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The Special Implication of Sex Hormones on Dendritic Cells During Pregnancy

Sabine E. Segerer and Ulrike Kämmerer University of Würzburg, Department of Obstetrics and Gynaecology, Würzburg, Germany

1. Introduction

Pregnancy bears a great challenge to the immune system: simultaneously, immune cells have to protect the reproductive tract against imminent infections, while the developing conceptus has to be tolerated. To face this problem, a distinct composition of immune cells has to be present in the decidualized endometrium (Gomez-Lopez, Guilbert *et al.*, 2010). The predominant subsets amongst those represent uterine natural killer cells (uNK cells) and cells of the monocyte/macrophage lineage like monocytes, macrophages and dendritic cells (DC) (Loke and King, 1995). Among the latter ones, DC as antigen-presenting cells (APC) are forming an important subgroup (Steinman, 2003).

DC represent a highly adaptive cell type, which can either be transformed into an immunostimulatory phenotype after exposure to inflammatory or infectious signals or into a tolerogenic phenotype preventing T cell activation when located in an adequate antiinflammatory microenvironment. Establishing contact with invading microorganisms or cells, DC acquire their antigens and process them into antigenic peptides in the context of MHC class I and class II molecules as ligands for antigen-specific T cell receptors. This action is accompanied by migration of DC to secondary lymphoid organs. Here, the processed antigens are presented together with co-stimulatory molecules like CD40, CD80 and CD86 as well as CD83 and MHC molecules (Inaba, Metlay *et al.*, 1990;Steinman, 2003) to select and activate naive T cells (Sallusto and Lanzavecchia, 1999).

Apart from antigens, DC differentiation and maturation is considerably influenced by cytokines, hormones and other soluble factors. Lacking infectious/danger- signals or localiezed in a distinct anti-inflammatory micromilieu (e.g. TGF beta, IL-10), a specific phenotype of DC is generated which prevents T cell activation and which is supposed to protect the semi-allogenic fetus (Rutella, Bonanno *et al.*, 2006).

In human endometrium only a small number of fully matured DC are detected (Rieger, Honig *et al.*, 2004). Thereby, the amount of mature, CD83+ DC is generally low in both pregnant and non-pregnant endometrium with a slight peak in late secretory phase endometrium. During pregnancy, a distinct DC subtype expressing CD14, CD68, HLA-DR and DC-SIGN is found in increased levels in decidua (Bonifaz, Bonnyay *et al.*, 2002;Kammerer, Eggert *et al.*, 2003). These DC, presumably an intertype between immature DC (iDC) and macrophages, are supposed to represent a "pro-fetal" tolerogenic population which is able to induce a TH2 pre-dominant state (Miyazaki, Tsuda *et al.*, 2003) and to suppress the activation of T cells (Kammerer, Eggert *et al.*, 2003).

There is evidence that female sex hormones contribute to the modulation of decidual immune cells into tolerogenic subtypes (for review see: (Kyurkchiev, Ivanova-Todorova *et al.*, 2010) and especially can impact on DC (Segerer, Muller *et al.*, 2009). Thereby, the "micromilieu" determines if the captured antigens are presented in a tolerogenic or immune activating way resulting also in different phenotypes of DC. In the following, we will highlight the impact of the characteristic pregnancy hormones on DC and their potent implication on the development of a tolerogenic subtype of DC.

2. hCG

Human chorionic gonadotropin (hCG) is a heterodimeric hormone of the glycoprotein hormone family and is composed by two subunits linked in a non-covalent way. While the alpha subunit does not differ from the other glycoprotein hormones, the beta subunit of hCG represents the biggest one of the glycoprotein hormones holding a large glycosylated domain which refers to its high stability. hCG represents the very early hormonal factor of pregnancy already produced by the trophoblast layer of the blastocyst before implantation (Bonduelle, Dodd *et al.*, 1988;Lopata and Hay, 1989) and after implantation by the syncytiotrophoblast in increasing amounts (Hoshina, Boothby *et al.*, 1985). The production of hCG reaches a peak between the 10th and 11th week of gestation and thereafter declines to a lower but constant level throughout pregnancy. Preventing luteolysis of the corpus luteum and stimulating the production of progesterone (Keay, Vatish *et al.*, 2004), hCG represents the essential factor to protect and to promote early pregnancy. Despite of these direct effects, hCG can also act as a paracrine factor modulating the proliferation of myometrial smooth muscle cells (Horiuchi, Nikaido *et al.*, 2000;Kornyei, Lei *et al.*, 1993) and increasing endometrial angiogenesis (Berndt, Blacher *et al.*, 2009).

There is also evidence that hCG contributes to maternal tolerance of the developing conceptus by influencing the surrounding immune cells. Thus, experiments on uNK cells revealed that hCG acts as a regulator of uNK cell proliferation (Kane, Kelly *et al.*, 2009). Despite of this direct effect on uNK cell proliferation, hCG can facilitate the adequate establishment and maintenance of pregnancy by inducing the secretion factors which promote angiogeneis by uNK cells and thus could even support placentation (Lash, Schiessl *et al.*, 2006).

Investigations on DC demonstrated hCG receptors to be constitutively expressed by these cells which allows a direct activation of DC via hCG (Yoshimura, Inaba *et al.*, 2003). Experiments on mouse DC revealed that hCG acts in an immunregulatory way by increasing the production of immunosuppressive factors like Interleukin-10 (IL-10) by DC and by reducing antigen-specific T-cell proliferation (Wan, Versnel *et al.*, 2008). In humans, hCG was also able to significantly decrease T-cell stimulatory capacity of DC (Huck, Steck *et al.*, 2005). In contrast, the phenotype of hCG-treated DC ressembeled that of matured DC expressing co-stimulatory molecules like CD 40, CD83 and CD 86 (Segerer, Muller *et al.*, 2009). Thus, hCG seems to be able to induce a tolerogenic subtype of DC even though co-stimulatory molecules of the mature subtype were expressed.

3. Estrogen

Estrogen receptors (ER) alpha and beta (ER α/β) are widely expressed in human endometrium but also in most immune cells (Lindzey, Wetsel *et al.*, 1998). Thereby, they are localized in different cellular compartments. The cytoplasmatic and membrane associated

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receptors confer to specific intracellular signaling pathways promoting genomic and nongenomic effects. Gene transcription and regulation is also mediated by nuclear ER acting as transcription factors and thus regulating long-term effects (Tamrazi, Carlson *et al.*, 2002;Biswas, Singh *et al.*, 2005). Recently, another estrogen receptor, GPR30, which represents an intracellular transmembrane G protein-coupled receptor was detected (Revankar, Cimino *et al.*, 2005). This receptor is proposed to initiate rapid non-genomic signaling effects and was found to be expressed in human endometrium throughout menstrual cycle and in early pregnancy decidua (Kolkova, Noskova *et al.*, 2010).

B and T lymphocytes were detected to be specific targets of estradiol (Peeva and Zouali, 2005;Nalbandian, Paharkova-Vatchkova *et al.*, 2005;Nalbandian and Kovats, 2005;Smithson, Couse *et al.*, 1998). Thereby, estrogen was able to modulate B lymphopoiesis and the production of immunoglobulins (Ig) where effects were mediated via ERa (Erlandsson, Jonsson *et al.*, 2003). T cell function was found to be modulated by estradiol changing the cytokine profile (Karpuzoglu and Zouali, 2011;Pernis, 2007;Salem, 2004).

Regarding DC, estradiol effects were mediated via ER α and β , both of which are expressed in DC. So far, the intracellular G-protein receptor GPR 30 has not been described in DC. Even though estradiol promotes the differentiation of DC (Paharkova-Vatchkova, Maldonado *et al.*, 2004;Segerer, Muller *et al.*, 2009) towards an immunostimulatory phenotype, expressing co-stimulatory molecules like CD40, CD83 and CD86, T-cell priming was significantly impaired in the presence of estradiol (Segerer, Muller *et al.*, 2009). In addition, investigations of the effects of E2 on murine spleen CD11c-positive dendritic cells revealed an increased stimulatory capacity of DCs and an elevated expression of the antiinflammatory cytokines like IL-10 (Yang, Hu *et al.*, 2006).

In autoimmune diseases, estrogens seem to have conflicting effects on immune cells. While a reduction of disease activity was seen during pregnancy in multiple sclerosis, it was reported that systemic lupus erythematodes (SLE) could frequently flare up during pregnancy and remit with menopause (Petri, Howard *et al.*, 1991).

It could be speculated, that the effect of estrogen on DC participates in the flare up of SLE during pregnancy: DC generated from monocytes which were isolated from patients suffering from SLE exhibited a matured, pro-inflammatory phenotype, expressing co-stimulatory molecules. In addition, these SLE-DC were very effective in activating T-cells (Ding, Mehta *et al.*, 2006). Perhaps, estrogen could even accelerate the maturation-process of SLE-DC and thus promote disease during ongoing pregnancy.

4. Progesterone

During the menstrual cycle, progesterone is produced by granulosa cells and the corpus luteum (Bachelot and Binart, 2005). In pregnancy, the corpus luteum is rescued 4-5 weeks after implantation. At that time, placental progesterone production becames sufficient to maintain pregnancy (Csapo and Pulkkinen, 1978). Several studies revealed that progesterone acts in an immunosuppressive way (Stites and Siiteri, 1983;Miyaura and Iwata, 2002b). Analysing the effects of progesterone on T lymphocytes, a direct and indirect inhibition of TH1 cell development was detected (Miyaura and Iwata, 2002a). DC cultured under the influence of progesterone changed their phenotype into an immunostimulatory maturated phenotype expressing co-stimulatory molecules (Ivanova, Kyurkchiev *et al.*, 2005;Segerer, Muller *et al.*, 2009). In contrast to effects seen for hCG and estradiol, the capacity of DC to stimulate T-cell proliferation was not significantly altered (Segerer, Muller *et al.*, 2009). However, progesterone had profound effects on rat mDC by suppressing the

production of pro-inflammatory cytokines like tumor necrosis factor alpha (TNF α) and interleukin 1beta (IL-1 β) as well as by inhibiting the DC-stimulated proliferation of T-cells (Butts, Shukair *et al.*, 2007). Thus, even though the expression of co-stimulatory molecules and T-cell stimulatory capacity was not affected progesterone seems to modulate immune responses by changing the cytokine secretion prolife.

5. Activin A and inhibin A

Activin A represents a distinct growth factor and member of the TGF beta superfamily (Chang, Brown *et al.*, 2002) which is composed by two activin β_A subunits. Its counterpart inhibin A is formed by a dimer consisting of an activin β subunit and a structurally different α subunit (Phillips, Jones *et al.*, 2005). Initially, the function of these glycoprotein hormones was defined as feedback factors regulating the release of the follicle-stimulating hormone (FSH) (Tong, Wallace *et al.*, 2003). Later, it was observed that activin A and inhibin A are endowed by diverse functions acting as cytokines in an autocrine or paracrine manner, too. Thus, activin A was detected to be involved in inflammatory responses (Jones, de Kretser *et al.*, 2004) and acting as a chemoattractant for monocytes (Eramaa, Hurme *et al.*, 1992;Shao, Frigon, Jr. *et al.*, 1992).

During pregnancy, activin A and inhibin A are locally produced by human placenta, decidua and fetal membranes resulting in serum levels in the low ng/ml range peaking at 10 weeks of gestation (Lockwood, Ledger *et al.*, 1997). Both glycoprotein hormones seem to have some impact on implantation and early embryo development (Birdsall, Ledger *et al.*, 1997;Muttukrishna, Jauniaux *et al.*, 2004;Jones, Salamonsen *et al.*, 2002). Furthermore, they have been found to be useful diagnostic markers for pregnancy success. While decreasing activin A levels indicated both an ongoing miscarriage or ectopic pregnancy (Florio, Severi *et al.*, 2007), decreasing inhibin A levels were used as a specific marker to reveal preclinical abortions (Muttukrishna, Jauniaux *et al.*, 2004;Prakash, Laird *et al.*, 2005).

Previous studies revealed that activin A is able to modulate immune responses via type I and II activin receptors on DC (Robson, Phillips *et al.*, 2008). Even though the expression of co-stimulatory molecules was not significantly affected neither by activin A nor inhibin A, both glycoprotein hormones were able to decrease the T-cell stimulatory capacity of DC. Thus, we propose that activin A and inhibin A are distinct modulating factors that could promote the generation of tolerance-inducing DC by affecting their T-cell stimulatory capacity (Segerer, Muller *et al.*, 2008).

	DC phenotype	T-cell stimulatory capacity	cytokines
hCG	immunostimulatory (CD40 ↑, CD83 ↑, CD86 ↑)	\rightarrow	IL-10 ↑
estrogen	immunostimulatory (CD40 ↑, CD83 ↑, CD86 ↑)	→	IL-10 ↑
progesterone	immunostimulatory (CD40 ↑, CD83 ↑, CD86 ↑)	↓ (in rats) → (in humans)	TNF- $\alpha \downarrow$, IL-1 $\beta \downarrow$ (in rats)
activin A and inhibin A	immunostimulatory (CD40 ↑, CD83 ↑, CD86 ↑)	↓	no significant effects

Table 1.

6. Summary

During pregnancy, the levels of the glycoprotein hormones activin A and inhibin A as well as female sex hormones (hCG, estradiol, progesterone) are substantially elevated in parallel with increased occurrence of cells of the monocyte/macrophage lineage like DC in the decidua. In vitro experiments revealed that all of them can induce a tolerogenic subtype of DC even though effects were found on different levels (reduction of the expression of co-stimulatory molecules, reduction of T-cell stimulation, tolerogenic cytokine-profile). Thus, this distinct mixture of hormonal factors seems to be indispensable to support the development of tolerance-inducing DC which could play a key role in the acceptance of the fetus.

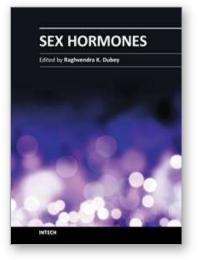
7. References

- [1] Bachelot A and Binart N (2005) Corpus luteum development: lessons from genetic models in mice. Curr Top Dev Biol, 68, 49-84.
- [2] Berndt S, Blacher S, Perrier dS, Thiry M, Tsampalas M, Cruz A, Pequeux C, Lorquet S, Munaut C, Noel A et al (2009) Chorionic gonadotropin stimulation of angiogenesis and pericyte recruitment. J Clin Endocrinol Metab, 94, 4567-4574.
- [3] Birdsall M, Ledger W, Groome N, Abdalla H, and Muttukrishna S (1997) Inhibin A and activin A in the first trimester of human pregnancy. J Clin Endocrinol Metab, 82, 1557-1560.
- [4] Biswas DK, Singh S, Shi Q, Pardee AB, and Iglehart JD (2005) Crossroads of estrogen receptor and NF-kappaB signaling. Sci STKE, 2005, e27.
- [5] Bonduelle ML, Dodd R, Liebaers I, Van SA, Williamson R, and Akhurst R (1988) Chorionic gonadotrophin-beta mRNA, a trophoblast marker, is expressed in human 8-cell embryos derived from tripronucleate zygotes. Hum Reprod, 3, 909-914.
- [6] Bonifaz L, Bonnyay D, Mahnke K, Rivera M, Nussenzweig MC, and Steinman RM (2002) Efficient targeting of protein antigen to the dendritic cell receptor DEC-205 in the steady state leads to antigen presentation on major histocompatibility complex class I products and peripheral CD8+ T cell tolerance. J Exp Med, 196, 1627-1638.
- [7] Butts CL, Shukair SA, Duncan KM, Bowers E, Horn C, Belyavskaya E, Tonelli L, and Sternberg EM (2007) Progesterone inhibits mature rat dendritic cells in a receptormediated fashion. Int Immunol, 19, 287-296.
- [8] Chang H, Brown CW, and Matzuk MM (2002) Genetic analysis of the mammalian transforming growth factor-beta superfamily. Endocr Rev, 23, 787-823.
- [9] Csapo AI and Pulkkinen M (1978) Indispensability of the human corpus luteum in the maintenance of early pregnancy. Luteectomy evidence. Obstet Gynecol Surv, 33, 69-81.
- [10] Ding D, Mehta H, McCune WJ, and Kaplan MJ (2006) Aberrant phenotype and function of myeloid dendritic cells in systemic lupus erythematosus. J Immunol, 177, 5878-5889.
- [11] Eramaa M, Hurme M, Stenman UH, and Ritvos O (1992) Activin A/erythroid differentiation factor is induced during human monocyte activation. J Exp Med, 176, 1449-1452.
- [12] Erlandsson MC, Jonsson CA, Islander U, Ohlsson C, and Carlsten H (2003) Oestrogen receptor specificity in oestradiol-mediated effects on B lymphopoiesis and immunoglobulin production in male mice. Immunology, 108, 346-351.

- [13] Florio P, Severi FM, Bocchi C, Luisi S, Mazzini M, Danero S, Torricelli M, and Petraglia F (2007) Single serum activin a testing to predict ectopic pregnancy. J Clin Endocrinol Metab, 92, 1748-1753.
- [14] Gomez-Lopez N, Guilbert LJ, and Olson DM (2010) Invasion of the leukocytes into the fetal-maternal interface during pregnancy. J Leukoc Biol, 88, 625-633.
- [15] Horiuchi A, Nikaido T, Yoshizawa T, Itoh K, Kobayashi Y, Toki T, Konishi I, and Fujii S (2000) HCG promotes proliferation of uterine leiomyomal cells more strongly than that of myometrial smooth muscle cells in vitro. Mol Hum Reprod, 6, 523-528.
- [16] 16. Hoshina M, Boothby M, Hussa R, Pattillo R, Camel HM, and Boime I (1985) Linkage of human chorionic gonadotrophin and placental lactogen biosynthesis to trophoblast differentiation and tumorigenesis. Placenta, 6, 163-172.
- [17] Huck B, Steck T, Habersack M, Dietl J, and Kammerer U (2005) Pregnancy associated hormones modulate the cytokine production but not the phenotype of PBMCderived human dendritic cells. Eur J Obstet Gynecol Reprod Biol, 122, 85-94.
- [18] Inaba K, Metlay JP, Crowley MT, Witmer-Pack M, and Steinman RM (1990) Dendritic cells as antigen presenting cells in vivo. Int Rev Immunol, *6*, 197-206.
- [19] Ivanova E, Kyurkchiev D, Altankova I, Dimitrov J, Binakova E, and Kyurkchiev S (2005) CD83 monocyte-derived dendritic cells are present in human decidua and progesterone induces their differentiation in vitro. Am J Reprod Immunol, 53, 199-205.
- [20] Jones KL, de Kretser DM, Patella S, and Phillips DJ (2004) Activin A and follistatin in systemic inflammation. Mol Cell Endocrinol, 225, 119-125.
- [21] Jones RL, Salamonsen LA, and Findlay JK (2002) Potential roles for endometrial inhibins, activins and follistatin during human embryo implantation and early pregnancy. Trends Endocrinol Metab, 13, 144-150.
- [22] Kammerer U, Eggert AO, Kapp M, McLellan AD, Geijtenbeek TB, Dietl J, van Kooyk Y, and Kampgen E (2003) Unique appearance of proliferating antigen-presenting cells expressing DC-SIGN (CD209) in the decidua of early human pregnancy. Am J Pathol, 162, 887-896.
- [23] Kane N, Kelly R, Saunders PT, and Critchley HO (2009) Proliferation of uterine natural killer cells is induced by human chorionic gonadotropin and mediated via the mannose receptor. Endocrinology, 150, 2882-2888.
- [24] Karpuzoglu E and Zouali M (2011) The multi-faceted influences of estrogen on lymphocytes: toward novel immuno-interventions strategies for autoimmunity management. Clin Rev Allergy Immunol, 40, 16-26.
- [25] Keay SD, Vatish M, Karteris E, Hillhouse EW, and Randeva HS (2004) The role of hCG in reproductive medicine. BJOG, 111, 1218-1228.
- [26] Kolkova Z, Noskova V, Ehinger A, Hansson S, Casslén B (2010) G protein-coupled estrogen receptor 1 (GPER, GPR 30) in normal human endometrium and early pregnancy decidua. Mol Hum Reprod, 16(10):743-51.
- [27] Kornyei JL, Lei ZM, and Rao CV (1993) Human myometrial smooth muscle cells are novel targets of direct regulation by human chorionic gonadotropin. Biol Reprod, 49, 1149-1157.
- [28] Kyurkchiev D, Ivanova-Todorova E, and Kyurkchiev SD (2010) New target cells of the immunomodulatory effects of progesterone. Reprod Biomed Online, 21, 304-311.

- [29] Lash GE, Schiessl B, Kirkley M, Innes BA, Cooper A, Searle RF, Robson SC, and Bulmer JN (2006) Expression of angiogenic growth factors by uterine natural killer cells during early pregnancy. J Leukoc Biol, 80, 572-580.
- [30] Lindzey J, Wetsel WC, Couse JF, Stoker T, Cooper R, and Korach KS (1998) Effects of castration and chronic steroid treatments on hypothalamic gonadotropin-releasing hormone content and pituitary gonadotropins in male wild-type and estrogen receptor-alpha knockout mice. Endocrinology, 139, 4092-4101.
- [31] Lockwood GM, Ledger WL, Barlow DH, Groome NP, and Muttukrishna S (1997) Measurement of inhibin and activin in early human pregnancy: demonstration of fetoplacental origin and role in prediction of early-pregnancy outcome. Biol Reprod, 57, 1490-1494.
- [32] Loke YW and King A (1995) Human Implantation. Cell Biology and Immunology., Cambridge University Press.
- [33] Lopata A and Hay DL (1989) The potential of early human embryos to form blastocysts, hatch from their zona and secrete HCG in culture. Hum Reprod, 4, 87-94.
- [34] Miyaura H and Iwata M (2002) Direct and indirect inhibition of Th1 development by progesterone and glucocorticoids. J Immunol, 168, 1087-1094.
- [35] Miyazaki S, Tsuda H, Sakai M, Hori S, Sasaki Y, Futatani T, Miyawaki T, and Saito S (2003) Predominance of Th2-promoting dendritic cells in early human pregnancy decidua. J Leukoc Biol, 74, 514-522.
- [36] Muttukrishna S, Jauniaux E, McGarrigle H, Groome N, and Rodeck CH (2004) In-vivo concentrations of inhibins, activin A and follistatin in human early pregnancy. Reprod Biomed Online, 8, 712-719.
- [37] Nalbandian G and Kovats S (2005) Understanding sex biases in immunity: effects of estrogen on the differentiation and function of antigen-presenting cells. Immunol Res, 31, 91-106.
- [38] Nalbandian G, Paharkova-Vatchkova V, Mao A, Nale S, and Kovats S (2005) The selective estrogen receptor modulators, tamoxifen and raloxifene, impair dendritic cell differentiation and activation. J Immunol, 175, 2666-2675.
- [39] Paharkova-Vatchkova V, Maldonado R, and Kovats S (2004) Estrogen preferentially promotes the differentiation of CD11c+ CD11b(intermediate) dendritic cells from bone marrow precursors. J Immunol, 172, 1426-1436.
- [40] Peeva E and Zouali M (2005) Spotlight on the role of hormonal factors in the emergence of autoreactive B-lymphocytes. Immunol Lett, 101, 123-143.
- [41] Pernis AB (2007) Estrogen and CD4+ T cells. Curr Opin Rheumatol, 19, 414-420.
- [42] Petri M, Howard D, and Repke J (1991) Frequency of lupus flare in pregnancy. The Hopkins Lupus Pregnancy Center experience. Arthritis Rheum, 34, 1538-1545.
- [43] Phillips DJ, Jones KL, Clarke IJ, Scheerlinck JP, and de Kretser DM (2005) Activin A: from sometime reproductive factor to genuine cytokine. Vet Immunol Immunopathol, 108, 23-27.
- [44] Prakash A, Laird S, Tuckerman E, Li TC, and Ledger WL (2005) Inhibin A and activin A may be used to predict pregnancy outcome in women with recurrent miscarriage. Fertil Steril, 83, 1758-1763.
- [45] Revankar CM, Cimino DF, Sklar LA, Arterburn JB, Prossnitz ER (2005) A transmembrane intracellular estrogen receptor mediates rapid cell signaling. Science, 307(5715):1625-30.

- [46] Rieger L, Honig A, Sutterlin M, Kapp M, Dietl J, Ruck P, and Kammerer U (2004) Antigen-presenting cells in human endometrium during the menstrual cycle compared to early pregnancy. J Soc Gynecol Investig, 11, 488-493.
- [47] Robson NC, Phillips DJ, McAlpine T, Shin A, Svobodova S, Toy T, Pillay V, Kirkpatrick N, Zanker D, Wilson K et al (2008) Activin-A: a novel dendritic cell-derived cytokine that potently attenuates CD40 ligand-specific cytokine and chemokine production. Blood, 111, 2733-2743.
- [48] Rutella S, Bonanno G, Procoli A, Mariotti A, de Ritis DG, Curti A, Danese S, Pessina G, Pandolfi S, Natoni F et al (2006) Hepatocyte growth factor favors monocyte differentiation into regulatory interleukin (IL)-10++IL-12low/neg accessory cells with dendritic-cell features. Blood, 108, 218-227.
- [49] Salem ML (2004) Estrogen, a double-edged sword: modulation of TH1- and TH2mediated inflammations by differential regulation of TH1/TH2 cytokine production. Curr Drug Targets Inflamm Allergy, 3, 97-104.
- [50] Sallusto F and Lanzavecchia A (1999) Mobilizing dendritic cells for tolerance, priming, and chronic inflammation. J Exp Med, 189, 611-614.
- [51] Segerer SE, Muller N, Brandt J, Kapp M, Dietl J, Reichardt HM, Rieger L, and Kammerer U (2008) The glycoprotein-hormones activin A and inhibin A interfere with dendritic cell maturation. Reprod Biol Endocrinol, 6, 17.
- [52] Segerer SE, Muller N, van den BJ, Kapp M, Dietl J, Reichardt HM, Rieger L, and Kammerer U (2009) Impact of female sex hormones on the maturation and function of human dendritic cells. Am J Reprod Immunol, 62, 165-173.
- [53] Shao L, Frigon NL, Jr., Young AL, Yu AL, Mathews LS, Vaughan J, Vale W, and Yu J (1992) Effect of activin A on globin gene expression in purified human erythroid progenitors. Blood, 79, 773-781.
- [54] Smithson G, Couse JF, Lubahn DB, Korach KS, and Kincade PW (1998) The role of estrogen receptors and androgen receptors in sex steroid regulation of B lymphopoiesis. J Immunol, 161, 27-34.
- [55] Steinman RM (2003) Some interfaces of dendritic cell biology. APMIS, 111, 675-697.
- [56] Stites DP and Siiteri PK (1983) Steroids as immunosuppressants in pregnancy. Immunol Rev, 75, 117-138.
- [57] Tamrazi A, Carlson KE, Daniels JR, Hurth KM, and Katzenellenbogen JA (2002) Estrogen receptor dimerization: ligand binding regulates dimer affinity and dimer dissociation rate. Mol Endocrinol, 16, 2706-2719.
- [58] Tong S, Wallace EM, and Burger HG (2003) Inhibins and activins: clinical advances in reproductive medicine. Clin Endocrinol (Oxf), 58, 115-127.
- [59] Wan H, Versnel MA, Leijten LM, van Helden-Meeuwsen CG, Fekkes D, Leenen PJ, Khan NA, Benner R, and Kiekens RC (2008) Chorionic gonadotropin induces dendritic cells to express a tolerogenic phenotype. J Leukoc Biol, 83, 894-901.
- [60] Yang L, Hu Y, and Hou Y (2006) Effects of 17beta-estradiol on the maturation, nuclear factor kappa B p65 and functions of murine spleen CD11c-positive dendritic cells. Mol Immunol, 43, 357-366.
- [61] Yoshimura T, Inaba M, Sugiura K, Nakajima T, Ito T, Nakamura K, Kanzaki H, and Ikehara S (2003) Analyses of dendritic cell subsets in pregnancy. Am J Reprod Immunol, 50, 137-145.



Sex Hormones Edited by Prof. Raghvendra Dubey

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Sex Hormones not only regulate reproductive function, but they also play a prominent role in the biology and physiology of several organs/tissues and in the pathophysiology of several diseases. During the last two decades, the information on the mechanisms of action of sex hormones, such as estrogens and androgens, has rapidly evolved from the conventional nuclear receptor dependent mechanisms to include additional non-nuclear, non-genomic and receptor-independent mechanisms. This highlights the need to update the current knowledge on sex hormones and their mode of action. Increasing evidence that exogenous/epigenetic factors can influence sex hormone production and action highlights the need to update our knowledge on the mechanisms involved. This book provides a systematic and updated overview of the male/female sex-hormones and their impact in the biology and physiology of various organs. Additionally, the book discusses their positive and negative association with the pathophysiology of various diseases (e.g. osteoporosis, cardiovascular-disease, hypogonadism, reproduction, cancer) and their therapeutic potential.

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