

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



# Silver Nanoparticles Impregnated Wound Dressings: Recent Progress and Future Challenges

*Atamjit Singh, Kirandeep Kaur, Komalpreet Kaur,  
Jaijeet Singh, Nitish Kumar, Neena Bedi  
and Preet Mohinder Singh Bedi*

## Abstract

Microbial infection remains all time and unresolved challenge in the management of burns and diabetic wounds. Especially in diabetic wounds infections are prominent reason of amputations. Microbial biofilms pose tough polymeric barrier that is difficult to cross by conventional antibiotics. Therefore, traditional approach of infection control using antibiotics is now failing at some extent that raised a need to shift this paradigm. Presently, silver nanoparticles incorporated scaffolds representing a new concept of nanoparticle dressings which is becoming popular in wound management. Recently developed silver nanoparticles functionalized wound dressings exhibited excellent profile in the management of wound infections and promotion of wound healing. This chapter throw light on the recent strategies used in the development of silver nanoparticles functionalized wound dressings and their outcomes along with potential benefits and future challenges in wound management.

**Keywords:** Silver nanoparticles, wound infections, biofilms, diabetic wounds, wound dressing

## 1. Introduction

A wound is any type of injury which can be in the form of a cut, bruise or contusion caused by some external force. Open wounds are susceptible of getting infected by microorganisms like bacteria, virus or fungi if left unattended. Infected wounds pose a major challenge for healthcare system due to its direct relation with mortality and morbidity of the affected patients [1]. When a wound is infected it causes stress (physiological and psychological) and slows down the wound healing process and in some cases worsens the situation [2]. Approximately 2% of all hospitalized patients worldwide have a chronic wound, and older adult patients are at highest risk, because aging impairs the healing process [3, 4]. As many as 70% of these wounds recur, and 34% are accompanied by infection [5]. A survey by Medicare beneficiaries in 2018 stated that approximately 8.2 million individuals were having some kind of wounds, with or without infection and they estimated the treatment cost to be near about 28.1 billion to 96.8 billion dollars making it a big

economic challenge as well [6]. Infectious wounds like surgical wounds, diabetic foot and ulcer are more problematic and are reported more frequently than other infectious wounds [7]. Silver since time memorial is used for its good antimicrobial potential due to its interference with the thiol group of the microbial membrane. It also increases production of reactive oxygen species with in the microbe resulting in damage of DNA and bacterial proteins [8]. Due to its broad spectrum antimicrobial activity, silver nanoparticles are getting a great attention especially in the form of dressing for the infected wounds. This chapter will throw light on the various applications and recent trends in the health care system for treatment of infectious wounds using silver nanoparticles.

## **2. Wound and wound infections**

In general, any form of breakage or harm to the surface of skin can be termed as a wound and broadly classified into two types i.e. acute wound and chronic wound. An acute wound is one in which the wound physiology tends to remain normal during the process of healing. Acute wounds are usually bites, minor burns, cuts and surgical wounds. It is seen that such kind of wounds heal within a predictable timespan depending upon the location, depth and type of wound. In case of chronic wounds the physiology of the wound is disturbed due to various endogenous mechanisms which results in damaging the integrity of the tissue. Examples of chronic wounds are ulcers, diabetic foot and pressure sores. These types of wounds aggravate by aging, malnutrition, diseases which results in immunosuppression within the patient like AIDS or in patients on immunosuppressant drugs [9, 10]. Wound is said to be infected when there is entry of a microbe from the breached skin, which slows down the healing process and results in appearance of signs and symptoms like pain, discoloration of the wounded area, edema, puss, abnormal smell, tenderness etc. [11]. Most of the hard to heal infected wounds are observed to possess biofilms in them [12]. Biofilms are surface linked microbial structural communities having sessile cells present in the matrix produced by the microbe itself, made of polysaccharides, deoxyribonucleic acid and other components which are an essential type of adaptation strategy for the survival of bacteria as it protects it from the harsh surroundings and several immune responses by the extracellular polymeric substance (EPS) [13]. Bacteria possessing biofilms has increased chance of gene transfer of antibiotic resistance gene to other bacterial species [14, 15]. Bacteria having biofilms are tough to treat even with higher doses as biofilms prevent antimicrobial agents to reach up to bacteria [16, 17]. Although there are many beliefs regarding the defensive mechanism of biofilms against the antimicrobial agents but till date its nature of defense is unknown [18].

## **3. Wound healing**

Replacement of damaged tissue by newly produced tissue is termed as wound healing [19]. Skin (epidermis and dermis) acts as a protective layer against the harsh environment and when this barrier is breached a sequence of biochemical events takes place to repair the damage [19, 20]. The process of wound healing is generally described in four phases i.e. blood clotting, inflammation, tissue growth and tissue remodeling [21]. In the first phase with in some time platelets starts covering the area of the affected site. They release a particular chemical signal which promotes clotting resulting in activation of fibrin which produces a mesh to which platelets bind and forms a clot. This phase is also known as hemostasis [22, 23]. In second

phase or inflammation stage clearing of debris and bacterial cells takes place via process of phagocytosis where white blood cells and macrophages engulf and destroys them [24]. In the next phase that is proliferation phase or tissue growth phase angiogenesis, collagen deposition, granulation tissue formation, epithelialization and wound contraction takes place [25]. At last the tissue enters the maturation or the remodeling phase in which collagen is realigned and the cells which are not needed are removed by apoptosis [26].

#### **4. Silver as an antimicrobial agent**

Since early times silver appeared in recorded history texts for its excellent antimicrobial action. The ancient Greek historian 'Herodotus' describes that at the times of war Persian kings among with his provisions used to take boiled water from silver jars [27, 28]. Raulin in 1869 was the first person who observed the antimicrobial activity of silver by observing that *Aspergillus niger* (most common type of fungus) was not able to grow in silver vessels [29, 30]. Carl Wilhelm von Nageli, a Swiss botanist came up with a term 'oligodynamics' (oligo means small; dynamics means power) which describes any metal having some antibacterial properties at very small concentration [31, 32]. Silver compounds are being used in the process of wound healing since 1970s, leading to discovery of silver sulphadiazine which has an effective, broad spectrum antibacterial activity [33]. The silver element in nature is inert and ionizes when it comes in contact with environment, producing  $\text{Ag}^+$  ions which are believed to show antimicrobial activity [34]. Because of its strong antimicrobial activity, silver is a mostly used as an adjunct therapy in wound care. However, it also has the potential to delay the process of wound healing by producing toxic effects on keratinocytes and fibroblasts [35].

#### **5. Silver nanoparticles**

Any small particle which ranges between 1 to 100 nanometers ( $10^{-9}$ ) in size is known as a nanoparticle. It cannot be detected with naked human eye and it exhibits different chemical and physical properties in comparison to their large material counterparts [36]. Among many inorganic nanoparticles, silver nanoparticles (AgNPs) have got researchers attention around the globe due its novel physical, chemical and biological properties as compared to their bulk form. They have particular chemical and physical properties like high electrical and thermal conductivity [37], surface enhanced Raman scattering effect [38], catalytic activity [39], chemical stability [40] and nonlinear optical behavior [41]. The above stated properties make it the material of choice to be used in electronics, and for medical use. Silver nanoparticles are also known for their antiviral, antibacterial and antifungal activity. Due to smaller size, surface area to volume ratio is increased which results in increase in the amount of atoms on the surface, that other forms. The net effect gives rise to unpredictable properties associated with nanoparticles [42]. Silver nanoparticles has shown proven antimicrobial activity in many in vivo and in vitro studies and have application in soaps, cosmetics, food packaging and wound dressing [43]. Due to epidermal keratin and phospholipids, and protein thiol groups, skin was thought to be impermeable to silver nanoparticles but some studies demonstrated, if any absorption beyond the stratum corneum [44, 45]. Silver wound dressings are in direct contact with damaged skin resulting in systemic absorption also associated with some toxicity [46].

## **6. Mechanistic insight of silver nanoparticles**

In an average human concentration of silver in plasma is less than 2 µg/mL which is derived from inhalation of particulate matter and diet [44]. Silver can enter human body by inhalation, oral ingestion and dermal absorption [34]. Pinocytosis and endocytosis are believed to be two processes by which the silver nanoparticles may enter the body. It is seen that the particles that are of nanoscale penetrate much deeper than those of regular size leading to a novel delivery therapy [47, 48]. Till now exact mechanism of action of silver nanoparticles is not clear but several actions have been proposed by the scientists for its antimicrobial activity. Continual release of silver ion is considered to be the main reason for its antimicrobial activity [49]. Due to sulfur protein affinity and electrostatic attraction silver ions adhere to the wall of cells and cytoplasmic membrane which increases its permeability and penetrability into the cytoplasmic membrane leading to disruption of the bacterial cell wall [50]. When the silver ion enters the cell it can deactivate the respiratory enzymes and can generate reactive oxygen species [51]. Reactive oxygen species acts as a key component and a major reason for cell membrane disruption and DNA damage (by interacting with sulfur and phosphorus of DNA) causing problem in DNA replication, reproduction results in death of the microbes. Silver ions also inhibit the synthesis of proteins by denaturation of ribosomes and cause interruption the production of ATP [52]. After anchoring the surface of the cell silver nanoparticles gets accumulated in the pits of the cellular wall of microbe resulting in cell membrane denaturation [53]. Due to nanosize they easily penetrate cell membrane, leading to rupture of cell organelles and even lysis. They also affect the bacterial transduction process by interfering with the phosphorylation of protein substrates which can result in cell apoptosis and cell multiplication [53, 54]. Gram-negative bacterial strains are more sensitive towards the effect of silver nanoparticles because the cellular walls of these bacteria are narrower than the gram positive bacteria [55]. One drawback of silver nanoparticles is that they are not much effective in the case of bacteria having biofilms. Biofilms protects the membrane from both nanoparticles and silver ions by altering their transport due to its complicated structure [56]. The pathway of the nanoparticles penetration is highly obstructed if the size is greater than 50 nm [57]. It is also observed that adsorption and accumulation of the silver nanoparticles on the biofilm results in reduced diffusion of nanoparticles in bacteria [58].

## **7. Silver nanoparticles wound dressings**

A dressing is a sterile material applied to a wound to promote healing and protect the wound from further harm [59, 60]. It has been designed in such a way that it is in direct contact with the wound, as distinguished from a bandage, which is most often used to hold a dressing in place. Silver dressings are used for both types of wounds (acute and chronic) and when there is risk of high level of bio burden or local infection for example in the case of burns [61]. Silver dressings helps in reducing the bioburden in infected or colonized wounds and also acts as a barrier to reduce any further chance of infection [62].

## **8. Synthesis of silver nanoparticles for wound dressing**

There are four types of silver nanoparticles synthesis, namely chemical, irradiation, green and thermal. In chemical synthesis, two types of synthesis methods are

used which are Brust-Schiffrin synthesis which is mainly used for golden nanoparticles and Turkevich method which is based on reduction of the boiling solution of silver salt with citrate salt solution [63, 64]. Irradiation synthesis is connected with radiation of precursors or intermediate products of reaction with electromagnetic radiation with different wavelengths [65–67]. In green synthesis plant, fungus or bacterial extract is mixed with silver ion usually silver nitrate, the bioactive molecules of extracts reduce silver ion to elementary silver and then it is precipitated in alcohol. Advantages of this synthesis are that cost involved in synthesis is low, environment friendly and plant extracts contain medicinal compounds which are used in conventional medicine [68–77]. Thermal synthesis is based on the principle of thermal reduction of silver salt. After the synthesis of the silver nanoparticles, they are incorporated in membrane or composite material, nanofibers, hydrogels, etc. and are used as a wound dressing.

### 8.1 Silver nanoparticles incorporated into membrane and composite material

Membrane and composite material-immobilized nanoparticles can have many functions including antimicrobial activity. Silver nanoparticles incorporated in membranes like polyethersulfone, acetate cellulose, polydopamine-coated poly(ether imide) etc. showed significant antimicrobial activity against diverse

Polymer used	Method of preparation of nanoparticles	Size (nm)	Result
Bacterial cellulose	Thermal method (thermal reduction at 80° C)	10–30	Reduction seen in <i>Staphylococcus aureus</i> ; 99%. Growth of cells observed with no cytotoxicity [79]
Chitin	Irradiation method, (gamma rays, <sup>60</sup> Co)	3–13	Significant bactericidal effect (p < 0.01) [80]
Bacterial cellulose	Green method (cellulose from <i>Acetobacter xylinum</i> )	50–150	Strong activity against <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [81]
Chitosan, Polyvinyl alcohol, Curcumine	Green method (chitosan)	16	Significant effects against <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> and <i>Candida albicans</i> [82]
Chitosan and Chitin	Green method ( <i>Camelia sinensis</i> )	60–150	Good healing activity [83]
Konjac Glucomannan	Green method (egg white)	8–32	Accelerates wound healing and fibroblast growth promotion [84]
Bacterial cellulose	Chemical method (NaBH <sub>4</sub> )	3–17	Inhibitor of growth of <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [85]
Chitin	Chemical method (sodium citrate)	5	Inhibitor of growth of <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [86]
Poly vinyl pyrrolidone chitosan	Chemical method (sodium citrate)	10–30	Reduces growth of <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [87]
Chitosan	Chemical method (NaBH <sub>4</sub> )	15	Facilitates cell proliferation and mitigate bacterial infection [88]
Bacterial cellulose	Chemical method (NaBH <sub>4</sub> )	5–14	Inhibition of growth of <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [89]

**Table 1.**  
 Silver nanoparticles based membrane composites for wound management.

Material used for preparation of clothing and dressings	Method of preparation of nanoparticles	Size (nm)	Result
Cotton fabrics	Chemical method (alkali solution of starch)	22–24	Inhibition of growth of <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [90]
Dressing material	Chemical method (NaBH <sub>4</sub> )	4–24	Reduction in wound inflammation and fibrogenic cytokines modulation [91]
Cotton fabrics	Green method ( <i>Fusarium oxysporum</i> )	2	Reduction of growth on <i>Staphylococcus aureus</i> [92]
Silver nanoparticles incorporated wound dressing	Green method ( <i>Aspergillus niger</i> )	200–800	Effective wound healing activity [93]

**Table 2.**  
Silver nanoparticles incorporated clothing and dressings for wound management.

Polymer used for preparation of nanofibers	Method of preparation of nanoparticles	Size (nm)	Result
Collagen	Chemical method (sodium citrate)	25–55	Accelerated wound healing [96]
Poly vinyl pyrrolidone	Chemical method (N,N- DMF)	3–5	Effective antibacterial action [97]
Poly methyl methacrylate-co-dopamine	Chemical method (Silver ion dipped in PMMDM)	<20	Effective antibacterial and wound healing action [98]
Plumbagine	Chemical method (PBG reduction)	60	High antibacterial activity [99]
Gelatine	Chemical method (Silver nitrate reduced with gelatin powder)	11–20	High anti antibacterial activity against Gram positive bacteria [100]
Poly ethylene oxide poly caprolactone	Chemical method (Silver nitrate reduced with PEO and DMF)	13–17	Good antibacterial potential [101]
Alginate	Chemical method (NaBH <sub>4</sub> )	5–17	Reduces the inflammatory phase and increased epidermal thickness [102]
Chitosan, glucose, Poly vinyl alcohol	Green method (chitosan, glucose)	10–30	Good antibacterial activity against gram negative bacteria [103]
Poly galacturonic acid	Green method (PGA, HA)	5–13	Maximum wound epithelization and collagen deposition [104]
Poly caprolactone	Green method ( <i>P. nigrum</i> )	5–20	Excellent antibacterial activity against <i>Escherichia coli</i> and <i>Staphylococcus aureus</i> [105]
Poly vinyl alcohol	Irradiation method	23–24	Significant antibacterial activity against <i>Staphylococcus aureus</i> and <i>Pseudomonas aeruginosa</i> [106]

**Table 3.**  
Silver nanoparticles incorporated nanofibers for wound management.

range of microbes thus have capability to sterile wound environment and promote healing process [78], representative examples are summarized in **Table 1**.

## 8.2 Powdered silver nanoparticle and topical application

Powdered silver nanoparticles are used for incorporation into different types of clothing and dressings. Representative examples are summarized in **Table 2**.

## 8.3 Nanofibers

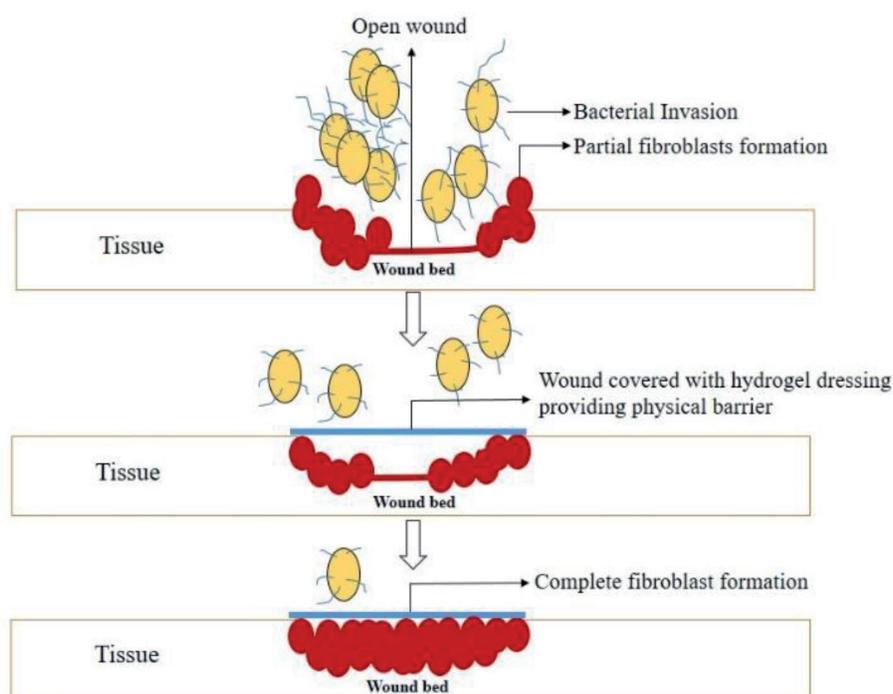
Nanofibers are emerged as an important structures with wide range of biological as well as physical applications like air filtration, immunoanalysis and as pseudo-enzymes etc. [81–84, 94, 95]. Apart from that active research is also ongoing for utilization of silver nanoparticles incorporated nanofibers for wound management. Examples of nanofibers are given in **Table 3**.

## 8.4 AgNPs-hydrogels

Hydrogels have excellent capacity to absorb wound exudates and at the same time maintain the moisture in wound environment to ensure proper healing. Hydrogels form impermeable physical barrier on wound surface and prevent bacterial invasion (**Figure 1**) and apart from that hydrogels also showcased its tendency to absorb wide range of metals [59, 96, 107, 108]. Some silver nanoparticles incorporated hydrogels showed excellent wound healing activity as shown in **Table 4**.

## 9. Future challenges

Silver nanoparticles functionalized wound dressings have significant antimicrobial activity and provide faster and effective tissue repair thus they are widely



**Figure 1.** Schematic layout of hydrogel membrane reducing bacterial invasion and accelerating wound healing process.

Polymer used for preparation of hydrogels	Method of preparation of nanoparticles	Size (nm)	Result
Poly acrylic acid and poly vinyl alcohol	Chemical method (NaBH <sub>4</sub> )	2–3	Significant antibacterial activity against <i>Escherichia coli</i> [109]
Beta- chitin	Chemical method (sodium citrate)	4–8	It showed inhibitory effects on the bacteria growth [110]
Chitosan	Green method (sericin and chitosan)	240–970	Bactericidal action [111]
2-acrylamide-2-methylpropane sulphonic acid sodium salt	Irradiation method (UV radiation)	—	Used as a burn wound dressing due to its good antibacterial activity [112]
Poly vinyl alcohol	Irradiation method (UV radiation, gamma rays <sup>60</sup> Co)	90	Antimicrobial activity against <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> and <i>Candida albicans</i> [113]
Collagen	Thermal method (reduction at 40°C)	5–14	Antimicrobial activity against <i>Escherichia coli</i> , <i>Staphylococcus aureus</i> and <i>Pseudomonas aeruginosa</i> [114]
Carboxymethylcellulose	Thermal method (reduction at 70–100°C)	7–21	Removes the exudates and prevents wound maceration [115]

**Table 4.**  
*Silver nanoparticles incorporated hydrogels for wound management.*

explored for wound healing activity. Beside this they are far away from clinical practice as well as commercialization. More rigorous preclinical investigations are still required to validate their capability for tissue regeneration. Apart from that work is also needed to be done on industrial scale up techniques for commercialization.

## 10. Conclusion

Nanomaterials now a days representing potential ways for combating microbial related diseases and disorders. Wounds especially chronic ones burdened with resistant microbes are posing serious challenge to healthcare system. For that purpose silver nanoparticles impregnated wound dressing due to their excellent antimicrobial potential are not less than a boon. Along with sterilization they are proven to fasten tissue repair in wounds. Presently more rigorous efforts are needed in their preclinical investigations to evaluate their efficacy verses safety ratio. They have capability to become potential wound dressing of future.

## Acknowledgements

The authors are also thankful to Guru Nanak Dev University, Amritsar for providing various facilities to carry out the work.

## Conflict of interest

The authors declare no conflict of interest.

IntechOpen

IntechOpen

### **Author details**

Atamjit Singh\*, Kirandeep Kaur, Komalpreet Kaur, Jaijeet Singh, Nitish Kumar,  
Neena Bedi and Preet Mohinder Singh Bedi  
Department of Pharmaceutical Sciences, Guru Nanak Dev University,  
Amritsar, Punjab, India

\*Address all correspondence to: [atamjitpharma.rsh@gndu.ac.in](mailto:atamjitpharma.rsh@gndu.ac.in)

### **IntechOpen**

---

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. 

## References

- [1] Wilkinson LJ, White RJ, Chipman JK. Silver and nanoparticles of silver in wound dressings: a review of efficacy and safety. *Journal of wound care*. 2011 Nov;20(11):543-549
- [2] Bowler PG, Duerden BI, Armstrong DG. Wound microbiology and associated approaches to wound management. *Clinical microbiology reviews*. 2001 Apr 1; 14(2):244-269.
- [3] Powers JG, Higham C, Broussard K, Phillips TJ. Wound healing and treating wounds: chronic wound care and management. *J Am Acad Dermatol* 2016;74(4):607-625.
- [4] Gottrup F, Holstein P, Jorgensen B, Lohmann M, Karlsmar T. A new concept of a multidisciplinary wound healing center and a national expert function of wound healing. *Arch Surg* 2001;136(7):765-772.
- [5] Wong SY, Manikam R, Muniandy S. Prevalence and antibiotic susceptibility of bacteria from acute and chronic wounds in Malaysian subjects. *J Infect Dev Ctries* 2015;9(9):936-944.
- [6] Nussbaum SR, Carter MJ, Fife CE, DaVanzo J, Haught R, Nussbaum M, Cartwright D. An economic evaluation of the impact, cost, and medicare policy implications of chronic nonhealing wounds. *Value in Health*. 2018 Jan 1; 21(1):27-32.
- [7] <https://www.apma.org/diabetic-woundcare>
- [8] Chaloupka, K., Malam, Y., Seifalian, A.M. Nanosilver as a new generation of nanoparticle in biomedical applications. *Trends Biotechnol*. 2010; 28: 11, 580-588.
- [9] [https://healthywa.wa.gov.au/Articles/U\\_Z/Wounds#:~:text=A%20wound%20is%20any%20damage%20to%20the%20legs%20and%20feet.](https://healthywa.wa.gov.au/Articles/U_Z/Wounds#:~:text=A%20wound%20is%20any%20damage%20to%20the%20legs%20and%20feet.)
- [10] Okur ME, Karantas ID, Şenyiğit Z, Okur NÜ, Siafaka PI. Recent trends on wound management: New therapeutic choices based on polymeric carriers. *Asian Journal of Pharmaceutical Sciences*. 2020 Jan 23.
- [11] Aftab S, Tarik MM, Siddique MA, Yusuf MA. Clinical and microbiological aspect of wound infection: a Review Update. *Bangladesh Journal of Infectious Diseases*. 2014;1(2):32-37.
- [12] Azeredo, J.; Azevedo, N.F.; Briandet, R.; Cerca, N.; Coenye, T.; Costa, A.R.; Desvaux, M.; Di Bonaventura, G.; Hébraud, M.; Jaglic, Z.; et al. Critical review on biofilm methods. *Crit. Rev. Microbiol*. 2017, 43, 313-351.
- [13] Chew, S. C., & Yang, L. (2016). *Biofilms*. *Encyclopedia of Food and Health*, 407-415.
- [14] Galloway W.R.J.D., Hodgkinson J.T., Bowden S., Welch M., Spring D.R. Applications of small molecule activators and inhibitors of quorum sensing in Gram-negative bacteria. 2012; 20:449-458.
- [15] Galloway W.R.J.D., Hodgkinson J.T., Bowden S.D., Welch M., Spring D.R. Quorum sensing in gram-negative Bacteria : small-molecule modulation of AHL and AI-2 quorum sensing pathways. *Chem. Rev*. 2011; 44:28-67.
- [16] Abraham W.-R. Going beyond the control of quorum-sensing to combat biofilm infections. *Antibiotics*. 2016; 5:3-16.
- [17] Hughes G., Webber M.A. Novel approaches to the treatment of bacterial biofilm infections. *Br. J. Pharmacol*. 2017; 174:2237-2246.

- [18] McCarty S, Woods E, Percival SL. Biofilms: from Concept to Reality. In *Biofilms in Infection Prevention and Control* 2014 Jan 1 (pp. 143-163). Academic Press.
- [19] Nguyen DT, Orgill DP, Murphy GF. The pathophysiologic basis for wound healing and cutaneous regeneration. In *Biomaterials for treating skin loss* 2009 Jan 1 (pp. 25-57). Woodhead Publishing.
- [20] Rieger S, Zhao H, Martin P, Abe K, Lisse TS. The role of nuclear hormone receptors in cutaneous wound repair. *Cell biochemistry and function*. 2015 Jan;33(1):1-3.
- [21] Stadelmann WK, Digenis AG, Tobin GR. Physiology and healing dynamics of chronic cutaneous wounds. *The American Journal of Surgery*. 1998 Aug 1;176(2):26S-38S.
- [22] Rasche H. Haemostasis and thrombosis: an overview. *European Heart Journal Supplements*. 2001 Dec 1; 3(suppl\_Q):Q3-7.
- [23] Versteeg HH, Heemskerk JW, Levi M, Reitsma PH. New fundamentals in coagulation. *Physiological Reviews*. 2013 Jan; 93:327-358.
- [24] Hirayama D, Iida T, Nakase H. The phagocytic function of macrophage-enforcing innate immunity and tissue homeostasis. *International journal of molecular sciences*. 2018 Jan;19(1):92.
- [25] Chang HY, Sneddon JB, Alizadeh AA, Sood R, West RB, Montgomery K, Chi JT, Van De Rijn M, Botstein D, Brown PO. Gene expression signature of fibroblast serum response predicts human cancer progression: similarities between tumors and wounds. *PLoS Biol*. 2004 Jan 13; 2(2):e7.
- [26] Medrado AR, Pugliese LS, Reis SR, Andrade ZA. Influence of low level laser therapy on wound healing and its biological action upon myofibroblasts. *Lasers in surgery and medicine*. 2003 Mar; 32(3):239-244.
- [27] E.H. Blakeney (ed) (1945) "The History of Herodotus" translated by G. Rawlinson; Dent, London
- [28] G. Sykes (1958) "Disinfection and Sterilization"; Spon, London.
- [29] J. Raulin (1869) *Sci. Nat.*, 11, 93. Berk (3) Abstr. 1
- [30] K.W. von Ngeli (1893) *Densch. schweiz natudorsch Ges.* 33, 174. Berk (3) Abstr. 5.
- [31] I.B. Romans (1968) in "Disinfection, Sterilization and Preservation" edited by C.A. Lawrence and S.S. Block; Lea and Febiger, London, pp372-400 and pp469-475.
- [32] Berk RG. Abstracts of articles on oligodynamic sterilization. The Engineer Board, US Army Corps of Engineers [Project WS 768], Fort Belvoir, Virginia. 1947.
- [33] Cooper, R., White, R.J.; Silver sulfadiazine: a review of the evidence. *Wounds-UK*; 2005; 1: 2, 51-61.
- [34] Lansdown AB. A pharmacological and toxicological profile of silver as an antimicrobial agent in medical devices. *Advances in pharmacological sciences*. 2010 Oct;2010.
- [35] Khansa I, Schoenbrunner AR, Kraft CT, Janis JE. Silver in wound care—friend or foe?: a comprehensive review. *Plastic and Reconstructive Surgery Global Open*. 2019 Aug;7(8).
- [36] <https://www.twi-global.com/technical-knowledge/faqs/what-are-nanoparticles>
- [37] Chan, K.L.; Mariatti, M.; Lockman, Z.; Sim, L.C. Effects of the Size and Filler Loading on the Properties of

- Copper- and Silver-Nanoparticle-Filled Epoxy Composites. *J. Appl. Polym. Sci.* 2011, 121, 3145-3152.
- [38] Stampelcoskie, K.G.; Scaiano, J.C.; Tiwari, V.S.; Anis, H. Optimal size of silver nanoparticles for surface-enhanced raman spectroscopy. *J. Phys. Chem. C* 2011, 115, 1403-1409
- [39] Zhang, P.; Shao, C.L.; Zhang, Z.Y.; Zhang, M.Y.; Mu, J.B.; Guo, Z.C.; Liu, Y.C. In Situ assembly of well-dispersed Ag nanoparticles (AgNPs) on electrospun carbon nanofibers (CNFs) for catalytic reduction of 4-nitrophenol. *Nanoscale* 2011, 3, 3357-3363.
- [40] Chinnapongse, S.L.; MacCuspie, R.I.; Hackley, V.A. Persistence of singly dispersed silver nanoparticles in natural freshwaters, synthetic seawater, and simulated estuarine waters. *Sci. Total Environ.* 2011, 409, 2443-2450.
- [41] Karimzadeh R, Mansour N. The effect of concentration on the thermo-optical properties of colloidal silver nanoparticles. *Optics & Laser Technology.* 2010 Jul 1;42(5):783-789.
- [42] Jones, C.F., Grainger, D.W. *In vitro* assessments of nanomaterial toxicity. *Adv Drug Deliv Rev.* 2009; 61:438-456.
- [43] Liao, C.Z.; Li, Y.C.; Tjong, S.C. Bactericidal and cytotoxic properties of silver nanoparticles. *Int. J. Mol. Sci.* 2019, 20, 449.
- [44] Lansdown, A.B. Physiological and toxicological changes in the skin resulting from the action and interaction of metal ions. *Crit Rev Toxicol.*1995; 25: 5, 397-462.
- [45] Stern, S.T., McNeil, S.E. Nanotechnology safety concerns revisited. *Toxicol Sci.* 2008; 101: 1, 4-21.
- [46] Johnston, H.J., Hutchison, G., Christensen, F.M., et al. A review of the in vivo and in vitro toxicity of silver and gold particulates: Particle attributes and biological mechanisms responsible for the observed toxicity. *Crit Rev Toxicol.* 2010; 40: 4, 328-346.
- [47] AshaRani, P.V., Low Kah Mun, G., Hande, M.P., Valiyaveetil, S. Cytotoxicity and genotoxicity of silver nanoparticles in human cells. *ACS Nano.* 2009; 24; 3: 2, 279-290.
- [48] Liu, X., Lee, P.Y., Ho, C.M., et al. Silver nanoparticles mediate differential responses in keratinocytes and fibroblasts during skin wound healing. *Chem Med Chem.* 2010; 1; 3, 468-475.
- [49] Bapat RA, Chaubal TV, Joshi CP, et al. An overview of application of silver nanoparticles for biomaterials in dentistry. *Mater Sci Eng C.* 2018;91:881-898. doi:10.1016/j.msec.2018.05.069
- [50] Khorrami S, Zarrabi A, Khaleghi M, Danaei M, Mozafari M. Selective cytotoxicity of green synthesized silver nanoparticles against the MCF-7 tumor cell line and their enhanced antioxidant and antimicrobial properties. *Int J Nanomedicine.* 2018;13:8013-8024. doi:10.2147/IJN.S189295
- [51] Ramkumar VS, Pugazhendhi A, Gopalakrishnan K, et al. Biofabrication and characterization of silver nanoparticles using aqueous extract of seaweed *Enteromorpha compressa* and its biomedical properties. *Biotechnol Rep.* 2017;14:1-7. doi:10.1016/j.btre.2017.02.001
- [52] Durán N, Nakazato G, Seabra A. Antimicrobial activity of biogenic silver nanoparticles, and silver chloride nanoparticles: an overview and comments. *Appl Microbiol Biotechnol.* 2016;100(15):6555-6570. doi:10.1007/s00253-016-7657-7
- [53] Liao C, Li Y, Tjong SC. Bactericidal and cytotoxic properties of silver nanoparticles. *Int J Mol Sci.*

2019;20(2):449. doi:10.3390/  
ijms20020449

[54] Li L, Li L, Zhou X, et al. Silver nanoparticles induce protective autophagy via Ca<sup>2+</sup>/CaMKK $\beta$ /AMPK/mTOR pathway in SH-SY5Y cells and rat brains. *Nanotoxicology*. 2019;13(3):369-391. doi:10.1080/17435390.2018.1550226

[55] Meikle T, Dyett BP, Strachan JB, White J, Drummond CJ, Conn CE. Preparation, characterization, and antimicrobial activity of cubosome encapsulated metal nanocrystals. *ACS Appl Mater Interfaces*. 2020;12(6):6944-6954. doi:10.1021/acsami.9b21783

[56] Saravanan M, Arokiyaraj S, Lakshmi T, Pugazhendhi A. Synthesis of silver nanoparticles from *Phenerochaete chrysosporium* (MTCC-787) and their antibacterial activity against human pathogenic bacteria. *Microb Pathog*. 2018;117:68-72. doi:10.1016/j.micpath.2018.02.008

[57] Yin IX, Yu OY, Zhao IS, et al. Developing biocompatible silver nanoparticles using epigallocatechin gallate for dental use. *Arch Oral Biol*. 2019;102:106-112. doi:10.1016/j.archoralbio.2019.03.022

[58] Pugazhendhi A, Prabakar D, Jacob JM, Karuppusamy I, Saratale RG. Synthesis and characterization of silver nanoparticles using *Gelidium amansii* and its antimicrobial property against various pathogenic bacteria. *Microb Pathog*. 2018;114:41-45. doi:10.1016/j.micpath.2017.11.013

[59] First Aid Equipment, Supplies, Rescue, and Transportation". *Hospital Corpsman*. Naval Education and Training Command. 2003. pp. 3-1

[60] Zarrintaj, Payam; Moghaddam, Abolfazl Salehi; Manouchehri, Saeed; Atoufi, Zhaleh; Amiri, Anahita; Amirkhani, Mohammad Amir; Nilforoushzadeh, Mohammad Ali; Saeb,

Mohammad Reza; Hamblin, Michael R; Mozafari, Masoud (October 2017). "Can regenerative medicine and nanotechnology combine to heal wounds? The search for the ideal wound dressing". *Nanomedicine*. 12 (19): 2403-2422. doi:10.2217/nnm-2017-0173

[61] International Wound Infection Institute (IWII) Wound infection in clinical practice. *Wounds International* 2016

[62] International consensus. Appropriate use of silver dressings in wounds. An expert working group consensus. London: Wounds International, 2012. Available to download from: [www.woundsinternational.com](http://www.woundsinternational.com)

[63] Lee, N.-Y.; Ko, W.-C.; Hsueh, P.-R. Nanoparticles in the treatment of infections caused by multidrug-resistant organisms. *Front. Pharmacol*. 2019, 10.

[64] Chugh, H.; Sood, D.; Chandra, I.; Tomar, V.; Dhawan, G.; Chandra, R. Role of gold and silver nanoparticles in cancer nano-medicine. *Artif. Cell. Nanomed. Biotechnol*. 2018, 46, S1210–S1220.

[65] Jia, X.W.; Yao, Y.C.; Yu, G.F.; Qu, L.L.; Li, T.X.; Li, Z.J.; Xu, C.P. Synthesis of gold-silver nanoalloys under microwave-assisted irradiation by deposition of silver on gold nanoclusters/triple helix glucan and antifungal activity. *Carbohydr. Polym*. 2020, 238, 7

[66] Yu, Z.L.; Wang, W.; Dhital, R.; Kong, F.B.; Lin, M.S.; Mustapha, A. Antimicrobial effect and toxicity of cellulose nanofibril/silver nanoparticle nanocomposites prepared by an ultraviolet irradiation method. *Colloid Surf. B Biointerfaces* 2019, 180, 212-220.

[67] Zhao, X.M.; Li, N.; Jing, M.L.; Zhang, Y.F.; Wang, W.; Liu, L.S.; Xu, Z.W.; Liu, L.Y.; Li, F.Y.; Wu, N.

- Monodispersed and spherical silver nanoparticles/graphene nanocomposites from gamma-ray assisted in-situ synthesis for nitrite electrochemical sensing. *Electrochim. Acta* 2019, 295, 434-443.
- [68] Saha, J.; Begum, A.; Mukherjee, A.; Kumar, S. A novel green synthesis of silver nanoparticles and their catalytic action in reduction of Methylene Blue dye. *Sustain. Environ. Res.* 2017, 27, 245-250.
- [69] Vorobyova, V.; Vasyliiev, G.; Skiba, M. Eco-Friendly "green" synthesis of silver nanoparticles with the black currant pomace extract and its antibacterial, electrochemical, and antioxidant activity. *Appl. Nanosci.* 2020, 12.
- [70] Bindhu, M.R.; Umadevi, M.; Esmail, G.A.; Al-Dhabi, N.A.; Arasu, M.V. Green synthesis and characterization of silver nanoparticles from *Moringa oleifera* flower and assessment of antimicrobial and sensing properties. *J. Photochem. Photobiol. B Biol.* 2020, 205, 7.
- [71] Jyoti, K.; Baunthiyal, M.; Singh, A. Characterization of silver nanoparticles synthesized using *Urtica dioica* linn. leaves and their synergistic effects with antibiotics. *J. Radiat. Res. Appl. Sci.* 2016, 9, 217-227.
- [72] Chinnasamy, G.; Chandrasekharan, S.; Bhatnagar, S. Biosynthesis of silver nanoparticles from *Melia azedarach*: Enhancement of antibacterial, wound healing, antidiabetic and antioxidant activities. *Int. J. Nanomed.* 2019, 14, 9823-9836.
- [73] Barbosa, A.; Silva, L.P.C.; Ferraz, C.M.; Tobias, F.L.; de Araujo, J.V.; Loureiro, B.; Braga, G.; Veloso, F.B.R.; Soares, F.E.D.; Fronza, M.; et al. Nematicidal activity of silver nanoparticles from the fungus *duddingtonia flagrans*. *Int. J. Nanomed.* 2019, 14, 2341-2348.
- [74] Hamedi, S.; Shojaosadati, S.; Shokrollahzadeh, S.; Hashemi-Najafabadi, S. Extracellular biosynthesis of silver nanoparticles using a novel and non-pathogenic fungus, *neurospora intermedia*: Controlled synthesis and antibacterial activity. *World J. Microbiol. Biotechnol.* 2014, 30, 693-704.
- [75] Saravanan, M.; Arokiyaraj, S.; Lakshmi, T.; Pugazhendhi, A. Synthesis of silver nanoparticles from *phenerochaete chrysosporium* (MTCC-787) and their antibacterial activity against human pathogenic bacteria. *Microb. Pathog.* 2018, 117, 68-72.
- [76] Li, F.S.; Weng, J.K. Demystifying traditional herbal medicine with modern approaches. *Nat. Plants* 2017, 3, 7. *Pharmaceutics* 2020, 12, 821-21 of 24
- [77] Sehnal, K.; Hosnedlova, B.; Docekalova, M.; Stankova, M.; Uhlirova, D.; Tothova, Z.; Kepinska, M.; Milnerowicz, H.; Fernandez, C.; Ruttkay-Nedecky, B.; et al. An assessment of the effect of green synthesized silver nanoparticles using sage leaves (*Salvia officinalis* L.) on germinated plants of maize (*Zea mays* L.). *Nanomaterials* 2019, 9, 1550.
- [78] Liang, M.; Su, R.X.; Huang, R.L.; Qi, W.; Yu, Y.J.; Wang, L.B.; He, Z.M. Facile in situ synthesis of silver nanoparticles on procyanidin-grafted eggshell membrane and their catalytic properties. *ACS Appl. Mater. Interfaces* 2014, 6, 4638-4649.
- [79] Wu, J.; Zheng, Y.; Song, W.; Luan, J.; Wen, X.; Wu, Z.; Chen, X.; Wang, Q.; Guo, S. In Situ synthesis of silver-nanoparticles/bacterial cellulose composites for slow-released antimicrobial wound dressing. *Carbohydr. Polym.* 2014, 102, 762-771.
- [80] Singh, R.; Singh, D. Chitin membranes containing silver nanoparticles for wound dressing

application. *Int. Wound J.* 2014, 11, 264-268.

[81] Hu, W.; Chen, S.; Li, X.; Shi, S.; Shen, W.; Zhang, X.; Wang, H. In Situ synthesis of silver chloride nanoparticles into bacterial cellulose membranes. *Mater. Sci. Eng. C* 2009, 29, 1216-1219.

[82] Kanikireddy, V.; Yallapu, M.; Varaprasad, K.; Nagireddy, N.; Ravindra, S.; Neppalli, S.; Raju, K. Fabrication of Curcumin Encapsulated Chitosan-PVA Silver Nanocomposite Films for Improved Antimicrobial Activity. *J. Biomater. Nanobiotechnol.* 2011, 2, 55-64.

[83] Ahamed, M.I.; Sankar, S.; Kashif, P.M.; Basha, S.K.; Sastry, T.P. Evaluation of biomaterial containing regenerated cellulose and chitosan incorporated with silver nanoparticles. *Int. J. Biol. Macromol.* 2015, 72, 680-686.

[84] Chen, H.; Lan, G.; Ran, L.; Xiao, Y.; Yu, K.; Lu, B.; Dai, F.; Wu, D.; Lu, F. A novel wound dressing based on a Konjac glucomannan/silver nanoparticle composite sponge effectively kills bacteria and accelerates wound healing. *Carbohydr. Polym.* 2018, 183, 70-80.

[85] Maneerung, T.; Tokura, S.; Rujiravanit, R. Impregnation of silver nanoparticles into bacterial cellulose for antimicrobial wound dressing. *Carbohydr. Polym.* 2008, 72, 43-51.

[86] Madhumathi, K.; Sudheesh, K.P.T.; Abhilash, S.; Sreeja, V.; Tamura, H.; Manzoor, K.; Nair, S.V.; Jayakumar, R. Development of novel chitin/nanosilver composite scaffolds for wound dressing applications. *J. Mater. Sci. Mater. Med.* 2010, 21, 807-813.

[87] Archana, D.; Singh, B.K.; Dutta, J.; Dutta, P.K. Chitosan-PVP-nano silver oxide wound dressing: In Vitro and in vivo evaluation. *Int. J. Biol. Macromol.* 2015, 73, 49-57.

[88] Levi-Polyachenko, N.; Jacob, R.; Day, C.; Kuthirummal, N. Chitosan wound dressing with hexagonal silver nanoparticles for hyperthermia and enhanced delivery of small molecules. *Colloids Surf. B Biointerfaces* 2016, 142, 315-324.

[89] Yang, G.; Xie, J.; Hong, F.; Cao, Z.; Yang, X. Antimicrobial activity of silver nanoparticle impregnated bacterial cellulose membrane: Effect of fermentation carbon sources of bacterial cellulose. *Carbohydr. Polym.* 2012, 87, 839-845.

[90] Hebeish, A.; El-Rafie, M.H.; El-Sheikh, M.A.; Seleem, A.A.; El-Naggar, M.E. Antimicrobial wound dressing and anti-inflammatory efficacy of silver nanoparticles. *Int. J. Biol. Macromol.* 2014, 65, 509-515.

[91] Tian, J.; Wong, K.; Ho, C.-M.; Lok, C.-N.; Yu, W.-Y.; Che, C.-M.; Chiu, J.-F.; Tam, P. Topical Delivery of Silver Nanoparticles Promotes Wound Healing. *Chem Med Chem* 2007, 2, 129-136.

[92] Duran, N.; Marcato, P.; Souza, G.; Alves, O.; Esposito, E. Antibacterial Effect of Silver Nanoparticles Produced by Fungal Process on Textile Fabrics and Their Euent Treatment. *J. Biomed. Nanotechnol.*

[93] Sundaramoorthi, C.; Mathews, D.; Sivanandy, D.P.; Kalaiselvan, V.; Rajasekaran, A. Biosynthesis of silver nanoparticles from *Aspergillus niger* and evaluation of its wound healing activity in experimental rat model. *Int. J. Pharm Tech Res.* 2009, 1, 1523-1529.

[94] Yun, B.J.; Koh, W.G. Highly-Sensitive SERS-based immunoassay platform prepared on silver nanoparticle-decorated electrospun polymeric fibers. *J. Ind. Eng. Chem.* 2020, 82, 341-348.

[95] Shen, B.L.; Zhang, D.Y.; Wei, Y.J.; Zhao, Z.H.; Ma, X.F.; Zhao, X.D.; Wang,

- S.; Yang, W.X. Preparation of Ag Doped Keratin/PA6 Nanofiber Membrane with Enhanced Air Filtration and Antimicrobial Properties. *Polymers* 2019, 11, 1511.
- [96] Rath, G.; Hussain, T.; Chauhan, G.; Garg, T.; Goyal, A.K. Collagen nanofiber containing silver nanoparticles for improved wound-healing applications. *J. Drug Target.* 2016, 24, 520-529.
- [97] Jin, W.-J.; Lee, H.; Jeong, E.; Park, W.H.; Youk, J. Preparation of Polymer Nanofibers Containing Silver Nanoparticles by Using Poly(N-vinylpyrrolidone). *Macromol. Rapid Comm.* 2005, 26, 1903-1907.
- [98] Ghavaminejad, A.; Unnithan, R.A.; Ramachandra, K.S.A.; Samarikhalaj, M.; Thomas, R.; Jeong, Y.; Nasser, S.; Murugesan, P.; Wu, D.; Park, C.; et al. Mussel-Inspired Electrospun Nanofibers Functionalized with Size Controlled Silver Nanoparticles for Wound Dressing Application. *ACS Appl. Mater. Interf.* 2015, 7.
- [99] Natarajan, D.; Lakra, R.; Srivatsan, K.; Usha, R.; Korrapati, P.; Kiran, M. Plumbagin caged silver nanoparticle stabilized collagen sca\_old for wound dressing. *J. Mater. Chem. B* 2014, 3.
- [100] Rujitanaroj, P.-O.; Pimpha, N.; Supaphol, P. Wound-Dressing materials with antibacterial activity from electrospun gelatin fiber mats containing silver nanoparticles. *Polymer* 2008, 49, 4723-4732.
- [101] Dubey, P.; Bhushan, B.; Sachdev, A.; Matai, I.; Kumar, U.; Packirisamy, G. Silver-Nanoparticle-Incorporated composite nanofibers for potential wound-dressing applications. *J. Appl. Polymer Sci.* 2015, 132.
- [102] Neibert, K.; Gopishetty, V.; Grigoryev, A.; Tokarev, I.; Al-Hajaj, N.; Vorstenbosch, J.; Philip, A.; Minko, S.; Maysinger, D. Wound-Healing with mechanically robust and biodegradable hydrogel fibers loaded with silver nanoparticles. *Adv. Healthc. Mater.* 2012, 1, 621-630.
- [103] Abdelgawad, A.M.; Hudson, S.M.; Rojas, O.J. Antimicrobial wound dressing nanofiber mats from multicomponent (chitosan/silver-NPs/polyvinyl alcohol) systems. *Carbohydr. Polym.* 2014, 100, 166-178.
- [104] El-Aassar, M.R.; Ibrahim, O.M.; Fouda, M.M.G.; El-Beheri, N.G.; Agwa, M.M. Wound healing of nanofiber comprising Polygalacturonic/Hyaluronic acid embedded silver nanoparticles: In-Vitro and in-vivo studies. *Carbohydr. Polym.* 2020, 238, 11. *Pharmaceutics* 2020, 12, 821 23 of 24
- [105] Augustine, R.; Kalarikkal, N.; Thomas, S. Electrospun PCL membranes incorporated with biosynthesized silver nanoparticles as antibacterial wound dressings. *Appl. Nanosci.* 2016, 6, 337-344.
- [106] Uttayarat, P.; Jetawattana, S.; Suwanmala, P.; Eamsiri, J.; Tangthong, T.; Pongpat, S. Antimicrobial electrospun silk fibroin mats with silver nanoparticles for wound dressing application. *Fiber. Polymer.* 2012, 13, 999-1006.
- [107] Dou, J.L.; Zhu, G.D.; Hu, B.; Yang, J.M.; Ge, Y.X.; Li, X.; Liu, J.Y. Wall thickness-tunable AgNPs-NCNTs for hydrogen peroxide sensing and oxygen reduction reaction. *Electrochim. Acta* 2019, 306, 466-476.
- [108] Warren, David S.; Sutherland, Sam P. H.; Kao, Jacqueline Y.; Weal, Geoffrey R.; Mackay, Sean M. (2017-04-20). "The Preparation and Simple Analysis of a Clay Nanoparticle Composite Hydrogel". *Journal of Chemical Education.* 94 (11): 1772-1779. Bibcode: 2017JChEd..94.1772W. doi:10.1021/acs.jchemed.6b00389

- [109] Varaprasad, K.; Yallapu, M.; Ravindra, S.; Nagireddy, N.; Kanikireddy, V.; Monika, K.; Bojja, S.; Raju, K. Hydrogel–Silver nanoparticle composites: A new generation of antimicrobials. *J. Appl. Polymer Sci.* 2010, 115, 1199-1207.
- [110] Kumar, P.T.S.; Abhilash, S.; Manzoor, K.; Nair, S.V.; Tamura, H.; Jayakumar, R. Preparation and characterization of novel  $\beta$ -chitin/nanosilver composite scaffolds for wound dressing applications. *Carbohydr. Polym.* 2010, 80, 761-767.
- [111] Verma, J.; Kanoujia, J.; Parashar, P.; Tripathi, C.B.; Saraf, S.A. Wound healing applications of sericin/chitosan-capped silver nanoparticles incorporated hydrogel. *Drug Deliv. Translat. Res.* 2017, 7, 77-88.
- [112] Boonkaew, B.; Kempf, M.; Kimble, R.; Supaphol, P.; Cuttle, L. Antimicrobial efficacy of a novel silver hydrogel dressing compared to two common silver burn wound dressings: Acticoat. and PolyMem Silver®. *Burns* 2014, 40, 89-96.
- [113] Oliveira, R.N.; Rouze, R.; Quilty, B.; Alves, G.G.; Soares, G.D.; Thire, R.M.; McGuinness, G.B. Mechanical properties and in vitro characterization of polyvinyl alcohol-nano-silver hydrogel wound dressings. *Interface Focus* 2014, 4.
- [114] Rattanuengsrikul, V.; Pimpha, N.; Supaphol, P. In Vitro efficacy and toxicology evaluation of silver nanoparticle-loaded gelatin hydrogel pads as antibacterial wound dressings. *J. Appl. Polymer. Sci.* 2012, 124.
- [115] Das, A.; Kumar, A.; Patil, N.B.; Viswanathan, C.; Ghosh, D. Preparation and characterization of silver nanoparticle loaded amorphous hydrogel of carboxymethylcellulose for infected wounds. *Carbohydr. Polym.* 2015, 130, 254-261.