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Chapter

Drugs for the Treatment of Muscle Atrophy

Linlin Chen, Hong Zhang, Mengyi Chi, Quanjun Yang and Cheng Guo

Abstract

Muscle mass is maintained through an interplay between anabolic and catabolic pathways. The ubiquitin-proteasome system plays an important role in the proteolysis progress during skeletal muscle atrophy which can be blocked by some proteasome inhibitors. But few studies have demonstrated the ability of these inhibitors to preserve muscle mass and architecture under catabolic condition in vivo. The insulin-like growth factor-1/phosphatidylinositide 3-kinases/protein kinase B/ mammalian target of rapamycin (IGF-1/PI3K/Akt/mTOR) pathway was associated with anabolic pathways. The activation of IGF-1 causes muscle hypertrophy; however, it cannot be used as a drug target. Myostatin pathway maintains activation that can induce skeletal muscle atrophy involved with various transcriptional and genetic factors. Skeletal muscle atrophy is a debilitating consequence of multiple chronic diseases and conditions that involve starvation. It reduces treatment options and positive clinical outcomes as well as compromising quality of life and increasing morbidity and mortality. Though considerable research has been undertaken to find the drug target and the molecular mechanisms that improve skeletal muscle atrophy, no drug was approved to treat skeletal muscle atrophy. However, these years, the signaling pathways involved in muscle atrophy were clarified and some effective treatments were currently available to prevent, attenuate, or reverse muscle atrophy for experiment research.

Keywords: muscle atrophy, sarcopenia, cachexia, anabolic, catabolic

1. Introduction

The pathophysiology of skeletal muscle atrophy is multifactorial, with cancer, sepsis, renal and cardiac failure, acquired immune deficiency syndrome (AIDS) and chronic obstructive pulmonary disease (COPD) as well as inactivity or during aging [1–3]. These factors gradually lead to muscle wasting and weakness by decreasing protein synthesis and accelerating protein degradation, which are characterized by substantial decrease in myonuclear number, muscle fiber cross-sectional area, muscle strength and protein content while increasing in fatigability and resistance to insulin [4, 5]. Muscle atrophy is recognized as an independent predictor of mortality and is associated with functional impairment and poor quality of life [6].

Studies have revealed that different types of molecular mediators/catabolic players such as pro-inflammatory cytokines i.e. tumor necrosis factor- α (TNF- α), interleukin-6 (IL-6), interleukin-1 (IL-1), interferon gamma (IFN- γ) and

TNF-like weak inducer of apoptosis (TWEAK), eicosanoids and transforming growth factor-β (TGF-β) family effectors (such as activin A and myostatin) are involved in skeletal muscle atrophy under above mentioned clinical settings [7–9]. These cytokines binding to their respective receptor results in activation of several catabolic pathways including nuclear factor-kappa B (NF-κB), Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathways and small mothers against decapentaplegic homolog 2/3 (SMAD2/3). In addition to cytokines, growth factors such as insulin-like growth factor-1 (IGF-1) signal through anabolic pathway (phosphatidylinositide 3-kinases/protein kinase B/mammalian target of rapamycin; PI3K/Akt/mTOR) to mediate functional repression of the transcription factors forkhead box protein O1 (FoxO1) or FoxO3 by inhibiting their nuclear translocation and overall levels, which together inhibit the transcription of muscle atrophy genes [10]. In spite of many promising therapeutic targets for improving skeletal muscle

In spite of many promising therapeutic targets for improving skeletal muscle atrophy, no treatment has been successful to date. In this chapter, we classify the potential drugs currently in laboratory/preclinical research into four categories and then discuss their mechanism of action.

2. Anabolic medications

2.1 Androgen/androgen receptor modulators

Testosterone treatments increase muscle protein synthesis and fat free mass, and its effects on muscle are modulated by nutrition and exercise [11]. Several studies have shown the beneficial effects of testosterone supplementation on sarcopenia characteristics such as decreases in the muscle mass [12] and grip strength [13]. A study recently demonstrated that testosterone administration for 3 years in older men (over 60 years old) significantly improved stair-climbing power, muscle mass and power [14, 15]. Similarly, lower doses of testosterone supplementation in women with hysterectomy or chronic heart failure significantly increases lean body mass, 6-m walk time, chest press power and maximal voluntary contraction [16]. Evident showed that the effect of testosterone in improving skeletal muscle atrophy is related to the positive regulation of IGF-1 [12], wnt [17] and myostatin [18]. Although testosterone and its analogs can induce muscle growth and increase muscle strength [19], its clinical use is substantially limited by severe side effects including the increased risk of developing prostate hypertrophy, cancer, sleep apnea, masculinization, thrombosis complication and behavioral abnormalities [20, 21].

Compared with testosterone, the selective androgen receptor modulator (SARM) binds to androgen receptors with differing levels of sensitivity [22], showed androgenic effects in some tissues (such as muscle and bone), and has no effect on other organs (such as prostate or skin), thereby limiting adverse reactions such as prostate hypertrophy or androgen production. Enobosarm (GTx-024), an orally bioavailable nonsteroidal SARM, has been shown to increase lean body mass in phase I and II clinical trials of cancer cachexia patients [23, 24]. Moreover, the stimulation of reproductive organs with enobosarm seems to be less pronounced compared to testosterone administration. However, the phase III clinical trial of enobosarm failed to meet its common primary endpoint of preserving lean body mass and physical function [25]. Phase I clinical trials using another SARM non-steroidal oral preparation LGD-4033/VK5211 also showed increased muscle mass, but there was no effect on fat mass [26]. The 4-aza steroidal drug MK0773 (TFM-4AS-1) is a dual SARM and an inhibitor of 5α -reductase. Studies have shown that it can improve IGF-1 levels and muscle function in women, however,

the trial was terminated due to increased cardiovascular risk [27]. GSK2881078, which is assessed for its impact on muscle growth and strength, has completed its phase I trial [28] and phase II trial for the treatment of weakness caused by COPD (NCT03359473). The development of SARM drugs still requires long-term follow-up and/or more effective and selective SARM trials to prove the safety and efficacy of SARM in improving physical function and health outcomes.

2.2 Ghrelin and its receptor agonist

Ghrelin is a growth hormone (GH)-releasing polypeptide that binds to the GH secretagogue receptor (GHSR-1α) and stimulates appetite by activating the neuropeptide Y (NY) in the hypothalamus and helps in regulation of body weight [29, 30]. Studies have shown that ghrelin can reduce dexamethasone, fasting, denervation, cancer and cisplatin-induced muscle atrophy [31, 32]. In cachexia induced by lung adenocarcinoma, ghrelin treatment can reduce the expression of TNF-α, IL-1β, IL-6 and C-reactive protein, and inhibit skeletal muscle atrophy by restoring the expressions of the p-Akt and p-FoxO1, and reducing the expressions of p-p38 mitogen-activated protein kinase and p-NF-κB in skeletal muscle of tumor-bearing mice [33]. A three-week clinical study of ghrelin therapy in cachexia patients with nausea, COPD and chronic heart failure (CHF) showed an increase in lean body mass and muscle strength [29, 34]. Although ghrelin plays a key role in stimulating appetite, gaining body weight and preventing muscle catabolism, its clinical efficacy is limited due to its half-life (0.5 h) and route of administration (intravenous) [35].

Ghrelin agonists (such as anamorelin) have the advantage of oral activity. Compared with ghrelin (0.5 h), it has a better half-life (7–12 h) [36]. A randomized, double-blind, placebo-controlled phase I clinical study showed that anamorelin gained body weight after 6 days of treatment [37]. In two phase II anamorelin trials in cachectic patients with advanced or incurable cancer [38] and two multinational phase III trials (ROMANA 1 and 2 trials) in cachectic patients with unresectable non-small cell lung cancer (NSCLC) [39], significant gains were recorded in lean body mass and body weight over 12 weeks, but there was no improvement in physical functions and hand-grip strength. Similarly, a multicenter, open-label, single-arm study investigated the efficacy and safety of anamorelin in advanced gastrointestinal cancer patients with cancer cachexia, and this study showed a positive effect of anamorelin on lean body mass, body weight, anorexia and patients' nutritional status [40]. Furthermore, anamorelin treatment was well tolerated over 12 weeks. Finally, two meta-analyses also strongly supported the positive effect of anamorelin on lean body mass and body weight [41, 42]. Recently, a single-center study on healthy young adults showed anamorelin elicited modest increases in hunger and achieved significant increases in hunger and caloric intake [43]. The findings are consistent with multi-center findings in cachectic cancer patients and expand the evidence supporting anamorelin as a potential intervention.

2.3 β-Adrenoceptor agonists

Muscle growth can also be stimulated by activation of G-protein coupled $\beta 2$ -adrenoreceptor ($\beta 2$ -AR), which causes protein kinase A activation [44] and thereby stimulating PI3K/Akt/mTOR signaling [45]. Formoterol is a $\beta 2$ -AR agonist, the administration of formoterol significantly increased the levels of follistatin and decreased the levels of myostatin and its receptors (activin receptor IIB, ActRIIB) in tumor-bearing rats, thereby regulating muscle mass loss [46, 47]. In addition to skeletal muscle, formoterol also shows a strong protective effect on the

heart muscle [48]. Clinical studies have also shown that formoterol treatment can increase the content of PGC-1 α and mtDNA in skeletal muscle of COPD patients to enhance the oxidation process of skeletal muscle and improve exercise ability [49]. Clenbuterol is another β 2-AR agonist and can improve skeletal muscle atrophy in a variety of muscle atrophy models dominated by denervation [50], immobilization [51] and spinal cord injury [52]. However, due to concerns about potential cardiovascular side effects [44, 53], such as cardiac arrhythmia, there has been little interest in the clinical applications of β 2-AR agonists for muscle atrophy treatment. Among them, espindolol may be a potentially attractive compound. It is a β 1 receptor antagonist, a partial β 2 receptor agonist and also has 5-HT1a receptor activities. In old rats, espindolol has been shown to significantly increase muscle mass, while reducing fat mass without negatively affecting heart function [54]. In addition, it has also shown very promising results in phase IIa cancer cachexia studies leading to increased muscle mass and grip strength [55, 56].

3. Enzyme inhibitors

3.1 Cox2 inhibitors

Cox2 is a bifunctional enzyme with cyclooxygenase and peroxidase activities. Cyclooxygenase activity is responsible for the synthesis of prostaglandins (PGE2) from arachidonic acid, while peroxidase activity can produce adjacent carcinogens. Both Cox2 and PGE2 are downstream effectors of cytokine activity and mediate cachexia [57]. A placebo-controlled study of celecoxib (Cox2 inhibitor) on cachectic patients with either head and neck or gastrointestinal cancer showed a significant increase of body mass and the quality of life [58]. In addition, a phase II non-randomized trial examined the efficacy and safety of celecoxib on cancer cachexia. Celecoxib administered at 300 mg/day for 4 months induced a significant increase of lean body mass, a decrease of serum TNF- α levels, and a trend toward a reduction of fatigue symptom [59]. Moreover, side effects such as grade 1/2 anemia, neuropathy and epigastralgia have been observed in only a few patients, and no grade 3/4 adverse events have been observed. Recently, a randomized double-blind clinical trial of combined treatment with megestrol acetate plus celecoxib versus megestrol acetate alone in cachexia-anorexia syndrome induced by gastrointestinal cancers was performed, however, this study failed to show that adding celecoxib (200 mg/day) to megestrol (320 mg/day) could enhance anti-cachexic effects of megestrol [60]. Meloxicam is another Cox2 inhibitor, and can suppress the expression of Cox2, Atrogin-1 and MuRF1 induced by lipopolysaccharide (LPS), and regulate the loss of muscle mass in rats by attenuating protein degradation [61]. In addition to cachexia, the administration of meloxicam can also inhibit the upregulation of Atrogin-1 and MuRF1 in the muscles of arthritis rats and improve the loss of muscle mass [62].

3.2 Histone deacetylase inhibitors

Trichostatin A (TSA) is a well-known class I and II histone deacetylase inhibitor. Published data indicate that TSA regulates atrogenes level and controls muscle mass by reducing HDAC4 activity and myogenin expression, and increasing Dach2 level under denervation condition (neuromuscular disorders) [63]. TSA treatment can improve body weight, myofiber cross-sectional area and myofiber number [64]. Recent report shows that TSA inactivates FoxO by inhibiting HDAC activity,

which leads to atrophy of skeletal muscle atrophy and contractile dysfunction [65]. In addition, under nutrition-deprived atrophy on C2C12 myotubes, TSA treatment leads to the suppression of FoxO target genes, including Lc3 (autophagy marker), MuRF1 and Atrogin-1 [66]. Similarly, TSA treatment can regulate muscle depletion by inhibiting the levels of Atrogin-1 and MuRF1 in dexamethasone-induced atrophic mice [63]. However, study shows that TSA treatment increased the expression level of follistatin (a negative regulator of skeletal muscle development), without retaining or increasing muscle mass in tumor-bearing mice [67]. Recent studies have shown that TSA can inhibit skeletal muscle atrophy and histomorphological alterations induced by unloading [68] and cigarette smoke (the main risk factor for COPD) [69]. Due to the contradictory findings, further research is needed to confirm the use of HDAC blockers to regulate atrophy.

3.3 PDE inhibitors

Torbafylline (HWA 448) is a xanthine derivative which acts as a phosphodiesterase (PDE) inhibitor [70]. Torbafylline treatment down-regulates the mRNA expression of cathepsin L, calpain and E3 ligases, and regulates the proteolytic pathway in burn-induced injury. In addition, the anti-atrophic effects of torbafylline have been demonstrated in casting, denervation or cancer induced cachexia models [70–72]. Torbafylline inhibits PDE activity leading to stimulation of the anti-proteolytic effect in PDE4/cAMP/Epac/PI3K/Akt pathway-mediated muscle atrophy [73]. Pentoxifylline (PTX) is another xanthine derivative that is non-selective in inhibiting PDE. Published data indicate that the administration of PTX under various pathological conditions in animal models (diabetes, tumors, sepsis) can stimulate the formation of cAMP, and by down-regulating calpain, cathepsin L and proteasome proteolytic system activity [74–76]. Other selective inhibitors of PDE, including rolipram and cilomilast have also been shown to reduce muscle atrophy in denervation and casting animal models [77, 78].

3.4 Angiotensin-converting enzyme inhibitors

ANGII induces muscle atrophy through several mechanisms including suppresses protein anabolism by reducing IGF-1 level and appetite, and promotes protein catabolism by increasing reactive oxygen species (ROS) and intermediate molecules (TNF-α, IL-6, glucocorticoids) in skeletal muscle [79]. In ACE-Is, enalapril treatment can reduce the risk of weight loss by>19% and delay the occurrence of cachexia by about 8 months [80]. Studies conducted in an old rat model show that the administration of enalapril can increase muscle strength and has a protective effect on age-related muscle degeneration [81]. Perindopril (an ACE inhibitor) has shown especially in a double-blind randomized controlled trial, which evaluated the effect of perindopril on the elderly 6-minute walking distance, thereby improving physical function, especially the 6-minute walk distance and reduced the incidence of hip fractures [82]. In subjects with dysfunction, perindopril improved exercise capacity to the extent reported after 6 months of exercise training [83]. However, the use of the perindopril in cachectic mice bearing colon-26 tumors to inhibit this pathway does not reduce muscle atrophy, nor does it increase the production of maximum muscle strength. Nonetheless, treatment with ACE inhibitors did enhance physical function and reduce fatigue of respiratory muscles. These effects appear to be due to a shift to a more oxidized muscle phenotype, as evident from increased oxidative enzyme capacity in the muscle cross-section [84].

4. Anti-inflammatory drugs

4.1 Thalidomide

Thalidomide is a glutamic acid derivative with various pharmacological activities, such as anti-inflammatory, immunomodulatory, anti-angiogenic, anti-emetic and sedative effects. Report shows that thalidomide and its derivatives can inhibit Cox2 and PGE2 synthesis induced by LPS in murine macrophages [85], and control systemic inflammation. In addition, evidence shows that thalidomide can reduce serum IL-6 and CRP levels in patients with cancer cachexia [86, 87]. Another study showed that thalidomide can maintain the fast-twitch type myofibers by reducing the expression of TNF- α and TGF- β 1 in soleus muscle of cholangiocarcinoma rats [88]. Down-regulation of NF-κB/iNOS pathway by chronic thalidomide treatment improves hepatopulmonary syndrome and skeletal muscle atrophy in rats with biliary cirrhosis [89]. In addition to anti-inflammatory and anti-cachectic activity, thalidomide treatment (Phase II trial) also showed an effect on appetite in 64% of patients with advanced stage of cancer [87]. Studies reported that thalidomide (100 mg/day and 200 mg/day) treatment showed a significant improvement in body weight and skeletal muscle atrophy in AIDS associated cachexia patients [90]. Similarly, another research team worked with pancreatic cachexia patients and observed a significant increase in body weight of patients treated with thalidomide [91]. The lack of benefits was mainly due to the drug toxicity of thalidomide including peripheral neuropathy, dizziness, constipation and rash, considering that 47% of patients receiving active treatment were unable to continue taking thalidomide due to side effects and disease-related morbidity [92].

4.2 Anti-IL-6/STAT3

Evidence has shown that antibodies against IL-6 or its receptor can effectively reduce skeletal muscle atrophy and cachexia in mouse models [93, 94]. Preliminary results of a phase II double-blind trial in patients with advanced NSCLC have shown that ALD518 (humanized IL-6 monoclonal antibody) can reverse fatigue and prevent muscle loss [95]. Tocilizumab is an IL-6 receptor (IL-6R) neutralizing antibody approved by the FDA for rheumatoid arthritis. It can destroy the binding of IL-6/IL-6R to GP130, and cause the decrease of JAK/STAT3 pathway activity, reduce B cell hyperactivity and lead to a dramatic normalization of the acute phase reactions [96, 97]. Pharmacologic inhibition of the IL-6R using tocilizumab antennas skeletal muscle atrophy and function loss during infection [98]. Recently, a case of 65-year-old man who underwent percutaneous coronary intervention for acute myocardial infarction received tocilizumab led to prompt remission of Takayasu arteritis activity and improvement of left ventricular function and skeletal muscle atrophy [99]. Ruxolitinib, a JAK1/2 inhibitor, may protect muscle through on-target effects because it significantly reduces IL-6-induced STAT3 activation and myotube atrophy in vitro [100]. However, due to the inability to recruit qualified patients, the clinical trial of cancer patient study (NCT02072057) that investigating whether blocking downstream signaling of IL-6 by ruxolitinib improves muscle atrophy were terminated. In addition, there is evidence that C188-9 (a small molecule of STAT3 inhibitor) can reduce skeletal muscle atrophy in tumor-bearing mice [101, 102], but there are no relevant clinical studies.

4.3 Anti-TNF-α

Studies have shown that the administration of anti-murine TNF IgG in rats bearing Yoshida AH-130 ascites hepatoma can reduce circulating TNF- α and

inhibit muscle protein degradation [103]. Similarly, injecting soluble TNF receptors (sTNFR1, a specific inhibitor of TNF- α) prevents the interaction of TNF- α with its receptor and attenuates ubiquitin transcription, reduce the waste of skeletal muscle and preserve body weight in cardiac cachexia [76]. A study reported the opposite effect of sTNFR1 on arthritic rat that it did not alter muscle mass and MuRF1 and Atrogin-1 gene expression [104]. Infliximab is a chimeric monoclonal antibody that blocks TNF- α action, thereby preventing its binding to cellular receptors and downstream immunological effects. A phase II study of the combined chemotherapy drugs gemcitabine and infliximab did not show the benefit of maintaining lean body mass or survival in pancreatic cancer cachexia patients [105]. Interestingly, in clinical trials of Crohn's disease patients with skeletal muscle atrophy or sarcopenia arising from chronic inflammation, significant gains were recorded in muscle volume and strength over 25 weeks of infliximab treatment [106, 107]. Etanercept is a recombinant fusion protein that acts as a decoy receptor to neutralize TNF- α , and has been used to treat inflammatory diseases including rheumatoid arthritis. In another study, significant weight gain was observed in rheumatoid arthritis patients who received etanercept twice a week for 12 consecutive months [108]. A phase I/II study compared the efficacy of etanercept with gemcitabine and gemcitabine alone for the treatment of advanced pancreatic cancer cachexia patients, and the results were also disappointing because the addition of etanercept did not improve symptoms of cancer cachexia [109].

4.4 Anti-IL-1α

MABp1 is a human antibody against IL-1 α (a chronic inflammatory mediator) and has anti-tumor activity. Intravenous MABp1 treatment for 8 weeks in adults with metastatic solid cancer showed increased lean body mass and improved quality of life (fatigue, pain, and loss of appetite), and has no toxic; however, there was no control group in this study [110]. A randomized, double-blind, placebo-controlled phase III clinical study showed that MABp1 improved the lean body mass, anorexia, fatigue and pain scores in advanced colorectal cancer patients [111]. Another phase I dose-escalation study evaluating the IL-1 α -targeted monoclonal antibody xilonix in patients with NSCLC showed increased lean body mass and improved symptoms, suggesting a clinically important response [112]. In view of this, a phase III placebocontrolled study of human antibodies against IL-1α has been conducted in patients with advanced colorectal cancer to assess the remission rate of the disease, muscle mass and appetite. Xilonix was very well tolerated by NSCLC patients, with the clinically significant reductions in pain, fatigue and improved lean body mass and appetite [113]. However, the primary limitation of this report is the small number of patients which made any comparisons statistically difficult.

4.5 TWEAK/Fn14 inhibition

The inflammatory cytokine TNF-like weak inducer of apoptosis (TWEAK) and its related receptor fibroblast growth factor-inducible 14 (Fn14) play multiple roles in proliferation, inflammation and wound repair. TWEAK/Fn14 signaling also negatively regulates muscle growth and function [8]. Report showed that TWEAK activates noncanonical NF-κB pathway and promotes myoblast fusion at low concentrations (10 or 100 ng/ml), and activates canonical NF-κB signaling to inhibit differentiation at high concentrations (500 ng/ml). Thus, TWEAK can maintain myoblast differentiation at physiological conditions; however, under pathological conditions (such as denervation and disuse), TWEAK/Fn14 system becomes activated and causes muscle atrophy [8]. Blocking antibodies against TWEAK antibody

can improve muscle function in mice caused by myotonic dystrophy and amyotrophic lateral sclerosis (ALS) [114, 115]. Consistent with these findings, colon-26 tumor-bearing mice treated with anti-Fn14 antibodies showed increased weight and muscle mass, improved muscle fatigue, and increased survival [116]. These results indicate that neutralizing antibodies against TWEAK and Fn14 should be further explored in various muscle atrophy models and clinical trials.

5. Other investigational drugs

5.1 Myostatin inhibition

Existing evidence indicates that members of the TGF- β superfamily, such as myostatin and activin A, are powerful catabolic stimuli that can inhibit muscle growth and promote muscle protein loss in various disease states [117]. It is reported that myostatin can improve the dystrophy phenotype of mdx mouse models, sarcopenia in aging mouse models and muscle atrophy in tumor-bearing mice [118, 119], which can significantly inhibit systemic inflammation and prolong the survival of tumor-bearing mice without affecting tumor growth [117]. There are currently two main strategies for targeting myostatin signals: First, neutralize myostatin directly by using humanized myostatin antibody (LY2495655), and second, block ActRII by using soluble ActRIIB (ACE-031) or ActRII antibody (bimagrumab/BMY338). LY2495655 treatment had mixed results in elderly subjects: the appendicular lean body mass and gait speed were slightly improved, and despite increased muscle mass, grip strength was not affected [120]. However, a randomized, phase II trial in patients with pancreatic cancer, LY2495655 treatment have no significant improvement in muscle volume or functional. Additionally, among possibly drug-related adverse events, fatigue, diarrhea, and anorexia were more common in LY2495655treated than in placebo-treated patients [121]. Soluble recombinant ActRIIB and other "ligand trap" interventions can generally inhibit TGF-β signaling and affect other tissues and processes, including reproduction and angiogenesis, with some causing severe off-target effects. Therefore, new strategies that target myostatin receptors and thereby reduce the activity of other ligands seem more promising. For example, after ACE-031 treatment, a group of 48 postmenopausal women gained weight and increased lean body mass [122]. However, in the phase II clinical trial conducted by ACE031 with Duchenne muscular dystrophy (DMD) patients and healthy volunteers, some participants experienced bleeding gums, nosebleeds, and skin vasodilation, which led to the interruption of the trial [123]. Blocking ActRII by administering BMY338 can greatly increase muscle mass and prevent dexamethasone-induced muscle atrophy in mice [124], and significantly improve patient's lean body mass, muscle mass, and 6-minute walking test in patients with myositis after 8 weeks of treatment. However, no significant differences were observed after 24 weeks of treatment [125]. In addition, there are no beneficial effects on these treatments were reported in cancer patients, while in COPD patients, muscle volume increases without affecting functional indicators, which is similar to the effect of BMY338 in sarcopenia patients [126, 127]. Therefore, these treatments seem to improve muscle mass and have less effect on muscle strength and other functional parameters [128].

5.2 Appetite stimulants

The FDA approved megestrol acetate (MA) as the treatment of cachexia caused by cancer and AIDS in 1993. More than 15 clinical trials have shown that this drug

can significantly improve appetite and lean body mass at a dose of 160-1600 mg/day. MA can be used alone or as a supplement along with meloxicam in patients with cancer cachexia, showing a positive effect in controlling weight loss [129]. Although the mechanism of appetite stimulation/weight gain is unclear, studies have shown that it is related to the involvement of neuropeptide Y and the inhibition of pro-inflammatory cytokines such as IL-1, IL-6 and TNF- α [130, 131]. However, a newer meta-analysis started in 2015, which studied the use of non-cancer cachexia (HIV, COPD, renal failure and geriatric cachexia) and concluded that progesterone therapy (MA or medroxyprogesterone acetate) has a negligible effect on weight gain when treat of non-cancer cachexia [132]. MA treatment can also cause serious side effects such as thromboembolism, peripheral edema, hyperglycemia, hypertension, adrenal suppression and adrenal insufficiency [132].

Previous studies have shown that cannabinoids have the potential to improve appetite, body weight and fat mass, as well as amelioration of quality of life in several chronic diseases including cancer [133]. The results of a pilot study conducted in adult patients with advanced solid tumors showed that patients receiving delta-9-tetrahydrocannabinol (THC) treatment had a marked increase in appetite [134]. However, the study did not record changes in participants' body weight and lean body mass, and a larger trial was needed to study the effect of cannabinoids on skeletal muscle atrophy. A pilot study in patients with advanced NSCLC showed food intake and quality of life in patients treated with nabilone (a tetrahydrocannabinol) have improved significantly [135]. However, another randomized, double-blind placebo-controlled trial showed that nabilone did not improve the symptoms of nausea during radiotherapy in head and neck cancer patients, nor did it have significant benefits for the appetite and body weight [136].

5.3 Natural compounds

Recently, growing evidence has shown that natural products play a key role in the prevention and treatment of skeletal muscle atrophy. Numerous studies conducted in vitro and in vivo confirmed that resveratrol treatment can prevent proteolysisinducing factor (PIF), angiotensin I and II, phorbol ester, 12-O-tetradecanoylphorbol 13-acetate (TPA), and dexamethasone-induced protein degradation [137]. In addition, resveratrol has been shown to protect muscle atrophy under various catabolic conditions, including cachexia and disuse [138, 139]. Salidroside is one of the main phenylpropane glycosides found in *Rhodiola rosea*. Research shows that salidroside treatment can effectively maintain body weight, reduce fat and gastrocnemius muscle loss in CT26 and LLC models. Additionally, in combination chemotherapy, salidroside can synergistically enhance the anti-tumor activity of cisplatin, especially reduce or eliminate cachexia caused by chemotherapy. Further analysis showed that salidroside can significantly increase the expression of p-mTOR and MyHC in the gastrocnemius muscle [140]. Matrine improves skeletal muscle atrophy in CT26 induced cachexia via inhibiting the production of TNF-α and IL-6 and activating the Akt/mTOR/FoxO3α signaling pathway [141]. Other natural medicines reported to improve skeletal muscle atrophy include imperatorin [142], parthenolide [143], ursolic acid [144] and cryptotanshinone [145], but more research is still needed to prove the anti-muscular atrophy effect of these compounds.

6. Conclusions

Up-regulation of muscle protein catabolic is a sign of atrophy, so most potential drugs target the proteolytic system to cure or prevent skeletal muscle atrophy.

Due to the multifactorial pathogenesis of muscle atrophy, combining new drugs with multimodal transport interventions including exercise methods and nutritional interventions may be the most promising approach; however, few clinical trials have investigated this approach. In this light, a better understanding of the contributing factors and underlying mechanisms of muscle atrophy is essential for the development of targeted therapies, and new methods of combination therapy for muscle atrophy treatment are needed.

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Conflict of interest

The authors declare no conflict of financial interest.



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References

- [1] Jackman RW, Kandarian SC. The molecular basis of skeletal muscle atrophy. American Journal of Physiology. Cell Physiology. 2004;287:C834-C843. DOI: 10.1152/ajpcell.00579.2003
- [2] Lecker SH, Goldberg AL, Mitch WE. Protein degradation by the ubiquitin-proteasome pathway in normal and disease states. Journal of the American Society of Nephrology: JASN. 2006;17:1807-1819. DOI: 10.1681/ asn.2006010083
- [3] Scott D. Sarcopenia in older adults. Journal of Clinical Medicine. 2019;8: 1844. DOI: 10.3390/jcm8111844
- [4] Dupont-Versteegden EE. Apoptosis in muscle atrophy: Relevance to sarcopenia. Experimental Gerontology. 2005;**40**:473-481. DOI: 10.1016/j. exger.2005.04.003
- [5] Biolo G, Cederholm T, Muscaritoli M. Muscle contractile and metabolic dysfunction is a common feature of sarcopenia of aging and chronic diseases: From sarcopenic obesity to cachexia. Clinical Nutrition (Edinburgh, Scotland). 2014;33:737-748. DOI: 10.1016/j.clnu.2014.03.007
- [6] Fearon K, Strasser F, Anker SD, et al. Definition and classification of cancer cachexia: An international consensus. The Lancet Oncology. 2011;12:489-495. DOI: 10.1016/s1470-2045(10)70218-7
- [7] Llovera M, Carbó N, López-Soriano J, et al. Different cytokines modulate ubiquitin gene expression in rat skeletal muscle. Cancer Letters. 1998;**133**:83-87. DOI: 10.1016/s0304-3835(98)00216-x
- [8] Mittal A, Bhatnagar S, Kumar A, et al. The TWEAK-Fn14 system is a critical regulator of denervation-induced skeletal muscle atrophy in

- mice. The Journal of Cell Biology. 2010;**188**:833-849. DOI: 10.1083/jcb.200909117
- [9] Frost RA, Lang CH. Protein kinase B/Akt: A nexus of growth factor and cytokine signaling in determining muscle mass. Journal of Applied Physiology (Bethesda, MD: 1985). 2007;103:378-387. DOI: 10.1152/japplphysiol.00089.2007
- [10] Schiaffino S, Dyar KA, Ciciliot S, Blaauw B, Sandri M. Mechanisms regulating skeletal muscle growth and atrophy. The FEBS Journal. 2013;280:4294-4314. DOI: 10.1111/febs.12253
- [11] Bhasin S, Woodhouse L, Storer TW. Proof of the effect of testosterone on skeletal muscle. The Journal of Endocrinology. 2001;**170**:27-38. DOI: 10.1677/joe.0.1700027
- [12] Ferrando AA, Sheffield-Moore M, Yeckel CW, et al. Testosterone administration to older men improves muscle function: Molecular and physiological mechanisms. American Journal of Physiology. Endocrinology and Metabolism. 2002;**282**:E601-E607. DOI: 10.1152/ajpendo.00362.2001
- [13] Bakhshi V, Elliott M, Gentili A, Godschalk M, Mulligan T. Testosterone improves rehabilitation outcomes in ill older men. Journal of the American Geriatrics Society. 2000;48:550-553. DOI: 10.1111/j.1532-5415.2000. tb05002.x
- [14] Storer TW, Basaria S, Traustadottir T, et al. Effects of testosterone supplementation for 3 years on muscle performance and physical function in older men. The Journal of Clinical Endocrinology and Metabolism. 2017;**102**:583-593. DOI: 10.1210/ jc.2016-2771

- [15] Iellamo F, Volterrani M, Caminiti G, et al. Testosterone therapy in women with chronic heart failure: A pilot double-blind, randomized, placebocontrolled study. Journal of the American College of Cardiology. 2010;56:1310-1316. DOI: 10.1016/j.jacc.2010.03.090
- [16] Huang G, Basaria S, Travison TG, et al. Testosterone dose-response relationships in hysterectomized women with or without oophorectomy: Effects on sexual function, body composition, muscle performance and physical function in a randomized trial. Menopause (New York, NY). 2014;21:612-623. DOI: 10.1097/gme.00000000000000003
- [17] Singh R, Bhasin S, Braga M, et al. Regulation of myogenic differentiation by androgens: Cross talk between androgen receptor/beta-catenin and follistatin/transforming growth factor-beta signaling pathways. Endocrinology. 2009;**150**:1259-1268. DOI: 10.1210/en.2008-0858
- [18] Mendler L, Baka Z, Kovács-Simon A, Dux L. Androgens negatively regulate myostatin expression in an androgen-dependent skeletal muscle. Biochemical and Biophysical Research Communications. 2007;361:237-242. DOI: 10.1016/j.bbrc.2007.07.023
- [19] Ferrando AA, Sheffield-Moore M, Paddon-Jones D, Wolfe RR, Urban RJ. Differential anabolic effects of testosterone and amino acid feeding in older men. The Journal of Clinical Endocrinology and Metabolism. 2003;88:358-362. DOI: 10.1210/jc.2002-021041
- [20] Khoo TK. Adverse events associated with testosterone administration. The New England Journal of Medicine. 2010;363:1865-1866; author reply 1866-1867. DOI: 10.1056/NEJMc1009326
- [21] Curran MJ, Bihrle W III. Dramatic rise in prostate-specific antigen after

- androgen replacement in a hypogonadal man with occult adenocarcinoma of the prostate. Urology. 1999;53:423-424. DOI: 10.1016/s0090-4295(98)00348-3
- [22] Mohler ML, Bohl CE, Jones A, et al. Nonsteroidal selective androgen receptor modulators (SARMs): Dissociating the anabolic and androgenic activities of the androgen receptor for therapeutic benefit. Journal of Medicinal Chemistry. 2009;52:3597-3617. DOI: 10.1021/jm900280m
- [23] Kim J, Wu D, Hwang DJ, Miller DD, Dalton JT. The para substituent of S-3-(phenoxy)-2-hydroxy-2-methyl-N-(4-nitro-3-trifluoromethyl-phenyl)-propionamides is a major structural determinant of in vivo disposition and activity of selective androgen receptor modulators. The Journal of Pharmacology and Experimental Therapeutics. 2005;315:230-239. DOI: 10.1124/jpet.105.088344
- [24] Dobs AS, Boccia RV, Croot CC, et al. Effects of enobosarm on muscle wasting and physical function in patients with cancer: A double-blind, randomised controlled phase 2 trial. The Lancet Oncology. 2013;14:335-345. DOI: 10.1016/s1470-2045(13)70055-x
- [25] Crawford J, Prado CM, Johnston MA, et al. Study design and rationale for the phase 3 clinical development program of enobosarm, a selective androgen receptor modulator, for the prevention and treatment of muscle wasting in cancer patients (POWER trials). Current Oncology Reports. 2016;18:37. DOI: 10.1007/s11912-016-0522-0
- [26] Basaria S, Collins L, Dillon EL, et al. The safety, pharmacokinetics, and effects of LGD-4033, a novel nonsteroidal oral, selective androgen receptor modulator, in healthy young men. The Journals of Gerontology Series A Biological Sciences and Medical Sciences. 2013;68:87-95. DOI: 10.1093/gerona/gls078

- [27] Papanicolaou DA, Ather SN, Zhu H, et al. A phase IIA randomized, placebo-controlled clinical trial to study the efficacy and safety of the selective androgen receptor modulator (SARM), MK-0773 in female participants with sarcopenia. The Journal of Nutrition, Health and Aging. 2013;17:533-543. DOI: 10.1007/s12603-013-0335-x
- [28] Neil D, Clark RV, Magee M, et al. GSK2881078, a SARM, produces dose-dependent increases in lean mass in healthy older men and women. The Journal of Clinical Endocrinology and Metabolism. 2018;103:3215-3224. DOI: 10.1210/jc.2017-02644
- [29] Nagaya N, Itoh T, Murakami S, et al. Treatment of cachexia with ghrelin in patients with COPD. Chest. 2005;**128**:1187-1193. DOI: 10.1378/chest.128.3.1187
- [30] Barazzoni R, Zhu X, Deboer M, et al. Combined effects of ghrelin and higher food intake enhance skeletal muscle mitochondrial oxidative capacity and AKT phosphorylation in rats with chronic kidney disease. Kidney International. 2010;77:23-28. DOI: 10.1038/ki.2009.411
- [31] Porporato PE, Filigheddu N, Reano S, et al. Acylated and unacylated ghrelin impair skeletal muscle atrophy in mice. The Journal of Clinical Investigation. 2013;**123**:611-622. DOI: 10.1172/jci39920
- [32] Chen JA, Splenser A, Guillory B, et al. Ghrelin prevents tumour- and cisplatin-induced muscle wasting: Characterization of multiple mechanisms involved. Journal of Cachexia, Sarcopenia and Muscle. 2015;6:132-143. DOI: 10.1002/jcsm.12023
- [33] Tsubouchi H, Yanagi S, Miura A, Matsumoto N, Kangawa K, Nakazato M. Ghrelin relieves cancer cachexia associated with the development of

- lung adenocarcinoma in mice. European Journal of Pharmacology. 2014;**743**:1-10. DOI: 10.1016/j.ejphar.2014.09.025
- [34] Nagaya N, Moriya J, Yasumura Y, et al. Effects of ghrelin administration on left ventricular function, exercise capacity, and muscle wasting in patients with chronic heart failure. Circulation. 2004;**110**:3674-3679. DOI: 10.1161/01. Cir.0000149746.62908.Bb
- [35] Strasser F, Lutz TA, Maeder MT, et al. Safety, tolerability and pharmacokinetics of intravenous ghrelin for cancer-related anorexia/cachexia: A randomised, placebo-controlled, double-blind, double-crossover study. British Journal of Cancer. 2008;**98**:300-308. DOI: 10.1038/sj.bjc.6604148
- [36] Pietra C, Takeda Y, Tazawa-Ogata N, et al. Anamorelin HCl (ONO-7643), a novel ghrelin receptor agonist, for the treatment of cancer anorexia-cachexia syndrome: Preclinical profile. Journal of Cachexia, Sarcopenia and Muscle. 2014;5:329-337. DOI: 10.1007/s13539-014-0159-5
- [37] Garcia JM, Friend J, Allen S. Therapeutic potential of anamorelin, a novel, oral ghrelin mimetic, in patients with cancer-related cachexia: A multicenter, randomized, doubleblind, crossover, pilot study. Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer. 2013;21:129-137. DOI: 10.1007/s00520-012-1500-1
- [38] Garcia JM, Boccia RV, Graham CD, et al. Anamorelin for patients with cancer cachexia: An integrated analysis of two phase 2, randomised, placebocontrolled, double-blind trials. The Lancet Oncology. 2015;**16**:108-116. DOI: 10.1016/s1470-2045(14)71154-4
- [39] Temel JS, Abernethy AP, Currow DC, et al. Anamorelin in patients with non-small-cell lung cancer and cachexia (ROMANA

- 1 and ROMANA 2): Results from two randomised, double-blind, phase 3 trials. The Lancet Oncology. 2016;17:519-531. DOI: 10.1016/ s1470-2045(15)00558-6
- [40] Hamauchi S, Furuse J, Takano T, et al. A multicenter, open-label, single-arm study of anamorelin (ONO-7643) in advanced gastrointestinal cancer patients with cancer cachexia. Cancer. 2019;125:4294-4302. DOI: 10.1002/cncr.32406
- [41] Nishie K, Yamamoto S, Nagata C, Koizumi T, Hanaoka M. Anamorelin for advanced non-small-cell lung cancer with cachexia: Systematic review and meta-analysis. Lung Cancer (Amsterdam, Netherlands). 2017;112:25-34. DOI: 10.1016/j. lungcan.2017.07.023
- [42] Bai Y, Hu Y, Zhao Y, et al. Anamorelin for cancer anorexiacachexia syndrome: A systematic review and meta-analysis. Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer. 2017;25:1651-1659. DOI: 10.1007/s00520-016-3560-0
- [43] Blum RA, Mair S, Duus EM. Appetite and food intake results from phase I studies of anamorelin. Journal of Cachexia, Sarcopenia and Muscle. 2019;10:1027-1035. DOI: 10.1002/jcsm.12439
- [44] Lynch GS, Ryall JG. Role of beta-adrenoceptor signaling in skeletal muscle: Implications for muscle wasting and disease. Physiological Reviews. 2008;88:729-767. DOI: 10.1152/physrev.00028.2007
- [45] Sandri M. Signaling in muscle atrophy and hypertrophy. Physiology (Bethesda, Md.). 2008;**23**:160-170. DOI: 10.1152/physiol.00041.2007
- [46] Quanjun Y, Genjin Y, Lili W, et al. Serum metabolic profiles reveal

- the effect of formoterol on cachexia in tumor-bearing mice. Molecular BioSystems. 2013;**9**:3015-3025. DOI: 10.1039/c3mb70134d
- [47] Busquets S, Toledo M, Marmonti E, et al. Formoterol treatment downregulates the myostatin system in skeletal muscle of cachectic tumour-bearing rats. Oncology Letters. 2012;3:185-189. DOI: 10.3892/ol.2011.442
- [48] Toledo M, Springer J, Busquets S, et al. Formoterol in the treatment of experimental cancer cachexia: Effects on heart function. Journal of Cachexia, Sarcopenia and Muscle. 2014;5:315-320. DOI: 10.1007/s13539-014-0153-y
- [49] D'Agostino B, Polverino M, Cirino G, et al. Exercise capacity and cytochrome oxidase activity in muscle mitochondria of COPD patients. Respiratory Medicine. 2010;**104**:83-90. DOI: 10.1016/j.rmed.2009.07.016
- [50] Gonçalves DA, Silveira WA, Lira EC, et al. Clenbuterol suppresses proteasomal and lysosomal proteolysis and atrophy-related genes in denervated rat soleus muscles independently of Akt. American Journal of Physiology. Endocrinology and Metabolism. 2012;302:E123-E133. DOI: 10.1152/ ajpendo.00188.2011
- [51] Suzuki H, Yoshikawa Y, Tsujimoto H, Kitaura T, Muraoka I. Clenbuterol accelerates recovery after immobilization-induced atrophy of rat hindlimb muscle. Acta Histochemica. 2020;122:151453. DOI: 10.1016/j. acthis.2019.151453
- [52] Ung RV, Rouleau P, Guertin PA. Functional and physiological effects of treadmill training induced by buspirone, carbidopa, and L-DOPA in clenbuterol-treated paraplegic mice. Neurorehabilitation and Neural Repair. 2012;**26**:385-394. DOI: 10.1177/1545968311427042

- [53] Brett J, Dawson AH, Brown JA. Clenbuterol toxicity: A NSW poisons information centre experience. The Medical Journal of Australia. 2014;200:219-221. DOI: 10.5694/mja13.10982
- [54] Pötsch MS, Tschirner A, Palus S, et al. The anabolic catabolic transforming agent (ACTA) espindolol increases muscle mass and decreases fat mass in old rats. Journal of Cachexia, Sarcopenia and Muscle. 2014;5:149-158. DOI: 10.1007/s13539-013-0125-7
- [55] Lainscak M, Laviano A. ACT-ONE ACTION at last on cancer cachexia by adapting a novel action beta-blocker. Journal of Cachexia, Sarcopenia and Muscle. 2016;7:400-402. DOI: 10.1002/jcsm.12136
- [56] Stewart Coats AJ, Ho GF, Prabhash K, et al. Espindolol for the treatment and prevention of cachexia in patients with stage III/IV non-small cell lung cancer or colorectal cancer: A randomized, double-blind, placebocontrolled, international multicentre phase II study (the ACT-ONE trial). Journal of Cachexia, Sarcopenia and Muscle. 2016;7:355-365. DOI: 10.1002/ jcsm.12126
- [57] Baumgarten AJ, Fiebig HH, Burger AM. Molecular analysis of xenograft models of human cancer cachexia--Possibilities for therapeutic intervention. Cancer Genomics & Proteomics. 2007;4:223-231
- [58] Lai V, George J, Richey L, et al. Results of a pilot study of the effects of celecoxib on cancer cachexia in patients with cancer of the head, neck, and gastrointestinal tract. Head & Neck. 2008;30:67-74. DOI: 10.1002/hed.20662
- [59] Mantovani G, Macciò A, Madeddu C, et al. Phase II nonrandomized study of the efficacy and safety of COX-2 inhibitor celecoxib

- on patients with cancer cachexia. Journal of Molecular Medicine (Berlin, Germany). 2010;88:85-92. DOI: 10.1007/s00109-009-0547-z
- [60] Kouchaki B, Janbabai G, Alipour A, Ala S, Borhani S, Salehifar E.
 Randomized double-blind clinical trial of combined treatment with megestrol acetate plus celecoxib versus megestrol acetate alone in cachexia-anorexia syndrome induced by GI cancers.
 Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer.
 2018;26:2479-2489. DOI: 10.1007/s00520-018-4047-y
- [61] Martin AI, Nieto-Bona MP, Castillero E, et al. Effect of cyclooxygenase-2 inhibition by meloxicam, on atrogin-1 and myogenic regulatory factors in skeletal muscle of rats injected with endotoxin. Journal of Physiology and Pharmacology: An Official Journal of the Polish Physiological Society. 2012;63:649-659
- [62] Granado M, Martín AI, Villanúa MA, López-Calderón A. Experimental arthritis inhibits the insulin-like growth factor-I axis and induces muscle wasting through cyclooxygenase-2 activation. American Journal of Physiology. Endocrinology and Metabolism. 2007;292:E1656-E1665. DOI: 10.1152/ajpendo.00502.2006
- [63] Bricceno KV, Sampognaro PJ, Van Meerbeke JP, Sumner CJ, Fischbeck KH, Burnett BG. Histone deacetylase inhibition suppresses myogenindependent atrogene activation in spinal muscular atrophy mice. Human Molecular Genetics. 2012;**21**:4448-4459. DOI: 10.1093/hmg/dds286
- [64] Avila AM, Burnett BG, Taye AA, et al. Trichostatin A increases SMN expression and survival in a mouse model of spinal muscular atrophy. The Journal of Clinical Investigation. 2007;117:659-671. DOI: 10.1172/jci29562

- [65] Beharry AW, Sandesara PB, Roberts BM, Ferreira LF, Senf SM, Judge AR. HDAC1 activates FoxO and is both sufficient and required for skeletal muscle atrophy. Journal of Cell Science. 2014;127:1441-1453. DOI: 10.1242/jcs.136390
- [66] Tang H, Goldman D. Activity-dependent gene regulation in skeletal muscle is mediated by a histone deacetylase (HDAC)-Dach2-myogenin signal transduction cascade. Proceedings of the National Academy of Sciences of the United States of America. 2006;103:16977-16982. DOI: 10.1073/pnas.0601565103
- [67] Bonetto A, Penna F, Minero VG, et al. Deacetylase inhibitors modulate the myostatin/follistatin axis without improving cachexia in tumor-bearing mice. Current Cancer Drug Targets. 2009;**9**:608-616. DOI: 10.2174/156800909789057015
- [68] Dupré-Aucouturier S, Castells J, Freyssenet D, Desplanches D. Trichostatin A, a histone deacetylase inhibitor, modulates unloaded-induced skeletal muscle atrophy. Journal of Applied Physiology (Bethesda, MD: 1985). 2015;119:342-351. DOI: 10.1152/japplphysiol.01031.2014
- [69] Ding J, Li F, Cong Y, et al. Trichostatin A inhibits skeletal muscle atrophy induced by cigarette smoke exposure in mice. Life Sciences. 2019;235:116800. DOI: 10.1016/j. lfs.2019.116800
- [70] Combaret L, Tilignac T, Claustre A, et al. Torbafylline (HWA 448) inhibits enhanced skeletal muscle ubiquitin-proteasome-dependent proteolysis in cancer and septic rats. The Biochemical Journal. 2002;**361**:185-192. DOI: 10.1042/0264-6021:3610185
- [71] Breuillé D, Farge MC, Rosé F, Arnal M, Attaix D, Obled C.

- Pentoxifylline decreases body weight loss and muscle protein wasting characteristics of sepsis. The American Journal of Physiology. 1993;265:E660-E666. DOI: 10.1152/ajpendo.1993.265.4.E660
- [72] Baviera AM, Zanon NM, Carvalho Navegantes LC, Migliorini RH, do Carmo Kettelhut I. Pentoxifylline inhibits Ca²⁺-dependent and ATP proteasome-dependent proteolysis in skeletal muscle from acutely diabetic rats. American Journal of Physiology. Endocrinology and Metabolism. 2007;292:E702-E708. DOI: 10.1152/ajpendo.00147.2006
- [73] Joshi R, Kadeer N, Sheriff S, Friend LA, James JH, Balasubramaniam A. Phosphodiesterase (PDE) inhibitor torbafylline (HWA 448) attenuates burn-induced rat skeletal muscle proteolysis through the PDE4/cAMP/EPAC/PI3K/Akt pathway. Molecular and Cellular Endocrinology. 2014;**393**:152-163. DOI: 10.1016/j. mce.2014.06.012
- [74] Arcaro CA, Assis RP, Zanon NM, et al. Involvement of cAMP/EPAC/Akt signaling in the antiproteolytic effects of pentoxifylline on skeletal muscles of diabetic rats. Journal of Applied Physiology (Bethesda, MD: 1985). 2018;124:704-716. DOI: 10.1152/japplphysiol.00499.2017
- [75] Deval C, Mordier S, Obled C, et al. Identification of cathepsin L as a differentially expressed message associated with skeletal muscle wasting. The Biochemical Journal. 2001;360:143-150. DOI: 10.1042/0264-6021:3600143
- [76] Steffen BT, Lees SJ, Booth FW. Anti-TNF treatment reduces rat skeletal muscle wasting in monocrotaline-induced cardiac cachexia. Journal of Applied Physiology (Bethesda, MD: 1985). 2008;105:1950-1958. DOI: 10.1152/japplphysiol.90884.2008

- [77] Lira EC, Gonçalves DA, Parreiras ESLT, Zanon NM, Kettelhut IC, Navegantes LC. Phosphodiesterase-4 inhibition reduces proteolysis and atrogenes expression in rat skeletal muscles. Muscle & Nerve. 2011;44:371-381. DOI: 10.1002/ mus.22066
- [78] Hinkle RT, Dolan E, Cody DB, Bauer MB, Isfort RJ. Phosphodiesterase 4 inhibition reduces skeletal muscle atrophy. Muscle & Nerve. 2005;32:775-781. DOI: 10.1002/mus.20416
- [79] Kackstein K, Teren A, Matsumoto Y, et al. Impact of angiotensin II on skeletal muscle metabolism and function in mice: Contribution of IGF-1, Sirtuin-1 and PGC-1α. Acta Histochemica. 2013;**115**:363-370. DOI: 10.1016/j. acthis.2012.09.009
- [80] Anker SD, Negassa A, Coats AJ, et al. Prognostic importance of weight loss in chronic heart failure and the effect of treatment with angiotensin-converting-enzyme inhibitors: An observational study. Lancet (London, England). 2003;**361**:1077-1083. DOI: 10.1016/s0140-6736(03)12892-9
- [81] Marzetti E, Calvani R, DuPree J, et al. Late-life enalapril administration induces nitric oxide-dependent and independent metabolic adaptations in the rat skeletal muscle. Age (Dordrecht, Netherlands). 2013;35:1061-1075. DOI: 10.1007/s11357-012-9428-4
- [82] Sumukadas D, Band M, Miller S, et al. Do ACE inhibitors improve the response to exercise training in functionally impaired older adults? A randomized controlled trial. The Journals of Gerontology Series A Biological Sciences and Medical Sciences. 2014;69:736-743. DOI: 10.1093/gerona/glt142
- [83] Sumukadas D, Witham MD, Struthers AD, McMurdo ME. Effect

- of perindopril on physical function in elderly people with functional impairment: A randomized controlled trial. CMAJ: Canadian Medical Association Journal (Journal de l'Association medicale canadienne). 2007;177:867-874. DOI: 10.1503/ cmaj.061339
- [84] Murphy KT, Chee A, Trieu J, Naim T, Lynch GS. Inhibition of the renin-angiotensin system improves physiological outcomes in mice with mild or severe cancer cachexia. International Journal of Cancer. 2013;133:1234-1246. DOI: 10.1002/ijc.28128
- [85] Fujita J, Mestre JR, Zeldis JB, Subbaramaiah K, Dannenberg AJ. Thalidomide and its analogues inhibit lipopolysaccharide-mediated induction of cyclooxygenase-2. Clinical Cancer Research: An Official Journal of the American Association for Cancer Research. 2001;7:3349-3355
- [86] Kedar I, Mermershtain W, Ivgi H. Thalidomide reduces serum C-reactive protein and interleukin-6 and induces response to IL-2 in a fraction of metastatic renal cell cancer patients who failed IL-2-based therapy. International Journal of Cancer. 2004;110:260-265. DOI: 10.1002/ijc.20089
- [87] Davis M, Lasheen W, Walsh D, Mahmoud F, Bicanovsky L, Lagman R. A phase II dose titration study of thalidomide for cancer-associated anorexia. Journal of Pain and Symptom Management. 2012;43:78-86. DOI: 10.1016/j.jpainsymman.2011.03.007
- [88] Liu KH, Liao LM, Ro LS, Wu YL, Yeh TS. Thalidomide attenuates tumor growth and preserves fast-twitch skeletal muscle fibers in cholangiocarcinoma rats. Surgery. 2008;143:375-383. DOI: 10.1016/j. surg.2007.09.035

- [89] Li TH, Lee PC, Lee KC, et al. Down-regulation of common NFκB-iNOS pathway by chronic thalidomide treatment improves hepatopulmonary syndrome and muscle wasting in rats with biliary cirrhosis. Scientific Reports. 2016;**6**:39405. DOI: 10.1038/srep39405
- [90] Kaplan G, Thomas S, Fierer DS, et al. Thalidomide for the treatment of AIDS-associated wasting. AIDS Research and Human Retroviruses. 2000;**16**:1345-1355. DOI: 10.1089/08892220050140892
- [91] Gordon JN, Trebble TM, Ellis RD, Duncan HD, Johns T, Goggin PM. Thalidomide in the treatment of cancer cachexia: A randomised placebo controlled trial. Gut. 2005;54:540-545. DOI: 10.1136/gut.2004.047563
- [92] Wilkes EA, Selby AL, Cole AT, Freeman JG, Rennie MJ, Khan ZH. Poor tolerability of thalidomide in end-stage oesophageal cancer. European Journal of Cancer Care. 2011;**20**:593-600. DOI: 10.1111/j.1365-2354.2011.01255.x
- [93] Tsujinaka T, Fujita J, Ebisui C, et al. Interleukin 6 receptor antibody inhibits muscle atrophy and modulates proteolytic systems in interleukin 6 transgenic mice. The Journal of Clinical Investigation. 1996;97:244-249. DOI: 10.1172/jci118398
- [94] Narsale AA, Carson JA. Role of interleukin-6 in cachexia: Therapeutic implications. Current Opinion in Supportive and Palliative Care. 2014;8:321-327. DOI: 10.1097/spc.0000000000000000091
- [95] Bayliss TJ, Smith JT, Schuster M, Dragnev KH, Rigas JR. A humanized anti-IL-6 antibody (ALD518) in nonsmall cell lung cancer. Expert Opinion on Biological Therapy. 2011;11:1663-1668. DOI: 10.1517/14712598.2011.627850
- [96] Jones SA, Scheller J, Rose-John S. Therapeutic strategies for the clinical

- blockade of IL-6/gp130 signaling. The Journal of Clinical Investigation. 2011;**121**:3375-3383. DOI: 10.1172/jci57158
- [97] Song SN, Yoshizaki K. Tocilizumab for treating rheumatoid arthritis: An evaluation of pharmacokinetics/pharmacodynamics and clinical efficacy. Expert Opinion on Drug Metabolism & Toxicology. 2015;11:307-316. DOI: 10.1517/17425255.2015.992779
- [98] Radigan KA, Nicholson TT, Welch LC, et al. Influenza A virus infection induces muscle wasting via IL-6 regulation of the E3 ubiquitin ligase atrogin-1. Journal of Immunology (Baltimore, Md.: 1950). 2019;**202**:484-493. DOI: 10.4049/jimmunol.1701433
- [99] Yano T, Osanami A, Shimizu M, et al. Utility and safety of tocilizumab in Takayasu arteritis with severe heart failure and muscle wasting. ESC Heart Failure. 2019;**6**:894-897. DOI: 10.1002/ehf2.12487
- [100] Bonetto A, Aydogdu T, Jin X, et al. JAK/STAT3 pathway inhibition blocks skeletal muscle wasting downstream of IL-6 and in experimental cancer cachexia. American Journal of Physiology. Endocrinology and Metabolism. 2012;303:E410-E421. DOI: 10.1152/ajpendo.00039.2012
- [101] Zhang L, Pan J, Dong Y, et al. Stat3 activation links a C/EBPδ to myostatin pathway to stimulate loss of muscle mass. Cell Metabolism. 2013;18:368-379. DOI: 10.1016/j.cmet.2013.07.012
- [102] Silva KA, Dong J, Dong Y, et al. Inhibition of Stat3 activation suppresses caspase-3 and the ubiquitin-proteasome system, leading to preservation of muscle mass in cancer cachexia. The Journal of Biological Chemistry. 2015;**290**:11177-11187. DOI: 10.1074/jbc.M115.641514
- [103] Llovera M, Carbó N, García-Martínez C, et al. Anti-TNF treatment

reverts increased muscle ubiquitin gene expression in tumour-bearing rats. Biochemical and Biophysical Research Communications. 1996;**221**:653-655. DOI: 10.1006/bbrc.1996.0651

[104] Granado M, Martín AI, Priego T, López-Calderón A, Villanúa MA. Tumour necrosis factor blockade did not prevent the increase of muscular muscle RING finger-1 and muscle atrophy F-box in arthritic rats. The Journal of Endocrinology. 2006;**191**:319-326. DOI: 10.1677/joe.1.06931

[105] Wiedenmann B, Malfertheiner P, Friess H, et al. A multicenter, phase II study of infliximab plus gemcitabine in pancreatic cancer cachexia. The Journal of Supportive Oncology. 2008;**6**:18-25

[106] Subramaniam K, Fallon K, Ruut T, et al. Infliximab reverses inflammatory muscle wasting (sarcopenia) in Crohn's disease. Alimentary Pharmacology & Therapeutics. 2015;**41**:419-428. DOI: 10.1111/apt.13058

[107] DeBoer MD, Lee AM, Herbert K, et al. Increases in IGF-1 after anti-TNF-α therapy are associated with bone and muscle accrual in pediatric Crohn disease. The Journal of Clinical Endocrinology and Metabolism. 2018;103:936-945. DOI: 10.1210/jc.2017-01916

[108] Chen CY, Tsai CY, Lee PC, Lee SD. Long-term etanercept therapy favors weight gain and ameliorates cachexia in rheumatoid arthritis patients: Roles of gut hormones and leptin. Current Pharmaceutical Design. 2013;19:1956-1964. DOI: 10.2174/1381612811319100014

[109] Wu C, Fernandez SA, Criswell T, et al. Disrupting cytokine signaling in pancreatic cancer: A phase I/II study of etanercept in combination with gemcitabine in patients with advanced disease. Pancreas. 2013;42:813-818. DOI: 10.1097/MPA.0b013e318279b87f

[110] Hong DS, Hui D, Bruera E, et al. MABp1, a first-in-class true human antibody targeting interleukin-1α in refractory cancers: An open-label, phase 1 dose-escalation and expansion study. The Lancet Oncology. 2014;**15**:656-666. DOI: 10.1016/s1470-2045(14)70155-x

[111] Hickish T, Andre T, Wyrwicz L, et al. MABp1 as a novel antibody treatment for advanced colorectal cancer: A randomised, double-blind, placebo-controlled, phase 3 study. The Lancet Oncology. 2017;18:192-201. DOI: 10.1016/s1470-2045(17)30006-2

[112] Hong DS, Janku F, Naing A, et al. Xilonix, a novel true human antibody targeting the inflammatory cytokine interleukin-1 alpha, in non-small cell lung cancer. Investigational New Drugs. 2015;33:621-631. DOI: 10.1007/s10637-015-0226-6

[113] Kurzrock R, Hickish T, Wyrwicz L, et al. Interleukin-1 receptor antagonist levels predict favorable outcome after bermekimab, a first-in-class true human interleukin- 1α antibody, in a phase III randomized study of advanced colorectal cancer. Oncoimmunology. 2019;8:1551651. DOI: 10.1080/2162402x.2018.1551651

[114] Yadava RS, Foff EP, Yu Q, et al. TWEAK/Fn14, a pathway and novel therapeutic target in myotonic dystrophy. Human Molecular Genetics. 2015;24:2035-2048. DOI: 10.1093/hmg/ddu617

[115] Bowerman M, Salsac C, Coque E, et al. Tweak regulates astrogliosis, microgliosis and skeletal muscle atrophy in a mouse model of amyotrophic lateral sclerosis. Human Molecular Genetics. 2015;24:3440-3456. DOI: 10.1093/hmg/ddv094

[116] Johnston AJ, Murphy KT, Jenkinson L, et al. Targeting of Fn14 prevents cancer-induced cachexia and prolongs survival. Cell. 2015;**162**:1365-1378. DOI: 10.1016/j. cell.2015.08.031

[117] Zhou X, Wang JL, Lu J, et al. Reversal of cancer cachexia and muscle wasting by ActRIIB antagonism leads to prolonged survival. Cell. 2010;**142**:531-543. DOI: 10.1016/j.cell.2010.07.011

[118] Roth SM, Walsh S. Myostatin: A therapeutic target for skeletal muscle wasting. Current Opinion in Clinical Nutrition and Metabolic Care. 2004;7:259-263. DOI: 10.1097/00075197-200405000-00004

[119] Benny Klimek ME, Aydogdu T, Link MJ, Pons M, Koniaris LG, Zimmers TA. Acute inhibition of myostatin-family proteins preserves skeletal muscle in mouse models of cancer cachexia. Biochemical and Biophysical Research Communications. 2010;**391**:1548-1554. DOI: 10.1016/j. bbrc.2009.12.123

[120] Becker C, Lord SR, Studenski SA, et al. Myostatin antibody (LY2495655) in older weak fallers: A proof-of-concept, randomised, phase 2 trial. The Lancet Diabetes & Endocrinology. 2015;3:948-957. DOI: 10.1016/s2213-8587(15)00298-3

[121] Golan T, Geva R, Richards D, et al. LY2495655, an antimyostatin antibody, in pancreatic cancer: A randomized, phase 2 trial. Journal of Cachexia, Sarcopenia and Muscle. 2018;**9**:871-879. DOI: 10.1002/jcsm.12331

[122] Attie KM, Borgstein NG, Yang Y, et al. A single ascending-dose study of muscle regulator ACE-031 in healthy volunteers. Muscle & Nerve. 2013;47:416-423. DOI: 10.1002/mus.23539

[123] Campbell C, McMillan HJ, Mah JK, et al. Myostatin inhibitor ACE-031 treatment of ambulatory boys with Duchenne muscular dystrophy: Results of a randomized, placebocontrolled clinical trial. Muscle & Nerve. 2017;55:458-464. DOI: 10.1002/ mus.25268

[124] Lach-Trifilieff E, Minetti GC, Sheppard K, et al. An antibody blocking activin type II receptors induces strong skeletal muscle hypertrophy and protects from atrophy. Molecular and Cellular Biology. 2014;34:606-618. DOI: 10.1128/mcb.01307-13

[126] Rooks D, Praestgaard J, Hariry S, et al. Treatment of sarcopenia with bimagrumab: Results from a phase II, randomized, controlled, proof-of-concept study. Journal of the American Geriatrics Society. 2017;65:1988-1995. DOI: 10.1111/jgs.14927

[127] Polkey MI, Praestgaard J, Berwick A, et al. Activin type II receptor blockade for treatment of muscle depletion in chronic obstructive pulmonary disease. A randomized trial. American Journal of Respiratory and Critical Care Medicine. 2019;**199**:313-320. DOI: 10.1164/rccm.201802-0286OC

[128] Mori-Yoshimura M, Yamashita S, Suzuki N, et al. Late phase II/III study of BYM338 in patients with sporadic inclusion body myositis (RESILIENT): Japanese cohort data. Rinsho Shinkeigaku (Clinical Neurology). 2019;59:806-813. DOI: 10.5692/clinicalneurol.cn-001325

[129] Pascual López A, Roqué i Figuls M, Urrútia Cuchi G, et al. Systematic review of megestrol acetate in the treatment of anorexia-cachexia syndrome. Journal of Pain and Symptom Management. 2004;27:360-369. DOI: 10.1016/j.jpainsymman.2003.09.007 [130] McCarthy HD, Crowder RE, Dryden S, Williams G. Megestrol acetate stimulates food and water intake in the rat: Effects on regional hypothalamic neuropeptide Y concentrations. European Journal of Pharmacology. 1994;**265**:99-102. DOI: 10.1016/0014-2999(94)90229-1

[131] Mantovani G, Macciò A, Massa E, Madeddu C. Managing cancer-related anorexia/cachexia. Drugs. 2001;**61**:499-514. DOI: 10.2165/00003495-200161040-00004

[132] Ronga I, Gallucci F, Riccardi F, Uomo G. Anorexia-cachexia syndrome in pancreatic cancer: Recent advances and new pharmacological approach. Advances in Medical Sciences. 2014;59:1-6. DOI: 10.1016/j. advms.2013.11.001

[133] Wang J, Wang Y, Tong M, Pan H, Li D. New prospect for cancer cachexia: Medical cannabinoid. Journal of Cancer. 2019;**10**:716-720. DOI: 10.7150/jca.28246

[134] Brisbois TD, de Kock IH, Watanabe SM, et al. Delta-9-tetrahydrocannabinol may palliate altered chemosensory perception in cancer patients: Results of a randomized, double-blind, placebocontrolled pilot trial. Annals of Oncology: Official Journal of the European Society for Medical Oncology. 2011;22:2086-2093. DOI: 10.1093/annonc/mdq727

[135] Turcott JG, Del Rocío Guillen Núñez M, Flores-Estrada D, et al. The effect of nabilone on appetite, nutritional status, and quality of life in lung cancer patients: A randomized, double-blind clinical trial. Supportive Care in Cancer: Official Journal of the Multinational Association of Supportive Care in Cancer. 2018;26:3029-3038. DOI: 10.1007/s00520-018-4154-9

[136] Côté M, Trudel M, Wang C, Fortin A. Improving quality of life with nabilone during radiotherapy treatments for head and neck cancers: A randomized doubleblind placebo-controlled trial. Annals of Otology, Rhinology, and Laryngology. 2016;125:317-324. DOI: 10.1177/0003489415612801

[137] Alamdari N, Aversa Z, Castillero E, et al. Resveratrol prevents dexamethasone-induced expression of the muscle atrophy-related ubiquitin ligases atrogin-1 and MuRF1 in cultured myotubes through a SIRT1dependent mechanism. Biochemical and Biophysical Research Communications. 2012;417:528-533. DOI: 10.1016/j. bbrc.2011.11.154

[138] Wang DT, Yin Y, Yang YJ, et al. Resveratrol prevents TNF-α-induced muscle atrophy via regulation of Akt/mTOR/FoxO1 signaling in C2C12 myotubes. International Immunopharmacology. 2014;**19**:206-213. DOI: 10.1016/j.intimp.2014.02.002

[139] Momken I, Stevens L, Bergouignan A, et al. Resveratrol prevents the wasting disorders of mechanical unloading by acting as a physical exercise mimetic in the rat. FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology. 2011;25:3646-3660. DOI: 10.1096/fj.10-177295

[140] Chen X, Wu Y, Yang T, et al. Salidroside alleviates cachexia symptoms in mouse models of cancer cachexia via activating mTOR signalling. Journal of Cachexia, Sarcopenia and Muscle. 2016;7:225-232. DOI: 10.1002/jcsm.12054

[141] Chen L, Chen L, Wan L, et al. Matrine improves skeletal muscle atrophy by inhibiting E3 ubiquitin ligases and activating the Akt/mTOR/ FoxO3 α signaling pathway in C2C12

myotubes and mice. Oncology Reports. 2019;**42**:479-494. DOI: 10.3892/ or.2019.7205

[142] Chen L, Xu W, Yang Q, et al. Imperatorin alleviates cancer cachexia and prevents muscle wasting via directly inhibiting STAT3. Pharmacological Research. 2020;158:104871. DOI: 10.1016/j.phrs.2020.104871

[143] Yang Q, Wan L, Zhou Z, et al. Parthenolide from *Parthenium integrifolium* reduces tumor burden and alleviate cachexia symptoms in the murine CT-26 model of colorectal carcinoma. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology. 2013;20:992-998. DOI: 10.1016/j. phymed.2013.04.020

[144] Yu R, Chen JA, Xu J, et al. Suppression of muscle wasting by the plant-derived compound ursolic acid in a model of chronic kidney disease. Journal of Cachexia, Sarcopenia and Muscle. 2017;8:327-341. DOI: 10.1002/jcsm.12162

[145] Chen L, Yang Q, Zhang H, et al. Cryptotanshinone prevents muscle wasting in CT26-induced cancer cachexia through inhibiting STAT3 signaling pathway. Journal of Ethnopharmacology. 2020;**260**:113066. DOI: 10.1016/j.jep.2020.113066