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Interleukin-6 in the Breast Tumor Microenvironment

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1. Introduction

Greater than 200,000 new cases of breast cancer cases were diagnosed in 2010 in the United States, with approximately 40,000 women succumbing to the disease (www.cancer.gov). Globally, an estimated 1.38 million new cases of breast cancer were diagnosed in 2008, with greater than 450,000 women succumbing to the disease (Jemal *et al.*, 2011). Despite our improved understanding of breast carcinogenesis, breast cancer remains the second most commonly diagnosed cancer in women behind non-melanoma skin cancer and the second leading cause of death in women behind lung cancer. These epidemiological statistics highlight the overwhelming clinical dilemma of breast cancer and emphasize the need for novel therapeutic targets and prevention strategies. Countless studies in the fields of mammary gland development and breast cancer have led to an appreciation of a breast tumor microenvironment that actively contributes to the heterogeneous nature of breast cancer. The current review will focus on the impact of IL-6 and STAT3 activation in the breast tumor microenvironment and subsequently present rationale for targeting the IL-6/STAT3 signaling pathway in this setting. IL-6 is a quintessential pleiotropic cytokine produced by a diverse number of cell populations, most of which can localize to the breast tumor microenvironment. Excessive IL-6 has been demonstrated in primary breast tumors and breast cancer patient sera and is associated with poor clinical outcomes in breast cancer. These clinical associations are corroborated by emerging preclinical data revealing that IL-6 is a potent growth factor and promotes an epithelial-mesenchymal (EMT) phenotype in breast cancer cells to indicate that IL-6 in the breast tumor microenvironment is clinically relevant. Numerous clinical reports have now demonstrated the safety and efficacy of IL-6 signaling antagonists in multiple diseases, which supports future investigations of these therapies in breast cancer.

Estrogen receptor-alpha (ER α) is a latent cytoplasmic ligand-activated transcription factor utilized by clinicians to subclassify the heterogeneous disease of breast cancer. ER α -positive breast cancer incidence increases up to age 51, the mean age of menopause, and continues to increase until age 80. Conversely, ER α -negative breast cancer incidence plateaus and even slightly decreases at age 51, while demonstrating an increase prior to age 50 comparable to that of ER α -positive disease. This discrepancy between the two incidence rates at menopause produces an inflection in the incidence rate of all breast cancer cases which has been termed Clemmesen's hook (Anderson and Matsuno, 2006). Whereas the prevalence of

ER α -positive cells within terminal duct lobular units of the breast of healthy premenopausal women has been reported at 7%, this number is estimated at 42% in postmenopausal women (Shoker *et al.*, 1999). In addition, approximately two-thirds of all breast cancers are diagnosed as ER α -positive, and 75% of postmenopausal breast cancers are ER α -positive (Macedo *et al.*, 2009). Progesterone receptor (PR) and epidermal growth factor receptor 2 (EGFR2; HER2; or ErbB2), a receptor tyrosine kinase involved in cellular proliferation, have also acquired much clinical attention following reports of dismal survival rates in “triple negative” (ER α -negative/PR-negative/HER2 not overexpressed) breast cancer patients. Triple negative breast cancer represents approximately 15 to 20% of all breast cancer cases and can only be treated with standard chemotherapy as it lacks current adjuvant therapeutic targets. Such breast tumors are highly proliferative with a high mitotic index, increased necrosis, elevated apoptosis, and typically are of higher tumor grade. *TP53* gene and p53 protein mutations as well as loss of the Rb tumor suppressor protein are common. Familial breast cancer patients with congenital BRCA1 mutations often present with triple negative breast cancer, as do relatively younger breast cancer patients and African American women. Currently, triple negative breast cancers are associated with a poor prognosis largely due to poor survival rates and early relapse. The fact that these breast tumors respond well if not completely to initial chemotherapy may seem counterintuitive, but enhanced invasiveness, consequent distant metastasis, and residual local recurrence eventually promote poor survival rates (Irvin and Carey, 2008).

Breast cancer most commonly metastasizes to bone, followed by lung, liver, and brain. Perhaps due to the heterogeneity across individual breast cancer cases, few prognostic molecular biomarkers have been demonstrated to accurately predict metastatic potential. One of the most important of these biomarkers is ER α , which is clinically exploited as a predictor of bone metastasis (Kominsky and Davidson, 2006). Whereas ER α -positive breast cancers have a strong tendency to metastasize to bone if at all (James *et al.*, 2003), their ER α -negative counterparts favor visceral organs such as lung and liver (Hess *et al.*, 2003). Primary mammary tumor cell dissemination has been quantified at 3 to 4 $\times 10^6$ primary tumor cells in circulation per 24 hours per gram of tumor in a rat mammary carcinoma model, which exemplifies the inefficient nature of metastasis (Butler and Gullino, 1975). Although metastasis has been generally accepted as a relatively late event throughout cancer progression, recent work has revealed evidence of early primary tumor cell dissemination, thus refuting this paradigm (Klein, 2009). In particular, it has now been demonstrated that untransformed triple transgenic (doxycycline-inducible K-ras, MYC, and polyoma middle T antigen) mammary epithelial cells are capable of lung colonization when tail vein-injected into immunocompromised female mice on doxycycline. This work showed that untransformed “normal” mammary epithelial cells can colonize ectopic lung tissue, and upon oncogene activation, disseminated mammary epithelial cells within circulation or a foreign host microenvironment are capable of forming tumors at the ectopic site (Podsypanina *et al.*, 2008). Additionally, reports of bone marrow cytokeratin-positive epithelial cells in up to 48% of breast cancer patients without overt metastases also offer support for early primary tumor cell dissemination. Decreased survival in patients with such cells was demonstrated in all studies (Braun *et al.*, 2000; Diel *et al.*, 1996; Gebauer *et al.*, 2001; Pantel *et al.*, 2003; Vannucchi *et al.*, 1998). Furthermore, only 8% of these patients with bone marrow micrometastases exhibited cytokeratin-positive/Ki67-positive cells, suggesting that lack of overt bone metastasis may be due to disseminated tumor cell dormancy (Pantel *et al.*, 2003).

2. The breast tumor microenvironment

A normal epithelial tissue can undergo hyperplasia and acquire tumorigenic properties that promote the development of a benign, non-invasive solid tumor known as carcinoma *in situ*. Normal epithelial tissues and non-invasive carcinoma *in situ* tumors are separated from a supportive stromal compartment by an intact basement membrane. Ultimately, carcinoma *in situ* can progress to a malignant, invasive carcinoma, the most common form of human cancer. The panoply of published investigations between the fields of mammary gland development and breast cancer has led to an appreciation for a supportive non-epithelial mammary stroma that mechanically and biologically restrains tumorigenesis. However, tumors of the breast and other epithelial tissues obviously overcome these growth restraints and exploit this stroma to sculpt a vastly divergent tumor stroma. Tumor stroma is generally divided into four main components: tumor vasculature, inflammatory leukocytes, extracellular matrix (ECM) and soluble growth factors, and fibroblasts. Malignant carcinoma cells and tumor stromal cells bi-directionally communicate with one another through paracrine signaling and intercellular contacts in a disorganized ECM to constitute a tumor microenvironment. Tumor-associated fibroblasts (TAF), the predominant stromal cell population within the tumor microenvironment, acquire and sustain an “activated” phenotype that promotes tumor progression (Rasanen and Vaheri, 2010). TAF are capable of enhancing breast tumor growth and metastasis by means of promoting angiogenesis (Orimo *et al.*, 2005), epithelial-mesenchymal transition (EMT) (Martin *et al.*, 2010; Radisky *et al.*, 2005), and progressive genetic instability (Kurose *et al.*, 2001; Moinfar *et al.*, 2000). In contrast, a normal mammary microenvironment can act in a dominant manner to inhibit tumor growth and “revert” the malignant phenotype of breast cancer cells (Kenny and Bissell, 2003). While resident breast tissue fibroblasts can inhabit breast tumors as TAF, breast tumors also recruit distant cell populations that engraft within the breast tumor microenvironment where they actively contribute as TAF. For example, mesenchymal stem cells (MSC), a bone marrow-derived stromal cell population, home to breast cancer cell xenograft tumors and persist as TAF (Spaeth *et al.*, 2009).

3. Cancer-associated inflammation

Although highly characterized for their protective capacity against infection, inflammatory leukocytes also reside within the tumor microenvironment. In fact, various immune cells are capable of eliminating transformed cells and thus preventing tumorigenesis in a process termed immunosurveillance (Dunn *et al.*, 2004). Whereas acute inflammation may prevent tumorigenesis by promoting an immune response directed against transformed cells, chronic inflammation promotes tumorigenesis. Rudolf Virchow is credited with making the seminal link between chronic inflammation and cancer by noting that human tumor biopsies were often infiltrated with inflammatory cells (Balkwill and Mantovani, 2001). Leukocytes can be detected in non-malignant tumors and carcinomas, including breast cancer (DeNardo and Coussens, 2007), which suggests an ongoing antitumor immune response. Despite the infiltration of leukocytes such as cytotoxic T-cells and NK-cells, the persistence of a tumor demonstrates immune evasion and highlights the local and systemic immune suppressive state of the tumor microenvironment and the tumor-bearing host, respectively.

4. Interleukin-6: A quintessential pleiotropic cytokine

Interleukin-6 (IL-6) is an inflammation-associated cytokine and major inducer of C-reactive protein (CRP) throughout the acute phase inflammatory response. *IL6* gene expression is nuclear factor-kappaB (NF- κ B)-dependent (Chauhan *et al.*, 1996) and produces a 26 kDa IL-6 protein product. First characterized as a T-cell-derived factor that induced proliferation, differentiation, and immunoglobulin production in B-cells, IL-6 was originally named B-cell stimulating factor-2 (BSF-2). It was later thought to be a novel interferon (IFN- β_2) due to studies demonstrating the ability of IL-6 to activate signal transducer and activator of transcription 3 (STAT3) (Kishimoto, 2006). Complementary DNA encoding the human IL-6 gene was subsequently cloned, and human IL-6 transgenic mice demonstrated a polyclonal IgG1 plasmacytosis phenotype (Suematsu *et al.*, 1989). Next, IL-6 knockout (IL-6^{-/-}) mice were generated and characterized. IL-6^{-/-} mice underwent normal development, but adult animals exhibited reduced numbers of peripheral T-cells and impaired antiviral cytotoxic T-cell activity (Kopf *et al.*, 1994). In addition, IL-6 is a critical factor during hematopoiesis and subsequent lymphocyte differentiation and activation. Multiple diverse cell populations including fibroblasts, T and B-cells, monocytes, macrophages, endothelial cells, keratinocytes, astrocytes, and smooth muscle cells all have the potential to produce constitutive or inducible IL-6 (Kishimoto, 2006).

Depending on cellular context, IL-6 can signal through multiple kinase-dependent proliferation and anti-apoptosis pathways including the mitogen-activated protein kinase (MAPK) pathway, the phosphatidylinositol-triphosphate kinase (PI-3K)/Akt pathway, and perhaps the most commonly evaluated in breast cancer, the Janus kinase (JAK)/signal transducer and activator of transcription-3 (STAT3) pathway (Hodge *et al.*, 2005). To do so, a plasma membrane-associated IL-6 receptor (IL-6R/CD126) homodimer first ligates two soluble IL-6 molecules, which leads to gp130 (CD130) homodimer ligation. Whereas IL-6R is only expressed on hepatocytes, osteoclasts, and most immune cells under normal physiological conditions, gp130 is a ubiquitous and promiscuous receptor involved in multiple cytokine signaling pathways (e.g., IL-11, leukemia inhibitory factor (LIF), oncostatin M (OSM), and ciliary neurotrophic factor (CNTF)) (Rose-John *et al.*, 2006). To initiate classical JAK/STAT3 signal transduction, JAK are recruited to the intracellular domain of the gp130 receptor where they bind and autophosphorylate. Subsequent gp130 phosphorylation via activated JAK offers docking sites for STAT3 and other receptor-associated proteins. Once bound to the intracellular domain of gp130, STAT3 is specifically phosphorylated (pSTAT3) by adjacent JAK on a C-terminal tyrosine residue (Y705), which grants its disengagement from the receptor. Dissociation of pSTAT3^{Y705} from gp130 facilitates its homodimerization within the cytoplasm, and the pSTAT3^{Y705} homodimer translocates to the nucleus. There, pSTAT3^{Y705} binds to specific promoters whereby it initiates the transcription of multiple downstream target genes (Clevenger, 2004). Under normal physiological conditions, an inhibitory feedback loop maintains rapid and transient STAT3 activation. Following activation in normal cells, STAT3 induces suppressors of cytokine signaling (SOCS) and protein inhibitors of activated STATs (PIAS) expression. While SOCS-1 specifically inhibits JAK function, SOCS-3 binds the IL-6R complex to inhibit IL-6 signal transduction. PIAS-3 directly interacts with STAT3 to inhibit all STAT3 target gene expression (Kishimoto, 2006). In contrast, many human cancers, including breast cancer, exhibit constitutive STAT3 activity. Recent studies have demonstrated that unphosphorylated STAT3 (U-STAT3) accumulates in tumor cells with constitutively active

STAT3 where it forms a complex with NF- κ B to activate a subset of NF- κ B target genes (Yang and Stark, 2008). Alternatively, IL-6 *trans*-signaling describes an IL-6 signaling pathway whereby an IL-6 soluble receptor (IL-6sR) binds IL-6 and subsequently ligates gp130 to stimulate STAT3 activation in cells that only express gp130. IL-6sR is naturally produced by either proteolytic cleavage of the membrane-bound IL-6R or alternative splicing of IL-6R mRNA (Rose-John *et al.*, 2006). Whereas IL-6 serum levels continue to increase with age, levels of serum IL-6sR rise until approximately age 70 at which time they gradually decline (Giuliani *et al.*, 2001). Furthermore, IL-6sR expression has been demonstrated in human breast cancer cell lines (Crichton *et al.*, 1996; Oh *et al.*, 1996; Singh *et al.*, 1995), suggesting that IL-6 *trans*-signaling mediates the effects of IL-6 in breast cancer cells. In contrast, an endogenous soluble gp130 (sgp130) specifically antagonizes IL-6 *trans*-signaling by exclusively ligating the IL-6/IL-6sR complex, thus having no effect on cells that express the membrane-bound IL-6R (Rose-John *et al.*, 2006) (Figure 1).

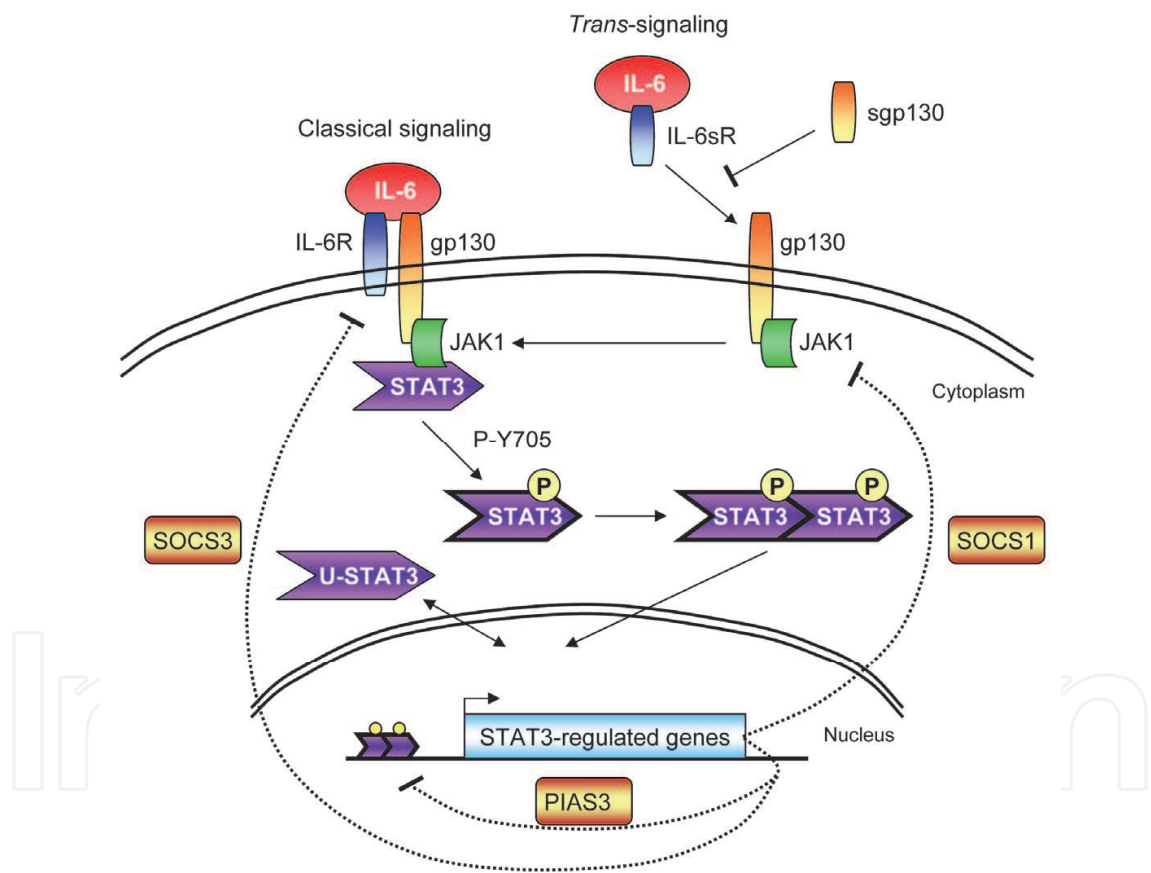


Fig. 1. The IL-6/STAT3 signaling pathway

5. Excessive IL-6 in human breast cancer

Aberrantly elevated IL-6 is associated with a poor prognosis in breast cancer (Bachelot *et al.*, 2003; Salgado *et al.*, 2003; Zhang and Adachi, 1999). Human breast tumors produce more IL-6 when compared to matched healthy breast tissue, and tumor IL-6 levels concurrently increase with tumor grade. In addition, increased serum IL-6 has been demonstrated in

breast cancer patients compared to normal donors and correlates with advanced breast tumor stage (Kozlowski *et al.*, 2003) and increased number of metastatic sites (Salgado *et al.*, 2003). Furthermore, a single nucleotide polymorphism (SNP) exists at position -174 in the IL-6 gene promoter region, noted as IL-6 (-174 G>C), with the following allele frequency in a Caucasian population: 36% G/G, 44% G/C, and 18% C/C. An inflammatory stimulus such as *Salmonella typhii* vaccination induced higher serum IL-6 in those individuals with the G/G allele (Bennermo *et al.*, 2004). Although the IL-6 (-174 G>C) SNP is not associated with increased risk of developing breast cancer (Gonzalez-Zuloeta Ladd *et al.*, 2006; Litovkin *et al.*, 2007; Yu *et al.*, 2009b), it is significantly associated with disease-free and overall survival in breast cancer patients (DeMichele *et al.*, 2003).

ER α is expressed in luminal subtype breast tumors (Perou *et al.*, 2000) and therefore associated with improved patient survival (Buyse *et al.*, 2006; Sorlie *et al.*, 2001). A clear and well-characterized inverse correlation exists between breast cancer ER α status and IL-6. In fact, ER α directly binds to NF- κ B, thus preventing transactivation of *IL6* gene expression (Galien and Garcia, 1997), which demonstrates a direct mechanism for such a correlation. Furthermore, ER α -negative human breast tumors produce more IL-6 than tumors that express ER α (Chavey *et al.*, 2007), and IL-6 serum levels are higher in ER α -negative breast cancer patients compared to ER α -positive patients (Jiang *et al.*, 2000). Likewise, ER α -negative breast cancer cell lines produce autocrine IL-6 whereas ER α -positive breast cancer cell lines do not (Sasser *et al.*, 2007). Therefore, this strongly suggests that ER α -negative breast cancer cells would exploit both paracrine (i.e., stromal cell-derived) and autocrine IL-6 signaling, whereas ER α -positive breast cancer cells could only utilize paracrine IL-6 signaling. In addition, ER α -negative breast cancer patients, whose tumors produce more IL-6 than those that express ER α (Chavey *et al.*, 2007), showed no difference in survival between the G/G allele (higher inducible serum IL-6) and any C allele (lower inducible serum IL-6) at the IL-6 (-174 G>C) promoter SNP. In contrast, ER α -positive breast cancer patients with any C allele at the IL-6 (-174 G>C) promoter SNP demonstrated improved disease-free and overall survival compared to those with the G/G allele (DeMichele *et al.*, 2003).

6. IL-6 promotes breast cancer cell growth

Stromal fibroblasts isolated from multiple types of tumors (i.e., TAF) or cancers (i.e., CAF) are now appreciated as influential players in cancer progression and metastasis (Orimo and Weinberg, 2006). CAF derived from multiple cancer types, including murine mammary cancers, exhibit an activated, proinflammatory phenotype with increased IL-6 production (Erez *et al.*, 2010). Furthermore, work from our laboratory has demonstrated that fibroblasts isolated from breast tissue and common sites of breast cancer metastasis such as bone and lung enhance the growth of breast cancer cells in an IL-6-dependent manner, and IL-6 is the major fibroblast-derived soluble factor that induced STAT3 activation in breast cancer cells (Sasser *et al.*, 2007; Studebaker *et al.*, 2008). MDA-MB-231 breast cancer cells are commonly utilized to model triple negative breast cancer and produce autocrine IL-6. MDA-MB-231 cells expressing a dominant negative isoform of gp130 lacked constitutively active STAT3 and exhibited impaired tumorigenicity in an orthotopic xenograft model (Selander *et al.*, 2004), thus suggesting that IL-6 may drive tumor progression in this model. In addition, STAT3 is estimated to be constitutively activated in more than half of primary breast cancers due to IL-6 signaling (Berishaj *et al.*, 2007).

Mesenchymal stem cells (MSC) are a bone marrow-derived fibroblast cell population that can be recruited to the breast tumor stroma, acquire a TAF phenotype, and produce high levels of IL-6. MSC enhance the growth of ER α -positive breast cancer cells, which do not express IL-6 or activated STAT3. In contrast, MSC have no effect on IL-6-producing ER α -negative breast cancer cells, which express constitutively activated STAT3. Moreover, ER α -positive breast cancer cells orthotopically co-injected with MSC or MSC conditioned medium and ER α -positive breast cancer cells that ectopically express IL-6 demonstrate enhanced xenograft tumor growth in the absence of exogenous 17 β -estradiol (Sasser *et al.*, 2007). Similar differential growth enhancement was demonstrated *in vivo* with ER α -positive and ER α -negative breast cancer cells co-injected with MSC, which also promoted metastasis (Karnoub *et al.*, 2007). Interestingly, IL-6 has been reported to facilitate the recruitment of MSC to hypoxic breast tumor microenvironments (Rattigan *et al.*, 2010). Likewise, IL-6 secreted from breast cancer cells has been shown to contribute to a recently characterized phenomenon termed “self-seeding” in which aggressive circulating tumor cells engraft within their original xenograft tumor (Kim *et al.*, 2009). MSC have also been shown to mediate the self-renewal capacity of breast cancer stem cells, in part, through a reciprocal IL-6 loop (Liu *et al.*, 2010). Taken together, preceding evidence strongly suggests that IL-6 promotes breast cancer cell growth by activating STAT3, which culminates with the upregulation of proliferative oncogenes such as c-Myc and cyclin D1 and growth factors such as IL-6, hepatocyte growth factor (HGF), vascular endothelial growth factor (VEGF), and epidermal growth factor (EGF) (Yu *et al.*, 2009a).

7. IL-6 promotes epithelial-mesenchymal transition in breast cancer cells

Normal polarized epithelial cells exhibit ‘cobblestone’ homophilic morphology and express E-cadherin, which is required for epithelial cell polarization, phenotype, and consequent homeostasis (Jeanes *et al.*, 2008). E-cadherin is a key prognostic molecular biomarker clinically utilized to predict the metastatic propensity of breast cancer. Whereas very few studies have failed to demonstrate E-cadherin as an independent prognostic biomarker in breast cancer patients (Lipponen *et al.*, 1994; Parker *et al.*, 2001), the overwhelming majority of relevant studies have revealed E-cadherin as one of the strongest predictors of patient survival. Specifically, impaired E-cadherin expression in human breast tumors correlates with enhanced invasiveness, metastatic potential (Oka *et al.*, 1993), and decreased breast cancer patient survival (Heimann and Hellman, 2000; Pedersen *et al.*, 2002). While appropriate E-cadherin function is essential to the maintenance of epithelial cell morphology, phenotype, and homeostasis, regulation of E-cadherin expression is of equal importance. *CDH1*, the gene that encodes E-cadherin, is located on human chromosome 16q22.1 (Rakha *et al.*, 2006) and is susceptible to inactivation by promoter hypermethylation, somatic mutation, or aberrant overexpression of repressive transcription factors including Twist, Snail, and Slug among others (Hirohashi, 1998). Likewise, E-cadherin loss of function can arise due to extracellular domain-specific proteolytic cleavage. Although uncommon, germline mutations of *CDH1* predispose individuals to hereditary diffuse gastric cancer (HDGC) syndrome, and a proportion of these patients present with other cancers, including breast cancer (Guilford, 1999).

E-cadherin was initially termed uvomorulin in mice and L-CAM in chicks following its discovery as a 120 kDa calcium-dependent trypsin-labile cell surface glycoprotein required for intercellular adhesion in mouse blastomeres (Hyafil *et al.*, 1981) and chick embryos

(Brackenbury *et al.*, 1981). It now represents the best studied member of the cadherin family of tissue-specific homophilic intercellular adhesion molecules. E-cadherin knockout studies have demonstrated early embryonic lethality due to impaired maintenance of epithelial polarity and failure to form an intact epithelium in E-cadherin^{-/-} embryos (Larue *et al.*, 1994). E-cadherin is localized on the cell surface of epithelial cells, and each E-cadherin protein consists of an amino-terminal extracellular domain, a single-pass transmembrane segment, and a carboxy-terminal intracellular domain. Five calcium-binding repeated subunits comprise an extracellular domain that promotes homophilic interaction to ultimately form anti-parallel trans-E-cadherin dimers between adjacent cells (Guilford, 1999). The intracellular domain is comprised of a juxtamembrane p120-catenin binding subdomain and a C-terminal beta (β)-catenin binding subdomain. β -catenin, a potent transcription factor, binds E-cadherin and alpha (α)-catenin subsequently binds β -catenin. Although contentious (Weis and Nelson, 2006), it is generally acknowledged that α -catenin interacts with F-actin and thereby, facilitates the linkage of E-cadherin to the cytoskeleton. This E-cadherin-catenin-actin complex localizes to epithelial intercellular junctions called adherens junctions and is critical to epithelial cell adhesion, polarity, and morphology (Hartsock and Nelson, 2008). Furthermore, E-cadherin sequesters β -catenin at the cell surface as one means to inhibit β -catenin nuclear translocation and consequent expression of β -catenin responsive genes (Perez-Moreno *et al.*, 2003).

Another prominent role of E-cadherin is that of an invasion/metastasis suppressor protein. Upon loss of E-cadherin and subsequent dissociation of adherens junctions, epithelial cells acquire enhanced invasive capability (Behrens *et al.*, 1989). MDA-MB-231 cells, an ER α -negative breast cancer cell line, lack E-cadherin, whereas MCF-7 cells, an ER α -positive breast cancer cell line express high levels of E-cadherin (Kenny *et al.*, 2007), and MDA-MB-231 cells exhibit enhanced invasive capability compared to MCF-7 cells (Sommers *et al.*, 1991). Naturally, E-cadherin expression and consequent invasive capacity regulate the propensity of breast cancer metastasis. Multiple signaling pathways are activated following loss of E-cadherin protein, which promote transformed human breast epithelial cell metastasis in a xenograft model. Interestingly, Twist, a transcriptional repressor of *CDH1*, is induced upon loss of E-cadherin and is necessary for metastasis in this model. Furthermore, the E-cadherin binding partner, β -catenin, was shown to be necessary but not sufficient for the EMT phenotype induced following loss of E-cadherin (Onder *et al.*, 2008). Ectopic expression of murine E-cadherin in highly metastatic human MDA-MB-231 cells significantly reduced osteolytic bone metastases in a murine intracardiac dissemination model (Mbalaviele *et al.*, 1996). Likewise, aberrant cytoplasmic or diminished to negative E-cadherin immunostaining patterns are commonly detected in invasive poorly differentiated breast carcinomas compared to noninvasive well-differentiated breast carcinomas and are associated with increased probability of breast carcinoma metastasis (Oka *et al.*, 1993). The finding that distant metastases often express E-cadherin even in patients which exhibit primary breast carcinomas which lack E-cadherin suggests that ultimate re-expression may be necessary for colonization of secondary tissues (Kowalski *et al.*, 2003; Saha *et al.*, 2007).

Loss of E-cadherin is a prerequisite for epithelial-mesenchymal transition (EMT), a highly conserved process which exemplifies the aberrant activation of an embryonic gene expression program during carcinoma progression. EMT is critical for multiple steps of developmental metazoan cellular morphogenesis as demonstrated in well-characterized

Drosophila and *Xenopus* models. Throughout embryonic development, EMT whereby epithelial cells give rise to more motile mesenchymal cells is essential for mesoderm and neural crest formation. Importantly, this is a transient process and mesenchymal-epithelial transition (MET) allows for cellular reversion (Yang and Weinberg, 2008).

Whereas EMT has been extensively studied for its essential role in embryogenesis, the concept of EMT-like cellular changes in human cancers has gained acceptance as a major mechanism to promote primary tumor cell invasion and subsequent tumor metastasis. A carcinoma cell must first detach from the primary tumor and invade through the basement membrane into the underlying tissue parenchyma to initiate the metastatic cascade. Although cancer-associated EMT was considered a controversial notion even in recent years (Tarin *et al.*, 2005), it has been demonstrated in multiple human carcinomas, including breast cancer (Cheng *et al.*, 2008; Heimann and Hellman, 2000; Moody *et al.*, 2005; Sarrio *et al.*, 2008), and is now recognized as a putative mediator of tumor metastasis. An EMT phenotype including impaired E-cadherin expression with concomitant induction of Vimentin, Alpha-smooth-muscle-actin, and/or N-cadherin is associated with the basal breast cancer subtype, suggesting that EMT may promote characteristic aggressiveness in these tumors and contribute to poor breast cancer patient survival (Sarrio *et al.*, 2008). Likewise, relatively noninvasive ER α -positive MCF-7 cells express E-cadherin, consistent with a characteristic epithelial phenotype, and are classified as luminal subtype, whereas highly invasive ER α -negative MDA-MB-231 cells lack E-cadherin and are classified as basal subtype (Blick *et al.*, 2008). Furthermore, ER α directly correlates with E-cadherin in primary human breast tumors (Ye *et al.*, 2010). While EMT may enhance carcinoma cell invasion and subsequent dissemination which would increase metastatic potential, it is not synonymous with metastasis in all models. For example, Lou, *et al.* demonstrated that EMT alone was insufficient for spontaneous murine mammary carcinoma metastasis (Lou *et al.*, 2008). Yet, Weinberg and colleagues described the promotion of metastasis with loss of E-cadherin and a consequent EMT phenotype in transformed human breast epithelial cells (Onder *et al.*, 2008).

Our laboratory has previously demonstrated that exogenous IL-6 exposure induced an EMT phenotype in a panel of human ER α -positive breast cancer cells, which included E-cadherin repression and concomitant induction of Vimentin, N-cadherin, Snail, and Twist. In addition, ectopic expression of IL-6 in ER α -positive MCF-7 breast cancer cells promoted an EMT phenotype and enhanced invasiveness. Likewise, MCF-7 cells with ectopic Twist expression exhibit an EMT phenotype (Mironchik *et al.*, 2005), autocrine IL-6 production, and constitutive STAT3 activation (Sullivan *et al.*, 2009).

8. Therapeutic targeting of the IL-6/STAT3 pathway

IL-6 levels are increased in human breast tumors and breast cancer patient sera, and excessive IL-6, both circulating and within the breast tumor microenvironment, is associated with poor clinical outcomes in breast cancer. STAT3, a critical downstream mediator of IL-6 signaling, is constitutively activated in more than half of human cancers and promotes the expression of proliferative, anti-apoptotic, immune suppressive, and pro-angiogenic target genes, which all potentiate carcinogenesis. Whereas the IL-6 signaling network has been targeted in numerous autoimmune diseases and cancers, this therapeutic strategy has yet to be clinically employed for breast cancer. Increased preclinical reports have revealed novel

mechanisms underlying IL-6/STAT3 signaling in breast cancer cells such as enhanced growth, induction of EMT, multidrug resistance, and recruitment of peripheral fibroblasts. Taken together, accumulating preclinical and clinical data emphasize IL-6 as a highly attractive therapeutic target in breast cancer. It is therefore imperative that more work be done to evaluate current therapeutics and develop novel agents that target IL-6/STAT3 signaling in breast cancer models.

Multiple strategies could be utilized to target the IL-6/STAT3 pathway, but first and most obvious would be anti-IL-6 neutralizing antibodies. One such anti-IL-6 monoclonal antibody is Siltuximab (CANTO 328). The safety and efficacy of Siltuximab has been demonstrated in preclinical studies and phase I/II clinical trials of diverse human pathologies and malignancies including Castleman's disease (van Rhee *et al.*, 2010), multiple myeloma (Hunsucker *et al.*, 2011; Voorhees *et al.*, 2007), prostate cancer (Cavarretta *et al.*, 2007; Cavarretta *et al.*, 2008; Dorff *et al.*, 2010; Karkera *et al.*, 2011), renal cell carcinoma (Puchalski *et al.*, 2010; Rossi *et al.*, 2010), non-small cell lung cancer (Song *et al.*, 2010), and ovarian cancer (Guo *et al.*, 2010). Furthermore, IL-6R can be targeted with tocilizumab, an anti-IL-6R monoclonal antibody that has shown promising results in IL-6-driven autoimmune diseases (Tanaka *et al.*, 2011) and was recently approved by the FDA for the treatment of rheumatoid arthritis. The promiscuous IL-6 coreceptor, gp130, also has an endogenous soluble form (sgp130) that exclusively inhibits IL-6 *trans*-signaling, thus preserving classical IL-6 signaling. Therapeutic sgp130 would potentially be more targeted toward breast cancer cells, which generally lack membrane-associated IL-6R and therefore utilize IL-6 *trans*-signaling through IL-6sR. Recombinant soluble gp130 (sgp130-Fc) has been shown to inhibit murine colon carcinogenesis (Becker *et al.*, 2004), suggesting that it may prove effective in breast cancer as well. Finally, a growing number of non-selective kinase inhibitors and recent focus on specific JAK and STAT3 inhibitor development will provide further insight into the roles of JAK and STAT3 in breast cancer.

9. Conclusions

Breast cancer is a heterogeneous disease and thus, highly variable across individual patients. This heterogeneity arises not only due to the diversity of genetic and molecular aberrations in primary breast cancer cells but also due to the diversity of cellular populations that inhabit the breast tumor microenvironment. Although IL-6 levels are higher in breast tumors and patient sera, the precise source of this IL-6 remains elusive. Importantly, many breast tumor stromal cells provide a paracrine source of IL-6 for breast cancer cells within the breast tumor microenvironment. In addition, certain clinical subtypes of breast cancers and research models, such as ER α -negative primary breast cancers and ER α -negative breast cancer cell lines, produce excessive IL-6 (Figure 2). Therefore, ER α -negative breast cancer cells may supply the tumor microenvironment with IL-6 by means of autocrine IL-6 production to exacerbate the poor prognosis associated with this clinical subtype. It will be critical to determine the specific cellular source of breast tumor-associated IL-6 to advance our understanding of this pleiotropic cytokine in breast cancer progression and metastasis. Moreover, this knowledge will facilitate the validation and subsequent clinical utility of current and novel targeted antagonists of the IL-6/STAT3 signaling network in breast cancer.

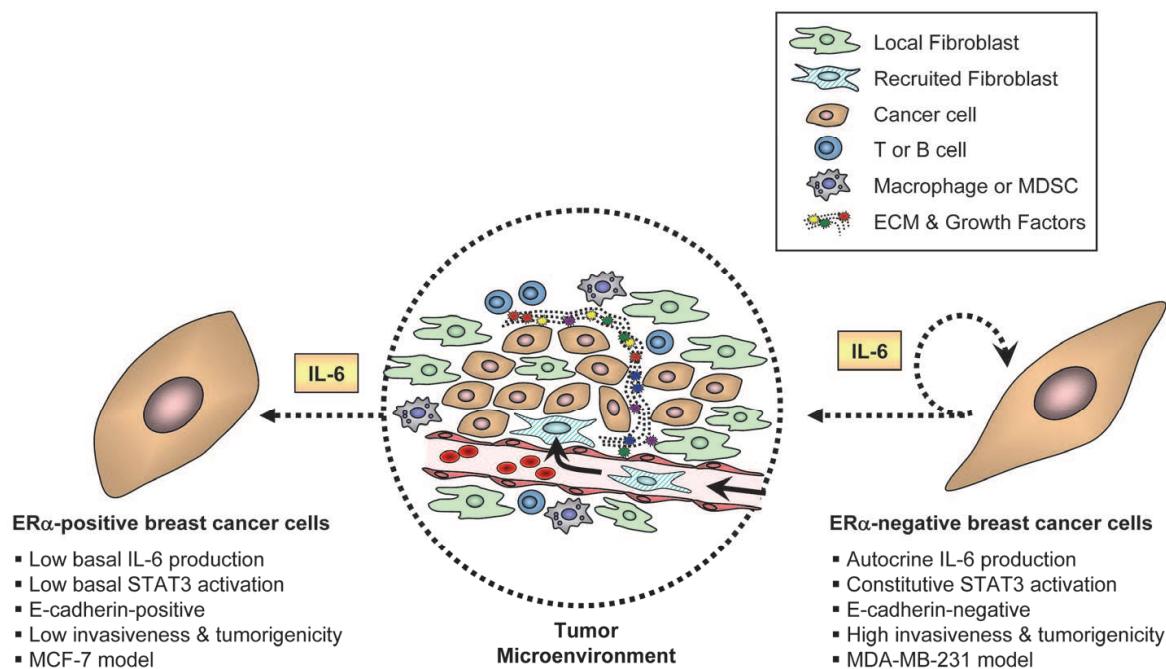


Fig. 2. Breast cancer cell ER α status dictates paracrine vs. autocrine IL-6 utilization.

10. References

- Anderson WF, Matsuno R (2006) Breast cancer heterogeneity: a mixture of at least two main types? *J Natl Cancer Inst* 98:948-951.
- Bachelot T, Ray-Coquard I, Menetrier-Caux C, Rastkha M, Duc A, Blay JY (2003) Prognostic value of serum levels of interleukin 6 and of serum and plasma levels of vascular endothelial growth factor in hormone-refractory metastatic breast cancer patients. *Br J Cancer* 88:1721-1726.
- Balkwill F, Mantovani A (2001) Inflammation and cancer: back to Virchow? *Lancet* 357:539-545.
- Becker C, Fantini MC, Schramm C, Lehr HA, Wirtz S, Nikolaev A, *et al.* (2004) TGF-beta suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. *Immunity* 21:491-501.
- Behrens J, Mareel MM, Van Roy FM, Birchmeier W (1989) Dissecting tumor cell invasion: epithelial cells acquire invasive properties after the loss of uvomorulin-mediated cell-cell adhesion. *J Cell Biol* 108:2435-2447.
- Bennermo M, Held C, Stemme S, Ericsson CG, Silveira A, Green F, *et al.* (2004) Genetic predisposition of the interleukin-6 response to inflammation: implications for a variety of major diseases? *Clin Chem* 50:2136-2140.
- Berishaj M, Gao SP, Ahmed S, Leslie K, Al-Ahmadie H, Gerald WL, *et al.* (2007) Stat3 is tyrosine-phosphorylated through the interleukin-6/glycoprotein 130/Janus kinase pathway in breast cancer. *Breast Cancer Res* 9:R32.
- Blick T, Widodo E, Hugo H, Waltham M, Lenburg ME, Neve RM, *et al.* (2008) Epithelial mesenchymal transition traits in human breast cancer cell lines. *Clin Exp Metastasis* 25:629-642.

- Brackenbury R, Rutishauser U, Edelman GM (1981) Distinct calcium-independent and calcium-dependent adhesion systems of chicken embryo cells. *Proc Natl Acad Sci U S A* 78:387-391.
- Braun S, Pantel K, Muller P, Janni W, Hepp F, Kentenich CR, *et al.* (2000) Cytokeratin-positive cells in the bone marrow and survival of patients with stage I, II, or III breast cancer. *N Engl J Med* 342:525-533.
- Butler TP, Gullino PM (1975) Quantitation of cell shedding into efferent blood of mammary adenocarcinoma. *Cancer Res* 35:512-516.
- Buyse M, Loi S, van't Veer L, Viale G, Delorenzi M, Glas AM, *et al.* (2006) Validation and clinical utility of a 70-gene prognostic signature for women with node-negative breast cancer. *J Natl Cancer Inst* 98:1183-1192.
- Cavarretta IT, Neuwirt H, Untergasser G, Moser PL, Zaki MH, Steiner H, *et al.* (2007) The antiapoptotic effect of IL-6 autocrine loop in a cellular model of advanced prostate cancer is mediated by Mcl-1. *Oncogene* 26:2822-2832.
- Cavarretta IT, Neuwirt H, Zaki MH, Steiner H, Hobisch A, Nemeth JA, *et al.* (2008) Mcl-1 is regulated by IL-6 and mediates the survival activity of the cytokine in a model of late stage prostate carcinoma. *Adv Exp Med Biol* 617:547-555.
- Chauhan D, Uchiyama H, Akbarali Y, Urashima M, Yamamoto K, Libermann TA, *et al.* (1996) Multiple myeloma cell adhesion-induced interleukin-6 expression in bone marrow stromal cells involves activation of NF-kappa B. *Blood* 87:1104-1112.
- Chavey C, Bibeau F, Gourgou-Bourgade S, Burlinchon S, Boissiere F, Laune D, *et al.* (2007) Oestrogen receptor negative breast cancers exhibit high cytokine content. *Breast Cancer Res* 9:R15.
- Cheng GZ, Zhang WZ, Sun M, Wang Q, Coppola D, Mansour M, *et al.* (2008) Twist is transcriptionally induced by activation of STAT3 and mediates STAT3 oncogenic function. *J Biol Chem* 283:14665-14673.
- Clevenger CV (2004) Roles and regulation of stat family transcription factors in human breast cancer. *Am J Pathol* 165:1449-1460.
- Crichton MB, Nichols JE, Zhao Y, Bulun SE, Simpson ER (1996) Expression of transcripts of interleukin-6 and related cytokines by human breast tumors, breast cancer cells, and adipose stromal cells. *Mol Cell Endocrinol* 118:215-220.
- DeMichele A, Martin AM, Mick R, Gor P, Wray L, Klein-Cabral M, *et al.* (2003) Interleukin-6 -174G-->C polymorphism is associated with improved outcome in high-risk breast cancer. *Cancer Res* 63:8051-8056.
- DeNardo DG, Coussens LM (2007) Inflammation and breast cancer. Balancing immune response: crosstalk between adaptive and innate immune cells during breast cancer progression. *Breast Cancer Res* 9:212.
- Diel IJ, Kaufmann M, Costa SD, Holle R, von Minckwitz G, Solomayer EF, *et al.* (1996) Micrometastatic breast cancer cells in bone marrow at primary surgery: prognostic value in comparison with nodal status. *J Natl Cancer Inst* 88:1652-1658.
- Dorff TB, Goldman B, Pinski JK, Mack PC, Lara PN, Jr., Van Veldhuizen PJ, Jr., *et al.* (2010) Clinical and correlative results of SWOG S0354: a phase II trial of CNT0328 (siltuximab), a monoclonal antibody against interleukin-6, in chemotherapy-pretreated patients with castration-resistant prostate cancer. *Clin Cancer Res* 16:3028-3034.

- Dunn GP, Old LJ, Schreiber RD (2004) The immunobiology of cancer immunosurveillance and immunoediting. *Immunity* 21:137-148.
- Erez N, Truitt M, Olson P, Arron ST, Hanahan D (2010) Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting Inflammation in an NF-kappaB-Dependent Manner. *Cancer Cell* 17:135-147.
- Galien R, Garcia T (1997) Estrogen receptor impairs interleukin-6 expression by preventing protein binding on the NF-kappaB site. *Nucleic Acids Res* 25:2424-2429.
- Gebauer G, Fehm T, Merkle E, Beck EP, Lang N, Jager W (2001) Epithelial cells in bone marrow of breast cancer patients at time of primary surgery: clinical outcome during long-term follow-up. *J Clin Oncol* 19:3669-3674.
- Giuliani N, Sansoni P, Girasole G, Vescovini R, Passeri G, Passeri M, et al. (2001) Serum interleukin-6, soluble interleukin-6 receptor and soluble gp130 exhibit different patterns of age- and menopause-related changes. *Exp Gerontol* 36:547-557.
- Gonzalez-Zuloeta Ladd AM, Arias Vasquez A, Witteman J, Uitterlinden AG, Coebergh JW, Hofman A, et al. (2006) Interleukin 6 G-174 C polymorphism and breast cancer risk. *Eur J Epidemiol* 21:373-376.
- Guilford P (1999) E-cadherin downregulation in cancer: fuel on the fire? *Mol Med Today* 5:172-177.
- Guo Y, Nemeth J, O'Brien C, Susa M, Liu X, Zhang Z, et al. (2010) Effects of siltuximab on the IL-6-induced signaling pathway in ovarian cancer. *Clin Cancer Res* 16:5759-5769.
- Hartsock A, Nelson WJ (2008) Adherens and tight junctions: structure, function and connections to the actin cytoskeleton. *Biochim Biophys Acta* 1778:660-669.
- Heimann R, Hellman S (2000) Individual characterisation of the metastatic capacity of human breast carcinoma. *Eur J Cancer* 36:1631-1639.
- Hess KR, Pusztai L, Buzdar AU, Hortobagyi GN (2003) Estrogen receptors and distinct patterns of breast cancer relapse. *Breast Cancer Res Treat* 78:105-118.
- Hirohashi S (1998) Inactivation of the E-cadherin-mediated cell adhesion system in human cancers. *Am J Pathol* 153:333-339.
- Hodge DR, Hurt EM, Farrar WL (2005) The role of IL-6 and STAT3 in inflammation and cancer. *Eur J Cancer* 41:2502-2512.
- Hunsucker SA, Magarotto V, Kuhn DJ, Kornblau SM, Wang M, Weber DM, et al. (2011) Blockade of interleukin-6 signalling with siltuximab enhances melphalan cytotoxicity in preclinical models of multiple myeloma. *Br J Haematol* 152:579-592.
- Hyafil F, Babinet C, Jacob F (1981) Cell-cell interactions in early embryogenesis: a molecular approach to the role of calcium. *Cell* 26:447-454.
- Irvin WJ, Jr., Carey LA (2008) What is triple-negative breast cancer? *Eur J Cancer* 44:2799-2805.
- James JJ, Evans AJ, Pinder SE, Gutteridge E, Cheung KL, Chan S, et al. (2003) Bone metastases from breast carcinoma: histopathological - radiological correlations and prognostic features. *Br J Cancer* 89:660-665.
- Jeanes A, Gottardi CJ, Yap AS (2008) Cadherins and cancer: how does cadherin dysfunction promote tumor progression? *Oncogene* 27:6920-6929.
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. *CA Cancer J Clin* 61:69-90.

- Jiang XP, Yang DC, Elliott RL, Head JF (2000) Reduction in serum IL-6 after vaccination of breast cancer patients with tumour-associated antigens is related to estrogen receptor status. *Cytokine* 12:458-465.
- Karkera J, Steiner H, Li W, Skradski V, Moser PL, Riethdorf S, *et al.* (2011) The anti-interleukin-6 antibody siltuximab down-regulates genes implicated in tumorigenesis in prostate cancer patients from a phase I study. *Prostate*.
- Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, *et al.* (2007) Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 449:557-563.
- Kenny PA, Bissell MJ (2003) Tumor reversion: correction of malignant behavior by microenvironmental cues. *Int J Cancer* 107:688-695.
- Kenny PA, Lee GY, Myers CA, Neve RM, Semeiks JR, Spellman PT, *et al.* (2007) The morphologies of breast cancer cell lines in three-dimensional assays correlate with their profiles of gene expression. *Mol Oncol* 1:84-96.
- Kim MY, Oskarsson T, Acharyya S, Nguyen DX, Zhang XH, Norton L, *et al.* (2009) Tumor self-seeding by circulating cancer cells. *Cell* 139:1315-1326.
- Kishimoto T (2006) Interleukin-6: discovery of a pleiotropic cytokine. *Arthritis Res Ther* 8 Suppl 2:S2.
- Klein CA (2009) Parallel progression of primary tumours and metastases. *Nat Rev Cancer* 9:302-312.
- Kominsky SL, Davidson NE (2006) A "bone" fide predictor of metastasis? Predicting breast cancer metastasis to bone. *J Clin Oncol* 24:2227-2229.
- Kopf M, Baumann H, Freer G, Freudenberg M, Lamers M, Kishimoto T, *et al.* (1994) Impaired immune and acute-phase responses in interleukin-6-deficient mice. *Nature* 368:339-342.
- Kowalski PJ, Rubin MA, Kleer CG (2003) E-cadherin expression in primary carcinomas of the breast and its distant metastases. *Breast Cancer Res* 5:R217-222.
- Kozlowski L, Zakrzewska I, Tokajuk P, Wojtukiewicz MZ (2003) Concentration of interleukin-6 (IL-6), interleukin-8 (IL-8) and interleukin-10 (IL-10) in blood serum of breast cancer patients. *Rocz Akad Med Bialymst* 48:82-84.
- Kurose K, Hoshaw-Woodard S, Adeyinka A, Lemeshow S, Watson PH, Eng C (2001) Genetic model of multi-step breast carcinogenesis involving the epithelium and stroma: clues to tumour-microenvironment interactions. *Hum Mol Genet* 10:1907-1913.
- Larue L, Ohsugi M, Hirchenhain J, Kemler R (1994) E-cadherin null mutant embryos fail to form a trophoblast epithelium. *Proc Natl Acad Sci U S A* 91:8263-8267.
- Lipponen P, Saarelainen E, Ji H, Aaltomaa S, Syrjanen K (1994) Expression of E-cadherin (E-CD) as related to other prognostic factors and survival in breast cancer. *J Pathol* 174:101-109.
- Litovkin KV, Domenyuk VP, Bubnov VV, Zaporozhan VN (2007) Interleukin-6 -174G/C polymorphism in breast cancer and uterine leiomyoma patients: a population-based case control study. *Exp Oncol* 29:295-298.
- Liu S, Ginestier C, Ou SJ, Clouthier SG, Patel SH, Monville F, *et al.* (2010) Breast cancer stem cells are regulated by mesenchymal stem cells through cytokine networks. *Cancer Res* 71:614-624.

- Lou Y, Preobrazhenska O, auf dem Keller U, Sutcliffe M, Barclay L, McDonald PC, *et al.* (2008) Epithelial-mesenchymal transition (EMT) is not sufficient for spontaneous murine breast cancer metastasis. *Dev Dyn* 237:2755-2768.
- Macedo LF, Sabnis G, Brodie A (2009) Aromatase inhibitors and breast cancer. *Ann N Y Acad Sci* 1155:162-173.
- Martin FT, Dwyer RM, Kelly J, Khan S, Murphy JM, Curran C, *et al.* (2010) Potential role of mesenchymal stem cells (MSCs) in the breast tumour microenvironment: stimulation of epithelial to mesenchymal transition (EMT). *Breast Cancer Res Treat* 124:317-326.
- Mbalaviele G, Dunstan CR, Sasaki A, Williams PJ, Mundy GR, Yoneda T (1996) E-cadherin expression in human breast cancer cells suppresses the development of osteolytic bone metastases in an experimental metastasis model. *Cancer Res* 56:4063-4070.
- Mironchik Y, Winnard PT, Jr., Vesuna F, Kato Y, Wildes F, Pathak AP, *et al.* (2005) Twist overexpression induces in vivo angiogenesis and correlates with chromosomal instability in breast cancer. *Cancer Res* 65:10801-10809.
- Moinfar F, Man YG, Arnould L, Bratthauer GL, Ratschek M, Tavassoli FA (2000) Concurrent and independent genetic alterations in the stromal and epithelial cells of mammary carcinoma: implications for tumorigenesis. *Cancer Res* 60:2562-2566.
- Moody SE, Perez D, Pan TC, Sarkisian CJ, Portocarrero CP, Sterner CJ, *et al.* (2005) The transcriptional repressor Snail promotes mammary tumor recurrence. *Cancer Cell* 8:197-209.
- Oh JW, Revel M, Chebath J (1996) A soluble interleukin 6 receptor isolated from conditioned medium of human breast cancer cells is encoded by a differentially spliced mRNA. *Cytokine* 8:401-409.
- Oka H, Shiozaki H, Kobayashi K, Inoue M, Tahara H, Kobayashi T, *et al.* (1993) Expression of E-cadherin cell adhesion molecules in human breast cancer tissues and its relationship to metastasis. *Cancer Res* 53:1696-1701.
- Onder TT, Gupta PB, Mani SA, Yang J, Lander ES, Weinberg RA (2008) Loss of E-cadherin promotes metastasis via multiple downstream transcriptional pathways. *Cancer Res* 68:3645-3654.
- Orimo A, Gupta PB, Sgroi DC, Arenzana-Seisdedos F, Delaunay T, Naeem R, *et al.* (2005) Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell* 121:335-348.
- Orimo A, Weinberg RA (2006) Stromal fibroblasts in cancer: a novel tumor-promoting cell type. *Cell Cycle* 5:1597-1601.
- Pantel K, Muller V, Auer M, Nusser N, Harbeck N, Braun S (2003) Detection and clinical implications of early systemic tumor cell dissemination in breast cancer. *Clin Cancer Res* 9:6326-6334.
- Parker C, Rampaul RS, Pinder SE, Bell JA, Wencyk PM, Blamey RW, *et al.* (2001) E-cadherin as a prognostic indicator in primary breast cancer. *Br J Cancer* 85:1958-1963.
- Pedersen KB, Nesland JM, Fodstad O, Maelandsmo GM (2002) Expression of S100A4, E-cadherin, alpha- and beta-catenin in breast cancer biopsies. *Br J Cancer* 87:1281-1286.
- Perez-Moreno M, Jamora C, Fuchs E (2003) Sticky business: orchestrating cellular signals at adherens junctions. *Cell* 112:535-548.

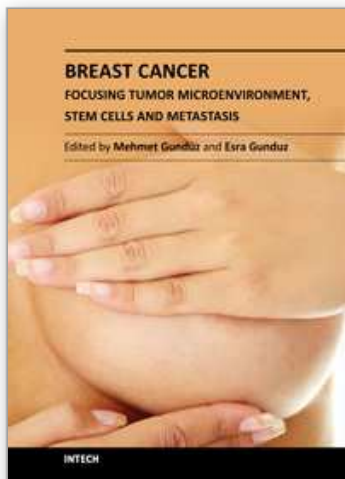
- Perou CM, Sorlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, *et al.* (2000) Molecular portraits of human breast tumours. *Nature* 406:747-752.
- Podsypanina K, Du YC, Jechlinger M, Beverly LJ, Hambardzumyan D, Varmus H (2008) Seeding and propagation of untransformed mouse mammary cells in the lung. *Science* 321:1841-1844.
- Puchalski T, Prabhakar U, Jiao Q, Berns B, Davis HM (2010) Pharmacokinetic and pharmacodynamic modeling of an anti-interleukin-6 chimeric monoclonal antibody (siltuximab) in patients with metastatic renal cell carcinoma. *Clin Cancer Res* 16:1652-1661.
- Radisky DC, Levy DD, Littlepage LE, Liu H, Nelson CM, Fata JE, *et al.* (2005) Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. *Nature* 436:123-127.
- Rakha EA, Green AR, Powe DG, Roylance R, Ellis IO (2006) Chromosome 16 tumor-suppressor genes in breast cancer. *Genes Chromosomes Cancer* 45:527-535.
- Rasanen K, Vaheri A (2010) Activation of fibroblasts in cancer stroma. *Exp Cell Res* 316:2713-2722.
- Rattigan Y, Hsu JM, Mishra PJ, Glod J, Banerjee D (2010) Interleukin 6 mediated recruitment of mesenchymal stem cells to the hypoxic tumor milieu. *Exp Cell Res* 316:3417-3424.
- Rose-John S, Scheller J, Elson G, Jones SA (2006) Interleukin-6 biology is coordinated by membrane-bound and soluble receptors: role in inflammation and cancer. *J Leukoc Biol* 80:227-236.
- Rossi JF, Negrier S, James ND, Kocak I, Hawkins R, Davis H, *et al.* (2010) A phase I/II study of siltuximab (CNTO 328), an anti-interleukin-6 monoclonal antibody, in metastatic renal cell cancer. *Br J Cancer* 103:1154-1162.
- Saha B, Chaiwun B, Imam SS, Tsao-Wei DD, Groshen S, Naritoku WY, *et al.* (2007) Overexpression of E-cadherin protein in metastatic breast cancer cells in bone. *Anticancer Res* 27:3903-3908.
- Salgado R, Junius S, Benoy I, Van Dam P, Vermeulen P, Van Marck E, *et al.* (2003) Circulating interleukin-6 predicts survival in patients with metastatic breast cancer. *Int J Cancer* 103:642-646.
- Sarrio D, Rodriguez-Pinilla SM, Hardisson D, Cano A, Moreno-Bueno G, Palacios J (2008) Epithelial-mesenchymal transition in breast cancer relates to the basal-like phenotype. *Cancer Res* 68:989-997.
- Sasser AK, Sullivan NJ, Studebaker AW, Hendey LF, Axel AE, Hall BM (2007) Interleukin-6 is a potent growth factor for ER-alpha-positive human breast cancer. *Faseb J* 21:3763-3770.
- Selander KS, Li L, Watson L, Merrell M, Dahmen H, Heinrich PC, *et al.* (2004) Inhibition of gp130 signaling in breast cancer blocks constitutive activation of Stat3 and inhibits in vivo malignancy. *Cancer Res* 64:6924-6933.
- Shoker BS, Jarvis C, Clarke RB, Anderson E, Hewlett J, Davies MP, *et al.* (1999) Estrogen receptor-positive proliferating cells in the normal and precancerous breast. *Am J Pathol* 155:1811-1815.
- Singh A, Purohit A, Wang DY, Duncan LJ, Ghilchik MW, Reed MJ (1995) IL-6sR: release from MCF-7 breast cancer cells and role in regulating peripheral oestrogen synthesis. *J Endocrinol* 147:R9-12.

- Sommers CL, Thompson EW, Torri JA, Kemler R, Gelmann EP, Byers SW (1991) Cell adhesion molecule uvomorulin expression in human breast cancer cell lines: relationship to morphology and invasive capacities. *Cell Growth Differ* 2:365-372.
- Song L, Rawal B, Nemeth JA, Haura EB (2010) JAK1 Activates STAT3 Activity in Non-Small-Cell Lung Cancer Cells and IL-6 Neutralizing Antibodies Can Suppress JAK1-STAT3 Signaling. *Mol Cancer Ther* 10:481-494.
- Sorlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, *et al.* (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci U S A* 98:10869-10874.
- Spaeth EL, Dembinski JL, Sasser AK, Watson K, Klopp A, Hall B, *et al.* (2009) Mesenchymal stem cell transition to tumor-associated fibroblasts contributes to fibrovascular network expansion and tumor progression. *PLoS One* 4:e4992.
- Studebaker AW, Storci G, Werbeck JL, Sansone P, Sasser AK, Tavolari S, *et al.* (2008) Fibroblasts isolated from common sites of breast cancer metastasis enhance cancer cell growth rates and invasiveness in an interleukin-6-dependent manner. *Cancer Res* 68:9087-9095.
- Suematsu S, Matsuda T, Aozasa K, Akira S, Nakano N, Ohno S, *et al.* (1989) IgG1 plasmacytosis in interleukin 6 transgenic mice. *Proc Natl Acad Sci U S A* 86:7547-7551.
- Sullivan NJ, Sasser AK, Axel AE, Vesuna F, Raman V, Ramirez N, *et al.* (2009) Interleukin-6 induces an epithelial-mesenchymal transition phenotype in human breast cancer cells. *Oncogene* 28:2940-2947.
- Tanaka T, Narazaki M, Kishimoto T (2011) Anti-interleukin-6 receptor antibody, tocilizumab, for the treatment of autoimmune diseases. *FEBS Lett.*
- Tarin D, Thompson EW, Newgreen DF (2005) The fallacy of epithelial mesenchymal transition in neoplasia. *Cancer Res* 65:5996-6000; discussion 6000-5991.
- van Rhee F, Fayad L, Voorhees P, Furman R, Lonial S, Borghaei H, *et al.* (2010) Siltuximab, a novel anti-interleukin-6 monoclonal antibody, for Castleman's disease. *J Clin Oncol* 28:3701-3708.
- Vannucchi AM, Bosi A, Glinz S, Pacini P, Linari S, Saccardi R, *et al.* (1998) Evaluation of breast tumour cell contamination in the bone marrow and leukapheresis collections by RT-PCR for cytokeratin-19 mRNA. *Br J Haematol* 103:610-617.
- Voorhees PM, Chen Q, Kuhn DJ, Small GW, Hunsucker SA, Strader JS, *et al.* (2007) Inhibition of interleukin-6 signaling with CNTO 328 enhances the activity of bortezomib in preclinical models of multiple myeloma. *Clin Cancer Res* 13:6469-6478.
- Weis WI, Nelson WJ (2006) Re-solving the cadherin-catenin-actin conundrum. *J Biol Chem* 281:35593-35597.
- Yang J, Stark GR (2008) Roles of unphosphorylated STATs in signaling. *Cell Res* 18:443-451.
- Yang J, Weinberg RA (2008) Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell* 14:818-829.
- Ye Y, Xiao Y, Wang W, Yearsley K, Gao JX, Shetuni B, *et al.* (2010) ERalpha signaling through slug regulates E-cadherin and EMT. *Oncogene* 29:1451-1462.
- Yu H, Pardoll D, Jove R (2009a) STATs in cancer inflammation and immunity: a leading role for STAT3. *Nat Rev Cancer* 9:798-809.

- Yu KD, Di GH, Fan L, Chen AX, Yang C, Shao ZM (2009b) Lack of an association between a functional polymorphism in the interleukin-6 gene promoter and breast cancer risk: a meta-analysis involving 25,703 subjects. *Breast Cancer Res Treat* 122:483-488.
- Zhang GJ, Adachi I (1999) Serum interleukin-6 levels correlate to tumor progression and prognosis in metastatic breast carcinoma. *Anticancer Res* 19:1427-1432.

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Cancer is the leading cause of death in most countries and its consequences result in huge economic, social and psychological burden. Breast cancer is the most frequently diagnosed cancer type and the leading cause of cancer death among females. In this book, we discussed characteristics of breast cancer cell, role of microenvironment, stem cells and metastasis for this deadly cancer. We hope that this book will contribute to the development of novel diagnostic as well as therapeutic approaches.

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