We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

186,000

200M

Download

154
Countries delivered to

Our authors are among the

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Chapter

Plant Growth Hormones

Amira Shawky Soliman

Abstract

Many factors can cause and affect cell growth in the plant such as external (environmental) and internal factors; one of the most important internal factors is plant growth hormones. Many hormones required for cell growth, such as auxins, gibberellins, brassinosteroids, ethylene, jasmonates, salicylic acid, strigolactones and cytokinins which able to accelerate or promote growth, but, some hormone-like abscisic acid has an adverse effect on growth which increases seed dormancy by inhibiting cell growth. Also, plant hormones are able to breakdowns dormancy for many plants and can alleviate abiotic stress (salinity, extreme temperatures and, drought,...) which led to enhance germination and improve growth for many plants, whether naturally occurring in the plant or by adding it to the plant in its artificially formed or in the form of bio- or nano-fertilization in order to increase the productivity and improve its efficiency under extreme conditions. Therefore, this chapter will highlight and will provide data for the positive or/and negative effect of these hormones on many plants to achieve a rapid germination method. It will also shed light on the relationship of these hormones to some enzymes to accelerate growth.

Keywords: plant hormones, seed germination, dormancy, cell growth, inhibition

1. Introduction

Plant hormones (phytohormones) are not nutrients, but chemicals and not all plant cells respond to hormones, but those cells that do are programmed to respond at specific points in their growth cycle. The greatest effects occur at specific stages during the cell's life, with diminished effects occurring before or after this period [1].

Plants need hormones at very specific times during plant growth and at specific locations. They also need to disengage the effects that hormones have when they are no longer needed. The production of hormones occurs very often at sites of active growth within the meristems, before the cells have fully differentiated. After production, they have sometimes moved to other parts of the plant, where they cause an immediate effect; or they can be stored in cells to be released later. Plants can also break down hormones chemically, effectively destroying them. Plant hormones frequently regulate the concentrations of other plant hormones [2, 3].

2. Importance of plant hormones

The small amounts of plant hormones promote, control, influence and develop the growth from embryo to reproductive development, also, stress tolerance and pathogen defense. According to the importance of plant hormones in this chapter will be divided into two main points: first: the effect of plant hormones on germination and growth of plants under internal or external suitable conditions, second: the effect of plant hormones on the germination and growth of plants under internal or external unsuitable environmental conditions.

2.1 The effect of plant hormones on germination and growth of plants under internal or external suitable conditions

Seed germination is attracted to the effective growth of the embryo when appropriate environmental conditions are present, leading to seed rupture and the appearance of a small plant. There are five basic steps to germination: water imbibition, enzyme activation, initiation of embryo growth, rupture of the seed coat and emergence of seedling, seedling establishment [4, 5].

In the second step stage of germination (enzyme activation), after the absorption of water through the natural openings in the casing of the seed and spread through the tissues of the seed, gibberellins which activate the formation of the hydrolytic enzymes, mainly α - amylase in the aleurone cells, which are responsible for hydrolysis of storage macro-molecules such as starch and proteins and convert them into available forms to the embryo, usage to increase in size, and raise the osmotic content of the seed, to increase water potential [6, 7].

In addition, plant hormones have an important role in plant growth not only germination such as cytokinins (CKs) which influence cell division, the formation of shoot and helping in delay tissues senescence [8, 9]. Also, the ratios of Cytokinins and auxins affect most major growth periods during a plant's lifetime [10]. Also, Peptide hormones, control of cell division, expansion, and play crucial roles in plant growth and development [11]. Furthermore, gibberellins (Gas) strongly promote cell elongation in seedlings [12, 13]. It can also affect cell cycling in plant [14]. Meanwhile, the responses of nitric oxide (NO) are in germination, cell death [15] and regulate plant cell organelle functions (e.g. mitochondria and ATP synthesis in chloroplasts) [16].

For enhances and increases plant hormones production in the plants, many studies have proved the need to add plant hormones either directly (GA3, kinetin and cytokinins) [17, 18] or indirectly (humic substances, manures, magnetite, natural zeolites, *Moringa* extract and bio-fertilization) to increase or accelerate the productivity of plant hormones in the plant [19] indicated that, the presence of organic matter represented in compost which a source of hormones like substances as auxin-like activity and gibberellin-like activity. Similar results were obtained from [20]. Ref. [21] concluded that it is also possible that the production of plant hormones influences symbiotic bacteria, such as nodule N₂ fixing bacteria. During the establishment of the soybean (*Glycine max* L.) and *Bradyrhizobium japonicum* N₂-fixing symbiosis, the production of plant hormones can determine the bacterial population in the nodules by, for example affecting the available substrate for the use of rhizobium. The other significant and interesting view of the effects of soil bacteria on the production of plant hormones is the alteration they may reason in plant signaling pathways, resulting in the output of plant hormones from the host plant [22, 23]. Ref. [24] concluded that the magnetic treatments have the same affected for phytohormone production. Ref. [25] reported that the highest mean values of IAA, GA, and CK (12.70, 13.71, 11.06 μg/g FW), respectively were recorded with compost and zeolite mixture in comparison with control. Ref. [26] concluded that the addition of a mixture of organic fertilizers and soil amendments led to significant increment in indigenous hormones characterized in **i**ndole **a**cetic acid (IAA), gibberellic acid (GA3), and cytokinins (CK), which led to a significant increase in morphological growth, floral characteristics and chemical composition of Oenothera biennis. In contrast [27] found that the application of HA inhibits

indoleacetic acid (IAA) oxidase, thereby hindering the destruction of this plant growth hormone.

2.2 The effect of plant hormones on the germination and growth of plants under internal or external unsuitable environmental conditions

Sometimes even under favorable germination conditions (an adequate water supply, a suitable temperature and the normal composition of the atmosphere) seeds do not germinate. In this case, seeds are considered dormant. Seed dormancy is defined as an inactive phase in which the growth and development are deferred and the respiration is greatly reduced [28, 29]. Seed coat dormancy involves the mechanical restriction of the seed coat. GA releases this dormancy by increasing the embryo growth potential, and/or weakening the seed coat so the radical of the seedling can break through the seed coat. ABA affects the testa or seed coat growth characteristics, including thickness, and affects the GA-mediated embryo growth potential [5].

Hormones also can mediate endosperm dormancy: Endosperm in most seeds is composed of living tissue that can actively respond to hormones generated by the embryo. The endosperm often acts as a barrier to seed germination, playing a part in seed coat dormancy or in the germination process. Living cells respond to and also affect the ABA:GA ratio, and mediate cellular sensitivity; GA thus increases the embryo growth potential and can promote endosperm weakening. GA also affects both ABA-independent and ABA-inhibiting process within the endosperm [30]. In addition, [33] concluded that the prevented germination of some seeds of tomato [31], iris [32], and some varieties of cabbage was due to the present of inhibitors (ABA, parasorbic acid, and coumarin) which cases distributed in plants and to possess the property of inhibiting seed germination and other growth phenomena [5, 34].

Plant hormones affect seed germination and dormancy by acting on different parts of the seed such as [35] found that the inhibitors in seeds of peach were at least one of the factors controlling in germination by preventing or retarding cell division of the radical. In *Lupinus angustifolius*, the contents of auxins increased through the 5th day of germination and started to decrease on the 7th day. Oppositely, gibberellins contents were decreased first then increased later, so it was clear that there was inversely related between auxins and gibberellins [36]. The germination percentage and germination rate of four studied Acacias (*A. saligna*, *A. sophorae*, *A. cyclopis*, and *A. melanoxylon*) were correlated positively with endogenous promoting and negative with endogenous inhibiting substances in their cotyledons plus embryo [37].

The promotion of germination by gibberellin and cytokinins has been demonstrated in many seed species [38, 39]. Ref. [40] treated the seeds of *Acacia longifolia*, with GA3 at 100 and 200 ppm and found that the higher GA3 concentration (200 ppm) was more effective in increasing germination while the concentration of 500 ppm was the best in the case of *Acacia catechu* [41].

Ref. [42] found that fresh seed of *Acacia nilotica* and *Acacia albida* were fully germinated when soaked in a solution of GA₃ at 200 ppm for 12 h. While soaking seeds of *Acacia nilotica* in gibberellic acid (100 or 300 ppm for 16 h) was the best [43]. Ref. [44] studied the effect of GA3 at a concentration of (50 ppm) on 16 species (four Acacia species), and found a high germination percentage for all species.

The effect of gibberellic acid and cytokinins were also recognized on the germination of other plant species seeds. Ref. [45] studied the effect of Kinetin at different concentrations on the seed germination of *Acer tataricum*, and found the highest germination percentage at the concentration of 500 ppm. Ref. [46] found the best germination percentage on soaking the seeds of *Trifolium pratense* in 50 ppm 6-benzylaminopurine (6-BAP). Ref. [47] studied the effect of Kinetin at different conc. (10, 25, 50 and 100 ppm) in the seeds of *Cassia sophera*, and found the highest

germination percentage at 100 ppm. The treatment of freshly harvested and 1 year old seeds of soybean (*Glycine max*) with, 1 ppm 6-BAP increased the germination percentage from 50 to 85% in freshly harvested seeds and to 75% in the older seeds [48]. The effect of kinetin and 6-BAP on the seed germination of *Vicia faba* were studied, [49] found an increase in its germination percentage at the concentration of 100 ppm kinetin. While [50] found that, the highest germination percentage for faba bean (*Vicia faba* L.) was achieved at the concentration of 100 ppm 6-BAP. Also [37, 51] reported that the storage has an adverse effect on the hormone within the seeds of *Acacia saligna*, *Acacia Cyclopes*, *Acacia nilotica* and *Acacia albida*, which contained the lowest value of GA3, IAA and the highest content of phenols.

Plant hormones can also alleviate abiotic stress such as drought, extreme temperatures, and salinity [52, 53]. The action of these hormones in response to situations of stress can be developed through synergistic or antagonistic activities [54]. Also, [55] concluded that the plant growth regulators like ABA, JA, and ethylene are involved in the regulation of the plant response to abiotic stress. Cytokinins are also able to enhance seed germination by the alleviation of stresses such as salinity, drought, heavy metals and oxidative stress [56–59]. Ref. [60] found that GA₃ plays an important role in the growth and metabolism of microalgae *Chlorella vulgaris* exposed to heavy metal stress and its adaptation ability to a low-level polluted aquatic environment. Meanwhile, gibberellin leads to enhancement for *Zea mays* seedling growth and establishment under saline soil conditions by improving nutrient levels and membrane permeability [61]. Also, hormonal interactions between plant and rhizosphere bacteria can affect plant tolerance to stress. As such, the plant and bacteria can be genetically modified so that they can perform more optimally under a range of conditions, including stress [62].

The decreased cytokinin and gibberellic acid (GA3) and increased abscisic acid contents are often observed responding in plants subjected to environmental stresses [63, 64]. Exogenous application of plant growth regulators [such as cytokinin or antioxidants (ascorbic acid) [65], Moringa (Moringa oleifera) leaves extract [66, 67], humic acid (HA) [68], or seaweed extract (SE) [69] could be an alternative strategy to ameliorate, minimizing or alleviating the adverse effects of abiotic stress factors on plant growth which led to promoting plant growth and development metabolism in plants. Several studies also indicated that results on wheat [70]; and on spinach [71]. Ref. [72] reported that the foliar application of Moringa (Moringa oleifera) leaves extract MLE is proved to be the most effective PGR in reducing plant (Lagerstroemia indica L. seedlings) exposure to salinity stress.

Also, bio-fertilization has beneficial microorganisms that increasing plant hormones, which led to enhances yield, plant growth and nutrient uptake under various environmental conditions such as salinity [73–76], drought and low fertility supply [77–79], especially that some endomycorrhizal fungi (Arbuscular mycorrhizal fungi) have been proven to improve drought stress; they colonize bio-trophically the root cortex and develop an extra-metrical mycelium that helps the plants to acquire mineral nutrients from the soil particularly those, which are immobile. They can under drought conditions stimulate growth-regulating substances, increase photosynthesis, improve osmotic adjustment, optimize hormonal balance and enhance water uptake [80].

Numerous studies have found also, that it can be alleviation of salt stress on peanut [81]; on pumpkin plants [82]; on *Moringa peregrina* plants [83] by using foliar application of nano-fertilizers. Also, [84] reported that nano Zn-Fe oxide plays a significant role importance in alleviating salt stress, oxidative damages on plant cells by activation of certain antioxidant enzymes. In addition, [85] reported that the application of nano-oxide and bio-fertilizer reduced the negative effects of salinity due to its contributed to produce hormones.

3. Conclusions

This chapter was indicated by many studies that the plant hormones, including IAA, cytokinins, ethylene, gibberellins, and brassinosteroids, can positively affect seed germination and seedling growth, for many plants as mentioned previously in the chapter, under favorable conditions. While ABA has an adverse on affect seed germination and the growth.

Also, this chapter sheds the light on the important role of soil bacteria in the production of plant hormones or as an alternative in the case of the low rate of plant hormones in the plant, which led to hence seed germination, growth, and hence crop production.

In addition, this chapter provided many studies that prove that the plant hormones very important to overcome dormancy or growth under stress condition. Also, shed the lights on the importance of the exogenous application of plant growth regulators (cytokinin or antioxidants, Moringa leaves extract, humic acid, or seaweed extract, bio- or nano fertilizers) for enhancing the productivity of plant hormones which led to increased cell growth.

Finally, it can be stated that the plant hormones are essential for cell growth, whether under normal conditions or under stress conditions.

Conflict of interest

The author declares that she does not have any conflict of interest.



Amira Shawky Soliman Natural Resources Department, Faculty of African Postgraduate Studies, Cairo University, Giza, Egypt

*Address all correspondence to: sitamira2000@yahoo.com

IntechOpen

© 2019 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. CC BY

References

- [1] Swarup R, Perry P, Hagenbeek D, Van Der Straeten D, Beemster GT, Sandberg G, et al. Ethylene upregulates auxin biosynthesis in Arabidopsis seedlings to enhance inhibition of root cell elongation. The Plant Cell. 2007;19(7):2186-2196. DOI: 10.1105/tpc.107.052100
- [2] Campbell N, Reec JB. Biology. 6th ed. San Francisco: Benjamin Cummings; 2002. ISBN: 978-0805366242
- [3] Davies PJ. The plant hormones: Their nature, occurrence, and functions. Plant Hormones: Physiology, Biochemistry and Molecular Biology. 2010:1-15. DOI: 10.1007/978-1-4020-2686-7_1
- [4] Copeland LO. Principles of Seed Science and Technology. USA: Burgess Publishing Company; 1976. pp. 55-58
- [5] Meyer BS, Anderson DB. Plant Physiology. 2nd ed. Canada: D. Van Nostrand Company, Ltd; 1952. pp. 709-715
- [6] Ikuma I, Thimann KV. Metabolic control of germination. In: The Physiology and Biochemistry of Seed Dormancy and Germination. Amsterdam, New York, Oxford: North Holland Publ. Co.; 1963
- [7] Öpik H, Rolfe SA, Willis AJ, Street HE. The Physiology of Flowering Plants. 4th ed. Cambridge University Press; 2005. p. 191. ISBN 978-0-521-66251-2
- [8] To JP, Kieber JJ. Cytokinin signaling: Two-components and more. Trends in Plant Science. 2008;**13**:85-92
- [9] Santner A, Calderon-Villalobos L, Estelle M. Plant hormones are versatile chemical regulators of plant growth. Nature Chemical Biology. 2009;5:301-307
- [10] Sipes DL, Einset JW. Cytokinin stimulation of abscission in lemon

- pistil explants. Journal of Plant Growth Regulation. 1983;**2**(1-3):73-80. DOI: 10.1007/BF02042235
- [11] Lindsey K, Casson S, Chilley P. Peptides: New signalling molecules in plants. Trends in Plant Science. 2002;7(2):78-83. DOI: 10.1016/S0960-9822(01)00435-3
- [12] Tsai FY, Lin CC, Kao CH. A comparative study of the effects of abscisic acid and methyl jasmonate on seedling growth of rice. Plant Growth Regulation. 1997;21(1):37-42. DOI: 10.1023/A:1005761804191
- [13] Van der Knaap E, Kim JH, Kende H. A novel gibberellins induced gene from rice and its potential regulatory role in stem growth. Plant Physiology. 2000;122:695-704
- [14] Hauvermale AL, Ariizumi T, Steber C. Gibberellin signaling: A theme and variations on DELLA repression. Plant Physiology. 2012;**160**:83-92
- [15] Shapiro AD. Nitric oxide signaling in plants. Vitamins and Hormones. 2005;**72**:339-398. DOI: 10.1016/S0083-6729(05)72010-0
- [16] Roszer T. Nitric oxide synthesis in the chloroplast. In: Roszer T, editor. The Biology of Subcellular Nitric Oxide. New York, London, Heidelberg, Springer; 2012. ISBN 978-94-007-2818-9
- [17] Afroz A, Chaudhry Z, Khan R, Rashid H, Khan SA. Effect of GA3 on regeneration response of three tomato cultivars (*Lycopersicon esculentum*). Pakistan Journal of Botany. 2009;**41**(1):143-151
- [18] Hyunggook K, Donggeun C, Inkyu K. Effect of growth regulator treatments on quality and growth in 'Gailiangmeru' grape (*Vitis* spp.). Acta Horticulturae. 2008;772:319-322

- [19] Marek K, Skorupska A. Production of B-group vitamins by plant growth promoting *Pseudomonas fluorescens* strain 267 and importance of vitamins in the colonization and nodulation of red clover. Biology and Fertility of Soils. 2001;33:146-151
- [20] Khalil FA, EL-Aref KAO. Biofertilizers as partner with mineral N-fertilizer for fertilizing wheat crop cultivar Sids 6. Journal of Agricultural Science, Mansoura University. 2001;**26**:8287-8295
- [21] Ikeda S, Okubo T, Anda M, Nakashita H, Yasuda M, Sato S, et al. Community- and genome-based views of plant-associated bacteria: Plant-bacterial interactions in soybean and rice. Plant & Cell Physiology. 2010;51:1398-1410
- [22] Cui H, Wang Y, Xue L, Chu J, Yan C, Fu J, et al. *Pseudomonas syringae* effector protein AvrB perturbs *Arabidopsis* hormone signaling by activating MAP kinase. Cell Host & Microbe. 2010;7:164-175
- [23] Miransari M, Abrishamchi A, Khoshbakht K, Niknam V. Plant hormones as signals in arbuscular mycorrhizal symbiosis. Critical Reviews in Biotechnology. 2014;34(2):123-133. DOI: 10.3109/07388551
- [24] Maheshwari LB. Magnetic treatment of irrigation water: Evaluation of its effects on vegetable crop yield and water productivity [thesis]. New South Wales: Western Sydney University; 2009
- [25] Soliman AS, Abdelwahab MM. Response of *Adansonia digitata* to compost and zeolite in replacement of chemical fertilization. American-Eurasian Journal Of Agricultural & Environmental Sciences. 2013;**13**(2):198-206
- [26] Abdelwahab MM, Soliman AS. Comparative study on the influence of

- organic fertilizer and soil amendments on evening primrose (*Oenothera biennis* L.). International Journal of Agricultural Research. 2017;**12**:52-63
- [27] Mato MC, Olmedo MG, Mendez J. Inhibition of indoleacetic acidoxidase by soil humic acids fractionated on sephadex. Soil Biology and Biochemistry. 1972;4:469-473
- [28] Bewley and Black. Seeds Physiology of Development and Germination. 2nd ed. New York, USA: Plenum Press; 1994
- [29] Crawley MJ. Plant Ecology. 2nd ed. UK: Blackwell Science Ltd; 1997
- [30] Gerhard W. Seed Dormancy. In: The Seed Biology Place. Royal Holloway University of London; 2000
- [31] Evenari M. Germination inhibitors. The Botanical Review. 1949;**15**:153-194
- [32] Randolph LF, Cox LG. Factors influencing the germination of Iris seed and the relation of inhibiting substances to embryo dormancy. Proceedings of the American Society for Horticultural Science. 1943;43:284
- [33] Cox Herald R, Clancy CF, Wolfe DM, Vanderscheer J. Report of Preliminary Studies on Tsutsugamushi (*Scrub typhus*). submitted to Natl. Research Council; 1945
- [34] Veldstra H, Havinga E. Enzymologia. Amsterdam. 1945;**11**:373
- [35] El-Khodary SES, Moustafa SM. Effect of a potent germination inhibitor of dormant Peach seeds on division of root cells. Proc. Soudi Bio. Soci. 1979;3:149-166
- [36] Denisova GM, Dmitrieve TA, Viktorova VE. On the kinetics of endogenous growth substances in organs of seedlings of *Lupinus angustifolius*. In: Fiziologiya Rostovykh

- Protsessov. Moscow: USSR; 1980. pp. 41-48
- [37] Youssef EMA, Badawy ESM, H eikal EA, Sakr SS. Studies on the germination of different *Acacia* species. Bulletin of Faculty of Pharmacy, Cairo University. 1991;42(3):849-868
- [38] Mayer AM, Poljakoff-Mayber A. The Germination of Seeds. 3rd ed. Oxford, UK: Pergamon Press; 1982
- [39] Briggs DE. In: Milbrorrow BV, editor. Biosynthesis and Its Control in Plants. New York: Academics Press; 1973. p. 219
- [40] AL-Kinany P. Effect of some pre-treatment on seed germination and subsequent development at *Acacia longifolia* seedling. Pakistan Journal of Forestry. 1981;**31**(3):81-88
- [41] Singh SS, Sharma BN, Paliwal GS. Effect of Niagara and GA3 on seed germination, early seedling growth, and cotyledonary expansion of *Acacia catechu* Willd. Indian Journal of Forestry. 1985;8(1):41-46
- [42] Krishnasamy V, Palaniappan M. Studies on seed dormancy in brinjal var. 'Annamalai'. South Indian horticulture. 1994;38(1):42-44
- [43] Patel VS, Kukadia MU, Vashi BG, Jadeja DB. The improvement of germination and seedling growth of *Acacia nilotica* (Linn.) Wild. by scarification methods. Gujarat Agricultural Universities Research Journal. 1999;**21**(2):149-151
- [44] Bell DT, King LA, Plummer JA. Ecophysiological effect of light quality and nitrate on seed germination in species from Western Australia. Australian Journal of Ecology. 2003;24(1):2-10
- [45] Pozdova LM. Changes of cytokinin activity during breaking dormancy

- in seeds. Soviet Plant Physiology. 1985;**32**(2):368-374
- [46] Kovalev VM, Shipova EV. Increasing the adaptive capacity of plants using physiologically active substances. In: Vestinik Sel'skohozyaistvennoi Nauki. Vol. 2. Moscow: USSR; 1987. pp. 74-78
- [47] Chhavi T, Thakur C. Influence of different growth regulators on seed germination and seedling growth of *Cassia sophera* L. Acta Botanica Indica. 1989;**17**(2):187-191
- [48] Fujiki T, Naito T, Akihama T. Promotion of germination and reduction of decrease of t RNA content in the embryo of aged seeds of soybean (*Glycine max* M.) treated with 6-benzylaminopurine. Japanese Journal of Breeding. 1989;**39**(1):1-7
- [49] Gupta M. Effect of growth regulators on foliar stomata of *Vicia faba* L. Advances in Plant Sciences. 1996;5(2):483-495
- [50] Khan AA, Ansari M, Talat K, Neelu K, Khursheed T, Kalra N. Studies on the effect of 6-benzyl aminopurine on meiosis in feba bean (*Vicia faba* L.). Advances in Plant Sciences. 2000;**11**(2):121-125
- [51] Soliman AS. Studies on ssed germination and seedling growth of some African Acacias [thesis]. Institute of African Research and Studies, Cairo Univ.; 2004. pp. 6-11
- [52] Bartsch M, Bednarek P, Vivancos PD, Schneider B, von Roepenack-Lahaye E, Foyer CH, et al. Accumulation of isochorismatederived 2,3-dihydroxybenzoic 3-O-β-D-xyloside in Arabidopsis resistance to pathogens and aging of leaves. The Journal of Biological Chemistry. 2010;**285**(33):25654-25665
- [53] Lumba S, Cutler S, McCourt P. Plant nuclear hormone receptors:

- A role for small molecules in proteinprotein interactions. Annual Review of Cell and Developmental Biology. 2010;**26**:445-469
- [54] Fujita M, Fujita Y, Noutoshi Y, Takahashi F, Narusaka Y, Yamaguchi-Shinozaki K, et al. Crosstalk between abiotic and biotic stress responses: A current view from the points of convergence in the stress signaling networks. Current Opinion in Plant Biology. 2006;**9**(4):436-442
- [55] Awan K, Khurshid MY, Mehmood A. Plant growth regulators and their role in abiotic stress management Fahad. The International Journal of Innovative Biosciences Research. 2017;1(1):9-22
- [56] Khan MA, Ungar IA. Alleviation of seed dormancy in the desert forb *Zygophyllum simplex* L. from Pakistan. Annals of Botany. 1997;**80**:395-400
- [57] Atici O, Agar G, Battal P. Changes in phytohormone contents in chickpea seeds germinating under lead or zink stress. Biologia Plantarum. 2005;49:215-222
- [58] Nikolic R, Mitic N, Miletic R, Neskovic M. Effects of cytokinins on in vitro seed germination and early seedling morphogenesis in *Lotus corniculatus* L. Journal of Plant Growth Regulation. 2006;25:187-194
- [59] Peleg Z, Blumwald E. Hormone balance and abiotic stress tolerance in crop plants. Current Opinion in Plant Biology. 2011;14:290-295
- [60] Falkowska M, Pietryczuk A, Piotrowska A, Bajguz A, Grygoruk A, Czerpak R. The effect of gibberellic acid (GA3) on growth, metal biosorption and metabolism of the green algae chlorella vulgaris (Chlorophyceae) Beijerinck exposed to cadmium and lead stress. Polish Journal of Environmental Studies. 2011;20(1):53-59

- [61] Tuna AL, Kaya C, Dikilitas M, Higgs D. The combined effects of gibberellic acid and salinity on some antioxidant enzyme activities, plant growth parameters and nutritional status in maize plants. Environmental and Experimental Botany. 2008;**62**(1):1-9
- [62] Ghanem M, Hichri I, Smigocki A, Albacete A, Fauconnier M, Diatloff E, et al. Root-targeted biotechnology to mediate hormonal signaling and improve crop stress tolerance. Plant Cell Reports. 2011;30:807-823
- [63] Atanasova L, Pissarska MG, Popov GS, Georgiev GI. Growth and endogenous cytokinins of juniper shoots as affected by high metal concentrations. Biologia Plantarum. 2004;48:157
- [64] Bajguz A. Brassinosteroid enhanced the level of abscisic acid in Chlorella vulgaris subjected to short-term heat stress. Journal of Plant Physiology. 2009;**166**:882
- [65] Sadak MS, Dawood MG. Role of ascorbic acid and α tocopherol in alleviating salinity stress on flax plant (*Linum usitatissimum* L.). Journal of Stress Physiology & Biochemistry. 2014;**10**(1):93-111
- [66] Rajanandh MG, Satishkumar MN, Elango K, Suresh B. *Moringa oleifera* Lam. a herbal medicine for hyperlipidemia: A preclinical report. Asian Pacific Journal of Tropical Disease. 2012;**2**:S790-S795
- [67] Moussa HR, Hassan MA-E. Growth enhancers to mitigate salinity stress in *Vicia faba*. International Journal of Vegetable Science. 2016;**22**(3):243-250
- [68] Rady MM, Abd El-Mageed TA, Abdurrahman HA, Mahdi AH. Humic acid application improves field performance of cotton (*Gossypium barbadense* L.) under saline conditions. Journal of Animal and Plant Sciences. 2016;**26**(2):487-493

- [69] Battacharya D, Babgohari MZ, Rathor P, Prithiviraj B. Seaweed extracts as biostimulants in horticulture. Scientia Horticulturae. 2015;**196**:39-48
- [70] Azra Y, Basra SMA, Muhammad F, Hafeez ur R, Nazim H, Habib RA. Exogenous application of moringa leaf extract modulates the antioxidant enzyme system to improve wheat performance under saline conditions. Plant Growth Regulation. 2013;69:225-233
- [71] Aslam M, Sultana B, Anwar F, Munir H. Foliar spray of selected plant growth regulators affected the biochemical and antioxidant attributes of spinach in a field experiment. Turkish Journal of Agriculture and Forestry. 2015;39:1-10
- [72] Soliman AS, Shanan NT. The role of natural exogenous foliar applications in alleviating salinity stress in *Lagerstroemia indica* L. seedlings. Journal of Applied Horticulture. 2017;**19**(1):35-45
- [73] Kang SM, Khan A, Waqas M, You YH, Kim JH, Kim JG, et al. Plant growth-promoting rhizobacteria reduce adverse effects of salinity and osmotic stress by regulating phytohormones and antioxidants in *Cucumis sativus*. Journal of Plant Interactions. 2014;**9**(1):673-682
- [74] Hussein MMM, Haggag AA. Effect of irrigation intervals and salt concentrations on the growth and chemical composition of *Asclepias curassavica* L. Annals of Agricultural Sciences. 2003;48(1):307-327
- [75] Gupta G, Parihar SS, Ahirwar NK, Snehi SK, Singh V. Plant growth promoting rhizobacteria (PGPR): Current and future prospects for development of sustainable agriculture. Journal of Microbial and Biochemical Technology. 2015;7:096-102
- [76] Soliman AS, Sakr WRA. Improving tolerance of *Vachellia farnesiana* plants

- to irrigation water salinity by using bio-inocula under sandy soil conditions. American-Eurasian Journal of Agricultural & Environmental Sciences. 2017;17(1):01-21
- [77] Soliman AS. Effect of Rhizobia isolated from some Acacias of on growth of *Acacia nilotica* under some stress conditions in North Africa [thesis]. Department of Natural Resources, Institute of African Research and Studies, Cairo University; 2008
- [78] Aroca R. Ruiz-Lozano JM Induction of plant tolerance to semiarid environments by beneficial soil microorganisms—A review. Sustainable Agriculture Reviews. 2009;2:121-135
- [79] Soliman AS, Morsy EM, Massoud ON. Tolerance of bio-fertilized *Delonix regia* seedlings to irrigation intervals. Journal of Horticulture and Forestry. 2015;**6**(3):73-83
- [80] Colla LM, Reinehr CO, Reichert C, Costa JAV. Production of biomass and nutracenutical compounds by *Spirulina platensis* under different temperature, nitrogen sources. Bioresource Technology. 2007;**98**:1489-1493
- [81] Liu XM, Zhang FD, Zhang SQ, He XS, Fang R, Feng Z, et al. Effects of nano-ferric oxide on the growth and nutrients absorption of peanut. Plant Nutrition and Fertilizer Science. 2005;11:14-18
- [82] Zhu H, Han J, Xiao JQ, Jin Y. Uptake, translocation and accumulation of manufactured iron oxide nanoparticles by pumpkin plants. Journal of Environmental Monitoring. 2008;**10**:713-717
- [83] Soliman AS, El-feky SA, Darwish E. Alleviation of salt stress on *Moringa peregrina* using foliar application of nanofertilizers. Journal of Horticulture and Forestry. 2015;7(2):36-47

Plant Growth Hormones
DOI: http://dx.doi.org/10.5772/intechopen.84350

[84] Ghaffari H, Razmjoo J. Response of durum wheat to foliar application of varied sources and rates of iron fertilizers. Journal of Agricultural Science and Technology. 2015;**17**:321-331

[85] Babaei K, Sharifi RS, Pirzad A, Khalilzadeh R. Effects of bio fertilizer and nano Zn-Fe oxide on physiological traits, antioxidant enzymes activity and yield of wheat (*Triticum aestivum* L.) under salinity stress. Journal of Plant Interactions. 2017;**12**(1):381-389

