We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Progression of β-Lactam Resistance in *Staphylococcus aureus*

Antresh Kumar and Manisha Kaushal

Abstract

Staphylococcus aureus is a notorious human pathogen that causes superficial and invasive infections both in nosocomial and community-acquired settings. The prevalence of staphylococcal infections became more challenging after emerging resistance against topical antibiotics. S. aureus evolved resistance to β -lactam antibiotics due to modification and expression of penicillin-binding proteins (PBP), inactivation of drug by β -lactamase synthesis, limiting uptake of drug by biofilm formation, and reducing uptake by expression of efflux pump. The wave of resistance was first observed in penicillin by β -lactamase production and PBPs modification. The second wave of resistance emerged to methicillin by appearing methicillin-resistant S. aureus (MRSA) strains. Cephalosporin has long been used as the last resort for preventing MRSA infections, but resistant strains appeared during treatment. In progression to control MRSA or related infections, carbapenems have been used but strains developed resistance. S. aureus is among the high-priority resistance organisms that need renewed efforts for the research and development of new antibiotics and innovative preventive approaches. However, a lot of toiling is involved in devising an effective treatment against drug resistant *S. aureus*. This chapter aim is to retrospectively determine the progression of resistance in S. *aureus*, against different β -lactam antibiotics and their challenges of medication.

Keywords: *Staphylococcus aureus*, Drug resistance, β-lactam antibiotics, Penicillin, Methicillin, Cephalosporin

1. Introduction

Infections caused by a variety of bacterial, fungal, viral, and other infectious microorganisms are considered to be the world's most leading problem. Infectious diseases are considered to be the world most leading cause of death, with almost 50,000 deaths per day [1]. Bacterial and fungal infections are the major cause of morbidity and mortality in both developed and developing countries [2]. *Staphylococcus aureus* is a gram-positive, coagulase-positive opportunistic bacterial pathogen, commonly found in the human nasal mucosa in the approximately 20–40% population [3, 4]. It causes a wide range of infections such as skin infections, including abscesses, impetigo, and necrotizing fasciitis; tissue infections, including osteomyelitis and endocarditis; and toxicities, including toxic shock syndrome, pneumonia, sepsis, and surgical site infections [5–7]. The superficial and invasive infections caused by *S. aureus* continue to raise serious health challenges globally as it notoriously exhibits resistance [8, 9]. These infections have rapidly developed resistance against most of the available antimicrobials, which pose serious threats [10–13]. Infections caused by *S. aureus* are associated with significantly higher mortality, because of the limitations of available antimicrobial therapies, difficulties in making a rapid and accurate diagnosis, and the development of multidrug resistance (MDR) [14]. The acute and chronic staphylococcal infections have now become more problematic after emerging multidrug resistance (MDR) against various frontline antibiotics [15, 16]. Antibiotics are small molecules that selectively inhibit the growth of a plethora of bacterial and other infections. These heterogeneous group molecules continue to be save many lives from different bacterial infections. Antibiotics are either naturally synthesized by microorganisms or chemically modified into exciting drugs. β -lactam antibiotics (β -LA) are considered to be the most successful and frequently used antibiotics against a number of bacterial infections. The underlying reason behind this is their wide spectrum activity, oral availability, excellent pharmacokinetics, lack of toxicity, and bactericidal action [17]. Due to the widespread and prolonged practice of β -LA emerged resistance to these resort and became an alarming and emerging problem to the public health. The microbial pathogens tend to adopt different resistance mechanism to skip the cytotoxic effect of β -LA. The progression in β -LA drug resistance to emerge multiple antibiotic-resistant microorganisms has made it difficult to manage many infectious diseases using common anti-infective drugs. In this chapter, we focus on emerging trends of drug resistance in *S. aureus* to the different β -LA.

2. β-Lactam antibiotics (β-LA)

The landmark discovery the beta-lactam penicillin has been developed with the remarkable weapon to control bacterial infections during the Second World War [18]. It was naturally synthesized from *Penicillium chrysogenum* (also known as *Penicillium notatum*). Penicillin G was the first β -lactam antibiotic (β -LA) discovered in 1944, which began the era of antibiotics against a wide range of infectious microorganisms [19]. The development of penicillin led to search its different derivatives (amoxicillin and methicillin) for the betterment of their efficacy, bioavailability, solubility, stability, and other pharmacokinetic properties and to evade steadily emerging problem of multidrug resistance (MDR). Structurally, penicillin is composed of a thiazolidine ring attached to a side chain of a four-membered betalactam ring. All penicillins are derivatives of 6-aminopenecillinic acid, which sometimes differ in their side-chain structure. Many β -LA have lactam ring as an integral part of a molecule such as cephalosporins, monobactams, cephamycins, and the carbapenems (imipenem and meropenem). These β -LA antibiotics came into the light to rescue mankind from different Gram –ve and Gram +ve bacterial infections including *S. aureus*. β -LA are the most available and over 34 β -LA approved by the FDA, which together constitute ~50% of all antibiotic prescriptions worldwide. Now, β -LA share the annual consumption of over \$15 billion, which contribute almost 65% of the total antibiotics [20].

The β -LA primarily target the cell wall of a bacterial pathogen. Peptidoglycan or murien present in the cell wall provides the mechanical strength to the bacterial cell membrane, which is composed of an alternating unit of *N*-acetylglucosamine (NAG) and *N*-acetylmuramic acid (NAM) residues, joined together by β -1 \rightarrow 4 linkage. The NAM is further linked with a pentapeptide stem, which is composed of L-Ala-D-Glu- L-Lys-D-Ala-D-Ala. The order and type of amino acids are almost similar in Gram –ve and Gram +ve bacterial with some slight variations. The last D-Ala is lost during maturation and glycan assembly is cross-linked to form a bridge

with the carboxyl group of D-Ala at position 4 and the amino group of the amino acid at position 3. Mechanistically, β -LA acts upon a 4-membered "beta-lactam" ring, which shows a resemblance to D-Ala-D-Ala sequence of the cell wall [21]. The primary function of PBP is in the elongation of the cell wall, which is composed of two distinct components termed as PBP1–4. The radioactive analysis revealed that penicillin specifically interacts with PBP protein *via* covalent interactions [22]. The tight binding of β -LA to the transpeptidase domain of PBP (penicillin-binding protein) thereby inhibits the peptidoglycan synthesis by acylating transpeptidase, involved in crosslinking peptide to form peptidoglycan [23].

3. β-Lactam resistance in S. aureus

According to the European Centre for Diseases Control (ECDC), antimicrobial resistance is the single biggest threat facing the world in the area of infectious diseases. With the progression of antibiotics discoveries and their prophylactic usages have emerged drug resistance to single or multiple drugs. Antibiotic resistance is a natural selection process when microorganisms are treated with different antibiotics, and microorganisms tend to escape this selection pressure with greater competency to survive and thus show antibiotics resistance. In contrast, bacteria with a susceptible nature are killed with exposed antibiotics. Emerging resistance to β -LA is a serious health concern that causes a major hurdle in the treatment of bacterial infections. The condition of drug resistance is primarily developed by increasing and indiscriminate usage of antibiotics in clinical ailments, unregulated sales of antibiotics, a long course of medication, and poor public health infrastructure. According to a hospital survey, over 80% of clinical samples of S. aureus were established resistance to the frontline antibiotics including methicillin [24, 25]. It has been reported that 70% of nosocomial bacterial pathogens have emerged resistance to more than one antibiotic during medication of chronic infections. In contrast, an alarming increase in resistance of community-acquired bacteria has also been observed with significant high rate both in acute and chronic bacterial infections. The emergence of drug-resistant strains of Gram-positive (Staphylococcus, Enterococcus, Streptococcus sp) and Gram-negative (Pseudomonas, Klebsiella, Enterobacter, Acinetobacter, Salmonella sp) bacteria is the more serious in the present therapeutic scenario. S. aureus clearly represents one of the most challenging pathogenic bacteria. Resistance in S. aureus strains has been continuously increasing; thus, the ability of these pathogens to spread in both hospital and community settings increased. Bacteria remarkably developed resistant to antimicrobial drugs in several ways. Upon antibiotics treatment, bacteria tend to overcome the selection pressure of the drug by morphological and genetic alterations or drug inactivation. Alterations of membrane integrity and transfer of resistance genes from one strain to another are the common examples of β -LA-mediated resistance in *S. aureus*. β -LA, Penicillin was initially succeeded in the treatment of S. aureus infections but widespread and prolonged uses of penicillin were no longer be effective and resistance has been emerged soon after in the 1950s [19]. Antibiotic resistance can be a typical feature of a bacterial species (intrinsic resistance) or acquired by the individual organism that is naturally susceptible (acquired resistance). The acquired resistance is the consequence of chromosomal mutations or acquisition of resistance genes by horizontal gene transfer [26]. Resistance to multiple β -LA can be acquired by individual strains, resulting in multidrug-resistant phenotypes. The high prevalence of drug resistance is primarily adopted by unregulated sales of antibiotics without prescription, a long course of medication, indiscriminate usage of drugs, and poor health infrastructure. The mobility and mortality caused by drug resistance in public

health are difficult to evaluate. In 2013, Center for Disease Control and Prevention (CDC) reported more than 11,000 deaths in the USA had a methicillin-resistant *S. aureus* (MRSA)-related infection (CDC 2013). This represents almost 50% of all causalities caused by antibiotic-resistant bacteria. As per WHO report, the MRSA remains among the high-priority multidrug-resistant organisms that need renewed efforts for the research and development of new antibiotics and innovative preventive approaches.

4. Mechanism of β -lactam resistance in S. aureus

Different mechanisms of drug resistance in bacterial pathogens are the major hurdle in their treatment. With emerging resistance, it became a serious concern to look into drug resistance mechanism, which can help us to prescribe a specific medication to effectively overcome the problem of resistance.

Several biochemical mechanisms are responsible for β -LA resistance, including enzymatic (β -lactamase) production inactivation of the drug (drug inactivation), modifications of drug target in penicillin-binding protein (PBPs) (target modifications), limiting uptake of drug by biofilm formation (reduced drug uptake), and active efflux of the drug (drug efflux) as shown in Figure 1 [27, 28]. Bacterial pathogens resist the inhibitory action of antibiotics primarily due to the presence of an enzyme that inactivates the antibiotic or modified antibiotic target by mutation or by the post-translational mechanism, which reduces binding of the antibiotic to the target or bypass of the function dependent on the antibiotic target by an alternative enzyme that is not inhibited by the antibiotic. Moreover, overexpression of drug efflux pumps rendered to reduce uptake of the antibiotic inside the cell, by pumping out the antibiotics from the cell. In contrast, encapsulation of biofilm over the cell boundary reduces the cell permeability to resist antibiotics entry into the cell. The expression of chromosomal β -lactamase can be induced by either producing the plasmid-encoded penicillinase (β -lactamase) enzyme that hydrolyzes β -lactam ring or expression of PBP2a, and a penicillin-binding protein (PBP) encoded by gene mecA spread through horizontal gene transfer with low affinity to β -lactam antibiotics is primarily responsible for penicillin resistance [17]. The penicillin-binding cascade induces the blaZ-encoded penicillinase in *S. aureus*, which is transcriptionally regulated by regulatory genes blaI and blaR1 [26, 29].



Figure 1. β -Lactam resistance mechanism of S. aureus.

5. Methicillin resistance in S. aureus

Methicillin was introduced in clinical practice for the effective treatment of penicillin-resistant *S. aureus* infections [30]. After 2 years, the second wave of resistance against methicillin came into the light and the first report on methicillin resistance S. aureus (MRSA) strain was published by MP Jevons in 1961 [31]. Statistically, incidences of methicillin-sensitive S. aureus (MSSA), methicillinresistant S. aureus (MRSA), and vancomycin-resistant S. aureus (VRSA) infections have increased up to 54% in both hospital-acquired (HA) and community-acquired (CA) [32]. These antimicrobial-resistant infections cause a significant economic burden on public health. The economic burden of antibiotic resistance in Europe was estimated at almost 1.5 billion euros. However, USA spent more than 55 billion dollars each year on the treatment of antibiotic-resistant infections [9]. It was found that acquisition of methicillin resistance in *S. aureus* was primarily contributed by the integration of a *mecA* gene encoded for low-affinity penicillin-binding protein 2a or 2' (PBP2a or PBP2') into the staphylococcal chromosomal cassette (SCCmec) element of methicillin-sensitive S. aureus (MRSA) [33]. The expression of mecA in MRSA is induced by the interaction of methicillin and other antibiotics to the regulatory network. MecIR a regulatory protein, homologous to the BlaIR proteins, controls the expression of *mecA*. It is under the control of MecIR regulatory proteins that are homologous to the BlaIR proteins that regulate BlaZ expression [34, 35]. The SSC*mec* is located specifically with an unknown gene (orfX) of the staphylococcal chromosomal. The function of the unknown gene is mediated by two recombinases termed as *ccrA* and *ccrB* that help in the site-specific integration or excision of DNA elements from the staphylococcal chromosomal [36, 37]. The insertion sequence, transposon (Tn554) or erythromycin- and spectinomycin-encoded resistance genes, and tobramycin and kanamycin resistance-encoded pUB110 plasmid can be additionally jumped in the SSC*mec* region. Typing of SSC*mec* elements is fundamental for the molecular epidemiology of MRSA and categorized majorly into five types, that is, type I-V [38]. The SSCmec-type I-III elements are present in hospital-acquired MRSA strains, which are typically resistant to non- β -lactam antibiotics. In contrast, SSCmec-type IV-V are only resistance to methicillin, which are primarily present in community-acquired MRSA (CA-MRSA). Different studies revealed that multiple insertions of SCCmec elements in the staphylococcal chromosome of MSSA strains yield a MRSA lineage. The mecC gene, homolog to mecA gene, exhibits 68.7% nucleotide identity is identified in S. aureus, Staphylococcus sciuri, and Staphylococcus xylosus strains [39]. The recent studies revealed that mecC carrying S. aureus contributes in methicillin resistance in the human population by up to 2.8% of MRSA strains [40-42], while no report was found on mecB-carrying S. aureus resistance to methicillin. In many MRSA strains, the expression of mecA is also affected either by the synthesis of truncated MecIR regulatory proteins or by repression by β -lactamase regulators BlaI and BlaR. The Mec and Bla regulatory proteins can alter the functional behavior and expression of PBP2a-encoded gene in MRSA strains. In a short period, MRSA strains have been identified all around the globe particularly Asia, USA, and Europe [43]. In spite of the rapidly spreading of methicillin resistance, MRSA exhibited broad-spectrum drug resistance against methicillin, penicillins, cephalosporins, and carbapenems. The MRSA cases were increased in hospitals and other healthcare facilities (hospital-acquired), and in communities (community-acquired infections). People with immediate surgeries or stay in healthcare facilities are at MRSA higher risk. Infection also spreads if a medical device has been put in their body or when they come close to contact with MRSA-infected patient. MRSA spreads in communities through uncovered or draining wounds mostly associated with crowded living, sharing personal items,

recent stays in healthcare facilities, etc. In 2017, CDC reported that more than a 0.3 million cases and over 10,000 deaths from MRSA-related infections are estimated in-hospital patients with more than 1.7 billion healthcare burdens in the United States. This figure represents mere a 50% of all the mortalities caused by antibiotic-resistant bacteria. The prevalence of MRSA infections in India has been reported to increase from 29% in 2009 to 47% in 2014 [35].

6. Cephalosporin resistance in S. aureus

Similar to penicillin or other β -lactams, cephalosporins also target to bind penicillin-binding proteins (PBPs) to inhibit peptidoglycan formation in bacteria. These are effectively used in the treatment of superficial (skin and soft tissue) infections, and nosocomial and community-acquired pneumonia. Different strains of *S. aureus* strains have evolved resistance to cephalosporins, which evolved by reducing the binding affinity of cephalosporins to transpeptidase of PBPs, and also, β -lactamases are produced by bacteria having encoded plasmid for inactivation of therapeutics effect of cephalosporins. The plasmid-mediated β -lactamase resistance is corroborated by the amount and activity of the enzyme produced in bacteria.

Recent studies revealed that the prevalence of cephalosporins resistance in *S. aureus* is comparable to the β -lactamase-resistant penicillin, which accounts for 30–35% [44, 45]. Ceftaroline is the fifth-generation antibiotics, approved by the FDA in 2010, which has a broad-spectrum activity against a plethora of bacterial pathogens. Ceftaroline is active against methicillin-resistant *S. aureus* (MRSA) and has been successfully used for the treatment of different invasive bacterial infections with low adverse effects. This potent third-generation drug was found resistance in MRSA-ST293 strain in different geographical regions. Ceftaroline had the higher affinity to PBP but nonsense or missense mutations in the *mecA* gene alter the amino acid sequence of PBP protein, which causes alteration in the ceftaroline binding to PBPs. In addition, alteration of the promoter sequence of PBP4 by mutation increases PBP4 production that leads to resistance to ceftaroline [46].

7. Carbapenem-resistance

The β -lactam antibiotic carbapenems are the last resort, potent, broad-spectrum antibiotic against Gram +ve and Gram –ve bacterial pathogens. They contain a carbapenem structure linked together with a beta-lactam ring, which primarily targets to bind with PBPs of the cell wall. Due to high potency, low adverse effect appeals to prefer the use of carbapenems. Prolonged and widespread uses of the drug have developed carbapenems resistance, which is contributed by a different mechanism. The resistance that arises to carbapenems is due to β -lactamase gene transfer/production, mutational alteration in PBPs, and expression of efflux pump systems [47, 48]. The carbapenem resistance is mainly contributed by β -lactamase production.

8. Future perspective

Emerging resistance in *S. aureus* is a serious human health problem, which continuously increasing mortality and morbidity rates in both nosocomial and acquired infections. The constant evolution of resistance to topical antibiotics

including the continuing appearance of new resistance mechanisms and complexity in multidrug-resistant phenotypes are appealing to find new diagnostic tools and therapeutic strategies to get rid of this problem. However, a lot of toiling has continued to devise a workable treatment against staphylococcal infections particularly for the elimination of MRSA and VRSA pathogens. Emerging MDR in S. aureus has evolved major challenges in research and need to expend research to the next level to understand the progression of drug resistance pathways and infections pattern of *S. aureus*. The new search of therapeutics targets and bioactive molecules and their judicious use may be proven significantly to prevent the problem of drug resistance [2, 49]. Reducing the outer membrane permeability of bacterial cells can circumvent the problem of drug resistance. Iron conjugated with the antibiotic method may help to selectively interact to the outer membrane to active transport of antibiotic inside the cell [50]. Another possible approach has been targeted to inhibit quorum sensing that is primarily related to the virulence factors release and associated with the microbial pathogenesis. Chemically, virulence factors are toxic to the host cells that disrupt immune response, along with host cell disruption and cell adhesion. SarA and agr are two main quorum sensing mechanisms of *S*. aureus, which can be targeted to block the quorum sensing for controlling S. aureus infections. In addition, bacteriophage therapy is one of the potential methods for controlling the drug resistance in S. aureus infection. Phage therapy has many advantages over chemotherapy, for example, very specific, no side effect, environmental friendly, no allergenic effects, and harmless to the eukaryotic host [51]. Phage has been used to eliminate MRSA infections but is still immature in clinical application [51]. The phage-based treatment of resistant S. aureus will further be helpful to select the gene responsible for its control. These strategies will pave a way to develop a vaccine in future against the S. aureus.

9. Conclusion

It is very clear that bacterium including Staphylococcus aureus shows extraordinary adaptability to cope with antibiotic effect and emerge drug resistance against antibiotics. The phenomenon of drug resistance was first observed when β-lactam antibiotics became ineffective after indiscriminative uses and plasmidresponsive β -lactamase (penicillinase) synthesis. The second wave of resistance against methicillin has been primarily contributed by the stable integration of a mecA gene-encoded penicillin-binding protein and penicillin-binding protein 2a or 2' (PBP2a or PBP2') into the staphylococcal chromosomal cassette (SCCmec) element. Cephalosporins have been proven as an effective drug preventing MRSA infections but failed. In progression to antibiotics, carbapenems have been used for preventing S. aureus infections, but multidrug resistance (MDR) strains developed. The common cause of bacterial resistance involves horizontal gene transfer, target alteration by point mutations, and expression of efflux pump, which made a variety of antibiotics ineffective and induces persistent infections in both hospital and community settings. Moreover, the prolonged and widespread use of different antibiotics, lack of awareness, and insanitation, primarily contribute in rapidly developing multiple drug resistance (MDR) in developing countries that causes a major financial burden in the treatment of infectious diseases. Though a lot of toiling is involved in devising an effective treatment against staphylococcal infections particularly for the elimination of MRSA and VRSA, the new search of bioactive molecules and their judicious use may be proven significantly to prevent the problem of drug resistance.

IntechOpen

Author details

Antresh Kumar^{1*} and Manisha Kaushal²

1 Department of Biochemistry, Central University of Haryana, Mahendergarh, Haryana, India

2 District Hospital Compound, Amroha, India

*Address all correspondence to: antreshkumar@cuh.ac.in

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Holmes KK, Bertozzi S, Bloom BR, Prabhat Jha P, Gelband H, DeMaria LM, et al. Major infectious diseases: Key messages from disease control priorities. In: Holmes KK, Bertozzi S, Bloom BR, Jha P, editors. Major Infectious Diseases. 3rd ed. Washington (DC): The International Bank for Reconstruction and Development/The World Bank; 2017

[2] Kumar S, Kumar A, Kaushal M, Kumar P, Mukhopadhyay K, Kumar A. Fungal-derived xenobiotic exhibits antibacterial and antibiofilm activity against *S. aureus*. Drug Discoveries & Therapeutics. 2018;**12**(4):214-223 (IF1.27)

[3] Gorwitz RJ, Kruszon-Moran D, McAllister SK, McQuillan G, McDougal LK, Fosheim GE, et al. Changes in the prevalence of nasal colonization with *Staphylococcus aureus* in the United States, 2001-2004. Journal of Infectious Diseases. 2008;**197**(9): 1226-1234. DOI: 10.1086/533494

[4] Adèle S, Fabienne B, Jean-Louis M, Jean-Marc R, Olivier B. *Staphylococcus aureus* nasal colonization: An update on mechanisms, epidemiology, risk factors, and subsequent infections. Frontier Microbiology. 2018;**9**:2419. DOI: 10.3389/fmicb.2018.02419

[5] Odell CA. Community-associated methicillin-resistant *Staphylococcus aureus* (CA-MRSA) skin infections. Current Opinion in *Pediatrics*. 2010;**22**:273-277

[6] Mohammad H, Thangamani S, Seleem MN. Antimicrobial peptides and peptidomimetics-potent therapeutic allies for staphylococcal infections. Current Pharmaceutical Design. 2015;**21**:2073-2088

[7] Peters PJ, Brooks JT, McAllister SK, Limbago B. Methicillin resistant *Staphylococcus aureus* colonization of the groin and risk for clinical infection among HIV infected adults. Emerging Infectious Diseases. 2013;**19**:623-629

[8] World Health Organization. WHO Antimicrobial Resistance: Global Report on Surveillance. Geneva, Switzerland. Available from: http:// www.who.int/drugresistance/ documents/surveillancereport/en/: World Health Organization; 2014 [Accessed: 01 January 2019]

[9] Antibiotic resistance threats in the United States. Centers for Disease Control and Prevention. Atlanta, GA. Publisher US Department of Health and Human Services. Available from: https://www. cdc.gov/drugresistance/threatreport-2013/pdf/ar-threats-2013-508.pdf; 2013 [Accessed: 01 January 2019]

[10] Sunagar R, Hegde NR, Archana GJ, Sinha AY. Prevalence and genotype distribution of methicillin-resistant *Staphylococcus aureus* (MRSA) in India. Journal of Global Antimicrobial Resistance. 2016;7:46-52

[11] Bouchiat C, El-Zeenni N,
Chakrakodi B, Nagaraj S, Arakere G,
Etienne J. Epidemiology of
Staphylococcus aureus in Bangalore,
India: emergence of the ST217 clone and
high rate of resistance to erythromycin
and ciprofloxacin in the community.
New Microbes and New Infections.
2015;7:15-20

[12] Thati V, Shivannavar CT, Gaddad SM. Vancomycin resistance among methicillin resistant Staphylococcus aureus isolates from intensive care units of tertiary care hospitals in Hyderabad. The Indian Journal of Medical Research. 2011;**134**:704-708

[13] Uematsu H, Yamashita K, Kunisawa S, Fushimi K, Imanaka Y. Estimating the disease burden of methicillin-resistant *Staphylococcus aureus* in Japan: Retrospective database study of Japanese hospitals. PLoS One. 2017;**12**:e0179767

[14] Koziel J, Maciag-Gudowska A, Mikolajczyk T, Bzowska M, Sturdevant DE, Whitney AR, et al. Phagocytosis of *Staphylococcus aureus* by macrophages exerts cytoprotective effects manifested by the upregulation of antiapoptotic factors. PLoS One. 2009;**4**:e5210

[15] Duin DV, Paterson D. Multidrug resistant Bacteria in the community: Trends and lessons learned. Infectious Disease Clinics of North America.
2016;**30**:377-390

[16] David MZ, Daum RS. Communityassociated methicillin-resistant *Staphylococcus aureus*: Epidemiology and clinical consequences of an emerging epidemic. Clinical Microbiology Reviews. 2010;**23**:616-687

[17] Bush K, Bradford PA. β-Lactams and β-lactamase inhibitors: An overview.
Cold Spring Harbor Perspectives in Medicine. 2016;6(8):a025247.
DOI: 10.1101/cshperspect.a025247

[18] Neushal P. Science, government and the mass production of penicillin. Journal of History of Medicine and Allied Sciences. 1993;**48**(4):371-395

[19] Walsh CT, Wencewicz TA. Antibiotics: Challenges, Mechanisms, Opportunities. Washington, DC: ASM Press; 2016

[20] Thakurla B, Lahon K. The beta lactam antibiotics as an empirical therapy in a developing country: An update on their current status and recommendations to counter the resistance against them. Journal of Clinical and Diagnostic Research. 2013;7:1207-1214

[21] Zango UU, Ibrahim M, Shawai SAA, Shamsuddin IM. A review on β -lactam

antibiotic drug resistance. MOJ Drug Design Development & Therapy. 2019;**3**:52-58

[22] Cooper PD, Rowley D. Investigations with radioactive penicillin. Nature. 1949;**163**:480-481

[23] Yocum RR, Rasmussen JR, Strominger JL. The mechanism of action of penicillin: Penicillin acylates the active site of *Bacillus stearothermophilus* d-alanine carboxypeptidase. Journal of Biological Chemistry. 1980;**255**(9): 3977-3986

[24] Turner NA, Sharma-Kuinkel BK, Maskarinec SA, Eichenberger EM, Shah PP, Carugati M, et al. Methicillinresistant *Staphylococcus aureus*: an overview of basic and clinical research. Nature Reviews Microbiology. 2019;**17**:203-218

[25] Wanda CR. An overview of the antimicrobial resistance mechanisms of bacteria. AIMS Microbiology.2018;4(3):482-501. DOI: 10.3934/ microbiol.2018.3.482

[26] Dever LA, Dermody TS. Mechanisms of bacterial resistance to antibiotics. Archives of Internal Medicine. 1991;**151**(5):886-895

[27] Ibrahim ME, Abbas M, Al-Shahrai AM, Elamin BK. Phenotypic characterization and antibiotic resistance patterns of extendedspectrum β -lactamase- and AmpC β -lactamase-producing gram-negative bacteria in a referral hospital, Saudi Arabia. Canadian Journal of Infectious Diseases and Medical Microbiology. 2019;**2019**:6054694. DOI: 10.1155/ 2019/6054694

[28] Pfeifer Y, Cullik A, Witte W.
Resistance to cephalosporins and carbapenems in Gram-negative bacterial pathogens. International Journal of Medical Microbiology. 2010;**300**: 371-379

[29] Guo Y, Song G, Sun M, Wang J, Wang Y. Prevalence and therapies of antibiotic-resistance in *Staphylococcus aureus*. Frontiers in Cellular and Infection Microbiology. 2020;**10**: article107

[30] Jevons M. "Celbenin"-resistant Staphylococci. British Medical Association. 1961;**1**:124-125

[31] Vysakh PR, Jeya M. A comparative analysis of community acquired and hospital acquired methicillin resistant *Staphylococcus aureus*. Journal of Clinical and Diagnostic Research. 2013;7(7): 1339-1342

[32] Schulte RH, Munson E. *Staphylococcus aureus* resistance patterns in Wisconsin. 2018 Surveillance of Wisconsin organisms for trends in antimicrobial resistance and epidemiology (SWOTARE) program report. Clinical Medicine & Research. 2019;**17**(3-4):72-81. DOI: 10.3121/ cmr.2019.1503

[33] Peacock SJ, Paterson GK. Mechanisms of methicillin resistance in *Staphylococcus aureus*. Annual Review of Biochemistry. 2015;**84**:577-601

[34] Fisher JF, Mobashery S. beta-lactam resistance mechanisms: Gram-positive bacteria and *Mycobacterium tuberculosis*. Cold Spring Harbor Perspectives in Medicine. 2016;**6**:a025221

[35] Hiramatsu K, Ito T, Tsubakishita S, Sasaki T, Takeuchi F, Morimoto Y, et al. Genomic basis for methicillin resistance in *Staphylococcus aureus*. Infection & Chemotherapy. 2013;**45**:117-136

[36] Chambers HF, DeLeo FR. Waves of Resistance: *Staphylococcus aureus* in the Antibiotic Era. Nature Reviews Microbiology. 2009;7(9):629-641. DOI: 10.1038/nrmicro2200

[37] Chongtrakool P, Ito T, Ma XX, Kondo Y, Trakulsomboon S, Tiensasitorn C, et al. Staphylococcal cassette chromosome *mec* (SCCmec) typing of methicillin-resistant *Staphylococcus aureus* strains isolated in 11 Asian countries: A proposal for a new nomenclature for SCCmec elements. Antimicrobial Agents and Chemotherapy. 2006 Mar;**50**(3):1001-1012. DOI: 10.1128/AAC.50.3.1001-1012. 2006

[38] Urushibara N, Paul SK, Hossain MA, Kawaguchiya M, Kobayashi N. Analysis of Staphylococcal cassette chromosome *mec* in *Staphylococcus haemolyticus* and *Staphylococcus sciuri*: Identification of a novel *ccr* gene complex with a newly identified *ccrA* allotype (*ccrA7*). Microbial Drug Resistance. 2011;**17**(2):291-297. DOI: 10.1089/ mdr.2010.0144

[39] Deplano A, Vandendriessche S, Nonhoff C, Denis O. Genetic diversity among methicillin-resistant *Staphylococcus aureus* isolates carrying the *mecC* gene in Belgium. Journal of Antimicrobial Chemotherapy. 2014;**69**: 1457-1460

[40] Cuny C, Layer F, Strommenger B, Witte W. Rare occurrence of methicillin resistant *Staphylococcus aureus* CC130 with a novel *mecA* homologue in humans in Germany. PLoS One. 2011;**6**:e24360

[41] Paterson GK, Morgan FJ, Harrison EM, Cartwright EJ, Torok ME, Zadoks RN, et al. Prevalence and characterization of human *mecC* methicillin-resistant *Staphylococcus aureus* isolates in England. Journal of Antimicrobial Chemotherapy. 2014;**69**:907-910

[42] Lakhundi S, Zhang K. Methicillin-Resistant *Staphylococcus aureus*: Molecular characterization, evolution, and epidemiology. Clinical Microbiology Reviews. 2018;**31**:e00020-e00018. DOI: 10.1128/ CMR.00020-18

[43] Kulkarni AP, Nagvekar VC, Veeraraghavan B, Warrier AR, Deepak TS, Ahdal J, et al. Current perspectives on treatment of grampositive infections in India: What is the way forward? Interdisciplinary Perspectives on Infectious Diseases. 2019;**2019**:7601847

[44] Deyno S, Fekadu S, Astatkie A. Resistance of Staphylococcus aureus to antimicrobial agents in Ethiopia: A meta-analysis. Antimicrobial Resistance and Infection Control. 2017;**6**:85. DOI: 10.1186/s13756-017-0243-7

[45] Alexander J, Chatterjee SS, Hamilton SM, Eltis LD, Chambers HF, Strynadka N. Structural and kinetic analyses of penicillin-binding protein 4 (PBP4)-mediated antibiotic resistance in *Staphylococcus aureus*. Journal of Biological Chemistry. 2018;**293**: 19854-19865

[46] Meletis G. Carbapenem resistance: Overview of the problem and future perspectives. Therapeutic Advances in Infectious Disease. 2016;**3**:15-21

[47] Meletis G, Exindari M, Vavatsi N, Sofianou D, Diza E. Mechanisms responsible for the emergence of carbapenem resistance in *Pseudomonas aeruginosa*. Hippokratia. 2012;**16**: 303-307

[48] Kumari N, Kumari V, Kumar V, Kumar S, Kumar A. Ouabain potentiates the antimicrobial activity of aminoglycosides against *Staphylococcus aureus*. BMC Complementary and alternative Medicine. 2019;**19**:119-129

[49] Carver PL. The battle for iron between humans and microbes. Current Medicinal Chemistry. 2018;**25**:85-96

[50] Golkar Z, Bagasra O, Pace DG. Bacteriophage therapy: A potential solution for the antibiotic resistance crisis. Journal of Infection in Developing Countries. 2014;**8**:129-136

[51] Dias RS, Eller MR, Duarte VS, Pereira AL, Silva CC, Mantovani HC, et al. Use of phages against antibioticresistant *Staphylococcus aureus* isolated from bovine mastitis. Journal of Animal Science. 2013;**91**:3930-3939

