We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Pathogenesis and Pathology of Chagas' Chronic Myocarditis

Julián González, Roberto Guerri-Guttenberg, Daniel Grana, Francisco Azzato and José Milei Instituto de Investigaciones Cardiológicas Prof. Dr. A. Taquini – UBA – CONICET Argentina

1. Introduction

Chronic chagasic cardiomyopathy (CCC) is the most serious manifestation of the chronic phase of Chagas' disease and constitutes the most common type of chronic myocarditis in the world (Guerri-Guttenberg, et al., 2008, Milei, et al., 1996a, Milei, et al., 2009, Milei, et al., 1992a, Storino, et al., 1992). Chagas' disease, a chronic illness caused by the flagellate parasite *Trypanosoma cruzi* (*T. cruzi*), was first described in 1909 by the Brazilian physician Carlos Chagas (Chagas C, 1909). The insect vectors of the disease are present throughout most of South and Central America, and their zone of distribution extends across the southern United States (Rassi, et al., 2010). It was estimated by year 2000, that in endemic areas 40 million people were considered to be at risk of infection, being 20 million already infected. Every year near 200,000 new cases are expected to happen, and 21,000 deaths per year occur (WHO, 2005).

Although always considered to be confined to Latin America, due to migratory movements from endemic countries to Europe and North America, Chagas' disease is being detected more frequently in developed countries. Europe is estimated to have from 24,001 to 38,708 (lower or upper limit of estimate, respectively) immigrants with *T. cruzi* infection (Guerri-Guttenberg, et al., 2008). In the United States, six autochthonous cases, five transfusion related cases and five transplant related cases have been reported, but migratory movements still remain the main source of Chagas' disease. It has been estimated that around 89,221 to 693,302 infected Latin Americans migrated to the United States in the period 1981 to 2005 (Milei, et al., 2009).

Two phases of the disease can be distinguished: (1) acute phase, with transiently high concentration of parasites in tissue and blood, nonspecific symptoms, and a 5% myocarditis incidence, lasting 4 – 8 weeks; and (2) chronic phase, lasting lifelong. Chronic phase can be presented as indeterminate form, characterized by lack of symptoms and normal ECG and normal radiographic examination of the chest, esophagus and colon. Approximately 60 – 70% of patients remain in this form for the rest of their lives. Only 20-40% of infected individuals, 10-30 years after the original acute infection, will develop cardiac, digestive or mixed form of the disease, characterized by the appearance of megavicera (dilated cardiomyopathy, megaesophagus and/or megacolon). It poses a substantial public health burden due to high morbidity and mortality (Milei, et al., 2009, Rassi, et al., 2000, Rassi, et al., 2010).

CCC is manifested by a chronic, diffuse, progressive fibrosing myocarditis that involves not only the working myocardium but also the atrioventricular (AV) conduction system, autonomic nervous system and microcirculation (Andrade, 1985, Marin-Neto, et al., 2007, Milei, et al., 1991b). This leads to cardiomegaly, cardiac failure, arrhythmias, thromboembolism, and death (Milei, et al., 1991b). Colon and esophagus are also commonly affected by Chagas' disease, being megacolon with constipation and megaesofagus with achalasia also features of the disease (Rassi, et al., 2010).

2. Pathogenesis of Chagas' myocarditis

Milei et al. proposed a combined theory that could explain the pathogenic mechanism in chronic chagasic myocarditis (Milei, et al., 1996a, Storino & Milei, 1994). This theory is based on three ingredients: the parasite, host immune system and fibrosis. These ingredients are proposed as being the primary causative agents of damage on myocardial tissue, conduction system, autonomic ganglia and nerves and microvasculature.

2.1 First ingredient: the parasite

The role of *T. cruzi* in the chronic phase has been previously underestimated due to the fact that its presence was believed to be scarce and unrelated to the inflammatory infiltrate present at this stage. Nowadays, the involvement of the parasite in the chronic phase has been well documented. Using dissimilar methods, different authors demonstrated either the persistence of *T. cruzi* or parasite antigens in mice (Younées-Chennoufi, et al., 1988), the parasite DNA sequence amplified by the polymerase chain reaction (PCR) (Jones, et al., 1993, Schijman, et al., 2004), *T. cruzi* antigens from inflammatory lesions in human chagasic cardiomyopathy (Higuchi, Brito et al. 1993), or the immunohistochemical finding of the parasite in endomyocardial biopsies with PCR confirmation (Añez, Carrasco et al. 1999). This would suggest a direct role for the parasite in the perpetuation of myocardial inflammation. In other words, the antigen stimulation would persist throughout the chronic stage, even though the parasites are not morphologically detectable by light microscopy (Andrade 1992).

The role of parasitemia is more controversial. High parasitemia correlated with severity of disease in one report (Basquiera, et al., 2003), but showed no association in another (Castro C., et al., 2005).

Interestingly, it has been observed that immunosuppression reactivates rather than ameliorates the disease, as seen in patients receiving immunosuppressive therapy to prevent transplant rejection and in AIDS patients. Accordingly, many experimental models where strains of genetically manipulated mice lacking various immune functions showed increased susceptibility to develop the disease (Tarleton & Zhang, 1999).

2.1.1 Life cycle of *Trypanosoma cruzi*

When a reduviid bug feeds from an infected mammal, it takes up circulating trypomastigotes, which reach then the bug's gut. There, they differentiate to amastigotes, which proliferate and start to differentiate into epimastigotes. In this process, when amastigote is still sphere-shaped but has developed its flagellum, some authors call this stage spheromastigotes. Then, it elongates its cell body and flagellum, taking the classical epimastigote shape. At this stage, the parasite undergoes metacyclogenesis, differentiating in metacyclic trypomastigotes, the infective form for mammals. When the bug feeds again,

120

it excretes trypomastigotes with feces, which in turn reach blood torrent through bug's wound. Trypomastigotes can infect a wide variety of host cells, within them it differentiate into amastigotes and proliferate. Then, they can differentiate into trypomastigotes again, reach circulation and infect new cells. If an uninfected bug feeds from the animal in the moment of parasitemia, cycle starts again (Tyler & Engman, 2001).



Fig. 1. Life cycle of Trypanosoma cruzi.

2.1.2 Genetic variability of Trypanosoma cruzi and its relation to its pathogenesis

The genetics of *T. cruzi* caught the attention of researchers in late 80' and early 90'. First studies on variability were performed analyzing electrophoretic variants on cellular enzymes. The groups resulting were called zymodemes and were named Z1, Z2, Z3. Only Z2 was associated with domestic transmission cycle.

The development of PCR based techniques, allowed the study of new variant regions and the characterization of multiple variants of a great number of genes. All these variants showed significant correlation with each other, suggesting the existence of two subtypes of *T. cruzi* based on these data (Macedo, et al., 2004). Moreover, *T. cruzi* II which is clearly linked to human pathology, being *T. cruzi* I mainly related to infection of wild sylvatic mammals. Even, applying LSSP-PCR to the study of the variable region of kinetoplast minicircle from *T. cruzi* provided evidence of a differential tissue distribution of genetically diverse *T. cruzi* populations in chronic Chagas' disease, suggesting that the genetic variability of the parasite is one of the determining factors of the clinical form of the disease (Vago, et al., 2000).

2.1.3 Cell host invasion and intracellular survival by Trypanosoma cruzi

Once *T. cruzi* reaches blood torrent, it invades a great variety of cells in the host. When parasiting non phagocytic cells, *T. cruzi* uses some surface glycoproteins to attach to cell: gp82, gp30 and gp35/50. All three glycoproteins are known to induce calcium mobilization from intracellular reservoirs. Gp82 is linked to the phospholipase C (PLC) and inositol 1,4,5 – triphosphate (IP3). Gp 35/50 is associated to increasing intracellular levels of cyclic AMP. On the other side, cruzipain, a protein known to be secreted by *T. cruzi*, acts on kininogen and produces bradykinin, which binds to its receptor, further increasing intracellular calcium. Increased intracellular calcium produces modifications in cytoskeleton that lead to parasite endocytosis (Yoshida & Cortez, 2008).

In the parasitoforous vacuole, mainly by the action of gp85/TS a glycoprotein with transsialidase action, and TcTox, a protease, the parasite degrades the membrane of the vacuole, escapes from it and proliferates within the cell (Alves & Colli, 2007).

2.1.4 Molecular mimicry

The induction of autoimmunity by similarities between *T. cruzi* and host epitopes has been long proposed as a mechanism that leads to tissue damage in the chronic phase of the disease. Both humoral and cellular autoimmune responses have been described, but we will discuss them in more detail in the section of immune system. The real importance of molecular mimicry in the pathogenesis of chagasic myocarditis is still a matter of debate (Girones, et al., 2005).

Although it seems that in some cases this mechanism triggers autoimmunity, in many others, autoimmunity seems to be an epiphenomenon of cellular destruction, with exposition of intracellular epitopes not normally exposed to the immune system. This, in turn may activate autoreactive lymphocytes leading to the appearance of autoantibodies that are not the cause of damage, rather a consequence (Girones, et al., 2005).

The most important cross reacting epitopes of *T. cruzi* and the correspondent epitopes in humans are listed in table 1, as well as the kind of immune response they elicit.

2.2 Second ingredient: host immune system

When the three ingredients theory was first proposed (Storino and Milei 1994, Milei, et al. 1996), second ingredients were mainly T lymphocytes and macrophages. In the subsequent years some evidence grew about the participation of humoral immune system through autoantibodies in the pathogenesis. As a consequence, the whole immune system of the host is now considered as the second ingredient.

As described earlier, mononuclear cells persist in the chronic stage of the disease, contributing to the inflammation through its products of secretion or through its own cytotoxicity (suppressor T cells) and cytolytic action (macrophages) (Storino & Milei, 1994).

As previously stated, molecular mimicry may be the main explanation of autoimmunity, triggering both cellular and humoral autoreactivity (Girones, et al., 2005). Figure 2 summarizes the most important immune events in CCC pathogenesis.

2.2.1 Innate immunity

In recent years innate immunity came to the attention of researchers of Chagas' disease pathogenesis. The role of NK cells has been particularly studied in early and late indeterminate forms of the disease and in CCC patients. In early indeterminate patients,

122

Pathogenesis and Pathology of Chagas' Chronic Myocarditis

Parasite antigen	Human Antigen	Immune reaction
B13	Cardiac myosin heavy chain	Autoantibodies Autoreactive T cells
R13 (ribosomal protein)	Ribosomal protein β1-adrenergic receptor M2-muscarinic receptor 38-kDa heart antigen	Autoantibodies
Ribosomal protein PO	β_1 -adrenergic receptor	Autoantibodies
FL-160	47-kDA neuron protein	Autoantibodies
Shed acute-phase antigen (SAPA)	Cha antigen	Autoreactive T cells
TENU2845/36 kDa	Cha antigen	Autoantibodies
Calcireticulin	Calcireticulin	Autoantibodies Autoreactive T cells
Galactosyl-cerebrosides	Galactosyl-cerebrosides	Autoantibodies
Unknown	Neurons, liver, kidney, testis	Autoantibodies
Sulphated glycolipids	Neurons	Autoantibodies
150-kDa protein	Smooth and striated muscle	Autoantibodies
Cruzipain	Cardiac myosin heavy chain M ₂ -muscarinic receptor	Autoantibodies
Microsomal fraction	Heart and skeletal muscle	Autoantibodies
Cytoskeleton	95-kDa myosin tail	Autoantibodies
SRA	Skeletal muscle Ca ²⁺ dependent SRA	Autoantibodies
MAP	MAP (brain)	Autoantibodies
Soluble extract	Myelin basic protein	Autoantibodies Autoreactive T cells
55-kDa membrane protein	28-kDa Lymphocyte membrane protein	Autoantibodies

Table 1. Examples of cross-reacting epitopes (Girones, et al., 2005, Marin-Neto, et al., 2007).

compared to non infected people, increased values of pre-natural killer (NK)-cells (CD3-CD16⁺ CD56⁻), and higher values of proinflammatory monocytes (CD14⁺ CD16⁺ HLA-DR⁺⁺) were found. The higher values of activated B lymphocytes (CD19⁺ CD23⁺) contrasted with impaired T cell activation, indicated by lower values of CD4⁺ CD38⁺ and CD4⁺ HLA-DR⁺ lymphocytes, a lower frequency of CD8⁺ CD38⁺ and CD8⁺ HLA-DR⁺ cells; a decreased frequency of CD4⁺ CD25^{HIGH} regulatory T cells was also observed. All these data suggest a rather proinflammatory profile (Vitelli-Avelar, et al., 2006). This profile may be useful to limit parasitemia and confine infection to tissues. In fact, it has been demonstrated that NK cells are important in defense against the spread of parasitic infection (Brener & Gazzinelli, 1997), and are an important source of INF- γ , a key cytokine to activate macrophages and help with parasite clearance (Camargo, et al., 1997).

In late indeterminate form, CD3-CD16⁺CD56⁺ and CD3-CD16⁺CD56^{DIM} NK cells are increased but are in normal range in CCC patients, suggesting a protective role for them (Vitelli-Avelar, et al., 2005). NK cells showing CD56^{DIM} may play a role in the down

modulation of cytotoxic deleterious T CD8⁺ response reported in CCC patients (Sathler-Avelar, et al., 2009).

Monocytes display different cytokine profile. In indeterminate patients they produce more IL-10 (Gomes, et al., 2003) while in CCC patients they produce more TNF- α (Vitelli-Avelar, et al., 2008), leading to a proinflammatory profile that could be responsible for chronic myocarditis.

Toll-like receptors (TLR) are also implied in the response to acute infection with *T. cruzi*. TLR-2 has been shown to recognize GPI surface molecules from the parasite. In vitro and in vivo studies have demonstrated that macrophages stimulated with GPIs through TLR-2/CD14 receptors produce NO, TNF- α and IL-12 (Campos & Gazzinelli, 2004).

2.2.2 Cellular adaptative immunity

The role of immune cells in the pathogenesis of Chagas' heart disease has been the dominant hypothesis for many years. The paucity of parasite cells in the inflamed myocardium and the presence throughout the evolution of the disease of macrophages and lymphocytes in patched infiltrates lead to this hypothesis. As early as in 1929, Magariños Torres, observing those infiltrates postulated an "allergic" mechanism for CCC. Further, Mazza and Jörg followed this thought and supported the "allergic" theory (Storino & Milei, 1994).

The study of circulating lymphocytes in peripheral blood of chagasic patients showed an increase in the percentages and actual numbers of double-positive cells of the phenotype CD3+/HLA-DR+, as well as decrease in the percentage of CD45RA+/CD4+ and CD45RA+/CD8+ T cells, indicating greater numbers of activated T cells circulating. Consistent parallel increases were seen also in the B lymphocyte subset which stained double-positive for CD19/CD5 (Dutra W. O., et al., 1994). These results were similar for both indeterminate and CCC patients. Moreover, activated T cells lacking the co-stimulatory molecule CD28 are increased in chagasic patients (Menezes, et al., 2004) and express high levels of HLA-DR molecules (Dutra, et al., 2000). Some interesting differences were demonstrated between indeterminate and CCC patients. CD28- T cells in indeterminate patients showed expression of CTLA-4, which recognizes the same ligands as CD28, but instead of inducing cell activation it causes down modulation of T cells. On the contrary, T cells in CCC patients do not up-regulate CTLA-4 (Souza P. E. A., et al., 2007). It is particularly interesting that CD8+CD28- cells are increased in CCC patients compared to indeterminate patients, and that these cells do not require co-stimulation to exert their cytotoxic functions. More strikingly, CD4+CD28- cells behave differently in indeterminate and CCC patients. In the formers, they are closely related to IL-10 levels, while in CCC patients they correlate with INF- γ levels (Menezes, et al., 2004).

Another interesting difference has been found in cellular response between indeterminate and CCC patients. CD4+ cells from CCC patients had an increased expression of V β 5+-TCR, not found in indeterminate patients. When CCC patients mononuclear cells from peripheral blood were cultured in the presence of trypomastigotes antigens, a selective expansion of CD4+ V β 5+ cells was obtained; while when cultured in the presence of epimastigotes antigens, an expansion of CD8+ V β 5+ cells was also noted. These findings could not be repeated in indeterminate patients. Trypomastigote stimulation led to the expansion of CD4+ V β 17+ in indeterminate patients, not seen in CCC patients. This suggests that CCC patients and indeterminate patients respond to different antigen repertoires (Costa, et al., 2000).

124

Monocytes from indeterminate patients, when infected *in vitro* with *T. cruzi*, express low levels of HLA-DR and high levels of CD80, a ligand for CTLA-4 (Souza P. E., et al., 2004). The interaction of these monocytes with CTLA-4⁺ T cells leads to the expression of IL-10, a cytokine known to down-modulate inflammatory responses (Gomes, et al., 2003). This is not observed in CCC patients. CD28⁻ T cells, not expressing CTLA-4, express TNF- α and INF- γ (Menezes, et al., 2004).

In the same direction, CD4-CD8- $\gamma\delta$ T cells are found to be increased in indeterminate patients compared with CCC ones. These cells are also linked to the production of IL-10 and a down modulator effect on inflammation (Villani, et al., 2010).

Cells infiltrating myocardium have also been studied. As demonstrated with immunostaining of endomyocardial biopsies by our group, leukocytes infiltrating myocardium in Chagas' disease were approximately 50% macrophages, and 50% lymphocytes, mainly T lymphocytes (Milei, et al., 1992b). Further immunohistochemical characterization of these cells with CD45R for lymphocytes, CD20 and lambda and kappa light chains for B lymphocytes, CD45R0 for T lymphocytes and CD68 for macrophages, confirmed these findings (Milei, et al., 1996a).

Autoreactive T cells have caught the attention of many investigators. In experimental models, CD4⁺ T cells from infected mice showed a proliferative response to the exposition to human cardiac myosin heavy chain and to *T. cruzi* B13 protein. They also arrested the beating of fetal heart cells and, more importantly, induced myocarditis in immunized mice and promoted rejection of transplanted normal hearts in the absence of *T. cruzi* (Ribeiro-Dos-Santos, et al., 2001). Also, it has been described that T cells infiltrating the myocardium of chagasic patients cross react with human cardiac myosin heavy chain and to *T. cruzi* B13 protein and express high levels of INF- γ and low levels of IL-4, switching to a Th1 profile (Cunha-Neto Edecio & Kalilf, 2001).

In recent years, Treg cells have come to attention in relation to Chagas' disease pathogenesis. These cells are characterized by the expression of CD4, CD25 and FOXP3 (Ziegler & Buckner, 2009). Treg cells are increased in indeterminate patients compared to CCC, which correlates negatively with levels of activated CD8⁺ (Vitelli-Avelar, et al., 2005). A second subset of T CD4⁺ cells, recently described, the Th17 cells, resulted important in Chagas' disease pathogenesis. These cells, mainly linked to autoimmune pathology, are characterized by the expression of CD4⁺, ROR γ t, and secrete IL-17 (Di Jin, et al., 2008). They were increased in a murine model of acute myocarditis induced by *T. cruzi* infection, as well as by immunization with heat-killed *T. cruzi* antigens (Bonney, et al., 2011). Both cell subsets seem to be related, as they require TGF- β to differentiate. In the presence of proinflammatory cytokines, differentiation to Th17 cell prevails and a pro-autoimmune profile develops (Ziegler & Buckner, 2009).

An additional mechanism is the bystander activation. This is the activation of autoreactive lymphocytes by antigen presenting cells in a proinflammatory environment (Fujinami, et al., 2006). This kind of autoreactive T cells activation has been described in Chagas' disease (Fedoseyeva, et al., 1999).

2.2.3 Humoral adaptative immunity

The importance of humoral immunity in controlling *T. cruzi* acute infection has been clearly established. Mice lacking B lymphocytes rapidly succumb to infection (Kumar & Tarleton, 1998). But the fact that attracted most attention is the production of autoantibodies.



Fig. 2A. The immune pathogenesis of Chagas disease in indeterminate patients. The presence on numerous down regulating mechanisms shift the response towards an anti-inflammatory profile.



Fig. 2B. The immune pathogenesis of Chagas disease in CCC patients. Cells evolve towards a proinflammatory profile, with development of autoimmunity.

The first autoantibody to be described was one that reacted to endocardium, blood vessels and interstitium of skeletal muscle (EVI) (Cossio, et al., 1974), but was the same group of investigators who recognized the heterophil nature of the antibody and realized that had no pathogenic role (Khoury, et al., 1983).

Another autoantibody, studied by our group, was anti-laminin antibody (Sanchez, Milei et al. 1993, (Milei, et al., 1993). These antibodies were shown to react against *T. cruzi* amastigotes and trypomastigotes and human laminin (Szarfman, et al., 1982) and deposition of this antibody in marked thickened basement membranes of myocytes, endothelial cells, and vascular smooth muscle cells was shown by us with light microscopy, electron microscopy and immunohistochemical techniques in endomyocardial biopsies of chagasic patients (Sanchez, et al., 1993) but then we found that only 50% of patients had the antibody on their sera and no correlation with disease severity could be established (Milei, et al., 1993).

Anti-myosin antibodies were postulated by some authors to be generated through molecular mimicry with two *T. cruzi* antigens: B13 protein (Gruber & Zingales, 1993) and cruzipain (Giordanengo Laura, et al., 2000a, Giordanengo Laura, et al., 2000b). Although cruzipain antibodies mainly react to skeletal muscle myosin, they can cause conduction disturbances when transferred to uninfected mice and, when transferred to pregnant animals, they caused conduction disturbances in pups (Giordanengo Laura, et al., 2000b). On the other hand, immunossuppresed mice did not mount any humoral response when immunized with myosin but still develop myocarditis (Neu, et al., 1990). This fact made some authors doubt on the molecular mimicry hypothesis and rather consider antibodies to myosin a consequence of myocyte damage (Kierszenbaum, 2003).

Antibodies that react with muscarinic receptors were intensely studied. In early 1990's IgG from chagasic patients was observed to bind to muscarinic M2 receptors and activate them (Sterin-Borda L, et al., 1991). These anti-muscarinic antibodies were found to increase intracellular cGMP and decrease cAMP (Goin J., et al., 1997) and were positively related to the presence of dysautonomia (Goin J. C., et al., 1994). These antibodies also caused accumulation of inositol phosphate and nitric oxide synthase stimulation, with a negative inotropic effect on myocardium (Sterin-Borda Leonor, et al., 1997). As mentioned before, anti-muscarinic autoantibodies are positively related to the presence of dysautonomia (Goin J. C., et al., 1997). As mentioned before, anti-muscarinic autoantibodies are positively related to the presence of dysautonomia (Goin J. C., et al., 1994), the presence of achalasia in chagasic patients (Goin J. C., et al., 1999), sinus node dysfunction (Altschuller, et al., 2007), but are not related with the degree of myocardial dysfunction (Altschuller, et al., 2007, Talvani Andre, et al., 2006), nor with the presence of brain lesions (Py, et al., 2009). In fact, patients with cardiomyopathy and left ventricular dysfunction but without autonomic dysfunction show low levels of anti-muscarinic antibodies (Sterin-Borda Leonor & Borda, 2000).

Antibodies against β_1 -adrenergic receptors were also deeply studied. Described in early 1980's (Borda E., et al., 1984) these antibodies increased cAMP in mouse atrial fibers, increasing the release of PGE₂ and TXB₂ causing diminished contractility (Gorelik, et al., 1990). Increased cAMP activates PKA and then increases the intracellular calcium concentration. This causes in turn inhibition of the Na⁺/K⁺-ATPase and stimulates Ca²⁺-ATPase activity leading to intracellular depletion of K⁺ and further increase in Ca²⁺. These alteration alter contractility and electric impulse generation and conduction (Borda E. S. & Sterin-Borda, 1996). Antiadrenergic autoantibodies titers could not be related to the severity of left ventricular dysfunction (Talvani Andre, et al., 2006) and patients with overt cardiomyopathy but without autonomic dysfunction show low levels of these antibodies (Sterin-Borda Leonor & Borda, 2000). Antibodies against β_2 -adrenergic receptors have also been described but are mainly related to megacolon (Wallukat, et al., 2010).

Autoantibody	Hypothetic pathogenic role	Reference
Anti-Cerebroside	Probably related to neurologial symptoms	(Avila & Rojas, 1990)
Anti-Gal	Apparently protective	(Gazzinelli, 1991)
Anti-Brain Microtubules	Unknown	(Kerner, et al., 1991)
Anti-Ribosome	Unknown	(Levitus, et al., 1991, Skeiky, et al., 1992)
Anti- UsnRNPs	Unkwnown	(Bach-Elias, et al., 1998)
Anti-Sulfatides	May cause myocarditis and induce arrhythmias	(Garcia, et al., 1998)
Anti-Galectin-1	Increased in CCC patients	(Giordanengo L., et al., 2001)
Anti-Cha R3	Specific of CCC	(Girones, et al., 2001a)
Anti-Desmoglein-1	Related to Penphigus foliaceum	(Diaz, et al., 2004)
Anticardiolipin	Unknown	(Pereira De Godoy, et al., 2005)
Anti- TrkA, TrkB and TrkC	Prevents apoptosis of neurons and helps cellular invasion	(Lu, et al., 2010)
Anti-MBP	Related to gastrointestinal form	(Oliveira E. C., et al., 2009)

Table 2. Less studied autoantibodies in Chagas' disease.

Antibodies against AV node and sinus auricular node tissues have been studied as markers of CCC. When compared in chronic chagasic cardiopathy patients, non-chagasic cardiopathy patients, indeterminate chagasic subjects and healthy blood donors as controls, they more frequently found in chronic chagasic cardiopathy, but not enough to be good markers for chagasic cardiopathy group. Besides, no clear association with complex rhythm or conduction alterations was found (Arce-Fonseca, et al., 2005).

Many other autoantibodies have been described (table 2) but are not so widely studied and their role in pathogenesis of chagasic myocarditis is not clear.

2.2.4 Genetic factors

Human leukocyte antigens (HLA) have shown some relation to the development of CCC. HLA-B40 and Cw3 combination was protective for CCC (Llop, et al., 1991), as resulted DRB1*14, DQB1*0303 (Fernandez-Mestre, et al., 1998), HLA-DQB1*06 (Deghaide, et al., 1998) and HLA-A68 (Cruz-Robles, et al., 2004). On the other hand, HLA-C*03 (Layrisse, et al., 2000), DRB1*1503 (Garcia Borras, et al., 2009), DRB1*01, DRB1*08, DQB1*0501 (Fernandez-Mestre, et al., 1998) and HLA-DR16 alelles (Cruz-Robles, et al., 2004) were positively related to the development of CCC.

A number of other genes related to immune system have been studied in order to determine their relation to a predisposition to develop CCC. In table 3 we list those positively related to the appearance of CCC (Cunha-Neto E., et al., 2009).

Gene	Polymorphism
CCL2/MCPI	- 2518
CCR5	+ 53029
TNF-α	- 308
LTA	+ 80, + 252
BAT-1	- 22, - 348
NFkBIL-1	- 62, - 262
IL-1B	- 31, + 3954, + 5810
IL-10	- 1082
IL-12B	+ 1188
MAL/TRIAP	S180L

Table 3. Genetic polymorphisms related to CCC. Adapted from (Cunha-Neto E., et al., 2009).

2.2.5 The cytokines and chemokines

Although proinflammatory cytokines seem to be necessary for controlling parasitemia during acute phase of the disease (Cunha-Neto E., et al., 2009), CCC patients display a rather proinflammatory cytokine while indeterminate patients display a down modulator one. CCC patients had higher levels of TNF- α and CCL2 than indeterminate patients (Ferreira, et al., 2003, Talvani A., et al., 2004). Infiltrating macrophages from CCC patients expressed INF- γ , TNF- α and IL-6 but showed low levels of IL-2, IL-4 and IL-10 (Abel, et al., 2001, Reis D. D., et al., 1993, Reis M. M., et al., 1997). Also CCR5, CXCR3 and CCR7 and their ligands were increased in hearts of CCC patients, as well as monocytes expressing CXCR3, CCR5, CXCL9 and CCL5 (Cunha-Neto E., et al., 2009). It has been shown that INF- γ and CCL2 induceed myocytes to secrete arial natriuretic factor and caused hypertrophy (Cunha-Neto E., et al., 2005), and IL-18 and CCR7 ligands, which are increased in CCC, caused cardiomyocyte hypertrophy and fibrosis (Reddy, et al., 2008, Riol-Blanco, et al., 2005, Sakai, et al., 2006).

2.3 The third ingredient: fibrosis

Fibrosis is one of the most striking characteristics of CCC. In our patients with CCC in endomyocardial biopsies, fibrosis had replaced between 8.2 and 49% of contractile myocardium, with only one patient having less than 10% (Milei, et al., 1992b). While in autopsies, fibrosis was more extensive reaching in the conduction system than in the contracting myocardium ($51.5 \pm 18\%$ vs $43.4 \pm 8\%$, p < 0.05) (Milei, et al., 1996a).

The deposition of laminin in extracellular and basement membranes has been implicated in the pathogenesis of inflammatory process, as laminin is able to bind proinflammatory citokines (Savino, et al., 2007). The inflammatory infiltrate in CCC was related to the production of citokines such as INF- γ , TNF- α , IL-18, CCL2 and CCL21, that may have modulator actions on fibrotic process (Cunha-Neto E., et al., 2009).

3. A combined theory that could explain the pathogenic mechanism in chronic chagasic myocarditis

With the perpetuation of inflammation, necrosis and scarring fibrosis, damage to all histological components of myocardium occurs. Damage to contracting myocardial fibers

130

determines contractile failure as well as electrophysiological disturbances. Conduction system, nervous autonomic system and microvasculature are also damaged and as a consequence they cause further damage to contractile myocardium and produce electrical instability. Figure 3 illustrates with a flow chart the interactive network of different elements in the pathogenesis of CCC.



Fig. 3. Schematic representation of the integrated theory of multiple factors that determine myocardial damage in CCC.

4. Pathophysiological consequences of organ damage

4.1 Dysautonomia

As early as 1922 Carlos Chagas noted that the chronotropic response to atropine was altered in chagasic patients (Chagas C. & Vilella, 1922), but it was not until late 1950's that Köberle published his works showing impressive neuronal depopulation in microscopic sections obtained from the intercaval atrial strip in chagasic patients using a standardized technique of cardiac intramural neuronal counting developed by himself (Köberle, 1956a, 1956b). These findings led to the "neurogenic hypothesis" (Köberle, 1959), which explained all megas in Chagas' disease as a consequence of neuronal depletion.

Although many other authors claimed to have confirmed this finding (Mott & Hagstrom, 1965, Oliveira J. S., 1985), other authors called to attention about the criteria used to diagnose neuronal depletion because of the great variability in the number of neurons in autonomic ganglia (Rossi L., et al., 1994) and they also remarked that the only right criterion to establish neuronal depletion is the presence of proliferation of satellite cells, with the formation of Terplan's nodules, a characteristic lesion described as proliferating satellite cells which replace degenerating neurons, forming nodular structures. These lesions, once considered patognomonic, can be found in other cardiomyopathies (Rossi L., et al., 1994). The same author could not confirm the loss of neurons or denervation in CCC (Rossi L., 1988). Finally, it was demonstrated that, using Terplan's nodules as diagnostic criterion,

CCC patients with heart failure had more neuronal depletion than patients with dilated cardiomyopathy of other causes (Oliveira J. S., 1985). In our experience the neuroganglionic involvement was variable in autopsies of chagasic hearts (Milei, et al., 1991b).

According to pioneer neurogenic hypothesis (Köberle, 1959), early and irreversible damage to the parasympathetic system during acute phase of the disease causes a cathecolaminergic cardiomyopathy, but this point of view has been debated and evidence is contradictory. Functional test performed in CCC patients demonstrated impaired parasympathetic heart rate regulation (metaraminol, phenylephrine and atropine intravenous injections, facial immersion, Valsalva maneuver, head-up and head-down tilt tests, respiratory sinus arrhythmia, handgrip, graded dynamic exercise, and spectral analysis of Holter recordings) (Amorim, et al., 1968, Amorim, et al., 1973, Gallo, et al., 1975, Guzzetti, et al., 1991, Junqueira Junior, et al., 1985, Manço, et al., 1969, Marin-Neto, et al., 1975, Sousa, et al., 1987). However, a careful analysis of these data showed that many patients had normal autonomic function and most patients had heart failure, that could explain autonomic dysfunction *per se* (Davila, et al., 1998).

On the other hand, the study of indeterminate patients has shown conflicting results. While some authors could demonstrate impaired autonomic function (Molina, et al., 2006, Vasconcelos & Junqueira, 2009) others could demonstrate that autonomic function was normal in patients without myocardial damage and that abnormalities in autonomic dysfunction was proportional to heart dysfunction, leading these authors to propose that these abnormalities arise as a compensating mechanism for the progressive left ventricular dilatation (Davila, et al., 1991, Davila Spinetti, et al., 1999). These findings led to a new "neurogenic theory", which considers autonomic dysfunction as secondary to ventricular dilatation and hemodynamic alterations, but once installed, acts synergistically with parasitism and inflammation to cause further myocardial damage (Davila, et al., 2004).

4.2 Microvascular damage

Microcirculation abnormalities in CCC have been firstly pointed out by Jorg as an angiographic anarchy due to capillary loss (Jörg, 1974) and furtherly demonstrated in experimental models as well as in clinical practice (Rossi M. A., et al., 2010).

Many investigators have found abnormal myocardial perfusion using isonitrile-99mtechnetium (Castro R., et al., 1988) and thallium-201 (Hagar & Rahimtoola, 1991, Marin-Neto, et al., 1992) scintigraphy in chagasic patients with normal epicardial coronary arteries. Furthermore, the progression of left ventricular systolic dysfunction is associated with both, the presence of reversible perfusion defects and the increase in perfusion defects at rest (Hiss, et al., 2009, Schwartz & Wexler, 2009). Anatomopathological studies in humans also provided evidence of microvascular damage in CCC. In late 1950's first reports showing collapse of arterioles and intimal proliferation (Torres, 1960) caught the attention of investigators. Also, microthrombi have been described (Rossi M. A., et al., 1984). As said, in endomyocardial biopsies thickening of capillary basement membranes was also found (Milei, et al., 1992b).

Additional evidence of microvascular damage was obtained from experimental models. Vascular constriction, microaneurysms, dilatation and proliferation of microvessels has been demonstrated (Factor & Sonnenblick, 1982, Morris, et al., 1989, Tanowitz H. B., et al., 1996, Tanowitz Herbert B., et al., 1992b).

Many factors have been advocated in the genesis of these lesions. First, the parasite itself. It was shown that *T. cruzi* produces a neuraminidase that removes sialic acid from de surface

of endothelial cells. This results in thrombin binding and platelet aggregation (Libby, et al., 1986). *T. cruzi* also produces tromboxane A_2 (TXA₂), specially during amastigote state (Ashton, et al., 2007), also favouring platelet aggregation and vascular spasm. Direct parasitism of endothelial cells by *T. cruzi* has also been demonstrated, and this causes the activation of the NF- κ B pathway increasing the expression af adhesion molecules (Huang, et al., 1999a), and secreting proinflammatory citokines (Tanowitz Herbert B., et al., 1992a) and iNOS (Huang, et al., 1999b).

Endothelin-1 (ET-1) is another proposed pathogenic element. Elevated levels of mRNA for preproendothelin-1, endothelin converting enzyme and endothelin-1 were observed in the infected myocardium (Petkova Stefka B., et al., 2000), and elevated levels of ET-1 have been found in CCC patients (Salomone, et al., 2001). Mitogen-activated protein kinases and the transcription factor activator-protein-1 regulate the expression of endothelin-1, and both are shown to be increased in myocardium, interstitial cells and vascular and endocardial endothelial cells (Petkova S. B., et al., 2001). Besides, treatment with phosphoramidon, an inhibitor of endothelin converting enzyme, decreases heart size and severity of lesions in an experimental model of Chagas' disease (Jelicks, et al., 2002).

Inflammation also produces dysfunction of endothelial cells. Macrophages secrete TXA₂ and platelet activating factor (PAF) act on endothelium causing vasoconstriction (Rossi M. A. & Carobrez, 1985). Endothelial cells infected *in vitro* with *T. cruzi* lose their antithrombotic properties in response to interleukin 1 β (IL-1 β) (Bevilacqua, et al., 1984, Nachman, et al., 1986).

It is remarkable that, although the data presented, endothelial function seems to be normal in CCC patients without heart failure, as measured by increases in blood flow in response to acetylcholine and sodium nitroprusside (Consolim-Colombo, et al., 2004). Further, in our concept, microvascular damage found in CCC, seems to be secondary to fibrosis and distortion of myocardial fiber arrangement by necrosis and chronic infiltrates, but as once established, may contribute to the perpetuation of myocardial damage.

5. Pathology

Pathological findings are described mostly according to our own findings.

5.1 Macroscopic features

The most striking characteristic of CCC is enlargement of heart with variable degrees of dilatation of chambers (Andrade, 1985) (Figure 4A). In autopsy series, hearts were overweighted (Andrade, 1985, Baroldi, et al., 1997, Bestetti, et al., 1993, Lopes, et al., 1981) compared with indeterminate chagasic patients and non-chagasic subjects. Marked cardiomegalies reached up to 500 grams. Right ventricle (RV) and atrium (RA) were generally more compromised than left chambers, being RV the most dilated in one paper (Laranja, et al., 1956) but RA was in other (Andrade, 1985).

A second remarkable feature is the thinning of the left ventricular apical wall, resulting in apical aneurysm, a very characteristic lesion in CCC (Figure 4B) (Moia, et al., 1955).

Other lesions described are flattening of the papillary muscles and a marked subendocardial sclerosis, parietal and/or aneurismal thrombosis and fibrotic plaques in pericardium (Milei, et al., 1996a, Milei, et al., 1991b, Storino & Milei, 1994).

Myocarditis



Fig. 4. A. High grade heart dilatation. Thining of the apical wall of the left ventricle (white arrow) and cavitary thrombus (black arrow). B. Characteristic apical aneurysm. A from Milei, et al., 1996b, B from Milei, et al., 2008.

5.2 Histological features

Microscopically, myocardial lesions consisted of a chronic inflammatory process with fibrotic scars and extensive mononuclear infiltrates. Such infiltrates were more prominent in the working myocardium and in the specialized cells of the left branch of the His bundle than in the AV node and in the right hisian branch, showing a microfocal disposition (Figure 5A). The percentage of fibrosis was variable and ranged between 8.2 to 49% (Milei, et al., 1996a, Milei, et al., 1992b) (Figure 5B).

Extensive myocytolysis and spotty contraction band necrosis were observed. Cell hypertrophy in the apparently preserved myocytes was revealed by hypertrophic bizarre nuclei. Dilated lymphatic channels widespread in the ventricular septum and in the AV node, His bundle, and in the root of the right and left bundles branches were observed. In the case of apical aneurysm of the left ventricle, dilated lymphatic were distributed subepicardically (Milei, et al., 1996a).

The serial sectioning of the conducting system showed prominent lesions. Sino-atrial node presented mononuclear infiltrates, necrosis of specialized fibers, and intense fibrosis (Milei, et al., 1991b). In the remaining specialized system lesions consisted of mild to moderate diffuse fibrosis of the AV node and of the penetrating and branching portions of the His bundle, complete destruction of the proximal segments of the right and left bundles branch by varying degrees of replacement by dense collagen tissue (Figure 5A). The remaining specialized fibers presented atrophy and mild fatty infiltration and were surrounded in most cases by infiltrates consisting mainly of lymphocytes and macrophages. The subendocardial Purkinje fibers were usually damaged by chronic inflammation and fibrosis (Milei, et al., 1991b) (Figure 5B). These vast fibrosis in the conduction system (Figure 5C) showed severe conduction alterations in electrocardiograms, although curiously in one revision, there were needed sophisticated electrophysiological studies to demonstrate electrical abnormalities in these patients (Andrade, et al., 1988)

134



Fig. 5. A. Extensive mononuclear infiltrates, myocytes loss, and subendocardial fibrosis. Hematoxylin and eosin stain, X25. B. Atrophic myocardial fibres (red) separated by thick bands of fibrous tissue (blue). Mallory trichrome, X 25. C. Bifurcating His bundle showing severe fibrosis at the left branch (between arrows). The right branch (asterisk) is intramyocardial and surrounded by connective tissue. Mallory trichrome, X25. A and C from Milei, 1996a. B from Milei, 2008.

Myocarditis



Fig. 6. A. Detail of the left bundle of His is shown. Immunostaining for T lymphocyte. Positive cells express CD45R0 antigen (brown); specialized myocardial cells have almost disappeared. Extensive mononuclear infiltrate, the majority of them being T lymphocytes. X20. B. Double immunostaining for the simultaneous demonstration of T lymphocytes (CD45RO) and macrophages (CD68). T lymphocytes (brown) in close contact with a macrophage (pinky cytoplasm). X1000. C. Immunostaining to show endothelial cells. Capillaries and small vessels are clearly showed by the expression of CD31. Vessels are midly or moderately disorted because of the surrounding fibrosis. X100. From Milei, 1996a.

In our studies in endomyocardial biopsies, infiltrates were approximately 50% lymphocytes and 50% macrophages. Almost 80% of lymphocytic population were T lymphocytes, being only 20% B lymphocytes. Eosinophils were scarce in infiltrates reaching 5%, and were associated with areas of necrotic myocardium. Mast cells also were scarce or absent in specialized and in contracting myocardium. (Milei, et al., 1996a, Milei, et al., 1992b)

Histological study of aneurisms showed a thinned wall 2-4 mm, with sclerotic plaques of thickened endocardium of up to 92% of total tissue and extensive mononuclear chronic inflammatory infiltrates and widespread fibrosis in myocardium. Myocytes were organized in thin bands or atrophic units surrounded by fibrotic tissue (Figure 5B) (Milei, et al., 1991a).

Autonomic ganglia showed above described Terplan's nodules, with satellite cell proliferation replacing degenerated autonomic neurons. As stated, these lesions, once considered patognomonic, can be found in other cardiomyopathies (Rossi L., et al., 1994).

5.3 Immunohistochemical findings

Immunophenotyping of infiltrates allowed a better characterization of the cells participating in the inflammatory infiltrates, mainly macrophages (CD68⁺) and lymphocytes (CD45R⁺). In our works 26.5% percent of them were T lymphocytes (CD45R⁺, CD45R0⁺) and 10.5% were B lymphocytes (CD20⁺, light chains kappa and/or lambda⁺) (Figure 6A). Thirty percent of the infiltrate was composed of macrophages (CD68⁺). The remaining infiltrate was composed of mononuclear cells resembling macrophages and CLA-negative mononuclear cells. Contacts between CD68 positive cells and T lymphocytes were frequently found

(Figure 6B). CD31 antibodies clearly pointed out normal endothelial cells, in either normal or damaged vessels (Figure 6C) (Milei, et al., 1996a).

5.4 Ultrastructural features

Myocardial fibers showed nuclear enlargement, nuclear membrane invaginations, lipofuscin deposits, myofibrils derangements and loss, swelling, mitochondrial atrophy, dilatation of sarcotubular system, and interstitial fibrosis (Carrasco, et al., 1982, Palacios-Prü, et al., 1982). These findings have been confirmed by our group in endomyocardial biopsies (Ferrans, et al., 1988, Milei, et al., 1992b). Platelet thrombi can be demonstrated within capillaries (Figure 7B).

Other striking alteration in these specimens was the thickening of the basement membranes of cardiac myocytes (Figures 7A, 7C), endothelium Figure 7C) and vascular smooth muscle up to 20 times their normal thickness of 500 Å (Ferrans, et al., 1988). The thickened basement membranes appeared structurally homogeneous, without being multilayered or subdivided into a lamina rara and a lamina densa. They were of relatively low electron density, had a finely fibrillar appearance at high magnification and measured up to 1 μ m in thickness. Using gold-conjugated antibodies, we could demonstrate the presence of laminin in the thickened basal membranes of myocytes and endothelium (Sanchez, et al., 1993).

Regarding the ultrastructure of aneurysms resected from chagasic patients we observed, hypertrophy of myocytes, with swelling, partial or complete loss of myofibrils, swelling of mitochondria, disruption of mitochondrial cristae, lipofuscin granules, and intact sarcolemmas. Basement membranes were thickened, as previously described (Milei, et al., 1991a)



Fig. 7. A. Myocardial fibre with thickened basement membrane. B. Platelet thrombus within a capillary. C. Thickened basement membranes in a myocardial fibre and a capillary. From Milei, et al., 2008.

6. Conclusions

As shown across the sections of this chapter, the numerous hypothesis about pathogenic pathways of CCC have supporting data and pitfalls. Finally all proposals interact with each other, giving us the idea that none of these theories explains the very complex development of CCC by itself. Rather, it seems more feasible that all these hypothesis conform a network of damaging elements, and that all ingredients cause and/or enhances each other. The triggering factor is obviously the interaction between parasite and host's immune system. Cell parasitism, the inflammatory process and consequent necrosis and fibrosis cause damage to contracting myocardium, autonomic system, conduction system and microcirculation. Autonomic damage causes impaired regulation of microvasculature and further alterations in blood flow. Ischemia causes more myocardial damage. Necrosis exposes intracellular epitopes and causes autoantibodies production with more necrosis, fibrosis and so on. It seems that, if adequate down modulator immune mechanisms work properly, this vicious circle stops and patients do not develop cardiomyopathy, rather they remain in the indeterminate form lifelong.

7. References

- Abel L. C., Rizzo L. V., Ianni B., Albuquerque F., Bacal F., Carrara D., Bocchi E. A., Teixeira H. C., Mady C., Kalil J. & Cunha-Neto E. (2001). Chronic Chagas' disease cardiomyopathy patients display an increased IFN-gamma response to Trypanosoma cruzi infection. *Journal of Autoimmunity*. Vol. 17, No. 1, pp. 99-107, ISSN 0896-8411
- Altschuller M. B., Pedrosa R. C., Pereira Bde B., Correa Filho W. B., Medeiros A. S., Costa P. C. & de Carvalho A. C. (2007). Chronic Chagas disease patients with sinus node dysfunction: is the presence of IgG antibodies with muscarinic agonist action independent of left ventricular dysfunction? *Revista da Sociedade Brasileira de Medicina Tropical*. Vol. 40, No. 6, pp. 665 671, ISSN 0037-8682
- Alves M. J. M. & Colli W. (2007). Trypanosoma cruzi: Adhesion to the Host Cell and Intracellular Survival. *IUBMB Life*. Vol. 59, No. 4 - 5, pp. 274 - 279, ISSN 1521-6551
- Amorim D. S., Godoy R. A., Manco J. C., Tanaka A. & Gallo L., Jr. (1968). Effects of acute elevation in blood pressure and of atropine on heart rate in Chagas' disease. A preliminary report. *Circulation*. Vol. 38, No. 2, pp. 289-294, ISSN 0009-7322
- Amorim D. S., Mello de Oliveira J. A., Manco J. C., Gallo L., Jr. & Meira de Oliveira J. S. (1973). Chagas' heart disease. First demonstrable correlation between neuronal degeneration and autonomic impairment. *Acta Cardiologica*. Vol. 28, No. 4, pp. 431-440, ISSN 0001-5385
- Andrade Z. A. (1985). A patologia da doenca de Chagas no honen. *Annales de la Societe Belge de Medecine Tropicale*. Vol. 65, No. 1, pp. 15-30
- Andrade Z. A., Cámara E. J. N., Sadigursky M. & Andrade S. G. (1988). Envolvimento do nodulo sinusal na doença de Chagas. Arquivos Brasileiros de Cardiologia. Vol. 50, No. pp. 153
- Arce-Fonseca M., Ballinas-Verdugo M. A., Reyes P. A., Aranda-Fraustro A. & Monteon V. M. (2005). Autoantibodies to human heart conduction system in Chagas' disease. *Vector Borne and Zoonotic Diseases.* Vol. 5, No. 3, pp. 233-236, ISSN 1530-3667
- Ashton A. W., Mukherjee S., Nagajyothi F., Huang H., Braunstein V. L., Desruisseaux M. S., Factor S. M., Lopez L., Berman J. W., Wittner M., Scherer P. E., Capra V., Coffman

T. M., Serhan C. N., Gotlinger K., Wu K. K., Weiss L. M. & Tanowitz H. B. (2007). Thromboxane A2 is a key regulator of pathogenesis during Trypanosoma cruzi infection. *The Journal of Experimental Medicine*. Vol. 204, No. 4, pp. 929-940

- Avila J. L. & Rojas M. (1990). Elevated cerebroside antibody levels in human visceral and cutaneous leishmaniasis, Trypanosoma rangeli infection, and chronic Chagas' disease. *The American Journal of Tropical Medicine and Hygiene*. Vol. 43, No. 1, pp. 52-60, ISSN 0002-9637
- Bach-Elias M., Bahia D., Teixeira D. C. & Cicarelli R. M. (1998). Presence of autoantibodies against small nuclear ribonucleoprotein epitopes in Chagas' patients' sera. *Parasitology Research*. Vol. 84, No. 10, pp. 796-799, ISSN 0932-0113
- Baroldi G., Oliveira S. J. M. & Silver M. D. (1997). Sudden and unexpected death in clinically silent' Chagas' disease. A hypothesis. *International Journal of Cardiology*. Vol. 58, No. 3, pp. 263-268, ISSN 0167-5273
- Basquiera A. L., Sembaj A., Aguerri A. M., Omelianiuk M., Guzman S., Moreno Barral J., Caeiro T. F., Madoery R. J. & Salomone O. A. (2003). Risk progression to chronic Chagas cardiomyopathy: influence of male sex and of parasitaemia detected by polymerase chain reaction. *Heart.* Vol. 89, No. 10, pp. 1186-1190, ISSN 1468-201X
- Bestetti R. B., Freitas O. C., Muccillo G. & Oliveira J. S. (1993). Clinical and morphological characteristics associated with sudden cardiac death in patients with Chagas' disease. *European Heart Journal*. Vol. 14, No. 12, pp. 1610-1614, ISSN 0195-668X
- Bevilacqua M. P., Pober J. S., Majeau G. R., Cotran R. S. & Gimbrone M. A. (1984). Interleukin 1 (IL-1) induces biosynthesis and cell surface expression of procoagulant activity in human vascular endothelial cells. *The Journal of Experimental Medicine*. Vol. 160, No. 2, pp. 618-623
- Borda E., Pascual J., Cossio P., De La Vega M., Arana R. & Sterin-Borda L. (1984). A circulating IgG in Chagas' disease which binds to beta-adrenoceptors of myocardium and modulates their activity. *Clinical & Experimental Immunology*. Vol. 57, No. 3, pp. 679 - 686
- Borda E. S. & Sterin-Borda L. (1996). Antiadrenergic and mucarinic receptor antibodies in Chagas' cardiomyopathy. *International Journal of Cardiology*. Vol. 54, No. pp. 149-156
- Brener Z. & Gazzinelli R. T. (1997). Immunological control of Trypanosoma cruzi infection and pathogenesis of Chagas' disease. *International Archives of Allergy and Immunology*. Vol. 114, No. 2, pp. 103-110, ISSN 1018-2438
- Camargo M. M., Andrade A. C., Almeida I. C., Travassos L. R. & Gazzinelli R. T. (1997). Glycoconjugates isolated from Trypanosoma cruzi but not from Leishmania species membranes trigger nitric oxide synthesis as well as microbicidal activity in IFNgamma-primed macrophages. *The Journal of Immunology*. Vol. 159, No. 12, pp. 6131-6139
- Campos M. & Gazzinelli R. (2004). Trypanosoma cruzi and its components as exogenous mediators of inflammation recognized through Toll-like receptors. *Mediators of Inflammation*. Vol. 13, No. 3, pp. 139-143, ISSN 0962-9351
- Carrasco G. H. A., Palacios-Prü E., Scorza C., Rangel A., Inglessis G., Sanoja C. L., Molina C. & Fuenmayor A. (1982). La biopsia endomiocárdica: ¿un recurso diagnóstico? Experiencia clínica en pacientes chagásicos y con otras miocardiopatías. *Revista Latinoamericana de Cardiología.* Vol. 3, No. 105, pp. 105

- Castro C., Prata A. & Macedo V. (2005). The influence of the parasitemia on the evolution of the chronic Chagas' disease. *Revista da Sociedade Brasileira de Medicina Tropical*. Vol. 38, No. 1, pp. 1-6, ISSN 0037-8682
- Castro R., Kuschnir E. & Sgammini H. (1988). Evaluación de la performance cardíaca y perfusión miocárdica con radiotrazadores en la cardiopatía chagásica crónica. *Revista de la Federación Argentina de Cardiología*. Vol. 17, No. pp. 226 231
- Chagas C. (1909). Nova tripanozomiaze humana. Estudos sobre a morfolojia e o ciclo evolutivo do Schizotripannum cruzi n. gen., n. sp., ajente etiolojico de nova entidade mobida do homem. *Memorias do Instituto Oswaldo Cruz*. Vol. 1, No. 2, pp. 159 - 218
- Chagas C. & Vilella E. (1922). Cardiac form of American trypanosomiasis. . *Memorias do Instituto Oswaldo Cruz*. Vol. 14, No. 1, pp. 5 - 61
- Consolim-Colombo F. M., Lopes H. F., Rosetto E. A., Rubira M. C., Barreto-Filho J. A. S., Baruzzi A. C. A., Rocha N. N., Mady C., Irigoyen M. C. & Krieger E. M. (2004). Endothelial Function Is Preserved in Chagas' Heart Disease Patients Without Heart Failure. *Endothelium*. Vol. 11, No. 5-6, pp. 241-246
- Cossio P. M., Diez C., Szarfman A., Kreutzer E., Candiolo B. & Arana R. M. (1974). Chagasic Cardiopathy: Demonstration of a Serum Gamma Globulin Factor Which Reacts with Endocardium and Vascular Structures. *Circulation*. Vol. 49, No. 1, pp. 13-21
- Cruz-Robles D., Reyes P. A., Monteon-Padilla V. M., Ortiz-Muniz A. R. & Vargas-Alarcon G. (2004). MHC class I and class II genes in Mexican patients with Chagas disease. *Human Immunology*. Vol. 65, No. 1, pp. 60-65, ISSN 0198-8859
- Cunha-Neto E., Dzau V. J., Allen P. D., Stamatiou D., Benvenutti L., Higuchi M. L., Koyama N. S., Silva J. S., Kalil J. & Liew C. C. (2005). Cardiac gene expression profiling provides evidence for cytokinopathy as a molecular mechanism in Chagas' disease cardiomyopathy. *The American Journal of Pathology*. Vol. 167, No. 2, pp. 305-313, ISSN 0002-9440
- Cunha-Neto E. & Kalilf J. (2001). Heart-infiltrating and Peripheral T Cells in the Pathogenesis of Human Chagas' Disease Cardiomyopathy. *Autoimmunity*. Vol. 34, No. 3, pp. 187-192
- Cunha-Neto E., Nogueira L. G., Teixeira P. C., Ramasawmy R., Drigo S. A., Goldberg A. C., Fonseca S. G., Bilate A. M. & Kalil J. (2009). Immunological and non-immunological effects of cytokines and chemokines in the pathogenesis of chronic Chagas disease cardiomyopathy. *Memorias do Instituto Oswaldo Cruz*. Vol. 104 Suppl 1, No. pp. 252-258, ISSN 1678-8060
- Davila D. F., Donis J. H., Torres A. & Ferrer J. A. (2004). A modified and unifying neurogenic hypothesis can explain the natural history of chronic Chagas heart disease. *International Journal of Cardiology*. Vol. 96, No. 2, pp. 191-195, ISSN 0167-5273
- Davila D. F., Donis J. H., Torres A., Gottberg C. F. & Rossell O. (1991). Cardiac parasympathetic innervation in Chagas' heart disease. *Medical Hypotheses*. Vol. 35, No. 2, pp. 80-84, ISSN 0306-9877
- Davila D. F., Inglessis G. & Mazzei de Davila C. A. (1998). Chagas' heart disease and the autonomic nervous system. *International Journal of Cardiology*. Vol. 66, No. 2, pp. 123-127, ISSN 0167-5273

Pathogenesis and Pathology of Chagas' Chronic Myocarditis

- Davila Spinetti D. F., Inglessis G. & Mazzei de Davila C. A. (1999). Chagas cardiomyopathy and the autonomic nervous system. Clinical studies. *Archivos del Instituto de Cardiologia de Mexico*. Vol. 69, No. 1, pp. 35-39, ISSN 0020-3785
- Deghaide N. H., Dantas R. O. & Donadi E. A. (1998). HLA class I and II profiles of patients presenting with Chagas' disease. *Digestive Diseases and Sciences*. Vol. 43, No. 2, pp. 246-252, ISSN 0163-2116
- Diaz L. A., Arteaga L. A., Hilario-Vargas J., Valenzuela J. G., Li N., Warren S., Aoki V., Hans-Filho G., Eaton D., dos Santos V., Nutman T. B., de Mayolo A. A., Qaqish B. F., Sampaio S. A. & Rivitti E. A. (2004). Anti-desmoglein-1 antibodies in onchocerciasis, leishmaniasis and Chagas disease suggest a possible etiological link to Fogo selvagem. *The Journal of Investigative Dermatology*. Vol. 123, No. 6, pp. 1045-1051, ISSN 0022-202X
- Dutra, Colley, Pinto D., Gazzinelli, Brener, Pereira, Coffman, Correa O. & Carvalho P. (2000). Self and Nonself Stimulatory Molecules Induce Preferential Expansion of CD5+ B Cells or Activated T Cells of Chagasic Patients, Respectively. *Scandinavian Journal of Immunology*. Vol. 51, No. 1, pp. 91-97, ISSN 1365-3083
- Dutra W. O., Martins-Filho O. A., Cancado J. R., Pinto-Dias J. C., Brener Z., Freeman Junior G. L., Colley D. G., Gazzinelli G. & Parra J. C. (1994). Activated T and B lymphocytes in peripheral blood of patients with Chagas' disease. *International Immunology*. Vol. 6, No. 4, pp. 499-506, ISSN 0953-8178
- Factor S. M. & Sonnenblick E. H. (1982). Hypothesis: Is congestive cardiomyopathy caused by a hyperreactive myocardial microcirculation (Microvascular spasm)? *The American Journal of Cardiology.* Vol. 50, No. 5, pp. 1149-1152, ISSN 0002-9149
- Fedoseyeva E. V., Zhang F., Orr P. L., Levin D., Buncke H. J. & Benichou G. (1999). De Novo Autoimmunity to Cardiac Myosin After Heart Transplantation and Its Contribution to the Rejection Process. *The Journal of Immunology*. Vol. 162, No. 11, pp. 6836-6842
- Fernandez-Mestre M. T., Layrisse Z., Montagnani S., Acquatella H., Catalioti F., Matos M., Balbas O., Makhatadze N., Dominguez E., Herrera F. & Madrigal A. (1998). Influence of the HLA class II polymorphism in chronic Chagas' disease. *Parasite immunology*. Vol. 20, No. 4, pp. 197-203, 0141-9838
- Ferrans V. J., Milei J., Tomita Y. & Storino R. A. (1988). Basement membrane thickening in cardiac myocytes and capillaries in chronic Chagas' Disease. American Journal of Cardiology. Vol. 61, No. pp. 1137-1140
- Ferreira R. C., Ianni B. M., Abel L. C., Buck P., Mady C., Kalil J. & Cunha-Neto E. (2003). Increased plasma levels of tumor necrosis factor-alpha in asymptomatic/"indeterminate" and Chagas disease cardiomyopathy patients. *Memorias do Instituto Oswaldo Cruz.* Vol. 98, No. 3, pp. 407-411, ISSN 0074-0276
- Fujinami R. S., von Herrath M. G., Christen U. & Whitton J. L. (2006). Molecular Mimicry, Bystander Activation, or Viral Persistence: Infections and Autoimmune Disease. *Clinical Microbiological Reviews*. Vol. 19, No. 1, pp. 80-94
- Gallo L., Jr., Neto J. A., Manco J. C., Rassi A. & Amorim D. S. (1975). Abnormal heart rate responses during exercise in patients with Chagas' disease. *Cardiology*. Vol. 60, No. 3, pp. 147-162, ISSN 0008-6312
- Garcia Borras S., Racca L., Cotorruelo C., Biondi C., Beloscar J. & Racca A. (2009). Distribution of HLA-DRB1 alleles in Argentinean patients with Chagas' disease

cardiomyopathy. Immunological Investigations. Vol. 38, No. 3-4, pp. 268-275, ISSN 1532-4311

- Garcia R., Avila J. L., Rojas M., Martinez A., Garcia W. & Bergel P. (1998). Anti-sulfatide antibody titers in patients with chronic Chagas disease and other forms of cardiopathy. *Revista Panamericana de Salud Publica*. Vol. 3, No. 4, pp. 249-256, ISSN 1020-4989
- Gazzinelli R. T. (1991). Natural anti-Gal antibodies prevent, rather than cause, autoimmunity in human Chagas' disease. *Research in Immunology*. Vol. 142, No. 2, pp. 164-167, ISSN 0923-2494
- Giordanengo L., Fretes R., Díaz H., Cano R., Bacile A., Vottero-Cima E. & Gea S. (2000a). Cruzipain induces autoimmune response against skeletal muscle and tissue damage in mice. *Muscle & Nerve*. Vol. 23, No. 9, pp. 1407-1413, ISSN 1097-4598
- Giordanengo L., Gea S., Barbieri G. & Rabinovich G. A. (2001). Anti-galectin-1 autoantibodies in human Trypanosoma cruzi infection: differential expression of this beta-galactoside-binding protein in cardiac Chagas' disease. *Clinical and Experimental Immunology*. Vol. 124, No. 2, pp. 266-273, ISSN 0009-9104
- Giordanengo L., Maldonado C., Rivarola H. W., Iosa D., Girones N., Fresno M. & Gea S. (2000b). Induction of antibodies reactive to cardiac myosin and development of heart alterations in cruzipain-immunized mice and their offspring. *European Journal* of Immunology. Vol. 30, No. 11, pp. 3181-3189, ISSN 1521-4141
- Girones N., Cuervo H. & Fresno M. (2005). Trypanosoma cruzi-induced molecular mimicry and Chagas' disease. *Current Topics in Microbiology and Immunology*. Vol. 296, No. pp. 89-123, ISSN 0070-217X
- Girones N., Rodriguez C. I., Basso B., Bellon J. M., Resino S., Munoz-Fernandez M. A., Gea S., Moretti E. & Fresno M. (2001a). Antibodies to an epitope from the Cha human autoantigen are markers of Chagas' disease. *Clinical and Diagnostic Laboratory Immunology*. Vol. 8, No. 6, pp. 1039-1043, ISSN 1071-412X
- Girones N., Rodriguez C. I., Carrasco-Marin E., Hernaez R. F., de Rego J. L. & Fresno M. (2001b). Dominant T- and B-cell epitopes in an autoantigen linked to Chagas' disease. *The Journal of Clinical Investigation*. Vol. 107, No. 8, pp. 985-993, ISSN 0021-9738
- Goin J., Leiros C., Borda E. & Sterin-Borda L. (1997). Interaction of human chagasic IgG with the second extracellular loop of the human heart muscarinic acetylcholine receptor: functional and pathological implications. *The FASEB Journal*. Vol. 11, No. 1, pp. 77-83
- Goin J. C., Borda E., Leiros C. P., Storino R. & Sterin-Borda L. (1994). Identification of antibodies with muscarinic cholinergic activity in human Chagas' disease: pathological implications. *Journal of the Autonomic Nervous System.* Vol. 47, No. 1-2, pp. 45-52, ISSN 0165-1838
- Goin J. C., Sterin-Borda L., Bilder C. R., Varrica L. M., Iantorno G., Ríos M. C. & Borda E. (1999). Functional implications of circulating muscarinic cholinergic receptor autoantibodies in chagasic patients with achalasia. *Gastroenterology*. Vol. 117, No. 4, pp. 798-805, ISSN 0016-5085
- Gomes J. A., Bahia-Oliveira L. M., Rocha M. O., Martins-Filho O. A., Gazzinelli G. & Correa-Oliveira R. (2003). Evidence that development of severe cardiomyopathy in human

Pathogenesis and Pathology of Chagas' Chronic Myocarditis

Chagas' disease is due to a Th1-specific immune response. *Infection and Immunity*. Vol. 71, No. 3, pp. 1185-1193, ISSN 0019-9567

- Gorelik G., Genaro A. M., Sterin-Borda L., Gonzalez Cappa S. & Borda E. S. (1990). Antibodies bind and activate beta adrenergic and cholinergic lymphocyte receptors in Chagas' disease. *Clinical Immunology and Immunopathology*. Vol. 55, No. 2, pp. 221-236, ISSN 0090-1229
- Gruber A. & Zingales B. (1993). Trypanosoma cruzi: characterization of two recombinant antigens with potential application in the diagnosis of Chagas' disease. *Experimental Parasitology*. Vol. 76, No. 1, pp. 1-12, ISSN 0014-4894
- Guerri-Guttenberg R. A., Grana D. R., Ambrosio G. & Milei J. (2008). Chagas cardiomyopathy: Europe is not spared![†]. *European Heart Journal*. Vol. 29, No. 21, pp. 2587-2591
- Guzzetti S., Iosa D., Pecis M., Bonura L., Prosdocimi M. & Malliani A. (1991). Impaired heart rate variability in patients with chronic Chagas' disease. *American Heart Journal*. Vol. 121, No. 6 Pt 1, pp. 1727-1734, ISSN 0002-8703
- Hagar J. M. & Rahimtoola S. H. (1991). Chagas' Heart Disease in the United States. *New England Journal of Medicine*. Vol. 325, No. 11, pp. 763-768
- Hiss F. C., Lascala T. F., Maciel B. C., Marin-Neto J. A. & Simoes M. V. (2009). Changes in myocardial perfusion correlate with deterioration of left ventricular systolic function in chronic Chagas' cardiomyopathy. *JACC. Cardiovascular Imaging.* Vol. 2, No. 2, pp. 164-172, ISSN 1876-7591
- Huang H., Calderon T. M., Berman J. W., Braunstein V. L., Weiss L. M., Wittner M. & Tanowitz H. B. (1999a). Infection of Endothelial Cells with Trypanosoma cruzi Activates NF-kappa B and Induces Vascular Adhesion Molecule Expression. *Infection and Immunity*. Vol. 67, No. 10, pp. 5434-5440
- Huang H., Chan J., Wittner M., Jelicks L. A., Morris S. A., Factor S. M., Weiss L. M., Braunstein V. L., Bacchi C. J., Yarlett N., Chandra M., Shirani J. & Tanowitz H. B. (1999b). Expression of Cardiac Cytokines and Inducible Form of Nitric Oxide Synthase (NOS2) inTrypanosoma cruzi-infected Mice. *Journal of Molecular and Cellular Cardiology*. Vol. 31, No. 1, pp. 75-88, ISSN 0022-2828
- Jelicks L. A., Chandra M., Shirani J., Shtutin V., Tang B., Christ G. J., Factor S. M., Wittner M., Huang H., Weiss L. M., Mukherjee S., Bouzahzah B., Petkova S. B., Teixeira M. M., Douglas S. A., Loredo M. L., D'Orleans-Juste P. & Tanowitz H. B. (2002). Cardioprotective effects of phosphoramidon on myocardial structure and function in murine Chagas' disease. *International Journal for Parasitology*. Vol. 32, No. 12, pp. 1497-1506, ISSN 0020-7519
- Jones E. M., Colley D. G., Tostes S., Reis Lopez E., Vnencak-Jones C. L. & McCurley T. L. (1993). Amplification of Trypanosoma cruzi DNA sequence from inflammatory lesions in human chagasic cardiomyopathy. *American Journal of Tropical Medicine* and Hygiene. Vol. 48, pp. 348-357
- Jorg M. E. (1974). Tripanosomiasis cruzi: anarquía angiotopográfica por descapilarización mesenquimorreactiva, cofactor patogénico de la miocardiopatía crónica. *Prensa Médica Argentina*. Vol. 61, pp. 94.
- Junqueira Junior L. F., Gallo Junior L., Manco J. C., Marin-Neto J. A. & Amorim D. S. (1985). Subtle cardiac autonomic impairment in Chagas' disease detected by baroreflex

sensitivity testing. *Brazilian Journal of Medical and Biologival Research*. Vol. 18, No. 2, pp. 171-178, ISSN 0100-879X

- Kerner N., Liegeard P., Levin M. J. & Hontebeyrie-Joskowicz M. (1991). Trypanosoma cruzi: antibodies to a MAP-like protein in chronic Chagas' disease cross-react with mammalian cytoskeleton. *Experimental Parasitology*. Vol. 73, No. 4, pp. 451-459, ISSN 0014-4894
- Khoury E. L., Diez C., Cossio P. M. & Arana R. M. (1983). Heterophil nature of EVI antibody in Trypanosoma cruzi infection. *Clinical Immunology and Immunopathology*. Vol. 27, No. 2, pp. 283-288, ISSN 0090-1229
- Kierszenbaum F. (2003). Views on the autoimmunity hypothesis for Chagas disease pathogenesis. FEMS Immunology and Medical Microbiology. Vol. 37, No. 1, pp. 1-11, ISSN 0928-8244
- Köberle F. (1956a). Chagas disease: a disease of the peripheral autonomic nervous system. *Wien Klin Woschenschr.* Vol. 68, No. 17, pp. 333 339,
- Köberle F. (1956b). Pathological findings in muscular hollow organs in experimental Chagas disease. *Zentralbl Allg Pathol.* Vol. 95, No. 7 8, pp. 321 329,
- Köberle F. (1959). Cardiopathia parasympathicopriva. *Münch Med Wochenschr*. Vol. 101, No. pp. 1308-1310, ISSN 0027-2973
- Kumar S. & Tarleton R. L. (1998). The relative contribution of antibody production and CD8+ T cell function to immune control of Trypanosoma cruzi. *Parasite Immunology*. Vol. 20, No. 5, pp. 207-216, ISSN 1365-3024
- Laranja F. S., Dias E., Nobrega G. & Miranda A. (1956). Chagas' Disease: A Clinical, Epidemiologic, and Pathologic Study. *Circulation*. Vol. 14, No. 6, pp. 1035-1060
- Layrisse Z., Fernandez M. T., Montagnani S., Matos M., Balbas O., Herrera F., Colorado I. A., Catalioti F. & Acquatella H. (2000). HLA-C(*)03 is a risk factor for cardiomyopathy in Chagas disease. *Human Immunology*. Vol. 61, No. 9, pp. 925-929, ISSN 0198-8859
- Levitus G., Hontebeyrie-Joskowicz M., Van Regenmortel M. H. & Levin M. J. (1991). Humoral autoimmune response to ribosomal P proteins in chronic Chagas heart disease. *Clinical and Experimental Immunology*. Vol. 85, No. 3, pp. 413-417, ISSN 0009-9104
- Libby P., Alroy J. & Pereira M. E. (1986). A neuraminidase from Trypanosoma cruzi removes sialic acid from the surface of mammalian myocardial and endothelial cells. *The Journal of Clinical Investigation*. Vol. 77, No. 1, pp. 127-135, ISSN 0021-9738
- Llop E., Rothhammer F., Acuna M., Apt W. & Arribada A. (1991). HLA antigens in Chagas cardiomyopathy: new evidence based on a case-control study. *Revista Médica de Chile*. Vol. 119, No. 6, pp. 633-636, ISSN 0034-9887
- Lopes E. R., Chapadeiro E., Andrade Z. A., Almeida Hde O. & Rocha A. (1981). Pathological anatomy of hearts from asymptomatic Chagas disease patients dying in a violent manner. *Memorias do Instituto Oswaldo Cruz*. Vol. 75, No. 2, pp. 189 - 197
- Lu B., Luquetti A. O., Rassi A. & PereiraPerrin M. (2010). Autoantibodies to neurotrophic receptors TrkA, TrkB and TrkC in patients with acute Chagas' disease. *Scandinavian Journal of Immunology*. Vol. 71, No. 3, pp. 220-225, ISSN 1365-3083
- Macedo A. M., Machado C. R., Oliveira R. P. & Pena S. D. J. (2004). Trypanosoma cruzi: Genetic Structure of Populations and Relevance of Genetic Variability to the Pathogenesis of Chagas Disease. *Memorias do Instituto Oswaldo Cruz.* Vol. 99, No. 1, pp. 1 - 12

- Manço J. C., Gallo L., Jr., Godoy R. A., Fernandes R. G. & Amorim D. S. (1969). Degeneration of the cardiac nerves in Chagas' disease. Further studies. *Circulation*. Vol. 40, No. 6, pp. 879-885, ISSN 0009-7322
- Marin-Neto J. A., Cunha-Neto E., Maciel B. C. & Simoes M. V. (2007). Pathogenesis of Chronic Chagas Heart Disease. *Circulation*. Vol. 115, No. 9, pp. 1109-1123
- Marin-Neto J. A., Gallo L., Jr., Manço J. C., Rassi A. & Amorim D. S. (1975). Postural reflexes in chronic Chagas's heart disease. Heart rate and arterial pressure responses. *Cardiology*. Vol. 60, No. 6, pp. 343-357, 0008-6312
- Marin-Neto J. A., Marzullo P., Marcassa C., Gallo Junior L., Maciel B. C., Bellina C. R. & L'Abbate A. (1992). Myocardial perfusion abnormalities in chronic Chagas' disease as detected by thallium-201 scintigraphy. *The American Journal of Cardiology*. Vol. 69, No. 8, pp. 780-784, ISSN 0002-9149
- Menezes C. A. S., Rocha M. O. C., Souza P. E. A., Chaves A. C. L., Gollob K. J. & Dutra W. O. (2004). Phenotypic and functional characteristics of CD28+ and CD28- cells from chagasic patients: distinct repertoire and cytokine expression. *Clinical & Experimental Immunology*. Vol. 137, No. 1, pp. 129-138, ISSN 1365-2249
- Milei J., Fernandez Alonso G., Vanzulli S., Storino R. A., Maturri L. & Rossi L. (1996a). Myocardial Inflammatory infiltrate in human chronic Chagasic cardiomyopathy: Immunohistochemical findings. *Cardiovascular Pathology*. Vol. 5, No. 4, pp. 209-219
- Milei J., Storino R.A., Matturri L., Rossi L. (1996b). Studio anatomo-clinico ed epidemiologico della malattia di Chagas. *Pathologica*. Vol. 88, pp. 117, 127
- Milei J., Guerri-Guttenberg R. A., Grana D. R. & Storino R. (2009). Prognostic impact of Chagas disease in the United States. *American Heart Journal*. Vol. 157, No. 1, pp. 22-29, ISSN 0002-8703
- Milei J., Mautner B., Storino R. A., Sanchez J. A. & Ferrans V. J. (1992a). Does Chagas' disease exist as an undiagnosd form of cardiomyopathy in the United States? *American Heart Journal*. Vol. 123, No. 6, pp. 1732-1735
- Milei J., Pesce R., Valero E., Muratore C., Beigelman R. & Ferrans V. J. (1991a). Electrophysiologic-structural correlations in chagasic aneurysms causing malignant arrhythmias. *International Journal of Cardiology*. Vol. 32, No. 1, pp. 65-73, ISSN 0167-5273
- Milei J., Sánchez J., Storino R., Yu Z.-X., Denduchis B. & Ferrans V. J. (1993). Antibodies to laminin and immunohistochemical localization of laminin in chronic chagasic cardiomyopathy: a review. *Molecular and Cellular Biochemistry*. Vol. 129, No. 2, pp. 161-170, ISSN 0300-8177
- Milei J., Storino R. A., Beigelman R., Fernandez Alonso G., Maturri L. & Rossi L. (1991b). Histopathology of specialized and ordinary myocardium and nerves in chronic Chagas disease, with a morphometric study of inflammation and fibrosis. *Cardiologia*. Vol. 36, No. 2, pp. 107-115
- Milei J., Storino R. A., Fernandez Alonso G., Beigelman R., Vanzulli S. & Ferrans V. J. (1992b). Endomyocardial biopsies in chronic chagasic cardiomyopathy. Immunohistochemical and ultrastructural findings. *Cardiology*. Vol. 80, No. 5-6, pp. 424-437
- Milei J., Guerri-Guttenberg R. A., Azzato F. & Storino R. (2008). Chagasic Cardiomyopathy: New trends for an old burden, Nova Science Publishers, 978-1-60692-193-7

- Moia B., Rosenbaum M. & Hojman D. (1955). Aneurismas ventriculares en la miocarditis crónica chagásica. *Revista Argentina de Cardiología*. Vol. 22, No. pp.
- Molina R. B. G., Matsubara B. B., Hueb J. C., Zanati S. G., Meira D. A., Cassolato J. L., Paiva S. A. R. & Zornoff L. A. M. (2006). Dysautonomia and ventricular dysfunction in the indeterminate form of Chagas disease. *International Journal of Cardiology*. Vol. 113, No. 2, pp. 188-193, ISSN 0167-5273
- Morris S. A., Weiss L. M., Factor S., Bilezikian J. P., Tanowitz H. & Wittner M. (1989). Verapamil ameliorates clinical, pathologic and biochemical manifestations of experimental chagasic cardiomyopathy in mice. *Journal of the American College of Cardiology*. Vol. 14, No. 3, pp. 782-789, ISSN 0735-1097
- Mott K. E. & Hagstrom J. W. C. (1965). The Pathologic Lesions of the Cardiac Autonomic Nervous System in Chronic Chagas' Myocarditis. *Circulation*. Vol. 31, No. 2, pp. 273-286
- Nachman R. L., Hajjar K. A., Silverstein R. L. & Dinarello C. A. (1986). Interleukin 1 induces endothelial cell synthesis of plasminogen activator inhibitor. *The Journal of Experimental Medicine*. Vol. 163, No. 6, pp. 1595-1600
- Neu N., Ploier B. & Ofner C. (1990). Cardiac myosin-induced myocarditis. Heart autoantibodies are not involved in the induction of the disease. *The Journal of Immunology*. Vol. 145, No. 12, pp. 4094-4100
- Oliveira E. C., Fujisawa M. M., Hallal Longo D. E., Farias A. S., Contin Moraes J., Guariento M. E., de Almeida E. A., Saad M. J., Langone F., Toyama M. H., Andreollo N. A. & Santos L. M. (2009). Neuropathy of gastrointestinal Chagas' disease: immune response to myelin antigens. *Neuroimmunomodulation*. Vol. 16, No. 1, pp. 54-62, 1423-0216
- Oliveira J. S. (1985). A natural human model of intrinsic heart nervous system denervation: Chagas' cardiopathy. *American heart journal*. Vol. 110, No. 5, pp. 1092 - 1098
- Palacios-Prü E., Carrasco G. H. A., Scorza C. & Espinosa R. (1982). Ultraestructura miocárdica en la enfermedad de Chagas. Diagnóstico diferencial con la miocardiopatía dilatada o congestiva y con las miocardiopatías arrítmicas. *Revista Latinoamericana de Cardiología*. Vol. 3, No. pp. 115
- Pereira de Godoy M. R., Cacao J. C., Pereira de Godoy J. M., Brandao A. C. & Silva Rossi Souza D. (2005). Chagas disease and anticardiolipin antibodies in older adults. *Archives of Gerontology and Geriatrics*. Vol. 41, No. 3, pp. 235-238, ISSN 0167-4943
- Petkova S. B., Huang H., Factor S. M., Pestell R. G., Bouzahzah B., Jelicks L. A., Weiss L. M., Douglas S. A., Wittner M. & Tanowitz H. B. (2001). The role of endothelin in the pathogenesis of Chagas' disease. *International Journal for Parasitology*. Vol. 31, No. 5-6, pp. 499-511, ISSN 0020-7519
- Petkova S. B., Tanowitz H. B., Magazine H. I., Factor S. M., Chan J., Pestell R. G., Bouzahzah B., Douglas S. A., Shtutin V., Morris S. A., Tsang E., Weiss L. M., Christ G. J., Wittner M. & Huang H. (2000). Myocardial Expression of Endothelin-1 in Murine Trypanosoma cruzi Infection. *Cardiovascular Pathology*. Vol. 9, No. 5, pp. 257-265, ISSN 1054-8807
- Py M. O., Maciel L., Pedrosa R. C., Nascimento J. H. & Medei E. (2009). The presence of antiautonomic membrane receptor antibodies do not correlate with brain lesions in Chagas' disease. *Arquivos Brasileiros de Cardiologia*. Vol. 67, No. 3A, pp. 633 638

- Rassi A., Jr., Rassi A. & Little W. C. (2000). Chagas' heart disease. *Clinical cardiology*. Vol. 23, No. 12, pp. 883-889, ISSN 0160-9289
- Rassi A., Jr., Rassi A. & Marin-Neto J. A. (2010). Chagas disease. *Lancet.* Vol. 375, No. 9723, pp. 1388-1402, 1474-547X
- Reddy V. S., Harskamp R. E., van Ginkel M. W., Calhoon J., Baisden C. E., Kim I. S., Valente A. J. & Chandrasekar B. (2008). Interleukin-18 stimulates fibronectin expression in primary human cardiac fibroblasts via PI3K-Akt-dependent NF-kappaB activation. *Journal of Cell Physiology*. Vol. 215, No. 3, pp. 697-707, ISSN 1097-4652
- Reis D. D., Jones E. M., Tostes S., Jr., Lopes E. R., Gazzinelli G., Colley D. G. & McCurley T. L. (1993). Characterization of inflammatory infiltrates in chronic chagasic myocardial lesions: presence of tumor necrosis factor-alpha+ cells and dominance of granzyme A+, CD8+ lymphocytes. *The American Journal of Tropical Medicine and Hygiene*. Vol. 48, No. 5, pp. 637-644, ISSN 0002-9637
- Reis M. M., Higuchi Mde L., Benvenuti L. A., Aiello V. D., Gutierrez P. S., Bellotti G. & Pileggi F. (1997). An in situ quantitative immunohistochemical study of cytokines and IL-2R+ in chronic human chagasic myocarditis: correlation with the presence of myocardial Trypanosoma cruzi antigens. *Clinical Immunology and Immunopathology*. Vol. 83, No. 2, pp. 165-172, ISSN 0090-1229
- Ribeiro-Dos-Santos R., Mengel J. O., Postol E., Soares R. A. O., Ferreira-Fernandez E., Soares M. B. P. & Pontes-De-Carvalho L. C. (2001). A heart-specific CD4+ T-cell line obtained from a chronic chagasic mouse induces carditis in heart-immunized mice and rejection of normal heart transplants in the absence of Trypanosoma cruzi. *Parasite Immunology*. Vol. 23, No. 2, pp. 93-101, ISSN 1365-3024
- Riol-Blanco L., Sanchez-Sanchez N., Torres A., Tejedor A., Narumiya S., Corbi A. L., Sanchez-Mateos P. & Rodriguez-Fernandez J. L. (2005). The chemokine receptor CCR7 activates in dendritic cells two signaling modules that independently regulate chemotaxis and migratory speed. *Journal of Immunology*. Vol. 174, No. 7, pp. 4070-4080, ISSN 0022-1767
- Rossi L. (1988). Neuroanatomopathology of the cardiovascular system., In: *Neurocardiology.*, Kulbertus, H. E. and Franck, G., pp. Futura Publishing Co. Inc., Mount Kisco, N.Y.
- Rossi L., Storino R., Milei J. & Matturri L. (1994). Depleción neuronal en la enfermedad de Chagas: todo debería revisarse. *Revista Argentina de Cardiología*. Vol. 62, No. 3, pp. 239 - 246
- Rossi M. A. & Carobrez S. G. (1985). Experimental Trypanosoma cruzi cardiomyopathy in BALB/c mice: histochemical evidence of hypoxic changes in the myocardium. *British Journal of Experimental Pathology.* Vol. 66, No. 2, pp. 155 - 160
- Rossi M. A., Gonçalves S. & Ribeiro-dos-Santos R. (1984). Experimental Trypanosoma cruzi Cardiomyopathy in BA4LB/c Mice. The Potential Role of Intravascular Platelet Aggregation in its Genesis. *American Journal of Pathology.* Vol. 114, No. 2, pp. 209 -216
- Rossi M. A., Tanowitz H., Malvestio L. M., Celes M. R., Campos E. C., Blefari V. & Prado C. M. (2010). Coronary Microvascular Disease in Chronic Chagas Cardiomyopathy Including an Overview on History, Pathology, and Other Proposed Pathogenic Mechanisms. *PLoS Neglected Tropical Diseases*. Vol. 4, No. 8, pp. e674
- Sakai N., Wada T., Yokoyama H., Lipp M., Ueha S., Matsushima K. & Kaneko S. (2006). Secondary lymphoid tissue chemokine (SLC/CCL21)/CCR7 signaling regulates

fibrocytes in renal fibrosis. *Proceedings of the National Academy Science U S A*. Vol. 103, No. 38, pp. 14098-14103, ISSN 0027-8424

- Salomone O. A., Caeiro T. F., Madoery R. J., Amuchástegui M., Omelinauk M., Juri D. & Kaski J. C. (2001). High plasma immunoreactive endothelin levels in patients with Chagas' cardiomyopathy. *The American Journal of Cardiology*. Vol. 87, No. 10, pp. 1217-1220, ISSN 0002-9149
- Sanchez J. A., Milei J., Yu Z. X., Storino R., Wenthold R. J. & Ferrans V. J. (1993). Immunohistochemical localization of laminin in the hearts of patients with chronic chagasic cardiomyopathy: relationship to thickening of basement membranes. *American Heart Journal*. Vol. 126, No. 6, pp. 1392 - 1401
- Sathler-Avelar R., Vitelli-Avelar D. M., Teixeira-Carvalho A. & Martins-Filho O. A. (2009). Innate immunity and regulatory T-cells in human Chagas disease: what must be understood? *Memorias do Instituto Oswaldo Cruz.* Vol. 104 Suppl 1, No. pp. 246-251, ISSN 1678-8060
- Savino W., Villa-Verde D. M., Mendes-da-Cruz D. A., Silva-Monteiro E., Perez A. R., Aoki M. d. P., Bottasso O., Guiñazú N., Silva-Barbosa S. D. & Gea S. (2007). Cytokines and cell adhesion receptors in the regulation of Immunity to Trypanosoma cruzi. *Cytokine & Growth Factor Reviews*. Vol. 18, No. pp. 107 - 124
- Schijman A. G., Vigliano C. A., Viotti R. J., Burgos J. M., Brandariz S., Lococo B. E., Leze M. I., Armenti H. A. & Levin M. J. (2004). Trypanosoma cruzi DNA in cardiac lesions of Argentinean patients with end-stage chronic chagas heart disease. *The American Journal of Tropical Medicine and Hygiene*. Vol. 70, No. 2, pp. 210-220, ISSN 0002-9637
- Schwartz R. G. & Wexler O. (2009). Early Identification and Monitoring Progression of Chagas' Cardiomyopathy With SPECT Myocardial Perfusion Imaging. JACC: Cardiovascular Imaging. Vol. 2, No. 2, pp. 173-175, ISSN 1936-878X
- Skeiky Y. A., Benson D. R., Parsons M., Elkon K. B. & Reed S. G. (1992). Cloning and expression of Trypanosoma cruzi ribosomal protein P0 and epitope analysis of anti-P0 autoantibodies in Chagas' disease patients. *The Journal of Experimental Medicine*. Vol. 176, No. 1, pp. 201-211, ISSN 0022-1007
- Sousa A. C., Marin-Neto J. A., Maciel B. C., Gallo L., Jr. & Amorim D. S. (1987). Cardiac parasympathetic impairment in gastrointestinal Chagas' disease. *Lancet.* Vol. 1, No. 8539, pp. 985, ISSN 0140-6736
- Souza P. E., Rocha M. O., Rocha-Vieira E., Menezes C. A., Chaves A. C., Gollob K. J. & Dutra W. O. (2004). Monocytes from patients with indeterminate and cardiac forms of Chagas' disease display distinct phenotypic and functional characteristics associated with morbidity. *Infection and immunity*. Vol. 72, No. 9, pp. 5283-5291, ISSN 0019-9567
- Souza P. E. A., Rocha M. O. C., Menezes C. A. S., Coelho J. S., Chaves A. C. L., Gollob K. J. & Dutra W. O. (2007). Trypanosoma cruzi Infection Induces Differential Modulation of Costimulatory Molecules and Cytokines by Monocytes and T Cells from Patients with Indeterminate and Cardiac Chagas' Disease. *Infection and. Immunity*. Vol. 75, No. 4, pp. 1886 - 1894
- Sterin-Borda L. & Borda E. (2000). Role of Neurotransmitter Autoantibodies in the Pathogenesis of Chagasic Peripheral Dysautonomia. Annals of the New York Academy of Sciences. Vol. 917, No. 1, pp. 273-280, ISSN 1749-6632

- Sterin-Borda L., Gorelik G. & Borda E. S. (1991). Chagasic IgG binding with cardiac muscarinic cholinergic receptors modifies cholinergic-mediated cellular transmembrane signals. *Clin Immuol Immunopathol.* Vol. 61, No. pp. 387-397
- Sterin-Borda L., Leiros C. P., Goin J. C., Cremaschi G., Genaro A., Echagüe A. V. & Borda E. (1997). Participation of Nitric Oxide Signaling System in the Cardiac Muscarinic Cholinergic Effect of Human Chagasic IgG. *Journal of Molecular and Cellular Cardiology*. Vol. 29, No. 7, pp. 1851-1865, ISSN 0022-2828
- Storino R. A., Barragan H. & Milei J. (1992). Aspectos epidemiologicos de la enfermedad de Chagas en la Argentina y America Latina. *Revista Federación Argentina de Cardiología.* Vol. 21, No. 3, pp. 239-246
- Storino R. A. & Milei J. (1994). Enfermedad de Chagas, Mosby-Doyma, Buenos Aires
- Szarfman A., Terranova V. P., Rennard S. I., Foidart J. M., de Fatima Lima M., Scheinman J. I. & Martin G. R. (1982). Antibodies to laminin in Chagas' disease. *The Journal of Experimental Medicine*. Vol. 155, No. 4, pp. ISSN 1161-1171
- Talvani A., Rocha M. O., Barcelos L. S., Gomes Y. M., Ribeiro A. L. & Teixeira M. M. (2004). Elevated concentrations of CCL2 and tumor necrosis factor-alpha in chagasic cardiomyopathy. *Clinical Infectious Disease*. Vol. 38, No. 7, pp. 943-950, ISSN 1537-6591
- Talvani A., Rocha M. O. C., Ribeiro A. L., Borda E., Sterin-Borda L. & Teixeira M. M. (2006). Levels of anti-M2 and anti-[beta]1 autoantibodies do not correlate with the degree of heart dysfunction in Chagas' heart disease. *Microbes and Infection.* Vol. 8, No. 9-10, pp. 2459-2464, 1286-4579
- Tanowitz H. B., Gumprecht J. P., Spurr D., Calderon T. M., Ventura M. C., Raventos-Suarez C., Kellie S., Factor S. M., Hatcher V. B., Wittner M. & Berman J. (1992a). Cytokine Gene Expression of Endothelial Cells Infected with Trypanosoma cruzi. *Journal of Infectious Diseases*. Vol. 166, No. 3, pp. 598-603
- Tanowitz H. B., Kaul D. K., Chen B., Morris S. A., Factor S. M., Weiss L. M. & Wittner M. (1996). Compromised Microcirculation in Acute Murine Trypanosoma cruzi Infection. *The Journal of Parasitology*. Vol. 82, No. 1, pp. 124 - 130
- Tanowitz H. B., Morris S. A., Factor S. M., Weiss L. M. & Wittner M. (1992b). Parasitic diseases of the heart I: Acute and chronic Chagas' disease. *Cardiovascular Pathology*. Vol. 1, No. 1, pp. 7-15, ISSN 1054-8807
- Tarleton R. & Zhang L. (1999). Chagas Disease Etiology: Autoimmunity or Parasite Persistence? *Parasitology Today*. Vol. 15, No. 3, pp. 94 - 99
- Torres C. M. (1960). Miocitólise e fibrose do miocárdio na doença de Chagas. *Memorias do Instituto Oswaldo Cruz.* Vol. 58, No. 2, pp. 161 182
- Tyler K. M. & Engman D. M. (2001). The life cycle of Trypanosoma cruzi revisited. International Journal for Parasitology. Vol. 31, No. pp. 472 - 481
- Vago A. R., Andrade L. O., Leite A. A., d'Avila Reis D., Macedo A. M., Adad S. J., Tostes S., Jr., Moreira M. C., Filho G. B. & Pena S. D. (2000). Genetic characterization of Trypanosoma cruzi directly from tissues of patients with chronic Chagas disease: differential distribution of genetic types into diverse organs. *The American Journal of Pathology*. Vol. 156, No. 5, pp. 1805-1809, ISSN 0002-9440
- Vasconcelos D. F. & Junqueira L. F., Jr. (2009). Distinctive impaired cardiac autonomic modulation of heart rate variability in chronic Chagas' indeterminate and heart diseases. *Journal of Electrocardiology*. Vol. 42, No. 3, pp. 281-289, ISSN 1532-8430

- Villani F. N., Rocha M. O., Nunes Mdo C., Antonelli L. R., Magalhaes L. M., dos Santos J. S., Gollob K. J. & Dutra W. O. (2010). Trypanosoma cruzi-induced activation of functionally distinct alphabeta and gammadelta CD4- CD8- T cells in individuals with polar forms of Chagas' disease. *Infection and Immunity*. Vol. 78, No. 10, pp. 4421-4430, ISSN 1098-5522
- Vitelli-Avelar D. M., Sathler-Avelar R., Dias J. C. P., Pascoal V. P. M., Teixeira-Carvalho A., Lage P. S., Elói-Santos S. M., Corrêa-Oliveira R. & Martins-Filho O. A. (2005). Chagasic Patients with Indeterminate Clinical Form of the Disease have High Frequencies of Circulating CD3+CD16-CD56+ Natural Killer T Cells and CD4+CD25High Regulatory T Lymphocytes. *Scandinavian Journal of Immunology*. Vol. 62, No. 3, pp. 297-308, ISSN 1365-3083
- Vitelli-Avelar D. M., Sathler-Avelar R., Massara R. L., Borges J. D., Lage P. S., Lana M., Teixeira-Carvalho A., Dias J. C., Eloi-Santos S. M. & Martins-Filho O. A. (2006). Are increased frequency of macrophage-like and natural killer (NK) cells, together with high levels of NKT and CD4+CD25high T cells balancing activated CD8+ T cells, the key to control Chagas' disease morbidity? *Clinical and Experimental Immunology*. Vol. 145, No. 1, pp. 81-92, ISSN 0009-9104
- Vitelli-Avelar D. M., Sathler-Avelar R., Teixeira-Carvalho A., Pinto Dias J. C., Gontijo E. D., Faria A. M., Eloi-Santos S. M. & Martins-Filho O. A. (2008). Strategy to assess the overall cytokine profile of circulating leukocytes and its association with distinct clinical forms of human Chagas disease. *Scandinavian Journal of Immunology*. Vol. 68, No. 5, pp. 516-525, ISSN 1365-3083
- Wallukat G., Munoz Saravia S. G., Haberland A., Bartel S., Araujo R., Valda G., Duchen D., Diaz Ramirez I., Borges A. C. & Schimke I. (2010). Distinct patterns of autoantibodies against G-protein-coupled receptors in Chagas' cardiomyopathy and megacolon. Their potential impact for early risk assessment in asymptomatic Chagas' patients. *Journal of the American College of Cardiology*. Vol. 55, No. 5, pp. 463-468, ISSN 1558-3597
- Yoshida N. & Cortez M. R. (2008). *Trypanosoma cruzi: parasite and host cell signaling during the invasion process*, Springer Science, Houton
- Younées-Chennoufi A. B., Hontebeyre-Joskowicz M., Tricottet V., Eisen H., Reynes M. & Said G. (1988). Persistence of Trypanosoma cruzi antigens in the inflammatory lesions of chronically infected mice. *Trans Roy Soc Trop Med Hyg.* Vol. 82, No. pp. 77



Myocarditis Edited by Dr. Daniela Cihakova

ISBN 978-953-307-289-0 Hard cover, 428 pages **Publisher** InTech **Published online** 19, October, 2011 **Published in print edition** October, 2011

Myocarditis, the inflammation of the heart muscle, could be in some cases serious and potentially fatal disease. This book is a comprehensive compilation of studies from leading international experts on various aspects of myocarditis. The first section of the book provides a clinical perspective on the disease. It contains comprehensive reviews of the causes of myocarditis, its classification, diagnosis, and treatment. It also includes reviews of Perimyocarditis; Chagas' chronic myocarditis, and myocarditis in HIV-positive patients. The second section of the book focuses on the pathogenesis of myocarditis, discussing pathways and mechanisms activated during viral infection and host immune response during myocarditis. The third, and final, section discusses new findings in the pathogenesis that may lead to new directions for clinical diagnosis, including use of new biomarkers, and new treatments of myocarditis.

How to reference

In order to correctly reference this scholarly work, feel free to copy and paste the following:

Julián González, Roberto Guerri-Guttenberg, Daniel Grana, Francisco Azzato and José Milei (2011). Pathogenesis and Pathology of Chagas' Chronic Myocarditis, Myocarditis, Dr. Daniela Cihakova (Ed.), ISBN: 978-953-307-289-0, InTech, Available from: http://www.intechopen.com/books/myocarditis/pathogenesis-andpathology-of-chagas-chronic-myocarditis



InTech Europe

University Campus STeP Ri Slavka Krautzeka 83/A 51000 Rijeka, Croatia Phone: +385 (51) 770 447 Fax: +385 (51) 686 166 www.intechopen.com

InTech China

Unit 405, Office Block, Hotel Equatorial Shanghai No.65, Yan An Road (West), Shanghai, 200040, China 中国上海市延安西路65号上海国际贵都大饭店办公楼405单元 Phone: +86-21-62489820 Fax: +86-21-62489821 © 2011 The Author(s). Licensee IntechOpen. This is an open access article distributed under the terms of the <u>Creative Commons Attribution 3.0</u> <u>License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

IntechOpen

IntechOpen