsis, in a predicting manner. But more information and clinical trials are needed in order TNFR to be used in clinical routine. Soluble TREM-1 is a reliable marker of bacterial infection, while it participates in the septic process. A decrease in soluble TREM-1 concentration may be indicative of treatment effectiveness. On the contrary, when sTREM-1 levels are persistently increased the therapy administered to patient is not effective or patients are not in a good condition. Moreover, sTREM-1 concentration is indicative of the prognosis of the patients and they can be used in predicting septic patient outcome. The receptors levels on the 28th day from diagnosis are significantly different in patients with good prognosis from those with not such a good prognosis, which may lead them to death (Rittisch et al, 2008, Annane et al, 2005, Gibot, 2005, Williams et al, 2003, McCuskey et al, 1996 Ertel, 1994).

6. Cell marker related biomarkers

Cells that bear on their membrane surface special proteins have been used in experiments in sepsis trials, in order to find some special patterns of expression of those proteins during sepsis and try to combine these findings with sepsis diagnosis and prognosis. So, CD10, CD11, CD14, CD18, CD25, CD28, CD40, CD48, CD54, CD60, CD80 and CD163 are the cell marker proteins which levels have been correlated to sepsis and septic shock prognosis. From those CD10 and CD11c are found in decreased levels in septic patients. Neutrophil CD11b and CD64 appear to be promising markers for diagnosis of early and late onset infections. CD11b is normally expressed in low concentration on the surface of neutrophils and its expression is increased 2-4 times more in infants and adults with positive blood culture for sepsis. CD11b seems to be very useful in early diagnosis of neonatal sepsis. CD14, CD25, CD28, CD40 and CD163 are significant different between septic patients with good prognosis and those with very bad prognosis in the 28th day from sepsis diagnosis. CD69 and CD48 are increased in septic patients (Giamarelos et al, 2010, Nolan et al, 2008, Kaneko et al, 2003, Mishra, 2006).

7. Vascular and endothelial damage dysfunction biomarkers

Activated macrophages in sepsis produce early response cytokines TNF- α and IL-1 which stimulate vascular endothelial cells to express Intercellular Adhesion Molecules-1 (ICAM-1), E-selectin and tissue factor. In addition, the adhesion of neutrophils to endothelium involves interaction between endothelial ICAM-1 and beta2 integrin of neutrophils. Adhesion molecules expression is increased in the most severely ill septic patients and it reaches its peak in patients with Multi Organ Failure (MOF). According to these findings, vascular cell adhesion molecule (VCAM-1), E-selectin, P-selectin and endocan are fine prognostic biomarkers in the development of multi-organ dysfunction (MODS) and septic shock in septic patients. Also, L-laminin shows a significant increase in septic patients and ADAMTS-13 is increased in septic patients without disseminated intravascular coagulopathy (DIC) compared to septic patients with DIC. Additionally, there is an existing increase in other biomarkers of this category like cellular and soluble endothelial leukocyte adhesion molecule (ELAM)-1 and vascular endothelial growth factor (VEGF) in septic patients with very poor prognosis, but further clinical studies must be demonstrated. (Drake et al 1993, Mimuro et al 2008, Seidelin et al 2002, Lopez et al 1999, Whalen et al 2000, Cowley et al, 1994, Moss et al, 1996).

8. Vasodilation related biomarkers

There are several biomarkers of sepsis related to vasodilation which have been already identified in patients. Firstly, C-type natriuretic peptide (CNP), anandamide, angiotensin converting enzyme (ACE), 2- arachidonoglycerol and neuropeptide Y are shown to be significantly increased in septic patients comparing to the healthy population, although further clinical studies must be done to ensure this finding. Also, 47 kD high molecular weight kininogen (HK) shows a critical correlation with more severe sepsis. Furthermore, proadrenomedullin and adrenomedullin are correlated with development of sepsis. But, the most critical vasodilatory factor which is correlated with development of sepsis is nitric oxide and its derivatives. In the metabolic pathway of inducible (or induced) NO synthase (Inducible Nitric Oxide Synthetase: iNOS), the inducible NO synthetase overproduces nitric oxide by reacting with L-Arginine, which results to relaxation of smooth muscle cells. Its concentration increases as a result of gene activation, which in turn produces high levels of nitric oxide (NO). NO activates guanylyl cyclase enzyme. Guanylyl cyclase increases the production of cyclic guanosine monophosphate (cGMP). However, if NO production is inhibited by blockading the NOS enzyme, then it is associated with poor prognosis in septic patients. Also, nitric oxide is being hugely produced after the activation of iNOS gene by Hypoxia Inducible Factor-1 (HIF-1). Finally, there are tetrahydrobiopterin which is shown increased in critically ill septic patients and vasoactive intestinal peptide (VIP) which is shown quite increased in tissues of patients with severe peritonitis. Elastin is shown decreased in septic patients compared to healthy population (Jacob et al 2007, Levy, et al 2005, Faury et al, 2005, Jang et al, 2004, Hama et al 1994, Beer et al 2002, Amalich et al 1995, Deitz et al, 1987).

9. Organ dysfunction biomarkers

Specific organ dysfunction biomarkers which are shown significantly increased in septic patients compared to healthy population are myocardial angiotensin II, pancreatitis-associated protein-I, pre B cell colony-enhancing factor (PBEF), glial fibrillary acidic protein

(GFAP) and Gc-globulin. Even most, Gc-globulin - which acts as an actin scavenger- has a good prognostic value in the development of multi-organ dysfunction, as it is found in low concentrations in patients developing sepsis and respiratory failure (Hsu et al 2008, Ji et al 1996, Tribl et al 2004, Ye et al 2005).

In sepsis it is highly possible that the balance between inflammatory and anti-inflammatory mechanisms becomes unbalanced so organ dysfunction and cardiovascular dysfunction, metabolic disturbances, renal dysfunction and haematological dysfunction are possible to happen. All the above clinical conditions combined are leading to multiple organ dysfunction. So, after an insult, the balance between inflammatory response and immunoparalysis determine the prognosis of the septic patient, which makes the above biomarkers of vital importance for them (Paterson et al, 2000, Poderoso et al, 1996, Neumann et al, 1997, Baudo et al, 1998, Karima et al, 1999, Grover et al, 1999,) (**figure 4**).

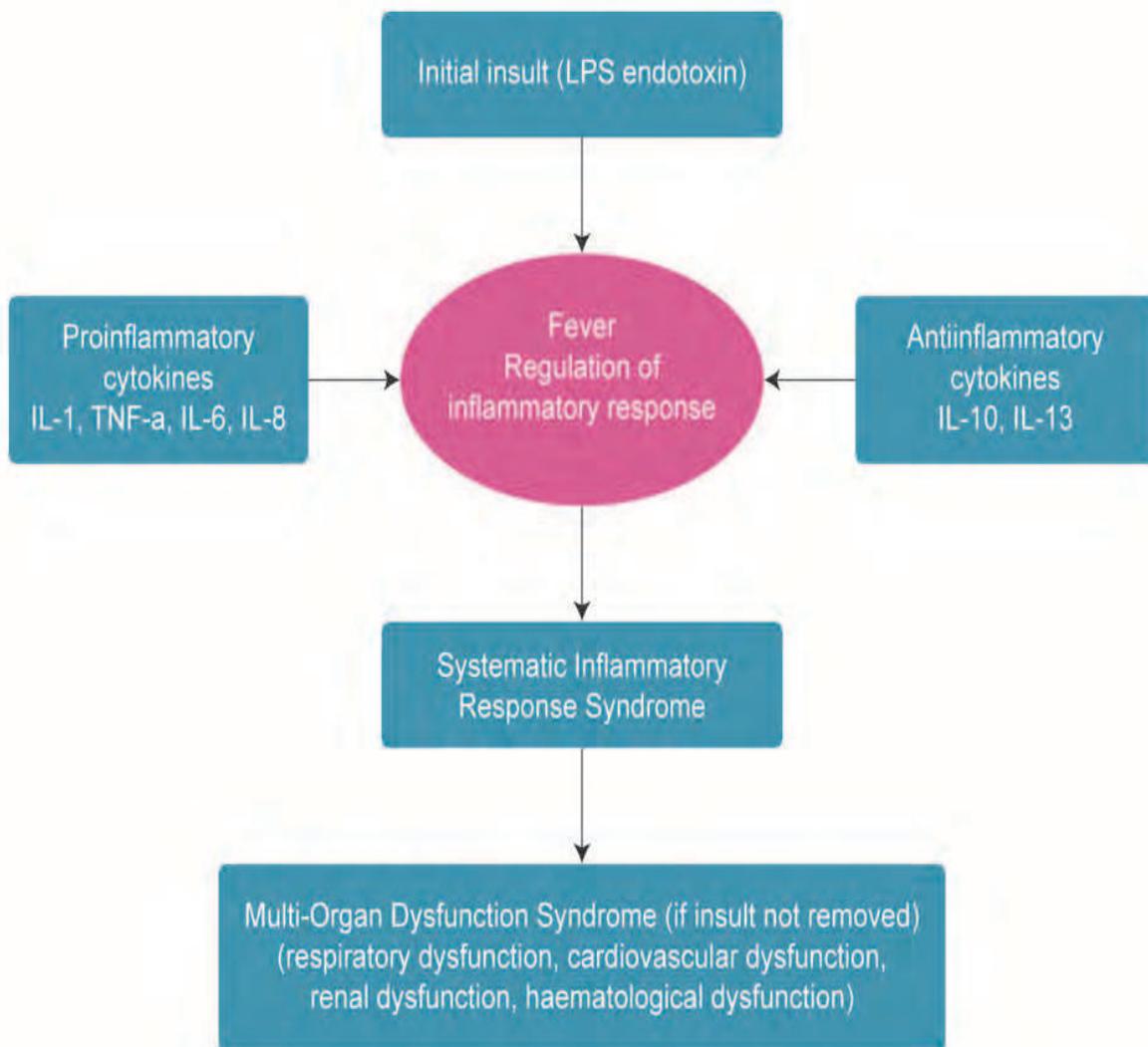


Fig. 4. The process of sepsis

10. Genetic biomarkers-single nucleotide polymorphisms

As in almost every other disease, genetic predisposition seems to play an important role in susceptibility in sepsis. Despite modern treatment of sepsis (eg, hydrocortisone) important genetic factors may be responsible even for death by sepsis. Also, along with genetic predisposition, comorbidity and age seem to play a significant role (Alberti et al 2002, Yende et al 2008, Pachot et al 2006, Wong et al 2007, Calvano et al 2005, Annane et al.1998, Bollaert et al.1998, Briegel et al.1999).

In recent years, there have been many clinical studies using modern methods and techniques of molecular genetics, such as genotyping to correlate gene expression with inflammatory response. Specific polymorphisms in some genes express proteins involved in the cascade of systematic inflammatory response and they have an important role in sepsis. These polymorphisms participating in sepsis are Single nucleotide polymorphisms (SNPs). SNPs of a gene is a DNA sequence variation which occurs whenever there is a difference in a single nucleotide position in genome and this one differs between members of a species. The simplest change of this kind is the substitution of one nucleotide by another which is not similar to the previous one. Single nucleotide polymorphisms have a minor allele frequency of >1% (or 0.5% etc.) A large number of SNPs in several genes involved in inflammatory response have been found to vary in their clinical outcomes during infection (Arcaroli et al 2005, Waterer et al 2003, Holmes et al 2003, Stuber et al 1996, Barber et al 2006). The major genetic agents that have been studied are the following:

Toll-like receptors: Polymorphisms found in TLR5 and TLR2 are correlated with bacterial infections, and those in TLR4 are correlated with susceptibility in sepsis (Sutherland et al. 2005, Skinner et al 2005, Hawn et al. 2003).

Collectins: There are collectins, such as Mannose binding lectine (MBL) whose polymorphisms associate with increased incidence of systemic inflammatory syndrome caused by infectious and noninfectious causes, increased severity of sepsis, and increased mortality although the results of some studies on this topic are contradictory (Garred et al. 2003, Roy et al. 2002, Gong et al 2007, Kronborg et al. 2002).

Tumor Necrosis Factor- α (TNF- α): TNF- α gene is polymorphic and there are various polymorphisms in its products. TNF-308 polymorphism with A-allele substitution is of high importance (the most common is G-allele) as it is correlated with increasing risk of death in septic patients. On the other hand, some studies state that there is no correlation between TNF polymorphism and sepsis (O'Keefe et al. 2002, Nakada et al. 2005, Azim et al. 2007, Schueller et al. 2006, Waterer et al. 2001).

Interleukin-1 (IL-1) Family: Polymorphisms in IL1 β may correlate with sepsis according to its cause, as in some cases they are associated with sepsis development and in some cases they are not. Also, there are conflicting studies of variants of IL-1 receptor antagonist gene (IL-1RN) and its correlation with mortality in patients with septic shock (Arnalich et al. 2002, Ma et al. 2002, Fang et al. 1999, Turner et al. 1997).

Interleukin-10 (IL-10): IL-10-1082 G polymorphic allele presence is correlated to sensitivity to pneumonia and susceptibility to septic shock as well as with multiorgan dysfunction (Stanilolva et al. 2006, Gallagher et al. 2003, Schaaf et al. 2003).

Furthermore, SNPs have also been found in genes expressing proteins participating into the inflammatory response. These genes, which SNPs may show correlation with septic shock are: *Lymphotoxin Alpha (LTA) gene*, *Immunoglobulin Receptors genes*, *Heat shock protein gene*, *Protein C gene* and *Plasminogen activation inhibitor (PAI) -1 gene*, *Lippopolysaccharide binding*

protein (LBP) gene, CD14 gene and Myeloid differentiation protein-2 (MD-2) gene (Gu et al. 2007, Temple et al. 2004, Yuan et al. 2003, Ye et al. 1995, Binder et al. 2007, Geishofer et al. 2005, Haralambous et al. 2003, Hermans et al. 1999, Van der Poll et al. 2001, Hubacek et al. 2001, Waterer et al. 2001).

As discussed in this chapter there are many biomarkers of sepsis. All of them are very promising in sepsis diagnosis and prognosis, but more evidence is needed so as to be valuable for in every day clinical routine. Sepsis severity and its high incidence to patient's death make extremely difficult every decision in using one and only one clue as the absolute biomarker for sepsis diagnosis and prognosis. Sepsis is such a complex syndrome that every oversimplified approach may become lethal for patients. Multi-biomarker control is the best approach till now. As research continues newer and more advanced biomarkers may be found helping in sepsis diagnosis and therapeutics.

11. References

- Akira S, Sato S (2003). Toll-like receptors and their signalling mechanisms. *Scand J Infect Dis*; 35:555-562.
- Alberti C, Brun-Buisson C, Burchardi H, Martin C, Goodman S, Artigas A, Sicignano A, Palazzo M, Moreno R, Boulmé R, Lepage E, Le Gall R (2002). Epidemiology of sepsis and infection in ICU patients from an international multicentre cohort study. *Intensive Care Med*; 28:108-121.
- Annane D, Bellissant E, Cavaillon J-M (2005). Septic shock. *Lancet*; 365:63-78.
- Annane D, Bellissant E, Sebille V, Lesieur O, Mathieu B, Raphael JC, Gajdos P (1998). Impaired pressor sensitivity to noradrenaline in septic shock patients with and without impaired adrenal function reserve. *Br J Clin Pharmacol*; 46:589-597.
- Arcaroli J, Fessler MB, Abraham E (2005). Genetic polymorphisms and sepsis. *Shock*; 24(4):300-312.
- Arnalich F, Sanchez JF, Martinez M, Jimenez M, Lopez J, Vazquez JJ, Hernanz A (1995). Changes in plasma concentrations of vasoactive neuropeptides in patients with sepsis and septic shock. *Life Sci*, 56:75-81.
- Arnalich F, Lopez-Maderuelo D, Codoceo R, Lopez J, Solis-Garrido LM, Capiscol C, Fernandez-Capitan C, Madero R, Montiel C (2002). Interleukin-1 receptor antagonist gene polymorphism and mortality in patients with severe sepsis. *Clin Exp Immunol*; 127:331-336.
- Assicot M, Gendrel D (1993). High serum procalcitonin concentrations in patients with sepsis and infection. *Lancet*; 341:515-518.
- Azim K, McManus R, Brophy K, Ryan A, Kelleher D, Reynolds JV (2007). Genetic polymorphisms and the risk of infection following esophagectomy. Positive association with TNF-alpha gene -308 genotype. *Ann Surg*; 246:122-128.
- Barber RC, Chang LY, Arnoldo BD, Purdue GF, Hunt JL, Horton JW, Aragaki CC (2006). Innate immunity SNPs are associated with risk for severe sepsis after burn injury. *Clin Med Res*; 4(4):250-255.
- Baudo F, Caimi TM, deCataldo F (1998). Antithrombin III (ATIII) replacement therapy in patients with sepsis and/or postsurgical complications: a controlled double-blind, randomized, multicenter study. *Intensive Care Med*; 24:336-342.
- Becker KL, Snider R, Nylén ES (2008). Procalcitonin assay in systemic inflammation, infection, and sepsis: clinical utility and limitations. *Crit Care Med*; 36:941-952.

- Beer S, Weighardt H, Emmanuilidis K, Harzenetter MD, Matevossian E, Heidecke CD, Bartels H, Siewert JR, Holzmann B (2002). Systemic neuropeptide levels as predictive indicators for lethal outcome in patients with postoperative sepsis. *Crit Care Med*; 30:1794-1798.
- Binder A, Endler G, Muller M, Mannhalter C, Zenz W (2007). 4G4G genotype of the plasminogen activator inhibitor-1 promoter polymorphism associates with disseminated intravascular coagulation in children with systemic meningococemia. *J Thromb Haemost*; 5:2049-54.
- Biomarkers Definitions Working Group (2001): Biomarkers and surrogate endpoints: Preferred definitions and conceptual framework. *Clin Pharmacol Ther*; 69:89-95.
- Bollaert PE, Charpentier C, Debouverie M, Audibert G, Larcan A (1998). Reversal of late septic shock with supraphysiologic doses of hydrocortisone. *Crit Care Med*; 26:645-650.
- Bone RC, Balk RA, Cerra FB, Dellinger RP, Fein AM, Knaus WA, Schein RM, Sibbald WJ; ACCP/SCCM Consensus Conference Committee (1992). ACCP/SCCM Consensus Conference. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Chest*; 101:1644-1655.
- Briegleb J, Forst H, Haller M, Schelling G, Kilger E, Kuprat G, Hemmer B, Hummel T, Lenhart A, Heyduck M, Stoll C, Peter K (1999). Stress doses of hydrocortisone reverse hyperdynamic septic shock: A prospective, randomized, double-blind, single-center study. *Crit Care Med*; 27:723-732.
- Brunkhorst FM, Heinz U, Forycki ZF (1998). Kinetics of procalcitonin in iatrogenic sepsis. *Intensive Care Med*; 24:888-889.
- Brunn GJ, Platt JL (2006). The aetiology of sepsis: turned inside out. *Trends Mol Med*; 12:10-16.
- Calvano SE, Xiao W, Richards DR, Felciano RM, Baker HV, Cho RJ, Chen RO, Brownstein BH, Cobb JP, Tschoeke SK, Miller-Graziano C, Moldawer LL, Mindrinos MN, Davis RW, Tompkins RG, Lowry SF; Inflamm and Host Response to Injury Large Scale Collab. Res. Program (2005). A network-based analysis of systemic inflammation in humans. *Nature*; 437:1032-1037.
- Casey LC, Balk RA, Bone RC (1993). Plasma cytokines and endotoxin levels correlate with survival in patients with the sepsis syndrome. *Ann Intern Med*; 119:771-778.
- Castelli GP, Pognani C, Meisner M, Stuardi A, Bellomi D, Sgarbi L (2004). Procalcitonin and C-reactive protein during systemic inflammatory response syndrome, sepsis and organ dysfunction. *Crit Care*; 8:R234-242.
- Chiesa C, Pellegrini G, Panero A, Osborn JF, Signore F, Assumma M, Pacifico L (2003). C-reactive protein, interleukin-6, and procalcitonin in the immediate postnatal period: influence of illness severity, risk status, antenatal and perinatal complications, and infection. *Clin Chem*; 49:60-68.
- Cicarelli DD, Vieira JE, Bensenor FE (2008). Comparison of C-reactive protein and serum amyloid a protein in septic shock patients. *Mediators Inflamm*; 2008:631414.
- Claeys R, Vinken S, Spapen H, Ver Elst K, Decochez K, Huyghens L, Gorus FK (2002). Plasma procalcitonin and C-reactive protein in acute septic shock: clinical and biological correlates. *Crit Care Med*; 30:757-762.
- Clec'h C, Ferriere F, Karoubi P (2004). Diagnostic and prognostic value of procalcitonin in patients with sepsis and septic shock. *Crit Care Med*; 32:1166-1169.

- Cornell Timothy T, Wynn James, Shanley Thomas P, Wheeler Derek S, Wong Hector R (2010). Mechanisms and Regulation of the Gene-Expression Response to Sepsis. *Pediatrics*; 125:1248.
- Couto RC, Barbosa JA, Pedrosa TM, Biscione FM (2007). C-reactive protein-guided approach may shorten length of antimicrobial treatment of cultureproven late-onset sepsis: an intervention study. *Braz J Infect Dis*; 11:240-245.
- Cowley HC, Heney D, Gearing AJ, Hemingway I, Webster NR (1994). Increased circulating adhesion molecules concentration in patients with the systemic inflammatory response syndrome: a prospective cohort study. *Crit Care Med*; 22: 651-657.
- De Werra I, Jaccard C, Corradin SB, Chioleró R, Yersin B, Gallati H, Assicot M, Bohuon C, Baumgartner JD, Glauser MP, Heumann D (1997). Cytokines, nitrite/nitrate, soluble tumor necrosis factor receptors, and procalcitonin concentrations: comparisons in patients with septic shock, cardiogenic shock, and bacterial pneumonia. *Crit Care Med*; 25:607-613.
- Deitz DM, Swartz KR, Wright M, Murphy E, Connell RS, Harrison MW (1987). Effects of E. coli endotoxin on rat plasma angiotensin converting enzyme activity in vitro and in vivo. *Circ Shock*; 21:23-29.
- Dellinger RP, Levy MM, Carlet JM, Bion J, Parker MM, Jaeschke R, Reinhart K, Angus DC, Brun-Buisson C, Beale R, Calandra T, Dhainaut JF, Gerlach H, Harvey M, Marini JJ, Marshall J, Ranieri M, Ramsay G, Sevransky J, Thompson BT, Townsend S, Vender JS, Zimmerman JL, Vincent JL (2008). Surviving Sepsis Campaign: international guidelines for management of severe sepsis and septic shock: 2008. *Crit Care Med*; 36:296-327.
- Delogu G, Casula MA, Mancini P, Tellan G, Signore L (1995). Serum neopterin and soluble interleukin-2 receptor for prediction of a shock state in gramnegative sepsis. *J Crit Care*; 10:64-71.
- Dhar R, Diringer MN (2008). The burden of the systemic inflammatory response predicts vasospasm and outcome after subarachnoid hemorrhage. *Neurocrit Care*; 8(3):404-412.
- Drake TA, Cheng J, Chang A, Taylor FB Jr (1993). Expression of tissue factor, thrombomodulin, and E-selectin in baboons with lethal Escherichia coli sepsis. *Am J Pathol*; 142:1458-1470.
- Ertel W, Scholl FA, Gallati H, Bonaccio M, Schildberg FW, Trentz O (1994). Increased release of soluble tumor necrosis factor receptors into blood during clinical sepsis. *Arch Surg*; 129:1330-1336.
- Fang XM, Schroder S, Hoefft A, Stuber F (1999). Comparison of two polymorphisms of the interleukin-1 gene family: interleukin-1 receptor antagonist polymorphism contributes to susceptibility to severe sepsis. *Crit Care Med*; 27:1330-1334.
- Faury G, Wynnychenko TM, Cand F, Leone M, Jacob MP, Verdetti J, Boyle WA (2005). Decreased circulating elastin peptide levels in humans with sepsis. *Pathol Biol (Paris)*; 53:443-447.
- Fisher CJ Jr, Yan SB (2000). Protein C levels as a prognostic indicator of outcome in sepsis and related diseases. *Crit Care Med*; 28:S49-S56.
- Freitas A, Alves-Filho JC, Victoni T, Secher T, Lemos HP, Sônego F, Cunha FQ, Ryffel B (2009). IL-17 receptor signaling is required to control polymicrobial sepsis. *J Immunol*; 182(12):7846-7854.

- Gallagher PM, Lowe G, Fitzgerald T, Bella A, Greene CM, McElvaney NG, O'Neill SJ (2003). Association of IL-10 polymorphism with severity of illness in community acquired pneumonia. *Thorax*; 58:154-156.
- Garred P, Strom J, Quist L, Taaning E, Madsen H (2003). Association of mannose-binding lectin polymorphisms with sepsis and fatal outcome, in patients with systemic inflammatory response syndrome. *J Infect Dis*; 188:1394-1403.
- Geishofer G, Binder A, Muller M, Zohrer B, Resch B, Muller W, Faber J, Finn A, Endler G, Mannhalter C, Zenz W, C. E. M. G. S. Group (2005). 4G/5G promoter polymorphism in the plasminogen-activator-inhibitor-1 gene in children with systemic meningococcaemia. *Eur J Pediatr*; 164:486-490.
- Giamarellos-Bourboulis EJ (2010). What is the pathophysiology of the septic host upon admission? *Int J Antimicrob Agents*; S2:S2-S5.
- Gibot S, Cravoisy A, Kolopp-Sarda MN, Bene MC, Faure G, Bollaert PE, Levy B (2005). Time-course of sTREM (soluble triggering receptor expressed on myeloid cells)-1, procalcitonin, and C-reactive protein plasma concentrations during sepsis. *Crit Care Med*; 33:792-796.
- Gong MN, Zhou W, Williams PL, Thompson BT, Pothier L, Christiani DC (2007). Polymorphisms in the mannose binding lectin-2 gene and acute respiratory distress syndrome. *Crit Care Med* 35:48-56.
- Grover R, Lopez A, Lorente J (1999). Multi-center, randomized, placebo-controlled, double blind study of the nitric oxide synthase inhibitor 546C88: effect on survival in patients with septic shock. *Crit Care Med*; 27: A33.
- Gu W, Shan Y, Zhou J, Jiang D, Zhang L, Du D, Wang Z, Jiang J (2007). Functional significance of gene polymorphisms in the promoter of myeloid differentiation-2. *Ann Surg*; 246:151-158.
- Hack CE, Aarden LA, Thijs LG (1997). Role of cytokines in sepsis. *Adv Immunol*; 66: 101-195.
- Halter J, Steinberg J, Fink G, Lutz C, Picone A, Maybury R, Fedors N, DiRocco J, Lee HM, Nieman G (2005). Evidence of systemic cytokine release in patients undergoing cardiopulmonary bypass. *J Extra Corpor Technol*; 37: 272-277.
- Hama N, Itoh H, Shirakami G, Suga S, Komatsu Y, Yoshimasa T, Tanaka I, Mori K, Nakao K (1994). Detection of C-type natriuretic peptide in human circulation and marked increase of plasma CNP level in septic shock patients. *Biochem Biophys Res Commun*; 198(3):1177-1182.
- Haralambous E, Hibberd ML, Hermans PW, Ninis N, Nadel S, Levin M (2003). Role of functional plasminogen-activator-inhibitor-1 4G/5G promoter polymorphism in susceptibility, severity, and outcome of meningococcal disease in Caucasian children. *Crit Care Med*; 31:2788-2793.
- Harbarth S, Holeckova K, Froidevaux C, Pittet D, Ricou B, Grau GE, Vadas L, Pugin J; Geneva Sepsis Network (2001). Diagnostic value of procalcitonin, interleukin-6, and interleukin-8 in critically ill patients admitted with suspected sepsis. *Am J Respir Crit Care Med*; 164:396-402.
- Hawn TR, Verbon A, Lettinga KD, Zhao LP, Li SS, Laws RJ, Skerrett SJ, Beutler B, Schroeder L, Nachman A, Ozinsky A, Smith KD, Aderem A (2003). A common dominant TLR5 stop codon polymorphism abolishes flagellin signaling and is associated with susceptibility to legionnaires' disease. *J Exp Med*; 198:1563-1572.

- Hermans PW, Hibberd ML, Booy R, Daramola O, Hazelzet JA, de Groot R, Levin M (1999). 4G/5G promoter polymorphism in the plasminogen-activator-inhibitor-1 gene and outcome of meningococcal disease. Meningococcal Research Group. *Lancet*; 354 (9178):556-560.
- Hildeman DA, Zhu Y, Mitchell TC, Kappler J, Marrack P (2002). Molecular mechanisms of activated T cell death in vivo. *Curr Opin Immunol*; 14:354-359.
- Holmes CL, Russell JA, Walley KR (2003). Genetic polymorphisms in sepsis and septic shock: role in prognosis and potential for therapy. *Chest*; 124(3):1103-1111.
- Hoover L, Bochicchio GV, Napolitano LM, Joshi M, Bochicchio K, Meyer W, Scalea TM (2006). Systemic inflammatory response syndrome and nosocomial infection in trauma. *J Trauma*; 61(2):310-316.
- Hotchkiss RS, Tinsley KW, Karl IE (2003). Role of apoptotic cell death in sepsis. *Scand J Infect Dis*; 35:585-592.
- Hotchkiss RS, Coopersmith CM, Karl IE (2005). Prevention of lymphocyte apoptosis--a potential treatment of sepsis? *Clin Infect Dis*; 41 Suppl 7:S465-469.
- Hotchkiss RS, Karl IE (2003). The pathophysiology and treatment of sepsis. *N Engl J Med*; 348:138-150.
- Hsu AA, Fenton K, Weinstein S, Carpenter J, Dalton H, Bell MJ (2008). Neurological injury markers in children with septic shock. *Pediatr Crit Care Med*; 9:245-251.
- Hubacek JA, Stuber F, Frohlich D, Book M, Wetegrove S, Ritter M, Rothe G, Schmitz G (2001). Gene variants of the bactericidal/permeability increasing protein and lipopolysaccharide binding protein in sepsis patients: gender-specific genetic predisposition to sepsis. *Crit Care Med*; 29:557-561.
- Iglesias J, Marik PE, Levine JS (2003). Elevated serum levels of the type I and type II receptors for tumor necrosis factor-alpha as predictive factors for ARF in patients with septic shock. *Am J Kidney Dis*; 41:62-65.
- Jacob P, Mueller MH, Hahn J, Wolk I, Mayer P, Nagele U, Hennenlotter J, Stenzl A, Konigsrainer A, Glatzle J (2007). Alterations of neuropeptides in the human gut during peritonitis. *Langenbecks Arch Surg*; 392:267-271.
- Jaffer U, Wade RG, Gourlay T (2010). Cytokines in the systemic inflammatory response syndrome: a review. Hammersmith Hospital, London, UK; University of East Anglia, Norwich, Norfolk, UK; University of Strathclyde, Glasgow, Scotland. *HSR Proceedings in Intensive Care and Cardiovascular Anesthesia*; 2:161-175.
- Jensen JU, Heslet L, Jensen TH, Espersen K, Steffensen P, Tvede M (2006). Procalcitonin increase in early identification of critically ill patients at high risk of mortality. *Crit Care Med*; 34:2596-2602.
- Ji Y, Ren X, Zhao Y, Dong L, Wu L, Su J (1996). Role of intracardiac angiotensin II in cardiac dysfunction of rat during septic shock. *Chin Med J (Engl)*; 109:864-867.
- Jiang W, Jiang HF, Cai DY, Pan CS, Qi YF, Pang YZ, Tang CS (2004). Relationship between contents of adrenomedullin and distributions of neutral endopeptidase in blood and tissues of rats in septic shock. *Regul Pept*; 118:199-208.
- Joshi VD, Kalvakolanu DV, Cross AS (2003). Simultaneous activation of apoptosis and inflammation in pathogenesis of septic shock: a hypothesis. *FEBS Letters*; 555:180-184.

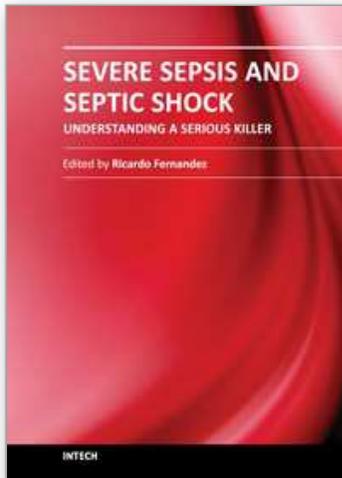
- Kalil AC, Coyle SM, Um JY, LaRosa SP, Turlo MA, Calvano SE, Sundin DP, Nelson DR, Lowry SF (2004). Effects of drotrecogin alfa (activated) in human endotoxemia. *Shock*; 21:222-229.
- Kan H, Xie Z, Finkel MS (1999). TNF-alpha enhances cardiac myocyte NO production through MAP kinasemediated NF-kappaB activation. *Am J Physiol*; 277:1641-1646.
- Kaneko T, Stearns-Kurosawa DJ, Taylor F Jr, Twigg M, Osaki K, Kinasewitz GT, Peer G, Kurosawa S (2003). Reduced neutrophil CD10 expression in nonhuman primates and humans after in vivo challenge with E. coli or lipopolysaccharide. *Shock*; 20:130-137.
- Karima R, Matsumoto S, Hidemitsu H, Matsushima K (1999). The molecular pathogenesis of endotoxic shock and organ failure. *Mol Med Today*; 98: 123-133.
- Kronborg G, Weis N, Madsen H, Pedersen S, Wejse C, Nielsen H, Skinhøj P, Garred P (2002). Variant mannose- binding lectin alleles are not associated with susceptibility to or outcome of invasive pneumococcal infection in randomly included patients. *J Infect Dis*; 185:1517-1520.
- Lever A, Mackenzie I (2007). Sepsis: definition, epidemiology, and diagnosis. *BMJ*; 335:879-883.
- Levy MM, Fink MP, Marshall JC, Abraham E, Angus D, Cook D, Cohen J, Opal SM, Vincent JL, Ramsay G, SCCM/ESICM/ACCP/ATS/SIS (2003). "2001 SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference". *Crit. Care Med.* 31 (4): 1250-1256.
- Levy RM, Prince JM, Billiar TR (2005). Nitric oxide: a clinical primer. *Crit Care Med*; 33:S492-S495.
- Li H, Rose MJ, Tran L, Zhang J, Miranda LP, James CA, Sasu BJ (2009). Development of a method for the sensitive and quantitative determination of hepcidin in human serum using LC-MS/MS. *J Pharmacol Toxicol Methods*; 59(3):171-180.
- Lopez S, Prats N, Marco AJ (1999). Expression of E-selectin, P-selectin, and intercellular adhesion molecule-1 during experimental murine listeriosis. *Am J Pathol*; 155:1391-1397.
- Luzzani A, Polati E, Dorizzi R, Rungatscher A, Pavan R, Merlini A (2003). Comparison of procalcitonin and C-reactive protein as markers of sepsis. *Crit Care Med*; 31:1737-1741.
- Ma P, Chen D, Pan J, Du B (2002). Genomic polymorphism within interleukin-1 family cytokines influences the outcome of septic patients. *Zhonghua Yi Xue Za Zhi*; 82:1237-1241.
- Madoiwa S, Nunomiya S, Ono T, Shintani Y, Ohmori T, Mimuro J, Sakata Y (2006). Plasminogen activator inhibitor 1 promotes a poor prognosis in sepsis-induced disseminated intravascular coagulation. *Int J Hematol*; 84:398-405.
- Marshall JC, Reinhart K (2009). Biomarkers of sepsis. *Crit Care Med*; 37:2290-2298.
- Martin C, Boisson C, Haccoun M, Thomachot L, Mege JL (1997). Patterns of cytokine evolution (tumor necrosis factor-alpha and interleukin-6) after septic shock, hemorrhagic shock, and severe trauma. *Crit Care Med*; 25:1813-1819.
- Mather T, Oganessyan V, Hof P, Huber R, Foundling S, Esmon C, Bode W -December (1996). "The 2.8 Å crystal structure of Gla-domainless activated protein C". *EMBO J.* 15 (24): 6822-6831.
- Matzinger P (2002). The danger model: A renewed sense of self. *Science*; 96:301-305.

- McCuskey RS, Urbaschek R, Urbaschek B (1996). The microcirculation during endotoxemia. *Cardiol Res*; 32:752-763.
- McDonald TE, Grinman MN, Carthy CM, Walley KR (2000). Endotoxin infusion in rats induces apoptotic and survival pathways in hearts. *Am J Physiol Heart Circ Physiol*; 79:H2053- H2061.
- Mimuro J, Niimura M, Kashiwakura Y, Ishiwata A, Ono T, Ohmori T, Madoiwa S, Okada K, Matsuo O, Sakata Y (2008). Unbalanced expression of ADAMTS13 and von Willebrand factor in mouse endotoxemia. *Thromb Res*; 122:91-97.
- Mishra UK, Jacobs SE, Doyle LW, Garland SM (2006). Newer approaches to the diagnosis of early onset neonatal sepsis. *Arch Dis Child Fetal Neonatal*; 91(3):F208-212.
- Mokart D, Capo C, Blache JL, Delpero JR, Houvenaeghel G, Martin C, Mege JL (2002). Early postoperative compensatory anti-inflammatory response syndrome is associated with septic complications after major surgical trauma in patients with cancer. *Br J Surg*; 89: 1450-1456.
- Mori K, Nakao K (1994). Detection of C-type natriuretic peptide in human circulation and marked increase of plasma CNP level in septic shock patients. *Biochem Biophys Res Commun*; 198:1177-1182.
- Mosnier LO, Zlokovic BV, Griffin JH (2007). The cytoprotective protein C pathway". *Blood*; 109 (8): 3161-3172.
- Moss M, Gillespie MK, Ackerson L, Moore FA, Moore EE, Parsons PE (1996). Endothelial cell activity varies in patients at risk for the adult respiratory distress syndrome. *Crit Care Med*; 24: 1782-1786.
- Muller B, Becker KL, Schachinger H, Rickenbacher PR, Huber PR, Zimmerli W, Ritz R (2000). Calcitonin precursors are reliable markers of sepsis in a medical intensive care unit. *Crit Care Med*; 28:977-983.
- Munford RS, Pugin J (2001). Normal responses to injury prevent systemic inflammation and can be immunosuppressive. *Am J Respir Crit Care Med*; 163:316-21.
- Nakada TA, Hirasawa H, Oda S, Shiga H, Matsuda KI, Nakamura M, Watanabe E, Abe R, Hatano M, Tokuhisa T (2005). Influence of toll-like receptor 4, CD14, tumor necrosis factor, and interleukine-10 gene polymorphisms on clinical outcome in Japanese critically ill patients. *J Surg Res*; 129:322-328.
- Neumann F, Ott I, Marx N, Marx N, Luther T, Kenngott S, Gawaz M, Kotzsch M, Schömig A (1997). Effect of human recombinant interleukin-6 and interleukin-8 on monocyte procoagulant activity. *Arterioscler Thromb Vasc Biol*;17:3399-3405
- Nicolaes GA, Dahlbäck B (2003). Congenital and acquired activated protein C resistance. *Semin Vasc Med*; 3 (1): 33-46.
- Nolan A, Weiden M, Kelly A, Hoshino Y, Hoshino S, Mehta N, Gold JA (2008). CD40 and CD80/86 act synergistically to regulate inflammation and mortality in polymicrobial sepsis. *Am J Respir Crit Care Med*; 177:301-308.
- O'Keefe GE, Hybki DL, Munford RS (2002). The G→A single nucleotide polymorphism at the -308 position in the tumor necrosis factor- α promoter increases the risk for severe sepsis after trauma. *J Trauma*; 52:817-825. discussion 825-826.
- O'Neill S, O'Neill AJ, Conroy E, Brady HR, Fitzpatrick JM, Watson RW (2000). Altered caspase expression results in delayed neutrophil apoptosis in acute pancreatitis. *J Leukoc Biol*; 68:15-20.

- Okazaki Y, Matsukawa A (2009). Pathophysiology of sepsis and recent patents on the diagnosis, treatment and prophylaxis for sepsis. *Recent Pat Inflamm Allergy Drug Discov*; 3(1):26-32.
- Oude Nijhuis CS, Vellenga E, Daenen SM, Graaf van der WT, Gietema JA, Groen HJ, Kamps WA, De Bont ES (2003). Lipopolysaccharide-binding protein: a possible diagnostic marker for Gram-negative bacteremia in neutropenic cancer patients. *Intensive Care Med*; 29:2157-2161.
- Pachot A, Lepape A, Vey S, Bienvenu J, Mouglin B, Monneret G (2006). Systemic transcriptional analysis in survivor and nonsurvivor septic shock patients: A preliminary study. *Immunol Lett*; 106:63-71.
- Panacek EA, Kaul M (1999). IL-6 as a marker of excessive TNF- α activity in sepsis. *Sepsis*; 3:65-73.
- Parrillo J (1993). Pathogenetic Mechanisms of Septic Shock. *N Engl J Med* 1993; 328:1471-1477.
- Paterson RL, Webster NR (2000). Sepsis and the systemic inflammatory response syndrome. *J R Coll Surg Edinb*; 45(3):178-182.
- Pettila V, Pentti J, Pettila M, Takkunen O, Jousela I (2002). Predictive value of antithrombin III and serum C-reactive protein concentration in critically ill patients with suspected sepsis. *Crit Care Med*; 30:271-275.
- Pezzilli R, Zerbi A, Di Carlo V, Bassi C, Delle Fave GF (2010). Working Group of the Italian Association for the Study of the Pancreas on Acute Pancreatitis. Practical guidelines for acute pancreatitis. *Pancreatology*; 10(5):523-535.
- Pierrakos C, Vincent JL (2010). Sepsis biomarkers: a review. *Crit Care*; 14(1):R15.
- Pinsky MR, Vincent JL, Deviere J, Alegre M, Kahn RJ, Dupont E (1993). Serum cytokine levels in human septic shock. Relation to multiple-system organ failure and mortality. *Chest*; 103:565-575.
- Poderoso JJ, Carreras MC, Lisdero C (1996). Nitric oxide inhibits electron transfer and increases superoxide radical production in rat mitochondria and submitochondrial particles. *Arch Biochem Biophys*; 328: 85-92.
- Povoa P, Coelho L, Almeida E, Fernandes A, Mealha R, Moreira P, Sabino H (2004). C-reactive protein as a marker of infection in critically ill patients. *Clin Microbiol Infect*; 11:101-108.
- Remick DG (2007). Pathophysiology of sepsis. *Am J Pathol*; 170(5):1435-44
- Rittisch D, Flierl MA, Ward PA (2008). Harmful molecular mechanisms in sepsis. *Nat Immunol*; 8:776-786.
- Rodríguez-Gaspar M, Santolaria F, Jarque-Lopez A, González-Reimers E, Milena A, de la Vega MJ, Rodríguez-Rodríguez E, Gómez-Sirvent JL (2001). Prognostic value of cytokines in SIRS general medical patients. *Cytokine*; 15:232-236.
- Roy S, Knox K, Segal S, Griffiths D, Moore C, Welsh K, González-Reimers E, Milena A, de la Vega MJ, Rodríguez-Rodríguez E, Gómez-Sirvent JL (2002). MBL genotype and risk of invasive pneumococcal disease: a case-control study. *Lancet*; 359(9317):1569-1573.
- Schaaf BM, Boehmke F, Esnaashari H, Seitzer U, Kothe H, Maass M, Zabel P, Dalhoff K (2003). Pneumococcal septic shock is associated with the interleukin-10-1082 gene promoter polymorphism. *Am J Respir Crit Care Med*; 168:476-480.

- Schmit X, Vincent JL (2008). The time course of blood C-reactive protein concentrations in relation to the response to initial antimicrobial therapy in patients with sepsis. *Infection*; 36:213-219.
- Schueller AC, Heep A, Kattner E, Kroll M, Wisbauer M, Sander J, Bartmann P, Stuber F (2006). Prevalence of two tumor necrosis factor gene polymorphisms in premature infants with early onset sepsis. *Biol Neonate*; 90:229-232.
- Seidelin JB, Nielsen OH, Strom J (2002). Soluble L-selectin levels predict survival in sepsis. *Intensive Care Med*; 28:1613-1618.
- Seller-Perez G, Herrera-Gutierrez ME, Lebron-Gallardo M, Toro-Peinado I, Martin-Hita L, Porras-Ballesteros JA (2005): [Serum C-reactive protein as a marker of outcome and infection in critical care patients]. *Med Clin (Barc)*; 125:761-765.
- Skinner NA, MacIsaac CM, Hamilton JA, Visvanathan K (2005). Regulation of toll-like receptor (TLR)2 and TLR4 on CD14dimCD16+ monocytes in response to sepsis-related antigens. *Clin Exp Immunol*; 141:270-278.
- Sobieski MA, Graham JD, Pappas PS, Tatoes AJ, Slaughter MS (2008). Reducing the effects of the systemic inflammatory response to cardiopulmonary bypass: can single dose steroids blunt systemic inflammatory response syndrome? *ASAIO J*; 54:203-206.
- Socha LA, Gowardman J, Silva D, Correcha M, Petrosky N (2006). Elevation in interleukin 13 levels in patients diagnosed with systemic inflammatory response syndrome. *Intensive Care Med*; 32:244-250.
- Sohn SJ, Rajpal A, Winoto A (2003). Apoptosis during lymphoid development. *Curr Opin Immunol*; 15:209-216.
- Spriggs DR, Sherman ML, Frei E 3rd, Kufe DW. (1987). Clinical studies with tumour necrosis factor. *Ciba Found Symp*; 131:206-227.
- Stanilova SA, Miteva LD, Karakolev ZT, Stefanov CS (2006). Interleukin-10-1082 promoter polymorphism in association with cytokine production and sepsis susceptibility. *Intensive Care Med*; 32:260-266.
- Stuber F, Petersen M, Bokelmann F, Schade U (1996). A genomic polymorphism within the tumor necrosis factor locus influences plasma tumor necrosis factor-alpha concentrations and outcome of patients with severe sepsis. *Crit Care Med*; 24:381-384.
- Sutherland AM, Russell JA (2005). Issues with polymorphism analysis in sepsis. *Clin Infect Dis*; 41(7 Suppl):S396-S402.
- Temple SE, Cheong KY, Ardlie KG, Sayer D, Waterer GW (2004). The septic shock associated HSPA1B1267 polymorphism influences production of HSPA1A and HSPA1B. *Intensive Care Med*; 30:1761-1767.
- Tribl B, Filipp D, Bodeker H, Yu P, Hammerrmuller I, McKerlie C, Keim V, Sibbald WJ (2004). Pseudomonas pneumonia-mediated sepsis induces expression of pancreatitis-associated protein-I in rat pancreas. *Pancreas*; 29:33-40.
- Turner DM, Williams DM, Sankaran D, Lazarus M, Sinnott PJ, Hutchinson IV (1997). An investigation of polymorphism in the interleukin-10 gene promoter. *Eur J Immunogenet*; 24:1-8.
- Van der Pol WL, Huizinga TW, Vidarsson G, Van der Linden MW, Jansen MD, Keijsers V, de Straat FG, Westerdal NA, de Winkel JG, Westendorp RG (2001). Relevance of Fc gamma receptor and interleukin-10 polymorphisms for meningococcal disease. *J Infect Dis*; 184:1548-1555.

- Van der Poll T, Opal SM (2008). Host-pathogen interactions in sepsis. *Lancet Infect Dis*; 8:32-43.
- Waterer GW, ElBahlawan L, Quasney MW, Zhang Q, Kessler LA, Wunderink RG (2003). Heat shock protein 70-2+1267 AA homozygotes have an increased risk of septic shock in adults with community-acquired pneumonia. *Crit Care Med*; 31:1367-1372.
- Waterer GW, Quasney MW, Cantor RM, Wunderink RG (2001). Septic shock and respiratory failure in community-acquired pneumonia have different TNF polymorphism associations. *Am J Respir Crit Care Med*; 163:1599-1604.
- Whalen MJ, Doughty LA, Carlos TM, Wisniewski SR, Kochanek PM, Carcillo JA (2000). Intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 are increased in the plasma of children with sepsis-induced multiple organ failure. *Crit Care Med*; 28:2600-2607.
- Whitlock RP, Young E, Noora J, Farrokhyar F, Blackall M, Teoh KH (2006). Pulse low dose steroids attenuate post-cardiopulmonary bypass SIRS; *SIRS I J Surg Res*; 132:188-194.
- Willatts SM, Speller DC, Winter RJ (1994). Incidence of gram-negative bacteraemia in sepsis syndrome. *Implications for immunotherapy. Anaesthesia*; 49:751-754.
- Williams DL, Ha T, Li C, Kalbfleisch JH, Schweitzer J, Vogt W, Browder IW (2003). Modulation of tissue Toll-like receptor 2 and 4 during the early phases of polymicrobial sepsis correlates with mortality. *Crit Care Med*; 31:1808-1818.
- Wong HR, Shanley TP, Sakthivel B, Lin R, Allen GL, Thomas NJ, Doctor A, Kalyanaraman M, Tofil NM, Penfil S, Monaco M, Tagavilla MA, Odoms K, Dunsmore K, Barnes M, Aronow BJ; Genomics of Pediatric SIRS/Septic Shock Investigators (2007). Genome-level expression profiles in pediatric septic shock indicate a role for altered zinc homeostasis in poor outcome. *Physiol Genomics*; 30:146-155.
- Ye S, Green FR, Scarabin PY, Nicaud V, Bara L, Dawson SJ, Humphries SE, Evans A, Luc G, Cambou JP, et al (1995). The 4G/5G genetic polymorphism in the promoter of the plasminogen activator inhibitor-1 (PAI-1) gene is associated with differences in plasma PAI-1 activity but not with risk of myocardial infarction in the ECTIM study. *Etude Cas-Temoins de l'infarctus du Myocarde. Thromb Haemost*; 74:837-841.
- Ye SQ, Simon BA, Maloney JP, Zambelli-Weiner A, Gao L, Grant A, Easley RB, McVerry BJ, Tudor RM, Standiford T, Brower RG, Barnes KC, Garcia JG (2005). Pre-Bcell colony-enhancing factor as a potential novel biomarker in acute lung injury. *Am J Respir Crit Care Med*; 171:361-370.
- Yende S, D'Angelo G, Kellum JA, Weissfeld L, Fine J, Welch RD, Kong L, Carter M, Angus DC, GenIMS Investigators (2008). Inflammatory markers at hospital discharge predict subsequent mortality after pneumonia and sepsis. *Am J Respir Crit Care Med*; 177:1242-1247.
- Yuan FF, Wong M, Pererva N, Keating J, Davis AR, Bryant JA, and Sullivan JS (2003). FcγRIIA polymorphisms in Streptococcus pneumoniae infection. *Immunol Cell Biol* 81:192-195.
- Zakariah AN, Cozzi SM, Van Nuffelen M, Clausi CM, Pradier O, Vincent JL (2008). Combination of biphasic transmittance waveform with blood procalcitonin levels for diagnosis of sepsis in acutely ill patients. *Crit Care Med*; 36:1507-1512.
- Zambon M, Ceola M, Almeida-de-Castro R, Gullo A, Vincent JL (2008). Implementation of the Surviving Sepsis Campaign guidelines for severe sepsis and septic shock: we could go faster. *J Crit Care*; 23:455-460.



Severe Sepsis and Septic Shock - Understanding a Serious Killer

Edited by Dr Ricardo Fernandez

ISBN 978-953-307-950-9

Hard cover, 436 pages

Publisher InTech

Published online 10, February, 2012

Published in print edition February, 2012

Despite recent advances in the management of severe sepsis and septic shock, this condition continues to be the leading cause of death worldwide. Some experts usually consider sepsis as one of the most challenging syndromes because of its multiple presentations and the variety of its complications. Various investigators from all over the world got their chance in this book to provide important information regarding this deadly disease. We hope that the efforts of these investigators will result in a useful way to continue with intense work and interest for the care of our patients.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

D. Paramythiotis, E. Digkas and G. Dryllis (2012). Biomarkers and Physiological Agents in Severe Sepsis and Septic Shock, *Severe Sepsis and Septic Shock - Understanding a Serious Killer*, Dr Ricardo Fernandez (Ed.), ISBN: 978-953-307-950-9, InTech, Available from: <http://www.intechopen.com/books/severe-sepsis-and-septic-shock-understanding-a-serious-killer/biomarkers-and-physiological-agents-in-severe-sepsis-and-septic-shock>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2012 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen