We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900
Open access books available

185,000
International authors and editors

200M
Downloads

Our authors are among the

154
Countries delivered to

TOP 1%
most cited scientists

12.2%
Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



11

Brine Shrimp Diversity in China Based on DNA Barcoding

Hangxiao Zhang, Jing Sun, Weiwei Wang and Jun Yu
CAS Key Laboratory of Genome Sciences and Information,
Beijing Institute of Genomics, Chinese Academy of Sciences
China

1. Introduction

Taxonomy, the science that deals with the study of identifying, grouping, and naming organisms according to their established natural relationship, is the basis of all biological studies. Biological and observation-based classification is still generally the best known form of taxonomy since 1735 when Carl Linnaeus published the great book - Species Plantarum, and it is an empirical science mostly based on morphological difference. With the development of science and technology, scientists have discovered many methods to identification new species and other tools or definitions for species classification, such as biochemical identification (Farmer et al. 1985), cytotaxonomic identification (Le Berre et al. 1985), chromosomal DNA fingerprinting (Owen 1989), restriction fragment length polymorphism (RFLP) (Sakaoka et al. 1992), and PCR-based DNA fingerprints (Matsuki et al. 2003). Among others, molecular or genetic approaches to identify species have been proposed and extensively used (Yamamoto 1992; Zhou et al. 2003).

1.1 DNA barcoding

The study of biodiversity lays the foundation for all biological studies, especially the classification of species, and the ways to do it have never stopped since Linnaeus. Traditional morphology-based taxonomy has its limitations, such as when facing mimetic polymorphism, and it mainly depends on the expertise of taxonomists, and there is little doubt that evidence at molecular levels should be complementary and of necessary. As the development of molecular biology, the idea of molecular taxonomy has been propounded and gradually accepted by related scientific communities. The standard molecular identification system was initiated during 1990s by using PCR-based and sequencing-based approaches (Frézal et al. 2008). Taken the advantage of the two powerful technologies in accuracy and convenience, DNA sequence signatures provide adequate "barcodes" for species identification, and "DNA barcoding" has been widely used in studies for speciation (Ghebremedhin et al. 2008; Sullivan et al. 1996), phylogenetics andevolution (Göker et al. 2009;Wood et al. 2000), and molecular ecology (Govan et al. 1996; Valentini et al. 2009) as well as for the classification of both pathogenic microbes (Beckmann 1999) and normal microbiomes (Holzapfel et al. 2001).

DNA barcoding is an ultimate and direct approach for molecular taxonomy, depending on the complexity of sequence signatures used, especially in distinguishing species with nearly

identical morphological features, thereby helping to establish legitimate phylogenetic relationships and to reveal evolutionary histories. The concept of DNA barcoding was first advocated by Arnot in 1993 (Arnot et al. 1993) and its new era began in 2003, marked by the establishment of the Consortium for the Barcode of Life (CBOL, http://barcoding.si.edu). This initiation was put forward and promoted by researchers at the University of Guelph in Ontario, Canada in 2004. The aim of this project is to create a universal protocol for an eukaryotic species inventory based on a standard molecular approach. Up to this date, the Consortium has more than 150 member organizations from 45 countries, including natural history museums, zoos, botanical gardens, university departments as well as private companies and governmental organizations (Frézal & Leblois 2008; Schindel et al. 2005). The DNA Barcode of Life Database (BOLD, http://www.boldsystems.org), an informatic workbench aiding the acquisition, storage, analysis, and publication of DNA barcode records, has been developed since 2004 and was officially established in 2007 (Ratnasingham et al. 2007). This database provides an integrated bioinformatic platform for the acquaintance, collection, and analysis of basic barcoding data and facilitates the development of DNA barcoding.

1.1.1 What is DNA barcoding?

In theory, nucleotide sequences of nuclear and organellar origins are natural 'barcodes' that are unique to each organism on earth. Therefore, a 15-bp nucleotide sequence creates 4^{15} (1 billion) combinations that would be sufficient for the differentiation of the estimated 10-15 million species (Butchart et al. 2010; Perrings 1996). However, practically, species are related and their genome sequences are often homologous, depending on their evolutionary distances. In addition, the rates of molecular evolution vary dramatically across taxa and even at different positions in a given genome. In the latter case, the main task of DNA barcoding is to find a sequence fragment that is evolutionarily less selected and serves as a unique barcode for species identification.

There had not been a universal barcode sequence for all species yet, especially across distant lineage boundaries, but several candidate genes are commonly used for phylogenetic analysis, such as mitochondrial 16S rRNA gene, mitochondrial cytochrome *b* gene, and the mitochondrial cytochrome c oxidase subunit 1(*COI*) which can serve as the core of global bio-identification system for animals (Hebert et al. 2003a, 2003b). A 648-bp segment at 58-705 from the 5'end of this gene is chosen as the barcode segment. *COI* gene is an ideal model to evaluate the evolution rate, as its third-position nucleotides show a high incidence of base substitutions but its amino acid sequence changes rather slowly as compared to other mitochondrial genes. As a result, on the one hand, the evolution of this gene is rapid enough for identification of not only closely related species, but also phylogeographic groups within a single species, and on the other hand, it is possible to place an unidentified species into higher taxonomic categories (from phyla to orders) (Hebert et al. 2003a). Although a unified opinion has not been reached on a single barcoding DNA segment chosen for taxological studies (Lin et al. 2009), the *COI* gene-based identification system has been proven superior within taxonomic groups of Protista (Chantangsi et al. 2007; Evans et al. 2007) and animals, including gastropods (Hebert et al. 2003b; Remigio et al. 2003), ants (Smith et al. 2005), butterflies (Hebert et al. 2004a), birds (Hebert et al. 2004b), spiders (Greenstone et al. 2005), fish (Ward et al. 2005), worms (Ferri et al. 2009; Jennings et al. 2010), Crustacea (Lefébure et al. 2006), and very recently primates (Nijman et al. 2010).

1.1.2 Examples of DNA barcoding applications

DNA barcoding has been successfully used for the taxonomy of invertebrate and vertebrate animals as well as microbes, including bacteria (Siddall et al. 2009), fungi (Kelly et al. 2011; Stockinger et al. 2010), Protista (Chantangsi et al. 2007; Evans et al. 2007), and algae taxonomies (Saunders 2005). In the past three years, an increasing number of studies has been focused on DNA-barcoding of plants (He et al. 2010; Kress et al. 2007; Lahaye et al. 2008). Since there is not yet a universally accepted DNA barcode for plants, many strategies have been proposed, based either on a single chloroplast segment (Hollingsworth et al. 2009; Lahaye et al. 2008) or a combination of multiple segments (He et al. 2010; Kress & Erickson 2007). Examples of DNA barcoding studies are summarized in Table 1 including DNA barcodes for animals, plants, fungi, and protists. As mentioned previously, there are advantages and limitations among the barcodes with respect to specific applications.

Organism group	DNA barcode	References
Animals	COI, 28SrRNA, cob	(Hebert et al. 2004a; Hogg et al. 2004; Ward et al. 2005; Zhang et al. 2011)
Plants	COI, trnL, matK, rbcL, trnH-psbA, ITS	(Kress et al. 2005; Savolainen et al. 2008; Shaw et al. 2011; Specht et al. 2007)
Fungi	COI, ITS, LSU, mtSSU, beta-tubulin	(Porter et al. 2008; Schussler et al. 2010; Seifert et al. 2007; Summerbell et al. 2007; Tedersoo et al. 2008)
Protists	COI, ITS	(Brodie et al. 2006; Keeling et al. 2010; Pawlowski et al. 2010; Saunders 2005; Stern et al. 2010)

Table 1. Applications of DNA barcoding technology

1.1.3 Advantages and drawbacks of DNA barcoding

There are several obvious advantages in the currently used DNA barcoding system. First, it uses a standard procedure that can be applied universally to relevant research fields. It is of great utility in conservation biology and can also be applied to samples where traditional morphological methods are unable to define, including species identification based on eggs and larval (Wang et al. 2008) and analysis of stomach contents or excreta to determine food webs. Another advantage of DNA barcoding comes from the rapid and cost-efficient acquisition of molecular data, enabling large-scale species identification (Frézal & Leblois 2008), whereas conventional taxonomy is time consuming, and in some cases it is almost impossible to apply (Rusch et al. 2007). Therefore, it is important to be able to improve large surveys aiming at unknown species detection and identification of pathogenic species with medical, ecological, and agronomical significance (Ball et al. 2008; Barth et al. 2006). Particularly, DNA barcoding becomes necessary when morphological traits do not adequately discriminate species (Caron et al. 2009; Guo et al. 2010; Kauffman et al. 2003; Kumar et al. 2006) or if species have polymorphic life cycles and/or exhibit pronounced phenotypic plasticity (Pegg et al. 2006; Randrianiaina et al. 2007).

However, controversies about DNA barcoding still remain. Although DNA barcoding was proposed initially as a method for species identification, to better achieve this goal, it needs

be validated intensively, especially in choosing the best candidate sequences that are both universal and highly variable among species. The first question is: what are these sequences: nuclear, mitochondrial, or chloroplast? An idea to use a simple sequence from mtDNA has been dismissed. It is not adequate to be used as a sole source for species-definition due to following genetic factors: reduced effective population size and introgression, maternal inheritance, recombination, inconsistent mutation rate, heteroplasmy, and compounding evolutionary processes (Meier et al. 2006; Rubinoff et al. 2006). Until now, there has not been an universal DNA barcode for all organisms and we have not found a single gene that is conserved enough and also exhibits appropriate divergence for all species regardless where they come from (Hickerson et al. 2006; Rubinoff et al. 2006; Song et al. 2008). The validity of DNA barcoding therefore lies on establishing reference sequences from taxonomically confirmed specimens, which will acquire an integration of morphological and molecular based taxonomy data, as well as decent cooperation among sample collection, such as museums, zoos, and research institutes (De Hoog et al. 2008). This approach is closest to what has been termed "integrative taxonomy" (Dayrat 2005; Will et al. 2005). DNA sequences in combination with traditional character sets are used in a complementary fashion to define and describe species (Heethoff et al. 2011; Padial et al. 2010; Pereira et al. 2010).

1.1.4 Recent progresses in DNA barcoding

Recently, the approach of DNA barcoding has been greatly revived to increase accuracy and sensitivity, and the major improvements are focused on using more than one barcoding strategies for a better identification of specific species (Aliabadian et al. 2009; Ferri et al. 2009; Lin et al. 2009; Nassonova et al. 2010). Shatters et al improved DNA barcoding by using different regions of *COI* gene to do biotype-specific barcoding (Shatters et al. 2009). As the sequencing technology developed rapidly in the past few years, sequence-based DNA barcoding also advanced rapidly, such as cap analysis of gene expression (CAGE) using an ultra high-throughput sequencer (Maeda et al. 2008), to show biodiversity (Creer 2010; Fonseca et al. 2010; Mitsui et al. 2010), and the ArkChip strategy for highly-resolved patterns of intraspecific evolution and a multi-species (Carr et al. 2008). Several new techniques have been implemented, and all based on the sequencing of individual DNA molecules (with or without an amplification step) in massive and parallel ways (Table 2, Figure 1). The high accuracy, throughput, and efficiency make the identification of genome sequences unique to different species and life forms easy.

The processes that apply next-generation sequencers to DNA barcoding are expected to be more complex than what has been anticipated. For instance, the classical DNA barcode is defined to be a fragment around 650bp but the effective read lengths of the next-G sequencers are actually shorter than it at present time. Progress has been made in recent studies, where smaller DNA fragments, called mini-barcode, of *COI* gene or rDNA were used for accurate species identification (Hajibabaei et al. 2006; Pawlowski & Lecroq 2010). Researches show that more than 90% and 95% success rates were achieved by using 100-bp and 250-bp barcodes, respectively (Meusnier et al. 2008). Although biodiversity studies based on next-G sequencing technologies were emerged in 2006, (Ley et al. 2006; Sogin et al. 2006), most of the studies have been done with the Roche/454 system (Hajibabaei et al. 2011; Meyer et al. 2007; Porazinska et al. 2009) and mainly for environmental samples (Deagle et al. 2010;Fire et al. 2007; Hajibabaei et al. 2011). More recently, the upgrading

speed of different sequencing platforms, such as those of Illumina and Life Technologies, has been very impressive and the read lengths of these new versions of sequencers are getting longer (Table 2). They may also one day be used for biodiversity study when their read length is increased to ~100bp and more.

Sequencer	Company	Read length	Reads per run	Total output	Time per run
Solexa	Illumina	75 bases	60 million	4 Gb	6.5 days
SOLiD	Life Technologies	50 bases	85 million	4 Gb	6 days
454 GS FLX	Roche Diagnostics	500 bases	1 million	0.5 Gb	8 h

Note: The recent machine and software upgrades from Illumina (such as HiSeq2000) and Life Technologies (such as 5500XL) promise ~100-fold increases in the total outputs of raw data.

Table 2. Comparison of next-generation sequencing platforms.

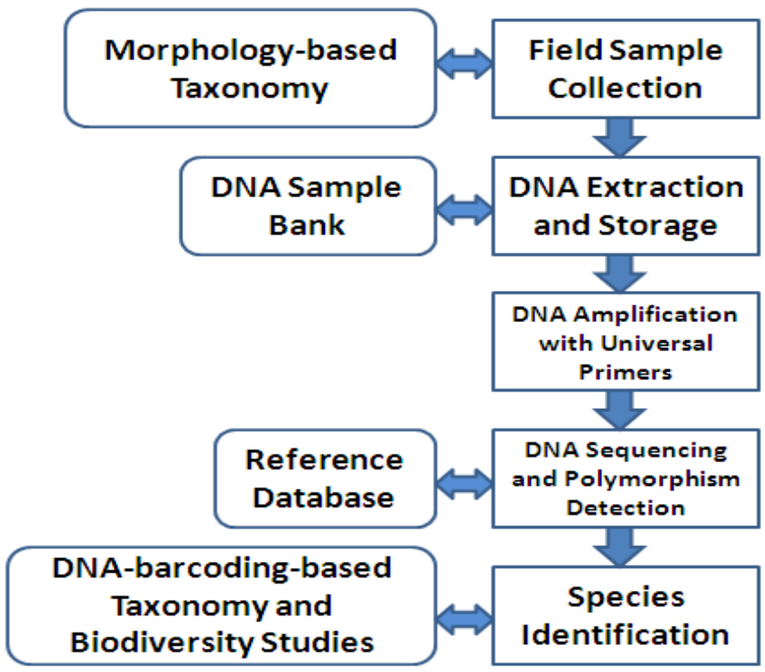


Fig. 1. Methodology for analyzing biodiversity based on high-throughtput DNA sequencing.

1.2 A case study on *Artemia* (Crustacea, Anostraca) in China

Brine shrimp or *Artemia* (Crustacea, Anostraca) is a worldwide living species well-adapted to survive in very harsh hypersaline environments, such as salty lakes and lagoons (Clegg et

al. 2009), it typically shows enormous diversity at the genus level in terms of their ability to survive under different ionic compositions, climatic conditions, and altitudes. In this case study, *Artemia* species are served as ideal model organisms for biodiversity study in inland hypersaline lakes (Camargo et al. 2005; Castro et al. 2006; Hand SC 1982; Maniatsi et al. 2009). In addition, the morphological variations displayed among *Artemia* populations also provide excellent materials for studying adaptive genetic polymorphisms at molecular levels. During the past two decades phylogenetic relationships among *Artemia* species have been established by combined studies based on cross-breeding, morphological differentiation, cytogenetics, nuclear (including allozymes and other nuclear DNA sequences) (Badaracco G 1995; Baxevanis et al. 2006; Sun Y 2000) and mitochondrial (mtDNA) DNA markers (Badaracco G 1995).

Seven sexual species have been described thus far, as well as numerous parthenogenetic populations. Five species are found in Eurasia: *A. salina* (Mediterranean area), *A. urmiana* (Iran), *A. tibetiana* (Tibet), *A. sinica* (van Wely et al.), and *A. spp* (Old World). The New World species are *A. franciscana* and *A. Persimilis*; the former are widely distributed in most part of America, while A. persimilis is restricted to certain locations in Chile and Argentina (Clegg et al. 2009). *A.franciscana, A.tibetiana,* and *A.sinica* are the main *Artemia* species that inhabit in China (Figure. 2). *A.tibetiana* dwells in the Tibetan Plateau, with the altitude of ~ 4,500m above the sea level. Living under the harsh condition of hypoxia, low temperature, high solar radiation, and lack of biological production, it requires a modified and adapted energy metabolism for survival. In 1980s, a large quantity of *A.franciscana* was released in the most part of salt field in the Bohai Bay. As a dominant species, *A.franciscana* replaced the local species, *A.sinica*, rapidly and has become the primary species in the Bohai Bay since. As a result, *A.sinica* is almost disappeared completely in sea shores of Eastern China.

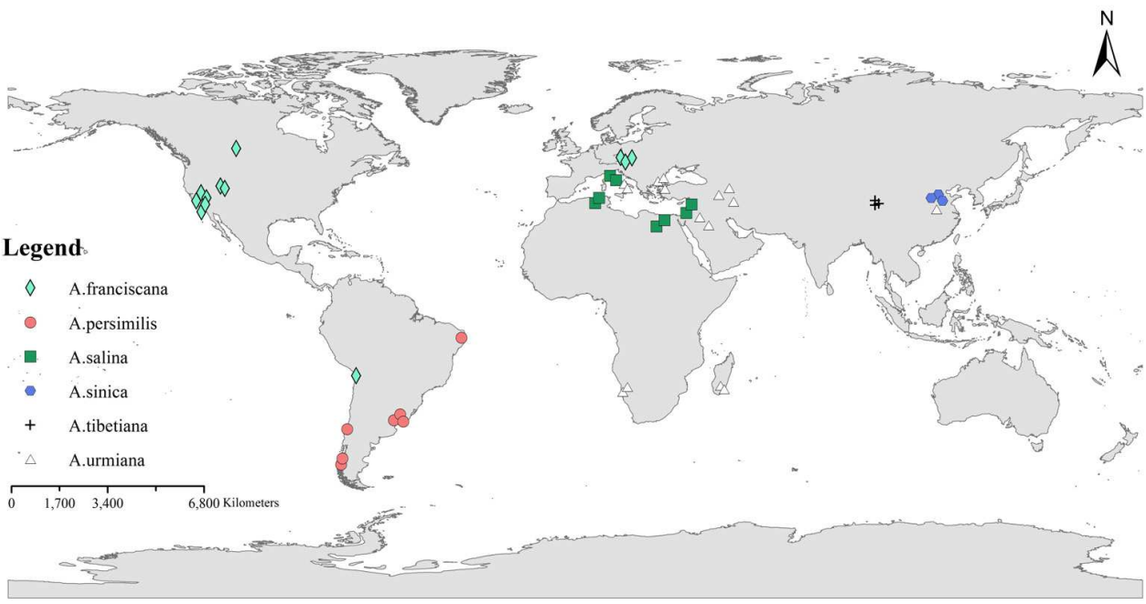


Fig. 2. The distribution of *Artemia* in the world.

2. Biodiversity of *Artemia* populations in China

The phylogeny of various *Artemia* samples from different habitats around the world was reported previously, and our focus now is on the biodiversity of *Artemia* species in China, especially that of the Tibetan Plateau. All strains used in this study are also kept as cysts at the Laboratory of Aquaculture & Artemia Reference Center (ARC) with ARC code numbers (Wang et al. 2008), including six populations represented five salt lakes from Nima (ARC 1609), Yangnapeng Co (ARC 1610), Lagkor Co (ARC 1348), Jingyu lake (ARC 1524), and Co Qen Lakes (ARC 1526 and ARC 1612) of the Tibetan Plateau (Table 3).

ARC #	Location	Year of harvesting
1348	Lagkor Co, Tibet, China	1996
1524	Jingyu Lake, Xinjiang, china	2001
1526	Co Qen,Tibet, China	2001
1612	Co Qen, Tibet, China	2001
1609	Nima, Tibet, China	1999
1610	Yangnapengco, Tibet, China	2002

Table 3. List of Artemia species in China and their locality and ARC codes.

2.1 Phylogenetic analysis of *Artemia* species in China based on *COI* gene barcoding

A 648-bp segment of the mitochondrial *COI* gene was selected as the standard barcode to establish phylogenetic relationships among *Artemia* species from major habitats, including species from the Tibetan Plateau (Figure 3, Wang et al. 2008). We built a phylogenetic tree based on *COI* gene, which separates the populations into five stable clades. Three of them are composed of species from China, and the first clade contains genotypes from populations collected in the Bohai Bay areas of Eastern China and also one sample from Vinh Chau, Vietnam, which shows a high sequence similarity to *A. franciscana*. It is evident for a large-scale invasion of *A. franciscana* in the Bohai Bay (Van Stappen 2007). The second clade is made of *A.tibetiana* genotypes from populations in Tibet and Southwestern China, with high sequence similarity to *A. urmiana*. The third clade belongs to *A. sinica*, which mainly contains populations from Inner Mongolia in the Central North of China. The fourth and fifth clades correspond to *A. persimilis* and *A. Salina,* respectively, and they are not found in this study as major populations in China.

Investigating the amino acid variations, we found two consistent amino acid changes in COI between high and low altitude species we collected in China: 153A/V and 183L/F. These sequence alterations may provide clues for further functional studies such as to determine if the adaptation to high altitude had resulted in the fixation of such mutations. We also used Ka/Ks calculator to estimate Ka/Ks (Zhang et al. 2006) with the aim to reveal sequence signatures of natural selection in *COI* gene. When using *A. franciscana* as a reference, *A. tibetiana* has significantly higher Ka/Ks ratios, which imply relatively stronger selective pressure on this species. Two variations that alter amino-acid sequences between the high and low altitude populations shared by the high altitude group were also detected. The sequence from sample 1612 has the highest Ka/Ks ratio, and its mutation spectra suggests a relatively stronger selection posed on this population and its synonymous mutations provide clues that the population is rapidly diverging, which is most likely due to environmental changes during last three million years rather than genetic drift.

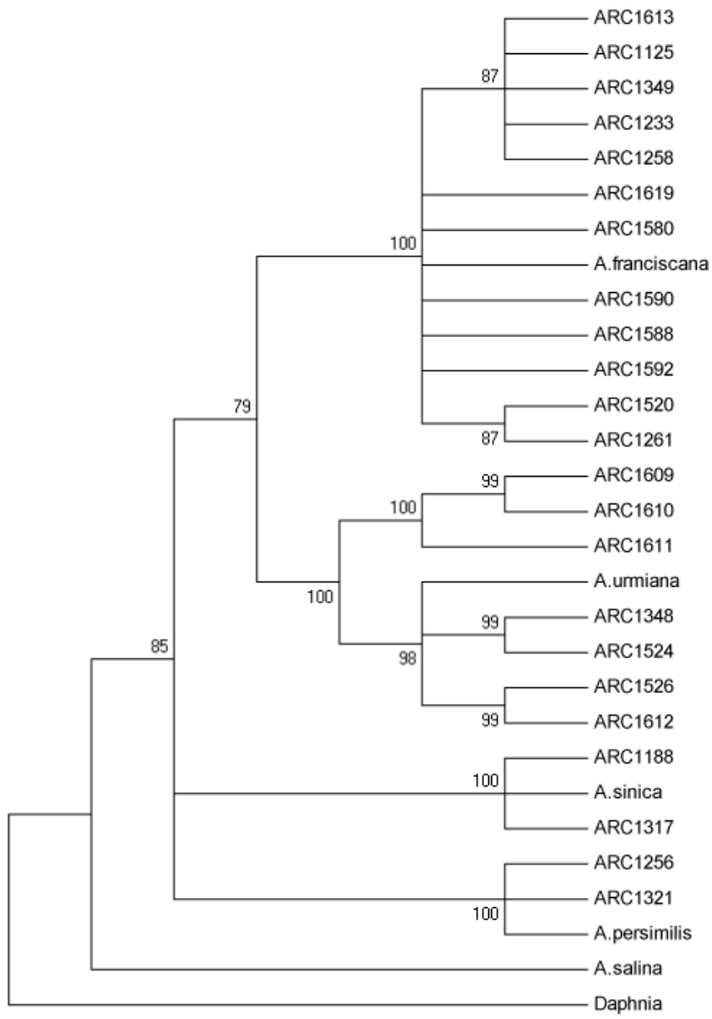


Fig. 3. A phylogenetic tree based on neighbor-joining method. The tree is constructed based on a sequence fragment of *COI* gene (Adapted from Wang et al.).

We further obtained high-quality sequences from individual adults of the six Tibetan populations and calculated the Kimura-2-Parameter distances (Table 4). For phylogentic tree construction, we used the consensus sequences when sequence heterogeneities are encountered among a minor set of samples.

	1348	1524	1526	1609	1610	1612
N	20	18	9	20	20	8
Min	0	0	0.38	0	0	2.55
Max	7.76	1.31	11.07	4.92	9.56	12.01
Mean	2.3	0.51	4.17	2.49	3.42	7.07
S.D.	0.34	0.17	0.52	0.45	0.5	0.79

Table 4. Kimura-2-Parameter distances of samples from Tibet (Adapted from Wang et al 2008).

2.2 Sequencing and comparative analysis of *Artemia* mitochondrial genomes

Based on the obvious divergence of *COI* gene, we speculated that environmental selection may bring more variations to other mitochondrial encoding genes involved in energy metabolism during the long-term selection that may affect structures and activities of the ATPase subunits or even other components of the mitochondrial respiratory chain complexes. Therefore, we decided to take *Artemia* species in Asia as our model and acquired whole mitochondrial genome sequences of *Artemia tibetiana* collected from the Tibetan Plateau and carried out comparative analysis involving other lower altitude *Artemia* species, *A. franciscana, A. urmiana*, and *A.sinica*, and aim to observe specific characteristics of the mt genome sequences of *A. tibetiana*.

We indeed acquired and annotated five mitochondrial genomes, including two ecotypes of *A. tibetiana*, one each from *A. urmiana*, *A.franciscana*, and *A.sinica*. The *A. tibetiana* samples were collected from Nima (ARC 1609) and Yangnapeng Counties (ARC 1610) of the Tibetan Plateau with the altitude higher than 4,000m. *A. urmiana*, which had a very close phylogenetic relationship with *A. tibetiana* based on previous DNA barcoding study, were collected from Urmia Lake of Iran (ARC 1227) at an altitude of 1275m above the sea level. *A.sinica*, another native species in China which is collected from Yimeng of Inner Mongolia (ARC 1188), where it has an altitude of ~1000m above the sea level and a climate of dry, windy, and sandy. We also have one ecotype of *A.franciscana* is collected from Huangnigou, Shangdong in China (ARC 1590). The length variations are mainly found in the non-coding region (known as the D-loop region).All *Artemia* mitochondrial genomes encode 37 genes including 2 rRNAs, 22 tRNAs, and 13 polypeptides that are subunits of the respiratory chain complexes residing on the inner mitochondrial membrane.

Comparative analysis of mitochondrial DNA (mtDNA) of these *Artemia* species shows that the nucleotide variation ratio is higher between *A. tibetiana* and *A. franciscana* and much lower between *A. tibetiana* and *A. urmiana* or *A.sinica*. Among the 13 protein-coding genes, *ND* gene family has more nucleotide variations than other genes. ND6 varies the most both between *A. tibetiana* and *A. franciscana* (T-F) and between *A. tibetiana* and *A. urmiana* (T-U), and the same situation is observed between *A. tibetiana* and *A.sinica* (T-S). When analyzing the amino-acid changes, *ATP8* gene has higher variation rates, second only to the *ND* gene family. In addition, COI is the most conservative protein in amino-acid sequence among the 13 polypeptides. The complexes IV and V contain more variations than other complexes. With Ka/Ks Calculator, ATP8 has a high Ka/Ks ratio, just lower than that of *ND4* when *A. tibetiana* and *A. urmiana* are separated from *A. franciscana*, while *ATP6* possesses higher evolutionary rate between *A. tibetiana* and *A. Urmiana* (data not shown)

3. Conclusion

Consequently, our results on DNA barcoding and comparative analysis reveal the current distribution of *Artemia* species in China and phylogenetic relationship among them, providing insights into the adaptive evolution of DNA sequences of *Artemia*. Based on phylogenetic and divergence analyses of the selected samples from different regions of the world, it is possible that the high altitude group of *Artemia* are descendents of a local ancestral species in the Himalayas which diverged genetically as the Tibetan Plateau arose stepwise over approximately the last three million years (Tapponnier et al. 2001).

The comparative studies among different *Artemia* species reveal complex sequence diversities that are expected to have functional relevance, such as energy metabolism and environmental

adaptation. The highest number of adaptive variations in ATP8 implies that it is under selective pressure during long-term geographical isolation when *A. tibetiana* separated from their common ancestor together with the rise of Himalaya Mountains. It was reported that the *ATP8* gene encodes a core subunit of F0 in ATPase that synthesizes ATP based on a proton-gradient that results from H+ pumping into the intermembrane space (da Fonseca, Johnson et al. 2008). It was also suggested that ATP8 may play regulatory roles in ATP synthesis among different species since it has highly variable sites in the protein-coding sequence (da Fonseca, Johnson et al. 2008). Moreover, the Ka/Ks ratio in ATP6 is also relatively high when we compared the 13 protein-coding mitochondrial genes of *A. tibetiana* to those of *A. urmiana* and *A.sinica*. It is known that ATP6 plays an important role in the assembly of F0 (Hadikusumo, Meltzer et al. 1988) and the highly variable sites are found in the predicted loop regions where the sequences are less selected in terms of its overall function. The high variation rates found among the ATPase subunits imply a strong selective pressure on the *Artemia* energy metabolism system from the high plateau environment.

4. Acknowledgment

The work received financial support by the National Natural Science Foundation of China (30221004) awarded to Weiwei Wang, grant (KSCX2-SW-331) from the Chinese Academy of Sciences awarded to Jun Yu, and the National Basic Research Program (973 Program) from the Ministry of Science and Technology of the People's Republic of China (2011CB944100) awarded to Jun Yu.

5. References

M. Aliabadian, M. Kaboli, et al. (2009). Molecular Identification of Birds: Performance of Distance-Based DNA Barcoding in Three Genes to Delimit Parapatric Species, *PLoS ONE*, Vol. 4, No. issue 1, pp.e4119,

D. E. Arnot, C. Roper, et al. (1993). Digital codes from hypervariable tandemly repeated DNA sequences in the Plasmodium falciparum circumsporozoite gene can genetically barcode isolates, *Molecular and Biochemical Parasitology*, Vol. 61, No. issue 1, pp.15-24, 0166-6851

B. M. Badaracco G, Landsberger N. (1995). Phylogenetic study of bisexual Artemia using random amplified polymorphic DNA, *Journal of molecular evolution*, Vol. 41, No. issue 2, pp.150-154,

S. L. BallK. F. Armstrong. (2008). Rapid, one-step DNA extraction for insect pest identification by using DNA barcodes, *J Econ Entomol*, Vol. 101, No. issue 2, 2008/05/08 pp.523-32, 0022-0493 (Print)

D. Barth, S. Krenek, et al. (2006). Intraspecific genetic variation in Paramecium revealed by mitochondrial cytochrome C oxidase I sequences, *J Eukaryot Microbiol*, Vol. 53, No. issue 1, 2006/01/31 pp.20-5, 1066-5234 (Print)

A. D. Baxevanis, I. Kappas, et al. (2006). Molecular phylogenetics and asexuality in the brine shrimp Artemia, *Molecular Phylogenetics and Evolution*, Vol. 40, No. issue 3, pp.724-738, 1055-7903

J. S. Beckmann. (1999). Disease taxonomy – monogenic muscular dystrophy, *British Medical Bulletin*, Vol. 55, No. issue 2, pp.340-357

J. Brodie, L. Robba, et al. (2006). Assessing the use of the mitochondrial cox1 marker for use in DNA barcoding of red algae (Rhodophyta), *American Journal of Botany*, Vol. 93, No. issue 8, pp.1101-1108, 0002-9122

S. H. M. Butchart, M. Walpole, et al. (2010). Global Biodiversity: Indicators of Recent Declines, *Science*, Vol. 328, No. issue 5982, pp.1164-1168,

W. Camargo, G. Duran, et al. (2005). Determination of biological and physicochemical parameters of Artemia franciscana strains in hypersaline environments for aquaculture in the Colombian Caribbean, *Saline Systems*, Vol. 1, No. issue 1, pp.9, 1746-1448

S. M. Carr, H. D. Marshall, et al. (2008). Phylogeographic genomics of mitochondrial DNA: Highly-resolved patterns of intraspecific evolution and a multi-species, microarray-based DNA sequencing strategy for biodiversity studies, *Comparative Biochemistry and Physiology D-Genomics & Proteomics*, Vol. 3, No. issue 1, pp.1-11, 1744-117X

T. Castro, G. Gajardo, et al. (2006). A biometric and ecologic comparison between Artemia from Mexico and Chile, *Saline Systems*, Vol. 2, No. issue 1, pp.13, 1746-1448

J. S. Clegg, G. Gajardo. (2009). Two highly diverged New World Artemia species, A. franciscana and A. persimilis, from contrasting hypersaline habitats express a conserved stress protein complement, *Comparative Biochemistry and Physiology - Part A: Molecular & Integrative Physiology*, Vol. 153, No. issue 4, pp.451-456, 1095-6433

S. Creer. (2010). Second-generation sequencing derived insights into the temporal biodiversity dynamics of freshwater protists, *Molecular Ecology*, Vol. 19, No. issue 14, pp.2829-2831, 0962-1083

B. Dayrat. (2005). Towards integrative taxonomy, *Biological Journal of the Linnean Society*, Vol. 85, No. issue 3, pp.407-415, 0024-4066

G. S. De HoogJ. S. Zeng. (2008). Exophiala spinifera and its allies: diagnostics from morphology to DNA barcoding, *Medical Mycology*, Vol. 46, No. issue 3, pp.193-208, 1369-3786

B. E. Deagle, A. Chiaradia, et al. (2010). Pyrosequencing faecal DNA to determine diet of little penguins: is what goes in what comes out?, *Conservation Genetics*, Vol. 11, No. issue 5, pp.2039-2048, 1566-0621

K. M. Evans, A. H. Wortley, et al. (2007). An Assessment of Potential Diatom "Barcode" Genes (cox1, rbcL, 18S and ITS rDNA) and their Effectiveness in Determining Relationships in Sellaphora (Bacillariophyta), *Protist*, Vol. 158, No. issue 3, pp.349-364, 1434-4610

J. J. Farmer, 3rd, B. R. Davis, et al. (1985). Biochemical identification of new species and biogroups of Enterobacteriaceae isolated from clinical specimens, *J Clin Microbiol*, Vol. 21, No. issue 1, 1985/01/01 pp.46-76, 0095-1137 (Print)

E. Ferri, M. Barbuto, et al. (2009). Integrated taxonomy: traditional approach and DNA barcoding for the identification of filarioid worms and related parasites (Nematoda), *Frontiers in Zoology*, Vol. 6, No. issue 1, pp.1, 1742-9994

A. Z. Fire, P. Parameswaran, et al. (2007). A pyrosequencing-tailored nucleotide barcode design unveils opportunities for large-scale sample multiplexing, *Nucleic Acids Research*, Vol. 35, No. issue 19, 0305-1048

V. G. Fonseca, G. R. Carvalho, et al. (2010). Second-generation environmental sequencing unmasks marine metazoan biodiversity, *Nature Communications*, Vol. 1, No. issue pp.-, 2041-1723

L. FrézalR. Leblois. (2008). Four years of DNA barcoding: Current advances and prospects, *Infection, Genetics and Evolution*, Vol. 8, No. issue 5, pp.727-736, 1567-1348

M. Göker, G. García-Blázquez, et al. (2009). Molecular Taxonomy of Phytopathogenic Fungi: A Case Study in <italic>Peronospora</italic>, *PLoS ONE*, Vol. 4, No. issue 7, pp.e6319,

B. Ghebremedhin, F. Layer, et al. (2008). Genetic Classification and Distinguishing of Staphylococcus Species Based on Different Partial gap, 16S rRNA, hsp60, rpoB, sodA, and tuf Gene Sequences, *J. Clin. Microbiol.*, Vol. 46, No. issue 3, pp.1019-1025,

J. R. W. Govan, J. E. Hughes, et al. (1996). Burkholderia cepacia: medical, taxonomic and ecological issues, *J Med Microbiol*, Vol. 45, No. issue 6, pp.395-407,

M. H. Greenstone, D. L. Rowley, et al. (2005). Barcoding generalist predators by polymerase chain reaction: carabids and spiders, *Molecular Ecology*, Vol. 14, No. issue 10, pp.3247-3266, 1365-294X

H. Y. Guo, W. W. Wang, et al. (2010). DNA barcoding provides distinction between Radix Astragali and its adulterants, *Science China-Life Sciences*, Vol. 53, No. issue 8, pp.992-999, 1674-7305

M. Hajibabaei, M. A. Smith, et al. (2006). A minimalist barcode can identify a specimen whose DNA is degraded, *Molecular Ecology Notes*, Vol. 6, No. issue 4, pp.959-964, 1471-8286

M. H. Hajibabaei, M., S. Shokralla, et al. (2011). Environmental Barcoding: A Next-Generation Sequencing Approach for Biomonitoring Applications Using River Benthos, *Plos One*, Vol. 6, No. issue 4, 1932-6203

C. F. Hand SC. (1982). Larval brine shrimp malate dehydrogenase: biosynthesis and temporal pattern related to environmental salinity, *The journal of experimental zoology*, Vol. 219, No. issue 1, pp.17-27,

J. He, K.-L. Wong, et al. (2010). Identification of the Medicinal Plants in Aconitum L. by DNA Barcoding Technique, *Planta Med*, Vol. 76, No. issue EFirst, 09.03.2010 pp.1622, 1628, 0032-0943

P. D. N. Hebert, A. Cywinska, et al. (2003). Biological identifications through DNA barcodes, *Proceedings of the Royal Society of London. Series B: Biological Sciences*, Vol. 270, No. issue 1512, pp.313-321,

P. D. N. Hebert, E. H. Penton, et al. (2004). Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator, *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 101, No. issue 41, pp.14812-14817, 0027-8424

P. D. N. Hebert, S. Ratnasingham, et al. (2003). Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species, *Proceedings of the Royal Society of London. Series B: Biological Sciences*, Vol. 270, No. issue Suppl 1, pp.S96-S99,

P. D. N. Hebert, M. Y. Stoeckle, et al. (2004). Identification of Birds through DNA Barcodes, *PLoS Biol*, Vol. 2, No. issue 10, pp.e312,

M. Heethoff, M. Laumann, et al. (2011). Integrative taxonomy: Combining morphological, molecular and chemical data for species delineation in the parthenogenetic Trhypochthonius tectorum complex (Acari, Oribatida, Trhypochthoniidae), *Frontiers in Zoology*, Vol. 8, No. issue pp.-, 1742-9994

M. J. Hickerson, C. P. Meyer, et al. (2006). DNA barcoding will often fail to discover new animal species over broad parameter space, *Systematic Biology*, Vol. 55, No. issue 5, pp.729-739, 1063-5157

I. D. HoggP. D. N. Hebert. (2004). Biological identification of springtails (Hexapoda : Collembola) from the Canadian Arctic, using mitochondrial DNA barcodes, *Canadian Journal of Zoology-Revue Canadienne De Zoologie*, Vol. 82, No. issue 5, pp.749-754, 0008-4301

P. M. Hollingsworth, M. L. Hollingsworth, et al. (2009). Selecting barcoding loci for plants: evaluation of seven candidate loci with species-level sampling in three divergent

groups of land plants, *Molecular Ecology Resources*, Vol. 9, No. issue 2, pp.439-457, 1755-098X

W. H. Holzapfel, P. Haberer, et al. (2001). Taxonomy and important features of probiotic microorganisms in food and nutrition, *The American Journal of Clinical Nutrition*, Vol. 73, No. issue 2, pp.365S-373S,

R. M. Jennings, A. Bucklin, et al. (2010). Barcoding of Arrow Worms (Phylum Chaetognatha) from Three Oceans: Genetic Diversity and Evolution within an Enigmatic Phylum, *PLoS ONE*, Vol. 5, No. issue 4, pp.e9949,

E. B. Kauffman, S. A. Jones, et al. (2003). Virus detection protocols for West Nile virus in vertebrate and mosquito specimens, *Journal of Clinical Microbiology*, Vol. 41, No. issue 8, pp.3661-3667, 0095-1137

P. J. Keeling, G. H. Gile, et al. (2010). DNA Barcoding of Chlorarachniophytes Using Nucleomorph Its Sequences1, *Journal of Phycology*, Vol. 46, No. issue 4, pp.743-750, 0022-3646

L. J. Kelly, P. M. Hollingsworth, et al. (2011). DNA barcoding of lichenized fungi demonstrates high identification success in a floristic context, *New Phytologist*, Vol. No. issue pp.no-no, 1469-8137

W. J. KressD. L. Erickson. (2007). A Two-Locus Global DNA Barcode for Land Plants: The Coding <italic>rbcL</italic> Gene Complements the Non-Coding <italic>trnH-psbA</italic> Spacer Region, *PLoS ONE*, Vol. 2, No. issue 6, pp.e508,

W. J. Kress, K. J. Wurdack, et al. (2005). Use of DNA barcodes to identify flowering plants, *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 102, No. issue 23, pp.8369-8374, 0027-8424

M. KumarP. K. Shukla. (2006). Single-stranded conformation polymorphism of large subunit of ribosomal RNA is best suited to diagnosing fungal infections and differentiating fungi at species level, *Diagn Microbiol Infect Dis*, Vol. 56, No. issue 1, 2006/05/03 pp.45-51, 0732-8893 (Print)

R. Lahaye, M. van der Bank, et al. (2008). DNA barcoding the floras of biodiversity hotspots, *Proceedings of the National Academy of Sciences*, Vol. 105, No. issue 8, pp.2923-2928,

R. Le BerreG. K. Fiasorgbor. (1985). [Importance of the cytotaxonomic identification of species of the Simulium damnosum complex in onchocercosis control], *Parassitologia*, Vol. 27, No. issue 1-2, 1985/01/01 pp.179-85, 0048-2951 (Print)

T. Lefébure, C. J. Douady, et al. (2006). Relationship between morphological taxonomy and molecular divergence within Crustacea: Proposal of a molecular threshold to help species delimitation, *Molecular Phylogenetics and Evolution*, Vol. 40, No. issue 2, pp.435-447, 1055-7903

R. E. Ley, P. J. Turnbaugh, et al. (2006). Microbial ecology: Human gut microbes associated with obesity, *Nature*, Vol. 444, No. issue 7122, pp.1022-1023, 0028-0836

S. Lin, H. Zhang, et al. (2009). High-Level Diversity of Dinoflagellates in the Natural Environment, Revealed by Assessment of Mitochondrial cox1 and cob Genes for Dinoflagellate DNA Barcoding, *Appl. Environ. Microbiol.*, Vol. 75, No. issue 5, pp.1279-1290,

N. Maeda, H. Nishiyori, et al. (2008). Development of a DNA barcode tagging method for monitoring dynamic changes in gene expression by using an ultra high-throughput sequencer, *Biotechniques*, Vol. 45, No. issue 1, pp.95-97, 0736-6205

S. Maniatsi, I. Kappas, et al. (2009). Sharp Phylogeographic Breaks and Patterns of Genealogical Concordance in the Brine Shrimp Artemia franciscana, *International Journal of Molecular Sciences*, Vol. 10, No. issue 12, pp.5455-5470, 1422-0067

T. Matsuki, K. Watanabe, et al. (2003). Genus- and species-specific PCR primers for the detection and identification of bifidobacteria, *Curr Issues Intest Microbiol*, Vol. 4, No. issue 2, 2003/09/25 pp.61-9, 1466-531X (Print)

R. Meier, K. Shiyang, et al. (2006). DNA barcoding and taxonomy in diptera: A tale of high intraspecific variability and low identification success, *Systematic Biology*, Vol. 55, No. issue 5, pp.715-728, 1063-5157

I. Meusnier, G. Singer, et al. (2008). A universal DNA mini-barcode for biodiversity analysis, *BMC Genomics*, Vol. 9, No. issue 1, pp.214, 1471-2164

M. Meyer, U. Stenzel, et al. (2007). Targeted high-throughput sequencing of tagged nucleic acid samples, *Nucleic Acids Research*, Vol. 35, No. issue 15, 0305-1048

J. Mitsui, Y. Fukuda, et al. (2010). Multiplexed resequencing analysis to identify rare variants in pooled DNA with barcode indexing using next-generation sequencer, *Journal of Human Genetics*, Vol. 55, No. issue 7, pp.448-455, 1434-5161

E. Nassonova, A. Smirnov, et al. (2010). Barcoding Amoebae: Comparison of SSU, ITS and COI Genes as Tools for Molecular Identification of Naked Lobose Amoebae, *Protist*, Vol. 161, No. issue 1, pp.102-115, 1434-4610

V. NijmanM. Aliabadian. (2010). Performance of distance-based DNA barcoding in the molecular identification of Primates, *Comptes Rendus Biologies*, Vol. 333, No. issue 1, pp.11-16, 1631-0691

R. J. Owen. (1989). Chromosomal DNA fingerprinting--a new method of species and strain identification applicable to microbial pathogens, *J Med Microbiol*, Vol. 30, No. issue 2, 1989/10/01 pp.89-99, 0022-2615 (Print)

J. M. Padial, A. Miralles, et al. (2010). The integrative future of taxonomy, *Frontiers in Zoology*, Vol. 7, No. issue pp.-, 1742-9994

J. PawlowskiB. Lecroq. (2010). Short rDNA Barcodes for Species Identification in Foraminifera, *Journal of Eukaryotic Microbiology*, Vol. 57, No. issue 2, pp.197-205, 1066-5234

G. G. Pegg, B. Sinclair, et al. (2006). MtDNA barcode identification of fish larvae in the southern Great Barrier Reef, Australia, *Scientia Marina*, Vol. 70, No. issue pp.7-12, 0214-8358

T. J. Pereira, G. Fonseca, et al. (2010). Diversity of free-living marine nematodes (Enoplida) from Baja California assessed by integrative taxonomy, *Marine Biology*, Vol. 157, No. issue 8, pp.1665-1678, 0025-3162

C. Perrings. (1996). Economics, ecology and the global biodiversity assessment, *Trends in Ecology & Evolution*, Vol. 11, No. issue 6, pp.270-270, 0169-5347

D. L. Porazinska, R. M. Giblin-Davis, et al. (2009). Evaluating high-throughput sequencing as a method for metagenomic analysis of nematode diversity, *Molecular Ecology Resources*, Vol. 9, No. issue 6, pp.1439-1450, 1755-098X

T. M. Porter, J. E. Skillman, et al. (2008). Fruiting body and soil rDNA sampling detects complementary assemblage of Agaricomycotina (Basidiomycota, Fungi) in a hemlock-dominated forest plot in southern Ontario, *Molecular Ecology*, Vol. 17, No. issue 13, pp.3037-3050, 0962-1083

R. D. Randrianiaina, F. Glaw, et al. (2007). Descriptions of the tadpoles of two species of Gephyromantis, with a discussion of the phylogenetic origin of direct development in mantellid frogs, *Zootaxa*, Vol. No. issue 1401, pp.53-61, 1175-5326

S. RatnasinghamP. D. N. Hebert. (2007). bold: The Barcode of Life Data System (http://www.barcodinglife.org), *Molecular Ecology Notes*, Vol. 7, No. issue 3, pp.355-364, 1471-8286

E. A. RemigioP. D. N. Hebert. (2003). Testing the utility of partial COI sequences for phylogenetic estimates of gastropod relationships, *Molecular Phylogenetics and Evolution*, Vol. 29, No. issue 3, pp.641-647, 1055-7903

D. Rubinoff, S. Cameron, et al. (2006). A genomic perspective on the shortcomings of mitochondrial DNA for "barcoding" identification, *J Hered*, Vol. 97, No. issue 6, 2006/12/01 pp.581-94, 0022-1503 (Print)

D. Rubinoff, S. Cameron, et al. (2006). A genomic perspective on the shortcomings of mitochondrial DNA for "barcoding" identification, *Journal of Heredity*, Vol. 97, No. issue 6, pp.581-594, 0022-1503

D. B. Rusch, A. L. Halpern, et al. (2007). The Sorcerer II Global Ocean Sampling expedition: northwest Atlantic through eastern tropical Pacific, *PLoS Biol*, Vol. 5, No. issue 3, 2007/03/16 pp.e77, 1545-7885 (Electronic)

H. SakaokaY. Eizuru. (1992). [Identification of species and discrimination among their individuals by restriction enzyme cleavage of the genomes], *Nippon Rinsho*, Vol. 50 Suppl, No. issue 1992/07/01 pp.125-35, 0047-1852 (Print)

G. W. Saunders. (2005). Applying DNA barcoding to red macroalgae: a preliminary appraisal holds promise for future applications, *Philosophical Transactions of the Royal Society B-Biological Sciences*, Vol. 360, No. issue 1462, pp.1879-1888, 0962-8436

V. Savolainen, R. Lahaye, et al. (2008). DNA barcoding the floras of biodiversity hotspots, *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 105, No. issue 8, pp.2923-2928, 0027-8424

D. E. SchindelS. E. Miller. (2005). DNA barcoding a useful tool for taxonomists, *Nature*, Vol. 435, No. issue 7038, pp.17-17, 0028-0836

A. Schussler, H. Stockinger, et al. (2010). DNA barcoding of arbuscular mycorrhizal fungi, *New Phytologist*, Vol. 187, No. issue 2, pp.461-474, 0028-646X

K. A. Seifert, R. A. Samson, et al. (2007). Prospects for fungus identification using C01 DNA barcodes, with Penicillium as a test case, *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 104, No. issue 10, pp.3901-3906, 0027-8424

R. G. Shatters, C. A. Powell, et al. (2009). Improved DNA Barcoding Method for Bemisia tabaci and Related Aleyrodidae: Development of Universal and Bemisia tabaci Biotype-Specific Mitochondrial Cytochrome c Oxidase I Polymerase Chain Reaction Primers, *Journal of Economic Entomology*, Vol. 102, No. issue 2, pp.750-758, 0022-0493

P. C. Shaw, M. Li, et al. (2011). Identification of herbal medicinal materials using DNA barcodes, *Journal of Systematics and Evolution*, Vol. 49, No. issue 3, pp.271-283, 1674-4918

M. E. Siddall, F. M. Fontanella, et al. (2009). Barcoding Bamboozled by Bacteria: Convergence to Metazoan Mitochondrial Primer Targets by Marine Microbes, *Systematic Biology*, Vol. 58, No. issue 4, pp.445-451,

M. A. Smith, B. L. Fisher, et al. (2005). DNA barcoding for effective biodiversity assessment of a hyperdiverse arthropod group: the ants of Madagascar, *Philosophical Transactions of the Royal Society B: Biological Sciences*, Vol. 360, No. issue 1462, pp.1825-1834,

M. L. Sogin, H. G. Morrison, et al. (2006). Microbial diversity in the deep sea and the underexplored “rare biosphere”, *Proceedings of the National Academy of Sciences*, Vol. 103, No. issue 32, pp.12115-12120,

H. Song, J. E. Buhay, et al. (2008). Many species in one: DNA barcoding overestimates the number of species when nuclear mitochondrial pseudogenes are coamplified, *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 105, No. issue 36, pp.13486-13491, 0027-8424

C. D. Specht, C. Sass, et al. (2007). DNA Barcoding in the Cycadales: Testing the Potential of Proposed Barcoding Markers for Species Identification of Cycads, *Plos One*, Vol. 2, No. issue 11, 1932-6203

R. F. Stern, A. Horak, et al. (2010). Environmental Barcoding Reveals Massive Dinoflagellate Diversity in Marine Environments, *Plos One*, Vol. 5, No. issue 11, 1932-6203

H. Stockinger, M. Krüger, et al. (2010). DNA barcoding of arbuscular mycorrhizal fungi, *New Phytologist*, Vol. 187, No. issue 2, pp.461-474, 1469-8137

D. J. Sullivan, M. C. Henman, et al. (1996). Molecular genetic approaches to identification, epidemiology and taxonomy of non-albicans Candida species, *J Med Microbiol*, Vol. 44, No. issue 6, pp.399-408,

R. C. Summerbell, M. K. Moore, et al. (2007). ITS barcodes for Trichophyton tonsurans and T-equinum, *Medical Mycology*, Vol. 45, No. issue 3, pp.193-200, 1369-3786

S. W. Sun Y, Zhong YC, Zhang RS, Chen RY. (2000). Phylogenetic study of Artemia from China using RAPD and AFLP markers, *Yi Chuan Xue Bao*, Vol. 27, No. issue 3, pp.210-218,

P. Tapponnier, X. Zhiqin, et al. (2001). Oblique stepwise rise and growth of the Tibet plateau, *Science*, Vol. 294, No. issue 5547, pp.1671-7, 0036-8075 (Print)

L. Tedersoo, T. Jairus, et al. (2008). Strong host preference of ectomycorrhizal fungi in a Tasmanian wet sclerophyll forest as revealed by DNA barcoding and taxon-specific primers, *New Phytologist*, Vol. 180, No. issue 2, pp.479-490, 0028-646X

A. Valentini, F. Pompanon, et al. (2009). DNA barcoding for ecologists, *Trends in Ecology & Evolution*, Vol. 24, No. issue 2, pp.110-117, 0169-5347

B. J. van Wely, S. Teerenstra, et al. (2011). Systematic review of the effect of external beam radiation therapy to the breast on axillary recurrence after negative sentinel lymph node biopsy, *Br J Surg*, Vol. 98, No. issue 3, 2011/01/22 pp.326-33, 1365-2168 (Electronic)

W. Wang, Q. Luo, et al. (2008). Phylogenetic analysis of brine shrimp (Artemia) in China using DNA barcoding, *Genomics Proteomics Bioinformatics*, Vol. 6, No. issue 3-4, 2009/03/31 pp.155-62, 1672-0229 (Print)

R. D. Ward, T. S. Zemlak, et al. (2005). DNA barcoding Australia's fish species, *Philosophical Transactions of the Royal Society B: Biological Sciences*, Vol. 360, No. issue 1462, pp.1847-1857,

K. W. Will, B. D. Mishler, et al. (2005). The perils of DNA barcoding and the need for integrative taxonomy, *Systematic Biology*, Vol. 54, No. issue 5, pp.844-851, 1063-5157

B. Wood, B. G. Richmond. (2000). Human evolution: taxonomy and paleobiology, *Journal of Anatomy*, Vol. 197, No. issue 1, pp.19-60, 1469-7580

H. Yamamoto. (1992). [Detection and identification of Legionella species by PCR], *Nippon Rinsho*, Vol. 50 Suppl, No. issue 1992/07/01 pp.394-9, 0047-1852 (Print)

Y. Z. Zhang, S. L. Si, et al. (2011). DNA barcoding of endoparasitoid wasps in the genus Anicetus reveals high levels of host specificity (Hymenoptera: Encyrtidae), *Biological Control*, Vol. 58, No. issue 3, pp.182-191, 1049-9644

Z. Zhang, J. Li, et al. (2006). Computing Ka and Ks with a consideration of unequal transitional substitutions, *BMC Evol Biol*, Vol. 6, No. issue pp.44, 1471-2148 (Electronic)

B. Zhou, L. Zhang, et al. (2003). [DNA-based technique and species identification], *Fa Yi Xue Za Zhi*, Vol. 19, No. issue 4, 2003/12/16 pp.245-8, 252, 1004-5619 (Print)



Changing Diversity in Changing Environment

Edited by PhD. Oscar Grillo

ISBN 978-953-307-796-3
Hard cover, 392 pages
Publisher InTech
Published online 14, November, 2011
Published in print edition November, 2011

As everybody knows, the dynamic interactions between biotic and abiotic factors, as well as the anthropic ones, considerably affect global climate changes and consequently biology, ecology and distribution of life forms of our planet. These important natural events affect all ecosystems, causing important changes on biodiversity. Systematic and phylogenetic studies, biogeographic distribution analysis and evaluations of diversity richness are focal topics of this book written by international experts, some even considering economical effects and future perspectives on the managing and conservation plans.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Hangxiao Zhang, Jing Sun, Weiwei Wang and Jun Yu (2011). Brine Shrimp Diversity in China Based on DNA Barcoding, Changing Diversity in Changing Environment, PhD. Oscar Grillo (Ed.), ISBN: 978-953-307-796-3, InTech, Available from: http://www.intechopen.com/books/changing-diversity-in-changing-environment/brine-shrimp-diversity-in-china-based-on-dna-barcoding

INTECH
open science | open minds

InTech Europe
University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China
Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the Creative Commons Attribution 3.0 License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen