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Chapter Thymic Senescence

Krisztian Kvell

Abstract

Thymic senescence develops in every person, although at different pace. Thymic senescence significantly lowers the production of naive T cells, leading to increased incidence of infections, cancer and autoimmune diseases. Certain external factors can accelerate thymic senescence. These include chemicals (copper-chelators), hormones (androgens), infections (viruses, fungi, protozoa). Others may slow the aging process of the thymus including perturbations to the hormonal (sex-steroid) system, genetic alterations (PPARgamma deficiency) or chemical compounds (PPARgamma antagonists). Thymic senescence research may provide insight to underlying molecular events and potentially appoint novel therapeutic targets for senescence intervention strategies. These hold promise to postpone thymus senescence and enhance T cell production. That would result in a decreased incidence of infections, cancer and autoimmune diseases, currently affecting the elderly. The attributed drop in health-care costs and gain in quality of life share tremendous economic and social interest.

Keywords: thymus, senescence, adipose tissue

1. The aging thymus

Transcription factor TBX-1 is a mastermind in the formation of the third pharyngeal pouch involved in thymus organogenesis during embryonic development [1]. Patients with 22q11.2DS that impairs TBX-1 often present thymus hypoplasia. Similarly, Tbx-1^{null} mice develop hypoplasia of the thymus [2, 3]. In both cases, defective thymus organogenesis leads to impaired thymocyte development [4]. However, as reported recently, the role of TBX-1 in thymus organogenesis is not straightforward. Ectopic forced expression of TBX-1 can inhibit transcription factor FoxN1, the mastermind of thymic epithelial identity thus indirectly impair thymus identity via sustained presence [5]. The thymus contains developing T cells (aka thymocytes) along with the non-lymphoid thymic stromal elements comprising the microenvironment that promotes thymocyte differentiation. Stromal elements include thymic epithelial cells (aka TECs), mesenchymal cells, endothelial cells as well as non-lymphoid hematopoietic cells (e.g., dendritic cells or macrophages). TECs constitute the main functional stromal cell type necessary to promote thymocyte differentiation [6, 7]. Soon after birth the thymus expands to increase the output of naive T cells, in order to colonize available niches in the periphery [8–10]. Cortical TECs (aka cTECs) are required for T lineage commitment, along with thymocyte expansion and differentiation, and positive selection. Medullary TECs (mTECs) are necessary for the induction of central tolerance and subsequent stages of thymocyte maturation before leaving the thymus. Of note, in order to maintain the well organized cortical and medullary compartments active (reverse) intercellular signaling is also required from developing thymocytes towards TECs

(aka cross-talk) [11, 12]. At a vaguely defined time point, the thymus begins to show involution, resulting in adipose degeneration of the organ; hence the process termed adipose involution. This senescent process is accompanied by the stepwise disorganization of thymic compartments, also shifting TEC subset ratios and reducing naive T cell production. Although the detailed mechanisms triggering these processes remain to be fully elucidated, they finally deteriorate thymus structure and function, severely impairing the output of fresh naive T cells. Decline of fresh naive T cells results in the inverse increase of memory T cell representation due to aging [13–15]. The observed bias in cTEC:mTEC ratio, and the fading of the most differentiated MHC class II-expressing TEC subsets leads to the development of a less complex medullary architecture along with the blurring of the cortico-medullary junction. This is followed by the focal disappearance of epithelial cells, gradually being replaced by adipose tissue in the perivascular spaces [16–20]. There is mounting evidence that adipose cells may have an thymic origin. Thymic adipose cells also produce an array of cytokines and signaling molecules that directly affect (impair) thymopoiesis [21–25]. As a result, although the appearance of thymic adipocytes may not trigger involution, their increasing presence with senescence can indirectly facilitate or perhaps even directly deteriorate thymus function. Thymus involution likely develops as a sum of failure of thymocyte progenitors and the inappropriate function of TEC compartments. It has been reported that the number of early T lineage precursors (aka ETPs) shows a gradual decline with senescence [26]. Reconstitution experiments of senescent thymi with bone marrow precursors from young donors cannot restore thymic compartments nor rescue impaired thymopoiesis. The opposite, however, reconstitution of young recipients using senescent bone marrow cells does not impair thymopoiesis [27, 28]. The genetic inactivation of cell cycle inhibitor p27 (aka Cdkn1b) also leads to the development of an enlarged thymus and enhances fresh naive T cell output along with normal stromal organization [29–33]. Recent thymic emigrants (aka RTEs) show a decrease upon enhanced expression of LIF, SCF, IL-6, and M-CSF [34, 35].

2. Characterization of thymic adipose tissue

There are significant differences between adipose tissue subtypes. At least three subtypes are distinguished: white adipose tissue (WAT), brown adipose tissue (BAT) and the recently described beige adipose tissue. White adipose tissue stores energy, brown adipose tissue generates heat (via NST or non-shivering thermogenesis), while beige adipocytes act as intermediates. It has currently been described that thymic adipose involution yields beige adipose tissue based on its gene expression, miRNA, histology and metabolic profile [36]. In terms of gene expression and histology characteristic epithelial markers show down-regulation (FoxN1, EPCAM1, MHCII, Wnt4) (see **Figure 1**). Considering the miRNA profile beige-adipose tissue-associated miRNA species show supportive changes (miR27a, miR106b, miR155) (see Figure 1). While PPARgamma is the mastermind of all adipose tissue subtypes, TBX-1 has been acknowledged as a key and specific marker of beige adipose tissue development [17–20]. Beige adipocytes respond to adrenergic stimuli by thermogenesis via mitochondrial uncoupling of biochemical degradation and energy production [21]. Along with TBX-1 other beige-indicative markers have also been reported. These include mitochondrial uncoupling proteins (mostly UCP-1), EAR2 (also known as Nr2f6) and CD137 (also known as Tnfrsf9) [22]. The above-mentioned adipose and beige markers show up-regulation along (see Figure 1). The adult thymus expresses PPARgamma, TBX-1 and UCP-1 in the epithelial compartment, and latter two have been reported to initiate beige adipose

Epithelial identity

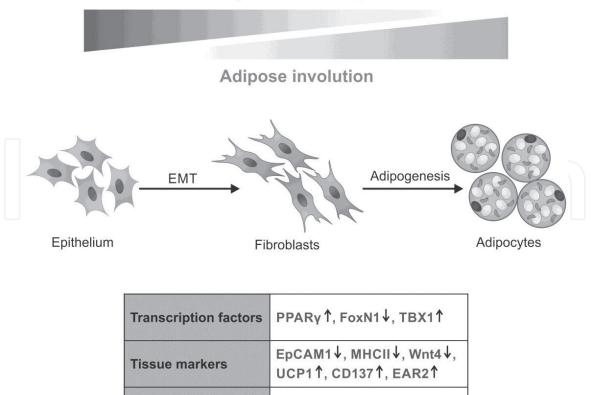


Figure 1.

Key molecular events of thymic senescence.

miRNA species

tissue development. Thymic adipose tissue may also bee classified based on cellular analysis from an adipocyte perspective [23–26]. Thymus tissue appears to be unique expressing TBX-1 during embryonic development and also during senescence embedded in different contexts. It is appreciable that TBX-1 plays a role in thymus organogenesis (immune peak) and thymic adipose involution (metabolic peak). This suggests an intersection of immunity and metabolism, and a dual role of TBX-1 showing bimodal expression [36].

27a↓, 106b↑, 155↓

3. Natural resistance to senescence

A medical condition termed persistent thymus has been known for long [37]. In the affected population the thymus is rescued from involution. These individuals, however, have severe defects in their hormonal system, affecting the level of sex steroids. It is the lack of androgen-effect that prevents thymus involution on one hand, but hampers the endocrine system on the other hand. Recently another medical condition termed FPLD3 (familial partial lipodystrophy type 3) has also been associated with the lack of thymus involution [38]. FPLD3 also derails the hormonal system by affecting PPARgamma activity. As for all adipose tissue subtypes, PPARgamma plays a crucial role during thymus adipose involution as well [39]. It has been suggested by others previously based on direct fate-mapping experiments that with senescence thymic adipose tissue develops from the thymic stromal or epithelial compartment [22]. In further support, epithelial to adipose trans-differentiation has been reported to occur as indicated by the presence by EpCAM-1/PPARgamma doublepositive cells at a given time point during thymus senescence. Such cells express cell surface markers as memories of their fading thymic epithelial identity (EpCAM-1), yet already show signs of their novel adipocyte differentiation program in their nuclei (PPARgamma). Further experiments showed that the medullary compartment is rescued from age-related shrinking in case of PPARgamma deficiency. Prolonged survival of thymus stromal niche provides permissive environment for sustained fresh naïve T cell production as indicated by increased mTrec values. Thymocyte subpopulations were equally supported by PPARgamma deficiency and fresh naive T cells outnumbered memory T cells despite age. The sustained support of fresh naive T cells provides functional advantages even at elevated ages. Oral consumption of foreign T-depended antigen initiates immune tolerance to block potential immune response, even along with parallel immunization. Unfortunately, this tolerance is impaired at old age [40–42] Loss of oral tolerance is a potential link to increasing food intolerance prevalence [43–46]. However, tolerance is rescued by PPARgamma deficiency at senior age [38]. In senior individuals protection from seasonal flu strains declines despite annual vaccination [47–49]. The cause: low levels of neutralizing antibody titers due to lacking naïve T-cells required for T-B cooperation. This, however, is also rescued by PPARgamma deficiency [38].

4. Induced rejuvenation

It has been reported early on that the thymus may be regenerated by a variety of interventions (aka thymic rebound) [50]. FoxN1 (a forkhead class transcription factor) is the mastermind of TEC differentiation [51–55]. FoxN1 has also been shown to promote proliferation [56]. Reducing (but not fully diminishing) FoxN1 expression early on triggers premature thymus involution (aka thymus progeria). The opposite, however, over-expression of FoxN1 efficiently postpones thymus involution [57]. Among secreted factors, Wnt4 and keratinocyte growth factor (aka KGF) have also verified as key factors of both thymic senescence and rebound [58–61]. The onset of adolescence presents a frequently proposed physiological cause for thymic degeneration. In accordance, both chemical and surgical castration that result in sex-steroid ablation (SSA) yield thymic rebound [62, 63]. SSA-triggered thymic rebound correlates with both increased thymus size and thymocyte number leading to increased fresh naive T cell production. At the histological level this is suggested by the recovery of the cortico-medullary junction [64]. Accordingly, systemic hormonal changes associated with senescence partly explain changes observed during thymic senescence. In harmony, deletion of the androgen receptor results in an enlarged thymus and resistance to androgen induced thymus atrophy [65]. This is also in line with reports showing that the thymus reaches peak size and productivity early after birth, and not later at puberty [66–68]. Unfortunately, castrationinduced rebound is only a transient phenomenon, and the thymus re-involutes within a couple of weeks. Apparently, although SSA may trigger the expansion of the thymus, yet does not rejuvenate it [69]. In the case of the thymus, in comparison with other organs, little is known about the molecular and cellular mechanisms that control its development and maintenance. FoxN1 certainly is a mastermind linking development and maintenance of the thymic microenvironment throughout life, yet some TEC differentiation also occurs independent from Foxn1 [70].

5. Novel trends of rejuvenation

Transcription factor FoxN1 - the mastermind of thymus organogenesis and identity - is a known as the molecular target of the glycolipoprotein Wnt4 [71, 72].

For this reason Wnt4 plays a crucial role in thymus development and maintenance [73–77]. With senescence thymic epithelial cells secrete less Wnt4, however, their Frizzled receptors (Fz4 and Fz6) become up-regulated as compensatory mechanism [78]. It is his loss of Wnt4 expression that allows for thymic adipose involution to develop due to PPARgamma-effect [79, 80]. The Wnt/b-catenin pathway and PPARgamma act as inhibitors of each other hence exogenous Wnt4 can reinforce thymic epithelial identity [79–84]. Wnt4 loses activity when purified, because the Wnt molecules travel in extracellular vesicles (EVs, or exosomes in this case) or on their surfaces [85, 86]. It has been reported that miR27b also specifically inhibits PPARgamma activity [87, 88]. The miRNA species are known travel in EVs and in exosomes as well [89, 90]. The thymus is a rich source of exosomes with immunological relevance in e.g. thymocyte selection [91–94]. As a combination of the above, artificially produced (transgenic) exosomes containing Wnt4 and miR27b in excess can block PPARgamma-effect in thymic epithelial cells thus efficiently counteracting senescence observed as thymic adipose involution [95].

World population is approaching 7.7 billion as of 2019 [96]. Global population increases due to increasing life expectancy, rather than increasing birth rate. However, increasing lifespan is not proportionally attributed with increasing health-span. As a result social expenses rise and novel solutions are urged. Central immune (thymus) senescence research based novel solutions can potentially improve senior immune fitness through decreasing the incidence of infections, malignant and autoimmune disorders. These could also thus alleviate the current burden on healthcare systems and increase quality of life in the elderly. An ultimate goal is to prolong immune fitness and realign it with constantly increasing lifespan.

With aging the thymus shows adipose involution. During this process thymic epithelial cells trans-differentiate into beige adipocytes through an intermediate fibroblast stage. Key molecular events are summarized at the level of transcription factors, tissue markers and miRNA species.

Acknowledgements

The project has been supported by the European Union, cofinanced by the European Social Fund: Comprehensive Development for Implementing Smart Specialization Strategies at the University of Pécs (EFOP-3.6.1.-16-2016-00004).

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References

[1] Gao S, Li X, Amendt BA. Understanding the role of Tbx1 as a candidate gene for 22q11.2 deletion syndrome. Current Allergy and Asthma Reports. 2013;**13**:613-621. DOI: 10.1007/ s11882-013-0384-6

[2] Farley AM, Morris LX, Vroegindeweij
E, Depreter MLG, Vaidya H, Stenhouse
FH, et al. Dynamics of thymus
organogenesis and colonization in early
human development. Development.
2013;140:2015-2026. DOI: 10.1242/
dev.087320

[3] Jerome LA, Papaioannou VE. DiGeorge syndrome phenotype in mice mutant for the T-box gene, Tbx1. Nature Genetics. 2001;**27**:286-291. DOI: 10.1038/85845

[4] Sirianni MC, Businco L, Seminara R, Aiuti F. Severe combined immunodeficiencies, primary T-cell defects and DiGeorge syndrome in humans: Characterization by monoclonal antibodies and natural killer cell activity. Clinical Immunology and Immunopathology. 1983;**28**:361-370. Available at: http://www.ncbi.nlm.nih. gov/pubmed/6349883

[5] Reeh KAG, Cardenas KT, Bain VE, Liu Z, Laurent M, Manley NR, et al. Ectopic TBX1 suppresses thymic epithelial cell differentiation and proliferation during thymus organogenesis. Development. 2014;**141**:2950-2958. DOI: 10.1242/ dev.111641

[6] Chinn IK, Blackburn CC, Manley NR, Sempowski GD. Changes in primary lymphoid organs with aging. Seminars in Immunology. 2012;**24**(5):309-320

[7] Chaudhry MS, Velardi E, Dudakov JA, van den Brink MR. Thymus: The next (re)generation. Immunological Reviews. 2016;**271**(1):56-71 [8] Adkins B. T-cell function in newborn mice and humans. Immunology Today. 1999;**20**:330-335

[9] Adkins B. Peripheral CD4+ lymphocytes derived from fetal versus adult thymic precursors differ phenotypically and functionally. Journal of Immunology. 2003;**171**:5157-5164

[10] Min B, McHugh R, Sempowski GD, Mackall C, Foucras G, Paul WE. Neonates support lymphopeniainduced proliferation. Immunity. 2003;**18**:131-140

[11] Anderson G, Jenkinson WE, Jones T, Parnell SM, Kinsella FA, White AJ, et al. Establishment and functioning of intrathymic microenvironments. Immunological Reviews. 2006;**209**: 10-27

[12] Gill J, Malin M, Sutherland J, Gray D, Hollander G, Boyd R. Thymic generation and regeneration. Immunological Reviews. 2003;**195**:28-50

[13] Min H, Montecino-Rodriguez E, Dorshkind K. Reduction in the developmental potential of intrathymic T cell progenitors with age. Journal of Immunology. 2004;**173**:245-250

[14] Min H, Montecino-Rodriguez E, Dorshkind K. Reassessing the role of growth hormone and sex steroids in thymic involution. Clinical Immunology. 2006;**118**:117-123

[15] Haynes L, Swain SL. Why aging T cells fail: Implications for vaccination. Immunity. 2006;**24**:663-666

[16] Taub DD, Longo DL. Insightsinto thymic aging and regeneration.Immunological Reviews. 2005;205:72-93

[17] Gray DH, Seach N, Ueno T, Milton MK, Liston A, Lew AM, et al. Developmental kinetics, turnover, and

stimulatory capacity of thymic epithelial cells. Blood. 2006;**108**:3777-3785

[18] Chen L, Xiao S, Manley NR. Foxn1 is required to maintain the postnatal thymic microenvironment in a dosage-sensitive manner. Blood. 2009;**113**:567-574

[19] Griffith AV, Fallahi M, Venables T, Petrie HT. Persistent degenerative changes in thymic organ function revealed by an inducible model of organ regrowth. Aging Cell. 2012;**11**:169-177

[20] Takeoka Y, Chen SY, Yago H, Boyd R, Suehiro S, Shultz LD, et al. The murine thymic microenvironment: Changes with age. International Archives of Allergy and Immunology. 1996;**111**:5-12

[21] Youm YH, Yang H, Amin R, Smith SR, Leff T, Dixit VD. Thiazolidinedione treatment and constitutive-PPARgamma activation induces ectopic adipogenesis and promotes age-related thymic involution. Aging Cell. 2010;**9**:478-489

[22] Youm YH, Yang H, Sun Y, Smith RG, Manley NR, Vandanmagsar B, et al.
Deficient ghrelin receptor-mediated signaling compromises thymic stromal cell microenvironment by accelerating thymic adiposity. The Journal of Biological Chemistry.
2009;284:7068-7077

[23] Dixit VD. Thymic fatness and approaches to enhance thymopoietic fitness in aging. Current Opinion in Immunology. 2010;**22**:521-528

[24] Sempowski GD, Hale LP, Sundy JS, Massey JM, Koup RA, Douek DC, et al. Leukemia inhibitory factor, oncostatin M, IL-6, and stem cell factor mRNA expression in human thymus increases with age and is associated with thymic atrophy. Journal of Immunology. 2000;**164**:2180-2187

[25] Yang H, Youm YH, Dixit VD. Inhibition of thymic adipogenesis by caloric restriction is coupled with reduction in age-related thymic involution. Journal of Immunology. 2009;**183**:3040-3052

[26] Yang H, Youm YH, Vandanmagsar B, Rood J, Kumar KG, Butler AA, et al. Obesity accelerates thymic aging. Blood. 2009;**114**:3803-3812

[27] Heng TS, Goldberg GL, Gray DH, Sutherland JS, Chidgey AP, Boyd RL. Effects of castration on thymocyte development in two different models of thymic involution. Journal of Immunology. 2005;**175**:2982-2993

[28] Mackall CL, Punt JA, Morgan P, Farr AG, Gress RE. Thymic function in young/old chimeras: Substantial thymic T cell regenerative capacity despite irreversible age-associated thymic involution. European Journal of Immunology. 1998;**28**:1886-1893

[29] Zhu X, Gui J, Dohkan J, Cheng L, Barnes PF, Su DM. Lymphohematopoietic progenitors do not have a synchronized defect with age-related thymic involution. Aging Cell. 2007;**6**:663-672

[30] Klug DB, Crouch E, Carter C, Coghlan L, Conti CJ, Richie ER. Transgenic expression of cyclin D1 in thymic epithelial precursors promotes epithelial and T cell development. Journal of Immunology. 2000;**164**:1881-1888

[31] Nowell CS, Bredenkamp N, Tetelin S, Jin X, Tischner C, Vaidya H, et al. Foxn1 regulates lineage progression in cortical and medullary thymic epithelial cells but is dispensable for medullary sublineage divergence. PLoS Genetics. 2011;7:e1002348

[32] Nakayama K, Ishida N, Shirane M, Inomata A, Inoue T, Shishido N, et al. Mice lacking p27(Kip1) display increased body size, multiple organ hyperplasia, retinal dysplasia, and pituitary tumors. Cell. 1996;**85**:707-720 [33] Kiyokawa H, Kineman RD, Manova-Todorova KO, Soares VC, Hoffman ES, Ono M, et al. Enhanced growth of mice lacking the cyclin-dependent kinase inhibitor function of p27(Kip1). Cell. 1996;**85**:721-732

[34] Fero ML, Rivkin M, Tasch M, Porter P, Carow CE, Firpo E, et al. A syndrome of multiorgan hyperplasia with features of gigantism, tumorigenesis, and female sterility in p27(Kip1)-deficient mice. Cell. 1996;**85**:733-744

[35] Sempowski GD, Rhein ME, Scearce RM, Haynes BF. Leukemia inhibitory factor is a mediator of Escherichia coli lipopolysaccharideinduced acute thymic atrophy. European Journal of Immunology. 2002;**32**:3066-3070

[36] Banfai K, Ernszt D, Pap A, Bai P, Garai K, Belharazem D, et al. Beige' cross talk between the immune system and metabolism. Frontiers in Endocrinology. 18 June 2019. https:// doi.org/10.3389/fendo.2019.00369

[37] Booth FR. Thymus gland, persistent or overactive. Veterinary Medicine.1950;45(10):387-388

[38] Ernszt D, Banfai K, Kellermayer Z, Pap A, Lord JM, Pongracz JE, et al. PPARgamma deficiency counteracts Thymic senescence. Frontiers in Immunology. 2017;**8**:1515

[39] Kvell K, Varecza Z, Bartis D, Hesse S, Parnell S, Anderson G, et al. Wnt4 and LAP2alpha as pacemakers of thymic epithelial senescence. PLoS One. 2010;**5**(5):e10701

[40] de Faria AM, Ficker SM, Speziali
E, Menezes JS, Stransky B, Silva
Rodrigues V, et al. Aging affects
oral tolerance induction but not its
maintenance in mice. Mechanisms
of Ageing and Development.
1998;102(1):67-80

[41] Kato H, Fujihashi K, Kato R, Dohi T, Fujihashi K, Hagiwara Y, et al. Lack of oral tolerance in aging is due to sequential loss of Peyer's patch cell interactions. International Immunology. 2003;**15**(2):145-158

[42] Song F, Guan Z, Gienapp IE, Shawler T, Benson J, Whitacre CC. The thymus plays a role in oral tolerance in experimental autoimmune encephalomyelitis. Journal of Immunology. 2006;**177**(3):1500-1509

[43] Galipeau HJ, Verdu EF. Gut microbes and adverse food reactions: Focus on gluten related disorders. Gut Microbes. 2014;5(5):594-605. DOI: 10.4161/19490976.2014.969635

[44] Marietta EV, Murray JA. Animal models to study gluten sensitivity.
Seminars in Immunopathology.
2012;**34**(4):497-511. DOI: 10.1007/ s00281-012-0315-y

[45] MacDonald TT. Evidence for cell-mediated hypersensitivity as an important pathogenetic mechanism in food intolerance. Clinical and Experimental Allergy. 1995;**25**(Suppl 1): 10-13

[46] Ferguson A, Ziegler K, Strobel S.Gluten intolerance (coeliac disease).Annals of Allergy. 1984;53(6 Pt 2):637-642

[47] Cromer D, van Hoek AJ, Jit M, Edmunds WJ, Fleming D, Miller E. The burden of influenza in England by age and clinical risk group: A statistical analysis to inform vaccine policy. The Journal of Infection. 2014;**68**(4):363-371. DOI: 10.1016/j. jinf.2013.11.013

[48] Ang LW, Lim C, Lee VJ, Ma S, Tiong WW, Ooi PL, et al. Influenza-associated hospitalizations, Singapore, 2004-2008 and 2010-2012. Emerging Infectious Diseases. 2014;**20**(10):1652-1660. DOI: 10.3201/eid2010.131768

[49] Haq K, McElhaney JE. Immunosenescence: Influenza vaccination and the elderly. Current Opinion in Immunology. 2014;**29**:38-42. DOI: 10.1016/j.coi.2014.03.008

[50] Billard MJ, Gruver AL, Sempowski GD. Acute endotoxin-induced Thymic atrophy is characterized by intrathymic inflammatory and wound healing responses. PLoS One. 2011;**6**:e17940

[51] Hale JS, Boursalian TE, Turk GL, Fink PJ. Thymic output in aged mice. Proceedings of the National Academy of Sciences of the United States of America. 2006;**103**:8447-8452

[52] Kaestner KH, Knochel W, Martinez DE. Unified nomenclature for the winged helix/forkhead transcription factors. Genes & Development. 2000;**14**:142-146

[53] Nehls M, Pfeifer D, Schorpp M, Hedrich H, Boehm T. New member of the winged-helix protein family disrupted in mouse and rat nude mutations. Nature. 1994;**372**:103-107

[54] Nehls M, Kyewski B, Messerle M, Waldschütz R, Schüddekopf K, Smith AJ, et al. Two genetically separable steps in the differentiation of thymic epithelium. Science. 1996;**272**:886-889

[55] Blackburn CC, Augustine CL, Li R, Harvey RP, Malin MA, Boyd RL, et al. The nu gene acts cell-autonomously and is required for differentiation of thymic epithelial progenitors. Proceedings of the National Academy of Sciences of the United States of America. 1996;**93**:5742-5746

[56] Su DM, Navarre S, Oh WJ, Condie BG, Manley NR. A domain of Foxn1 required for crosstalkdependent thymic epithelial cell differentiation. Nature Immunology. 2003;**4**:1128-1135

[57] Itoi M, Kawamoto H, Katsura Y, Amagai T. Two distinct steps of immigration of hematopoietic progenitors into the early thymus anlage. International Immunology. 2001;**13**:1203-1211

[58] Zook EC, Krishack PA, Zhang S, Zeleznik-Le NJ, Firulli AB, Witte PL, et al. Overexpression of Foxn1 attenuates age-associated thymic involution and prevents the expansion of peripheral CD4 memory T cells. Blood. 2011

[59] Suniara RK, Jenkinson EJ, Owen JJ. An essential role for thymic mesenchyme in early T cell development. The Journal of Experimental Medicine. 2000;**191**:1051-1056

[60] Erickson M, Morkowski S, Lehar S, Gillard G, Beers C, Dooley J, et al. Regulation of thymic epithelium by keratinocyte growth factor. Blood. 2002;**100**:3269-3278

[61] Revest JM, Suniara RK, Kerr K, Owen JJ, Dickson C. Development of the thymus requires signaling through the fibroblast growth factor receptor r2-iiib. Journal of Immunology. 2001;**167**:1954-1961

[62] Rossi SW, Jeker LT, Ueno T, Kuse S, Keller MP, Zuklys S, et al. Keratinocyte growth factor (KGF) enhances postnatal T-cell development via enhancements in proliferation and function of thymic epithelial cells. Blood. 2007;**109**:3803-3811

[63] Fitzpatrick FT, Kendall MD, Wheeler MJ, Adcock IM, Greenstein BD. Reappearance of thymus of ageing rats after orchidectomy. The Journal of Endocrinology. 1985;**106**:R17-R19

[64] Hince M, Sakkal S, Vlahos K, Dudakov J, Boyd R, Chidgey A. The role of sex steroids and gonadectomy in the control of thymic involution. Cellular Immunology. 2008;**252**:122-138 [65] Olsen NJ, Olson G, Viselli SM, Gu X, Kovacs WJ. Androgen receptors in thymic epithelium modulate thymus size and thymocyte development. Endocrinology. 2001;**142**:1278-1283

[66] Steinmann GG. Changes in the human thymus during aging. Current Topics in Pathology. 1986;**75**:43-88

[67] Steinmann GG, Klaus B, Muller-Hermelink HK. The involution of the ageing human thymic epithelium is independent of puberty. A morphometric study. Scandinavian Journal of Immunology. 1985;**22**:563-575

[68] George AJ, Ritter MA. Thymic involution with ageing: Obsolescence or good housekeeping? Immunology Today. 1996;**17**:267-272

[69] Henderson J. On the relationship of the thymus to the sexual organs. Journal of Physiology. 1904;**31**:222-229

[70] Zamisch M, Moore-Scott B, Su DM, Lucas PJ, Manley N, Richie ER. Ontogeny and regulation of IL-7expressing thymic epithelial cells. Journal of Immunology. 2005;**174**:60-67

[71] Romano R, Palamaro L, Fusco A, Giardino G, Gallo V, Del Vecchio L, et al. FOXN1: A master regulator gene of Thymic epithelial development program. Frontiers in Immunology. 2013;4:187. DOI: 10.3389/ fimmu.2013.00187

[72] Kvell K, Varecza Z, Bartis D, Hesse S, Parnell S, Anderson G, et al. Wnt4 and LAP2alpha as pacemakers of thymic epithelial senescence. PLoS One. 2010;**5**:e10701. DOI: 10.1371/journal. pone.0010701

[73] Pongracz JE, Parnell SM, Jones T, Anderson G, Jenkinson EJ. Overexpression of ICAT highlights a role for catenin-mediated canonical Wnt signalling in early T cell development. European Journal of Immunology. 2006;**36**:2376-2383. DOI: 10.1002/ eji.200535721

[74] Kvell K, Fejes AV, Parnell SM, Pongracz JE. Active Wnt/beta-catenin signaling is required for embryonic thymic epithelial development and functionality ex vivo. Immunobiology. 2014;**219**:644-652. DOI: 10.1016/j. imbio.2014.03.017

[75] Heinonen KM, Vanegas JR, Brochu S, Shan J, Vainio SJ, Perreault C. Wnt4 regulates thymic cellularity through the expansion of thymic epithelial cells and early thymic progenitors. Blood. 2011;**118**:5163-5173. DOI: 10.1182/blood-2011-04-350553

[76] Taub DD, Longo DL. Insights into thymic aging and regeneration. Immunological Reviews. 2005;**205**:72-93. DOI: 10.1111/j.0105-2896.2005.00275.x

[77] Pongracz J, Hare K, Harman B, Anderson G, Jenkinson EJ. Thymic epithelial cells provide Wnt signals to developing thymocytes. European Journal of Immunology. 2003;**33**: 1949-1956. DOI: 10.1002/eji.200323564

[78] Varecza Z, Kvell K, Talabér G, Miskei G, Csongei V, Bartis D, et al. Multiple suppression pathways of canonical Wnt signalling control thymic epithelial senescence. Mechanisms of Ageing and Development. 2011;**132**:249-256. DOI: 10.1016/j.mad.2011.04.007

[79] Marinova TT. Epithelial framework reorganization during human thymus involution. Gerontology. 2005;**51**:14-18. DOI: 10.1159/000081429

[80] Talaber G, Kvell K, Varecza Z, Boldizsar F, Parnell SM, Jenkinson EJ, et al. Wnt-4 protects thymic epithelial cells against dexamethasone-induced senescence. Rejuvenation Research. 2011;**14**:241-248. DOI: 10.1089/ rej.2010.1110

[81] Gustafson B, Smith U. The WNT inhibitor Dickkopf 1 and bone morphogenetic protein 4 rescue adipogenesis in hypertrophic obesity in humans. Diabetes. 2012;**61**:1217-1224. DOI: 10.2337/db11-1419

[82] Lecarpentier Y, Claes V, Vallée A, Hébert J-L. Thermodynamics in cancers: Opposing interactions between PPAR gamma and the canonical WNT/ beta-catenin pathway. Clinical and Translational Medicine. 2017;**6**:14. DOI: 10.1186/s40169-017-0144-7

[83] Lecarpentier Y, Vallée A. Opposite interplay between PPAR gamma and canonical Wnt/Beta-catenin pathway in amyotrophic lateral sclerosis. Frontiers in Neurology. 2016;7:100. DOI: 10.3389/ fneur.2016.00100

[84] Kvell K, Pongracz JE. Central Immune Senescence, Reversal Potentials. Rijeka: InTech; 2012. Available at: http://www. ncbi.nlm.nih.gov/pubmed/28045481 [Accessed: 04-July-2018]

[85] Willert K, Brown JD, Danenberg E, Duncan AW, Weissman IL, Reya T, et al. Wnt proteins are lipid-modified and can act as stem cell growth factors. Nature. 2003;**423**:448-452. DOI: 10.1038/ nature01611

[86] Zhang L, Wrana JL. The emerging role of exosomes in Wnt secretion and transport. Current Opinion in Genetics & Development. 2014;27:14-19. DOI: 10.1016/j.gde.2014.03.006

[87] Wang T, Xu Z. miR-27 promotes osteoblast differentiation by modulating Wnt signaling. Biochemical and Biophysical Research Communications. 2010;**402**:186-189. DOI: 10.1016/j. bbrc.2010.08.031

[88] Jennewein C, von Knethen A, Schmid T, Brüne B. MicroRNA-27b contributes to lipopolysaccharidemediated peroxisome proliferatoractivated receptor gamma (PPARgamma) mRNA destabilization. The Journal of Biological Chemistry. 2010;**285**:11846-11853. DOI: 10.1074/jbc. M109.066399

[89] Gross JC, Chaudhary V, Bartscherer K, Boutros M. Active Wnt proteins are secreted on exosomes. Nature Cell Biology. 2012;**14**:1036-1045. DOI: 10.1038/ncb2574

[90] Xu D, Tahara H. The role of exosomes and microRNAs in senescence and aging. Advanced Drug Delivery Reviews. 2013;**65**:368-375. DOI: 10.1016/j.addr.2012.07.010

[91] Skogberg G, Telemo E, Ekwall O. Exosomes in the thymus: Antigen transfer and vesicles. Frontiers in Immunology. 2015;**6**:366. DOI: 10.3389/ fimmu.2015.00366

[92] Turiák L, Misják P, Szabó TG, Aradi B, Pálóczi K, Ozohanics O, et al. Proteomic characterization of thymocyte-derived microvesicles and apoptotic bodies in BALB/c mice. Journal of Proteomics. 2011;74:2025-2033. DOI: 10.1016/j.jprot.2011.05.023

[93] Skogberg G, Gudmundsdottir J, van der Post S, Sandström K, Bruhn S, Benson M, et al. Characterization of human thymic exosomes. PLoS One. 2013;8:e67554. DOI: 10.1371/journal. pone.0067554

[94] Lundberg V, Berglund M, Skogberg G, Lindgren S, Lundqvist C, Gudmundsdottir J, et al. Thymic exosomes promote the final maturation of thymocytes. Scientific Reports. 2016;**6**:36479. DOI: 10.1038/srep36479

[95] Banfai K, Garai K, Ernszt D, Pongracz JE, Kvell K. Transgenic exosomes for thymus regeneration. Frontiers in Immunology. 2019;**10**:862. DOI: 10.3389/fimmu.2019.00862

[96] Available from: https://population. io/