

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Role of Cytokines and Chemokines in HIV Infection

Vishwanath Venketaraman et al.*

College of Osteopathic Medicine of the Pacific, Western University of Health Sciences,
USA

1. Introduction

Human immunodeficiency virus (HIV) is the cause of acquired immunodeficiency syndrome (AIDS). Blood monocytes and resident macrophages are important *in vivo* cell targets for HIV infection and their role in AIDS pathogenesis are well documented. These cells of innate immune defenses usually survive HIV infection, serve as a major virus reservoir, and function as immunoregulatory cells through secretion of several pro-inflammatory cytokines and chemokines in response to HIV infection, thereby recruiting and activating new target cells for the virus, including CD4+ T cells. This review describes the alterations in the synthesis of host cytokines and chemokines following HIV infection thereby favoring successful survival of the virus inside the host and enhancing the susceptibility of the host to opportunistic infections.

2. HIV and chemokine receptors

HIV infects immune cells of the macrophage and T-cell lineage. Entry into these cells requires CD4 as a receptor in addition to a co-receptor which most frequently is either chemokine receptor CCR5 or CXCR4 (Gorry & Ancuta, 2011). Binding and entry into human cells requires the two HIV envelope glycoproteins gp120 and gp41. Gp41 possesses a transmembrane domain and is associated with the viral envelope while Gp120 is present in association with Gp41 but does not insert into or contact the viral membrane (Tagliamonte *et al*; 2010). These two viral glycoproteins are present in HIV as tetramers. Therefore, three Gp41 molecules associate within the viral membrane, while three molecules of Gp120 associate with Gp41 (Tagliamonte *et al*; 2010). To facilitate HIV-1 entry into human cells, Gp120 binds to human cellular CD4 with high affinity. Binding causes a conformational change in Gp120 that reveals a co-receptor binding site. Binding to one of the chemokine

*Devin Morris², Clare Donohou⁵, Andrea Sipin⁴, Steven Kung⁴, Hyoung Oh², Mesharee Franklin², John P. Murad³, Fadi T. Khasawneh³, Beatrice Saviola¹, Timothy Guilford⁶ and Clare Donahue⁶

¹College of Osteopathic Medicine of the Pacific, Western University of Health Sciences, USA

²Graduate College of Biomedical Sciences, USA

³College of Pharmacy, Western University of Health Sciences, USA

⁴California State Polytechnic University, USA

⁵Pitzer College, USA

⁶Your Energy Systems, USA

receptors is then facilitated which in turn induces a conformational change in the glycoprotein gp41 N-terminus (Tagliamonte *et al*; 2010). A fusion peptide portion of gp41 inserts into the host cell membrane and lowers energy that is required for fusion of the host and viral membranes (Tagliamonte *et al*; 2010). The viral core is then translocated into the cytoplasm of the host cell.

HIV-1 viral variants can in general use either the CCR5 or CXCR4 co-receptor for entry into human cells (Gorry & Ancuta, 2011). They may also at times use a variety of other chemokine receptors for entry (Gorry *et al*; 2007). The normal function of chemokine receptors is to bind chemokines that target immune cells to areas of inflammation within the human body. Certain HIV-1 viruses may have an increased ability to either bind the CCR5 receptor and are known as R5 viruses, bind to CXCR4 and are known as X4 viruses, or bind with mixed affinity to either receptor. This differential affinity lies within the specific alterations in amino acid sequence of the gp120 glycoprotein (Gorry & Ancuta, 2011). Although not correlating completely, macrophage tropic HIV-1 viruses generally are R5 and T-cell tropic viruses are X4 viruses (Gorry & Ancuta, 2011). Early during infection R5 viruses predominate, and it appears that there is some mechanism which selects these viruses during the transmission process (Grivel *et al*; 2010). For example, an HIV-1 naive individual may be exposed to both R5 and X4 virus particles from an infected individual, but only become infected with the R5 viral particles. There may be multiple factors which affect this process, including co-receptor availability and pH at the sites of infection. Acidic pH may act to disrupt the cationic charge present in gp120 proteins which bind to CXCR4 preferentially (Kwong *et al*; 2010, Edo-Matas *et al*; 2010). R5 viruses are also prominent during chronic infection. X4 viruses or R5X4 viruses which have mixed affinity can arise later during infection and often their presence precedes disease progression and immune cell depletion (Mariani, 2010).

Deletion of the CCR5 receptor can in many cases abrogate infection with HIV-1 completely. It has previously been identified that individuals homozygous for a 32 base pair deletion within the CCR5 gene resulting in a nonfunctional CCR5 molecule are resistant to infection with the HIV-1 virus, though there have been some instances where homozygous CCR5 Δ 32 individuals were infected with X4 HIV-1 (Samson, 1996). Additionally, people who carry one allele of CCR5 Δ 32 have a slower progression of the disease. This knowledge has led to the development of treatments for HIV-1 infection. Transplantation of stem cells from individuals homozygous for CCR5 Δ 32 into CCR5 HIV-1 positive individuals resulted in clearing of the virus from the infected patients (Hutter, 2009). Monoclonal antibodies against CCR5 to inhibit binding of HIV-1 to this co-receptor are a potential therapeutic to prevent viral entry and replication (Tenorio, 2011, Suleiman, 2010). In addition there are plans to use an individual's native stem cells as a target to disrupt the CCR5 receptor gene which can then be transplanted back into the HIV infected patient to effect elimination of the HIV-1 virus from the body (Cannon and June, 2011). Pitfalls of these therapies include the problem that the CCR5 chemokine receptor has a native function within the body, and that disrupting this receptor may cause unforeseen deficits in the immune system. In fact, lack of the CCR5 receptor gene has been associated with increased risk of severe infection with other viruses such as the West Nile Virus, and certain flaviviruses (Lin *et al*; 2008, Kindberg *et al*; 2008). Notwithstanding the previously mentioned caveat, interference with the CCR5 receptor may indeed be a promising target to treat those infected with HIV-1 as well as prevent infection for those exposed to the virus via sexual activity, needle sharing, or accidental hospital transmission.

3. Chemokine ligand-2 (CCL2)

CCL2 or monocyte chemotactic protein-1 (MCP-1), of the C-C chemokine family, is a cytokine with the ability to influence both innate and adaptive immune responses (Daly *et al*; 2003). This chemokine is produced by a variety of different cell types including endothelial cells, fibroblasts, epithelial cells, smooth muscle cells, mesangial cells, astrocytic cells, and microglial cells. However, despite the wide range of cell types that have the ability to manufacture CCL2, the majority of CCL2 is produced by macrophages and monocytes (Deshmane *et al*; 2009).

Although technically a chemokine, CCL2 is often classified as an inflammatory cytokine due to its ability to attract various leukocytes (monocytes, memory T cells, basophils, natural killer (NK) cells etc.) to sites of trauma, bacterial and mycobacterial infection, toxin exposure, and ischemia (Daly *et al*; 2003, Deshmane *et al*; 2009, Mahad *et al*; 2003,, Charo *et al*; 2006). Besides attracting various leukocytes, CCL2 also specifically regulates the infiltration of monocytes, memory T lymphocytes, and NK cells (Deshmane *et al*; 2009). In addition, CCL2 has been found to have a profound effect on the differentiation of naïve helper T cells (Daly *et al*; 2003). Interestingly, studies have found that CCL2 expression tends to lead to the development of a Th2 immune response. Taking this tendency into account, it seems likely that CCL2 concentrations in HIV patients, which Weiss *et al*. (Weiss *et al*; 1997) found were correlated with viral load, can be linked to the Th1 to Th2 cytokine response switch often observed in HIV-1-infected patients (Deshmane *et al*; 2009).

CCL2 has also been found to play other roles in HIV pathogenesis. Eugenin *et al*. (Eugenin *et al*; 2006) noted that CCL2 in the central nervous system (CNS) attracts HIV-infected leukocytes into the brain thereby increasing the rate of HIV-1-infected cell-dispersal and causing the eventual impairment of the blood-brain-barrier. In fact, multiple studies indicate that CCL2 is largely responsible for the development of HIV encephalitis (HIVE), HIV-1-associated dementia (HAD), and NeuroAIDS (Deshmane *et al*; 2009, Eugenin *et al*; 2006).

4. HIV and the Th1 to Th2 Cytokine shift

Under normal conditions, the immune system utilizes a Th1 subset response to viral infections. Activated antigen presenting cells (APC) secrete interleukin-12 (IL-12) which causes Th cell differentiation into the Th1 subset of cells (Clerci *et al*; 1993). These Th1 cells then secrete a characteristic Th1 profile of cytokines consisting of interleukin-2 (IL-2), interferon-gamma (IFN- γ), and tumor necrosis factor-beta (TNF- β). IL-2 induces proliferation of naïve Th cells (T₀), amplifying the Th response. IFN- γ induces further IL-12 production in activated APCs, amplifying the Th1 response, and suppressing any Th2 response. IFN- γ also plays an important role in the activation of cytotoxic T_C cells which destroy virally infected cells.

In individuals infected with HIV, the normal Th1 response to viral infection is shifted to a Th2 response (Klein *et al*; 1997, Osakwe *et al*; 2010). Measurement of the serum cytokine levels of HIV infected patients has revealed an increase in Th2 cytokines as well as a decrease in Th1 cytokines (Klein *et al*; 1997, Osakwe *et al*; 2010). Assays have shown elevated serum IL-4 levels in HIV seropositive individuals (Clerci *et al*; 1993). IL-4 in the presence of proliferating T₀ cells leads to their differentiation into the Th2 subset. Th2 cells promote B-cell proliferation, class switching, and eosinophil activation (Clerci *et al*; 1993). This Th2 response is not appropriate for control of intracellular pathogens such as HIV, and so allows it to persist and spread in CD4⁺ T-cells.

5. TNF- α and HIV infection

It has also been shown that HIV infection induces increased production of TNF- α by macrophages. TNF- α stimulates the production of free radicals. Moreover, enhanced levels of free radicals are likely to increase TNF- α in various cells. TNF- α consists of 233 amino acids and is expressed on all somatic cells, particularly on the cell membrane where it becomes hydrolyzed to its soluble form. TNF- α is considered as one of the most highly studied pro-inflammatory cytokines because it plays a critical role in the origin and progression of diseases such as HIV-1 (Bahia and Silakari, 2010). The immuno-regulatory response of the host influences the pathogenesis of HIV-1 infection, triggering monocytes, macrophages, and natural killer cells to produce TNF- α (Alfano and Poli, 2005). As a result, there is a positive correlation between HIV-1 viremia and TNF- α levels in serum of HIV-1 infected patients. This relationship suggests that reducing TNF- α levels may also reduce occurrence of HIV-1 viremia. In excess, TNF- α may cause severe inflammatory damage and toxicity, making control of its production and secretion highly important. Regulating its release serves as a potential means of therapy for HIV-1 and other diseases. TNF- α can also induce other pro-inflammatory cytokines such as IL-6 and IL-8, which aid in the upregulation of viral replication (Fernandez-Ortega et al; 2004). Studies have also shown the ability of TNF- α to stimulate production of anti-inflammatory cytokine IL-10, preventing further inflammation by causing TNF- α inhibition (Leghmari et al; 2008). TNF- α is secreted during the early phase of acute inflammatory diseases. Its pathogenic role in HIV-1 infection involves activation of nuclear factor κ B (NF- κ B), stimulating apoptosis of T lymphocytes. Tissue and plasma samples of hosts express high levels of TNF- α , contributing to fever, anorexia, and other symptoms of HIV/AIDS. TNF- α must be targeted at an appropriate time during production to prevent progression to the chronic stage. Local effect of the cytokine may be beneficial to the host, so monitoring its development is critical. Highly active antiretroviral therapy helps to reduce mortality rates, and development of potent antiretroviral drugs blocking HIV transcription continues to be successful. However, drug resistance and toxicity remains a challenge in this field of medicine (Fernandez-Ortega et al; 2004).

6. Interleukin 1 (IL-1) and HIV infection

HIV infection and its viral proteins can disturb the production of cytokines and disrupt their usual interactions resulting in disruption of the normal immune function. IL-1 and TNF- α are produced by activation of mononuclear phagocytes as well as microglia in the brain in response to normal immune stimuli such as immune complexes, lipopolysaccharides and phorbol esters (Burchett et al; 1998). It has been reported that IL-1 and TNF- α will be produced by either the binding of gp120 to the CD4 molecules on mononuclear phagocytes or infection with HIV (Merrill et al; 1989, Cheung et al; 2008).

IL-1 is the first discovered and most studied member of the cytokine family (Fantuzzi, 2003). IL-1 is a pro-inflammatory cytokine that plays a fundamental role in host defense by inducing acute and chronic inflammation through activation of the innate and acquired immune systems (Nambu and Nakae, 2010). IL-1 has been described as the prototypic pro-inflammatory cytokine as it was originally described as the first “endogenous” pyrogen due to its fever-inducing properties in both rabbits and humans (Dinarello, 1999). However, in spite of much research in the area of fever induction, the role of IL-1 in this area is still

undefined (Fantuzzi, 2003). IL-1 consists of two distinct ligands (IL-1 α and IL-1 β) with two indistinguishable biological activities that signal through the IL-1 receptor (IL-1R1) (Bujak and Frangogiannis, 2009). Both IL-1 α and IL-1 β can also bind the IL-1 receptor accessory protein (IL-1RAcP). Once bound to the receptor, the complex transduces a signal that initiates a wide variety of inflammatory genes by activating the NF- κ B system. The NF- κ B transcribed genes can produce a variety of inflammatory products including chemokines, pro-inflammatory cytokines, such as TNF- α , IL-6 or IL-8 (Nambu and Nakae, 2010), adhesion molecules, such as vascular cell adhesion molecule-1 (VCAM-1) (Marui et al; 1993), colony-stimulating factors, and mesenchymal growth factor genes (Bujak and Frangogiannis, 2009). In addition, expression of inducible nitric oxide synthase, type 2 cyclooxygenase (COX)-2, and type 2 phospholipase A₂ is exquisitely sensitive to IL-1 (Bujak and Frangogiannis, 2009). IL-1 has also been associated with augmentation of the mast cell activation and Th2 cytokine secretion, suggesting involvement of IL-1 in allergic diseases such as allergic asthma (Dinarello, 1999).

Knockouts of IL-1 have been used to study acute and chronic neurodegenerative conditions, in which a role for IL-1 has been well established (Fantuzzi, 2003). For example, in rodent studies the presence of IL-1 after occlusion of the middle cerebral artery will increase the ischemic damage area. It has been shown that caspase-1 cleaves the inactive pre-form to the active mature form of IL-1 β , which contributes to the damage from ischemia (Bujak and Frangogiannis, 2009). In resting cells, procaspase-1 is bound to an inhibitory molecule that prevents its activation. After damage to cells, conversion of procaspase-1 to caspase-1 is triggered by a molecular complex termed the "IL-1 β inflammasome" (Martinon, 2002).

Macrophages and dendritic cells produce IL-1, IL-12 and other cytokines that permit CD4 cells to reach the level of maturation needed to produce IL-2, which is needed for self-replication of the CD4 cells and for the growth and function of CD8 cells (Levy, 2007). Thus IL-1 plays a role in maintaining normal immune function.

Elevation of IL-1 and TNF- α has been demonstrated in the serum of some patients with HIV-1 (Lepe-Zuniga et al; 1987). High levels of IL-1 (Lepe-Zuniga et al; 1987, Weiss et al; 1989, Molina et al; 1989, Roux-Lombard et al; 1989, Emilie et al; 1990) and TNF- α (Roux-Lombard et al; 1989) are produced in the supernatant of cultured peripheral blood monocyte early in the onset of HIV disease. The levels of TNF- α and IL-1 in the serum were positively correlated in symptomatic versus asymptomatic individuals (Lepe-Zuniga et al; 1987). HIV virus is present in mononuclear phagocytes and in the blood and brain of AIDS patients. Production of IL-1 and TNF- α from mononuclear phagocytes after stimulation with HIV-1 may contribute to some of the symptoms of AIDS such as fever, cachexia and aseptic meningitis (Merrill et al; 1989).

Chronic infection and viral latency are typical of HIV-1 infection. Stimulation with IL-1 β as well as TNF- α can stimulate viral replication in chronically infected cells (Devadas et al; 2004). Monocytes are also major reservoirs for HIV-1 in infected tissue and vectors for virus transmission to target cells, as well as sources of potent cytokines that can affect cell function and virus replication (Devadas et al; 2004). It is thought that stimulation of viral replication in chronically infected cells is due to activation of NF- κ B. In addition to IL-1 and TNF- α , contact with macrophages as well as a number of stressors can stimulate NF- κ B, including phorbol esters, radical oxygen intermediates, and UV irradiation (Devadas et al; 2004).

Clinical manifestations of AIDS include both immunologic and neurologic disorders. In the brain it has been shown that IL-1 induces activation and proliferation of astrocytes, while TNF- α contributes to necrosis of cerebral blood vessels and possibly to demyelination

(Merrill et al; 1989). A feedback process has been described in which HIV-1 and TNF- α can each induce expression of the other. It has been proposed that IL-1 will participate in this feedback loop by inducing TNF- α or by direct T-cell activation, which is needed for HIV-1 replication (Merrill et al; 1989).

IL-1 has been implicated in the pathogenesis of HIV associated dementia (HAD) (Kaul et al; 2001). Both IL-1 β and TNF- α are highly expressed in the central nervous system (CNS) of individuals with HAD, correlate with neuronal injury and are implicated in the pathogenesis of HAD (Epstein and Gendelman, 1993,, Brabers and Nottet, 2006). HIV-1, recombinant gp120, and viral transactivator Tat can activate astrocytes to secrete pro-inflammatory cytokines TNF- α , IL-6, and IL-1 β (Corasaniti et al; 2001), which may contribute to the inflammatory environment in the brain (Herbein and Varin, 2010). Microglia and macrophages in the brain can release IL-1 β after stimulation with HIV-1 envelope protein gp120 (Merrill et al; 1992, Wahl et al; 1989), which is elevated in brain during HIV (Tyor et al; 1992) and has been shown to be elevated in the cerebral spinal fluid during HIV infection (Gallo et al; 1989).

About 25% of subjects with HIV will develop dementia, particularly HIV encephalitis (HIVE), which can occur in spite of the use of HAART (Levy, 2007). Macrophage inflammatory products including IL-1 β have been demonstrated in HIV related encephalitis in mouse and human brain tissue (Persidsky et al; 1997). It has been suggested that the release of neurotoxins, including L-cysteine, from macrophages in the brain is mediated by IL-1 following stimulation of the macrophages by the HIV membrane protein gp120.

L-Cysteine can be released from human monocyte derived macrophages stimulated by either gp120 (Lipton, 1998), or by IL-1 (Yeh et al; 2000). It has been suggested that cytokines including IL-1 may mediate the neurotoxic actions of gp120 (Yeh et al; 2000). Cysteine can act as an endogenous neurotoxin (Olney et al; 1990), which under both physiologic and pathophysiologic conditions stimulates N-methyl-D-aspartate subtype of glutamate receptor (NMDARs) and leads to neuronal apoptosis (Yeh et al; 2000). Thus, immune activation of macrophages in the brain without direct HIV infection may lead to neural damage (Yeh et al; 2000).

Autopsy evaluation of brain tissue from HIVE cases shows increased IL-1 β in the frontal white matter of all 11 of the brains evaluated (67). Additionally, IL-1 β , but not TNF- α expression was detected in HIVp24-positive cells in the HIVE patients, which indicates that IL-1 β is induced by HIV-1 infection. The authors concluded that a macrophage/microglia lineage is the main cell type to release cytokines in HIVE, and IL-1 β expression by HIV-1-infected cells may be one of the important factors for induction of HIVE (67).

7. Interleukin-6 (IL-6)

The family of IL-6-type cytokines comprises IL-6, IL-11, LIF (leukaemia inhibitory factor), OSM (oncostatin M), CNTF (ciliary neurotrophic factor), CT-1 (cardiotrophin-1) and CLC (cardiotrophin-like cytokine) (Heinrich et al; 2003). IL-6 is a pleiotropic cytokine that is commonly produced at local tissue sites and released into circulation in almost all situations of homeostatic perturbation typically including endotoxemia, endotoxic lung, trauma, and acute infections. In addition to its critical participation in the generation of immunity against chronic intracellular infections, circulating IL-6, together with other alarm cytokines TNF- α and IL-1, is known to be required for the induction of acute phase reactions composed of fever, corticosterone release, and hepatic production of acute phase proteins many of which

are protease inhibitors (Xing et al; 1998). They activate target genes involved in differentiation, survival, apoptosis and proliferation. The members of this cytokine family have pro- as well as anti-inflammatory properties and are major players in haematopoiesis, as well as in acute-phase and immune responses of the organism. IL-6-type cytokines bind to plasma membrane receptor complexes containing the common signal transducing receptor chain gp 130 (glycoprotein 130). Signal transduction involves the activation of JAK (Janus kinase) tyrosine kinase family members, leading to the activation of transcription factors of the STAT (signal transducers and activators of transcription) family. Another major signaling pathway for IL-6-type cytokines is the MAPK (mitogen-activated protein kinase) cascade (Heinrich et al; 2003). IL-6 was originally identified as $\beta 2$ (IFN- $\beta 2$), IL-1-inducible 26kD protein and as a factor that induces the differentiation of B cells to antibody producing plasma cells (Hibi et al; 1996).

The induction of IL-6 by live HIV preparations occurred in the absence of T cells and could be neutralized by human anti-HIV serum indicating that HIV was responsible for this IL-6 inducing activity. It has been demonstrated that IL-6 can be produced by a variety of cells upon various kinds of stimulation: T cells infected with HTLV-1, fibroblasts stimulated with polyI:C, IL-1, platelet-derived growth factor, TNF- α , FCS, or LPS, and monocyte/macrophages stimulated with LPS. Monocyte/macrophages, one of the target cells of HIV, produced IL-6 upon stimulation with both live and inactivated HIV (Nakajima et al; 1989). A study of women treated for cervical intraepithelial lesions showed that after treatment, there were increased levels of genital HIV, TNF- α , IL-6, and other activation markers in cervicovaginal lavage (Spear et al; 2008). In univariate analysis, genital tract HIV RNA was significantly associated with plasma HIV RNA and several of the cytokines, while in multivariate analysis, genital tract HIV RNA was significantly associated only with plasma HIV RNA and IL-6 (Spear et al; 2008). Another study was done to determine the effect of HIV on thymic stromal cells and the production of cytokines important in thymocyte development, three types of adherent thymic cultures were established and studied: thymic epithelial cells (TECs), macrophage-enriched, and mixed cultures of macrophages and TECs (M phi/TEC). M phi/TEC and macrophage-enriched cultures were infected by both HIV strains without cytopathic changes. The TECs grew well in culture exposed to HIV-1 strains HIV-1IIIB and HIV-1Ba-L for at least 6 weeks and showed no evidence of infection, cytopathology, or changes in cytokine production with HIV. Only cultures containing macrophages (M phi/TEC or macrophage enriched) showed changes in cytokine (IL-1 alpha, IL-1 beta, and IL-6) production with HIV. Unstimulated macrophage-enriched cultures produced small amounts of IL-6 that were increased by HIV 20-fold (Sandborg et al; 1994).

There are many studies showing the increase of IL-6 expression within HIV infected cells but not many studies suggesting what IL-6 does to HIV. In a study done by Miles in 1990, it was found that IL-6 might actually be a growth factor for the HIV virus (Miles et al; 1990). There was a proliferative response of the AIDS-Kaposi sarcoma (AIDS-KS) cells to high concentrations of hrIL-6 and the detection of IL-6-rRNA in the areas of the skin involved with Kaposi sarcoma. AIDS-KS cells synthesized, released, and responded to biologically active IL-6. AIDS-KS cells, in which IL-6 protein translation arrest was induced by an IL-6 anti sense oligodeoxynucleotide, did not proliferate optimally unless exogenous hrIL-6 was added (Miles et al; 1990).

8. Interleukin-17 (IL-17)

IL-17 is an inflammatory cytokine that is exclusively produced by a recently discovered subset of CD4⁺ T helper (Th) cells, referred to as Th17 cells (Crome et al; 2009). This cytokine has been found to help regulate the inflammatory response by activating fibroblasts, recruiting neutrophils, and acting on macrophages to promote both their recruitment and survival (Crome et al; 2009, Chang et al; 2007). In addition, IL-17 is thought to play a significant role in activating and inducing anti-microbial peptides and pro-inflammatory cytokines like IL-6, CCL2, and TNF- α (Crome et al; 2009, Chang et al; 2007). Furthermore, high levels of this cytokine have been linked to a number of inflammatory diseases including rheumatoid arthritis, multiple sclerosis (MS), and asthma. Low levels, on the other hand, are thought to cause both impaired host defense against mycobacterial infection and decreased antibacterial immunity (Crome et al; 2009, Brenchley et al; 2008).

Studies of the effects of HIV on IL-17 concentrations using flow cytometry have found that HIV-infected patients have significantly increased levels of IL-17 (Giorgio, 2003). Venketaraman *et al.* (unpublished data) was also able to show increased levels of IL-17 in HIV-infected blood plasma using ELISA assays. However, Brenchley *et al.*; 2008 noted that there were significantly fewer IL-17 producing Th17 cells in the gastrointestinal tract of HIV-infected patients. In fact, the study indicated that Th17 cells were preferentially targeted during HIV infection.

The decrease of IL-17 concentrations at the mucosal wall of the gastrointestinal tract could greatly increase the probability of bacterial infections, which could in turn have significant implications for the speed of HIV pathogenesis (Brenchley et al; 2008). As Levy et al; 2009 noted, chronic immune activation increases the production of pro-inflammatory cytokines (IL-6, IL-17, TNF- α , etc.). This up-regulation of pro-inflammatory cytokines often leads to the rapid loss of CD4⁺ T cells via apoptosis. Decreased IL-17 concentrations due to HIV infection can therefore ultimately lead to the general advancement of HIV by creating an environment favorable to opportunistic infection and chronic immune activation (Maek-A-Nantawat et al; 2007).

9. Interleukin-12 (IL-12)

IL-12 is a heterodimeric pro-inflammatory cytokine that is produced by dendritic cells and phagocytes during an infection (Giorgio, 2003). It is a cytokine identified as a master switch for leading the naïve CD4⁺ T cells towards the Th1 pathway and also activating NK cells (Villinger and Ansari, 2010). Not only does it directly induce T, NK, and NKT cell cytotoxicity, IL-12 also promotes macrophage activity via T- and NK-cell-produced IFN- γ (Giorgio, 2003, Egilmez et al; 2011). The pathway is antagonized in the presence of IL-10 (Villinger and Ansari, 2010).

IL-12 plays important roles in protecting the body from various microbial infections such as parasites, bacteria, and viruses (Yang et al; 2010). With mutations in genes of the IL-12, the cells are susceptible to intracellular pathogens such as tuberculosis, leprosy, HIV-1, hepatitis and malaria (Vannberg et al; 2011). One of the characteristics of HIV infection is the gradual deterioration of cellular effector responses. Studies has concluded that CD4⁺ and CD8⁺ T cell responses were enhanced *ex vivo* by the addition of IL-12, but that capacity to respond is decreased in patients with marked CD4 loss (Villinger and Ansari, 2010). Louis *et al.*; 2010, also added that IL-12 production required the presence of IFN- γ . Therefore, as HIV

progresses, decreased IFN- γ leads to decrease in IL-12 which leads to decreased CD4⁺ and CD8⁺ T cell response.

A decrease of IL-12 concentration increases the probability for opportunistic infections. Taoufik *et al*; 1997 and Mirani *et al*; 2002, showed IL-12 mRNA was diminished while IL-10 production was up-regulated in the presence of *Staphylococcus aureus* and HIV gp120, further inhibiting IL-12 cytokine production. Even though IL-12 is potent, Villinger and Ansari 2010, noted that when IL-12 therapy was administered in the late stages of HIV, it failed to restore normal levels of CD4 T cells and IFN- γ .

10. Additional effects of HIV on IFN- γ signaling

In addition to the Th1 subset response mediation mentioned earlier, IFN- γ normally acts on APCs to enhance their expression of major histocompatibility complex II (MHC-II), thereby enhancing their antigen presentation ability (Li *et al*; 2011). HIV transactivator protein (TAT) interferes with the intracellular signaling normally performed by the IFN- γ bound IFN- γ receptor (Cheng *et al*; 2009). In so doing, the TAT protein lowers the antigen presentation capacity of dendritic cells and macrophages, further limiting the immune response to the invading virus (Salgame *et al*; 2009).

11. The transforming growth factor β (TGF- β)

TGF- β cytokine family are closely related polypeptides which include tissue growth factors that have a diverse range of proteins that regulate many physiological processes including embryonic development, homeostasis, wound healing, chemotaxis, cell cycle control, cell proliferation, differentiation, apoptosis, adhesion, and migration (Leask and Abraham, 2004). TGF- β is one of the most immunosuppressive substances produced in the body and yet may inhibit or stimulate cell growth, depending on the cell type and culture conditions (Liu and Gaston Pravia, 2010). TGF- β is produced in many immune cells including lymphocytes, macrophages and dendritic cells (Liu and Gaston Pravia, 2010). Receptors for TGF- β have been found on all cell lines tested, allowing this cytokine to have effects on almost any tissue in the body (Leask and Abraham, 2004). It has also been shown to play a central role in tissue fibrosis (Leask and Abraham, 2004). Because of the multifunctional role played by TGF- β , it plays a central role in the pathogenesis of many diseases. (Leask and Abraham, 2004).

There are three forms of TGF- β (TGF- β 1, TGF- β 2, and TGF- β 3) in mammalian cells. TGF- β s are synthesized using inactive precursors and cannot bind receptors until they are activated. After release of TGF- β from cells they associate with latency-associated protein and form a small inactive complex. In the extracellular matrix, this complex is bound by latent TGF- β -binding protein (LTBP), a component of the extracellular matrix that is necessary for the secretion and storage of TGF- β (Letterio and Roberts, 1998). Intracellular activity of TGF- β is mediated by the actions of Smad transcription factors as well as independent factors (Letterio and Roberts, 1998). Active Smad complexes bind to DNA weakly and high affinity binding is achieved by the association of Smad proteins with a large number of transcription factor partners (Massague, 1992). The variations of Smad proteins in transcriptional regulations and the diversity of Smad-independent pathways allow the pleiotropic actions of TGF- β (Letterio and Roberts, 1998).

HIV infection leads to a variety of disturbances in cytokine expression that can lead to a state of chronic activation of B cells and release of cytokines that may actually play an important role in the pathogenesis of HIV infection (Li and Flavell, 2008). Early HIV-1 infection is associated with a massive oligoclonal expansion of CD8 T cells (Massague and Gomis, 2006), however despite the high number of circulating CD8⁺ T cells the cytotoxic T lymphocyte (CTL) response is highly variable among HIV-1 infected individuals (Poli and Fauci, 1993). It has also been shown that the immune dysfunction in the initial phase of HIV infection exceeds CD4⁺ T cell infection and loss (Pantaleo et al; 1994). It appears that the immunosuppression effect occurs almost immediately upon infection (Garba et al; 2002). The result is diminished T cell response to antigen stimulation and persistence of HIV replication (Pantaleo and Fauci, 1995).

HIV-1 products such as TAT, induce the transcription of cytokines with immunosuppressive effects, including TGF- β (Cohen et al; 1999). It has been reported that extracellular TAT can be taken up by bystander cells and that it is possible that exogenous TAT, not associated with direct infection of a cell, can induce TGF- β transcription in immune competent cells (Pantaleo et al; 1993). Macrophages appear to be very sensitive to TAT and are affected by TAT concentrations 1,000-fold lower (500 pM) than those that affect T cells (Cohen et al; 1999). Macrophages stimulated by TAT either by infection or by the uptake of soluble TAT (sTat) induce Fas ligand (FasL), which in turn can trigger the apoptosis of antigen-reacting, Fas-expressing helper T cells. This mechanism would suppress T-cell dependent cellular and humoral immune responses to both HIV and other antigens (Cohen et al; 1999).

The transactivating effect of HIV-1 TAT is mediated by activator protein-1, which is the same multimolecular complex that is activated by TGF- β (Cohen et al; 1999). HIV-1 can induce both the transcription and secretion of TGF- β (Reinhold et al; 1999) and the induction of TGF- β can increase the apoptosis of NK cells (Poggi and Zocchi, 2006). TGF- β and Tat have been detected in the sera of early HIV-1 infected individuals at levels that were biologically active *in vitro* (Reinhold et al; 1999).

Some HIV-infected individuals have been shown to lose the ability of their cytotoxic T lymphocytes CTL (CD8⁺) to control infection in cells that carry HIV as well as other infectious agents (Pantaleo et al; 1993). About 25% of HIV infected individuals have been shown to produce TGF- β 1 in response to stimulation with HIV proteins or peptides (Garba et al; 2002). It has been shown that the loss of CTL activity is related to the production of TGF- β 1 in sufficient amounts to significantly reduce the IFN- γ response of CD8⁺ cells to both HIV and other viral proteins such as vaccinia virus (Garba et al; 2002).

It has been established that feline CD4⁺CD25⁺ T regulatory (T reg) cells share phenotypic and functional characteristics with human and murine T reg cells (Vahlenkamp et al; 2004). Early in the infection with feline immunodeficiency virus (FIV), CD4⁺CD25⁺ T reg cells exhibit increased expression of a membrane TGF- β (mTGF- β) (Mexas et al; 2008). The appearance of TGF- β ⁺CD4⁺CD25⁺ lymphocytes within the lymph node of FIV⁺ cats occurs in both acute and chronic FIV, even though mTGF- β does not appear in the blood (Fogle et al; 2010). There is also evidence of increased expression of TGF- β RII, the receptor of TGF- β 1, on CD8⁺ lymphocytes in FIV⁺ cats that would make the CD8⁺ lymphocytes much more sensitive to TGF- β inhibition (Fogle et al; 2010). In FIV lentiviral infection, during both the acute and chronic stages of infection, CD4⁺CD25⁺ Tregs suppress CD8⁺ responses and the CD4⁺CD25⁺ Tregs use mTGF- β to suppress IFN- γ expression resulting in suppression of CD8⁺ lymphocyte function (Fogle et al; 2010). These findings help explain the paradox of chronic HIV-1 infection, in which CD8⁺ T cells display an activated phenotype but exhibit

reduced effector function (Fogle et al; 2010, Bucci et al; 1998, Tompkins and Tompkins 2008). IL-10 and TGF- β overlap with each other in many of their biological effects including inhibition of T cell proliferation and IFN- γ production (Othieno et al; 1999).

12. Interleukin-10 (IL-10)

IL-10 is an anti-inflammatory cytokine that essentially plays two regulatory roles in innate and adaptive immunity. It suppresses the up-regulation of various genes in macrophages and dendritic cells that are normally stimulated via toll-like receptors and promotes the proliferation of cytotoxic T cells, activates B cells, and induces the upregulation of specific genes in toll-like receptor activated phagocytic and dendritic cells (Trinchieri, 2007). In addition, a critical function of IL-10 is its ability to inhibit pro-inflammatory cytokines such as TNF- α , IFN- γ , IL-1, IL-6, IL-2, and IL-12 (Trinchieri, 2007). IL-10 decreases the production of pro-inflammatory cytokines by limiting the major histocompatibility class II and CD80/CD 86 expressed on monocytes and macrophage (Wang et al; 2005). IL-10 was believed to be produced by CD4⁺ Th2 cells; however, studies have shown that it is secreted by both Th1 and Th2 cells (Brockman et al; 2009). Also, cells from the myeloid lineage which include macrophages and dendritic cells also produce cytokine IL-10 (Hedrich and Bream, 2010). Furthermore, IL-10 is regulated both at the transcriptional and post-translational level and is involved in various signaling pathways (Couper et al; 2008).

There are several speculations of the role of IL-10 in HIV pathogenesis and the subject has been a popular interest in many studies. Ji *et al.*, 2005 reported that CD14⁺ monocytes are the main cells producing cytokine IL-10 in PBMCs after HIV-1 infection via interactions independent of CD4⁺ molecules, thus, concluding that IL-10 production is dependent on the presence of CD14⁺ monocytes. Moreover, as the patient progresses to advanced stages of HIV disease, the frequency of IL-10 producing cells increases significantly (118). On the other hand, Naicker *et al.*, 2009, stated that different stages of the HIV disease will govern what role IL-10 will play in infected individuals. For instance, in acute HIV-1 individuals, IL-10 may promote viral replication by inhibiting effector immune response from both arms of the innate and adaptive immunity (Naicker et al; 2009). Furthermore, it was proposed in a chronic phase, that IL-10 resembled a protective role by reducing immune activation, inhibiting virus replication in macrophages, and the increase in production of IL-10 levels lowered plasma viral load and increased CD4⁺ cell count (Naicker et al; 2009).

13. HIV and free radicals

It has been shown that HIV infection induces increased production of free radicals by macrophages. Free radical formation occurs as a byproduct of oxidative stress. Oxidative stress occurs when there is a disproportion between the reactive oxygen elements in the body versus the ability of the body to properly eliminate these reactive species. The presence of free radicals has been implicated in disturbing and damaging a number of biological processes (Karthikeyan et al; 2010). With regards to HIV infection the increase of oxidative stress has been seen to influence components in antioxidant defense in physiological antioxidants such as glutathione which are seen to decrease dramatically in HIV patients (Pace and Leaf, 1995). In addition to glutathione, vitamin A, C, and E at high doses as well as improving low levels of selenium were associated with assisting the prevention of HIV infection progression by working as antioxidants to remove free radicals

(Garland and Fawzi, 1999). The aforementioned studies may provide a low cost method for improving the prognosis of HIV infected patients in high risk, underprivileged areas of the world.

Chronic oxidative stress is often associated with HIV infection and research indicates a benefit for increased antioxidant vitamins and supplements in reduction of DNA base damage, which in turn can slow progression of infection (Jaruga et al; 2002). Neutrophils from asymptomatic HIV patients show increased oxygen radical production which can be modified by treating with N-acetyl cysteine, a compound used as an antioxidant (Smietana et al; 2008). The role of free radical oxidative stress on DNA damage is correlated with stimulated DNA repair mechanisms which activate enzymes associated with initiation of apoptosis such as poly ADP-ribose transferase and p53. Reduced NAD/NADH production would lower ATP synthesis that in turn correlates with a deficiency in glutathione; which as mentioned is an endogenous antioxidant important in resolving imbalance of free radicals (Dobmeyer et al; 1997).

The progression of HIV is correlated with a decreased immunity. One way in which this decreased immunity progresses is by free radical overload of monocytes and granulocytes which leads to deficiency of antioxidant mechanisms which may lead to the loss of CD4 cells often seen in the progression of HIV (Dobmeyer et al; 1997). The decreased immunity may also be related to the reactive oxygen species and free radical presence which is higher in HIV infected patients. With HIV infection progression there is an increased production of reactive oxygen species which leads to the theory of free radical mediated apoptosis of lymphocytes which reduces the ability for immune response to progressive HIV infections (Dobmeyer et al; 1997). With regards to CD4 cell counts the apoptosis of lymphocytes by free radicals leads to progression of immunodeficiency and makes for a quicker transition from HIV infection to AIDS (Bautista, 2001). It has been published that during HIV-1 infection, hematopoietic cells are exposed to high amounts of free radicals. Subsequently there is a reduction of leukocytopoiesis and increase susceptibility to further infections (Masutani, 2000). Furthermore, there is a link to lipid peroxidation observed in patients with HIV or AIDS to a deficiency of antioxidants which leads to free radical proliferation (Favier et al; 1994).

Rate of viral replication is a key process to the proliferation of HIV infection. The conditions in which viruses such as HIV will proliferate seem to correlate with the presence of oxidative stress/free radicals *in vitro* (Fuchs et al; 1991). There tends to be an increase in nuclear transcription factor and inflammatory cytokine activation of the immune system (Brach et al; 1992). The progression of the virus/infection will then allow for opportunistic infections which then would also promote more oxidative stress due to increased free radical elements, again improving viral replication and weakening antioxidant defense (Knysz, 2007).

Damage or altering of the DNA repair machinery is an important aspect of the progression of HIV infection pathogenesis (Olinski et al; 2002). There is a slow and deliberate degradation of cellular components such as membrane blebbing, chromatin condensation, and DNA cleavage ability. Additionally, there is evidence that shows oxidative DNA damage will lead to the apoptotic cell death in HIV infected patients. There appears to be an increase in oxidatively modified DNA bases in HIV infected patients leading to what is known as pyrimidine and purine derived lesions. One specific lesion labeled, 8-OH-Gua was found in isolated lymphocytes of HIV patients. The presence of this lesion leads to transversions of DNA base pairs unless repaired before replication. The number of these

lesions was seen to be reduced in response to antioxidant supplement and vitamin treatments, which correlates to free radical influence in DNA damage, and potential progression of HIV infection (Olinski et al; 2002).

14. Conclusion

Both HIV-1 and HIV-2 cause AIDS, but HIV-1 is found worldwide, whereas HIV-2 is found primarily in West Africa. Chemokine receptors, such as CXCR4 and CCR5 proteins, are required for the entry of HIV into CD4-positive cells. After establishing infection, HIV alters the synthesis of host cytokines and chemokines and kills CD4+ T lymphocytes thereby resulting in the loss of cell-mediated immunity and a high probability that the host will develop opportunistic infections.

15. References

- Alfano, M and Poli, G. 2005. Role of cytokines and chemokines in the regulation of innate immunity and HIV infection. *Molecular Immunology* 42: 161-182.
- Bahia, M.S and Silakari, O. 2010. Tumor Necrosis Factor Alpha Converting Enzyme: An Encouraging Target for Various Inflammatory Disorders. *Chemical Biology & Drug Design*. 75: 415-443.
- Bautista AP. 2001 Free radicals, chemokines, and cell injury in HIV-1 and SIV infections and alcoholic hepatitis. *Free Radical Biology and Medicine*. 31(12):1527-1532.
- Brabers NA and Nottet HS. 2006. Role of the pro-inflammatory cytokines TNF-alpha and IL-1beta in HIV-associated dementia. *European journal of clinical investigation*. 36(7):447-58.
- Brach MA, de Vos S, Arnold C, Gruss HJ, Mertelsmann R and Herrmann F. 1992. Leukotriene B4 transcriptionally activates interleukin-6 expression involving NK-chi B and NF-IL6. *Eur J Immunol*. Oct 22(10):2705-2711.
- Brenchley, Jason M., Paiardini, M., Knox, Kenneth S., Asher, Ava I., Cervasi, B., Asher, Tedi E., Scheinberg, P., Price, David A., Hage, Chadi A., Kholi, Lisa M., Khoruts, A., Frank, I., Else, J., Schacker, T., Silvestri, G and Daniel C. Douek. 2008. Differential Th17 CD4 T-cell depletion in pathogenic and nonpathogenic lentiviral infections. *Blood* 112(7): 2826-2835.
- Brockman MA, Kwon DS, Tighe DP, Pavlik DF, Rosato PC, Sela J, Porichis F, Le Gall S, Waring MT, Moss K, Jessen H, Pereyra F, Kavanagh DG, Walker BD and Kaufmann DE. 2009. IL-10 is up-regulated in multiple cell types during viremic HIV infection and reversibly inhibits virus-specific T cells. *Blood*. 114(2):346-56.
- Bucci JG, Gebhard DH, Childers TA, English RV, Tompkins MB and Tompkins WA. 1998. The CD8+ cell phenotype mediating antiviral activity in feline immunodeficiency virus-infected cats is characterized by reduced surface expression of the CD8 beta chain. *The Journal of infectious diseases*. 178(4):968-77.
- Bujak M and Frangogiannis NG. 2009. The role of IL-1 in the pathogenesis of heart disease. *Arch Immunol Ther Exp (Warsz)*. 57(3):165-76.
- Burchett SK, Weaver WM, Westall JA, Larsen A, Kronheim S and Wilson CB. 1998. Regulation of tumor necrosis factor/cachectin and IL-1 secretion in human mononuclear phagocytes. *Journal of immunology*. 140(10):3473-81.

- Cannon, P and June, C. 2011. Chemokine receptor 5 knockout strategies. *Current Opinion in HIV and AIDS*. 6: 74-79.
- Chang, Seon H., and Chen Dong. 2007. A novel heterodimeric cytokine consisting of IL-17 and IL-17F regulates inflammatory responses. *Cell Research* 17: 435-440.
- Charo, Israel F. and Richard M. Ransohoff. 2006. The many roles of chemokines and chemokine receptors in inflammation. *The New England Journal of Medicine*. 345: 610-621.
- Cheng SM, Li JCB, Lin SS, Lee DCW, Liu L, Chen Z, and Lau ASY. 2009. HIV-1 transactivator protein induction of suppressor cytokine signaling-2 contributes to dysregulation of IFN- γ signaling. *Blood*. 13: 5192-5201.
- Cheung R, Ravyn V, Wang L, Ptasznik A and Collman RG. 2008. Signaling mechanism of HIV-1 gp120 and virion-induced IL-1 β release in primary human macrophages. *Journal of immunology*. 180(10):6675-84.
- Clerici M, Frances TH, Venzon D J, Blatt S, Hendrix CW, Wynn TA and Shearer GM. 1993. Changes in Interleukin-2 and Interleukin-4 Production in Asymptomatic Human Immunodeficiency Virus-seropositive Individuals. *J Clin Invest*. 91:759-765.
- Cohen SS, Li C, Ding L, Cao Y, Pardee AB, Shevach EM, et al. 1999. Pronounced acute immunosuppression in vivo mediated by HIV Tat challenge. *Proceedings of the National Academy of Sciences of the United States of America*. 96(19):10842-7.
- Corasaniti MT, Bilotta A, Strongoli MC, Navarra M, Bagetta G and Di Renzo G. 2001. HIV-1 coat protein gp120 stimulates interleukin-1 β secretion from human neuroblastoma cells: evidence for a role in the mechanism of cell death. *British journal of pharmacology*. 134(6):1344-50.
- Couper, K. N., Blount, D. G and Riley, E. M. 2008. IL-10: the master regulator of immunity to infection. *J Immunol*. 180(9): 5771-5777.
- Crome, S.Q., Wang, A.Y and M.K. Levings. 2009. Translational Mini-Review Series on Th17 Cells: Function and regulation of human T helper 17 cells in health and disease. *The Journal of Translational Immunology*. 159: 109-119.
- Daly, Christine and Barrett J. Rollins. 2003. Monocyte Chemoattractant Protein-1 (CCL2) in Inflammatory Disease and Adaptive Immunity: Therapeutic opportunities and controversies. *Microcirculation*. 10: 247-257.
- Deshmane, Satish L., Kremlev, S., Amini, S and Bassel E. Sawaya. 2009. Monocyte chemoattractant protein-1 (MCP-1): an overview. *Journal of Interferon & Cytokine Research*. 29(6): 313-326.
- Devadas K, Hardegen NJ, Wahl LM, Hewlett IK, Clouse KA, Yamada KM, et al. 2004. Mechanisms for macrophage-mediated HIV-1 induction. *Journal of immunology*. 173(11):6735-44.
- Dinarello CA. 1999. Cytokines as endogenous pyrogens. *The Journal of infectious diseases*. 179 Suppl 2:S294-304.
- Dobmeyer TS, Findhammer S, Dobmeyer JM, et al. 1997. Ex vivo induction of apoptosis in lymphocytes is mediated by oxidative stress: role for lymphocyte loss in HIV infection. *Free Radic Biol Med*. 22(5):775-785.
- Edo-Matas D., van Dort KA, Setiawan LC, Schuitemaker H, Kootstra NA. 2011. Comparison of in vivo and in vitro evolution of CCR5 to CXCR4 co-receptor use of primary human Immunodeficiency type 1 variants. *Virology*. 412(2):269-77.

- Egilmez NK, Harden JL, Virtuoso LP, Schwendener RA and Kilinc MO. 2011. Nitric oxide short-circuits interleukin-12-mediated tumor regression. *Cancer Immunology, Immunotherapy* 2011; 1-7.
- Emilie D, Peuchmaur M, Maillot MC, Crevon MC, Brousse N, Delfraissy JF, et al. 1990. Production of interleukins in human immunodeficiency virus-1-replicating lymph nodes. *The Journal of clinical investigation*. 86(1):148-59.
- Epstein LG and Gendelman HE. 1993. Human immunodeficiency virus type 1 infection of the nervous system: pathogenetic mechanisms. *Annals of neurology*. 33(5):429-36.
- Eugenin, Eliseo A., Osiecki, K., Lopez, L., Goldstein, H., Calderon, Tina M and Joan W. Berman. 2006. CCL2/monocyte chemoattractant protein-1 mediates enhanced transmigration of human immunodeficiency virus (HIV)-infected leukocytes across the blood-brain-barrier: a potential mechanism of HIV-CNS invasion and NeuroAIDS. *The Journal of Neuroscience*. 26(4): 1098-1106.
- Fantuzzi G. 2003. Cytokine knockouts. Totawa, NJ: Humana Press; p. 471.
- Favier A, Sappey C, Leclerc P, Faure P and Micoud M. 1994. Antioxidant status and lipid peroxidation in patients infected with HIV. *Chem Biol Interact*. 91(2-3):165-180.
- Fernandez-Ortega, C., Dubed, M., Ramos, T., Navea, L., Alvarez, G., Lobaina, L., Lopez, L., Casilla, D and Rodriguez, L. 2004. Non-induced leukocyte extract reduces HIV replication and TNF secretion. *Biochemical and Biophysical Research Communications*. 325: 1075-1081.
- Fogle JE, Mexas AM, Tompkins WA and Tompkins MB. 2010. CD4(+)CD25(+) T regulatory cells inhibit CD8(+) IFN-gamma production during acute and chronic FIV infection utilizing a membrane TGF-beta-dependent mechanism. *AIDS research and human retroviruses*. 26(2):201-16.
- Fuchs J, Ochsendorf F, Schofer H, Milbradt R and Rubsamen-Waigmann H. 1991. Oxidative imbalance in HIV infected patients. *Med Hypotheses*. 36(1):60-64.
- Gallo P, Frei K, Rordorf C, Lazdins J, Tavolato B and Fontana A. 1989. Human immunodeficiency virus type 1 (HIV-1) infection of the central nervous system: an evaluation of cytokines in cerebrospinal fluid. *J Neuroimmunol*. 23(2):109-16.
- Garba ML, Pilcher CD, Bingham AL, Eron J and Frelinger JA. 2002. HIV antigens can induce TGF-beta(1)-producing immunoregulatory CD8+ T cells. *Journal of immunology*. 168(5):2247-54.
- Garland M and Fawzi WW. 1999. Antioxidants and progression of human immunodeficiency virus (HIV) disease. *Nutrition Research*. 19(8):1259-1276.
- Gorry, P.R. and Ancuta, P. 2011. Coreceptors and HIV-1 Pathogenesis. *Curr HIV/AIDS Rep*. 8(1):45-53.
- Gorry, P.R., et al. 2007. Changes in region of gp120 contribute to unusually broad co-receptor usage of an HIV-1 isolate from CCR5 Delta32 heterozygote. *Virology*. 362: 163-178.
- Grivel, J., et al. 2010. Selective transmission of R5 HIV-1 variants: where is the gatekeeper? *Journal of Transitional Medicine*. 9 (Suppl 1): S6.
- Hedrich, C. M., & Bream and J. H. 2010. Cell type-specific regulation of IL-10 expression in inflammation and disease. *Immunol Res*. 47(1-3): 185-206.
- Heinrich, Peter C., Iris Behrmann, Serge Haan, Heike M. Hermanns, Gerhard Müller-Newen and Fred Schaper. 2003. Principles of Interleukin (IL)-6-type Cytokine Signalling and Its Regulation. *Institut Für Biochemie*. 374: 1-20.

- Herbein G and Varin A. 2010. The macrophage in HIV-1 infection: from activation to deactivation? *Retrovirology*. 7:33.
- Hibi M K, Nakajima and T. Hirano. 1996. IL-6 Cytokine Family and Signal Transduction: a Model of the Cytokine System. *Journal of Molecular Medicine*. 74.1: 1-12.
- Hutter G. 2009. Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. *N. Engl J Med*. 360: 692-698.
- Jaruga P, Jaruga B, Gackowski D, et al. 2002. Supplementation with antioxidant vitamins prevents oxidative modification of DNA in lymphocytes of HIV-infected patients. *Free Radic Biol Med*. 32(5):414-420.
- Ji, J., Sahu, G. K., Braciale, V. L and Cloyd, M. W. 2005. HIV-1 induces IL-10 production in human monocytes via a CD4-independent pathway. *Int Immunol*. 17(6): 729-736.
- Karthikeyan R, Manivasagam T, Anantharaman P, Balasubramanian T and Somasundaram S. 2010. Chemopreventive effect of *Padina boerghesii* extracts on ferric nitrilotriacetate (Fe-NTA)-induced oxidative damage in Wistar rats. *Journal of Applied Phycology*. 1-7
- Kaul M, Garden GA and Lipton SA. 2001. Pathways to neuronal injury and apoptosis in HIV-associated dementia. *Nature*. 410(6831):988-94.
- Kindberg, E., Mickiene A, Ax C, Akerlind B, Vene S, Lindquist L, Lundkvist A, Svensson L. 2008. A deletion in the chemokine receptor 5 (CCR5) gene is associated with tickborne encephalitis. *J Infect Dis*. 197:266-9.
- Kwong P.D., Wyatt R, Sattentau QJ, Sodroski J, Hendrickson WA. 2000. Oligomeric modeling and electrostatic analysis of the gp120 envelope glycoprotein of human immunodeficiency virus. *J Virol*. 74:1961-1972.
- Klein SA, Dobmeyer JM, Dobmeyer TS, Pape M, Ottmann OG, Helm EB, Hoelzer D and Rossol R, 1997. Demonstration of the Th1 to Th2 cytokine shift during the course of HIV-1 infection using cytoplasmic cytokine detection on single cell level by flow cytometry. *AIDS*. 11:1111-1118
- Knysz B, Szetela B and Gladysz A. 2007. Pathogenesis of HIV-1 infection - chosen aspects. *HIV & AIDS Review*. 6(1):7-11.
- Leask A and Abraham DJ. 2004. TGF-beta signaling and the fibrotic response. *The FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 18(7):816-27.
- Leghmari, K., Bennasser, Y., Tkaczuk, J and Bahraoui, E. 2008. HIV-1 Tat protein induces IL-10 production by an alternative TNF-a-independent pathway in monocytes: Role of PKC- δ and p38 MAP kinase. *Cellular Immunology*. 253: 45-53.
- Lepe-Zuniga JL, Mansell PW and Hersh EM. 1987. Idiopathic production of interleukin-1 in acquired immune deficiency syndrome. *Journal of clinical microbiology*. 25(9):1695-700.
- Letterio JJ and Roberts AB. 1998. Regulation of immune responses by TGF-beta. *Annual review of immunology*. 16:137-61.
- Levy J. 2007. HIV and the pathogenesis of AIDS. Washington, DC: ASM Press.
- Levy and Jay A. 2009. HIV pathogenesis: 25 years of progress and persistent challenges. *AIDS*. 23:147-160.
- Li JCB, Au K, Fang J, Yim HCH, Chow K, Ho P and Lau ASY. 2011. HIV-1 trans-activator protein dysregulates IFN-g signaling and contributes to the suppression of autophagy induction. *AIDS*. 25:15-25.

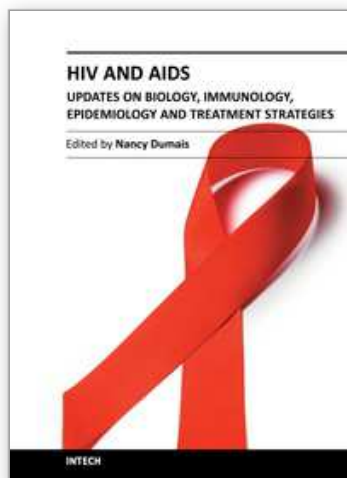
- Li MO and Flavell RA. 2008. TGF-beta: a master of all T cell trades. *Cell*. 134(3):392-404.
- Lim JK, Louie CY, Glaser C, Jean C, Johnson B, Johnson H, McDermott DH, Murphy PM. 2008. Genetic deficiency of chemokine receptor CCR5 is a strong risk factor for symptomatic West Nile virus infection: a meta-analysis of 4 cohorts in the US epidemic. *J Infect Dis*. 197:262-5.
- Lipton SA. 1998. Neuronal injury associated with HIV-1: approaches to treatment. *Annu Rev Pharmacol Toxicol*. 38:159-77.
- Liu RM and Gaston Pravia KA. 2010. Oxidative stress and glutathione in TGF-beta-mediated fibrogenesis. *Free radical biology & medicine*. 48(1):1-15.
- Louis S., Dutertre CA., Vimeux L., Fery L., Henno L., Diocous S., Kahi S., Deveau C., Meyer L., Goujard C and Hosmalin A. 2010. IL-23 and IL-12p70 production by monocytes and dendritic cells in primary HIV-1 infection. *Journal of Leukocyte Biology*. 87(4):645.
- Maek-A-Nantawat, W, Buranapraditkun, S, Klaewsongkram, J and Kiat Ruxrungthum. 2007. Increased interleukin-17 production both in helper T cell subset Th17 and CD4-negative T cells in human immunodeficiency virus infection. *Viral Immunology*. 20(1): 66-75.
- Mahad, Don J and Richard M. Ransohoff. 2003. The role of MCP-1 (CCL2) and CCR2 in multiple sclerosis and experimental autoimmune encephalomyelitis (EAE). *Seminars in Immunology*. 15: 23-32.
- Marui N, Offermann MK, Swerlick R, Kunsch C, Rosen CA, Ahmad M, et al. 1993. Vascular cell adhesion molecule-1 (VCAM-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. *The Journal of clinical investigation*. 92(4):1866-74.
- Martinon F, Burns K and Tschopp J. 2002. The inflammasome: a molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Molecular cell*. 10(2):417-26.
- Massague J. 1992. Receptors for the TGF-beta family. *Cell*. 69(7):1067-70.
- Massague J and Gomis RR. 2006. The logic of TGF-beta signaling. *FEBS letters*. 580(12):2811-20.
- Mariani, S.A. 2010. Asymmetric HIV-1 co-receptor use and replication in CD4+ T lymphocytes. *Journal of Translational Medicine*. 9 (Suppl 1):S8.
- Masutani H. 2000. Oxidative stress response and signaling in hematological malignancies and HIV infection. *Int J Hematol*. 71(1):25-32.
- Merrill JE, Koyanagi Y and Chen IS. 1989. Interleukin-1 and tumor necrosis factor alpha can be induced from mononuclear phagocytes by human immunodeficiency virus type 1 binding to the CD4 receptor. *Journal of virology*. 63(10):4404-8.
- Merrill JE, Koyanagi Y, Zack J, Thomas L, Martin F and Chen IS. 1992. Induction of interleukin-1 and tumor necrosis factor alpha in brain cultures by human immunodeficiency virus type 1. *Journal of virology*. 66(4):2217-25.
- Mexas AM, Fogle JE, Tompkins WA and Tompkins MB. 2008. CD4+CD25+ regulatory T cells are infected and activated during acute FIV infection. *Vet Immunol Immunopathol*. 126 (3-4):263-72.
- Miles, Steven A., Ahmad R. Rezai, Jesus F. Salazar-Gonzalez, Meta Vander Meyden, Ronald H. Stevens, Diane M. Logan, Ronald T. Mitsuyasu, Tetsuya Taga, Toshio Hirano, Tadimitsu Kishimoto and Otoniel Matinez-Maza. 1990. AIDS Kaposi Sarcoma-

- Derived Cells Produce and Respond to Interleukin 6. *Proceedings of the National Academy of Sciences*. 87.11: 4068-072.
- Mirani M, Elenkov I, Volpi S, Hiroi N, Chrousos GP and Kino T. 2002. HIV-1 protein Vpr suppresses IL-12 production from human monocytes by enhancing glucocorticoid action: potential implications of Vpr coactivator activity for the innate and cellular immunity deficits observed in HIV-1 infection. *Journal of Immunology*.169:6361.
- Molina JM, Scadden DT, Byrn R, Dinarello CA and Groopman JE. 1989. Production of tumor necrosis factor alpha and interleukin 1 beta by monocytic cells infected with human immunodeficiency virus. *The Journal of clinical investigation*. 84(3):733-7.
- Naicker, D. D., Werner, L., Kormuth, E., Passmore, J. A., Mlisana, K., Karim, S. A., et al. 2009. Interleukin-10 promoter polymorphisms influence HIV-1 susceptibility and primary HIV-1 pathogenesis. *J Infect Dis*. 200(3): 448-452.
- Nakajima K, Martínez-Maza O, Hirano T, Breen EC, Nishanian PG, Salazar-Gonzalez JF, Fahey JL and Kishimoto T. 1989. Induction of IL-6 (B cell stimulatory factor-2/IFN-beta 2) production by HIV. *J Immunol*.142(2):531-6
- Nambu A and Nakae S. 2010. IL-1 and Allergy. *Allergol Int*. 59(2):125-35.
- Olinski R, Gackowski D, Foksinski M, Rozalski R, Roszkowski K and Jaruga P. 2002. Oxidative DNA damage: assessment of the role in carcinogenesis, atherosclerosis, and acquired immunodeficiency syndrome. *Free Radical Biology and Medicine*. 33(2):192-200.
- Olney JW, Zorumski C, Price MT and Labruyere J. 1990. L-cysteine, a bicarbonate-sensitive endogenous excitotoxin. *Science (New York, NY)*. 248(4955):596-9.
- Osakwe CE, Bleotu C, Chifiriuc MC, Crancea C, Otelea D, Paraschiv S, Petrea S, Dinu M, Baicus C, Streinu-Cercel A and Lazar V. 2010. TH1/TH2 Cytokine Levels as an Indicator for Disease Progression in Human Immunodeficiency Virus Type 1 Infection and Response to Antiretroviral Therapy. *Roum Arch Microbiol Immunol*. 69(1):24-34.
- Othieno C, Hirsch CS, Hamilton BD, Wilkinson K, Ellner JJ and Toossi Z. 1999. Interaction of *Mycobacterium tuberculosis*-induced transforming growth factor beta1 and interleukin-10. *Infect Immun*. 67(11):5730-5.
- Pace GW and Leaf CD. 1995. The role of oxidative stress in HIV disease. *Free Radic Biol Med*. 19(4):523-528.
- Pantaleo G, Graziosi C, Demarest JF, Butini L, Montroni M, Fox CH, et al. 1993. HIV infection is active and progressive in lymphoid tissue during the clinically latent stage of disease. *Nature*. 362(6418):355-8.
- Pantaleo G, Demarest JF, Soudeyans H, Graziosi C, Denis F, Adelsberger JW, et al. 1994. Major expansion of CD8+ T cells with a predominant V beta usage during the primary immune response to HIV. *Nature*. 370(6489):463-7.
- Pantaleo G and Fauci AS. 1995. New concepts in the immunopathogenesis of HIV infection. *Annual review of immunology*. 13:487-512.
- Persidsky Y, Buttini M, Limoges J, Bock P and Gendelman HE. 1997. An analysis of HIV-1-associated inflammatory products in brain tissue of humans and SCID mice with HIV-1 encephalitis. *Journal of neurovirology*. 3(6):401-16.
- Poggi A and Zocchi MR. 2006. HIV-1 Tat triggers TGF-beta production and NK cell apoptosis that is prevented by pertussis toxin B. *Clinical & developmental immunology*. 13(2-4):369-72.

- Poli G and Fauci AS. 1993. Cytokine modulation of HIV expression. *Semin Immunol.* 5(3):165-73.
- Reinhold D, Wrenger S, Kahne T and Ansorge S. 1999. HIV-1 Tat: immunosuppression via TGF-beta1 induction. *Immunology today.* 20(8):384-5.
- Salgame P, Guan MX, Agahtehrani A and Henderson EE. 1998. Infection of T Cell Subsets by HIV-1 and the Effects of Interleukin-12. *J. Interferon and Cytokine Res.* 18:521-528.
- Samson, M. 1996. Resistance to HIV-1 infection in Caucasian individuals bearing mutant alleles of the CCR-5 chemokine receptor gene. *Nature.* 382: 722-725.
- Sandborg, Christy L, Karen L. Imfeld, Frank Zaldivar and Monique A. Berman. 1994. HIV Type 1 Induction of Interleukin 1 and 6 Production by Human Thymic Cells. *AIDS Research and Human Retroviruses.* 10.10: 1221-229.
- Smietana M, Clayette P, Mialocq P, Vasseur J-J and Oiry J. 2008. Synthesis of new N-isobutyryl-L-cysteine/MEA conjugates: Evaluation of their free radical-scavenging activities and anti-HIV properties in human macrophages. *Bioorganic Chemistry.* 36(3):133-140.
- Spear, Gregory T., M. Reza Zariffard, Hua Y. Chen, Joshua J. Anzinger, Kathryn Anastos, John Rusine, John Gatabazi, Audrey L. French, Mardge Cohen and Alan L. Landay. 2008. Positive Association between HIV RNA and IL-6 in the Genital Tract of Rwandan Women. *AIDS Research and Human Retroviruses.* 24.7: 973-76.
- Suleiman, J. 2010. Vicriviroc in combination therapy with an optimized regimen for treatment-experienced subjects: 48-week results of the VICTOR-E1 phase 2 trial. *J Infect Dis.* 201:590-9.
- Tagliamonte, M, Tornesello ML, Buonaguro FM, Buonaguro L. 2011. Conformational HIV-1 envelope on particulate structures: a tool for chemokine coreceptor binding studies. *Journal of Translational Medicine.* 9 (Suppl 1):S1.
- Taoufik Y, Lantz O, Wallon C, Charles A, Dussaix E and Delfraissy JF. 1997. Human immunodeficiency virus gp120 inhibits interleukin-12 secretion by human monocytes: an indirect interleukin-10-mediated effect. *Blood.* 89: 2842.
- Tenorio, A. R. 2011. The monoclonal CCR5 antibody PRO-140: the promise of once-weekly HIV therapy. *Curr HIV/ AIDS Rep.* 8(1):1-3.
- Tompkins MB and Tompkins WA. 2008. Lentivirus-induced immune dysregulation. *Vet Immunol Immunopathol.* 123(1-2):45-55.
- Trinchieri Giorgio. 2003. Interleukin-12 and the regulation of innate resistance and adaptive immunity. *Nature Reviews Immunology.* 3: 133.
- Trinchieri, G. 2007. Interleukin-10 production by effector T cells: Th1 cells show self control. *J Exp Med.* 204(2): 239-243.
- Tyor WR, Glass JD, Griffin JW, Becker PS, McArthur JC, Bezman L, et al. 1992. Cytokine expression in the brain during the acquired immunodeficiency syndrome. *Annals of neurology.* 31(4):349-60.
- Vahlenkamp TW, Tompkins MB and Tompkins WA. 2004. Feline immunodeficiency virus infection phenotypically and functionally activates immunosuppressive CD4+CD25+ T regulatory cells. *Journal of immunology.* 172(8):4752-61.
- Vannberg FO, Chapman SJ and Hill AVS. 2011. Human genetic susceptibility to intracellular pathogens. *Immunological Reviews.* 240:105.

- Villinger F and Ansari AA. 2010. Role of IL-12 in HIV infection and vaccine. *European Cytokine Network*. 21:215.
- Wahl LM, Corcoran ML, Pyle SW, Arthur LO, Harel-Bellan A and Farrar WL. 1989. Human immunodeficiency virus glycoprotein (gp120) induction of monocyte arachidonic acid metabolites and interleukin 1. *Proceedings of the National Academy of Sciences of the United States of America*. 86(2):621-5.
- Wang, Z. Y., Sato, H., Kusam, S., Sehra, S., Toney, L. M and Dent, A. L. 2005. Regulation of IL-10 gene expression in Th2 cells by Jun proteins. *J Immunol*. 174(4): 2098-2105.
- Weiss L, Haeffner-Cavaillon N, Laude M, Gilquin J and Kazatchkine MD. 1989. HIV infection is associated with the spontaneous production of interleukin-1 (IL-1) in vivo and with an abnormal release of IL-1 alpha in vitro. *AIDS (London, England)*. 3(11):695-9.
- Weiss L, Si-Mohamed A, Giral P, Castiel P, Ledur A, Blondin C, Kazatchkine MD and N Haeffner-Cavaillon. 1997. Plasma levels of monocyte chemoattractant protein-1 but not those of macrophage inhibitory protein-1 α and RANTES correlate with virus load in human immunodeficiency virus infection. *Journal of Infectious Diseases*. 176: 1621-1624.
- Xing HQ, Hayakawa H, Izumo K, Kubota R, Gelpi E, Budka H, et al. In vivo expression of proinflammatory cytokines in HIV encephalitis: an analysis of 11 autopsy cases.
- Xing, Z., J. Gauldie, G. Cox, H. Baumann, M. Jordana, X. F. Lei and M. K. Achong. 1998. IL-6 Is an Anti-inflammatory Cytokine Required for Controlling Local or Systemic Acute Inflammatory Responses. *Journal of Clinical Investigation*. 101.2: 311-20.
- Yang H, Wei J, Zhang H, Song W, Wei W, Zhang L, Qian K and He S. 2010. Upregulation of Toll-like Receptor (TLR) expression and release of cytokines from mast cells by IL-12. *Cell Physiol Biochem*. 26(3):337-46.
- Yeh MW, Kaul M, Zheng J, Nottet HS, Thylin M and Gendelman HE, et al. 2000. Cytokine-stimulated, but not HIV-infected, human monocyte-derived macrophages produce neurotoxic levels of L-cysteine. *Journal of immunology*. 164(8):4265-70.

IntechOpen



HIV and AIDS - Updates on Biology, Immunology, Epidemiology and Treatment Strategies

Edited by Dr. Nancy Dumais

ISBN 978-953-307-665-2

Hard cover, 694 pages

Publisher InTech

Published online 26, October, 2011

Published in print edition October, 2011

The continuing AIDS pandemic reminds us that despite the unrelenting quest for knowledge since the early 1980s, we have much to learn about HIV and AIDS. This terrible syndrome represents one of the greatest challenges for science and medicine. The purpose of this book is to aid clinicians, provide a source of inspiration for researchers, and serve as a guide for graduate students in their continued search for a cure of HIV. The first part of this book, “From the laboratory to the clinic,” and the second part, “From the clinic to the patients,” represent the unique but intertwined mission of this work: to provide basic and clinical knowledge on HIV/AIDS.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Vishwanath Venketaraman, Devin Morris, Clare Donohou, Andrea Sipin, Steven Kung, Hyoung Oh, Mesharee Franklin, John P. Murad, Fadi T. Khasawneh, Beatrice Saviola, Timothy Guilford and Clare Donahue (2011). Role of Cytokines and Chemokines in HIV Infection, HIV and AIDS - Updates on Biology, Immunology, Epidemiology and Treatment Strategies, Dr. Nancy Dumais (Ed.), ISBN: 978-953-307-665-2, InTech, Available from: <http://www.intechopen.com/books/hiv-and-aids-updates-on-biology-immunology-epidemiology-and-treatment-strategies/role-of-cytokines-and-chemokines-in-hiv-infection>

INTECH
open science | open minds

InTech Europe

University Campus STeP Ri
Slavka Krautzeka 83/A
51000 Rijeka, Croatia
Phone: +385 (51) 770 447
Fax: +385 (51) 686 166
www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai
No.65, Yan An Road (West), Shanghai, 200040, China
中国上海市延安西路65号上海国际贵都大饭店办公楼405单元
Phone: +86-21-62489820
Fax: +86-21-62489821

© 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the [Creative Commons Attribution 3.0 License](https://creativecommons.org/licenses/by/3.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen