

# We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

186,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index  
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?  
Contact [book.department@intechopen.com](mailto:book.department@intechopen.com)

Numbers displayed above are based on latest data collected.  
For more information visit [www.intechopen.com](http://www.intechopen.com)



---

# Dengue Fever: A General Perspective

---

Muhammad Kashif Zahoor, Azhar Rasul,  
Muhammad Asif Zahoor, Iqra Sarfraz,  
Muhammad Zulhussnain, Rizwan Rasool,  
Humara Naz Majeed, Farhat Jabeen and  
Kanwal Rania

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/intechopen.81277>

---

## Abstract

Dengue Fever or commonly known as Dengue, a mosquito-borne arboviral infection has emerged as havoc around the globe. Annually, about 50 million infections are reported, resulting in 22,000 deaths and almost 2.5 billion people are reported living at risk. Dengue infection is caused by Dengue Virus (DENV), which is a member of genus Flavivirus and comprised of ten proteins; three proteins, capsid (C), membrane (M), and envelope (E), play structural role and seven are identified as non-structural that direct DENV replication. Four distinct serotypes: DENV-1, DENV-2, DENV-3 and DENV-4 are transmitted via *Aedes* mosquitoes. Clinically, Dengue patients can be categorized into three groups according to WHO 2009 revised classification. Typical symptoms of dengue include: extreme fatigue; sudden fever (from 3-7 days), headache, joint, muscle, and back pain; vomiting and diarrhea, appetite loss; skin rash along minor bleeding. *Aedes aegypti* is geographically distributed in tropical areas and breeds in artificially filled water containers i.e. drums, tyres, flower vases plastic food containers, tin cans, etc. Due to four viral serotypes and non-availability of the model animal for dengue, producing vaccines is a challenging task. Thus, Dengue can be managed using various vector control strategies through physical, chemical and biological means.

**Keywords:** dengue fever, dengue hemorrhagic fever, dengue shock syndrome, *Aedes aegypti*, DEN virus

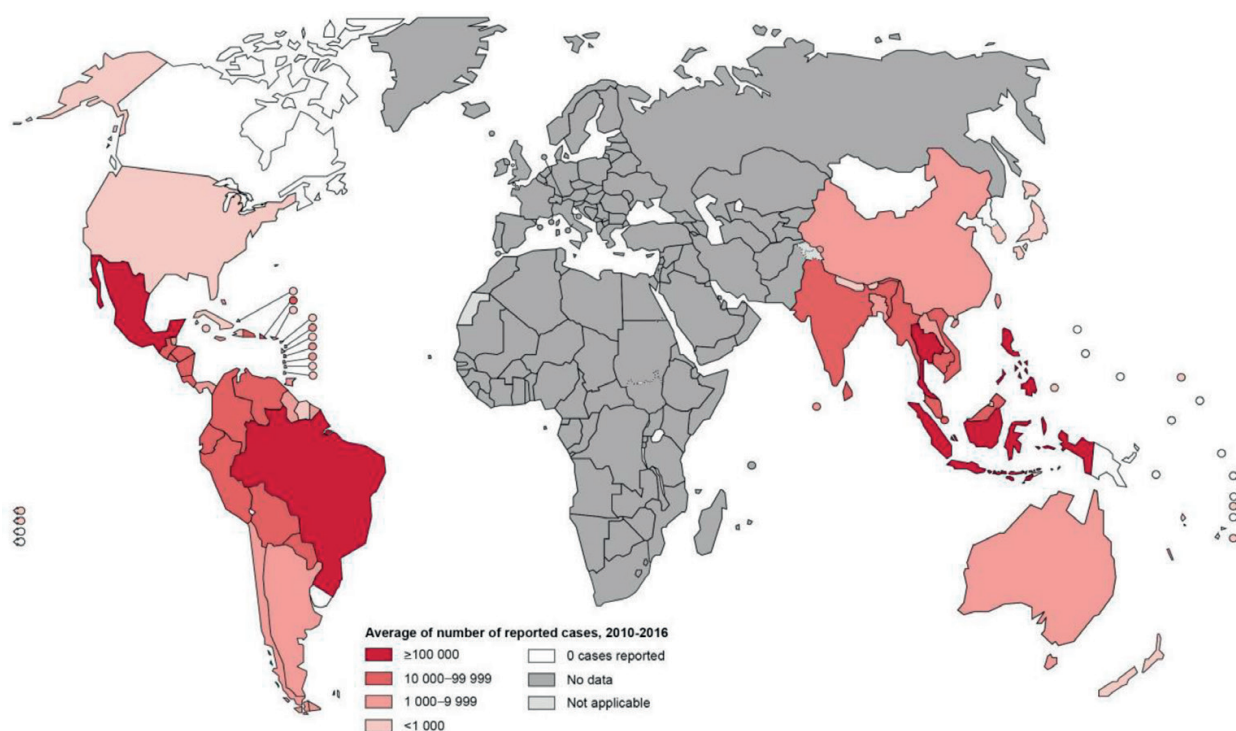
---

## 1. Historical background

The word “dengue” is known to be derived from Swahili language “ki denga pepo”, which illustrates the meaning as “sudden cramp like seizure”. The signs and indications that are suggestive of this viral disease can be tracked back to Chinese Chin Dynasty (265–420 AD) where this infection was believed to be a type of water poison and reported to be linked with insects and water [1]. Some of the historical accounts for dengue fever states that about 500–600 years ago, it appeared from Africa while the first and foremost outbreak of this deadly disease reached other parts of world in 1780s [2]. The detection and isolation of dengue virus date backed to the World War II and it was documented in Japan for the first time in 1942 [3]. Dengue-like symptoms have been reported in early Chinese manuscripts which can be traced back to 992 and to 1600s in the West Indies [4]. In another report, Benjamin Rush observed the first detailed symptoms of dengue shock syndrome (now severe dengue) in 1780 during an outbreak in Philadelphia near Delaware River [5]. Similar disease symptoms were observed in North America along Atlantic coast during eighteenth–nineteenth centuries, on the Caribbean Islands and the Mississippi basin [6]. Bancroft reported for the first time that *Aedes aegypti* mosquito is vector of dengue virus [7]. However, modern research about dengue virus was not started until 1943–1944. For the first time culturing and isolation of this virus was performed from suckling mice brain [8].

## 2. Geographic distribution

It is scientifically accepted that dengue viruses originated in monkeys and jumped to humans in Africa or Southeast Asia between 100 and 800 years ago. Dengue fever remained geographically



**Figure 1.** Distribution of dengue worldwide (taken from [www.who.int/denguecontrol/epidemiology/](http://www.who.int/denguecontrol/epidemiology/)).

restricted till 1950s. But due to the Second World War, transport of *Aedes* mosquitoes happened around the world which played a crucial role in the dissemination of the viruses. Now, approximately 2.5 billion people live in areas where there is a risk of dengue transmission [9–12].

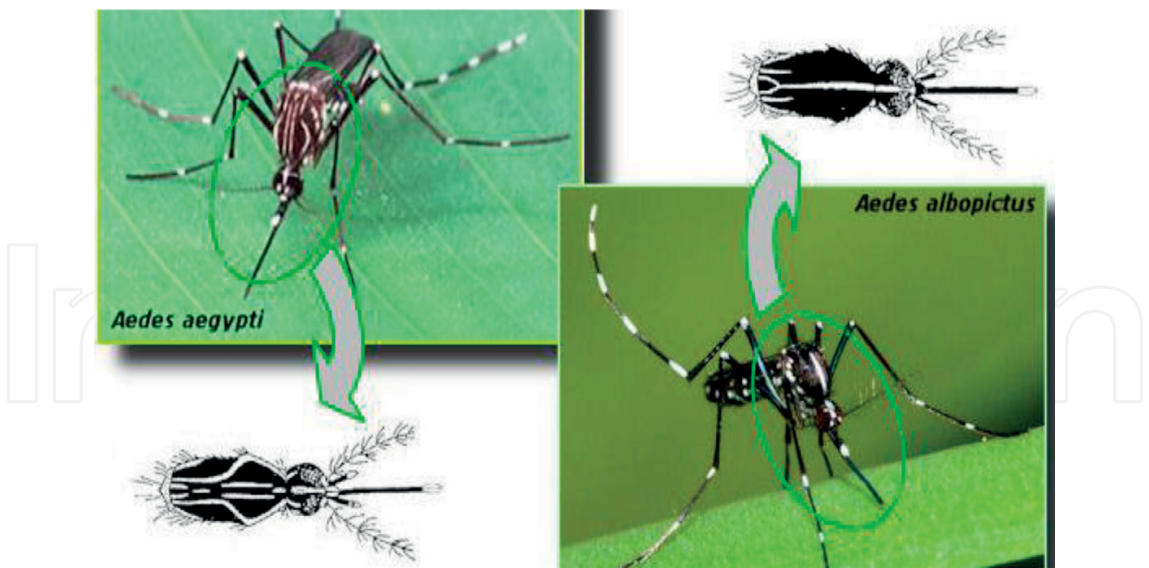
During 1850s, first case of dengue was documented in the Philippines and Thailand. Later, after 1980s large number of cases began to appear in the Caribbean and Latin America. Today, Dengue is endemic in at least 100 countries in Asia, the Pacific, the Americas, Africa, and the Caribbean. Dengue fever is reported to prevail in 26 states [13–15]. DENV-2 was the predominant serotype in dengue outbreaks that occurred before 2000 but DENV-3 was the predominant serotype between 2000 and 2009. After 2010, DENV-1 dominated global dengue outbreaks, and DENV-4 was the least frequently identified serotype [16, 17]. The geographical distribution of dengue with respect to countries has been shown in **Figure 1** which explains the current prevalence of this disease around the world [11].

### 3. The vector

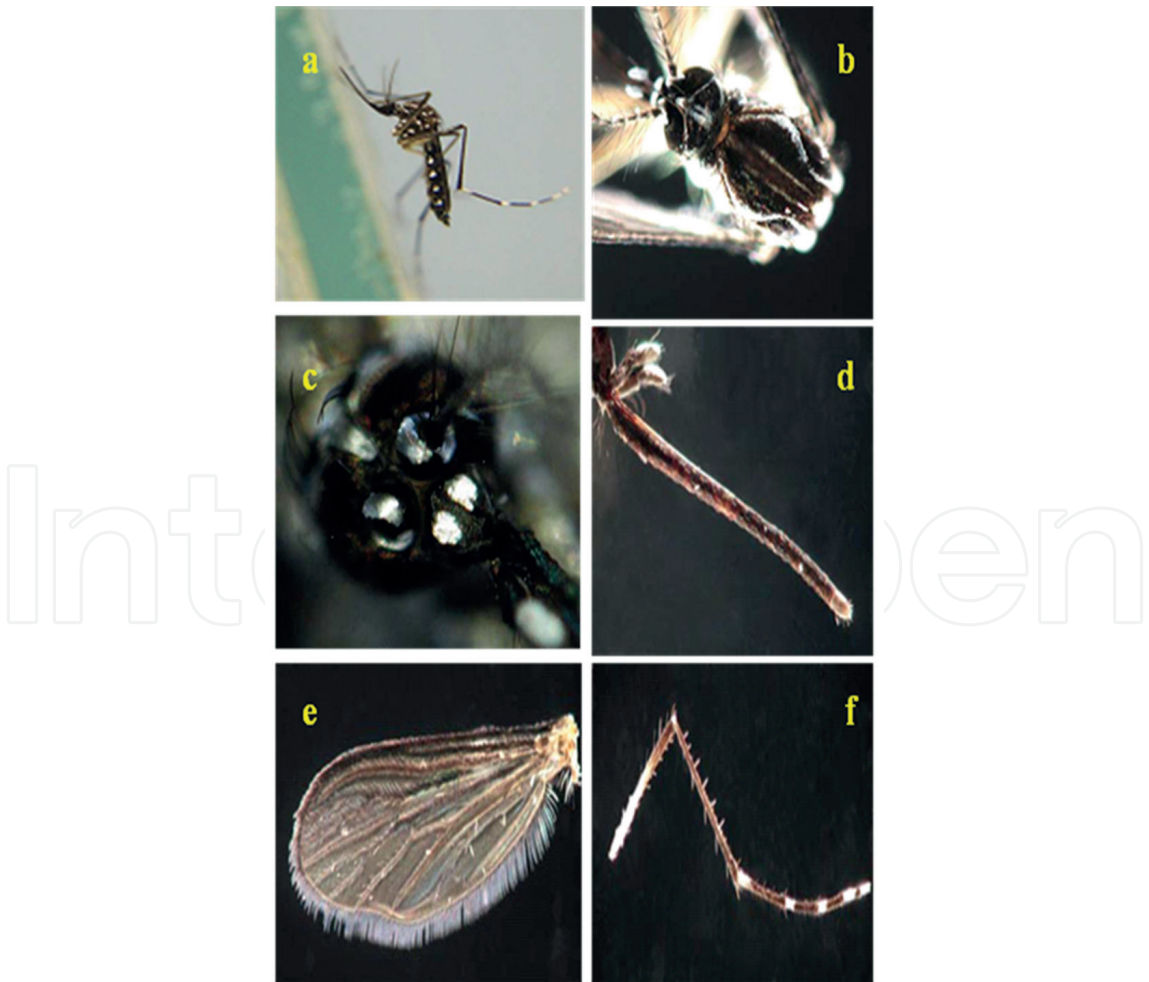
Dengue virus spreads due to infected females of genus *Aedes*, significantly from *Aedes aegypti* and *Aedes albopictus*. There has been a serious concern amid public health departments. In newly invaded countries, *Aedes albopictus* would cause severe epidemics of arbovirus diseases (it is considered as a competent vector transmitting about 22 arboviruses), especially all four serotypes of dengue; however generally it is transmitted by *Aedes aegypti*. *Aedes albopictus* persists to spread, taking the place of *Aedes aegypti* in some areas. [18] *Aedes albopictus* might serve as a maintenance vector of dengue in non-urban areas of Pacific islands and South-east Asia. *Aedes albopictus* is not considered an imperative urban dengue vector, but in a few countries where *Aedes aegypti* is not present, that is, the Seychelles, parts of China, Japan and Hawaii [18]. The biting females of *Aedes albopictus* were discovered firstly in 1999, in Southern Cameroon; it provoked survey in 2000 and then adults as well as breeding populations were identified in five major cities of the country mainly breeding in old tires imported from Nigeria and USA which were infested with the mosquitoes [19].

*Aedes* is best known vectors of dengue fever and yellow fever. Some species of *Aedes* are also vectors of viral disease and filariasis [20]. Several serotypes of the dengue virus are carried to human beings via the bites of *Aedes* mosquitoes infected with dengue virus. *Aedes aegypti* is considered one of the most crucial vector whereas *Aedes niveus*, *Aedes albopictus* and *Aedes polynesiensis* have been reported as secondary vectors in most of the regions of the world [9]. *Aedes aegypti* and *Aedes albopictus* are known as the two primary vectors for transmitting the dengue in most parts of South Asia, including India. As the distribution of this affliction is concerned in respect to geographically, it is characteristically parallel to that of the principal vector species, *Aedes aegypti* [21]. Dengue mosquito is a subtropical and tropical species having distribution throughout the world [22]. Dengue virus spreads due to infected females of genus *Aedes*, specifically through *Aedes aegypti* in urban settings and *Aedes albopictus* in sylvatic areas [18]. *Aedes albopictus* (Diptera: Culicidae), is basically endemic to Pacific and Indian Oceanic islands, and from South-east Asia, it spread to America, Europe and Africa in recent decades dormant eggs in the tires. Venereal and possibly vertical transmission of dengue virus takes place by infected female of *Aedes aegypti* to its progeny (transovarian) and also from infected male to the female during the process of copulation, respectively [23].





**Figure 2.** Difference between *Aedes aegypti* and *Aedes albopictus* (Source: <http://www.mdsauade.com/wp-content/uploads/2012/04/aedes-aegypti-e-aedes-albopictus.jpg>).



**Figure 3.** *Aedes aegypti* with its taxonomic characteristics. (a) *Aedes aegypti*; (b) Lyrix at thorax; (c) Clypeus; (d) Proboscis; (e) silvery scales on wing; (f) white stripes on leg.

The adult of yellow fever mosquito have approximately 4–7 mm size and it range from small to medium-sized mosquito. To the human eye, these mosquitoes are similar to the Asian tiger mosquito with a minor dissimilarity in thorax patterns and size. Adults of *Aedes aegypti* have white scales that form the shape of a violin or lyre, on the dorsal side of the thorax while the adults of *Aedes albopictus* is characterized by a white stripe to the middle at the top of the thorax region. Every tarsal portion of the hind legs exhibit white bands, this is what appear to be stripes. Abdomen is usually dark brown to black in color, but also exhibit white scales. Males are smaller than females, and can be discriminated by small palps tipped with white or silver scales. Males are characterized by plumose type of antennae; however, females possess sparse short hairs. Under a microscope, male mouthparts are viewed as modified structure for nectar feeding, and mouthparts of female are modified for feeding f blood. The proboscis from both sexes is darkly colored, and the segment above the proboscis which is known as clypeus has two clusters of white scales. A characteristic feature of all *Aedes* species is the pointed tip of the abdomen (**Figures 2 and 3**) [24].

#### 4. Life cycle of *Aedes aegypti*

*Aedes aegypti* is geographically distributed in tropical areas and it breeds in artificially filled water containers such as drums, tyres, flower vases such as plastic food containers, tin cans and old motor parts [4]. *Aedes aegypti* is a holometabolous type of insect, going through complete metamorphosis meaning four developmental stages from egg to adult stage. Life span of adult can range from 2 weeks to about 4 weeks but it depends on conditions of environment. A female mosquito lay eggs for about 4–5 times during her entire life span and average number of eggs in single spawn ranges from 10 to 100 eggs. *Aedes aegypti* are found in three different polytypic forms: sylvan, domestic, and peridomestic. The domestic type usually breeds in urban habitats, mostly inside or around houses. The sylvan type is rural form which breeds in tree holes, normally in forests, and the peridomestic form generally lives in environmentally-modified regions as coconut farms and groves (**Figure 4**) [24].

As the spread of mosquitoes is concerned, it occurs by active flight (adult) and passive transportation (immature stages) through international trades. Successive waves of invasion of *Aedes aegypti* and *Culex pipiens* have been aided by commercial passages also from fifteenth century to onward. *Aedes aegypti* substituted *Aedes albopictus* in Asian countries during the twentieth century [25, 26].

#### 5. The virus

Dengue infection is transmitted by dengue virus (DV) which is a member of genus *Flavivirus*. This Arbovirus group of viruses is specifically transmitted via insect vectors. Mature viral particles have diameter of 40–50 nm, spherical shape and 11 kb, having positive single stranded RNA which has a 5'-methyl cap with a single open reading frame. Genus *Flavivirus* has 4 antigenically associated but four distinct serotypes known as DENV-1, DENV-2, DENV-3 and DENV-4 [27, 28]. The serotypes are evolved from a common ancestor and all are considered

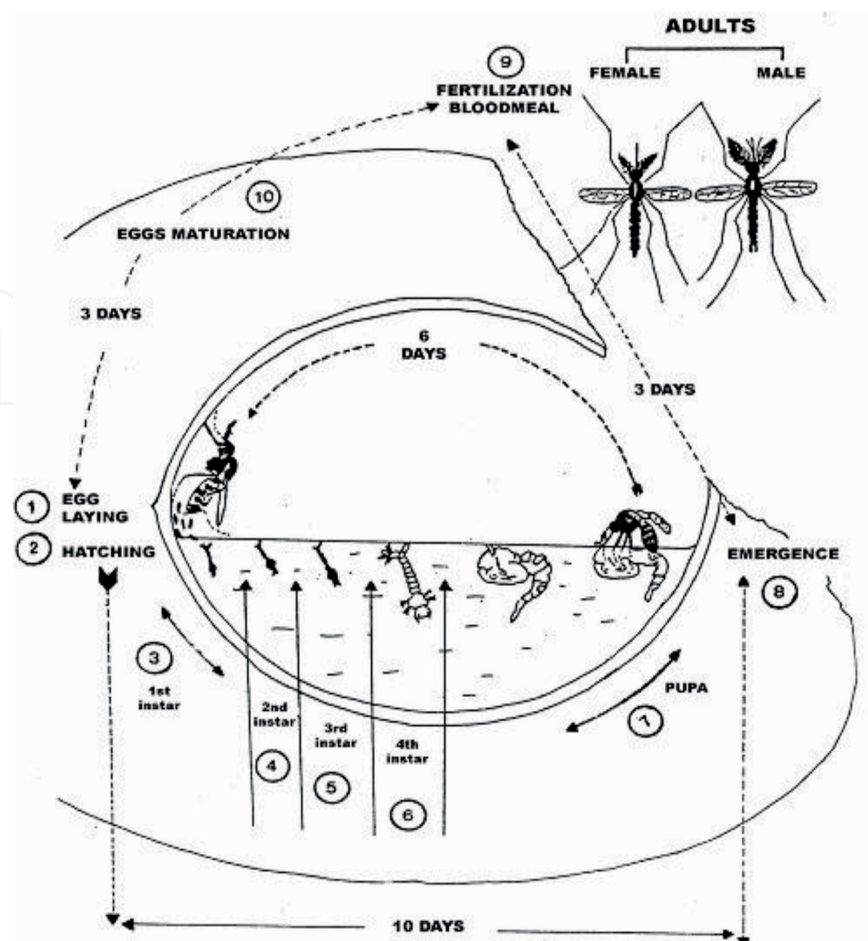


Figure 4. Lifecycle of *Aedes aegypti* ([http://www.ipnc.nc/FCKeditorFiles/Image/entomo\\_20.jpg](http://www.ipnc.nc/FCKeditorFiles/Image/entomo_20.jpg)).

causative agent of broadly analogous disease spectrum in humans [29–31]. It consists of ten proteins, out of which three proteins, capsid (C), membrane (M), and envelope (E), play structural role and seven (NS1, NS2A, NS2B, NS3, NS4A, NS4B and NS5) are identified as non-structural that direct DENV replication. Approximately 17% of these virions are lipid by weight which forms a lipid bilayer between E/M outer shell and the nucleocapsid core [32, 33].

Binding of dengue virus like most of the other viruses to its receptor is regulated by envelop protein (E). In mammalian cells, all the serotypes binds with nLc4Cer, DC-SIGN/L-SIGN, Heparan sulfate as well as Mannose receptors. Additionally DENV-2 serotype also show binding trend with HSP70/HSP90, CD14-associated protein, GRP78 and two other unknown protein receptors. On the other hand DENV 1–3 serotypes can also attach with Laminin receptor while DENV 2–4 serotypes are also found to bind with protein receptor which is unknown [34]. After binding with particular receptors through receptor-mediated endocytosis, virion fuses with acidic lysosomes. Then, the viral particle releases its RNA in the cytoplasm of host cell for the synthesis of viral proteins. After the synthesis of all the required proteins, viral RNA starts generating a minus strand, and then transcription of new plus stranded molecule occurs. Hundreds of copies of viral particles are generated from a single virus particle leading towards cellular damage and even death. RNA-dependent RNA polymerases (RdRps) encoded by the virus itself and other cellular factors catalyze the infection cycle of this virus

[35]. However the exact mechanism of vascular permeability and hemorrhagic fever is not clear. Studies are being oriented to understand these mechanisms specially focusing on the role played by T-cell immune response. High levels of interferon alpha were reported during secondary infection after 1–2 days of fever onset [36] while high concentration of soluble interleukin 2 receptor, interferon  $\gamma$ , soluble CD8 and soluble CD4 interleukin 2 were also described by researchers during the outset of vascular permeability [36, 37].

## 6. Clinical aspects

The most commonly occurring DENV infection transits through an asymptomatic or mildly symptomatic course [38]. Symptomatic dengue fever is usually accompanied by headache, malaise, retroorbital pain, arthralgia, and myalgia with a severity that honors “break-bone” fever alternative name of this disease. It lasts from three to 7 days. A small fraction of these patients evolve to the life-threatening clinical form of severe dengue, usually preceded by the appearance of warning signs (see below). All the four viral serotypes cause resembling disease symptoms.

In comparison to the previous 1997 version, the WHO revised classification of 2009. This makes more precision towards sensitivity and specificity of dengue cases. While being reported having changed dengue features with the passage of time during treatment; the dengue affected patients are categorized in Group A or Group B. More concern is recommended if symptoms becoming serious for next step of necessary hospitalization [39, 40]. As WHO 1997 version already includes plasma leakage and bleeding; but, however, the WHO 2009 classification entails target monitoring and organ impairment exhibiting the situation more clearly towards future dengue disease cure. Group C category of dengue patients has been explained in a better way in revised classification of WHO version 2009. This version, indeed is a practical guidance and very much helpful in dengue endemic areas especially where medical facilities are lacking. It is worth mentioning that WHO 2009 classification also highlights the other co-existing factors such as pregnancy, child and old age, diseases like diabetes and various social circumstances [41].

The WHO 2009 revised dengue classification stratifies disease into the following:

1. Without warning signs of dengue,
2. With warning signs of dengue (i.e., abdominal pain, vomiting, fluid accumulation, mucosal bleed, lethargic condition, liver enlargement >2 cm, and rapid decrease in platelet count), and
3. Severe symptoms of dengue.

Furthermore, three categories have been described in 2 WHO scheme of dengue-affected patients:

- i. Group A includes patients without warnings signs,
- ii. Group B includes patients with more than one warning sign, and annotated with certain coexisting conditions such as pregnancy, infancy, old age, obesity, diabetes, renal failure, and chronic hemolytic diseases), and with certain social circumstances,



- iii. Group C includes patients with severe plasma leakage, severe bleeding leading towards extreme condition of organ impairment [41].

The classification is meant to make it realized that the group is clearly identified so that patients are going to be treated keeping in view the relevant category.

## 7. Diagnosis of dengue fever

Dengue infection symptoms are the major tool for its diagnosis. However, this is not a reliable method for the confirmation of dengue infection but laboratory studies are needed [42, 43]. Dengue virus in the initial stages may cause fever to dengue fever or later on more it can result in severe dengue. Common tools for the detection of dengue infection in laboratory tests include; an identification of the particular viral serotype, genomic sequences, viral antigen, genomic sequence, and/or antibodies. Major advances in the diagnosis of this infection include IgM captured ELISA, dengue specific monoclonal antibodies, viral RNA detection by nucleic acid amplification tests (NAAT), and viral isolation from mosquito cell lines and also live mosquitoes, all these are reported to have major advances in dengue diagnosis. Diagnosis involves two levels of detection. At level one, the patient is in acute febrile phase, where NS1 antigens and viral RNA can be detected, and at level two is the stage in which IgG and IgM antibodies are abundant in blood with the post febrile period [44]. Acute stages of dengue may be represented by flu like fever in which diagnosis is made possible by identifying viral RNA/proteins in the patient's blood. Dengue viral RNA can also be identified in early stage of infection using RT-PCR. This technique is quite reliable but unaffordable for the poor people [45, 46]. ELISA test is also being utilized to identify primary as well as secondary infection by utilizing dengue-specific monoclonal NS1 antibody to identify NS1 in victim's blood [47–49]. MACELISA assays in combination with NS1 Ag can be utilized for the detection of dengue viruses in earlier stages of infection [50]. Commonly used laboratory methods include immune-fluorescence tests, capture ELISAs, and hemagglutination assays [51]. Nonetheless, it is important to consider that serological tests can be misleading due to cross-reactivity while there is more than flavivirus circulating in the region.

## 8. Control of dengue fever

Vaccines against dengue are difficult to develop. Nonetheless, as for December of 2015, CYD-TDV vaccine was approved for human use, and to date it remains as the sole vaccine with this status. As for specific treatments none is available, however various anti-viral natural entities are being evaluated for the elimination of dengue virus [32, 47, 48, 52–54].

There are several methods used to control dengue infection. The first and most important preventive measure is the prevention of contact with infected mosquitoes. *Aedes* mosquitoes usually have biting preferences during daytime and its contact can be minimized in various ways, for example, proper management of stored water and wastes, use of insecticides to eradicate the mosquitoes, use of mosquito nets and coils as well as repellents, use of wearing which minimize the exposed body surface. Insecticide treated nets (ITNs) are also available

in the market for the protection of people including young children, pregnant women, old people [55].

## 9. Control of vector *Aedes*

The best way to control dengue is to improve capabilities of mosquito abatement especially in the most populated areas where vector densities are high due to availability of hosts [18].

## 10. Public awareness

Public awareness counts in integrated pest management at a significant level, various examples can be cited from the literature when community efforts played a role for the eradication of disease agents. As *Aedes aegypti* was eliminated from countries various regions of the USA during the 1960s when relatively well funded eradication campaign supported by a high degree of political and community were involved. The effective collaboration of a well-educated society with the assistance of mosquito control well-trained staff will be the most compelling and economically reliable method for the removal of *Aedes albopictus* populations in rural and suburban regions [18].

## 11. Chemical control

The vector borne diseases are controlled worldwide, simply via controlling the vector. This thumb rule equally implies on dengue vector *Aedes* mosquitoes as well. Integrated Vector Management (IVM) mostly focuses on chemical control using insecticides; most frequently used are reported organophosphates and pyrethroids by WHO against dengue, malaria as well as yellow fever. These insecticides are affective against larvae, pupae and adults as well [25, 56, 57]. No new public health insecticides have been developed for mainstream vector control in disease-endemic countries (DECs) for the last three decades. Narrow range of public health insecticides necessitates new, safe, less expensive, environment friendly insecticides to replace those already being commercially used and mostly have been reported to develop resistance. Pyrethroid insecticides such as Permethrin, Deltamethrin, Cypermethrin, Cyhalothrin, etc. and DDVP organophosphate insecticides have been frequently used against mosquitoes and flies at household level. However, pyrethroid insecticides are reported to develop resistance. Hence, synergistic use of organophosphates and pyrethroid insecticides is being used now-a-days in order to combat this resistance menace [58]. New insecticides which are safe for health and environment as well demand investment. It is estimated that about US\$70 million amount is required to develop a new insecticide. Public health insecticide market encompasses about US\$151.2 million worldwide, hereby, shows the overall small size market. It is a dire need of time to engage commercial partners in the development of new insecticides. It has been suggested that both commercial and academic partners must collaborate and work together. In addition, community level health workers must be stimulated

to locate and target the investment so that safe, cost-effective, user-friendly vector control insecticides can be developed.

## 12. Biological control

Although, chemical use in the form of synthetic insecticides remains promising factor for the control of insect vectors; however, indiscriminate and overuse pose insecticide resistance issues [58]. Moreover, various health and environmental concerns make the use of insecticide questionable. Thus, it is imagined that in future only those techniques will be accepted which may overcome the problems related to chemical insecticides. Recently, non-chemical methods have been summed-up into “biopesticides”; meaning thereby simply to kill the pest using material originated from living things [59]. Hence, it necessitates to explore biological control agents like various predators and parasites, that is, viruses, fungi, bacteria, etc. to look for a potent agent for the development of safe control program. Various pathogens and predators have been reported to use against mosquitoes as biological control agents. Recently, in Vietnam, copepods were used to control larvae of *A. aegypti*. At local level, the control program was launched very successfully and showed good results [61, 62]. In addition, a bacterium *Wolbachia pipientis* which is an obligate intracellular bacterium and vertically transmitted from mother to their offsprings and causes cytoplasmic incompatibility. It has been reported to present in 60% populations of insects in field conditions. *Wolbachia* infects the gonads and ensures transmission to the next host generation and orchestrates various reproductive manipulations in host. The symbiont can also cause feminization of genetic males, parthenogenesis and male killing, depending on the host species [63]. Thus, via females the *Wolbachia* spreads in the host populations and ultimately hinders its increase in number in future. It is reported that *Wolbachia* infections spread upto 100 km per annum. The *Wolbachia* strains were manifested and manipulated successfully in 1967 in Burma against filariasis vectors, where *Wolbachia* infected male *Culex quinquefasciatus* were released in wild populations. In principle, *Wolbachia* infection affects the sperm and prevents the further reproduction as a measure of local mosquito population control [65].

The sterile insect technique (SIT) is widely tested strategy in insects; wherein, males are treated with either sterilizing chemicals or exposed to  $\gamma$ -irradiation producing random dominant lethal mutations; means only one locus containing the DNA damage can cause dominant effect in the form of lethality. The SIT males when mate with normal females results non-viable offsprings leading to elimination of the populations in successive generations [65]. Another approach is RIDL (release of insects carrying a dominant lethal mutation) which is an improved version of SIT using transgenic technique and specifically focuses on female-killing. For instance, gene specifically expressing in the flight muscles were made transgenically expressed low and the resulting females in the offsprings would not be able to fly properly which causes its non-feeding on human blood meal which ultimately leads towards low fecundity [67]. Specific transgenic approaches have been proved successful also in pupae and adults [68, 69]. This RIDL techniques is being exploited and deployed by Oxitec® in Brazil and Malaysia and reproduced appreciable results [70]. Subsequently, *Bacillus thuringiensis*

*israelensis* (Bti), methoprene and the insect growth hormone are also proven to be quite effective against *Aedes albopictus* in the laboratory as well as in the field [57, 71–75].

### 13. Botanical control

Plants as a whole and/or their certain parts plus various products originated from different plants have been incorporated in the control programs from long time ago. However, plant oils have been annotated with potentially good insecticidal properties [76, 77]. Plant extracts are reported as fumigant and caused ovicidal, larvicidal, and overall insecticidal activity against various insects. The plants derived insecticides; mostly mentioned as biopesticides are non-hazardous to the environment, cheap, and are considered safe to human as well as other animals. Black pepper extracts have been shown with significant potential as adulticide against *Aedes aegypti* and *Anopheles stephensi* [78].

It is thus, suggested that plant extracts have promising capability to control the mosquitoes. Being safe to human health and to the environment; these can be successfully incorporated in mosquito control programs [79]. In addition, a few plants extracts have been successfully tested against some viral diseases. Aforementioned wherein the life cycle of dengue virus; DENV makes an attachment with host via host receptors and envelope proteins; suggesting thereby that DENV infection can be controlled via inhibition of host-viral interactions using plants extracts. Moreover, NS2-NS3 protease and NS5 have also been reported as significant antiviral drug targets due to their impact on viral replication and other cellular processes as well [80, 81]. Several medicinal plants such as *Momordica charantia* and *Andrographis paniculata* have been reported in inhibiting the replication cycle of DENV. Few of the weed plants have been shown to cause insecticidal and enzyme inhibitory activities in insects [83, 84]. Further investigations are needed to develop potential dengue treatment [85].

### 14. Conclusion and future perspectives

Dengue viral disease is an emerging health concern in many regions of the world and has become a serious threat in many areas of the world including Southeast Asia and Pakistan. The control of the dengue is difficult as there is no vaccine available so far. Vaccine preparation against dengue requires a tetravalent vaccine but no such licensed compound has been prepared so far. However dengue viral envelope proteins can be targeted to make an effective drug against dengue as these proteins are involved in the entry of the virus into the host cell. Several medicinal plants have been identified so far which show significant inhibitory results against dengue but still there is a need of proper medicine which can show promising results. In future, exploration of interaction of *Aedes albopictus* and other mosquito species is required. *Toxorhynchites* mosquitoes should be searched out for their predatory role against *Aedes albopictus* and *Aedes aegypti*. Vectoral role of *Aedes albopictus* and *Aedes aegypti* must be regulated in and between countries. In addition, the predominant serotype in dengue outbreaks can be managed through respective vaccine especially against the documented serotype.



## Author details

Muhammad Kashif Zahoor<sup>1\*</sup>, Azhar Rasul<sup>1</sup>, Muhammad Asif Zahoor<sup>2</sup>, Iqra Sarfraz<sup>1</sup>, Muhammad Zulhussnain<sup>1</sup>, Rizwan Rasool<sup>1</sup>, Humara Naz Majeed<sup>3</sup>, Farhat Jabeen<sup>1</sup> and Kanwal Rania<sup>1</sup>

\*Address all correspondence to: kashif.zahoor@gcuf.edu.pk

<sup>1</sup> Department of Zoology, Government College University Faisalabad, Pakistan

<sup>2</sup> Department of Microbiology, Government College University Faisalabad, Pakistan

<sup>3</sup> Department of Biochemistry, Government College Women University, Faisalabad, Pakistan

## References

- [1] Anonymous. Etymologia dengue. *Emerging Infectious Diseases*. 2006;**12**:893
- [2] Mairuhu ATA, Wagenaar J, Brandjes DPM, van Gorp ECM. Dengue: An arthropod-borne disease of global importance. *European Journal of Clinical Microbiology & Infectious Diseases*. 2004;**23**:425-433
- [3] Hotta S. Experimental studies on dengue. I. Isolation, identification and modification of the virus. *The Journal of Infectious Diseases*. 1952;**90**:1-9
- [4] Gubler DJ. Dengue and dengue hemorrhagic fever: Its history and resurgence as a global public health problem. In: Gubler DJ, Kuno G, editors. *Dengue and Dengue Hemorrhagic Fever*. Wallingford, UK: CAB International; 1997. pp. 1-22
- [5] Rush B. An account of the bilious remitting fever: As it appeared in Philadelphia in the summer and autumn of the year 1780. *The American Journal of Medicine*. 1951;**11**(5):546-550
- [6] Siler JF, Hall MW, Hitchens AP. Dengue: Its history, epidemiology, mechanisms of transmission, etiology, clinical manifestations, immunity and prevention. *Philippine Journal of Science*. 1926;**29**:1-304
- [7] Bancroft TL. On the etiology of dengue fever. *Australasian Medical Gazette*. 1906;**25**:17-18
- [8] Kimura R, Hotta S. On the inoculation of dengue virus into mice. *Nippon Igaku*. 1944;**3379**:629-633
- [9] World Health Organization. *Weekly Epidemiological Monitor*. 2013;**6**:37
- [10] World Health Organization. *World Malaria Report*. Geneva: World Health Organization. 2009a
- [11] World Health Organization. Dengue and dengue haemorrhagic fever. In: *Fact Sheet No117*. Geneva: World Health Organization; 2009b

- [12] Idrees S, Ashfaq UA. A brief review on dengue molecular virology, diagnosis, treatment and prevalence in Pakistan. *Idrees and Ashfaq Genetic Vaccines and Therapy*. 2012;**10**:6
- [13] Hanna JN, Ritchie SA, Phillips DA, Serafin IL, Hills SL, van den Hurk AF. An epidemic of dengue 3 in far North Queensland, 1997-1999. *The Medical Journal of Australia*. 2001;**174**: 178-182
- [14] Cobelens FG, Groen J, Osterhaus AD, Leentvaar-Kuipers A, Wertheim-van Dillen PM, Kager PA. Incidence and risk factors of probable dengue virus infection among Dutch travellers to Asia. *Tropical Medicine & International Health*. 2002;**7**:331-338
- [15] Messer WB, Gubler DJ, Harris E, Sivananthan K, de Silva AM. Emergence and global spread of a dengue serotype 3, subtype III virus. *Emerging Infectious Diseases*. 2003;**9**: 800-809
- [16] Hasteed SB. Dengue fever and dengue Hemorrhagic fever. In: Kliegman RM, Behrman RE, Jenson HB, Stanton BF, editors. *Nelson Textbook of Pediatrics*. 18th ed. Philadelphia, USA: WB Saunders; 2007. pp. 1412-1414
- [17] Halstead SB. Etiologies of the experimental dengue of Siler and Simmons. *The American Journal of Tropical Medicine and Hygiene*. 1974;**23**:974-982
- [18] Gratz NG. Critical review of the vector status of *Aedes albopictus*. *Medical and Veterinary Entomology*. 2004;**18**:215-227
- [19] Fontenille D, Toto JC. *Aedes* (Stegomyia) *albopictus* (Skuse), a potential new dengue vector in southern Cameroon. *Emerging Infectious Diseases*. 2001;**7**:1066-1067
- [20] World Health Organization, Special Programme for Research, Training in Tropical Diseases, World Health Organization. Department of Control of Neglected Tropical Diseases, World Health Organization. Epidemic, & Pandemic Alert. Dengue: guidelines for diagnosis, treatment, prevention and control. World Health Organization. 2009. <http://apps.who.int/iris/bitstream/handle/10665/44188/9789241547871?sequence=1>
- [21] Jacobs MG, Robinson PJ, Bletchly C, Mackenzie JM, Young PR. Dengue virus nonstructural protein 1 is expressed in a glycosyl-phosphatidylinositol-linked form that is capable of signal transduction. *The FASEB Journal*. 2000;**14**:1603-1610
- [22] Gibbons RV, Vaughn DW. Dengue: An escalating problem. *BMJ*. 2002;**324**:1563-1566
- [23] Kow CY, Koon LL, Yin PF. Detection of dengue viruses in field caught male *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) in Singapore by type-specific PCR. *Journal of Medical Entomology*. 2001;**38**(4):475-479
- [24] Rey JR, O'Meara GF, O'Connell SM, Cutwa-Francis MM. Factors affecting mosquito production from storm water drains and catch basins in two Florida cities. *Journal of Vector Ecology*. 2006;**31**(2):334-343
- [25] Curtis CF, Jana-Kara B, Maxwell CA. Insecticide treated nets: Impact on vector populations and relevance of initial intensity of transmission and pyrethroid resistance. *Journal of Vector Borne Diseases*. 2003;**40**:1-9

- [26] Tripathi P, Kumar R, Tripathi S, Tambe JJ, Venkatesh V. Descriptive epidemiology of dengue transmission in Uttar Pradesh. *Indian Pediatrics*. 2008;**45**:315-318
- [27] Raheel U, Faheem M, Riaz MN, Kanwal N, Javed F, Sahar N, et al. Dengue fever in the Indian subcontinent: An overview. *Journal of Infection in Developing Countries*. 2011;**5**(4):239-247
- [28] Guzman MG, Kouri G. Dengue and dengue hemorrhagic fever in the Americas: Lessons and challenges. *Journal of Clinical Virology*. 2003;**27**:1-13
- [29] Wang WK, Sung TL, Lee CN, Lin TY, King CC. Sequence diversity of the capsid gene and the nonstructural gene NS2B of dengue-3 virus in vivo. *Virology*. 2002;**303**:181-191
- [30] Ross TM. Dengue virus. *Clinics in Laboratory Medicine*. 2010;**30**:149-160
- [31] Wang E, Ni H, Xu R, Barrett AD, Watowich SJ, Gubler DJ, et al. Evolutionary relationships of endemic/epidemic and sylvatic dengue viruses. *Journal of Virology*. **74**:3227-3234
- [32] Kuhn RJ, Zhang W, Rossmann MG, Pletnev SV, Corver J, Lenches E, et al. Structure of dengue virus: Implications for flavivirus organization, maturation, and fusion. *Cell*. 2001;**108**:717-725
- [33] Hanley KA, Lee JJ, Blaney JE Jr, Murphy BR, Whitehead SS. Paired charge-to-alanine mutagenesis of dengue virus type 4 NS5 generates mutants with temperature-sensitive, host range, and mouse attenuation phenotypes. *Journal of Virology*. 2002;**76**:525-531
- [34] Hidari KI, Suzuki T. Dengue virus receptor. *Tropical Medicine and Health*. 2011;**39**:37-43
- [35] Filomatori CV, Lodeiro MF, Alvarez DE, Samsa MM, Pietrasanta L, Gamarnik AV. A 5' RNA element promotes dengue virus RNA synthesis on a circular genome. *Genes & Development*. 2006;**20**:2238-2249
- [36] Kurane I, Innis BL, Nimmannitya S, Nisalak A, Meager A, Janus J, et al. Activation of T lymphocytes in dengue virus infections. High levels of soluble interleukin 2 receptor, soluble CD4, soluble CD8, interleukin 2, and interferon-gamma in sera of children with dengue. *The Journal of Clinical Investigation*. 1991;**88**:1473-1480
- [37] Kurane I, Innis BL, Nimmannitya S, Nisalak A, Meager A, Ennis FA. High levels of interferon alpha in the sera of children with dengue virus infection. *The American Journal of Tropical Medicine and Hygiene*. 1993;**48**:222-229
- [38] George R, Lum LC. Clinical spectrum of dengue infection. In: *Dengue and Dengue Haemorrhagic Fever*. New York: CAB International. 1997. pp. 89-113
- [39] Narvaez F, Gutierrez G, Pérez MA, Elizondo D, Nuñez A, Balmaseda A, et al. Evaluation of the traditional and revised WHO classifications of dengue disease severity. *PLoS Neglected Tropical Diseases*. 2011;**5**:e1397
- [40] Lee IK, Liu JW, Yang KD. Fatal dengue hemorrhagic fever in adults: Emphasizing the evolutionary pre-fatal clinical and laboratory manifestations. *PLoS Neglected Tropical Diseases*. 2012;**6**:e1532
- [41] World Health Organization. *Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control*. Geneva: World Health Organization. 2009

- [42] Burke DS, Nisalak A, Johnson DE, Scott RM. A prospective study of dengue infections in Bangkok. *The American Journal of Tropical Medicine and Hygiene*. 1988;**38**:172-180
- [43] Endy TP, Chunsuttiwat S, Nisalak A, Libraty DH, Green S, Rothman AL, et al. Epidemiology of inapparent and symptomatic acute dengue virus infection: A prospective study of primary school children in Kamphaeng Phet, Thailand. *American Journal of Epidemiology*. 2002;**156**:40-51
- [44] Libraty DH, Young PR, Pickering D, Endy TP, Kalayanarooj S, Green S, et al. High circulating levels of the dengue virus nonstructural protein NS1 early in dengue illness correlate with the development of dengue hemorrhagic fever. *The Journal of Infectious Diseases*. 2002;**186**:1165-1168
- [45] Kuno G, Cropp CB, Wong-Lee J, Gubler DJ. Evaluation of an IgM immunoblot kit for dengue diagnosis. *American Journal of Tropical Medicine and Hygiene*. 1998;**59**:757-762
- [46] Laue T, Emmerich P, Schmitz H. Detection of dengue virus RNA in patients after primary or secondary dengue infection by using the TaqMan automated amplification system. *Journal of Clinical Microbiology*. 1999;**37**:2543-2547
- [47] Shu PY, Chen LK, Chang SF, Yueh YY, Chow L, Chien LJ, et al. Comparison of capture immunoglobulin M (IgM) and IgG enzyme-linked immunosorbent assay (ELISA) and nonstructural protein NS1 serotype-specific IgG ELISA for differentiation of primary and secondary dengue virus infections. *Clinical and Diagnostic Laboratory Immunology*. 2003;**10**:622-630
- [48] Dussart P, Labeau B, Lagathu G, Louis P, Nunes MR, Rodrigues SG, et al. Evaluation of an enzyme immunoassay for detection of dengue virus NS1 antigen in human serum. *Clinical and Vaccine Immunology*. 2006;**13**:1185-1189
- [49] Khan AH, Hayat AS, Masood N, NM S, Shaikh TZ. Frequency and clinical presentation of dengue fever at Tertiary Care Hospital of Hyderabad/Jamshoro. *JLUMHS*. 2010;**09**(2): 88-93
- [50] Datta S, Wattal C. Dengue NS1 antigen detection: A useful tool in early diagnosis of dengue virus infection. *Indian Journal of Medical Microbiology*. 2010;**28**:107-110
- [51] Butt N, Abbassi A, Munir SM, Ahmad SM, Sheikh QH. Haematological and biochemical indicators for the early diagnosis of dengue viral infection. *Journal of the College of Physicians and Surgeons-Pakistan*. 2008;**282-285**(80):18
- [52] Chadwick D, Arch B, Wilder-Smith A, Paton N. Distinguishing dengue fever from other infections on the basis of simple clinical and laboratory features: Application of logistic regression analysis. *Journal of Clinical Virology*. 2006;**35**(2):147-153
- [53] Singh K, Lale A, Eong OE, Chiu L, Chow V, Tambyah P, et al. A prospective clinical study on the use of reverse transcription-polymerase chain reaction for the early diagnosis of dengue fever. *The Journal of Molecular Diagnostics*. 2006;**8**:613-616
- [54] <http://www.denguevirusnet.com/treatment.html>
- [55] Jahan F. Dengue fever (DF) in Pakistan. *Asia Pacific Family Medicine*. 2011;**10**:1. <https://doi.org/10.1186/1447-056X-10-1>



- [56] Zaim M, Jambulingam P. Global Insecticide Use for Vector Borne Disease Control. World Health Organization Communicable Disease Control, Prevention and Eradication, WHO Pesticide Evaluation Scheme (WHOPES). 2004. Geneva (No. WHO/CDS/WHOPES/GCDPP/2007.2)
- [57] Ali A, Nayar JK, Xue R-D. Comparative toxicity of selected larvicides and insect growth regulators to a Florida laboratory population of *Aedes albopictus*. Journal of the American Mosquito Control Association. 1995;**11**:72-76
- [58] Zaim M, Guillet P. Alternative insecticides: An urgent need. Trends in Parasitology. 2002;**18**(4):161-163
- [59] Broadhurst M. The influence of the molecular basis of resistance on insecticide discovery. Philosophical Transactions of the Royal Society of London B: Biological Sciences. 1998;**353**(1376):1723-1728
- [60] Amer A, Mehlhorn H. Repellency effect of forty-one essential oils against *Aedes*, *Anopheles*, and *Culex* mosquitoes. Parasitology Research. 2006;**99**:478-490
- [61] Nam VS, Yen NT, Duc HM, Tu TC, Thang VT, Le NH, et al. Community-based control of *Aedes aegypti* by using Mesocyclops in southern Vietnam. The American Journal of Tropical Medicine and Hygiene. 2012;**86**(5):850-859
- [62] Kay B, Nam VS. New strategy against *Aedes aegypti* in Vietnam. The Lancet. 2005;**365**(9459):613-617
- [63] O'Neill SL, Hoffmann AA, Werren JH, editors. Influential Passengers: Inherited Microorganisms and Arthropod Reproduction. USA, Oxford University Press; 1998. ISBN: 9780198501732
- [64] Charlat S, Hurst GD, Merçot H. Evolutionary consequences of Wolbachia infections. Trends in Genetics. 2003;**19**(4):217-223
- [65] Turelli M, Hoffmann AA. Cytoplasmic incompatibility in *Drosophila simulans*: Dynamics and parameter estimates from natural populations. Genetics. 1995;**140**(4):1319-1338
- [66] Alphey L, Benedict M, Bellini R, Clark GG, Dame DA, Service MW, et al. Sterile-insect methods for control of mosquito-borne diseases: An analysis. Vector-Borne and Zoonotic Diseases. 2010;**10**(3):295-311
- [67] Fu G, Lees RS, Nimmo D, Aw D, Jin L, Gray P, et al. Female-specific flightless phenotype for mosquito control. Proceedings of the National Academy of Sciences. 2010;**107**(10):4550-4554
- [68] Phuc HK, Andreasen MH, Burton RS, Vass C, Epton MJ, Pape G, et al. Late-acting dominant lethal genetic systems and mosquito control. BMC Biology. 2007;**5**(1):11
- [69] Bargielowski I, Nimmo D, Alphey L, Koella JC. Comparison of life history characteristics of the genetically modified OX513A line and a wild type strain of *Aedes aegypti*. PLoS One. 2011;**6**(6):e20699
- [70] Lacroix R, McKemey AR, Raduan N, Wee LK, Ming WH, Ney TG, et al. Open field release of genetically engineered sterile male *Aedes aegypti* in Malaysia. PLoS One. 2012;**7**(8):e42771

- [71] Farghal AI, Roe RM, Apperson CS. Evaluation of two insect growth regulators alone and in combination with *Bacillus thuringiensis* var. *israelensis* against *Culex quinquefasciatus* and *Aedes albopictus* larvae in the laboratory Assiut. *Journal of Agricultural Science*. 1988;**19**:284-303
- [72] Toma T, Kamiyama S, Fujihara S, Miyagi I. Effects of methoprene, a juvenile hormone analogue, on mosquito larvae from the Ryukyu Archipelago, Japan. *Japanese Journal of Sanitary Zoology*. 1990;**41**:99-103
- [73] Basci RS, Wright GB, Willis FS. Control of *Aedes albopictus* larvae using time release larvicide formulations in Louisiana. *Journal of the American Mosquito Control Association*. 1994;**10**:1-6
- [74] Sulaiman S, Jeffery J, Sohadi AR. Residual efficacy of triflumuron and methoprene against the dengue vector, *Aedes albopictus* (Skuse). *Bulletin of the Society for Vector Ecology*. 1994;**19**:111-114
- [75] Becnel JJ, Garcia J, Johnson M. Effects of three larvicides on the production of *Aedes albopictus* based on the removal of pupal exuviae. *Journal of the American Mosquito Control Association*. 1996;**12**:499-502
- [76] Adebayo TA, Gbolade AA, Olaifa JI. Comparative study of toxicity of essential oils to larvae of three mosquito species. *Nigerian Journal of Natural Products and Medicine*. 1999;**3**:74-76
- [77] Gbolade AA, Oyedele AO, Sosan MB, Adewoyin FB, Soyelu O. Mosquito repellent activities of essential oils from two Nigerian *Ocimum* species. *Journal of Tropical Medicinal Plants*. 2000;**1**:146-148
- [78] Trongtokit Y, Burivong P, Moore S, Hill N, Curtis CF. Plant Extracts in Prevention of Mosquito Attacks. *Botanical Medicine in Clinical Practice*. 2008. pp. 807-813
- [79] Nawaz R, Rathor RH, Bilal H, Hassan SA, Khan IA. Adulticidal activity of *Olea vera*, *Linum usitatissimum* and *Piper nigrum* against *Anopheles stephensi* and *Aedes aegypti* under laboratory conditions. *Journal of Arthropod-Borne Diseases*. 2011;**5**(2):2-9
- [80] Chow VT, Yong RY, Ngho BL, Chan YC. Automated type specific ELISA probe detection of amplified NS3 gene products of dengue viruses. *Journal of Clinical Pathology*. 1997;**50**:346-349
- [81] Falgout B, Miller RH, Lai CJ. Deletion analysis of dengue virus type 4 nonstructural protein NS2B: Identification of a domain required for NS2B-NS3 protease activity. *Journal of Virology*. 1993;**67**:2034-2042
- [82] Tang LI, Ling AP, Koh RY, Chye SM, Voon KG. Screening of anti-dengue activity in methanolic extracts of medicinal plants. *BMC Complementary and Alternative Medicine*. 2012;**12**:3
- [83] Sultana K, Zahoor MK, Sagheer M, Nasir S, Zahoor MA, Jabeen F, Riaz B. Insecticidal activity of weed plants, *Euphorbia prostrata* and *Chenopodium murale* against stored

grain insect pest *Trogoderma granarium* Everts, 1898 (Coleoptera: Dermestidae). Turkish Journal of Entomology. 2016;**40**(3):291-301

- [84] Riaz B, Zahoor MK, Zahoor MA, Majeed HN, Javed I, Ahmed A, Jabeen F, Zulhussnain M, Sultana K. Toxicity, phytochemical composition and enzyme inhibitory activities of some indigenous weed plant extracts in fruit fly, *Drosophila melanogaster*. Evidence-Based Complementary and Alternative Medicine. 2018;**2018**. Article ID: 2325659. DOI: 10.1155/2018/2325659
- [85] Talarico LB, Damonte EB. Interference in dengue virus adsorption and uncoating by carrageenans. Virology. 2007;**363**:473-485

IntechOpen