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Valorization Options of Strawberry Extrudate Agro-Waste. A Review

Juan Cubero-Cardoso, Antonio Serrano, Ángeles Trujillo-Reyes, Denys K. Villa-Gómez, Rafael Borja and Fernando G. Feroso

Abstract

This review summarizes and critically analyzes the different types of potential valorization options for strawberry extrudate in order to have a broader overview of the potential management of this waste. Animal feed is commonly used as a management option for the strawberry extrudate; however, most of the strawberry extrudate is disposed in landfills. Strawberry extrudate contains different bioactive compounds that encourage the use of an alternative management approach than landfilled. The present review offers a complete comparative, including the advantages and drawbacks of each reviewed technique, to facilitate the selection of the most suitable technology for the different valorization scenarios. This review has been structured in three sections: 1. Composition of the strawberry extrudate and strawberry especially focused on their content in bioactive compounds. 2. The different techniques of extraction and purification of bioactive compounds. 3. The handling and management of the resulting biomass after the extraction process of bioactive compounds.

Keywords: strawberry extrudate, bioactive compounds, bioproducts, extraction techniques, purification

1. Introduction

In 2016, 8 million tons of strawberry were produced in the world with a value of agricultural gross production of 17,739 million US\$ [1]. Besides its market as fresh product, strawberry is also used to produce many types of by-products, due to its peculiar flavor and aroma. Strawberry by-products are mainly formulated from a strawberry concentrate. The most common technology to obtain the strawberry concentrate is by extrusion. Strawberries are extruded by twin-screws up to several sieves with different mesh sizes. The sieves retain a residual fraction formed by the fibrous part and the achenes, named strawberry extrudate, which accounts about 7% of the manufactured strawberry [2].

Animal feed is commonly used as a management option for the strawberry extrudate, however, most of the strawberry extrudate is disposed in landfills, contributing to greenhouse emissions due to its high organic load [3]. Alternatives for strawberry extrudate management are required to avoid severe environmental

impacts that cause landfills, such as negative effects on agricultural soil quality, polluting of aquatic ecosystems and atmospheric contamination [4].

Similar to strawberry, strawberry extrudate contains substances of high interest such as bioactive compounds. Some of these bioactive compounds have beneficial health effects on cardiovascular, neurological or cancerous disorders [5]. Due to their health benefits, bioactive compounds have an economic interest for different commercial sectors, such as the pharmaceutical, food and chemical industries [6]. Added to bioactive compounds, strawberry extrudate could be used to obtain other types of resources such as bioenergy [7, 8]. It is also well known the high phenolic composition in the achenes and in the pulp of the strawberry [9].

A general biorefinery scheme as a management option for the strawberry extrudate should look for synergies between unitary processes of extraction of bioactive compounds, purification and the management of the final biomass of the strawberry extrudate after extraction (**Figure 1**).

The extraction of bioactive compounds in agro-waste materials, such as the strawberry extrudate can be performed through various extraction techniques [10]. The main objective at this step consists of solubilizing the compounds of interest, with less possible impurities and making it an economically profitable technique. Extraction techniques in literature can be clustered into two groups: conventional extraction techniques, e.g. hydrothermal treatments, which are widely used at lab and full scale [11], and in recent years, more innovative techniques, e.g. enzyme assisted extraction. Additionally, combined extraction techniques between conventional and innovative techniques are being carried out to achieve high extraction yield [12]. All these extraction techniques will be revised and analyzed in the present chapter.

Any of these extraction techniques usually generate a liquid phase, with the bioactive compounds of interest, and a solid phase with a high amount of organic matter. After purification process of the liquid phase, a new liquid phase remains without the extracted bioactive compounds. Therefore, just the recovery of compounds of interest from the strawberry extrudate does not solve the problem of stabilization of the biomass of the strawberry extrudate and the use of a subsequent

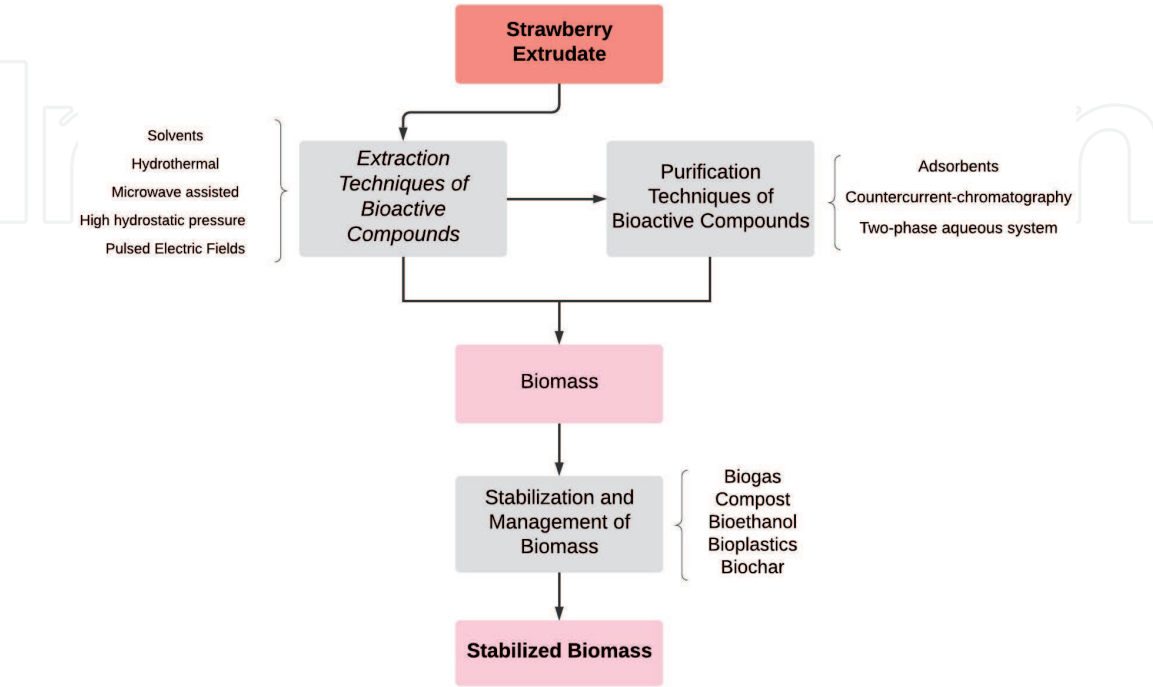


Figure 1.
General scheme of a biorefinery approach as a valorisation option for strawberry extrudate.

treatment is necessary for its stabilization [13]. The liquid phase after purification and the solid phase must undergo a new treatment for stabilization. In addition, extraction and purification processes consume energy which should be valued. The main options for assessing and stabilizing biomass after the extraction and purification process of the bioactive compounds should be focused on obtaining bioenergy and other bioproducts of interest [14].

The present chapter aims to summarize the bioactive compounds present in strawberries, to summarize and critically analyzes the different extraction and purification techniques for the recovery of these bioactive compounds, as well as the different options for the management and stabilization of the strawberry extrudate after the extraction process.

2. Bioactive compounds in strawberry extrudate and strawberries

2.1 Nutrients

Strawberry extrudate presents similar nutrients composition than strawberry [15]. The strawberry has high concentration of dietary fibrous (2 g fibrous/100 g raw strawberry), such as lignin, hemicellulose, cellulose, and pectin, containing small amounts of protein (0.4–0.5 g protein/100 g raw strawberry) and fat (0.1 g fat/100 g raw strawberry) [16, 17].

The strawberry contains high concentrations of vitamin C, contributing to 24% to the antioxidant capacity of strawberries [16]. The recommended daily intake of vitamins (100–150 mg/day) can be satisfied with an average of 100 g of strawberries per day [18]. Furthermore, strawberry is a source of many other vitamins in smaller amounts, such as vitamin E, vitamin A, vitamin B6, vitamin K, thiamine, riboflavin, folate acid, and niacin (0.01–0.4 g vitamin/100 g raw strawberry) [5, 19]. Strawberry is also rich in manganese, potassium, and a good source of iodine, magnesium, copper, iron, and phosphorus [5, 16].

The sugar composition of strawberries varies with the degree of maturity of the fruit [20], being glucose, fructose, and sucrose the main sugars in strawberries [5]. Sugars in strawberries are involved in the taste of the fruit and are responsible for the caloric value of the strawberries. Organic fatty acids such as citric acid, malic acid, succinic acid, tartaric acid, oxalic acid, and fumaric acid are ones of the response of the taste, texture, pH, and color of the strawberry, and can alter the sensory quality of this fruit [6].

2.2 Phytochemicals

Figure 2 shows a general scheme for the classification of phytochemical compounds that can be founded in the strawberry extrudate. Phytochemicals are widely studied, mainly due to the extensive types of compounds that have potential biological benefits in humans. The main phytochemicals in strawberries are the flavonoids, followed by the hydrolysable tannins and the phenolic acids and, as minor constituents, the condensed tannins [5].

2.2.1 Flavonoids

Flavonoids are divided into two groups, i.e. anthocyanins, and anthoxanthines. Anthoxanthines, in turn, are grouped into five subclasses, i.e. flavones, flavonols, flavanones, flavanols, and isoflavones [21]. The three main classes of flavonoids in strawberries are anthocyanis, flavonols, and flavanols [22].

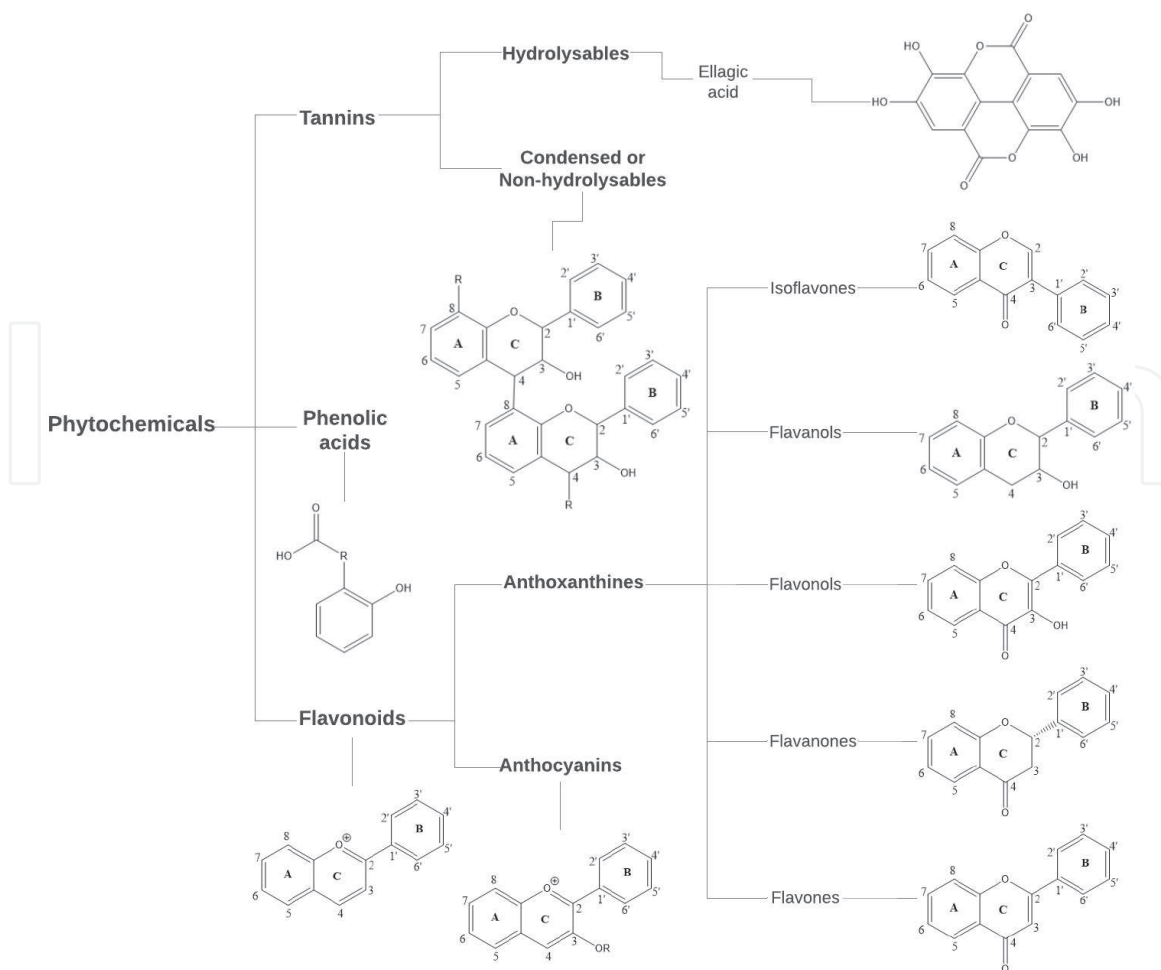


Figure 2.
General scheme of phytochemicals contained in strawberries.

The most relevant flavonoids present in strawberries are the anthocyanins due to their high concentration, approximately 20–47 mg/100 g raw strawberry [16]. More than 25 different pigments of anthocyanins have been described in the different varieties of strawberries [5]. Anthocyanins are responsible for the red color in strawberries [16]. The most important anthocyanins of the strawberry belong to the family of pelargonidin aglycones and cyanidin aglycones [23–25]. According to several studies, pelargonidin-3-glucoside is the dominant anthocyanin in strawberries [16, 24, 26–28]. The interest in anthocyanins have recently increased because of its pharmacological and therapeutic properties [5]. Anthocyanins have shown to positive effect toward reduction of coronary diseases, anticancer, antitumor, anti-inflammatory and anti-diabetic effects; as well as improving visual acuity and cognitive behavior [29]. These therapeutic effects of anthocyanins are connected to their high antioxidant activity [29]. In addition, anthocyanins can be used as a pigment in the food industry [29].

The second most important group of flavonoids in strawberries are flavonols, with approximately 1.5–3.4 mg/100 g raw strawberry [5, 16, 30, 31]. The most important flavonols of the strawberry belong to the family of quercetin and kaempferol, being the quercetin-derivatives the most abundant flavonols in strawberries [5, 17, 25–27]. Quercetin, in particular, is a potent antioxidant, cytoprotective, and anti-inflammatory [30].

Finally, the third group of flavonoids in strawberries are flavanols. Flavanols are the only class of flavonoids that do not naturally occur as glycosides. They are found in strawberries as monomeric compounds, such as catechins, and in

polymeric form, which are called condensed or non-hydrolysable tannins [5, 25]. These compounds can be difficult to measure in the strawberry because they are usually presented as part of a complex mixture of phenolic substances. Because of this, the amount of catechins present is sometimes overestimated [31]. At low concentrations flavanols particularly the catechins, are used as sweetening and/or flavoring additives. These flavanols improve taste and sweetness but are not substitutes for sweeteners and flavorings as they do not have taste and are a little astringent. Some authors have pointed out that their role is to make the receptors in the mouth more sensitive to sweeteners, thus lowering the levels of the sweeteners and flavorings used [21, 32].

2.2.2 Tannins

Tannins are classified into two groups: non-hydrolysable or condensed tannins and hydrolysable tannins (**Figure 2**). The condensed tannins are also called proanthocyanins, and are bound to the flavanols [33]. The content of condensed tannins in strawberries is approximately 54–163 mg/100 g raw strawberry [16]. In strawberries, the most relevant condensed tannins are procyanidins from catechin and its polymers. Condensed tannins are commonly found in the pulp of strawberries and achenes [5]. Due to the variety of physiological activities, they have been reported to possess, directly and indirectly, antioxidant, antimicrobial, anti-allergic and antihypertensive properties, as well as to inhibit the activities of some enzymes and physiological receptors [34].

The most common hydrolysable tannins in strawberries are ellagitannins, specifically sanguin H-6 and ellagic acid [5, 26–28]. The content of ellagitannins in strawberries is approximately 10–23 mg/100 g raw strawberry [16]. Ellagic acid is an ellagitannin present in the secondary metabolism of vegetables, its main characteristic is its antioxidant, antimicrobial, antimutagenic, anticarcinogenic and antiviral capacity [16]. The content of ellagic acid in strawberries is approximately 1–2 mg/100 g raw strawberry [16]. Due to the phenolic nature of ellagic acid, this compound tends to react by forming complexes with other molecules of proteins, alkaloids, and polysaccharides, so that it is usually found as ellagitannins esterified with glucose, because of this it is difficult to find it free [20]. The properties of ellagic acid are also exploited in the food industry, so it is used in the manufacture of nutraceutical drinks and food supplements. Likewise, the application of ellagic acid for food preservation is of great impact for the perishable food industry, using its antioxidant activity for microorganisms inhibition [35].

2.2.3 Phenolic acids

Strawberries contain a variety of phenolic acids which are presented as derivatives of the hydroxycinnamic acid, such as caffeic acid, and hydroxybenzoic acids such as gallic acid [5]. The content of phenolic acids in strawberries is approximately 0.8–6.7 mg/100 g raw strawberry [16]. The major hydroxycinnamic acid in strawberries is p-coumaroylhexose, but ferulic acid and caffeic acid glycosides have also been identified in strawberries [26, 27]. Hydroxycinnamic acid derivatives are responsible for the bitter taste of the strawberry, and it is used in the manufacture of creams [33, 36]. The primary derivative of hydroxybenzoic acid is p-hydroxybenzoic glycoside [28]. The p-hydroxybenzoic glycoside is widely used in the synthesis of organic compounds and their esters, known as parabens, which are used as preservatives in cosmetics [37].

3. Extraction and purification of bioactive compounds in strawberry extrudate

3.1 Extraction techniques

3.1.1 Solvents extraction

The extraction with solvents is a technique to isolate a substance from a solid or liquid mixture. This technique is currently used in combination with other techniques such as microwaves and ultrasound since the solvent only extracts soluble compound. Due to the strawberry extrudate nature, the solid–liquid extraction can be carried out with a Soxhlet extractor, which is one of the most commonly used conventional extraction techniques [38]. The extraction efficiency depends mainly on the choice of solvents [39]. The polar character of the bioactive compounds allows their solubility in various solvents, such as water, alcohols, and acetone [40].

Recently, numerous studies have explored the extraction of bioactive compounds using deep eutectic solvents from various groups of natural sources [41]. The formation of eutectic solvents is the result of the complexation of a halide salt, which acts as a hydrogen bond receptor, and a hydrogen bond donor [41]. Some eutectic solvents have been developed from the combination of primary metabolites and bio-renewable starting materials, e.g., sugars, alcohols, amino acids and organic acids [41]. Eutectic solvents produce less adverse effects on the environment, allowing to replace conventional chemical methods [41].

There is a long variety of studies on solvent extraction focusing on the extraction of bioactive compounds. An evaluation of the effect of different solvents and acids in the extraction of anthocyanins from strawberry fruits concluded that acetone provided an efficient and reproducible extraction, avoiding problems with pectins and allowing the concentration of the sample at low temperature [42]. In another study, it was observed that the acetone/acetic acid mixture (99:1, v/v) reached good results for the qualitative and quantitative evaluation of polyphenols present in strawberries [43].

3.1.2 Hydrothermal extraction

Hydrothermal extraction is a process in which the matter is treated by adding hot water or water vapor [38]. Steam explosion is another kind of hydrothermal treatment where the matter is treated with saturated water vapor at high pressure followed by rapid depressurization [44]. The disadvantage of using hydrothermal treatments is that they affect thermosensitive compounds and might form undesirable compounds [45].

Hydrothermal extraction at low temperature, i.e. ranging between 50 and 90°C, mainly induces the de-flocculation of macromolecules [46]. Hydrothermal extraction at medium temperature, i.e. ranging between 150 and 180°C, solubilizes cellulosic and hemicellulose biomass [47]. The steam explosion treatment, with temperature ranging between 180 and 260°C and increase in pressure of 0.69–4.83 MPa, it is able to solubilize lignocellulose biomass [44].

Several studies confirm the successful extraction of bioactive compounds by these hydrothermal extractions [48, 49]. Extraction of bioactive compounds in strawberry extrudate has been studied by applying hydrothermal treatments in the range of 90–200°C, [15]. Thermal treatment at 150°C for 60 minutes was the most efficient process based on the solubilization of sugars and phenols as well as the antioxidant capacity of the liquid phase produced [15].

3.1.3 Microwave assisted extraction

Microwaves are electromagnetic fields in a frequency range of 300 MHz to 30 GHz, which are generally operated at a frequency of 2.45 GHz [10]. Microwaves can access biological matrices and interact with polar molecules, such as water, which vibrate or rotate by the effect of microwaves and generate heat and can enhance the processes of extraction of bioactive compounds [10, 50]. Microwave assisted extraction has been successfully applied in anthocyanin extraction processes in grape skins [51], the recovery of pectins from press residues of various berries, i.e. red and black currant, raspberry and elderberry [52] and to extract phenolic antioxidants from peanut skins [53].

3.1.4 High hydrostatic pressure extraction

High hydrostatic pressure extraction is a method that works at high pressures ranging from 100 to 1000 MPa. These high pressures cause cell deformation, cell membrane damage, protein denaturation, deprotonation of charged groups, and the breakdown of bonds, making bioactive compounds more accessible for extraction [54]. High hydrostatic pressure extraction is considered to be a faster and more efficient technique than other conventional extraction methods [55, 56]. In addition, high hydrostatic pressure extraction has the advantage of not increasing the temperature during the processing time, so it would be an ideal method to extract thermosensitive compounds.

Several high hydrostatic pressure extraction studies have been carried out with strawberries for the extraction of bioactive compounds. The impact of high hydrostatic pressure extraction on total strawberry puree phenols was observed by Patras et al., [56], which reported that the amount of total phenols increases as the pressure in high hydrostatic pressure extraction increases [56]. In another study, the change in kaempferol, and quercetin quantity in strawberries pulps were tested at different pressures and for different processing times [57]. According to this study, the change in the amount of kaempferol was not very significant and the amount of quercetin increased with increasing pressure [57]. Another study showed that the nutritional and sensory qualities of strawberry puree after high-pressure processing at 500 MPa and 50°C for 15 min were much better than after a heat treatment at 90°C for 15 min [58].

3.1.5 Pulsed electric fields extraction

Pulsed electric fields or high intensity pulsed electric fields consist of a short time electrical treatment, between nanoseconds to milliseconds, in which the material located between two electrodes is exposed to a strong electric pulse of intensity field of 100 to 300 V/cm Pulsed Electric Fields or 20 to 80 kV/cm high intensity pulsed electric field, the operation parameters being the duration and number of pulses [59, 60]. Pulsed electric fields can produce the electrical rupture of the cell membranes producing the formation of pores, what is known as electroporation [60, 61]. Pore formation improves cell permeability allowing the recovery of bioactive compounds [62]. Compared to other non-thermal treatments such as the high hydrostatic pressure extraction method, pulsed electric field extraction methods require a much shorter processing time, higher extraction efficiency and these techniques can be easily applied in continuous operation [59]. Therefore, pulsed electric fields is a promising technique for different applications in the food industry because they can improve extraction capacity and recovery of nutritionally valuable compounds as well as the bioavailability of micronutrients and compounds in a wide range of foods [59].

Several studies on the extraction of antioxidant compounds in agri-foods show enhanced yields with pulsed electric fields. For instance, a comparison study between a heat treatment at 90°C for 60 or 30 seconds and high intensity pulsed electric field in strawberries juice, showed that strawberry juice treated with high intensity pulsed electric field maintained greater amount of phenolic acids and total anthocyanins than thermally treated juices [63, 64]. Likewise, the recovery of phenols from the shell of the pomegranate by pulsed electric field has been assayed, resulting in a similar antioxidant extraction yields and an energy saving of 50% compared to an ultrasound extraction technique [65].

3.1.6 Extraction techniques comparison

After reviewing the different extraction techniques that have been applied to the strawberry and strawberry extrudate, a summary describing their most interesting aspects is shown in **Table 1**. The aspects that have been compared are: the specificity of the extraction techniques with the bioactive compounds, the possibility of combining with other extraction techniques, the ability to release bioactive compounds, the potential degradation of bioactive compounds, possibility of intracellular attack, bonds breakage and whether the technique has a high operational and investment cost. The choice of the best technique for the strawberry extrudate is a tailor-made solution for each situation that will depend of the investment capacity, target compounds to be recovered or the required extraction yield.

3.2 Purification techniques of bioactive compounds

3.2.1 Adsorbents

There are many studies that show the properties of adsorbents to separate, concentrate and purify various compounds [66, 67]. Functionality, porosity, irregularities, surface area, tightly bonded impurities, internal porous structure, particle size, ionic strength, pH, and temperature all influence physical adsorption [66]. The temperature influences the adsorption in two ways, increasing the transport

Extraction technique	Specificity	Possibility of combination	Ability to release compounds	Degradation of bioactive compounds	Intracellular attack	Breaks of bonds	High cost
Solvents extraction	x	x					
Hydrothermal extraction		x	x	x		x	
Microwave assisted extraction		x	x	x	x	x	
High hydrostatic pressure extraction		x	x	x	x	x	x
Pulsed electric fields extraction		x	x	x	x	x	

Table 1.
Summary table of characteristics for comparing extraction techniques.

speed through the outer boundary layer and inside the pores due to the decrease in the viscosity of the solution, and changing the capacity of the adsorbent. However, high temperatures can promote irreversible interactions [67]. Another important parameter for purification with adsorbents is pH. For example, at acid pH, the adsorption of phenolic compounds by different adsorbents increases because the phenols are not dissociated and dispersion interactions predominate [66]. At alkaline pH, the adsorption decreases due to the dissociation of hydroxyl groups and carboxyl groups [66]. There are many types of adsorbents such as activated carbons, mineral adsorbents, synthetic polymeric adsorbents, ion exchange resins, lignin and lignocellulosic materials, adsorbents based on polysaccharides and others [66]. Among the available adsorbents Amberlite XAD adsorbents are widely used in the concentration of polyphenols [68]. Zhang et al. [69] reported the isolation and structural characterization of 10 phenolic compounds from strawberry extracts using a combination of Amberlite XAD-16 and C18 columns, HPLC-UV, and nuclear magnetic resonance spectroscopy methods.

3.2.2 Countercurrent-chromatography

Countercurrent chromatography is a technique widely used in the purification of natural products [70]. Countercurrent chromatography is a liquid-liquid partition chromatography process in which both the mobile phase and the stationary phase are liquid [70]. The main advantage of countercurrent chromatography, when compared to equivalent techniques such as low pressure liquid chromatography, is that there are no adsorption losses in the stationary phase [70]. The range of selectivity offered by chromatographic resins is equivalent to the range of selectivity offered by the different solvent systems [70].

Several studies have shown the importance of countercurrent chromatography for the purification of bioactive compounds from strawberry. The compound 2,5-dimethyl-4-hydroxy-3[2H]-furanone 6'-O-malonyl- β -D-glucopyranoside was isolated from a strawberry glycosidic extract (*Fragaria* \times *ananassa*, cv. Senga Sengana) by countercurrent chromatography [71]. Peonidin-3-glucoside and malvidin-3-glucoside were obtained from grapes in a single step, while in a second step, cyanidin-3-glucoside was isolated [72]. In another research, the separation of anthocyanin monomers of high purity from mulberry fruits was developed [73].

3.2.3 Two-phase aqueous system

Two-phase aqueous system is a liquid-liquid fractionation technique that is usually formed by mixing two polymers in aqueous media, for example, polyethylene glycol and dextran or maltodextrin, or by a polymer and a salt, such as polyethylene glycol and salts of phosphates, citrates or sulphates [74–76]. This method has advantages over other purification techniques due to a comparatively low consumption of energy and time, as well as the possibility to be designed for a continuous operation. Moreover, two-phase aqueous systems are effective for many types of substances, especially for the concentration and purification of bioactive compounds [74, 75]. Several studies have demonstrated the suitability of this technique for the purification of bioactive compounds such as phenolic compounds from fig fruits (*Ficus carica* L.) [76], or the purification of gallic acid from natural matrices with ionic liquids [77]. Furthermore, two-phase aqueous system has been applied for the purification of polyphenols from a model solution of gallic acid and three real samples of red and white wine, and orange juice in combination with macro and micro extractors [78]. Polyphenols have been also extracted from *Aronia melanocarpa* berries, using ultrasound-assisted extraction in combination with the two-phase aqueous system [79].

4. Stabilization of biomass by obtaining bioenergy and bioproducts

4.1 Biogas production

Anaerobic digestion is a microbiological process, in absence of oxygen, where organic matter is progressively degraded by an heterogeneous bacterial population to methane (55–70%) and carbon dioxide (30–45%) [80]. Anaerobic digestion presents some fundamental advantages such as the possibility of working at high rates of organic load, and the produced methane can be used as an energy source due to its heating value ($35,793 \text{ kJ/m}^3$, at 1 atm, 0°C), which equals to 1 kg of raw coal or 0.76 kg of standard coal [3, 11]. The use of biogas for energy supply reduces deforestation, soil erosion and environmental pollution [81, 82]. Also, it can improve the energy efficiency of various production processes due to the energetic contribution that provides [82]. In addition, a wet waste called digestate, which is a mixture of partially degraded organic matter, microbial biomass, and inorganic compounds, is produced during biomethanization and could be used as a base for fertilizers or organic amendments [82, 83].

Several studies on anaerobic digestion of strawberries extrudate have been carried out. The results of one these studies reveal that strawberries extrudates have a high level of anaerobic biodegradability (90% in VS, (total volatile solids)) and that a substantial amount of methane can be obtained in this way ($312 \text{ mL CH}_4 \text{ STP/g added VS}$) (STP: standard temperature and pressure conditions, i.e. 0°C , 1 atm) at an organic loading rate range of 2.04 to $3.51 \text{ kg VS/m}^3\cdot\text{d}$ [84]. In another study of anaerobic digestion of strawberry waste from supermarkets, using an organic loading rate of 0.55 – $4.4 \text{ (g/L}\cdot\text{d)}$, the experimental biogas and methane yields were 0.588 and 0.231 L/g , respectively [85]. It has been observed that sometimes it is necessary to co-digest strawberry extrudate with a substrate that provides alkalinity, such as sewage sludge [86, 87]. Co-digestion studies of strawberry extrudate with other substrates such as fish waste [3] and glycerol [83] have also been studied. Anaerobic digestion of strawberry extrudate is a promising technique but it should be further studied since low alkalinity of the extrudate together with formation of inhibitory compounds caused by the extraction process could negatively affect the digestion process.

4.2 Compost production

Composting has been proposed for a long time as a quite cheap option for agricultural waste management [2]. Composting has also been proposed as a post-treatment for the produced digestate after anaerobic digestion [88]. Composting is the bio-oxidative conversion of organic waste into an organic amendment. According to Gutiérrez et al. (2017) [2], the cost of composting varies in a wide range from \$40 to \$500 per throughput ton depending on the technology. Composting costs vary widely depending on the type of operation, which ranges from the most simple ones, such as opening windrows, to more complex procedures like in-vessel aerobic composting that allows smell emissions to be controlled and prevents environmental pollution [2]. The great disadvantage is that a considerable amount of offensive odors can be emitted during the process due to the generation of volatile organic compounds [89]. Other disadvantages are the long process time and the necessity of a proper monitoring [90]. Co-composting of a waste mixture containing strawberry extrudate, fish waste, sewage sludge and bulking agent has been successfully proven [2, 89].

4.3 Bioethanol production

Bioethanol is one of the most produced alcohols from the fermentation of sugars found in fruits and vegetables [7, 91–93]. Theoretically, any organic product with a

high content of sugars and starch, such as strawberry extrudate, may be susceptible to obtaining bioethanol [91]. Inedible sources from the strawberry extrudate such as lignocellulosic biomass, which mainly comprises cellulose, hemicellulose, and lignin, can be hydrolysed to produce a mixture of pentoses and hexoses that can be transformed into bioethanol [94]. Bioethanol from agro-waste, such as strawberry extrudate, could be a promising technology that involves four processes, pre-treatment, enzymatic hydrolysis, fermentation and distillation, this final step is crucial for the process to be economically viable on a commercial scale due to high energy consumption in the form of steam to increase the yield of bioethanol production when lignocellulose materials are used as raw material [93]. These processes have several challenges and limitations, such as the efficient pre-treatment process to eliminate lignin from the lignocellulosic agro-residues. The proper pre-treatment process can increase the concentrations of fermentable sugars after enzymatic hydrolysis, thus improving the efficiency of the entire process [92].

4.4 Bioplastics production

Fossil fuel depletion, global warming, and problems of pollution of the environment that provoke plastics in its life cycle are encouraging the development of biodegradable plastics [95, 96]. Agri-food waste are usually rich in many useful substances such as lipids, polysaccharides, and aromatics, which could be used for the manufacture of biodegradable polymeric materials. Bioplastics already play an important role in the sectors of packaging, agriculture, consumer electronics and motoring, but still have a very low share in the total production of plastics. Currently, about 1% of the annual tons of plastic are bioplastics [97]. Examples of such bioplastics are exopolysaccharides, polycaprolactone, polybutylene succinate, polybutylene adipate terephthalate, polyhydroxyalkanoates or polyhydroxybutyrates [97, 98]. For obtaining bioplastics from agri-food waste, the waste must be treated to extract or isolate specific macromolecules, such as cellulose, lignin, suberin, starch, or monomers, such as vegetable oils, tannins and terpenes [96, 99]. A study conducted on the production of bioplastics from Murta fruit extract, that is a native Chilean berry, showed the feasibility of using berries for bioplastic production [8].

4.5 Biochar production

Biochar is the solid carbonaceous residue produced through organic waste and used as a soil improver [100, 101]. Biochar is produced through several types of methods such as pyrolysis, torrefaction or hydrothermal carbonization [100, 101].

There are no studies reported in the literature dealing with the production of biochar from strawberry extrudates. However, the above-mentioned techniques (pyrolysis, torrefaction and hydrothermal carbonization) could be potentially applied for this substrate. Several studies have been carried out on the hydrothermal carbonization of other agri-food waste, such as olive cuttings and olive pulp [102]; grape marc [103]; olive mill waste, canned artichoke and orange waste [104].

5. Conclusions

This chapter has reviewed up-to-date literature on the bioactive compounds contained in strawberries, which have an important health and market value. Different extraction and purification techniques to obtain valuable compounds from strawberry extrudate have been reviewed and analyzed. The reviewed techniques present different advantages and drawbacks that were analyzed to facilitate the selection of the most suitable process for each valorisation scenario. Finally,

different stabilization options for the biomass remaining after extraction have also been reviewed. Stabilization is required to avoid severe environmental impacts, and additionally could be an economically beneficial aid for balancing the cost of the extraction of high value-added compounds. As usually for any waste management option, selection of the best extraction, purification and stabilization technique for the strawberry extruded is a tailor-made solution for each situation.

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Conflict of interest

The authors declare no conflict of interest.

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References

- [1] FAO. FAOSTAT: Food and agriculture organization of the united nations [Internet]. 2020 [cited 2020 Jul 17]. Available from: <http://www.fao.org/faostat/es/#data/QC/visualize>
- [2] Gutiérrez MC, Serrano A, Siles JA, Chica AF, Martín MA. Centralized management of sewage sludge and agro-industrial waste through co-composting. *Journal of Environmental Management*. 2017 Jul;196:387-93.
- [3] Serrano A, Siles JA, Chica AF, Martín MÁ. Agri-food waste valorization through anaerobic co-digestion: Fish and strawberry residues. *Journal of Cleaner Production*. 2013 Sep;54:125-32.
- [4] Aharonov-Nadborny R, Raviv M, Graber ER. Soil spreading of liquid olive mill processing wastes impacts leaching of adsorbed terbutylazine. *Chemosphere*. 2016;156(Supplement C): 220-7.
- [5] Giampieri F, Tulipani S, Alvarez-Suarez JM, Quiles JL, Mezzetti B, Battino M. The strawberry: Composition, nutritional quality, and impact on human health. *Nutrition*. 2012;28(1):9-19.
- [6] Dias MI, Barros L, Sousa MJ, Oliveira MBPP, Santos-Buelga C, Ferreira ICFR. Enhancement of nutritional and bioactive compounds by in vitro culture of wild *Fragaria vesca* L. vegetative parts. *Food Chemistry*. 2017;235:212-9.
- [7] Dias MOS, Ensinas A V, Nebra SA, Maciel Filho R, Rossell CE V, Maciel MRW. Production of bioethanol and other bio-based materials from sugarcane bagasse: Integration to conventional bioethanol production process. *Chemical Engineering Research and Design*. 2009 Sep 1;87(9):1206-16.
- [8] López de Dicastillo C, Bustos F, Guarda A, Galotto MJ. Cross-linked methyl cellulose films with murta fruit extract for antioxidant and antimicrobial active food packaging. *Food Hydrocolloids*. 2016;60(Supplement C):335-44.
- [9] Ariza MT, Reboredo-Rodríguez P, Mazzoni L, Forbes-Hernández TY, Giampieri F, Afrin S, et al. Strawberry achenes are an important source of bioactive compounds for human health. *International Journal of Molecular Sciences*. 2016;17(7):1103.
- [10] Wijngaard H, Hossain MB, Rai DK, Brunton N. Techniques to extract bioactive compounds from food by-products of plant origin. *Food Research International*. 2012;46(2): 505-13.
- [11] Fermoso FG, Serrano A, Alonso-Fariñas B, Fernández-Bolaños J, Borja R, Rodríguez-Gutiérrez G. Valuable Compound Extraction, Anaerobic Digestion, and Composting: A Leading Biorefinery Approach for Agricultural Wastes. *Journal of Agricultural and Food Chemistry American Chemical Society*; Aug 15, 2018 p. 8451-68.
- [12] Pérez-Loredo MG, Hernández- De Jesús L, Barragán-Huerta BE. Extracción de compuestos bioactivos de pitaya roja (*Stenocereus stellatus*) aplicando pretratamientos con microondas, ultrasonido y enzimáticos. *Agrociencia*. 2017;51(2):135-51.
- [13] Rajendran K, Drielak E, Sudarshan Varma V, Muthusamy S, Kumar G. Updates on the pretreatment of lignocellulosic feedstocks for bioenergy production—a review. Vol. 8, *Biomass Conversion and Biorefinery*. Springer Verlag; 2018. p. 471-83.
- [14] Machineni L. Lignocellulosic biofuel production: review of alternatives. *Biomass Conversion and Biorefinery*. Springer Verlag; 2019.

- [15] Rodríguez-Gutiérrez G, Cardoso JCJC, Rubio-Senent F, Serrano A, Borja R, Fernández-Bolaños J, et al. Thermally-treated strawberry extrudate: A rich source of antioxidant phenols and sugars. *Innovative Food Science and Emerging Technologies*. 2019 Jan 1;51:186-93.
- [16] Basu A, Nguyen A, Betts NM, Lyons TJ. Strawberry As a Functional Food: An Evidence-Based Review. *Critical Reviews in Food Science and Nutrition*. 2014;54(6):790-806.
- [17] Sójka M, Klimczak E, Macierzyński J, Kołodziejczyk K. Nutrient and polyphenolic composition of industrial strawberry press cake. *European Food Research and Technology*. 2013;237(6):995-1007.
- [18] Kafkas E, Koşar M, Paydaş S, Kafkas S, Başer KHC. Quality characteristics of strawberry genotypes at different maturation stages. *Food Chemistry*. 2007;100(3):1229-36.
- [19] Tulipani S, Romandini S, Suarez JMA, Capocasa F, Mezzetti B, Busco F, et al. Folate content in different strawberry genotypes and folate status in healthy subjects after strawberry consumption. *BioFactors*. 2008;34(1):47-55.
- [20] da Silva Pinto M, Lajolo FMFM, Genovese MIMI, Pinto MS, Lajolo FMFM, Genovese MIMI. Bioactive compounds and quantification of total ellagic acid in strawberries (*Fragaria x ananassa* Duch.). *Food Chemistry*. 2008;107(4):1629-35.
- [21] Ochoa M CI, Ayala A AA. Los Flavonoides: Apuntes Generales y su Aplicación en la Industria de Alimentos. *Revista Ingeniería y Competitividad*; Vol 6, Núm 2. 2004;
- [22] Robards K, Prenzler PD, Tucker G, Swatsitang P, Glover W. Phenolic compounds and their role in oxidative processes in fruits. *Food Chemistry*. 1999;66(4):401-36.
- [23] Böhm H. G. Mazza und E. Miniati: Anthocyanins in Fruits, Vegetables and Grains. 362 Seiten, zahlr. Abb. und Tab. CRC Press, Boca Raton, Ann Arbor, London, Tokyo 1993. Preis: 144.— £. Vol. 38, Food / Nahrung. CRC press; 1994. 343-343 p.
- [24] da Silva FL, Escribano-Bailón MT, Pérez Alonso JJ, Rivas-Gonzalo JC, Santos-Buelga C. Anthocyanin pigments in strawberry. *LWT - Food Science and Technology*. 2007;40(2):374-82.
- [25] da Silva Pinto M, Lajolo FM, Genovese MI. Bioactive compounds and quantification of total ellagic acid in strawberries (*Fragaria x ananassa* Duch.). *Food Chemistry*. 2008;107(4):1629-35.
- [26] Aaby K, Skrede G, Wrolstad RE. Phenolic composition and antioxidant activities in flesh and achenes of strawberries (*fragaria ananassa*). *Journal of Agricultural and Food Chemistry*. 2005;53(10):4032-40.
- [27] Aaby K, Ekeberg D, Skrede G. Characterization of phenolic compounds in strawberry (*Fragaria x ananassa*) fruits by different HPLC detectors and contribution of individual compounds to total antioxidant capacity. *Journal of Agricultural and Food Chemistry*. 2007;55(11):4395-406.
- [28] Klopotek Y, Otto K, Böhm V. Processing strawberries to different products alters contents of vitamin C, total phenolics, total anthocyanins, and antioxidant capacity. *Journal of Agricultural and Food Chemistry*. 2005;53(14):5640-6.
- [29] Garzón GA. Anthocyanins As Natural Colorants And Bioactive Compounds: A Review. *Acta Biológica Colombiana*. 2008;13(3):27-36.

- [30] Shaik-Dashagirisahab YB, Varvara G, Murmura G, Saggini A, Caraffa A, Antinolfi P, et al. Inhibitor effect of antioxidant flavanoids quercetin, and capsaicin in mast cell inflammation. *European Journal of Inflammation*. 2013;357(2):353-7.
- [31] Hannum SM. Potential Impact of Strawberries on Human Health: A Review of the Science. *Critical Reviews in Food Science and Nutrition*. 2004;44(1):1-17.
- [32] Kashket S. Sweetness and flavor enhanced compositions and method of preparing such compositions. 1990 Apr 7; Techniques for extraction of bioactive compounds from plant materials: A review. *Journal of Food Engineering*. 2013;117(4):426-36.
- [39] Cowan MM. *Plant Products as Antimicrobial Agents*, Clinical Microbiology Reviews, 565, 568, 570. Departement of Microbiology, Miami University. Oxford, Ohio; 1999.
- [40] Zapata LM, Heredia AM, Quinteros CF, Malleret AD, Clemente G, Cárcel JA. Optimización de la extracción de antocianinas de arándanos. *Ciencia, Docencia y Tecnología*. 2014;25(49):166-92.
- [33] Porras-Loaiza A p., López-Malo A. Importancia de los grupos fenólicos en los alimentos. *Temas Selectos de Ingeniería de Alimentos*. 2009;3(1):121-34.
- [41] Zainal-Abidin MH, Hayyan M, Hayyan A, Jayakumar NS. New horizons in the extraction of bioactive compounds using deep eutectic solvents: A review. *Analytica Chimica Acta*. 2017;979:1-23.
- [34] Santos-Buelga C, Scalbert A. Proanthocyanidins and tannin like compounds nature, occurrence, dietary intake and effects on nutrition and health. *Journal of the Science of Food and Agriculture*. 2000;80(May):1094-117.
- [42] Garcia-Viguera C, Zafrilla P, Tomás-Barberán FA. The use of acetone as an extraction solvent for anthocyanins from strawberry fruit. *Phytochemical Analysis*. 1998;9(6):274-7.
- [35] Saucedo-Pompa S, Rojas-Molina R, Aguilera-Carbó AF, Saenz-Galindo A, GarzaB H de La, Diana Jasso-Cantú, et al. Edible Films based on canadelilla wax to improve the shelr life and quality of hass avocado. *Food Research International*. 2009;42(4):511-515.
- [43] Kajdžanoska M, Petreska J, Stefova M. Comparison of different extraction solvent mixtures for characterization of phenolic compounds in strawberries. *Journal of Agricultural and Food Chemistry*. 2011 May 25;59(10):5272-8.
- [36] Joseph ALII, DiNardo JC. Methods of preparing and using botanical antioxidant compositions. Google Patents; 2017.
- [44] Rincón B, Rodríguez-Gutiérrez G, Bujalance L, Fernández-Bolaños J, Borja R. Influence of a steam-explosion pre-treatment on the methane yield and kinetics of anaerobic digestion of two-phase olive mil solid waste or alperujo. *Process Safety and Environmental Protection*. 2016;102(Supplement C):361-9.
- [37] Ma WL, Zhao X, Lin ZY, Mohammed MOA, Zhang ZF, Liu LY, et al. A survey of parabens in commercial pharmaceuticals from China and its implications for human exposure. *Environment International*. 2016;95(Supplement C):30-5.
- [45] Struck S, Plaza M, Turner C, Rohm H. Berry pomace - A review of processing and chemical analysis of its polyphenols. *International Journal*
- [38] Azmir J, Zaidul ISM, Rahman MM, Sharif KM, Mohamed A, Sahena F, et al.

of Food Science and Technology. 2016;51(6):1305-18.

[46] Jain S, Jain S, Wolf IT, Lee J, Tong YW. A comprehensive review on operating parameters and different pretreatment methodologies for anaerobic digestion of municipal solid waste. Renewable and Sustainable Energy Reviews. 2015;52:142-54.

[47] Hendriks ATWM, Zeeman G. Pretreatments to enhance the digestibility of lignocellulosic biomass. Bioresource Technology. 2009;100(1):10-8.

[48] Serrano A, Feroso FG, Alonso-FariñasB, Rodríguez-GutierrezG, Fernandez-Bolaños J, Borja R. Olive mill solid waste biorefinery: High-temperature thermal pre-treatment for phenol recovery and biomethanization. Journal of Cleaner Production. 2017 Apr 1;148:314-23.

[49] Mrabet A, Jiménez-Araujo A, Fernández-Bolaños J, Rubio-Senent F, Lama-Muñoz A, Sindic M, et al. Antioxidant phenolic extracts obtained from secondary Tunisian date varieties (*Phoenix dactylifera* L.) by hydrothermal treatments. Food Chemistry. 2016 Apr 1;196(Supplement C):917-24.

[50] Flores E. Extracción de Antioxidantes de las Bayas del Sauco (*Sambucus nigra* L. *subsp.* *peruviana*) con Ultrasonido, Microondas, Enzimas y Maceración para la Obtención de Zumos Funcionales. Informacion Tecnologica. 2017;28(1):121-32.

[51] Liazid A, Guerrero RF, Cantos E, PalmaM, Barroso CG. Microwave assisted extraction of anthocyanins from grape skins. Food Chemistry. 2011;124(3):1238-43.

[52] Bélaí-Bakó K, Cserjési P, Beszédes S, Csanádi Z, Hodúr C. Berry Pectins: Microwave-Assisted Extraction and Rheological Properties.

Food and Bioprocess Technology. 2012;5(3):1100-5.

[53] Ballard TS, Mallikarjunan P, Zhou K, O'Keefe S. Microwave-assisted extraction of phenolic antioxidant compounds from peanut skins. Food Chemistry. 2010;120(4):1185-92.

[54] Shouqin Z, Jim X, Changzheng W. Note: Effect of high hydrostatic pressure on extraction of flavonoids in propolis. Food Science and Technology International. 2005;11(3):213-6.

[55] Corrales M, Toepfl S, Butz P, Knorr D, Tauscher B. Extraction of anthocyanins from grape by-products assisted by ultrasonics, high hydrostatic pressure or pulsed electric fields: A comparison. Innovative Food Science and Emerging Technologies. 2008;9(1):85-91.

[56] Patras A, Brunton NP, Da Pieve S, Butler F. Impact of high pressure processing on total antioxidant activity, phenolic, ascorbic acid, anthocyanin content and colour of strawberry and blackberry purées. Innovative Food Science and Emerging Technologies. 2009 Jul;10(3):308-13.

[57] Cao X, Zhang Y, Zhang F, Wang Y, Yi J, Liao X. Effects of high hydrostatic pressure on enzymes, phenolic compounds, anthocyanins, polymeric color and color of strawberry pulps. Journal of the Science of Food and Agriculture. 2011;91(5):877-85.

[58] Marszałek K, Mitek M, Skąpska S. The effect of thermal pasteurization and high pressure processing at cold and mild temperatures on the chemical composition, microbial and enzyme activity in strawberry purée. Innovative Food Science and Emerging Technologies. 2015;27(Supplement C): 48-56.

[59] Yan L-G, He L, Xi J. High intensity pulsed electric field as an innovative

technique for extraction of bioactive compounds—A review. *Critical Reviews in Food Science and Nutrition*. 2017;57(13):2877-88.

[60] Parniakov O, Lebovka NI, Van Hecke E, Vorobiev E. Pulsed Electric Field Assisted Pressure Extraction and Solvent Extraction from Mushroom (*Agaricus Bisporus*). *Food and Bioprocess Technology*. 2013;7(1):174-83.

[61] Neuman E, Schaefer-Ridder M, Wang Y, Hofschneider PH. Gene transfer into mouselyoma celles by electroporation in high electric fields. *Embo J*. 1982 Jul 1;1(7):1841-.

[62] Angersbach A, Heinz V, Knorr D. Effects of pulsed electric fields on cell membranes in real food systems. *Innovative Food Science & Emerging Technologies*. 2000;1(2):135-49.

[63] Odriozola-Serrano I, Soliva-Fortuny R, Martín-Belloso O. Impact of high-intensity pulsed electric fields variables on vitamin C, anthocyanins and antioxidant capacity of strawberry juice. *LWT - Food Science and Technology*. 2009;42(1):93-100.

[64] Odriozola-Serrano I, Soliva-Fortuny R, Martín-Belloso O. Phenolic acids, flavonoids, vitamin C and antioxidant capacity of strawberry juices processed by high-intensity pulsed electric fields or heat treatments. *European Food Research and Technology*. 2008;228(2):239-48.

[65] Pan Z, Qu W, Ma H, Atungulu GG, McHugh TH. Continuous and pulsed ultrasound-assisted extractions of antioxidants from pomegranate peel. *Ultrasonics Sonochemistry*. 2012;19(2):365-72.

[66] Soto ML, Moure A, Domínguez H, Parajó JC. Recovery, concentration and purification of phenolic compounds by adsorption: A review. *Journal of Food Engineering*. 2011;105(1):1-27.

[67] Qiu N, Guo S, Chang Y. Study upon kinetic process of apple juice adsorption de-coloration by using adsorbent resin. *Journal of Food Engineering*. 2007;81(1):243-9.

[68] Ahmad A, Siddique JA, Laskar MA, Kumar R, Mohd-Setapar SH, Khatoon A, et al. New generation Amberlite XAD resin for the removal of metal ions: A review. *Journal of Environmental Sciences (China)*. 2015;31(Supplement C):104-23.

[69] Zhang Y, Seeram NP, Lee R, Feng L, Heber D. Isolation and identification of strawberry phenolics with antioxidant and human cancer cell antiproliferative properties. *Journal of Agricultural and Food Chemistry*. 2008;56(3):670-5.

[70] Valls J, Millán S, Martí MP, Borràs E, Arola L. Advanced separation methods of food anthocyanins, isoflavones and flavanols. *Journal of Chromatography A*. 2009;1216(43):7143-72.

[71] Roscher R, Herderich M, Steffen JP, Schreier P, Schwab W. 2,5-dimethyl-4-hydroxy-3[2H]-furanone 6'O-malonyl-beta-D- glucopyranoside in strawberry fruits. *Phytochemistry*. 1996;43(1):155-9.

[72] Renault J-H, Thepenier P, Zeches-hanrot M, Le Men-Olivier L, Durand A, Foucault A, et al. Preparative separation of anthocyanins by gradient elution centrifugal partition chromatography. *Journal of Chromatography A*. 1997;763(1):345-52.

[73] Chen Y, Du F, Wang W, Li Q, Zheng D, Zhang W, et al. Large-scale isolation of high-purity anthocyanin monomers from mulberry fruits by combined chromatographic techniques. *Journal of Separation Science*. 2017 Sep 1;40(17):3506-12.

[74] Iqbal M, Tao Y, Xie S, Zhu Y, Chen D, Wang X, et al. Aqueous two-phase system (ATPS): an overview and

advances in its applications. *Biological Procedures Online*. 2016;18(1):1-18.

[75] Molino JVD, Viana Marques D de A, Júnior AP, Mazzola PG, Gatti MSV. Different types of aqueous two-phase systems for biomolecule and bioparticle extraction and purification. *Biotechnology Progress*. 2013;29(6):1343-53.

[76] Feng YC, Li WL, He FM, Kong TT, Huang XW, Gao ZH, et al. Aqueous Two-Phase System as an Effective Tool for Purification of Phenolic Compounds from Fig Fruits (*Ficus carica* L.). *Separation Science and Technology (Philadelphia)*. 2015;50(12):1785-93.

[77] Cláudio AFM, Ferreira AM, Freire CSR, Silvestre AJD, Freire MG, Coutinho JAP. Optimization of the gallic acid extraction using ionic-liquid-based aqueous two-phase systems. *Separation and Purification Technology*. 2012;97(Supplement C):142-9.

[78] Tušek AJ, Šalić A, Zelić B. Mathematical modelling of polyphenol extraction by aqueous two-phase system in continuously operated macro- and micro-extractors. *Separation Science and Technology (Philadelphia)*. 2017;52(5):864-75.

[79] Xu YY, Qiu Y, Ren H, Ju DH, Jia HL. Optimization of ultrasound-assisted aqueous two-phase system extraction of polyphenolic compounds from *Aronia melanocarpa* pomace by response surface methodology. *Preparative Biochemistry and Biotechnology*. 2017;47(3):312-21.

[80] Monge O, Certucha Barragn MT, Almendariz Tapi FJ. Microbial Biomass in Batch and Continuous System. In: *Biomass Now - Sustainable Growth and Use*. InTech; 2013.

[81] Pérez I, Garfi M, Cadena E, Ferrer I. Technical, economic and environmental assessment of household biogas digester

for rural communities. *Renewable Energy*. 2014;62(Supplement C):7.

[82] Bożym M, Florczak I, Zdanowska P, Wojdalski J, Klimkiewicz M. An analysis of metal concentrations in food wastes for biogas production. *Renewable Energy*. 2015 May 1;77:467-72.

[83] Serrano A, Siles JA, Chica AF, Martín MA. Improvement of mesophilic anaerobic co-digestion of agri-food waste by addition of glycerol. *Journal of Environmental Management*. 2014 Jul;140:76-82.

[84] Siles JA, Serrano A, Martín A, Martín MA. Biomethanization of waste derived from strawberry processing: Advantages of pretreatment. *Journal of Cleaner Production*. 2013 Mar 1;42:190-7.

[85] Arhoun B, Gomez-Lahoz C, Abdala-Diaz RT, Rodriguez-Maroto JM, Garcia-Herruzo F, Vereda-Alonso C. Production of biogas from co-digestion of livestock and agricultural residues: A case study. *Journal of Environmental Science and Health - Part A Toxic/Hazardous Substances and Environmental Engineering*. 2017;52(9):856-61.

[86] Serrano A, Siles JA, Gutiérrez MC, Martín MÁ. Improvement of the biomethanization of sewage sludge by thermal pre-treatment and co-digestion with strawberry extrudate. *Journal of Cleaner Production*. 2015 Mar 1;90:25-33.

[87] Serrano A, Siles JA, Chica AF, Martín MÁ. Anaerobic co-digestion of sewage sludge and strawberry extrudate under mesophilic conditions. *Environmental Technology (United Kingdom)*. 2014 Dec 2;35(23):2920-7.

[88] Vaneeckhaute C, Lebuf V, Michels E, Belia E, Vanrolleghem PA, Tack FMG, et al. Nutrient Recovery from Digestate: Systematic Technology

Review and Product Classification.
 Waste and Biomass Valorization.
 2017;8(1):21-40.

[89] Toledo M, Gutiérrez MC, Siles JA, García-Olmo J, Martín MA. Chemometric analysis and NIR spectroscopy to evaluate odorous impact during the composting of different raw materials. *Journal of Cleaner Production*. 2017 Nov 20;167:154-62.

[90] Parthiba Karthikeyan O, Trably E, Mehariya S, Bernet N, Wong JWC, Carrere H. Pretreatment of food waste for methane and hydrogen recovery: A review. *Bioresource Technology*. 2018 Feb 1;249:1025-39.

[91] Goh CS, Tan KT, Lee KT, Bhatia S. Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresource Technology*. 2010;101(13):4834-41.

[92] Guerrero AB, Ballesteros I, Ballesteros M. The potential of agricultural banana waste for bioethanol production. *Fuel*. 2018;213(Supplement C):176-85.

[93] Gupta A, Verma JP. Sustainable bio-ethanol production from agro-residues: A review. *Renewable and Sustainable Energy Reviews*. 2015;41(Supplement C):550-67.

[94] Tan KT, Lee KT, Mohamed AR. Role of energy policy in renewable energy accomplishment: The case of second-generation bioethanol. *Energy Policy*. 2008;36(9):3360-5.

[95] Heredia-Guerrero JA, Athanassiou A. Editorial: Non-polysaccharide Plant Polymeric Materials. Vol. 3, *Frontiers in Materials*. Frontiers Media SA; 2016.

[96] Heredia-Guerrero JA, Heredia A, Domínguez E, Cingolani R, Bayer IS, Athanassiou A, et al. Cutin from agro-waste as a raw material for the production of bioplastics. *Journal*

of Experimental Botany. 2017 Nov 9;68(19):5401-10.

[97] Rujnić-Sokele M, Pilipović A. Challenges and opportunities of biodegradable plastics: A mini review. *Waste Management & Research*. 2017;35(2):132-40.

[98] Yaradoddi J, Patil V, Ganachari S, Banapurmath N, Hunashyal A, Shettar A, et al. Biodegradable Plastic Production From Fruit Waste Material and Its Sustainable Use for Green Applications. *International Journal of Pharmaceutical Research & Allied Sciences*. 2016;5(4):56-66.

[99] Gandini A, Lacerda TM, Carvalho AJF, Trovatti E. Progress of Polymers from Renewable Resources: Furans, Vegetable Oils, and Polysaccharides. *Chemical Reviews*. 2016;116(3):1637-69.

[100] de la Rosa JM, Rosado M, Paneque M, Miller AZ, Knicker H. Effects of aging under field conditions on biochar structure and composition: Implications for biochar stability in soils. *Science of the Total Environment*. 2018;613-614(Supplement C):969-76.

[101] Oliveira FR, Patel AK, Jaisi DP, Adhikari S, Lu H, Khanal SK. Environmental application of biochar: Current status and perspectives. *Bioresource Technology*. 2017;246(Supplement C):110-22.

[102] Volpe M, Fiori L. From olive waste to solid biofuel through hydrothermal carbonisation: The role of temperature and solid load on secondary char formation and hydrochar energy properties. *Journal of Analytical and Applied Pyrolysis*. 2017;124(Supplement C):63-72.

[103] Basso D, Patuzzi F, Weiss-Hortala E, Rada EC, Castello D, Baratieri M, et al. Waste to biofuel through hydrothermal carbonization. *Venice 2014: V*

International Symposium on Energy from Biomass and Waste. 2014;47(Part A):114-21.

[104] Benavente V, Calabuig E, Fullana A. Upgrading of moist agro-industrial wastes by hydrothermal carbonization. Journal of Analytical and Applied Pyrolysis. 2015;113 (Supplement C):89-98.