

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Gene Markers Representing Stem Cells and Cancer Cells for Quality Control

Shihori Tanabe

*National Institute of Health Sciences
Japan*

1. Introduction

Populations of cells have unique characteristics and gene markers representative of each cell type, and these features are useful for identifying cell characteristics. For example, the gene expression profile of cells differs at each stage of development and differentiation. This review focuses on gene expression in stem and cancer cells to investigate the possibility of identifying cancer stem cells by such markers.

Cancer stem cells show similarities to normal stem cells in terms of self-renewal and differentiation into multiple lineages. However, cancer stem cells have an indefinite potential for self-renewal that leads to malignant tumorigenesis. The origins of cancer stem cells are not completely clear but accumulation of gene mutations and cell niches are involved in their development. This article describes the gene expression patterns of stem and cancer cells with the aim of determining gene markers for diverse cell types and culture stages for quality control in cellular therapeutics.

2. The microarray quality control (MAQC) projects

Stem cells have varied gene and protein expression profiles and it is important to identify these profiles for quality control in disease treatment, as illnesses such as cancer may cause cell feature changes. The differentiation capacity of stem cells might be altered upon malignancy and there is the possibility that cancer comes from so-called cancer stem cells. Several methods are available to detect cell marker expression, such as surface protein marker detection, intracellular protein marker detection, and gene expression detection. The MAQC project, which is a collaborative effort conducted as part of the US Food and Drug Administration's Clinical Path Initiative for medical product development is useful to detect gene markers in cells (MAQC Consortium, 2006, 2010; Fan et al., 2010; Oberthuer et al., 2010; Huan et al., 2010; Luo et al., 2008; Parry et al., 2010; Shi et al., 2010; Miclaus et al., 2010; Hong et al., 2010; Tillinghast, 2010). It began in February 2005 and aims to describe the reliability and evaluate the performance of microarrays on several platforms.

MAQC-I mainly focuses on the technical aspects of gene expression analysis, whereas MAQC-II focuses on developing accurate and reproducible multivariate gene expression-based prediction models. Possible uses for gene expression data are vast, including diagnosis, early detection (screening), monitoring of disease progression, risk assessment,

prognosis, complex medical product characterisation and prediction of responses to treatment (with regard to safety or efficacy) with a drug or device labelling intent. The MAQC-II data model prediction is dependent upon endpoints, including preclinical toxicity, breast cancer, multiple myeloma and neuroblastoma. Some endpoints are highly predictive based on the nature of the data, and other endpoints are difficult to predict regardless of the model development protocol. Clear differences in proficiency exist between data analysis teams, and such differences are correlated with the level of team experience. The internal validation performance from well-implemented, unbiased cross-validation analyses shows a high degree of concordance with the external validation performance in a strictly blinded process, and many models with similar performance can be developed from a given data set (Table 1).

	MAQC-I	MAQC-II
Aim	To address the concerns about the reliability of microarray techniques	To develop and evaluate accurate and reproducible multivariate gene expression-based predictive model
Summary	The technical performance of microarrays as assessed in the project supports their continued use for gene expression profiling in basic and applied research and may lead to their use as a clinical diagnostic tool as well.	1) Model prediction performance was endpoint dependent. 2) There are clear differences in proficiency between data analysis teams (organisations). 3) The internal validation performance from well-implemented, unbiased cross-validation shows a high degree of concordance with the external validation performance in a strict blinding process. 4) Many models with similar performance can be developed from a given data set. 5) Application of good modelling practices appeared to be more important than the actual choice of a particular algorithm over the others within the same step in the modelling process.
Reference	MAQC Consortium (2006). The MicroArray Quality Control (MAQC) project shows inter- and intraplatform reproducibility of gene expression measurements, Nature Biotechnology, Vol.24, No.9, (September 2006), pp.1151-1161	MAQC Consortium (2010). The MicroArray Quality Control (MAQC)-II study of common practices for the development and validation of microarray-based predictive models, Nature Biotechnology, Vol.28, No.8, (August 2010), pp.827-838

Table 1. The Microarray Quality Control (MAQC) projects

Applying good modelling practice seems to be more important than the actual choice of a particular algorithm over the others within the same step in the modelling process. The order of the analysis process was as follows: design, pilot study or internal validation, and pivotal study or external validation. Observations based on an analysis of the MAQC-II dataset may be applicable to other diseases. (MAQC Consortium, 2010)

3. Gene markers for stem cells

3.1 Cell surface marker genes

The stem cell expression profile varies in differentiated cells. The expression pattern may change depending on differentiation or malignancy of the disease. Endothelial cells in glioblastomas have unique gene expression profiles, and the differences between glioblastomas and lower grade gliomas suggest a more complex ontogeny of the glioblastoma endothelium (Wang et al., 2010). Quantitative *in situ* hybridisation analyses have revealed that fluorescence-activated cell-sorted CD105+ (one of the human endothelial markers) cells with more than 3 copies of the epidermal growth factor receptor (EGFR) amplicon or the centromeric portion of chromosome 7 are similar to the proportion of tumour cells with similar aberrations. CD133 is a cell surface glycoprotein, which has been used as a possible cancer stem cell marker. CD133 is also expressed in haematopoietic stem cells.

3.2 Genes for mesenchymal stem cells

3.2.1 Genes expressed in mesenchymal stem cells

CD29, CD44, CD49a-f, CD51, CD54, CD71, CD73, CD90, CD105, CD106, CD166, Stro-1 and MHC class I molecules are positively expressed in human bone marrow derived mesenchymal stem cells (MSCs), whereas CD11b, CD14, CD18, CD19, CD31, CD34, CD40, CD45, CD56, CD79 α , CD80, CD86 and HLA-DR are not (Chamberlain et al., 2007; Kuroda et al., 2010; Pittenger et al., 1999; Kumar et al., 2008; Tsai et al., 2007) (Table 2). Specific markers for MSCs have not been identified. A combination of gene markers may be important to characterise the features of MSCs.

3.2.2 Genes representing the mesenchymal stem cell culture stage

MSCs are often used for treating graft-versus-host disease (GVHD) (Weng et al., 2010; Le Blanc et al., 2008), suggesting that an infusion of MSCs may be an effective therapy for patients with steroid-resistant acute GVHD. The necdin homologue (mouse) (*NDN*), EPH receptor A5 (*EPHA5*), nephroblastoma overexpressed gene (*NOV*) and runt-related transcription factor 2 (*RUNX2*) are possible markers to describe culture status, including growth capacity and differentiation (Tanabe et al., 2008). *EPHA5* and *NOV* are upregulated in late culture stage of human MSCs, whereas *NDN* and *RUNX2* are downregulated (GEO series, Tanabe et al., 2008, accession GSE7637 and GSE7888).

NOV expression in prostate cancer tends to be involved in cancer conditions, based on human prostate cancer gene expression data (Best et al., 2005). This expression is upregulated in androgen-independent primary human prostate cancer compared to untreated human prostate cancer (GEO series, Best, 2005, accession GSE2443). *NOV* might be a candidate marker for identifying the cancer state.

Human MSCs have been reported to promote growth of osteosarcomas, a common primary malignant bone tumour (Bian et al., 2010). In addition, interleukin-6 plays an important role

in maintaining the ‘stemness’ of human MSCs and the proliferation of Saos-2. It is possible that the secretion of interleukin-6 and interaction of human MSCs and Saos-2 through interleukin-6 are essential for their proliferation; this suggests that humoural factors participate in stem cell development.

	positive	negative
Human MSC	CD29, CD44, CD49a-f, CD51, CD54, CD71, CD73, CD90, CD105, CD106, CD166, Stro-1 and MHC class I molecules	CD11b, CD14, CD18, CD19, CD31, CD34, CD40, CD45, CD56, CD79 α , CD80, CD86, HLA-DR

Table 2. Positive and negative human mesenchymal stem cell genes

3.3 Marker genes in neural stem cells

Glial fibrillary acidic protein (GFAP), Musashi, nestin, excitatory amino acid transporter 1 (GLAST), PDGFR- α and CD133 are known to be expressed in neural stem cells and are generally used as their markers (Yadirgi & Marino, 2009; Jackson & Alvarez-Buylla, 2008; Gage, F.H., 2000). The expression of nestin, Dlx2, DVC, PSA-NCAM and β III tubulin has been reported to be altered during cell development. Another report has shown that Sox2, which is believed to be a marker of the nervous system, is expressed in embryonic neural stem cells and other multipotent cells, and that it plays an essential role in mouse brain neurogenesis (Ferri et al., 2004). Recently, it had been revealed that glia have the ability to act as stem cells (Robel et al., 2011). In this study, GFAP, vimentin, nestin, tenascin-C (TNC) and brain lipid-binding protein (BLBP) are described as immature markers; whereas, glycogen granules, glutamine synthetase, S100 β , GLAST and excitatory amino acid transporter 2 (GLT1), which is involved in glutamate uptake and conversion to glutamine, are indicated to be common glial markers. GLAST and GLT1 are shared markers in astrocytes, radial glial in the developing central nervous system, and neural stem cells in the adult mammalian brain. GFAP, S100 β and aldehyde dehydrogenase 1 family, member L1 (ALDL1H1) are expressed in both astrocytes and neural stem cells.

4. Genes for reprogramming stem cells

Recent findings have indicated that a set of 4 genes such as POU class 5 homeobox 1 (*POU5F1*, *OCT3/4*), sex-determining region Y-box 2 (*SOX2*), Kruppel-like factor 4 (gut) (*KLF4*) and v-myc myelocytomatosis viral oncogene homologue (avian) (*MYC*, *c-Myc*) induce fibroblast reprogramming into pluripotent stem cells (Takahashi et al., 2007; Park et al., 2008; Lowry et al., 2008). After determining that reprogramming genes actually exist to manipulate and modify human cells, these 4 genes, or some other set of genes such as *POU5F1* (*OCT4*), *SOX2* and *KLF4* have been used globally to produce induced pluripotent stem cells. Recently, somatic cells have been suggested to be directly reprogrammed without an induced pluripotent stem cell-mediated pathway but with culture condition modifications (Han et al, 2011). In that study, fibroblasts were infected with retrovirus expressing *Oct4*, *Sox2*, *Klf4* and *c-Myc*, and directly induced into epiblast stem cells by adding basic fibroblast

growth factor. The combination of gene expression and factors from outside the cells may play important roles in reprogramming cells.

4.1 Genes for generation of induced pluripotent stem (iPS) cells

Recently, it had been reported that *OCT4* is sufficient to induce alterations in the human keratinocyte differentiation pathway (Racila et al., 2011). Transfection of *OCT4*, using a plasmid, into human skin keratinocytes resulted in exhibited expression of endogenous embryonic genes and reduced genomic methylation. These *OCT4*-transfected cells could become neuronal and mesenchymal cell types. The cells have been shown to have characteristics of cultured smooth muscle or myofibroblast cells from a mesenchymal stem cell lineage. It is probable that partial reprogramming using several genes can induce transitions in cell phenotypes and features; hence, complete reprogramming of somatic cells into iPS cells would not always be required for the application of these cells in clinical therapy.

The characterization of human iPS cells, with respect to pluripotency and the ability for terminal differentiation, has been performed with 16 iPS cell lines (Boulting et al., 2011). This study revealed that all iPS cell lines examined, reprogrammed with *OCT4*, *SOX2* and *KLF4*, or *OCT4*, *SOX2*, *KLF4* and *c-MYC* showed the capacity to function as functional motor neurons after differentiation, although there was some variation in the expression of early pluripotency markers and the transgenes. iPS cell lines have been shown to express pluripotency markers, such as *NANOG*, *OCT4*, *SSEA3*, *SSEA4*, *TRA-1-60* and *TRA-1-81*.

4.2 Involvement of genome structure in reprogramming to iPS cells

Copy number variation has been reported to be involved in the reprogramming to pluripotency (Hussein et al., 2011). The comparison of copy number variations of different passages of human iPS cells with their fibroblast cell origins and with human embryonic stem (ES) cells revealed high copy number variation levels in early-passage human iPS cells. The number of copy number variations in human iPS cell lines decreases with an increase in the number of passage. This decrease during culture passages could be due to DNA repair mechanisms or mosaicism followed by selection. The authors proposed that *de novo* generated copy number variations create mosaicism that is followed by selection of less damaged cells during culturing, because DNA repair is not considered as a sufficient explanation of the rapid decrease in copy number variation.

4.3 Involvement of epigenetic modification and methylation in iPS cells

EMT has been shown to be associated with a stem cell phenotype (Mani et al., 2008; Battula et al., 2010; Polyak & Weinberg, 2009). The tumour suppressor *p53* has been suggested to regulate EMT and EMT-associated stem cell properties through transcriptional activation of miRNA (Chang et al., 2011). EMT and the reverse process, the mesenchymal-epithelial transition, are believed to be key elements in the regulation of embryogenesis. It has also been suggested that EMT activation is related to cancer progression and metastasis.

Recently, EMT has been shown to play a role in the acquisition of stem cell properties in normal and neoplastic cell populations. miRNAs are small non-coding RNA molecules and suppress gene expression by interacting with the 3'-untranslated regions (3' UTRs) of target mRNAs. miRNAs are known to be related to EMT and cancer. The study revealed that *p53* activates miR-200c, which is down-regulated in normal stem cell and neoplastic stem cell populations, and suppresses the EMT phenotype and stem cell properties represented in

CD24–CD44⁺ cell populations. The expression of mesenchymal stem cell markers, such as N-cadherin and ZEB1, has been shown to be suppressed by p53. The mRNA levels of KLF4 and BMI1, which are known as stemness-associated genes and RNA targets of miR-200c and miR-183, have been shown to be regulated by p53.

It has also been reported that the p53R175H mutant up-regulates Twist1 expression and promotes EMT in immortalized prostate cells (Kogan-Sakin et al., 2011). Inactivated or mutated p53 may result in the up-regulation of cell cycle progression genes, such as Twist1, which is a regulator of metastasis and EMT.

4.4 Epithelial-mesenchymal transition (EMT) and microRNAs (miRNAs)

iPS cells have been known to show reprogramming variability such as aberrant reprogramming of DNA methylation (Lister et al., 2010). From whole-genome, single-base-resolution DNA methylomic analyses of iPS cells and ES cells, the authors obtained new evidence showing that iPS cells are methylated during reprogramming, and the methylome of iPS cells generally resembles that of ES cells. In the study, a detailed interpretation of the data indicated that there were many differences in DNA methylation between ES cells and iPS cells. For example, many differentially methylated regions that were differentially methylated in either the iPS cell line or the ES cell line existed in several iPS cell lines.

5. Gene markers for cancer cells

5.1 Regulated genes in renal cancer

NCBI's Gene Expression Omnibus (GEO) database is a useful tool to profile gene expression and search for markers representing cell features (Edgar et al., 2002; Barrett, 2011). Renal tumour samples have been analysed using microarray (Yusenko et al., 2009). It was observed that loss of chromosomes 2, 10, 13, 17 and 21 discriminate chromophobe renal cell carcinomas from renal oncocytomas. These authors suggested that detecting chromosomal changes can be used for an accurate diagnosis in routine histology.

The gene expression profiles of the microarray data deposited in the GEO database (GEO series, Szponar et al., 2009, accession GSE11151) were analysed and caveolin 2 (CAV2), proteasome (prosome, macropain) subunit, beta type, 8 (large multifunctional peptidase 7) (PSMB8) major histocompatibility complexes, class I, F (HLA-F), major histocompatibility complex, class I, B (HLA-B), apoptosis enhancing nuclease (AEN), major histocompatibility complex, class I, G (HLA-G), and tumour necrosis factor receptor superfamily member 10b (TNFRSF10B) are upregulated by more than three-fold in renal tumours (n = 62) compared to normal kidney (n = 5) (Table 3).

The collaborative genome-wide study for renal cell carcinoma using SNP detection techniques has revealed that genome loci on 2p21 and 11q13.3 are genomic regions associated with renal cell carcinoma (Purdue et al., 2011). From this study, EPAS1, encoding hypoxia-inducible-factor-2 alpha at 2p21 and SCARB1, the scavenger receptor class B, member 1 at 12q24.31, were identified as feature genes that have single nucleotide polymorphism mutations in renal cell carcinoma.

5.2 Genes expressed in leukaemia

A model in which human cancers are believed to be generated hierarchically from self-renewing cancer stem cells has been reported. Human acute myeloid leukaemia (AML) is a disease that relates to the model, and AML stem cell-targeting therapy has been developed

(Majeti, 2011; Jin et al., 2006). CD25, CD32, CD44, CD47, CD96, CD123 and CLL-1 are expressed on the surface of AML stem cells. Of these genes, CD44 is suggested to be a cancer stem cell marker.

Gene Symbol	Gene Title	Chromosomal location	Entrez Gene	Function
CAV2	Caveolin-2	Chr7q31.1	858	Protein homodimerisation activity
PSMB8	Proteasome (prosome, macropain) subunit, beta type, 8 (large multifunctional peptidase 7)	Chr6p21.3	5696	ATP binding, MHC class I protein binding
HLA-F	Major histocompatibility complex, class I, F	Chr6p21.3	3134	MHC class I receptor activity
HLA-B	Major histocompatibility complex, class I, B	Chr6p21.3	3106	MHC class I receptor activity
AEN	Apoptosis enhancing nuclease	Chr15q26.1	64782	Exonuclease activity
HLA-G	Major histocompatibility complex, class I, G	Chr6p21.3	3135	MHC class I receptor activity
TNFRSF10B	Tumour necrosis factor receptor superfamily, member 10b	Chr8p22-p21	8795	TRAIL binding, caspase activator activity

Table 3. Upregulated genes in renal tumours (Ref. GSE11151)

The concept of cancer stem cell is important in explaining cancer development from the viewpoint of stem cells (Clevers, 2011; Wang & Shen, 2011). The cancer stem cells for leukaemia were identified from a study showing that CD34+CD38– fractions of cells derived from acute myeloid leukaemia had the capacity to initiate engraftment in immunodeficient mice (Lapidot et al., 1994; Bonnet & Dick, 1997). It is known that deletion or mutation of *IKZF1* (*IKAROS*), *PAX5*, *EBF1* and *CDKN2A/B* are involved in BCR-ABL1 lymphoblastic leukaemia (Mullighan et al., 2008; Mullighan et al., 2009). The function of human BCR-ABL1 lymphoblastic leukaemia-initiating cells in human lymphoblastic leukaemia has been studied from the point of view of genome diversity (Notta et al., 2011). Functional and genetic analysis of Philadelphia chromosome acute lymphoblastic leukaemia (Philadelphia-positive [Ph+] ALL) revealed that the frequencies of

genetic alterations in *IKZF1* (84%), *CDKN2A/B* (50%) and *PAX5* (50%) were consistent with those reported in previous studies. Complete deletion of *IKZF1* was observed in both aggressive and non-aggressive groups; whereas, there were differences in the frequencies of deletion of the *CDKN2A/B* and *PAX5* genes, which may provide markers for malignancy.

On the other hand, CD44 has been identified as a key regulator of leukaemic stem cells in AML (Jin et al., 2006). It was suggested that elimination of leukaemic stem cells, cells capable of initiating and maintaining the leukaemic clonal hierarchy, was required for a permanent cure of AML. Hence, stimulation with a CD44-specific antibody has been reported to result in the elimination of leukaemic stem cells.

5.3 Genes expressed in glioblastomas

Wang *et al.* hypothesised that the CD133+ fraction is related to the endothelial differentiation potential and analysed cells from a series of glioblastomas fractionated as follows: (1) CD144+ (endothelial cadherin)/CD133-, (2) CD144+/CD133+, (3) CD133+/CD144-, (4) CD133-/CD144-. The results of quantitative PCR with a reverse transcription analysis demonstrated that VEGFR2 and endothelial progenitor marker CD34 are enriched in the CD144+/CD133- and in the CD144+/CD133+ populations. CD105 was negative in the CD133+ and CD144+ fractions. It was also shown that mice who were administered an injection of CD133+/CD144- or CD133+/CD144+ cells from a primary glioblastoma revealed tumours.

A new mechanism was suggested in which tumour vascularisation occurs through endothelial differentiation of glioblastoma stem-like cells (Ricci-Vitiani et al., 2010). The differentiation of cancer stem-like cells may be involved in cancer malignancy, and it is possible to predict or diagnose the malignant stage of cancer using stem cell markers for quality control.

In a genome-wide association study (GWAS) of four case series on 2,251 test patients and 6,097 control patients of European ancestry, LIM domain only 1 (*LMO1*) at 11p15.4 was found to be associated with neuroblastoma and malignancy (Wang et al., 2011). An integrative genomics study to demonstrate that common genetic polymorphisms associated with cancer tendencies are also related to genomic regions that have possibility of somatic alterations which in turn influence tumour progression, revealed that mutation in *LMO1* may also be a candidate indicator of a malignant phenotype.

5.4 Surface markers for cancer stem cells

Several markers have been reported for identification of cancer stem cells (Clevers, 2011). CD19 as a surface marker for B cell malignancies, CD20 and ATP-binding cassette transporter B5 (ABCB5) for melanoma, and the following molecules for cancer stem cells in the respective cancer type have been reported: CD24 for pancreas/lung cancer, CD34 for hematopoietic malignancies, CD44 for breast/liver/head and neck/pancreas cancer, CD90 for liver cancer, CD133 for brain/colorectal/lung/liver cancer and epithelial cell adhesion molecule (EpCAM)/epithelial-specific antigen (ESA) for colorectal/pancreatic cancer (Ebben et al., 2010).

5.5 Cancer stem cell hypothesis

Cancer stem cells have capacity for self-renewal, which is also the feature to normal stem cells. Cancer stem cells are also capable of generating malignant tumours, and this property

may differentiate them from normal stem cells. The origin of the cancer stem cells has not been fully revealed, however, there is a model in which cancer stem cells occur by normal stem cells or normal cells by the accumulation of gene mutations. The process of cancer stem cell derivation is considered to be involved with niche which is microenvironment around normal stem cells.

There are two models to explain tumourigenesis. The first model is stochastic model in which all cells have capacity of tumourigenesis, but the probability to enter into tumourigenesis cell cycle is relatively low. The second model is hierarchy theory in which only small population of cells in cancer has capacity of tumourigenesis and generate tumour with high probability, which lead to cancer stem cell hypothesis.

It is also notable that cancer stem cells are not necessarily related to the cell of origin in a cancer (Visvader, 2011). Although the cell of origin for a particular tumour may have the capacity to differentiate into a mature cell, cancer stem cells have the ability to maintain tumourigenesis according to the cell-of-origin model.

6. Conclusion

The recent development in molecular biology and bioinformatics technology has revealed stem cell features and their candidate marker genes. Gene expression profiles change widely and dramatically with cell development, various culture conditions and disease status. The each cell type has different gene expression profile after being differentiated, and it is known that the expression pattern alters in each disease status. Even though it seems that the stemness has distinct feature in gene expression, the cell population show various gene expression patterns in each cell lineage or even in each subset of the cell.

Until recently, targeting cancer stem cells in cancer therapy was rare because the proportion of these cells in cancer was considered very low and retaining the feature of cancer stem cells in vitro was difficult. The stem cell-targeted therapy including cancer treatment will be expected to progress further in the near future, and the role of markers would become much greater. It is important to know the precise feature and gene expression pattern for quality control in the cell-targeted therapy.

7. Acknowledgments

The author acknowledges Dr. Yoji Sato, Dr. Takayoshi Suzuki, Dr. Kazuhiro Suzuki, Dr. Taku Nagao, Dr. Teruhide Yamaguchi, Dr. Yasuo Ohno, Dr. Eriko Uchida, Dr. Tadashi Oshizawa and Dr. Masahiro Nishijima.

8. References

- Barrett, T.; Troup, D.B.; Wilhite, S.E.; Ledoux, P.; Evangelista, C.; Kim, I.F.; Tomashevsky, M.; Marshall, K.A.; Phillippy, K.H.; Sherman, P.M.; Muetter, R.N.; Holko, M.; Ayanbule, O.; Yefanov, A. & Soboleva, A. (2011). NCBI GEO: archive for functional genomics data sets—10 years on, *Nucleic Acids Research*, Vol.39, Database issue, pp.D1005-D1010 (January 2011)
- Battula, V.L.; Evans, K.W.; Hollier, B.G.; Shi, Y.; Marini, F.C.; Ayyanan, A.; Wang, R-Y.; Briskin, C.; Guerra, R.; Andreeff, M. & Mani, S.A. (2010). Epithelial-mesenchymal transition-derived cells exhibit multilineage differentiation potential similar to mesenchymal stem cells, *Stem Cells*, Vol.28, No.8, (August 2010), pp.1435-1445

- Best, C.J.; Gillespie, J.W.; Yi, Y.; Chandramouli, G.V.; Perlmutter, M.A.; Gathright, Y.; Erickson, H.S.; Georgevich, L.; Tangrea, M.A.; Duray, P.H.; González, S.; Velasco, A.; Linehan, W.M.; Matusik, R.J.; Price, D.K.; Figg, W.D.; Emmert-Buck, M.R. & Chuaqui, R.F. (2005). Molecular Alterations in Primary Prostate Cancer after Androgen Ablation Therapy, *Clinical Cancer Research*, Vol.11, No.19, pp.6823-6834 (October 2005)
- Bian, Z-Y.; Fan, Q-M.; Li, G.; Xu, W-T. & Tang, T-T. (2010). Human mesenchymal stem cells promote growth of osteosarcoma: Involvement of interleukin-6 in the interaction between human mesenchymal stem cells and Saos-2, *Cancer Science*, Vol.101, No.12, (December 2010), pp.2554-2560
- Bonnet, D. & Dick, J.E. (1997). Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell, *Nature Medicine*, Vol.7, (July 1997), pp.730-737
- Boulting, G.L.; Kiskinins, E.; Croft, G.F.; Amoroso, M.W.; Oakley, D.H.; Wainger, B.J.; Williams, D.J.; Kahler, D.J.; Yamaki, M.; Davidow, L.; Rodolfa, C.T.; Dimos, J.T.; Mikkilineni, S.; MacDermott, A.B.; Woolf, C.J.; Henderson, C.E.; Wichterle, H. & Eggan, K. (2011). A functionally characterized test set of human induced pluripotent stem cells, *Nature Biotechnology*, Vol.29, (March 2011), pp.279-286
- Chamberlain, G.; Fox, J.; Ashton, B. & Middleton J. (2007). Differentiation Capacity, Immunological Features, and Potential for Homing, *Stem Cells*, Vol.25, pp.2739-2749 (November 2007)
- Chang, C-J.; Chao, C-H.; Xia, W.; Yang, J-Y.; Xiong, Y.; Li, C-W.; Yu, W-H.; Rehman, S.K.; Hsu, J.L.; Lee, H-H.; Liu, M.; Chen, C-T.; Yu, D. & Hung, M-C. (2011). P53 regulates epithelial-mesenchymal transition and stem cell properties through modulating miRNAs, *Nature Cell Biology*, Vol.13, No.3, (March 2011), pp.317-323
- Clevers, H. (2011). The cancer stem cell: premises, promises and challenges, *Nature Medicine*, Vol.17, No.3, (March 2011), pp.313-319
- Ebben, J.D.; Treisman, D.M.; Zomiak, M.; Kutty, R.G; Clark, P.A. & Kuo, J.S. (2010). The cancer stem cell paradigm: a new understanding of tumor development and treatment, *Expert Opinion on Therapeutic Targets*, Vol.14, No.6, (June 2010), pp.621-632
- Edgar, R.; Domrachev, M. & Lash, A.E. (2002). Gene Expression Omnibus: NCBI gene expression and dhybridization array data repository, *Nucleic Acids Research*, Vol.30, pp.207-210 (January 2002)
- Fan, X.; Lobenhofer, E.K.; Chen, M.; Shi, W.; Huang J.; Luo, J.; Zhang, J.; Walker, S.J.; Chu, T.M.; Li, L.; Wolfinger, R.; Bao, W.; Paules, R.S; Bushel, P.R.; Li, J.; Shi, T.; Nikolskaya, T.; Nikolsky, Y.; Hong, H.; Deng, Y.; Cheng, Y.; Fang, H.; Shi, L. & Tong, W. (2010). Consistency of predictive signature genes and classifiers generated using different microarray platforms, *Pharmacogenomics Journal*, Vol.10, No.4, pp.247-257 (August 2010)
- Ferri, A.L.; Cavallaro, M.; Braidà, D.; Di Cristofano, A.; Canta, A.; Vezzani, A.; Ottolenghi, S.; Pandolfi, P.P.; Sala, M.; DeBiasi, S. & Nicolis, S.K. (2004). Sox2 deficiency causes neurodegeneration and impaired neurogenesis in the adult mouse brain, *Development*, Vol.131, No.15, (August 2004), pp.3805-3819
- Gage, F.H. (2000). Mammalian neural stem cells, *Science*, Vol.287, (February 2000), pp.1433-1438

- Han, D.W.; Greber, B.; Wu, G.; Tapia, N.; Araúzo-Bravo, M.J.; Ko, K.; Bernemann, C.; Stehling, M. & Schöler, H.R (2011). Direct reprogramming of fibroblasts into epiblast stem cells, *Nature Cell Biology*, Vol.13, No.1, pp. 66-71 (January 2011)
- Hong, H.; Shi, L.; Su, Z.; Ge, W.; Jones, W.D.; Czika, W.; Miclaus, K.; Lambert, C.G.; Vega, S.C.; Zhang, J.; Ning, B.; Liu, J.; Green, B.; Xu, L.; Fang, H.; Perkins, R.; Lin, S.M.; Jafari, N.; Parl, K.; Ahn, T.; Chierici, M.; Furlanello, C.; Zhang, L.; Wolfinger, R.D.; Goodsaid, F. & Tong, W. (2010). Assessing sources of inconsistencies in genotypes and their effects on genome-wide association studies with HapMap samples, *Pharmacogenomics Journal*, Vol.10, No.4, pp.364-374 (August 2010)
- Huan, J.; Shi, W.; Zhang, J.; Chou, J.W.; Paules, R.S.; Gerrush, K.; Li, J.; Luo, J.; Wolfinger, R.D.; Bao, W.; Chu, T.M.; Nikolsky, Y.; Nikolskaya, T.; Dosymbekov, D.; Tsyganova, M.O.; Shi, L.; Fan, X.; Corton, J.C.; Chen, M.; Cheng, Y.; Tong, W.; Fnag, H. & Bushel, P.R. (2010). Genomic indicators in the blood predict drug-induced liver injury, *Pharmacogenomics Journal*, Vol.10, No.4, pp.267-277 (August 2010)
- Hussein, S.M.; Batada, N.N.; Vuoristo, S.; Ching, R.W.; Autio, R.; Närvä, E.; Ng, S.; Sourour, M.; Hämäläinen, R.; Olsson, C.; Lundin, K.; Mikkola, M.; Trokovic, R.; Peitz, M.; Brüstle, O.; Bazett-Jones, D.P.; Alitalo, K.; Lahesmaa, R.; Nagy, A. & Otonkoski, R. (2011). Copy number variation and selection during reprogramming to pluripotency, *Nature*, Vol.471, (March 2011), pp.58-62
- Jin, L.; Hope, K.J.; Zhani, Q.; Smadja-Joffe, F. & Dicke, J.E. (2006). Targeting of CD44 eradicates human acute myeloid leukemic stem cells, *Nature Medicine*, Vol.12, No.10, pp.1167-1174 (October 2006)
- Kogan-Sakin, I.; Tabach, Y.; Buganim, Y.; Molchadsky, A.; Solomon, H.; Madar, S.; Kamer, I.; Stambolsky, P.; Shelly, A.; Goldfinger, N.; Valsesia-Wittman, S.; Puisieux, A.; Zundeleovich, A.; Gal-Yam, E.N.; Avivi, C.; Barshack, I.; Brait, M.; Sidransky, D.; Domany, E. & Rotter, V. Mutant p53R175H upregulates Twist1 expression and promotes epithelial-mesenchymal transition in immortalized prostate cells, *Cell Death & Differentiation*, Vol.18, No.2, (February 2011), pp.271-281
- Kumar, S.; Chanda, D. & Ponnazhagan, S. (2008). Therapeutic potential of genetically modified mesenchymal stem cells, *Gene Therapy*, Vol.15, pp.711-715 (March 2008)
- Kuroda, Y.; Kitada, M.; Wakako, S.; Nishikawa, K.; Tanimura, Y.; Makinoshima, H.; Goda, M.; Akashi, H.; Inutsuka, A.; Niwa, A.; Shigemoto, T.; Nabeshima, Y.; Nakahata, T.; Nabeshima, Y.; Fujiyoshi, Y. & Dezawa, M. (2010). Unique multipotent cells in adult human mesenchymal cell populations, *Proceedings of the National Academy of Sciences of the United States of America*, Vol.107, No.19, pp.8639-8643 (May 2010)
- Lapidot, T.; Sirard, C.; Vormoor, J.; Murdoch, B.; Hoang, T.; Caceres-Cortes, J.; Minden, M.; Paterson, B.; Caligiuri, M.A.; Dick, J.E. (1994). A cell initiating human acute myeloid leukaemia after transplantation into SCID mice, *Nature*, Vol.367, (February 1994), pp.645-658
- Le Blanc, K.; Frassoni, R.; Ball, L.; Locatelli, F.; Roelofs, H.; Lewis, I.; Lanino, E.; Sundberg, B.; Bernardo, M.E.; Remberger, M.; Dini, G.; Egeler, R.M.; Bacigalupo, A.; Fibbe, W. & Ringdén, O. (2008). Mesenchymal stem cells for treatment of steroid-resistant, severe, acute graft-versus-host disease: a phase II study. *The Lancet*, Vol.371, pp.1579-1586 (May 2008)
- Lister, R.; Pelizzola, M.; Kida, Y.S.; Hawkins, R.D.; Nery, J.R.; Hon, G.; Antosiewicz-Bourgets, J.; O'Malley, R.; Castanon, R.; Klugman, S.; Downes, M.; Yu, R.; Stewart, R.; Ren, B.; Thomson, J.A.; Evans, R.M. & Ecker, J.R. (2010). Hotspots of

- aberrant epigenomic reprogramming in human induced pluripotent stem cells, *Nature*, Vol. 471, No.7336, (March 2011), pp.46-47
- Lowry, W.E.; Richter, L.; Yachechko, R.; Pyle, A.D.; Tchieu, J.; Sridharan, R.; Clark, A.T. & Plath, K. (2008). Generation of human induced pluripotent stem cells from dermal fibroblasts, *Proceedings of the National Academy of Sciences of the United States of America*, Vol.105, No.8, pp.2883-2888 (February 2008)
- Luo, J.; Schumacher, M.; Scherer, A.; Sanoudou, D.; Megherbi, D.; Davison, T.; Shi, T.; Tong, W.; Shi, L.; Hong, H.; Zhao, C.; Elloumi, F.; Shi, W.; Thomas, R.; Lin, S.; Tillinghast, G.; Liu, G.; Zhou, Y.; Herman, D.; Li, Y.; Deng, Y.; Fnag, H.; Bushel, P.; Woods, M. & Zhang, J. (2010). A comparison of batch effect removal methods for enhancement of prediction performance using MAQC-II microarray gene expression data, *Pharmacogenomics Journal*, Vol.10, No.4, pp.278-291 (August 2010)
- Majeti, R. (2011). Monoclonal antibody therapy directed against human acute myeloid leukemia stem cells, *Oncogene*, Vol.30, No.3, pp.1009-1019 (March 2011)
- Mani, S.A.; Guo, W.; Liao, M-J.; Eaton, E.N.; Ayyanan, A.; Zhou, A.Y.; Brooks, M.; Reinhard, F.; Zhang, C.C.; Shipitsin, M.; Campbell, L.L.; Polyak, K.; Briskin, C.; Yang, J. & Weinberg, R.A. (2009). The epithelial-mesenchymal transition generates cells with properties of stem cells, *Cell*, Vol.133, No.4, (May 2008), pp.704-715
- MAQC Consortium (2006). The MicroArray Quality Control (MAQC) project shows inter- and intraplatform reproducibility of gene expression measurements *Nature Biotechnology*, Vol.24, No.9, (September 2006), pp.1151-1161
- MAQC Consortium (2010). The MicroArray Quality Control (MAQC)-II study of common practices for the development and validation of microarray-based predictive models, *Nature Biotechnology*, Vol.28, No.8, (August 2010), pp.827-838
- Miclaus, K.; Wolfinger, R.; Vega, S.; Chierici, M.; Furlanello, C.; Lambert, C.; Hong, H.; Zhang, L.; Yin, S. & Goodsaid, F. (2010). Batch effects in the BRLMM genotype calling algorithm influence GWAS results for the Affymetrix 500K array, *Pharmacogenomics Journal*, Vol.10, No.4, pp.336-346 (August 2010)
- Mullighan, C.G.; Miller, C.B.; Radtke, I.; Phillips, L.A.; Dalton, J.; Ma, J.; White, D.; Hughes, T.P.; Le Beau, M.M.; Pui, C-H.; Relling, M.V.; Shurtleff, S.A. & Downing, J.R. (2008). BCR-ABL1 lymphoblastic leukaemia is characterized by the deletion of Ikaros, *Nature*, Vol.453, (May 2008), pp.110-114
- Mullighan, C.G.; Su, X.; Zhang, J.; Radtke, I.; Phillips, L.A.A.; Miller, C.B.; Ma, J.; Liu, W.; Cheng, C.; Schulman, B.A.; Harvey, R.C.; Chen, I-M.; Clifford, R.J.; Carroll, W.L.; Reaman, G.; Bowman, W.P.; Devidas, M.; Gerhard, D.S.; Yang, W.; Relling, M.V.; Pharm, D.; Shurtleff, S.A.; Campana, D.; Borowitz, M.J.; Pui, C-H.; Smith, M.; Hunger, S.P.; Willman, C.L.; Downing, J.R. (2009). Deletion of IKZF1 and prognosis in acute lymphoblastic leukemia, *New England Journal of Medicine*, Vol.360, No.5, (January 2009), pp.470-480
- Notta, F.; Mullighan, C.G.; Wang, J.C.; Poepl, A.; Doulatov, S.; Phillips, L.A.; Ma, J.; Minden, M.D.; Downing, J.R. & Dick, J.E. (2011). Evolution of human BCR-ABL1 lymphoblastic leukemia-initiating cells, *Nature*, Vol.469, (January 2011), pp.362-367
- Oberthuer, A.; Juraeva, D.; Li, L.; Kahlert, Y.; Westermann, F.; Eils, R.; Berthold, F.; Shi, L.; Wolfinger, R.D.; Fischer, M. & Brors, B. (2010). Comparison of performance of one-color and two-color gene-expression analyses in predicting clinical endpoints of neuroblastoma patients, *Pharmacogenomics Journal*, Vol.10, No.4, pp.258-266 (August 2010)

- Park, I.-H.; Zhao, R.; West, J.A.; Yabuuchi, A.; Huo, H.; Ince, T.A.; Lerou, P.H.; Lensch, M.W. & Daley, G.Q. (2008). Reprogramming of human somatic cells to pluripotency with defined factors, *Nature*, Vol.451, pp.141-146 (January 2008)
- Parry, R.M.; Jones, W.; Stokes, T.H.; Phan, J.H.; Moffitt, R.A.; Fnag, H.; Shi, L.; Oberthuer, A.; Fischer, M.; Tong, W. & Wang, M.D. (2010). K-Nearest neighbor models for microarray gene expression analysis and clinical outcome prediction, *Pharmacogenomics Journal*, Vol.10, No.4, pp.292-309 (August 2010)
- Pittenger, M.F.; Mackay, A.M.; Beck, S.C.; Jaiswal, R.K.; Dounglas, R.; Mosca, J.D.; Moorman, M.A.; Simonetti, D.W.; Craig, S. & Marshak, D.R. (1999). Multilineage Potential of Adult Human Mesenchymal Stem Cells, *Science*, Vol.184, pp.143-147 (Apr 1999)
- Polyak, K. & Weinberg R.A. (2009). Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits, *Nature Reviews Cancer*, Vol.9, No.4, (April 2009), pp.265-273
- Purdue, M.P.; Johansson, M.; Zelenika, D.; Toro, J.R.; Scelo, G.; Moore, L.E.; Prokhortchouk, E.; Wu, X.; Kiemeny, L.A.; Gaborieau, V.; Jacobs, K.B.; Chou, W-H.; Zaridze, D.; Matveev, V.; Lubinski, J.; Trubicka, J.; Szeszenia-Dabrowska, N.; Lissowska, J.; Rudnai, P.; Fabianova, E.; Bucur, A.; Bencko, V.; Foretova, L.; Janout, V.; Boffetta, P.; Colt, J.S.; Davis, F.G.; Schwartz, K.L.; Banks, R.E.; Selby, P.J.; Harnden, P.; Berg, C.D.; Hsing, A.W.; Grubb, R.L.III; Boeing, H.; Vineis, P.; Clavel-Chapelon, F.; Palli, D.; Tumino, R.; Krogh, V.; Panico, S.; Duell, E.J.; Quirós, J.R.; Sanchez, M-J.; Navarro, C.; Ardanaz, E.; Dorronsoro, M.; Khaw, K-T.; Allen, N.E.; Bueno-de-Mesquita, H.B.; Peeters, P.H.M.; Trichopoulos, D.; Linseisen, J.; Ljungberg, B.; Overvad, K.; Tjønneland, A.; Romieu, I.; Riboli, E.; Mukeria, A.; Shangina, O.; Stevens, V.L.; Thun, M.J.; Diver, W.R.; Gapstur, S.M.; Pharoah, P.D.; Easton, D.F.; Albanes, D.; Weinstein, S.J.; Virtamo, J.; Vatten, L.; Hveem, K.; Njølstad, I.; Tell, G.S.; Stoltenberg, C.; Kumar, R.; Koppova, K.; Cussenot, O.; Benhamou, S.; Oosterwijk, E.; Vermeulen, S.H.; Aben, K.K.H.; van der Marel, S.L.; Ye, Y.; Wood, C.G.; Pu, X.; Mazur, A.M.; Boulygina, E.S.; Chekanov, N.N.; Foglio, M.; Lechner, D.; Gut, I.; Heath, S.; Blanche, H.; Hutchinson, A.; Thomas, G.; Wang, Z.; Yeager, M.; Fraumeni, J.F.Jr; Skryabin, K.G.; McKay, J.D.; Rothman, N.; Chanock, S.J.; Lathrop, M. & Brennan, P. (2011). Genome-wide association study of renal cell carcinoma identifies two susceptibility loci on 2p21 and 11q13.3, *Nature Genetics*, Vol.43, No.1, (January 2011), pp.60-65
- Racila, D.; Winter, M.; Said, M.; Tomanek-Chalkley, A.; Wiechert, S.; Eckert, R.L. & Bickenbach, J.R. (2011). Transient expression of OCT4 is sufficient to allow human keratinocytes to change their differentiation pathway, *Gene Therapy*, Vol.18, No.3, (March 2011), pp.294-303
- Ricci-Vitiani, L.; Pallini, R.; Biffoni, M.; Todaro, M.; Invernici, G.; Cenci, T.; Maira, G.; Parati, E.A; Stassi, G.; Larocca, L.M. & Maria, R.D. (2010). Tumour vascularization via endothelial differentiation of glioblastoma stem-like cells, *Nature*, Vol.468, pp.824-828 (December 2010)
- Robel, S.; Berninger, B. & Götz, M. (2011). The stem cell potential of glia: lessons from reactive gliosis, *Nature Reviews Neuroscience*, Vol.12, (February 2011), pp.88-104
- Shi, W.; Bessarabova, M.; Dosymnekov, D.; Dezso, Z.; Nikolskaya, T.; Dudoladova, M.; Serebryiskaya, T.; Bugrim, A.; Guryanov, A.; Brennan, R.J.; Shah, R.; Dopazo, J.;

- Cheng, M.; Deng, Y.; Shi, T.; Jurman, G.; Furlanello, C.; Thomas, R.S.; Corton, J.C.; Tong, W.; Shi, L. & Nikolsky, Y. (2010). Functional analysis of multiple genomic signatures demonstrates that classification algorithms choose phenotype-related genes, *Pharmacogenomics Journal*, Vol.10, No.4, pp.310-323 (August 2010)
- Suh, H.; Consiglio, A.; Ray, J.; Sawai, T. & D'Amour, K.A. (2007). In vivo fate analysis reveals the multipotent and self-renewal capacities of Sox2+ neural stem cells in the adult hippocampus, *Cell Stem Cell*, Vol.1, (November 2007), pp.515-528
- Takahashi, K.; Tanabe, K.; Ohnuki, M.; Narita, M.; Ichisaka, T.; Tomoda, K. & Yamanaka, S. (2007). Induction of pluripotent stem cells from adult human fibroblasts by defined factors, *Cell*, Vol.131, No.5, pp.861-872 (November 2007)
- Tanabe, S.; Sato, Y.; Suzuki, T.; Suzuki, K.; Nagao, T. & Yamaguchi, T. (2008). Gene expression profiling of human mesenchymal stem cells for identification of novel markers in early- and late-stage cell culture, *Journal of Biochemistry*, Vol.144, No.3, pp.399-408 (September 2008)
- Tillinghast, G.W. (2010). Microarrays in the clinic, *Nature Biotechnology*, Vol.28, No.8, pp.810-812 (August 2010)
- Tsai, M.S.; Hwang, S.M.; Chen, K.D.; Lee, Y.S.; Hsu, L.W.; Chang, Y.J.; Wang, C.N.; Peng, H.H.; Chang, Y.L.; Chao, A.S.; Chang, S.D.; Lee, K.D.; Wang, T.H.; Wang, H.S. & Soong, Y.K. (2007). Functional Network Analysis of the Transcriptomes of Mesenchymal Stem Cells Derived from Amniotic Fluid, Amniotic Membrane, Cord Blood, and Bone Marrow, *Stem Cells*, Vol.25, No.10, pp.2511-2523 (October 2007)
- Visvader, J.E. (2011). Cells of origin in cancer, *Nature*, Vol.469, (January 2011), pp.314-322
- Wang, K. ; Diskin, S.J. ; Zhang, H. ; Attiyeh, E.F. ; Winter, C. ; Hou, C. ; Schnepp, R.W. ; Diamond, M. ; Bosse, K. ; Mayes, P.A. ; Glessner, J. ; Kim, C. ; Frackelton, E. ; Garriss, M. ; Wang, Q. ; Glaberson, W. ; Chiavacci, R. ; Nguyen, L. ; Jagannathan, J. ; Saeki, N. ; Sasaki, H. ; Grant, S.F.A., ; Iolascon, A. ; Mosse, Y.P. ; Cole, K.A. ; Li, H. ; Devoto, M. ; McGrady, P.W. ; London, W.B. ; Capasso, M. ; Rahman, N. ; Hakonarson, H. & Maris, J.M. (2011). Integrative genomics identifies LMO1 as a neuroblastoma oncogene, *Nature*, Vol.469, (January 2011), pp.216-220
- Wang, R.; Chadalavada, K.; Wilshire, J.; Kowalik, U.; Hovinga, K.E.; Geber, A.; Fligelman, B.; Leversha, M.; Brennan, C. & Tabar, V. (2010). Glioblastoma stem-like cells give rise to tumour endothelium, *Nature*, Vol.468, pp.829-833 (December 2010)
- Wang, Z.A. & Shen, M.M. (2011). Revisiting the concept of cancer stem cells in prostate cancer, *Oncogene*, Vol.30, No.11, (March 2011), pp.1261-1271
- Weng, J.Y.; Du, X.; Geng, S.X.; Peng, Y.W.; Wang, Z.; Lu, Z.S.; Wu, S.J.; Luo, C.W.; Guo, R.; Ling, W.; Deng, C.X.; Liao, P.J. & Xiang, A.P. (2010). Mesenchymal stem cell as salvage treatment for refractory chronic GVHD, *Bone Marrow Transplantation*, Vol.45, pp.1732-1740 (December 2010)
- Yadirgi, G. & Marino, S. (2009). Adult neural stem cells and their role in brain pathology, *Journal of Pathology*, Vol.217, (January 2009), pp.242-253
- Yusenko, M.V.; Kuiper, R.P.; Boethe, T.; Ljungberg, B.; van Kessel, A.G. & Kovacs, G. (2009). High-resolution DNA copy number and gene expression analyses distinguish chromophobe renal cell carcinomas and renal oncocytomas, *BMC Cancer*, Vol.9, No.152 (May 2009)



Wide Spectra of Quality Control

Edited by Dr. Isin Akyar

ISBN 978-953-307-683-6

Hard cover, 532 pages

Publisher InTech

Published online 07, July, 2011

Published in print edition July, 2011

Quality control is a standard which certainly has become a style of living. With the improvement of technology every day, we meet new and complicated devices and methods in different fields. Quality control explains the directed use of testing to measure the achievement of a specific standard. It is the process, procedures and authority used to accept or reject all components, drug product containers, closures, in-process materials, packaging material, labeling and drug products, and the authority to review production records to assure that no errors have occurred. The quality which is supposed to be achieved is not a concept which can be controlled by easy, numerical or other means, but it is the control over the intrinsic quality of a test facility and its studies. The aim of this book is to share useful and practical knowledge about quality control in several fields with the people who want to improve their knowledge.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Shihori Tanabe (2011). Gene Markers Representing Stem Cells and Cancer Cells for Quality Control, Wide Spectra of Quality Control, Dr. Isin Akyar (Ed.), ISBN: 978-953-307-683-6, InTech, Available from: <http://www.intechopen.com/books/wide-spectra-of-quality-control/gene-markers-representing-stem-cells-and-cancer-cells-for-quality-control>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen