We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



186,000

200M



Our authors are among the

TOP 1% most cited scientists





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



Chapter

Mixture Transition Distribution Modelling of Multivariate Time Series of Discrete State Processes: With an Application to Modelling Flowering Synchronisation with Respect to Climate Dynamics

Irene Hudson, Susan Won Sun Kim and Marie Keatley

Abstract

A new approach to assess synchronicity developed in this chapter is a novel bivariate extension of the generalised mixture transition distribution (MTDg) model (we coin this B-MTD). The aim of this chapter is to test MTDg an extended MTD with interactions model and its bivariate extension of MTD (B-MTD) to investigate synchrony of flowering of four Eucalypts species—E. leucoxylon, E. microcarpa, E. polyanthemos and E. tricarpa over a 31 year period. The mixture transition distribution (MTDg) is a method to estimate transition probabilities of high order Markov chains. Our B-MTD approach allows us the derive rules of thumb for synchrony and asynchrony between pairs of species, e.g. flowering of the four species. The latter B-MTD rules are based on transition probabilities between all possible on and off flowering states from previous to current time. We also apply MTDg modelling using lagged flowering states and climate covariates as predictors to model current flowering status (on/off) to assess synchronisation using residuals from the resultant models via our adaptation of Moran's classic synchrony statistic. We compare these MTDg (with covariates)-based synchrony measures with our B-MTD results in addition to those from extended Kalman filter (EKF)-based residuals.

Keywords: multivariate mixed transition distributions, Markov chains, synchrony, climate, eucalypt flowering

1. Introduction

Separation or lack of overlap of flowering time in eucalypts has been suggested as a mechanism for maintaining overall 'generic identity' of a plant species. If, for example flowering times and pollinators overlap in sympatric species, hybridization can occur between closely related eucalypts species. Therefore examination of longterm synchrony establishes a baseline of flowering behaviour which may assist in detecting recent or future changes. Although *Eucalyptus* as a genus dominates much of the Australian landscape [1, 2], few studies have quantified eucalypt flowering overlap within or between species, due to the shortage of phenological data in Australia [3, 4, 7]. This chapter examines flowering synchrony over a 31 year period, 1940–1970, at the population level among four eucalypts species—*Eucalyptus leucoxylon, E. microcarpa, E. polyanthemos* and *E. tricarpa* [3–8].

A new approach to assess synchronicity developed in this chapter is a novel bivariate extension of the of the MTDg model [6, 9] (we coin this B-MTD). The aim of this chapter is to test mixture transition distribution (MTD) and an extended MTDg with interactions; and a novel bivariate extension of MTD (B-MTD) to investigate synchrony in phenological data. The MTDg model [6, 9] was the first approach developed to study the multivariate relationship between the probability of flowering with two states of rain and mean temperature via a mixture transition distribution (MTD), assuming, however a different transition matrix from each lag to the present time (our MTDg analysis), thus generalising the MTD approach in [13], (see also [10]) which led to the development of the MARCH software to perform MTD without covariates [11, 12]. The MTDg model is different to MARCH not only in terms of incorporating interactions between the covariates but also in its minimization process, namely using the AD Model BuilderTM [14], which uses auto-differentiation as a minimisation tool. This is shown to be computationally less intensive than MARCH. The assumption Berchtold's MTD model, namely the assumed equality of the transition matrices among different lags, was a strong assumption, so the idea of the mixture transition distribution model was to consider independently the effect of each lag to the present instead of considering the effect of the combination of lags as in pure Markov chain processes. Specifically, an extended model for MTDg analysis which accommodates interactions was developed in [6], and applied to MTDg modelling of the flowering of four eucalyptus species studied in this chapter, as multivariate time series.

This work extends both MARCH and the work in [15, 16] to allow for differing transition matrices among the lags, i.e. our B-MTD method builds on this approach of the MTDg with interaction model [6, 9]. The MTDg model with interactions showed that the flowering of *E. leucoxylon* and *E. tricarpa* behave similarly with temperature (both flower at low temperature) and both have a positive relationship with flowering intensity 11 months ago. The flowering of *E. microcarpa* behaves differently in that *E. microcarpa* flowers at high temperature.

Our B-MTD approach developed in this chapter allows us the derive rules of thumb for synchrony and asynchrony between pairs of species. The latter B-MTD rules are based on transition probabilities between all possible on and off flowering states from previous to current time. Synchronisation is also tested using residuals from the resultant models via an adaptation of Moran's [17, 18] classical synchrony statistic, incorporating MTDg residuals [17–19].

We also apply the earlier MTDg modelling in [6] using climate covariates and lagged flowering states as predictors to model flowering states (on/off) and thus assess synchronisation using an adaptation of the approach of Moran to the resultant MTDg model and fitted residuals. We compare these MTDg (with covariates)-based synchrony measures with our B-MTD results in addition to those using the extended Kalman filter (EKF) [15, 19], based residuals obtained earlier in [21].

2. The mixture transition distribution (MTDg) and B-MTD models: mathematical formulations in brief

2.1 The MTDg model with interactions between the covariates

The MTD model with covariates was discussed in [6] and developed in [15, 19] to incorporate interactions between the covariates (e.g. rainfall, temperature

variants in the case study discussed). The high-order MTD transition probabilities are computed as follows:

$$P(X_{t} = i_{0} | X_{t-1} = i_{1}, ...X_{t-f} = i_{f}, C_{1} = c_{1}, ...C_{e} = c_{e}, M_{1} = m_{1}, ...M_{l} = m_{l})$$

$$= \sum_{g=1}^{f} \lambda_{gqi_{g}i_{0}} + \sum_{h=1}^{e} \lambda_{f+h} d_{hj_{h}i_{0}} + \sum_{u=1}^{l} \lambda_{f+e+u} s_{uv_{u}i_{0}}$$
(1)

where λ_{f+e+u} is the weight for the interaction term, $q_{i_g i_0}$ is the transition probability from modality i_g observed at time t-g and modality i_0 observed at time t in the transition matrix Q, $s_{uv_u i_0}$ is transition probability between covariate h_1 and covariate h_2 interaction term ($v_u = d_{h_1 j_{h_1}} \times d_{h_2 j_{h_2}}$) and X_t , and where $\sum_{g=1}^{f+e+l} \lambda_g = 1$ and where $\lambda_g \ge 0$.

We refer the reader to [6], and further works in the seminal book by Hudson and Keatley [7] for further mathematical details.

2.2 The bivariate mixture transition distribution (B-MTD)

Let $\{X_t\}$ and $\{Y_t\}$ be sequences of random variables (say two (flowering intensity) time series) taking values in the finite set $N = \{1, ..., k\}$. In a f^{th} -order Markov chain, the probability that $\{X_t, Y_t\} = \{i_0, i'_0\}$, $(i_0, i'_0 \in N)$ depends on the combination of values taken by X_{t-f} , ..., X_{t-1} , Y_{t-f} , ..., Y_{t-1} . In the MTD model, the contributions of the different lags are combined additively. Then a bivariate MTD model, which we denote by B-MTD, has the following formulation:

$$P\Big(\{X_t, Y_t\} = \{i_0, i'_0\} | X_{t-1} = i_1, ..., X_{t-f} = i_f, Y_{t-1} = i'_1, ..., Y_{t-f} = i'_f\Big)$$

$$= \sum_{g=1}^f \lambda_g q_{i_g, i'_g, i_0, i'_0}$$
(2)

where $i_f, ..., i_0, i'_f, ..., i'_0 \in N$, the probabilities q_{i_g, i'_g, i_0, i'_0} are elements of an $m^2 \times m^2$ transition matrix $Q = [q_{i_g, i'_g, i_0, i'_0}]$, each row of which is a probability distribution (i.e. each row sums to 1 and the elements are nonnegative) and $\lambda = (\lambda_f, ..., \lambda_1)'$ is a vector of lag parameters. To ensure that the results of the model are probabilities, that is, $0 \leq \sum_{g=1}^{f} \lambda_g q_{i_g i'_g i_0 i'_0} \leq 1$ the vector λ is subject to the constraints $\sum_{g=1}^{f} \lambda_g = 1$ and $\lambda_g \geq 0$.

Covariates and interaction terms can be added to the bivariate MTD (B-MTD) as follows:

$$P(\{X_{1,t},...,X_{n,t}\} = \{i_{1,0},...,i_{n,0}\} | X_{t-1} = i_{1},...,X_{t-f} = i_{f}, Y_{t-1} = i'_{1},...,Y_{t-f} = i'_{f}, C_{1} = c_{1},...,C_{e} = c_{e}, M_{1} = m_{1},...,M_{l} = m_{l})$$

$$= \sum_{g=1}^{f} \lambda_{g} q_{i_{1,g},...,i_{n,g},i_{1,0},...,i_{n,0}} + \sum_{h=1}^{e} \lambda_{f+h} d_{hj_{h},i_{1,0},...,i_{n,0}} + \sum_{u=1}^{l} \lambda_{f+e+u} s_{uv_{u}i_{1,0},...,i_{n,0}}$$
(3)

where λ_{f+e+u} is the weight for the interaction term, $s_{uv_u i_{1,0}, \dots, i_{n,0}}$ is the transition probability between covariate h_1 and covariate h_2 interaction term $(v_u = d_{h_1 j_{h_1}} \times d_{h_2 j_{h_2}})$ and (X_t, Y_t) , and where $\sum_{g=1}^{f+e+l} \lambda_g = 1$. For example, if both X_t and Y_t are time series that constitute random realizations of two states {0, 1} and the covariates C_1, \dots, C_e are also defined by bivariate states {0, 1}, then the set of all possible states for (X_t, Y_t) is {(0, 0), (0, 1), (1, 0), (1, 1)}. Hence the transition matrix $Q = \left[q_{i_g i'_g i_0 i'_0}\right]$ is a 4 × 4 matrix as specified below.

Previous state (X_{t-1}, Y_{t-1})				
	0, 0 (1)	0,1(2)	1, 0 (3)	1, 1 (4)
0, 0 (1)	(1, 1)	(1, 2)	(1, 3)	(1, 4)
0, 1 (2)	(2, 1)	(2, 2)	(2, 3)	(2, 4)
1, 0 (3)	(3, 1)	(3, 2)	(3, 3)	(3, 4)
1, 1 (4)	(4, 1)	(4, 2)	(4, 3)	(4, 4)

The transition matrices $D_h = \left[d_{hj_h i_0 i'_0} \right]$, h = 1, ..., e, are 2 × 4 matrices as below.

	Covaria	ite state
(X_t, Y_t)	0 (1)	1 (2)
0, 0 (1)	(1, 1)	(1, 2)
0, 1 (2)	(2, 1)	(2, 2)
1, 0 (3)	(3, 1)	(3, 2)
1, 1 (4)	(4, 1)	(4, 2)

2.3 Synchrony analysis using Moran's approach

Moran in [17, 18] suggested that if two series x_t and y_t are synchronous, and if x_t can be estimated by a model f(x), the residuals from series x_t fitted to f(x), and the residuals from series y_t , fitted with the same model, but with observations, y_t , then, f(y) will be positively correlated. The synchrony of two series can then be examined by testing the significance of the correlation of these two series of residuals (using the same model). Moran used an autoregressive integrated moving average (ARIMA) model to test synchrony. Moran's theorem suggests that if two (or more) populations sharing a common linear density-dependence (in a so-called renewal process) are disturbed with correlated noise, they will become synchronised with a correlation matching the noise correlation (see details in [4], and also [6, 15, 21]).

In this chapter we adopt the *k*th order linear stochastic difference to assess synchrony. Goodness of fit of the second order AR (k = 2) model is obtained. The series of residuals can then be found by subtracting the predicted (fitted species) value from the observed series. In summary, synchrony (or otherwise) of two series can be established by performing a test of significance on the correlation coefficient calculated from the two series of residuals as follows:

- Calculate the residuals for say, *E. leucoxylon* (Leu) using its AR (2) model. We denote this residual series by R1.
- Calculate residuals for say, *E. tricarpa* (Tri) using this same model. We denote this residual series by R2.
- Calculate the Pearson correlation coefficient between the residual series R1 and R2 and test for its significance at p < 0.05.

Further details on how Moran's method is used and adapted in the case of the MTDg-based models are given in [15] (see also Section 4.8). We use the functionals and parameterisations from the mixture transition distribution (MTD) analysis as the basis of our EKF modelling approach. EKF is likewise a method to estimate the past, present and future status of non-linear time series data by minimising the mean square error. We will also test whether EKF better detects asynchronous species pairs, given EKF estimates the Kalman gain and covariance matrix at each time point [15, 19].

3. Data

Flowering data were sourced from the Box-Ironbark Forest near Maryborough, Victoria, in particular the flowering records of *E. leucoxylon*, *E. microcarpa*, *E. polyanthemos* and *E. tricarpa* (1940 and 1971). Flowering intensity was calculated by using a rank score (from 0 to 5) based on the quantity and distribution of flowering [4, 20, 23].

Flowering intensity scores were dichotomised into two discrete states, namely on and off (1/0) flowering (**Figure 1**) as in [6]. One temperature variant, mean monthly diurnal temperature (MeanT), in addition to the monthly rainfall (Rain) were included as climate covariates in the MTDg models; along with the temperature by rain interaction effect. We used discrete state low/high (lower than median temperature *vs* higher than median temperature) for the temperature variable dichotomies and less/more (less than the median rainfall *vs* more than the median



Figure 1. *Flowering of the four eucalypts species.*

Climate variables	Low (less)	High (more)
Mean diurnal temp (°C)	≤13.84	>13.84
Rain (mm)	≤40.45	>40.45

Table 1.

Cut-points for climate variables based on medians.

rainfall) for the rainfall variable. The cut-points for the states or low/high categories of each climate covariate are shown in **Table 1**.

4. Results

4.1 Bivariate MTD (B-MTD) discrete states results

Four eucalypts species, *E. leucoxylon*, *E. microcarpa*, *E. polyanthemos* and *E. tricarpa* were modelled using the order 1 B-MTD model discussed in Section 2.2—without the inclusion of covariates (such as temperature (variants) and rainfall). These species were paired as follows: *E. leucoxylon* and *E. microcarpa* (LeuMic); *E. leucoxylon* and *E. polyanthemos* (LeuPol); *E. leucoxylon* and *E. tricarpa* (LeuTri) and so on; hence 6 pairs were modelled via B-MTD (see **Table 2**) for the corresponding bivariate transition probabilities (see also **Figure 2**).

The possible states for any pair of species is the set {(0, 0), (0, 1), (1, 0), (1, 1)}, where no flowering is represented as 0 (state = 0 = no flowering) and flowering is represented by a 1 (state = 1 = flowering). Since lag order 1 B-MTD models were used, the mixing probability λ is equal to 1.0.

The corresponding transition matrices for the 6 B-MTD models are given in **Table 2**. These transition profiles are also shown schematically as flow diagrams in **Figures 3**–4, and also as transition signatures in **Figures 5**–6. These shall be discussed in more detail later. The transitions to differing states (from **Table 2**) are shown as arrows (transitions A to F) in the schematic diagram of **Figure 2**. The exact probabilities of such transitions are given by the off diagonal elements of **Table 2** and also shown above or below the arrows in **Figures 3** and **4**.

The transitions have the following intuitive interpretation and associated probability (sum), which are derived from the subcomponents of the transition matrices Q (see **Table 2**).

- A: transition of both species off to one species on: q(0, 0; 0, 1) + q(0, 0, 1, 0)
- B: transition of both species on to one species off: q(1, 1; 0, 1) + q(1, 1, 1, 0)
- C: species switching states: q(0, 1;1, 0) + q(1, 0; 0, 1)
- D: transition of one species off to both species off: q(0, 1; 0, 0) + q(1, 0; 0, 0)
- E: transition of one species on to both species on: q(0, 1;1, 1) + q(1, 0;1, 1)
- F: transition of one species on/off to both species off/on: q(0, 0;1, 1) + q(1, 1;0, 0)

In this chapter we shall demonstrate that transitions that lead towards both species being off or both species being on (states D, E or F), are considered to be

Species	Previous state	Current state				
		(0, 0)	(0, 1)	(1, 0)	(1, 1)	
LeuMic	(0, 0)	0.6667	0.2280	0.1053	0.0000	
	(0, 1)	0.0000	0.6000	0.1333	0.2667	
	(1, 0)	0.0845	0.0376	0.8357	0.0423	
	(1, 1)	0.0000	0.0612	0.4490	0.4898	
LeuPol	(0, 0)	0.6970	0.0303	0.2626	0.0101	
	(0, 1)	0.4444	0.3889	0.0000	0.1667	
	(1, 0)	0.0562	0.0000	0.7921	0.1517	
	(1, 1)	0.1309	0.0952	0.1429	0.6310	
LeuTri	(0, 0)	0.6947	0.1263	0.1053	0.0737	
	(0, 1)	0.0455	0.3636	0.0000	0.5909	
	(1, 0)	0.2203	0.0085	0.7034	0.0678	
	(1, 1)	0.0069	0.0069	0.1736	0.8125	
MicPol	(0, 0)	0.7637	0.1429	0.0879	0.0055	
	(0, 1)	0.1818	0.6705	0.1023	0.0455	
	(1, 0)	0.2737	0.0000	0.6842	0.0421	
	(1, 1)	0.0714	0.2141	0.3572	0.3573	
MicTri	(0, 0)	0.7975	0.0316	0.1329	0.0380	
	(0, 1)	0.2232	0.7500	0.0179	0.0089	
	(1, 0)	0.1090	0.0182	0.5819	0.2909	
	(1, 1)	0.0000	0.4259	0.0000	0.5741	
PolTri	(0, 0)	0.7464	0.1739	0.0797	0.0000	
	(0, 1)	0.0719	0.7842	0.0360	0.1079	
	(1, 0)	0.3067	0.0400	0.6400	0.0133	
	(1, 1)	0.0370	0.1482	0.4074	0.4074	

Mixture Transition Distribution Modelling of Multivariate Time Series of Discrete State... DOI: http://dx.doi.org/10.5772/intechopen.88554

Table 2.



Figure 2.

Subcomponents of possible transitions.

synchronising. However, transitions that lead towards only one species being on or off (flowering) (A and B) and where within a species pair flowering switches (transitions C) are considered to be asynchronous.

Note that the probabilities of staying in the same state; e.g. both species continuing to be in a non-flowering state (a (0, 0) to (0, 0) transition); one species flowering off and the other species in the pair with flowering on, (a (0, 1) to (0, 1)



Figure 3. Diagram of transition probabilities for synchronous pairs: LeuTri and LeuPol.



Figure 4. Diagram of transition probabilities for asynchronous pairs: PolTri, LeuMic and MicPol.

transition); one species on the other in the pair off (a (1, 0) to (1, 0) transition); and both species continuing to flower (a (1, 1) to (1, 1) transition) are not shown on **Figure 2**. These to same states transitions, are given for each species, by the diagonal elements in the transition matrices (from previous to current states) in **Table 2**; and are also shown in **Figures 3** and **4** as numbers (positioned next to the 4 states as boxes).

An examination of the transition probabilities for the species pairs in **Table 2** shows that there is a significantly high propensity (probability) to remain in the

Mixture Transition Distribution Modelling of Multivariate Time Series of Discrete State... DOI: http://dx.doi.org/10.5772/intechopen.88554



Figure 5.



same (bivariate) state as the previous state (see highlighted transition probabilities on the diagonals). For synchronous species pairs, such as LeuPol, and LeuTri the likelihood of species switching flowering state (states C), i.e. transition from one species flowering in a pair previous state = (0, 1) to the other species flowering, current state = (1, 0) never occurs (transition probability = 0.0000); or the likelihood of the transition from one species flowering to the other species flowering (i.e. a (1, 0) to (0, 1) transition) is rare (0.0000 \leq transition probability \leq 0.0085). For asynchronous species pairs such as LeuMic, MicPol, and PolTri, their switching probabilities are significantly higher in that at least one of the transition probabilities from (0, 1) to (1, 0); or from (1, 0) to (0, 1) is greater than 0.036, with associated probability \geq 0.076.

Overall for synchronous pairs the probabilities of one species flowering to both or no species flowering, i.e. one off to both off, or one on to both on are high (>0.30). The latter are delineated by D and E transitions in **Figure 2** and **Table 4**. Overall for asynchronous pairs there are high probabilities of both off (or on) to one off (or on). The latter transitions are delineated by A and B in **Figure 2**, with probabilities given in **Tables 3** and **4**.

In summary the transitions that lead to both species being off (no flowering) or both species being on (flowering) (transitions D, E or F), are considered to be



Figure 6.

Transition probabilities from (0, 1) and (1, 0) states for 6 species pairs.

Transition names	Description	Probability: sum of subcomponents	Threshold for synchrony	Threshold for asynchrony	Rules	
А	Both off to one off	q(0, 0;0, 1) + q(0, 0;1, 0)	<0.30	≥0.30▲	P(A or B) >	
B	Both on to one on	q(1, 1;0, 1) + q(1, 1;1, 0)	<0.50	≥0.50	0.8 for asynchrony	
C^{\varphi}	Switching	q(0, 1;1, 0) + q(1,0;0, 1)	<0.05▼	≥0.05	7	
D¢	One off to both off	q(0, 1;0, 0) + q(1, 0;0, 0)	≥0.40▲	<0.40	P(D or E) >	
Е	One on to both on	q(0, 1;1, 1) + q(1, 0;1, 1)	≥0.40▲	<0.40	0.65 for synchrony	
F	Both on (off) to both off (on)	q(0, 0;1, 1) + q(1, 1;0, 0)	≥0.08▲	<0.08		

Table 3.

Descriptions and rules of (a) synchrony based on the transitions A-F.

synchronizing. However, transitions that lead to only one species being on or off (no flowering) (transitions A and B) and where a species pairs' flowering status switches (transitions C) are considered to be asynchronous.

We now provide a rule for synchrony (or asynchrony) based on subcomponent (sums) of the transition probabilities derived from the B-MTD model:

- Two species are synchronous if P(D or E) > 0.65, i.e. P(one on to both on) + P(one off to both off) > 0.65,
- Two species are asynchronous if P(A or B) > 0.80, i.e. P(both off to one off) + P(both on to one on) > 0.8.

The transitions have the following interpretation and probabilities (**Tables 3** and **4**):

- A: transition of both species off (in the past state) to one species flowering (on) in the current state;
- B: transition of both species on to one species off;
- C: species switching states;
- D: transition of one species off to both species off;
- E: transition of one species on to both species on;
- F: transition of one species on/off to both species off/on.

According to the rules given in **Table 3**, the synchronous pairs are LeuTri and LeuPol (with P(D or E) > 0.65); asynchronous pairs are: PolTri, LeuMic and MicPol (with P(A or B) > 0.80) and a species pair that is neither synchronous nor asynchronous is MicTri.

In summary we have a simple rule for (a) synchrony, which in agreement with the work of [6] (see also [25]), using the synchronisation theory of Moran that:

- *E. leucoxylon* flowering is synchronous with both *E. polyanthemos* and *E. tricarpa*, but asynchronous with *E. microcarpa*.
- *E. microcarpa* is synchronous with none of three species; specifically it is asynchronous with both *E. leucoxylon* and *E. polyanthemos* (and has no relationship with *E. tricarpa*).

Transition probability sums	P(A)	P(B)	P(A or B)	P(C)	P(D)	P(E)	P(D or E) P(F)
Synchronous (S) pairs							
LeuTri	0.232	0.181		0.008	0.266	0.659	0.081
LeuPol	0.293	0.238		0.000	0.501	0.318	0.141
Asynchronous (A) pairs							
PolTri	0.254	0.556		0.076	0.379	0.121	0.037
LeuMic	0.333	0.510		0.171	0.085	0.309	0.000
MicPol	0.231	0.571		0.102	0.455	0.088	0.077
Neither S nor A							
MicTri	0.165	0.426		0.036	0.332	0.300	0.038

Table 4.

Transition probabilities of events A to F for each species pair categorised into synchronous and asynchronous (or neither) species pairs.

- *E. polyanthemos* flowering is synchronous only with *E. leucoxylon*; and asynchronous with both *E. microcarpa* and *E. tricarpa*.
- *E. tricarpa* flowering is synchronous only with that of *E. leucoxylon*; and is asynchronous with *E. polyanthemos* (and has no relationship with *E. microcarpa*).

We can view **Figure 5** as the transition signatures from past states, where both species flowering is off or both species flowering is on, for synchronous pairings (LeuTri or LeuPol) and the asynchronous species pairs (PolTri, LeuMic and MicPol). **Figure 6** likewise delineates transition signatures from past states, where only one species of the pair is flowering. These signatures (**Figures 5** and **6**) distinctly differ according to whether a species pair is synchronous or asynchronous.

For MicTri the associated sum of the probabilities for transitions A and B (both off/on to one off/on) is 0.591 (see **Table 4**), which is close to the threshold for synchrony of 0.65. Note that the more sophisticated MTDg modelling approach in Section 4.2 which incorporates covariates (mean temperature and rainfall) with interactions, shows that indeed *E. microcarpa* and *E. tricarpa* are synchronous (**Tables 6** and 7), wherein the MTDg model allows for prior lag 1 to lag 12 month flowering effects and climate covariates (see also **Table 7** and **Figure 7**).

4.2 Moran tests on residuals of the MTDg models incorporating climatic covariates

In this section synchronisation among species pairs is tested using Moran's correlation method on the cross-residuals, based on MTDg models which incorporate both climate covariates and lagged effects of previous flowering. This work is based on [16], where MTDg models allowing interactions were fitted to the same four species. We present here only MTDg models with two covariates, namely, mean temperature and rainfall.

Parameters of the MTDg models are shown in **Table 5**. Significant lag effects of previous flowering states (*lag j*, where j = 1, ..., 12 months), and of the climatic covariates (*meanT* and *rain*) and their interaction (*meanT*rain*) are also given in **Table 5**. The estimated parameters for the MTDg models generally show a (positive) 1 month lag effect and 9, 11 and 12 months lag effects of previous flowering status (**Table 5**).

From **Tables 5** and **6** we observe that mean diurnal temperature (*meanT*) has a significant effect on flowering for all species; *rain* impacts significantly only on *E. tricarpa* (Tri) and an interaction effect between *rain* and *meanT* exists for *E. polyanthemos* (Pol). Overall, flowering increases as temperature (*MeanT*)

	Species	lag 1	lag 9	lag 10	lag 11	lag 12	Temp variable	Rain	$\mathbf{Temp}\times\mathbf{rain}$	
	E. mic	0.534	-	-	0.032^{φ}	0.275	0.136	-	-	
	E. poly	0.530	0.060	-	0.160	0.105	0.091	0.009	0.045	
	E. leu	0.611	-	-	0.124	0.042	0.202	-	-	
	E. tri	0.617	0.059	0.009	0.096	-	0.157	0.062	-	
,										

^{*b*}Covariate effects above 0.03 are considered significant.

Table 5.

MTDg mixing probabilities of MeanT and rain models.

⁻ indicates cells with zero probabilities.

Species	Climate effects	Prev flow	rious ering	Temp	erature	R	ain	Temperature by rain interac		n interaction	
	(temp/ rain)	Off	On	Low ¹	High ²	Less ³	More ⁴	Low/ less	Low/ more	High/ less	High/ more
E. mic	(+/-)	0.00	1.00	0.00	1.00	0.39	0.28	-	-	-	-
E. poly	Inter- action	0.01	1.00	0.00	0.34	0.94	0.03	0.88	0.12	0.20	0.96
E. leu	(_/+)	0.05	1.00	1.00	0.00	0.88	0.94	-	-	-	-
E. tri	(-/+)	0.00	1.00	1.00	0.00	0.00	1.00)}[-	-	
¹ Cut point for	r low temper	rature sta	ates: M	eanT 1	3.83°C.				()	(
² Cut point for	r high tempe	rature st	tates: N	1eanT >	13.84°C.						
³ Cut point for	r less rain: ra	ain 40.4	14 mm								
⁴ Cut point for Note that '-' i	r more rain: ndicates cell	rain >4 s with ze	0.45 n ero pro	ım. babilitie	<i>'s</i> .						

Table 6.

Transition probabilities of flowering for the meanT and rain MTDg models.

Model species	mic	pol	leu	tri			
Synchronous fitted species	tri (0.14)	leu (0.14)	pol (0.16)	mic (0.15)			
			tri (0.11)				
Asynchronous fitted species			mic (-0.14^{ϕ})				
^{<i>φ</i>} A negative and significant correlation indicates an asynchronous species pair.							

Table 7.

Significant Moran correlations (in brackets) from the MTDg models.



Figure 7.

Synchrony relationships among the four eucalypts species.

increases for *E. microcarpa*; and flowering decreases as temperature increases for both *E. leucoxylon* and *E. tricarpa*. Rainfall positively impacts the flowering of *E. tricarpa* (i.e. flowering increases with more rainfall). Interestingly *E. polyanthemos* exhibits increased flowering at low *meanT* when there is contemporaneous below average rainfall and at high *meanT* with above average rainfall (see the transition probabilities to flowering for the interaction effect of *E. polyanthemos* (i.e. (0.88, 0.12, 0.20, 0.96)) in **Table 6**.

In what follows we denote the species used to estimate the parameters for the MTD-based equation as the 'Model species' and the species fitted with these

estimated parameters as the 'Fitted species'. **Table** 7 gives the resultant significant Moran correlations based on the residual series from the MTDg-based model and fitted species equations. Significant Moran correlations from both the MTDg (and the EKF models show that (a)synchronous pairings found via the MTD and EKF models in [15–19] generally agree (**Tables 7** and **8**); refer also to **Figure 7**, where a solid line indicates synchronous pairs and a dashed line indicates asynchronous pairs of species.

Table 7 shows significant positive MTDg-based correlations (P < 0.006) for the following (model species: fitted species) pairs—(LeuPol), (PolLeu), (LeuTri), (MicTri) and (TriMic), indicating that *E. leucoxylon* is synchronous with *E. polyanthemos*, in agreement with the rules of synchrony described earlier (**Tables 3** and 4). *E. leucoxylon* is synchronous with *E. tricarpa*; and that *E. microcarpa* and *E. tricarpa* are synchronous. The synchrony of the latter species pair (MicTri) however, contrasts the results of Moran-based results on raw intensity profiles which indicate that *E. microcarpa* and *E. tricarpa* (i.e. MicTri or TriMic), the associated sum of the probabilities for transitions D and E (one species off/on to both species off/on) is 0.591 (**Table 4**), which is close to the threshold for synchrony of 0.65 (**Tables 3** and 4).

Tables 7 and **8** shows significant negative-based correlations (*P* < 0.001) for the following (model species: fitted species) pairs; (LeuMic), (PolMic) and (MicLeu) indicating that that *E. leucoxylon* is asynchronous with *E. microcarpa* and *E. microcarpa* is asynchronous with *E. polyanthemos* (only via the EKF-based residuals) (**Figure 7** RHS); in agreement with the rule for asynchrony (**Table 4**) and Moranbased AR analysis of the flowering intensities.

Both the MTDg- and EKF-based models show that *E. tricarpa* is not asynchronous with *E. polyanthemos* (**Tables 7** and **8**). Note that for this species pairing *E. tricarpa* and *E. polyanthemos* (i.e. TriPol and PolTri) the associated sum of the probabilities for transitions P(A) and P(B) (both species off/on to one species off/ on) is equal to 0.802 (**Table 4**), which is just above the to the threshold for asynchrony of 0.80.

4.3 Principal component analysis on the (λ, Q, d, s) parameters

In this section a novel approach which invokes a principal component analysis (PCA) of the resultant (λ, Q, d, s) parameters (Section 2.2) which details the weight λ , q, d and s parameters from the MTD (n = 4) models) is performed. The resultant two dimensional PCA axis plots (**Figure 8**) of the rotated (λ , Q, d, s)-based PCs provides an informative visualisation of the synchronous and asynchronous species groupings (of n > 2 species) allowing for interpretation of the main climate drivers and climatic profiles (e.g.+/- or (-/+)) detailed in **Table 6**.

Model species	mic	pol	leu	tri				
Synchronous fitted species	tri (0.12)	leu (0.19)	pol (0.18)	leu (0.26)				
			tri (0.33)					
Asynchronous fitted species leu (-0.17°) mic (-0.10°)								
A negative and significant correlation indicates an asynchronous species pair.								

Table 8.

Significant Moran correlations (in brackets) from the EKF models.



Figure 8. Distances in the (λ, Q, d, s) parameters among the 4 species—without interaction terms (left) or with interaction terms (right).

The resulting parameters estimated from the MTDg models with and without interaction terms can be compared among all four species using **Figure 8**, which shows that the separation of *E. tricarpa* (-/+) and *E. microcarpa* (+/-) from other species along the horizontal axis 1, is due to the effect of mean temperature. Although *E. leucoxylon* is affected by the similar lag 1 and 11 month flowering terms as *E. polyanthemos*, *E. leucoxylon* (-/+) commences flowering at low temperature and shuts down at high temperatures. *E. microcarpa* begins flowering at high temperature (+/-). **Figure 8** also displays the similarity (synchronicity) of *E. leucoxylon* and *E. polyanthemos*.

5. Discussion and conclusion

The highest degree of synchrony (via the B-MTD rules of synchrony, the MTD models and Moran AR method) occurs between *E. leucoxylon* and *E. tricarpa*; then followed by *E. polyanthemos* and *E. leucoxylon* which indicates the potential for intense competition for potential pollinators, and therefore the prospect for a high level of hybridization. Both these species pairs were shown to be synchronous by Keatley et al., [20]; with *E. leucoxylon* and *E. tricarpa* having 6 years of no overlap (and a long term mean synchrony value of 0.62); and *E. polyanthemos* and *E. leucoxylon* having 5 years of the 31 years (between 1940 and 1970) with no overlap (long term mean synchrony value of 0.51); as quantified in [20]. The degree of synchrony or overlap of flowering was however determined using the method outlined in [22] which measures the extent of overlapping in the flowering periods among pairs of individuals in a population.

E. leucoxylon is the only species to synchronise flowering with *E. tricarpa*, as shown by all three methods, namely the B-MTD rules of synchrony, MTD models and Moran's AR method. Synchrony between *E. leucoxylon* and *E. tricarpa*, may be explained in terms of niche/competition and also facilitation may be a factor, due to their different modes of flower production. This agrees with the findings of [20]. Interestingly the MTD models discussed here (see also [6, 16, 24]) show that the climatic drivers or signature of *E. leucoxylon* and *E. tricarpa* is similar with respect to temperature, in that both exhibit decreased flowering with increased temperature.

Likewise *E. leucoxylon* is the only species to synchronise flowering with *E. polyanthemos. E. leucoxylon* and *E. polyanthemos* sometimes occur in the same

geographical area; and earlier studies have shown they overlap significantly [20]. From the flowering behaviour indices of Keatley and Hudson in [23], *E. leucoxylon* and *E. polyanthemos* were shown to have temporally separated months of peak flowering, September and November, respectively; likewise their flowering commencement months May and October, respectively. These two species can occur in the same geographical area and their flowering period. Differentiation of these two species is based on their differing months of peak flowering as well as their separated months of most probable flowering; October and November, respectively. Likewise their flowering commencement months differ, May and October, respectively.

The least degree of synchrony (via the B-MTD rules of synchrony, the MTD models and Moran method) is shown in this chapter to occur between *E. leucoxylon* and *E. microcarpa*; then followed by *E. polyanthemos* and *E. microcarpa*. Our results agree with the findings in [20], which established that a cross between *E. leucoxylon* and *E. microcarpa* is impossible. In terms of climatic signatures: the flowering of *E. microcarpa* behaves differently from *E. leucoxylon* and *E. tricarpa*. *E. microcarpa* flowers at higher temperature and its flowering has a significant and positive relationship with flowering a year ago, refer also to the results reported in [23].

Eucalyptus tricarpa and *E. polyanthemos* were shown in this chapter also to be asynchronous (discordant or out of phase). This is in agreement with conclusions reported in [2]. The MTDg model found a significant interaction between two climate variables, mean temperature and rainfall on the flowering of *E. polyanthemos*. As flowering is viewed as either 'off' or 'on' this interaction appears to be delineating *E. polyanthemos*' flowering period. It usually commences flowering in late spring—as mean temperature is increasing and rainfall is decreasing and ceases in early summer; just prior to the warmest mean temperature and lowest rainfall.

Specific temperature thresholds for commencement and for the cessation of flowering for the four species studied here, have been established, see [5, 7, 8]. For example, *E. microcarpa* was shown to flower at high temperatures, and *E. leucoxylon* and *E. tricarpa* both at lower temperatures. The flowering of *E. polyanthemos* was shown to be impacted by both rainfall and temperature, with increased flowering when conditions were either cool and dry, or hot and wet—indicative of a rainfall by temperature interaction.

Moran residual analysis and the B-MTD analysis described in this chapter showed that *E. tricarpa* and *E. microcarpa* did not exhibit a significant synchronous nor an asynchronous relationship. However, for this species pairing, the associated sum of the probabilities for transitions A and B (both off/on to one off/on) is 0.591, which is close to the threshold for synchrony of 0.65. Indeed the more sophisticated MTDg modelling approach which incorporates covariates (mean temperature and rainfall) with interactions, showed that *E. microcarpa* and *E. tricarpa* are synchronous, wherein the MTDg model allows for prior lag 1 to lag 12 month flowering effects and climate covariates.

SOM-based clustering [4] and Moran AR (2) tests also found that *E. polyanthemos* was asynchronous to *E. microcarpa* and *E. tricarpa*, in agreement with the extended Kalman filter (EKF)-based synchrony measures in [15, 21]. Note also it was demonstrated in [20] that *E. polyanthemos* and *E. microcarpa* have 25 years with no overlap (with a long term mean synchrony value of 0.29). Note that the more sophisticated MTDg modelling approach which incorporates covariates (mean temperature and rainfall) with interactions, showed that indeed *E. microcarpa* and *E. tricarpa* are synchronous, wherein the MTDg model allows for prior lag 1 to lag 12 month flowering effects and climate covariates.

Recently synchronisation of eucalypt flowering is shown to be a complex mechanism that incorporates all the flowering elements—flowering duration, timing of

peak flowering, and the timing of start and finishing of flowering, as well as possibly specific climate drivers for flowering [4]. The four species studied were shown to be influenced by temperature and rainfall and as a consequence their flowering phenology will change in response to climate change. This in turn will have an impact on species interactions and community [4].

Extensions of the B-MTD models to allow for climate covariates and for the comparison of more than 2 species at a time (a so-called multivariate M-MTD) is the topic of future work. Other forthcoming research is to examine the timing and a/synchronisation of the within species phenostages of both budding and flowering. Refer to earlier work using wavelets [26] and Generalized Additive Model for Location, Scale and Shape (GAMLSS) [27] to model the relationship between climate (mean monthly minimum, maximum temperatures and rainfall) during bud development and the flowering cycles of *Eucalyptus leucoxylon* and *E. tricarpa* from the Maryborough region of Victoria between 1940 and 1962. Monthly behaviour (start, peak, finish, monthly intensity, duration and success) in budding and flowering was assessed using, as in this current chapter, the indices of Keatley in [23].

Author details

Irene Hudson^{1*}, Susan Won Sun Kim² and Marie Keatley³

1 Department of Mathematical Sciences, Royal Melbourne Institute of Technology, Melbourne, Australia

2 South Australian Health and Medical Research Institute, Adelaide, South Australia

3 School of Ecosystem and Forest Sciences, The University of Melbourne, Melbourne, Australia

*Address all correspondence to: irene.hudson@rmit.edu.au

IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

References

[1] Brooker MIH, Kleinig DA. South Eastern Australia. In: Field Guide to Eucalypts: Vol. 1. 3rd ed. Hawthorn: Bloomings Books; 2006. 383 p

[2] MacNally R, Horrocks G. Landscapescale conservation of an endangered migrant: The swift parrot (*Lathamus discolor*) in its winter range. Biological Conservation. 2000;**92**:335-343

[3] Hudson IL. Interdisciplinary approaches: Towards new statistical methods for phenological studies. Climatic Change. 2010;**100**:143-171. DOI: 10.1007/s10584-010-9859-9

[4] Hudson IL, Keatley MR, Lee L. Using self-organising maps (SOMs) to assess synchronies: An application to historical eucalypt flowering records. International Journal of Biometeorology. 2011;55: 879-904

[5] Hudson IL, Keatley MR, Kim SW. Climatic influences on the flowering phenology of four eucalypts: A GAMLSS approach. In: Hudson IL, Keatley MR, editors. Phenological Research: Methods for Environmental and Climate Change Analysis. Dordrecht: Springer; 2010. pp. 213-237. DOI: 10.1007/978-90-481-3335-2_10

[6] Hudson IL, Keatley MR, Kim SW. Modelling the flowering of four eucalypt species using new mixture transition distribution models. In: Hudson IL, Keatley MR, editors. Phenological Research: Methods for Environmental and Climate Change Analysis. Dordrecht: Springer; 2010b. pp. 315-340. DOI: 10.1007/978-90-481-3335-2_14

[7] Hudson IL, Keatley MR. Phenological Research: Methods for Environmental and Climate Change Analysis. Dordrecht: Springer; 2010

[8] Hudson IL, Keatley MR, Kang I. Wavelet characterization of eucalypt flowering and the influence of climate. Environmental and Ecological Statistics. 2011;**18**:513-533. DOI: 10.1007/ s10651-010-0149-5

[9] Kim SW, Hudson IL, Keatley MR, Anderssen RS, Braddock RD, Newham LTH. Modelling the flowering of four eucalypts species via MTDg with interactions. In: 18th World IMACS Congress and MODSIM09 International Congress on Modelling and Simulation.
13-17 July 2009; Cairns, Australia: Modelling and Simulation Society of Australia and New Zealand and International Association for Mathematics and Computers in Simulation. 2009. pp. 2625-2631

[10] Raftery AE. A model for high-order Markov chains. Journal of the Royal Statistical Society: Series B: Methodological. 1985;47:528-539

[11] Berchtold A. March V.2.01. Markovian Models Computation and Analysis Users Guide. 2004. Available from: http://www.andreberchtold.com/ march.html

[12] Berchtold A. March v.3.00 Markovian Models Computation and Analysis Users Guide. 2006. Available from: http://www.andreberchtold.com/ march.html

[13] Berchtold A, Raftery AE. The mixture transition distribution model for high-order Markov chains and nongaussian time series. Statistical Science. 2002;**17**:328-356

[14] Fournier DA. AD Model Builder, Version 5.0.1. Canada: Otter Research Ltd.; 2000

[15] Kim SW, Hudson IL, Keatley MR, Agrawal M, Eilers P. Modelling and synchronization of four eucalypt species via mixed transition distribution (MTD)

and extended Kalman filter (EKF). In: Proceedings of the 23rd International Workshop on Statistical Modelling, 23rd International Workshop on Statistical Modelling; 7-11 July 2008; Utrecht, Netherlands. 2008. pp. 287-292

[16] Kim S. Bayesian and non-Bayesian mixture paradigms for clustering multivariate data: Time series synchrony tests. Chapter 4.
University of South Australia; 2011.
pp. 60-118. Available from: http:// researchoutputs.unisa.edu.au/1959.8/
138604

[17] Moran PAP. The statistical analysis of the Canadian lynx cycle. I. Structure and prediction. Australian Journal of Zoology. 1953a;**1**:163-173

[18] Moran PAP. The statistical analysis of the Canadian lynx cycle. II.Synchronization and meteorology.Australian Journal of Zoology. 1953b;1: 291-298

[19] van der Merwe R. Quick-Start Guide for ReBel Toolkit. Oregon Health and Science University; 2004

[20] Keatley MR, Hudson IL,Fletcher TD. Long-term floweringsynchrony of box-ironbark eucalypts.Australian Journal of Botany. 2004;52:47-54. DOI: 10.1071/BT03017

[21] Kim S. Bayesian and non-Bayesian mixture paradigms for clustering multivariate data: Time series synchrony tests. Chapter 5. University of South Australia; 2011. pp. 119-123. Available from: http://researchoutputs. unisa.edu.au/1959.8/138604

[22] Augspurger CK. Flowering synchrony of neotropical plants. In: WG D'A, Correa MD, editors. The Botany and Natural History of Panama. Saint Louis: Missouri Botanical Garden; 1985. pp. 235-243 [23] Keatley MR, Hudson IL. A comparison of long-term flowering patterns of box-ironbark species in Havelock and Rushworth forests. Environmental Modeling and Assessment. 2007;**12**:279-292. DOI: 10.1007/s10666-006-9063-5

[24] Hudson IL, Kim SW, Keatley MR, Anderssen RS, Braddock RD, Newham LTH. Climatic influences on the flowering phenology of four eucalypts: A GAMLSS approach. In: 18th World IMACS Congress and MODSIM09 International Congress on Modelling and Simulation; 13-17 July 2009. Cairns, Australia: Modelling and Simulation Society of Australia and New Zealand and International Association for Mathematics and Computers in Simulation. 2009. pp. 2611-2617

[25] Hudson IL, Keatley MR, Kim SW, Kang I. Synchronicity in phenology:
From PAP Moran to now. In: Australian Statistical Conference/New Zealand
Statistical Association (ASC/NZSA)
Conference; 3-6 July 2006, Auckland, New Zealand. 2006

[26] Hudson IL, Kang I, Keatley MR, Weber T, Mcphee MJ, Anderssen RS. Wavelet characterization of eucalypt flowering and the influence of climate and budding. In: MODSIM 2015, 21st International Congress on Modelling and Simulation; Modelling and Simulation Society of Australia and New Zealand. 2015. pp. 1813-1819

[27] Hudson IL, Keatley MR, Piantadosi J, Anderssen RS, Boland J. Scoping the budding and climate impacts on eucalypt flowering: Nonlinear time series decomposition modelling. In: MODSIM2013, 20th International Congress on Modelling and Simulation; 1-6 December; Modelling and Simulation Society of Australia and New Zealand. 2013. pp. 1582-1588