We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



185,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Renin Angiotensin Aldosterone System Functions in Renovascular Hypertension

Abstract

Jose A. Gomez

The renin angiotensin aldosterone system (RAAS) plays a key function in renovascular hypertension induced by renal artery stenosis (RAS). RAS causes a decrease in renal perfusion in the stenosed kidney which in turn stimulates renin the rate limiting enzyme in RAAS. This stimulation triggers a series of events starting with renin release leading to Ang II production, decrease in sodium excretion, increase sympathetic tone; all contributing to the development of renovascular hypertension. In RAS increase of superoxide reduce nitric oxide in the afferent arteriole increasing vasoconstriction and a marked decrease in glomerular filtration rate. In renovascular hypertension prostaglandins mediate renin release in the stenosed kidney. Targeting different RAAS components is part of the therapy for renovascular hypertension, with other options including renal nerves denervation and revascularization. Different clinical studies had explored revascularization, RAAS blocking and renal nerves denervation as a therapy. We will discuss organ, cellular and molecular components of this disease.

Keywords: Renin angiotensin aldosterone system, renovascular hypertension, renin, renal nerves, oxidative stress

1. Introduction

Renal artery stenosis (RAS) is a common condition in patients suffering from atherosclerosis and fibromuscular dysplasia [1–6], with an overall prevalence disease rate of 15.4% [4]. Progression to severe stenosis is well documented and leads to hypertension and kidney damage [7–9]. Clinically, renovascular hypertension is one the most important causes of secondary hypertension and kidney damage. In patients with RAS, 65% are hypertensive and 26.5% suffer kidney failure [4, 6]. Advancement to end stage renal disease is known to increase cardiovascular events [10]. The clinical trials Angioplasty and Stenting for Renal Artery Lesions (ASTRAL) [11], and Cardiovascular Outcomes in Renal Atherosclerotic Lesions (CORAL) [12] targeted renal vascularization to improve disease outcomes but failed to show any improvement in renal function, cardiovascular events or mortality [11, 12]. Furthermore, prospective studies in ASTRAL and CORAL concluded that 15-22% of patients suffering from renovascular disease will progress to renal "end point" within 3 to 4 years [13]. The NHLBI Cardiovascular Health Study used a non-invasive screen and found that 6.8% elderly patients (both African American

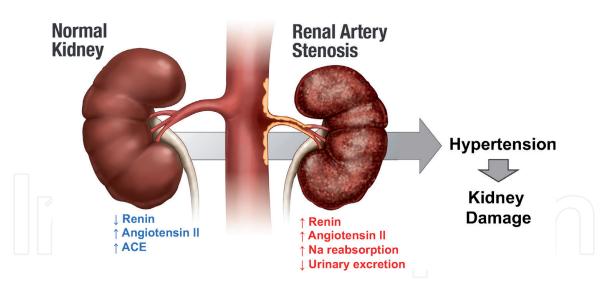


Figure 1.

Renin Angiotensin Aldosterone System (RAAS) key role in renal artery stenosis (RAS) induction of renovascular hypertension and kidney damage. Deterioration of renal perfusion in the stenosed kidney cause a decrease in renal pressure which in turn stimulates RAAS. This stimulation triggers a series of events starting with renin release leading to angiotensin II production; decrease in sodium excretion, increase sympathetic tone; ending in hypertension.

and white) had more than 60% RASten or renal artery occlusion [14, 15]. The renin angiotensin aldosterone system (RAAS) plays a key role in hypertension, with renin recognized as the driver of renovascular hypertension (**Figure 1**). In humans, plasma renin activity (PRA) is used as biomarker for the activation of RAAS in hypertension and in patients with atherosclerotic RAS, high PRA is associated with increased risk for cardiovascular events and high mortality [16]. These suggest an important function for RAAS in renovascular hypertension onset and the need to target different components of RAAS for therapy.

2. Renin angiotensin aldosterone system function in renal artery stenosis

Renal artery stenosis causes a decrease in renal perfusion in the stenosed kidney which in turn stimulates RAAS. This stimulation triggers a series of events starting with renin release leading to angiotensin II (Ang II) production, decrease in sodium excretion, increase sympathetic tone; all contributing to the development of hypertension (**Figure 1**) [17, 18]. When there is a need for renin expression and release, the number of renin expressing cells increase a process known as Juxtaglomerular (JG) cell recruitment [19–24] involving the trans differentiation of vascular smooth muscle cells into renin expressing cells along the afferent arteriole [20, 21, 23]. JG cell recruitment is well documented in this model [25-27]. Activation of the renal baroreceptor in RAS causes renovascular hypertension through RAAS activation [28]. In uni- and bi-lateral RAS aldosterone levels are upregulated [29–32]. Moreover, in renovascular hypertension prostaglandins mediate renin release in the stenosed kidney [33–36], and catecholamines mediated by an increase in cAMP and activation of protein kinase A (PKA) [37–39]. Decrease renal perfusion cause a decline in renal function and increase kidney injury [40, 41]. This decrease in renal function starts with endothelial damage, decrease in nitric oxide and increase in vasoconstrictors and oxidative species [42]. Reactive oxidative stress (ROS) increase renal vascular tone, tubuloglomerular feedback, and endothelial disfunction decreasing glomerular filtration rate [43].

Successful treatments for hypertension such as angiotensin converting enzyme (ACE) inhibitors and angiotensin receptor blockers (ARBs) alleviate hypertension,

but need close examining for kidney failure and hyperkalemia [4]. Aliskiren, a direct renin inhibitor, may still be a potential option for the treatment of high blood pressure in some forms of hypertension such as chronic kidney disease (CKD) and renovascular hypertension [44]. In a clinical study, aliskiren combined with olmesartan reduced proteinuria by about 40% from baseline in patients with CKD with persistent proteinuria [45]. In non-diabetic CKD patients, aliskiren combined with ARBs, safely reduced proteinuria and attenuated the decline in glomerular filtration rate (GFR) [46]. These results indicate that a complete treatment of renal artery stenosis induced renovascular hypertension and kidney damage may need targeting both the angiotensin II-dependent and the Ang II-independent arms of RAAS.

Renal artery stenosis is common in diabetic patients placing them at higher risk of end organ damage causing end stage renal disease [9, 47–49]. In older patients, RAS is the most common problem of end stage renal failure [50]. In RAS renin is recognized as the disease driver [6, 16, 51–54]. RAS is common in atherosclerotic patients and caused hypertension, oxidative stress, and kidney damage [7, 9]. Increased oxidative stress has been reported in humans as well as in two kidney one clip (2K1C) animal model and other hypertensive animal models [24, 55–60]. Changes in renal perfusion activate RAAS and increase the sympathetic activity of the afferent renal nerves contributing to renovascular hypertension and end-stage renal disease during RAS [61]. In the 2K1C model renal denervation decreases hypertension [62, 63]. Clinical trials (Renal Denervation in Patients With Refractory Hypertension (HTN-1) (Symplicity HTN-1), Renal Denervation in Patients With Uncontrolled Hypertension (Symplicity HTN-2), The Renal Denervation for Hypertension (DENERHTN), and Catheter-based renal denervation in patients with uncontrolled hypertension in the absence of antihypertensive medications (SPYRAL)) report that using renal denervation as therapy for hypertension has good outcomes [64–67]. The therapeutic effects of renal denervation have been attributed to removal of sympathetic efferent and/or afferent fibers [68]. Renin secretion is stimulated by renal efferent nerves, which also stimulate tubular sodium reabsorption [62] without perturbations to glomerular filtration rate or albumin urinary secretion [69]. These indicates that initially, renal artery stenosis induces RAAS and in later stages other organs involved in blood pressure homeostasis are involved in the induction of renovascular hypertension such as renal nerves and adrenal gland.

3. Central nervous system input in renal artery stenosis

Different experimental models of hypertension showed the crucial role play by the central nervous system (CNS) in this disease. Specifically, sympathetic efferent outflow augments during hypertension. It has been shown that both Ang II and aldosterone actions are mediated by the CNS [70, 71]. In experimental models of hypertension, ablation of the forebrain surrounding the anteroventral third cerebral ventricle (AV3V) inhibited hypertension [72, 73]. In the CNS the AV3V contains the median preoptic eminence, the organum vasculosum of the lateral terminalis, and the preoptic periventricular nucleus [74]. This forebrain region is responsible for cardiovascular regulation, and includes the subfornical organ, the organum vasculosum of the lamina terminalis, which are circumventricular organs lacking a blood-brain barrier [75]. Production of ROS in these brain regions strongly influences blood pressure [76]. Several reports showed that actions on these brain regions are responsible for Ang II hypertension and increase oxidative stress with NADPH oxidase playing a key role [77–80]. Renal vasculature and tubular segments are controlled by the efferent sympathetic renal nerves and promote arteriolar vasoconstriction and renin release and increases sodium reabsorption [81]. In the afferent arterioles Ang II activates the alpha1 adrenergic receptor, which increases oxidative stress and constriction of the afferent arterioles, reducing renal blood flow [82]. Contrary, activation of the b1-adrenergic receptor activation inhibits ROS generation promoting vasodilation [83]. In different hypertension animal models renal denervation inhibit the induction of hypertension, showing that ablation of renal efferent induction of ROS is important in hypertension development [84, 85]. These data indicate that oxidative stress control efferent and afferent renal nerve actions in the development of hypertension.

Renal artery stenosis activates RAAS and increases the activity of the afferent renal nerves resulting in hypertension and end-stage renal disease [61]. It is known that in the 2K1C model renal denervation decreases hypertension [62, 63]. Removal of sympathetic efferent and/or afferent fibers controls hypertension [68], and the renal efferent nerves stimulate renin secretion and tubular sodium reabsorption [62]. During renal artery stenosis, there is an increase in Neutrophil Oxidase Factor p47 (p47phox) and p67phox [86–88]. Furthermore, in renal artery stenosis generation of ROS induced renal damage [88, 89], with the main source of ROS being NADPH oxidase [90, 91].

In the induction of renovascular hypertension, the renal nerves as well as the renin angiotensin aldosterone system activation cause the increase in blood pressure and dysregulation of sodium secretion, with renal denervation alleviating the central nerve system input decreasing blood pressure.

4. Oxidative stress in renal artery stenosis

Oxidative stress in the kidney and vasculature contribute to hypertension development. NADPH oxidase is a major source of oxidative stress in mammalian cells [75]. Most of the renal cells express NADPH oxidase and there are several stimuli that cause its activation leading to organ injury and hypertension development [75, 92, 93]. Reactive oxygen species (ROS) produced by NADPH oxidase in the kidney cause vasoconstriction and organ injury. Specifically, increase of superoxide reduces nitric oxide (NO) in the afferent arteriole increasing vasoconstriction and a marked decrease in GFR. In rabbits, Ang II-induced hypertension increase the p22phox subunit of NADPH oxidase causing endothelial dysfunction in the afferent arteriole [94]. Moreover, in spontaneous hypertensive rats, superoxide is generated in the afferent arteriole in response to endothelin-1 (ET-1) [95, 96]. Podocytes are important components of the renal filtration system. Dahl salt-sensitive rats had increase glomerular expression of p22phos and NOX2 that increases oxidative stress causing podocyte injury, glomerular sclerosis and proteinuria, with the antioxidant tempol (4-Hydroxy-TEMPO) correcting this glomerular injury [97, 98]. Plasminogen causes podocyte injury through stimulation of NOX2 and NOX4 expression [99], Ang II stimulates ROS generation in the mitochondria stimulating autophagy [100], Ang II-induced ROS production caused glomerulosclerosis [101], and oxidative stress disrupts nephrin – caveolin-1 crosstalk in podocytes disrupting of glomerular filtration barrier [102]. In the vasculature, increased oxidative stress causes hypertension in different animal models [103–108]. During renal artery stenosis, generation of ROS is recognized as the main mechanism of renal damage [88, 89, 109, 110] with the activation of NADPH oxidase as the source of ROS [90, 91], and associated with an increase in p47phox and p67phox [19, 86–88].

It is important to recognize that renal artery stenosis increase the production of reactive oxygen species leading to renal damage. ROS production influences not only organ damage but also contributes to the increase in blood pressure. In the therapy of this disease multiple molecules are involved leading to increases in oxidative stress, blood pressure and renal injury and all start with the activation of the renin angiotensin aldosterone system.

5. Angiotensin II dependent and independent action in renal artery stenosis

In renal artery stenosis induction of renovascular hypertension, renin is recognized a key molecule, and as such in the therapy of renovascular hypertension Angiotensin Converting Enzyme (ACE) inhibitors and Angiotensin Receptor blockers (ARBs) are used [4]. Moreover, sympathetic nervous systems action in the kidney promotes renin secretion through renal efferent nerves, which also stimulate tubular sodium reabsorption [62], and in the 2K1C model denervation inhibit the onset of hypertension [62, 63]. Renal artery stenosis causes renovascular hypertension, which is associated with deterioration of kidney function [20]. Reduction in renal flow is recognize as a source of hypoxia during renovascular hypertension [21]. Arterial stenosis causes thrombosis, and ischemia in renovascular hypertension [22]. During renal artery stenosis generation of ROS is recognized as the main mechanism of renal damage [88, 89], causing increased in vasoconstrictors, cell death and decrease in the activity of nitric oxide [109, 110]. A swine model of renal artery stenosis presented an increase in ROS, renal and cardiac damage [23, 86–89, 111–113]. In renal artery stenosis activation of RAAS increase ROS generating by the activation of NADPH oxidase [90, 91], associated with is an increase in p47phox and p67phox [86–88]. Phosphorylation of p47phox by PKC is a key step in NADPH oxidase activation [114–118]. Hypertension is associated with PKC activation and increase oxidative stress [119], which caused endothelial nitric oxide synthase (eNOS) disfunction and uncoupling producing ROS instead of NO. This uncoupling is a key mechanism for endothelial dysfunction in angiotensin II-induced hypertension [120–122]. Increase in NOX2 activity requires increase NOX2 expression and p47phox association and activation of NOX2 [19]. Furthermore, increase in oxidative stress is well documented in 2K1C model [55–59, 123, 124]. All the actions mentioned above are Ang II mediated.

New evidence places (pro)renin receptor (PRR) as an effector molecule in the Ang II-independent RAAS [125]. PRR binds both renin and prorenin [125–129]. There is an association of PRR with different pathophysiology of diseases [130–135]. PRR binds renin causing an increase in Ang I [125] and it can activate prorenin by promoting a conformational change [125–129]. PRR mRNA is expressed in different organs such as kidney, heart, brain, eye, adipose tissue and vascular SMCs [125, 134], It has been proposed that PRR activates the Ang II-independent RAAS with tissue specificity [136]. My laboratory and others are uncovering new functions of the Ang II independent pathway in blood pressure, oxidative stress and organ damage. New studies will define the relevance of this arm of RAAS and possible define new molecular targets for therapy.

6. Concluding remarks and future perspectives

In the definition of the molecular pathways involved in the development of renovascular hypertension, the Goldblatt two kidney one clip animal model has been critical. This animal mode has been extensively used with different animals all showing that renal artery stenosis strongly stimulates renin overexpression and release promoting renovascular hypertensions and kidney injury. In renovascular hypertension renin is key and promotes the increase in Ang II leading to hypertension. Renin being the rate limiting step in the production of Ang II in RAAS, has been investigated as a possible target for the therapy. However, the main therapies used are angiotensin converting enzyme inhibitors and angiotensin receptor blockers. Direct renin inhibition by aliskiren, is potential therapy for hypertension in chronic kidney disease (CKD) and renovascular hypertension. Combination of aliskiren with olmesartan in the clinic, reduced proteinuria in patients with CKD with persistent proteinuria. In non-diabetic CKD patients, aliskiren combined with ARBs, reduced proteinuria and protected from the decline in glomerular filtration rate. We have shown here clinical and research data that indicates the during renal artery stenosis induced renovascular hypertension RAAS is activated and play a critical role in this pathology. It is important that a complete treatment of renovascular hypertension may need targeting both the angiotensin II-dependent and the Ang II-independent arms of RAAS.

Acknowledgements

We apologize to colleagues whose work has not been included in this chapter due to space limitations. This work was supported by NIH Grants: NHLBI Research Scientist Development Grant (1K01HL135461).

Intechopen

Author details

Jose A. Gomez Department of Medicine, Clinical Pharmacology Division, Vanderbilt University Medical Center, Nashville, TN, United States

*Address all correspondence to: jose.a.gomez@vumc.org

IntechOpen

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Jean WJ, al-Bitar I, Zwicke DL, Port SC, Schmidt DH, Bajwa TK. High incidence of renal artery stenosis in patients with coronary artery disease. Cathet Cardiovasc Diagn 1994;32:8-10.

[2] Olin JW, Melia M, Young JR, Graor RA, Risius B. Prevalence of atherosclerotic renal artery stenosis in patients with atherosclerosis elsewhere. Am J Med 1990;88:46N-51N.

[3] Silva JA. Evaluation and approach to treatment of renal artery stenosis in patients with diabetic nephropathy. Curr Diab Rep 2008;8:494-8.

[4] de Mast Q, Beutler JJ. The prevalence of atherosclerotic renal artery stenosis in risk groups: a systematic literature review. J Hypertens 2009;27:1333-40.

[5] Cooper EL, Xie Y, Nguyen H et al. Early Rapid Decline in Kidney Function in Medically Managed Patients With Atherosclerotic Renal Artery Stenosis. J Am Heart Assoc 2019;8:e012366.

[6] Textor SC, Lerman L. Renovascular hypertension and ischemic nephropathy. Am J Hypertens 2010;23:1159-69.

[7] Schreiber MJ, Pohl MA, Novick AC. The natural history of atherosclerotic and fibrous renal artery disease. Urol Clin North Am 1984;11:383-92.

[8] Crowley JJ, Santos RM, Peter RH et al. Progression of renal artery stenosis in patients undergoing cardiac catheterization. Am Heart J 1998;136:913-8.

[9] Caps MT, Perissinotto C, Zierler RE et al. Prospective study of atherosclerotic disease progression in the renal artery. Circulation 1998;98:2866-72.

[10] Al-Suraih M, Grande JP. Management of renal artery stenosis: What does the experimental evidence tell us? World J Cardiol 2014;6:855-60. [11] Investigators A, Wheatley K, Ives N et al. Revascularization versus medical therapy for renal-artery stenosis. N Engl J Med 2009;361:1953-62.

[12] Cooper CJ, Murphy TP, Cutlip DE et al. Stenting and medical therapy for atherosclerotic renal-artery stenosis. N Engl J Med 2014;370:13-22.

[13] Textor SC. Renal Arterial Disease and Hypertension. Med Clin North Am 2017;101:65-79.

[14] Hansen KJ, Edwards MS, Craven TE et al. Prevalence of renovascular disease in the elderly: a population-based study. J Vasc Surg 2002;36:443-51.

[15] Weber BR, Dieter RS. Renal artery stenosis: epidemiology and treatment. Int J Nephrol Renovasc Dis 2014;7:169-81.

[16] Daidoji H, Tamada Y, Suzuki S et al. Plasma Renin Activity Predicts the Improvement in Resistant Hypertension after Percutaneous Transluminal Renal Artery Angioplasty. Intern Med 2016;55:3421-3426.

[17] An epidemiological approach to describing risk associated with blood pressure levels. Final report of the Working Group on Risk and High Blood Pressure. Hypertension 1985;7:641-51.

[18] Border WA, Noble NA. Interactions of transforming growth factor-beta and angiotensin II in renal fibrosis. Hypertension 1998;31:181-8.

[19] Brandes RP, Weissmann N, Schroder K. Nox family NADPH oxidases: Molecular mechanisms of activation. Free Radic Biol Med 2014;76:208-26.

[20] Plouin PF, Rossignol P, Bobrie G. Atherosclerotic renal artery stenosis:

to treat conservatively, to dilate, to stent, or to operate? J Am Soc Nephrol 2001;12:2190-6.

[21] Liu J, Wei Q, Guo C et al. Hypoxia, HIF, and Associated Signaling Networks in Chronic Kidney Disease. Int J Mol Sci 2017;18.

[22] Yu W, Liu-Bryan R, Stevens S, Damanahalli JK, Terkeltaub R. RAGE signaling mediates post-injury arterial neointima formation by suppression of liver kinase B1 and AMPK activity. Atherosclerosis 2012;222:417-25.

[23] Rajagopalan S, Duquaine D, King S, Pitt B, Patel P. Mineralocorticoid receptor antagonism in experimental atherosclerosis. Circulation 2002;105:2212-6.

[24] Saleem M., Saavedra-Sánchez L., Barturen-Larrea P., and Gomez J.A. The transcription factor Sox6 controls renin expression during renal artery stenosis. Kidney360 March 2021, 10.34067/ KID.0002792020; DOI: https://doi. org/10.34067/KID.0002792020.

[25] Tomanek RJ, Schalk KA, Marcus ML, Harrison DG. Coronary angiogenesis during long-term hypertension and left ventricular hypertrophy in dogs. Circ Res 1989; 65:352-9.

[26] Braam B, Navar LG, Mitchell KD. Modulation of tubuloglomerular feedback by angiotensin II type 1 receptors during the development of Goldblatt hypertension. Hypertension 1995;25:1232-7.

[27] Gollan F, Richardson E, Goldblatt H. Hypertension in the systemic blood of animals with experimental renal hypertension. J Exp Med 1948;88: 389-400.

[28] Reinhold SW, Uihlein DC, Boger CA et al. Renin, endothelial NO synthase

and endothelin gene expression in the 2kidney-1clip Goldblatt model of long-term renovascular hypertension. Eur J Med Res 2009;14:520-5.

[29] Kotliar C, Inserra F, Forcada P et al. Are plasma renin activity and aldosterone levels useful as a screening test to differentiate between unilateral and bilateral renal artery stenosis in hypertensive patients? J Hypertens 2010;28:594-601.

[30] Maxwell MH, Lupu AN, Viskoper RJ, Aravena LA, Waks UA. Mechanisms of hypertension during the acute and intermediate phases of the one-clip, two-kidney model in the dog. Circ Res 1977;40:I24-8.

[31] McAreavey D, Brown JJ, Cumming AM et al. Inverse relation of exchangeable sodium and blood pressure in hypertensive patients with renal artery stenosis. J Hypertens 1983;1:297-302.

[32] Takabatake T, Ohta H, Yamamoto Y et al. Effect of angiotensin blockade and converting enzyme inhibition on renovascular hypertension: comparison between unilateral and bilateral renal artery stenosis. Angiology 1987;38: 434-9.

[33] Imanishi M, Kawamura M, Akabane S et al. Aspirin lowers blood pressure in patients with renovascular hypertension. Hypertension 1989;14:461-8.

[34] Schricker K, Hamann M, Kaissling B, Kurtz A. Role of the macula densa in the control of renal renin gene expression in two-kidney/one-clip rats. Pflugers Arch 1994;427:42-6.

[35] Tuttle KR. Charting new territory by simulated modeling of a clinical trial. Clin J Am Soc Nephrol 2010;5:750-2.

[36] Wang JL, Cheng HF, Harris RC. Cyclooxygenase-2 inhibition decreases

renin content and lowers blood pressure in a model of renovascular hypertension. Hypertension 1999;34:96-101.

[37] Friis UG, Jensen BL, Sethi S, Andreasen D, Hansen PB, Skott O. Control of renin secretion from rat juxtaglomerular cells by cAMP-specific phosphodiesterases. Circ Res 2002;90: 996-1003.

[38] Schramm A, Schweda F, Sequeira-Lopez MLS, Hofmann F, Sandner P, Schlossmann J. Protein Kinase G Is Involved in Acute but Not in Long-Term Regulation of Renin Secretion. Front Pharmacol 2019;10:800.

[39] Sparks MA, Crowley SD, Gurley SB, Mirotsou M, Coffman TM. Classical Renin-Angiotensin system in kidney physiology. Compr Physiol 2014;4: 1201-28.

[40] Fava C, Minuz P, Patrignani P, Morganti A. Renal artery stenosis and accelerated atherosclerosis: which comes first? J Hypertens 2006;24:1687-96.

[41] Chade AR, Lerman A, Lerman LO. Kidney in early atherosclerosis. Hypertension 2005;45:1042-9.

[42] Brodsky SV, Yamamoto T, Tada T et al. Endothelial dysfunction in ischemic acute renal failure: rescue by transplanted endothelial cells. Am J Physiol Renal Physiol 2002;282:F1140-9.

[43] Schnackenberg CG. Physiological and pathophysiological roles of oxygen radicals in the renal microvasculature. Am J Physiol Regul Integr Comp Physiol 2002;282:R335-42.

[44] Morisawa N, Sugano N, Yamakawa T, Kuriyama S, Yokoo T. Successful long-term effects of direct renin inhibitor aliskiren in a patient with atherosclerotic renovascular hypertension. CEN Case Rep 2017;6:66-73. [45] Moriyama T, Tsuruta Y, Kojima C et al. Beneficial effect of aliskiren combined with olmesartan in reducing urinary protein excretion in patients with chronic kidney disease. Int Urol Nephrol 2012;44:841-5.

[46] Li SY, Chen YT, Yang WC et al. Effect of add-on direct renin inhibitor aliskiren in patients with non-diabetes related chronic kidney disease. BMC Nephrol 2012;13:89.

[47] Bryfogle JW, Bradley RF. The vascular complications of diabetes mellitus; a clinical study. Diabetes 1957;6:159-67.

[48] Courreges JP, Bacha J, Aboud E, Pradier P. Prevalence of renal artery stenosis in type 2 diabetes. Diabetes Metab 2000;26 Suppl 4:90-6.

[49] Fatica RA, Port FK, Young EW. Incidence trends and mortality in end-stage renal disease attributed to renovascular disease in the United States. Am J Kidney Dis 2001;37:1184-90.

[50] Mailloux LU, Bellucci AG, Mossey RT et al. Predictors of survival in patients undergoing dialysis. Am J Med 1988;84:855-62.

[51] Covic A, Gusbeth-Tatomir P. The role of the renin-angiotensinaldosterone system in renal artery stenosis, renovascular hypertension, and ischemic nephropathy: diagnostic implications. Prog Cardiovasc Dis 2009;52:204-8.

[52] Safian RD, Textor SC. Renal-artery stenosis. N Engl J Med 2001;344:431-42.

[53] Postma CT, van Oijen AH, Barentsz JO et al. The value of tests predicting renovascular hypertension in patients with renal artery stenosis treated by angioplasty. Arch Intern Med 1991;151:1531-5. [54] Gloviczki ML, Glockner JF, Lerman LO et al. Preserved oxygenation despite reduced blood flow in poststenotic kidneys in human atherosclerotic renal artery stenosis. Hypertension 2010;55:961-6.

[55] Chen K, Xie F, Liu S et al. Plasma reactive carbonyl species: Potential risk factor for hypertension. Free Radic Res 2011;45:568-74.

[56] Wang W, Saad A, Herrmann SM et al. Changes in inflammatory biomarkers after renal revascularization in atherosclerotic renal artery stenosis. Nephrol Dial Transplant 2016;31:1437-43.

[57] Kinra M, Mudgal J, Arora D, Nampoothiri M. An insight into the role of cyclooxygenase and lipooxygenase pathway in renal ischemia. Eur Rev Med Pharmacol Sci 2017;21:5017-5020.

[58] Dias AT, Rodrigues BP, Porto ML et al. Sildenafil ameliorates oxidative stress and DNA damage in the stenotic kidneys in mice with renovascular hypertension. J Transl Med 2014;12:35.

[59] Zhang X, Eirin A, Li ZL et al. Angiotensin receptor blockade has protective effects on the poststenotic porcine kidney. Kidney Int 2013;84:767-75.

[60] Reckelhoff JF, Romero DG, Yanes Cardozo LL. Sex, Oxidative Stress, and Hypertension: Insights From Animal Models. Physiology (Bethesda) 2019;34:178-188.

[61] Grisk O, Rettig R. Interactions between the sympathetic nervous system and the kidneys in arterial hypertension. Cardiovasc Res 2004;61:238-46.

[62] DiBona GF, Kopp UC. Neural control of renal function. Physiol Rev 1997;77:75-197.

[63] Osborn JW, Foss JD. Renal Nerves and Long-Term Control of Arterial Pressure. Compr Physiol 2017;7:263-320.

[64] Azizi M, Sapoval M, Gosse P et al. Optimum and stepped care standardised antihypertensive treatment with or without renal denervation for resistant hypertension (DENERHTN): a multicentre, open-label, randomised controlled trial. Lancet 2015;385:1957-65.

[65] Krum H, Schlaich M, Whitbourn R et al. Catheter-based renal sympathetic denervation for resistant hypertension: a multicentre safety and proof-ofprinciple cohort study. Lancet 2009;373:1275-81.

[66] Krum H, Schlaich MP, Sobotka PA et al. Percutaneous renal denervation in patients with treatment-resistant hypertension: final 3-year report of the Symplicity HTN-1 study. Lancet 2014;383:622-9.

[67] Townsend RR, Mahfoud F, Kandzari DE et al. Catheter-based renal denervation in patients with uncontrolled hypertension in the absence of antihypertensive medications (SPYRAL HTN-OFF MED): a randomised, sham-controlled, proof-ofconcept trial. Lancet 2017;390:2160-2170.

[68] Osborn JW, Banek CT. Catheter-Based Renal Nerve Ablation as a Novel Hypertension Therapy: Lost, and Then Found, in Translation. Hypertension 2018;71:383-388.

[69] Mahfoud F, Cremers B, Janker J et al. Renal hemodynamics and renal function after catheter-based renal sympathetic denervation in patients with resistant hypertension. Hypertension 2012;60:419-24.

[70] Guyenet PG. The sympathetic control of blood pressure. Nat Rev Neurosci 2006;7:335-46.

[71] Peterson JR, Sharma RV, Davisson RL. Reactive oxygen species in

the neuropathogenesis of hypertension. Curr Hypertens Rep 2006;8:232-41.

[72] Brody MJ. Central nervous system and mechanisms of hypertension. Clin Physiol Biochem 1988;6:230-9.

[73] Gordon FJ, Haywood JR, Brody MJ, Johnson AK. Effect of lesions of the anteroventral third ventricle (AV3V) on the development of hypertension in spontaneously hypertensive rats. Hypertension 1982;4:387-93.

[74] Whyte DG, Johnson AK. Thermoregulatory role of periventricular tissue surrounding the anteroventral third ventricle (AV3V) during acute heat stress in the rat. Clin Exp Pharmacol Physiol 2005;32:457-61.

[75] Loperena R, Harrison DG. Oxidative Stress and Hypertensive Diseases. Med Clin North Am 2017;101:169-193.

[76] Zimmerman MC, Lazartigues E, Lang JA et al. Superoxide mediates the actions of angiotensin II in the central nervous system. Circ Res 2002;91:1038-45.

[77] Lob HE, Schultz D, Marvar PJ, Davisson RL, Harrison DG. Role of the NADPH oxidases in the subfornical organ in angiotensin II-induced hypertension. Hypertension 2013;61:382-7.

[78] Nagashima Y, Ohaki Y, Tanaka Y et al. Establishment of an epithelioid malignant schwannoma cell line (YST-1). Virchows Arch B Cell Pathol Incl Mol Pathol 1990;59:321-7.

[79] Zimmerman MC, Lazartigues E, Sharma RV, Davisson RL. Hypertension caused by angiotensin II infusion involves increased superoxide production in the central nervous system. Circ Res 2004;95:210-6.

[80] Zimmerman MC, Sharma RV, Davisson RL. Superoxide mediates angiotensin II-induced influx of extracellular calcium in neural cells. Hypertension 2005;45:717-23.

[81] Grassi G, Seravalle G, Brambilla G, Mancia G. The sympathetic nervous system and new nonpharmacologic approaches to treating hypertension: a focus on renal denervation. Can J Cardiol 2012;28:311-7.

[82] Wang D, Jose P, Wilcox CS. beta(1) Receptors protect the renal afferent arteriole of angiotensin-infused rabbits from norepinephrine-induced oxidative stress. J Am Soc Nephrol 2006;17:3347-54.

[83] Boivin V, Jahns R, Gambaryan S, Ness W, Boege F, Lohse MJ.
Immunofluorescent imaging of beta
1- and beta 2-adrenergic receptors in rat kidney. Kidney Int 2001;59:515-31.

[84] Campese VM, Ye S. A vitamin-Efortified diet reduces oxidative stress, sympathetic nerve activity, and hypertension in the phenol-renal injury model in rats. J Am Soc Hypertens 2007;1:242-50.

[85] DiBona GF, Esler M. Translational medicine: the antihypertensive effect of renal denervation. 2010;298:R245-53.

[86] Chade AR, Krier JD, Rodriguez-Porcel M et al. Comparison of acute and chronic antioxidant interventions in experimental renovascular disease. Am J Physiol Renal Physiol 2004;286:F1079-86.

[87] Chade AR, Rodriguez-Porcel M, Herrmann J et al. Antioxidant intervention blunts renal injury in experimental renovascular disease. J Am Soc Nephrol 2004;15:958-66.

[88] Chade AR, Rodriguez-Porcel M, Grande JP et al. Distinct renal injury in early atherosclerosis and renovascular disease. Circulation 2002;106:1165-71.

[89] Chade AR, Rodriguez-Porcel M, Grande JP et al. Mechanisms of renal

structural alterations in combined hypercholesterolemia and renal artery stenosis. Arterioscler Thromb Vasc Biol 2003;23:1295-301.

[90] Mazzolai L, Duchosal MA, Korber M et al. Endogenous angiotensin II induces atherosclerotic plaque vulnerability and elicits a Th1 response in ApoE-/- mice. Hypertension 2004;44:277-82.

[91] Welch WJ, Mendonca M, Aslam S, Wilcox CS. Roles of oxidative stress and AT1 receptors in renal hemodynamics and oxygenation in the postclipped 2K,1C kidney. Hypertension 2003;41:692-6.

[92] Cuevas S, Zhang Y, Yang Y et al. Role of renal DJ-1 in the pathogenesis of hypertension associated with increased reactive oxygen species production. Hypertension 2012;59:446-52.

[93] Liu R, Ren Y, Garvin JL, Carretero OA. Superoxide enhances tubuloglomerular feedback by constricting the afferent arteriole. Kidney Int 2004;66:268-74.

[94] Wang D, Chen Y, Chabrashvili T et al. Role of oxidative stress in endothelial dysfunction and enhanced responses to angiotensin II of afferent arterioles from rabbits infused with angiotensin II. J Am Soc Nephrol 2003;14:2783-9.

[95] Araujo M, Wilcox CS. Oxidative stress in hypertension: role of the kidney. Antioxid Redox Signal 2014;20:74-101.

[96] Chabrashvili T, Kitiyakara C, Blau J et al. Effects of ANG II type 1 and 2 receptors on oxidative stress, renal NADPH oxidase, and SOD expression. 2003;285:R117-24.

[97] Meng S, Cason GW, Gannon AW, Racusen LC, Manning RD, Jr. Oxidative stress in Dahl salt-sensitive hypertension. Hypertension 2003;41:1346-52. [98] Nagase M, Shibata S, Yoshida S, Nagase T, Gotoda T, Fujita T. Podocyte injury underlies the glomerulopathy of Dahl salt-hypertensive rats and is reversed by aldosterone blocker. Hypertension 2006;47:1084-93.

[99] Raij L, Tian R, Wong JS, He JC, Campbell KN. Podocyte injury: the role of proteinuria, urinary plasminogen, and oxidative stress. Am J Physiol Renal Physiol 2016;311:F1308-F1317.

[100] Jia J, Ding G, Zhu J et al. Angiotensin II infusion induces nephrin expression changes and podocyte apoptosis. Am J Nephrol 2008;28:500-7.

[101] Hua P, Feng W, Rezonzew G, Chumley P, Jaimes EA. The transcription factor ETS-1 regulates angiotensin II-stimulated fibronectin production in mesangial cells. Am J Physiol Renal Physiol 2012;302:F1418-29.

[102] Ren Z, Liang W, Chen C, Yang H, Singhal PC, Ding G. Angiotensin II induces nephrin dephosphorylation and podocyte injury: role of caveolin-1. Cell Signal 2012;24:443-50.

[103] Beswick RA, Dorrance AM, Leite R, Webb RC. NADH/NADPH oxidase and enhanced superoxide production in the mineralocorticoid hypertensive rat. Hypertension 2001;38:1107-11.

[104] Cai H, Li Z, Dikalov S et al. NAD(P)H oxidase-derived hydrogen peroxide mediates endothelial nitric oxide production in response to angiotensin II. J Biol Chem 2002;277: 48311-7.

[105] Landmesser U, Cai H, Dikalov S et al. Role of p47(phox) in vascular oxidative stress and hypertension caused by angiotensin II. Hypertension 2002;40:511-5.

[106] Suzuki H, Swei A, Zweifach BW, Schmid-Schonbein GW. In vivo

evidence for microvascular oxidative stress in spontaneously hypertensive rats. Hydroethidine microfluorography. Hypertension 1995;25:1083-9.

[107] Swei A, Lacy F, DeLano FA, Schmid-Schonbein GW. Oxidative stress in the Dahl hypertensive rat. Hypertension 1997;30:1628-33.

[108] Zhou X, Bohlen HG, Miller SJ, Unthank JL. NAD(P)H oxidase-derived peroxide mediates elevated basal and impaired flow-induced NO production in SHR mesenteric arteries in vivo. 2008;295:H1008-H1016.

[109] Lerman L, Textor SC. Pathophysiology of ischemic nephropathy. Urol Clin North Am 2001;28:793-803, ix.

[110] Napoli C, de Nigris F, Palinski W. Multiple role of reactive oxygen species in the arterial wall. J Cell Biochem 2001;82:674-82.

[111] Rodriguez-Porcel M, Lerman A, Herrmann J et al. Hypertension exacerbates the effect of hypercholesterolemia on the myocardial microvasculature. Cardiovasc Res 2003;58:213-21.

[112] Rodriguez-Porcel M, Krier JD, Lerman A et al. Combination of hypercholesterolemia and hypertension augments renal function abnormalities. Hypertension 2001;37:774-80.

[113] Rodriguez-Porcel M, Herrman J, Chade AR et al. Long-term antioxidant intervention improves myocardial microvascular function in experimental hypertension. Hypertension 2004;43:493-8.

[114] Benna JE, Dang PM, Gaudry M et al. Phosphorylation of the respiratory burst oxidase subunit p67(phox) during human neutrophil activation. Regulation by protein kinase C-dependent and independent pathways. J Biol Chem 1997;272:17204-8.

[115] Bouin AP, Grandvaux N, Vignais PV, Fuchs A. p40(phox) is phosphorylated on threonine 154 and serine 315 during activation of the phagocyte NADPH oxidase. Implication of a protein kinase c-type kinase in the phosphorylation process. J Biol Chem 1998;273:30097-103.

[116] el Benna J, Faust LP, Babior BM. The phosphorylation of the respiratory burst oxidase component p47phox during neutrophil activation. Phosphorylation of sites recognized by protein kinase C and by proline-directed kinases. J Biol Chem 1994;269:23431-6.

[117] El Benna J, Faust RP, Johnson JL, Babior BM. Phosphorylation of the respiratory burst oxidase subunit p47phox as determined by twodimensional phosphopeptide mapping. Phosphorylation by protein kinase C, protein kinase A, and a mitogenactivated protein kinase. J Biol Chem 1996;271:6374-8.

[118] Faust LR, el Benna J, Babior BM, Chanock SJ. The phosphorylation targets of p47phox, a subunit of the respiratory burst oxidase. Functions of the individual target serines as evaluated by site-directed mutagenesis. J Clin Invest 1995;96:1499-505.

[119] Li H, Wallerath T, Munzel T, Forstermann U. Regulation of endothelial-type NO synthase expression in pathophysiology and in response to drugs. Nitric Oxide 2002;7:149-64.

[120] Hink U, Li H, Mollnau H et al. Mechanisms underlying endothelial dysfunction in diabetes mellitus. Circ Res 2001;88:E14-22.

[121] Mollnau H, Wendt M, Szocs K et al. Effects of angiotensin II infusion on the expression and function of NAD(P)H oxidase and components of nitric oxide/ cGMP signaling. Circ Res 2002; 90:E58-65.

[122] Munzel T, Li H, Mollnau H et al. Effects of long-term nitroglycerin treatment on endothelial nitric oxide synthase (NOS III) gene expression, NOS III-mediated superoxide production, and vascular NO bioavailability. Circ Res 2000;86:E7-E12.

[123] Zou X, Kwon SH, Jiang K et al. Renal scattered tubular-like cells confer protective effects in the stenotic murine kidney mediated by release of extracellular vesicles. Sci Rep 2018;8:1263.

[124] Cavalcanti CO, Alves RR, de Oliveira AL et al. Inhibition of PDE5 Restores Depressed Baroreflex Sensitivity in Renovascular Hypertensive Rats. Front Physiol 2016;7:15.

[125] Nguyen G, Delarue F, Burckle C, Bouzhir L, Giller T, Sraer JD. Pivotal role of the renin/prorenin receptor in angiotensin II production and cellular responses to renin. J Clin Invest 2002;109:1417-27.

[126] Batenburg WW, Krop M, Garrelds IM et al. Prorenin is the endogenous agonist of the (pro)renin receptor. Binding kinetics of renin and prorenin in rat vascular smooth muscle cells overexpressing the human (pro)renin receptor. J Hypertens 2007;25:2441-53.

[127] Nabi AH, Kageshima A, Uddin MN, Nakagawa T, Park EY, Suzuki F. Binding properties of rat prorenin and renin to the recombinant rat renin/prorenin receptor prepared by a baculovirus expression system. Int J Mol Med 2006;18:483-8.

[128] Nurun NA, Uddin NM, Nakagawa T et al. Role of "handle" region of prorenin prosegment in the nonproteolytic activation of prorenin by binding to membrane anchored (pro) renin receptor. Front Biosci 2007;12:4810-7.

[129] Zhang J, Noble NA, Border WA, Owens RT, Huang Y. Receptordependent prorenin activation and induction of PAI-1 expression in vascular smooth muscle cells. Am J Physiol Endocrinol Metab 2008;295:E810-9.

[130] Burckle CA, Jan Danser AH, Muller DN et al. Elevated blood pressure and heart rate in human renin receptor transgenic rats. Hypertension 2006;47:552-6.

[131] Ichihara A, Hayashi M, Kaneshiro Y et al. Inhibition of diabetic nephropathy by a decoy peptide corresponding to the "handle" region for nonproteolytic activation of prorenin. J Clin Invest 2004;114:1128-35.

[132] Ichihara A, Sakoda M, Kurauchi-Mito A, Kaneshiro Y, Itoh H. Involvement of (pro)renin receptor in the glomerular filtration barrier. J Mol Med (Berl) 2008;86:629-35.

[133] Ichihara A, Sakoda M, Kurauchi-Mito A, Kaneshiro Y, Itoh H. Renin, prorenin and the kidney: a new chapter in an old saga. J Nephrol 2009;22:306-11.

[134] Ichihara A, Sakoda M, Mito-Kurauchi A, Itoh H. Activated prorenin as a therapeutic target for diabetic nephropathy. Diabetes Res Clin Pract 2008;82 Suppl 1:S63-6.

[135] Kaneshiro Y, Ichihara A, Sakoda M et al. Slowly progressive, angiotensin II-independent glomerulosclerosis in human (pro)renin receptor-transgenic rats. J Am Soc Nephrol 2007;18:1789-95.

[136] Nabi AH, Suzuki F. Biochemical properties of renin and prorenin binding to the (pro)renin receptor. Hypertens Res 2010;33:91-7.