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Chapter

Invasive Candidiasis: Epidemiology and Risk Factors

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Abstract

Invasive candidiasis is a severe infection caused by the yeast of the genus *Candida*. This highly lethal infection can affect any organs, but it is usually identified by the growth of the yeast in bloodstream samples. Although *C. albicans* was the most frequently found species, there has been a global trend to the non-albicans isolates. The appearance of *C. auris*, a newly identified species around the world, is a cause of concern because of resistance to antifungals. In this chapter, the epidemiology and risk factors for the acquisition of candidemia and other forms of invasive candidiasis are reviewed, while showing the current knowledge of worldwide epidemiology.

Keywords: *Candida*, *Candida albicans*, invasive, candidiasis, fungemia, candidemia, intensive care units, surgery, immunosuppression, microbiota

1. Introduction

Candidasis is the common name for diseases produced by the yeast of the genus *Candida*. This is the most frequently found yeast in human microbiome and is capable of causing disease at different sites of the human anatomy and with diverse severity [1]. Invasive candidiasis refers to severe fungal infections in which the yeast might be found in deep organs or blood [2]. Due to the difficulty of identifying *Candida* yeasts in tissues, since it requires a biopsy of the tissue compromised, invasive candidiasis in the literature has been primarily found as bloodstream infections, alone or with accompanying tissue compromise.

2. Microbiology and environment

Candida species are yeasts (i.e., they mainly have a unicellular form). They are small, with a size of 4–6 μ m, with a thin wall and an ovoid aspect, named blastospores [3]. They reproduce by budding. Using the microscope, these yeasts can be seen in the form of pseudohyphae, budding cells that do not separate, or truly hyphae (multicellular organisms). *Candida* organisms belong to the class Ascomycetes, order Saccharomycetales, and family Saccharomycetes [4]. There are around 200 species of *Candida;* however, a limited number has a pathogenic effect on humans [4]. **Table 1** shows the most frequently found species. Due to their previous prevalence and pathogenic significance, they were usually classified as *albicans*

Species	Characteristic	
C. albicans	Usually the most frequently found	
C. parapsilosis complex	C. parapsilosis, C. orthopsilosis, C. metapsilosis	
C. tropicalis	Related to cancer	
C. glabrata	Usually resistant to azoles, seen more frequently in developed scenarios and older patients	
C. guilliermondii	Less pathogenicity	
C. lusitaniae	Potentially resistant to amphotericin	
C. krusei	Intrinsically resistant to azoles	
C. dubliniensis	Difficult to differentiate from <i>C. albicans</i>	
C. auris	Responsible for a global outbreak	

Table 1.

Most frequently found Candida species in human disease.

versus *non-albicans Candida* species. However, due to changes in epidemiology, this overall classification might not be useful any more.

They grow in agar as colonies with a smooth, creamy, white appearance. The formal identification can be made by use of biochemical physiological reactions, which can differentiate an important number of isolates. The metabolic reactions include carbohydrate fermentation, nitrate use, and urease production.

Candida yeasts might be seen with direct stains like KOH with 10–20% concentrations, but also with others like Gram, Giemsa, Wright para amino-salicylic (PAS) acid, and Papanicolaou. In direct stains, *Candida* might be seen as big aggregates of blastoconidiae, with short and large pseudohyphae. Usual growth media include Sabouraud agar, brain infusion, heart, and yeast extract. While *C. albicans* and *C. dubliniensis* grow in usual Sabouraud agar with antibiotics, some species might be inhibited by cycloheximide [4]. Usual growth time is 2–3 days at 28–37°C. Chromogenic agars were developed more than 20 years ago and are capable of identifying the most commonly found species, and speciation is desirable due to pathogenic and susceptibility differences among them. There are several commercial methods using chromogenic agars. The sensitivity for detection of *Candida* yeast is over 95%, usually with a low number or no false positive results [5]. The finding of a positive culture does not imply an invasive infection, and a special consideration has to be made for isolates from sterile sites.

Candida species differ in their susceptibility to different antifungals available in different countries. Most frequently found isolates of *C. albicans* and *C. parapsilosis* are susceptible to all antifungals available. *C. tropicalis* might have some resistance to fluconazole, while maintaining susceptibility to equinocandins and amphotericin B. *C. glabrata* tends to have higher minimal inhibitory concentrations (MICs) to azoles, while remaining susceptible to equinocandins and amphotericin B. *C. lusitaniae* isolates can be found to be resistant to amphotericin B. The recently found that *C. auris* is frequently found multidrug resistant.

Susceptibility testing can be performed by different methods, including broth microdilution (recommended in the USA and Europe), but there are other different commercial methods available in hospitals. Two slightly different standards for susceptibility testing are currently available. One is suggested by the Clinical Laboratory Standards Institute (CLSI, in USA), while the other is proposed by the European Committee on Antimicrobial Susceptibility Testing (EUCAST), sponsored by the European Society of Clinical Microbiology and Infectious Diseases

(ESCMID). Basic differences between both methodologies include time and instrument to read the results. Different clinical breakpoints have been established for the most commonly found species, with the intention of differentiating the risk of clinical failure after treatment. The experience with fluconazole has allowed to develop better prediction models, in comparison with newer antifungals [6]. In summary, an isolate of Candida is exposed to different concentrations of the antifungal and the *in vitro* growth is observed. If there is no important growth, determined optically or by a spectrophotometer, a minimal inhibitory concentration (MIC) is established. As mentioned, data from clinical trials and observation cohorts with common species such as C. albicans and C. glabrata have allowed to identify clinically relevant breakpoints to differentiate isolates with low MICs (susceptible); intermediate MICs (also called susceptible dose dependent—SDD), in which an increase in the administered antifungal can control the infection; and high MICs, (resistant), for which a lower probability of success is expected. For some other uncommon species, only epidemiological breakpoints are available. These breakpoints are also MICs, but there is no clinical evidence of correlation with the clinical outcome after treatment. However, since MICs are higher than those in usual isolates, a worse outcome might be expected. These breakpoints are expected to identify isolates with natural or acquired mechanisms of antifungal resistance. The epidemiological breakpoints are based on the statistical distribution of MICs of the wild-type isolates (i.e., isolates without any previous resistant pressure). Commercial methods are modifications of the standard methods that use dyes to identify the growth (e.g., Alamar Blue) of the microorganisms. Examples include Sensititre[™] and YeastOne[™]. Other methods are based on agar, in which a gradient of the antifungal is diffused in the solid growth media, which allows to directly read the MIC (e.g., Etest [™]) [7].

Candida species are part of the human microbiota and they live in human mucosae and skin. Candida species can be found in the ground, animals, fruits and vegetables, and in the hospital environment. It is not considered a laboratory contaminant. It is considered an endogenous pathogen since around 60-75% of the people might have it in the mucosal epithelium, especially in the gastrointestinal and genital tracts [8]. In the hospital area, they have been found over inanimate surfaces, including percutaneous catheters and tubes. They might even be found in the hands of healthcare workers. Among patients in healthcare centers, the colonization of the mucosae has been related to antibiotic use and hospitalization time [9]. In patients in the intensive care unit (ICU), colonization might be found in different anatomical sites with ample variations [10, 11]. Pharyngeal colonization rate has been found to be between 34 and 65%, gastric colonization between 42 and 67%, rectal colonization between 21 and 40%, and colonization in other sites between 11 and 40% [10, 11]. These data show the possibility of colonization that has this microorganism in patients under stress conditions (in this case, severe disease). In the normal host, the colonization rate might reach over 50% in the mouth, 40% in the vaginal tissue in women, and 73% in any mucosa of the gastrointestinal or genital tracts [8].

3. Pathogenesis

Candida species have some characteristics that permit them to adapt to different environments and act as an opportunistic pathogen. These factors include adaptation to pH changes, permitting to survive in blood or some alkaline environments, as well as in the acidic environment of the vaginal tissue; these species have adhesins, mannoproteins with capacity to adhere to different cells and cell products. These adherence proteins allow the isolates to survive in tissues, but also over inanimate surfaces that have been exposed to plasma or inflammatory host proteins like urinary or endotracheal catheters. *Candida* species have also important enzymes as virulence factors, since some of them have keratinolytic, peptidase, hemolysin, and other effects. One of the most frequently mentioned virulence factors include the possibility of a morphologic transition, which has been extensively studied. It refers to the possibility of morphologic changes of blastoconidia to pseudohyphae to real hyphae. These changes are stimulated by environmental conditions. The filamentous forms are related to active infection in the host, except for *C. glabrata*. Other factors related to pathogenicity or virulence also include a phenotyping change, the possibility of adopting different phenotypes in the cultures (color or aspect of the colonies), and biofilm formation. A biofilm is a large community of symbiotic microorganisms adhered to a surface. This conformation allows the microorganisms to have a highly defensive capacity, persistence, and a highly antimicrobial resistance.

As mentioned before, Candida might be part of the human flora. The majority of infections are due to the interplay between the risk factors, that pose a risk to the individual, the interaction with other microorganisms present in the skin or mucosa and the total quantity of microorganisms present. This was demonstrated some years ago in an experiment [12]. An individual ingested directly from a C. albicans culture. After some hours, this immunocompetent individual began to have fever. After 12 hours, *Candida* isolates were found in the bloodstream and, after 16 hours, they were found in the urine. After 24 hours, Candida isolates were cleared from the body and the individual returned to the normal state. This experiment proved the importance of colonization. With posterior evidence, it has been demonstrated that the first step to have an infection is colonization by *Candida* especially in the gastrointestinal tract, but otherwise in contact with indwelling catheters, the skin, or wounds that may permit the entry of the yeast into the bloodstream. In another critical observation, patients in the ICU were followed with cultures. The colonization index (it is the proportion of positive cultures for the same Candida species taken from different anatomical places) increased over time and was correlated to the probability of developing an invasive candidiasis [9]. These studies suggest that in individuals with Candida colonization, those factors that promote the grow of the yeast, by eliminating the bacteria that can compete for the environment, that alter or facilitate the penetration of the yeast to the bloodstream (lesions in the gastrointestinal mucosa, indwelling catheters) will promote the entry of Candida yeast to the blood, while the net state of compromise of the immune system will affect the probability of fungal clearance and the possibility of seeding on specific organs.

4. Epidemiology

4.1 Risk factors

4.1.1 Candida infection in the intensive care unit

Patients in the ICU have the highest rate of *Candida* infections in the hospital. In comparison with patients in other wards, patients in the ICU have more frequent abdominal surgery, stay longer in the hospital, and are more severely ill [13]. They also have a worse prognosis in the long term, with increased mortality after one year of the event.

4.1.1.1 Vascular devices

Patients in the ICU have higher rates of *Candida* infection in comparison with patients in other wards. Critically ill patients often require multiple vascular and other indwelling devices for their management and candidemia has been related to catheter colonization in 20-80% of the cases [14, 15]. One study in Japan identified the presence of a solid tumor, the use of total parenteral nutrition, and the administration of anti-anaerobic agents as the main risk factors for the development of Candida infections [16]. As mentioned, Candida colonization of the catheter might provide a route for entering into the bloodstream without a heavy gastrointestinal colonization. Studies have shown that *Candida* catheter-related bloodstream infections have a shorter time to grow in comparison with those from other sources [14]. With a breakpoint of 30 hours, the time to grow in patients with Candida bloodstream infection might identify 100% of those catheter-related infection. Probably, patients with catheter-related infection have a higher inoculum, which would explain the faster time to grow and the fact that observational studies have shown a lower mortality when catheter is removed [17, 18]. On the other hand, patients with non-catheter-related candidemia were more seriously ill, had a higher mortality, and the removal of the catheter did not affect the outcome [17].

4.1.1.2 Parenteral nutrition

Another commonly identified risk factor is the use of parenteral nutrition or the length of its use [15, 19]. This group of patients shares several risk factors, but parenteral nutrition has been identified in multivariate analysis [20]. Usually, they have an abdominal procedure (see below) and they require parenteral nutrition for several days. Lack of appropriate measures to handle the nutrition, colonization of the catheter or the ports used to infuse it, and probably the availability of optimal growing conditions are conditions related to its use. But clearly, the use of parenteral nutrition leads to the development of mucosal atrophy and a loss of mucosal epithelial barrier function [21], which might affect the relationship between microorganisms in the gut and the possibility of gaining access to blood vessels. Total parenteral nutrition has also a profound effect in the gastrointestinal microbiome [22].

4.1.1.3 Surgical procedures

Several studies have shown the relationship between candidemia and a previous surgical procedure [19, 23], specially an abdominal surgery. There are several explanations to this observation, but gut manipulation, and the effect of resected sections over the gut microbiology, microbiota abundance, and epithelial function might contribute to the possibility of candidemia. Studies have shown that patients with high anastomotic leak, as well as those with recurrent gastrointestinal perforation, or acute necrotizing pancreatitis, have a higher risk of candidemia [15].

4.1.1.4 Antibiotic use

Almost all studies of candidemia have shown an extremely high use of antibiotics previous to the identification of bloodstream or tissue infection. The proportion of patients with antibiotic use is over 80% [24]. The number and spectrum of the antibiotics used might affect the risk of candidemia. Antimicrobials also have an

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effect over gut microbiota, and some studies have shown some impact from antibiotics with anti-anaerobic effect, and those with higher gastrointestinal concentration [25]. They contribute to the observed increased colonization over time observed in patients in the ICU. With more antibiotic effect, there is a net decrease in the number of species in the gastrointestinal tract, an increase in the number of patients colonized, and the proportion of them being heavily colonized [26].

4.1.1.5 Other risk factors

Studies have identified several risk factors that alone, or in combination, might increase the probability of having candidemia. The presence of renal failure, the use of antihistaminic blockers, the severity of illness, and the length of stay in the ICU contribute to colonization and development of candidemia [24, 27]. All these factors contribute to the acquisition of *Candida*, its colonization, or failure in the gastrointestinal epithelial function, favoring the entry of the yeast to the bloodstream.

4.1.1.6 Scores based on risk factors

The identification of risk factors lead to the use of some scores based in the presence of such factors to identify patients with higher risk of *Candida* infection. The first and most simple of those scores was introduced in mid-1990s. Pittet et al. in a surgical ICU followed prospectively patients admitted in the ward with cultures of several anatomical sites [9]. They defined the colonization index as previously stated, establishing that with an index of 0.5 or more (50% of the sites with the same species), there was an increase in the risk of candidemia. With a lower colonization index, the risk in the original study was 0%. They defined a second index based on the density of colonization, in which patients overpassing some thresholds in the number of colonies isolated per site, being able to improve the identification of the patients at risk.

A second score to identify risk factors in patients was developed in Spain by León and his collaborators [20]. They identified colonization (with a different definition from that used by Pittet et al.), surgery at ICU admission, and use of total parenteral nutrition as risk factors independently associated with candidemia. They also identified sepsis as independently related, but this is clearly more a clinical syndrome than a risk factor. A third score was developed by a multicenter collaboration group, in which they again identified the same risk factors [28]: antibiotic use, having an intravascular catheter, in conjunction with at least two additional risk factors such as any surgery, immunosuppressive use, pancreatitis, total parenteral nutrition, dialysis, or steroid use.

Common to these scores has been the presence of the aforementioned risk factors. The problem, however, is that such scores identify a huge number of patients at risk with a final intermediate risk of developing candidemia, in a range from 7 to 30% [29, 30]. The great advantage of the diagnostic scores relies in their high negative predictive value. Patients with a negative score have a low probability of candidemia, below a 1% probability.

4.1.2 Hematological malignancy, solid organ transplantation, and other immunosuppressive states

These disorders share a common factor: immunosuppression. However, different types of immunocompromise entail different risks for the patients. The incidence of candidemia among patients with cancer is higher in comparison with other patients in the hospital. In a multicenter study in Greece, patients with

hematological disease had an incidence of candidemia of 1.4 cases per 1000 admissions, while other patients hospitalized had an incidence of 0.83 cases per 1000 admissions [31]. A multicenter European study found an incidence of 1.2% cases of candidemia among patients with bone marrow transplantation (BMT) and leukemia [32]. An Italian multicenter study from a surveillance network showed a diminishing trend for candidemia among patients with cancer, especially among those with acute myeloid leukemia [33]. Whether this trend can be inferred to other European countries or not is not known, and the most likely explanation for this decrease in the number of cases could be related to the use of prophylaxis among those patients with acute leukemia with posaconazole. In general, non-albicans *Candida* species are more frequently found among these groups of patients [31].

4.1.2.1 Neutropenia

Neutropenia, a count of leukocytes in peripheral blood below 500 cells per μ l, is the common risk factor among patients with hematological disorders (i.e., leukemia, lymphoma, multiple myeloma among others) as well as those with bone marrow transplantation (BMT). Neutropenia might be a consequence of the activity of the hematological disease, an effect of chemotherapeutic strategies or side effect of multiple medications including antimicrobials. It also is a marker of the intensity of chemotherapy. Patients with chemotherapy-induced neutropenia accumulate various risk factors: they usually receive wide spectrum antibiotics for several days, they have serious gastrointestinal epithelial tissue dysfunction, usually with diarrhea and signs of mucosal damage, and the use of vascular catheters for the infusion of chemotherapeutic drugs and antibiotics [34]. Several studies have shown that isolates of *C. tropicalis* are more frequently found among patients with cancer [35]. A study that looked for risk factors identified underlying leukemia as one of the major risk factors, together with chronic lung disease [36].

In patients with prolonged neutropenia, a condition called hepatosplenic candidiasis might be seen. In it, seeding of yeasts occurs during the neutropenic phase which might be not clinically evident until neutropenia recovery. In these patients, fever persists and lesions can be seen in the liver, usually known as bull-eye lesions [37] (**Figure 1**).

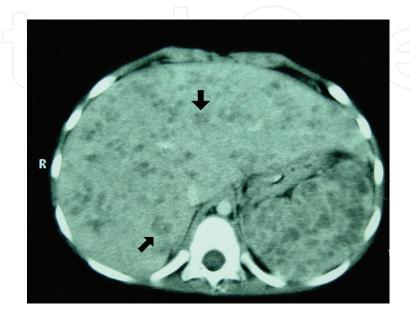


Figure 1.

Tomographic image of liver and spleen showing abscesses (bull's eye, arrows) and hypodense lesions in a patient with chronic disseminated candidiasis. Reproduced with permission from Cortés et al. [37].

4.1.2.2 Concurrent conditions in patients with cancer

In patients with cancer and candidemia, several factors were identified in comparison with those with cancer and bacterial infections [38]. Total parenteral nutrition over 5 days, urinary catheter for more than 2 days, distant metastasis of cancer, and gastrointestinal cancer were independent risk factors. Patients with solid tumors might accumulate factors as patients in critical care, since they have abdominal surgery (gastrointestinal neoplasm), require vascular catheters for extended periods of time (for chemotherapy or antibiotics), total parenteral nutrition and received antibiotics frequently [39]. A study to identify factors predicting catheter-related infections with *Candida* identified solid tumors and the use of antianaerobic antibiotics as risk factors [16].

Among patients with leukemia and BMT, the risk factors for occurrence of candidemia included bone marrow or cord blood stem cell source, T-cell depletion, use of total body irradiation, and acute graft versus host disease [32]. These data were derived from a huge multicenter registry of patients with cancer and transplantation, which allowed to identify more precisely the risk factors.

4.1.3 Neonates

Newborns have no gastrointestinal flora at birth and have to be colonized by enterobacteria and other microorganisms, which is made via maternal breast feeding. Any alteration in the normal process can lead to colonization by pathogenic microorganisms, including yeasts [40]. Neonates in the intensive care unit usually have limited breastfeeding, indwelling vascular catheters, total parenteral nutrition, and antibiotics [41]. Such combination of risk factors put. this group of patients at a higer risk of infection, reaching over 10% of patients in units with extreme prematures and low weight at birth (the group that requires more invasive interventions) [42]. Some studies have illustrated this relationship with proportion of candidemia between 3 and 10% among those with a weight of less than 1000 g while showing an incidence of less than 1% for those weighting over 2500 g [43]. In this scenario, disseminated candidemia can be found and near 10% of those with invasive disease can compromise the central nervous system. Another important risk factor includes the time that the patient has been in the unit [44]; clearly, patients with low weight, lower gestational age, and more comorbidity tend to spend more time in the neonatal ICU and to accumulate other risk factors (surgery, indwelling catheter, antibiotics, etc.) [45]. There are some high-risk units, in which the incidence of candidemia traditionally has been high, usually over 10% of the admitted cases. In this scenario, prophylaxis has been suggested for the prevention of infection [46].

4.1.4 Outbreaks

Candida yeast can survive in inanimate surfaces and in the hands of healthcare personnel, which confers the risk of outbreak and cross dissemination among high-risk units such as neonatal, intensive care, and surgical intensive care units [44, 47]. An interesting study in Iceland over a long period of time allowed to confirm the presence of clonal isolates of different *Candida* species among patients in the ICU and other wards [48]. The proportion of patients involved at one time with an outbreak of all patients with *Candida* isolates might be as high as 38%. Other study in Spain showed that clusters (of patients with candidemia) were possible with *C. albicans* and *C. parapsilosis*, and reached in a period as high as 40% of the isolates [49]. Besides, the use of chlorhexidine has been shown to diminish the number of

candidemia events in patients in the ICU, showing the importance of colonization and cross infection among high-risk patients and establishing this recommendation in the guidelines for the prevention of candidemia [50].

As shown, colonization is the preliminary step to infection. Besides, a number of interventions are common to immunosuppressed and critically ill patients including indwelling catheters (urinary and vascular), severity of illness, total parenteral nutrition, etc. These conditions predispose the patients to cross contamination. An outbreak among newborns was demonstrated to be due to poor practices of catheter ports disinfection [51].

A study in China in a cancer institute showed that 21 out of 36 episodes of candidemia were caused by two endemic genotypes [52]. In this study, gastrointestinal cancer and insertion of a nasogastric tube were related to infection. As mentioned before, cancer patients with solid and hematological tumors share several of the risk factors of colonization and infection.

4.2 Global epidemiology

Since 2013, the Leading International Fungal Education (LIFE) portal has facilitated an important effort to know the epidemiology and burden of fungal infections around the world and allowed a better understanding of their epidemiology in different countries [53]. The real incidence of candidemia is difficult to calculate due to differences in the approach. While studies based on hospitals might overestimate the importance of some groups of high-risk patients, they are difficult to compare. Data from population studies might reflect better the real situation, but this kind of information is scarce. Studies have shown ample differences in the incidence in some regions and at specific times [54].

4.2.1 Changing trend for non-albicans Candida

Traditionally, *C. albicans* had been the most frequently isolated species. However, a trend toward non-albicans species has been observed around the world in the last 15 years. In United States, *C. glabrata* has been identified as second in frequency, while *C. parapsilosis* or *C. tropicalis* dispute this place in other regions. **Table 2** shows the proportion of isolates in some studies around the world in the last 10 years [55–59].

Two studies deserve a detailed description. The first one is a multicenter study from the Southeast Asia region, including 25 hospitals from 6 countries: China, Hong Kong, India, Singapore, Taiwan, and Thailand [60]. They found differences between the countries that include the frequency of *C. tropicalis* isolation, being

Area and publication year	C. albicans (%)	C. glabrata (%)	C. tropicalis	C. parapsilosis	References
USA 2012	38	29	17	10	[49]
Latin America 2013	37.6	6.3	17.6	26.5	[48]
Spain 2014	45.4	13.4	7.7	24.9	[50]
Asia-Pacific region 2016	20–55	5–22	2–20	8–27	[51]
France 2014	56	18.6	9.3	11.5	[52]

Table 2.

Proportion of Candida species in selected studies of candidemia around the world.

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more commonly found in hematology-oncology wards and in tropical areas. This study confirmed the observed trend for a lower frequency of *C. albicans* isolates. The other study is the Latin-American surveillance study [55]. It involved patients from 20 centers in 7 seven countries: Argentina, Brazil, Colombia, Chile, Honduras, Mexico, and Venezuela. Important differences were seen among institutions, reflecting difference in healthcare systems, access, population types, and risk factors. However, in these two studies, the incidence of candidemia is higher than in developed countries in Europe and North America. In Latin America, *C. parapsilosis* frequency is over 30% of the isolates while this place is occupied by *C. tropicalis* in the Asian countries.

4.2.2 Epidemiology in Europe and North America

There are data from some population surveillance surveys in Europe and United States. In general, the incidence might be lower than in some other areas of the world. **Table 3** shows the incidence from data from North America and European countries [61–77]. In Europe, the highest incidence has been observed in Hungary, while in North America the highest incidence has been seen in some cities in United States.

4.2.3 Epidemiology in Central and South America and the Caribbean

This region has profound differences in healthcare systems, access to care, and medical technology development. With a transition toward a higher income, a growing number of institutions with capacity to attend cancer patients, and more

Country/region	Publication Year	Incidence (per 100.000 inhabitants)	References
Belgium	2015	5	[54]
Denmark	2008	10.4	[55]
Finland	2010	2.8	[56]
Germany	2015	4.6	[57]
Hungary	2015	11	[58]
Ireland	2015	7.3	[59]
Norway	2018	3.8	[60]
Portugal	2017	2.57	[61]
Romania	2018	6.8	[62]
Russia	2015	8.29	[63]
Serbia	2018	10	[64]
Spain	2015	8.1	[65]
Sweden	2013	4.2	[66]
Ukraine	2015	5.8	[67]
Canada	2017	2.91	[68]
México	2015	8.6	[69]
USA	2015	9.5–14.4	[70]

Table 3.

Estimated incidence of invasive candidiasis or candidemia in countries of the European or North American regions.

Country/region	Publication year	Incidence (per 100,000 inhabitants)	References	
Argentina	2018	6.25	[71]	
Brazil	2016	14.9	[72]	
Chile	2017	5.8	[73]	
Colombia	2018	14.7	[74]	
Ecuador	2017	7.2	[75]	
Guatemala	2017	6.4	[76]	
Jamaica	2015	5.8	[77]	
Perú	2017	5.8	[78]	
Trinidad and Tobago	2015	5.8	[79]	
Uruguay	2018	36.5	[80]	

Table 4.

Estimated incidence of invasive candidiasis or candidemia in countries of Central and South America and the Caribbean.

complex medical needs, the number of candidemia cases seems to be higher than in developed countries.

Ample information exists about the problem in Brazil, where a number of studies have been carried out in high-complexity hospitals in the main cities of the country [78, 79]. These studies show a higher frequency of invasive candidiasis in comparison with developed countries, an increased isolation of *C. glabrata* for the last period and an important exposition to fluconazole (which might have increased the selection for non-albicans species) [79]. Country-wise estimates for incidence are shown in **Table 4** [80–89].

4.2.4 Epidemiology in Africa and Asia

A multicenter in Asia gathered information from various countries, including nine hospitals from China [60]. The incidence rate among patients hospitalized was 0.38 per 1000 admissions, which is lower than that observed in the Latin-American region with 1.08 cases per 1000 admissions [55]. The estimated incidence of candidemia in countries in Asia is shown in **Table 5** [90–100]. In Asia, the highest incidence has been observed in Pakistan, followed by Qatar and Israel. In China, geographic variations in the causative species and susceptibilities were noted, with increasing isolates resistant to fluconazole [101]. The numbers for the African countries are lacking and for some countries like Algeria, Burkina Faso, Cameroon, Egypt, Malawi, Mozambique, and Tanzania, the estimated incidence is 5.8 cases per 100,000 inhabitants, a standard calculation based on previously reported incidence in other countries [102–108].

4.2.5 Azole resistance epidemiology

Azole-resistant *Candida* isolates have had an increased frequency over the years. Susceptibility changes with the species, and fluconazole use has been related to an increase in the frequency of *C. glabrata* and *C. krusei*, and a low increase in the number of resistant *C. albicans* or *C. tropicalis*. A large multicenter study in French ICUs identified the age and the exposure to antifungals as independent risk factors for resistance [109]. Patients with isolates resistant to fluconazole tended to be older than 15 years and to have been exposed to this drug, while those with

Country/region	Publication year Ir	Incidence (per 100,000 inhabitants)	References
Bangladesh	2017	5	[83]
Israel	2015	11	[84]
Jordan	2018	5.75	[85]
Kazakhstan	2018	4.3	[86]
Korea	2017	4.57	[87]
Malaysia	2018	5.8	[88]
Pakistan	2017	21	[89]
Philippines	2017	2.25	[90]
Qatar	2015	15.4	[91]
Thailand	2015	13.3	[92]
Uzbekistan	2017	5.93	[93]

Table 5.

Estimated incidence of invasive candidiasis or candidemia in countries of Africa and Asia.

equinocandin-resistant isolates were younger and found to have been exposed to equinocandin. In general, risk factors for resistance remain the same as in resistant bacteria: immunosuppression, previous use of antifungals [110, 111]. Other identified risk factors include chronic renal failure and anti-tuberculous treatment. This last one might be due to a medication interaction.

Among patients with cancer, not only are non-albicans *Candida* species more frequently found, but also resistance to azoles has increased. In a study in Greece, resistance to fluconazole among patients with cancer reached 27% [31]. Since azoles have been widely used in the prophylaxis against fungal infections among cancer patients [112, 113], this seems to be a natural consequence of their use. Among patients with cancer, isolates of *C. tropicalis*, *C. glabrata*, and *C. krusei* have increased resistant proportions [35].

4.2.6 Candida auris global outbreak

Up to 2009, there was no report on *C. auris*. In that year, a clinical case from Japan was published, and 2 years later three cases of candidemia were identified [114, 115]. During the following years, isolates of C. auris were responsible of outbreaks around the world, affecting hospitals in India, Pakistan, South Africa, England, and Venezuela [116–119]. It was detected in the USA in 2013 with growing frequency [120]. A worldwide alarm was raised in 2016 because of two problems related to this species. The first one was the difficulty in proper identification [121]. C. auris is most commonly identified as C. haemulonii and *Rhodotorula glutinis* by the commercial systems and sometimes as *C. famata*, *C.* guilliermondii, and C. parapsilosis [121]. The other problem is the higher frequency of resistance to multiple antifungals, including azoles and amphotericin [122]. Currently, C. auris has been isolated in several areas in the USA, continental Europe, and the Caribbean coast of South America, including the islands [123–125], and continue to extend to other areas, where reports are being published. A search for virulence factors in the isolates of *C. auris* has shown some different properties, specially the capacity for biofilm formation [126]. Molecular observations have diverse geographic dissemination caused by unique clades in each geographic region [127].

5. Outcomes

Patients with candidemia and cancer are considered to have higher mortality, but this issue has not been clearly assessed. Older studies showed an attributable mortality around 40%. Although mortality among patients with candidemia or invasive candidiasis is reported usually around 40–50%, they occur in patients with important comorbidity. A recent multicenter analysis showed a crude mortality for patients with candidemia of 53%, while those without candidemia had a mortality of 26% [128]. After adjusting in a propensity score analysis, the crude mortality was 51% for the candidemic patients and 37% for the others and the difference was not statistically significant. The study shows that an increase in mortality might exist for those patients with candidemia, but it is clear that patients with candidemia also have severe comorbidity and some of them can die with candidemia instead of because of it.

Risk factors for mortality among patients with candidemia include ascites, presence of septic shock, ICU admission, concomitant bacterial infection and catheterrelated infections [129]. Studies with diverse population have shown that elderly patients have higher mortality [130]. In these patients, a combination of comorbidity, poor clinical situation, and more pathogenic species might contribute to their mortality [131]. A pooled analysis from patients included in randomized clinical trials comparing micafungin and amphotericin B showed differences among geographic regions, severity of disease (measured with Apache score for patients critically ill), and catheter removal [132]. In those with abdominal candidiasis, the lack of control of the source of infection has been related to increased risk of death [133]. Among patients with cancer, risk factor for mortality includes infection by a *C*. tropicalis isolate, a high Charlson index score, neutropenia, and septic shock [35, 134]. One multicenter study identified tachypnea as a risk factor for mortality [135], while others identified respiratory failure and use of non-antifungal medications [39]. Besides, antifungal prophylaxis and remission of the underlying cancer had a protective effect over mortality [135].

The impact of the antifungal treatment in the mortality of patients with candidemia is not entirely clear. There are several constrains to identify the benefits of the antifungal treatment: An important proportion of patients did not receive antifungal treatment despite the identification of a bloodstream infection; of those that receive the treatment, some of them can receive it as empirical treatment, based on the risk factors, clinical condition, while others have an antifungal started upon detection of candidemia. Besides, some of them are infected with a resistant isolate and some do poorly, and an additional antifungal must be started. Although meta-analysis with patient-level data has showed the benefit of equinocandin use (in contrast to azole treatment) [136], neither the cohort data [137], nor the randomized trials have confirmed this finding [138]. There is an additional complication in understanding this relationship; the laboratory breakpoints for identification of susceptible versus resistant isolates have changed over the time, especially for azoles [130]. Among those patients with septic shock, the delay in the administration of the antifungal treatment has been associated with increased mortality.

6. Conclusion

Candidemia is the most frequently found form of invasive candidiasis. The *Candida* species might be found as part of the flora and patients with previous colonization are at risk of developing an infection. They share some common factors

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like antibiotic exposure, use of indwelling catheters, parenteral nutrition, and surgery. These factors affect the normal physiology of the gastrointestinal tract or provide access to the bloodstream to yeast in patients with some comorbidities, in critical care or with immunosuppressive states.

Conflict of interest



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References

[1] Kullberg BJ, Arendrup MC. Invasive candidiasis. The New England Journal of Medicine. 2015;**373**:1445-1456

[2] Quindos G. Epidemiology of candidaemia and invasive candidiasis. A changing face. Revista Iberoamericana de Micología. 2014;**31**:42-48

[3] Dadar M, Tiwari R, Karthik K, Chakraborty S, Shahali Y, Dhama K. *Candida albicans*—Biology, molecular characterization, pathogenicity, and advances in diagnosis and control—An update. Microbial Pathogenesis. 2018; **117**:128-138

[4] Howell SA, Hazen KC, Brandt ME. *Candida, Cryptococcus*, and other yeast of medical importance. En: Jorgensen JH, Pfaller MA, editors. Manual of Clinical Microbiology. Washington, D.C.: American Society of Microbiology. 2015:1984-2014

[5] Perry JD. A decade of development of chromogenic culture media for clinical microbiology in an era of molecular diagnostics. Clinical Microbiology Reviews. 2017;**30**:449-479

[6] Patel TS, Carver PL, Eschenauer GA. Are in vitro susceptibilities to azole antifungals predictive of clinical outcome in the treatment of candidemia? Journal of Clinical Microbiology. 2018

[7] Sanguinetti M, Posteraro B. Susceptibility testing of Fungi to antifungal drugs. Journal of Fungi (Basel). 2018;4:E110

[8] Odds FC. Candida infections: An overview. Critical Reviews in Microbiology. 1987;**15**:1-5

[9] Pittet D, Monod M, Suter PM, Frenk E, Auckenthaler R. Candida colonization

and subsequent infections in critically ill surgical patients. Annals of Surgery. 1994;**220**:751-758

[10] Magill SS, Swoboda SM, Johnson EA, Merz WG, Pelz RK, Lipsett PA, et al. The association between anatomic site of Candida colonization, invasive candidiasis, and mortality in critically ill surgical patients. Diagnostic Microbiology and Infectious Disease. 2006;**55**:293-301

[11] Leon C, Alvarez-Lerma F, Ruiz-Santana S, Leon MA, Nolla J, Jorda R, et al. Fungal colonization and/or infection in non-neutropenic critically ill patients: Results of the EPCAN observational study. European Journal of Clinical Microbiology & Infectious Diseases. 2009;**28**:233-242

[12] Krause W, Matheis H, Wulf K. Fungaemia and funguria after oral administration of *Candida albicans*. Lancet. 1969;**1**:598-599

[13] Ylipalosaari P, Ala-Kokko TI, Karhu J, Koskela M, Laurila J, Ohtonen P, et al. Comparison of the epidemiology, risk factors, outcome and degree of organ failures of patients with candidemia acquired before or during ICU treatment. Critical Care. 2012;**16**:R62

[14] Ben-Ami R, Weinberger M, Orni-Wasserlauff R, Schwartz D, Itzhaki A, Lazarovitch T, et al. Time to blood culture positivity as a marker for catheter-related candidemia. Journal of Clinical Microbiology. 2008;**46**: 2222-2226

[15] Blumberg HM, Jarvis WR, Soucie JM, Edwards JE, Patterson JE, Pfaller MA, et al. Risk factors for candidal bloodstream infections in surgical intensive care unit patients: The NEMIS prospective multicenter study. The National Epidemiology of Mycosis Survey. Clinical Infectious Diseases. 2001;**33**:177-186

[16] Nagao M, Hotta G, Yamamoto M, Matsumura Y, Ito Y, Takakura S, et al. Predictors of Candida spp. as causative agents of catheter-related bloodstream infections. Diagnostic Microbiology and Infectious Disease. 2014;**80**:200-203

[17] Arias S, Denis O, Montesinos I, Cherifi S, Miendje Deyi VY, Zech F. Epidemiology and mortality of candidemia both related and unrelated to the central venous catheter: A retrospective cohort study. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:501-507

[18] Garnacho-Montero J, Diaz-Martin A, Garcia-Cabrera E, Ruiz Perez de Pipaon M, Hernandez-Caballero C, Lepe-Jimenez JA. Impact on hospital mortality of catheter removal and adequate antifungal therapy in Candida spp. bloodstream infections. The Journal of Antimicrobial Chemotherapy. 2013; **68**:206-213

[19] Chow JK, Golan Y, Ruthazer R, Karchmer AW, Carmeli Y, Lichtenberg DA, et al. Risk factors for albicans and non-albicans candidemia in the intensive care unit. Critical Care Medicine. 2008;**36**:1993-1998

[20] Leon C, Ruiz-Santana S, Saavedra P, Almirante B, Nolla-Salas J, Alvarez-Lerma F, et al. A bedside scoring system ("Candida score") for early antifungal treatment in nonneutropenic critically ill patients with Candida colonization. Critical Care Medicine. 2006;**34**:730-737

[21] Yang H, Feng Y, Sun X, Teitelbaum DH. Enteral versus parenteral nutrition: Effect on intestinal barrier function. Annals of the New York Academy of Sciences. 2009;**1165**:338-346

[22] Pierre JF. Gastrointestinal immune and microbiome changes during parenteral nutrition. American Journal of Physiology. Gastrointestinal and Liver Physiology. 2017;**312**:G246-GG56

[23] Ortiz Ruiz G, Osorio J, Valderrama S, Alvarez D, Elias Diaz R, Calderon J, et al. Risk factors for candidemia in nonneutropenic critical patients in Colombia. Medicina Intensiva. 2016;**40**: 139-144

[24] Wey SB, Mori M, Pfaller MA, Woolson RF, Wenzel RP. Risk factors for hospital-acquired candidemia. A matched case-control study. Archives of Internal Medicine. 1989;**149**:2349-2353

[25] Samonis G, Gikas A, Anaissie EJ, Vrenzos G, Maraki S, Tselentis Y, et al. Prospective evaluation of effects of broad-spectrum antibiotics on gastrointestinal yeast colonization of humans. Antimicrobial Agents and Chemotherapy. 1993;**37**:51-53

[26] Guyton K, Alverdy JC. The gut microbiota and gastrointestinal surgery. Nature Reviews. Gastroenterology & Hepatology. 2017;**14**:43-54

[27] Bross J, Talbot GH, Maislin G, Hurwitz S, Strom BL. Risk factors for nosocomial candidemia: A case-control study in adults without leukemia. The American Journal of Medicine. 1989;**87**: 614-620

[28] Ostrosky-Zeichner L, Sable C, Sobel J, Alexander BD, Donowitz G, Kan V, et al. Multicenter retrospective development and validation of a clinical prediction rule for nosocomial invasive candidiasis in the intensive care setting. European Journal of Clinical Microbiology & Infectious Diseases. 2007;**26**:271-276

[29] Ostrosky-Zeichner L, Shoham S, Vazquez J, Reboli A, Betts R, Barron MA, et al. MSG-01: A randomized, double-blind, placebo-controlled trial of caspofungin prophylaxis followed by preemptive therapy for invasive

candidiasis in high-risk adults in the critical care setting. Clinical Infectious Diseases. 2014;**58**:1219-1226

[30] Leon C, Ruiz-Santana S, Saavedra P, Galvan B, Blanco A, Castro C, et al. Usefulness of the "Candida score" for discriminating between Candida colonization and invasive candidiasis in non-neutropenic critically ill patients: A prospective multicenter study. Critical Care Medicine. 2009;**37**:1624-1633

[31] Gamaletsou MN, Walsh TJ, Zaoutis T, Pagoni M, Kotsopoulou M, Voulgarelis M, et al. A prospective, cohort, multicentre study of candidaemia in hospitalized adult patients with haematological malignancies. Clinical Microbiology and Infection. 2014;**20**:O50-O57

[32] Cesaro S, Tridello G, Blijlevens N, Ljungman P, Craddock C, Michallet M, et al. Incidence, risk factors and longterm outcome of acute leukemia patients with early candidemia after allogeneic stem cell transplantation. A study by the acute leukemia and infectious diseases working parties of EBMT. Clinical Infectious Diseases. 2018;**67**(4):564-572

[33] Pagano L, Dragonetti G, Cattaneo C, Marchesi F, Veggia B, Busca A, et al. Changes in the incidence of candidemia and related mortality in patients with hematologic malignancies in the last ten years. A SEIFEM 2015-B report. Haematologica. 2017;**102**:e407-ee10

[34] Raad I, Hanna H, Boktour M, Girgawy E, Danawi H, Mardani M, et al. Management of central venous catheters in patients with cancer and candidemia. Clinical Infectious Diseases. 2004;**38**: 1119-1127

[35] Wu PF, Liu WL, Hsieh MH, Hii IM, Lee YL, Lin YT, et al. Epidemiology and antifungal susceptibility of candidemia isolates of non-albicans Candida species from cancer patients. Emerging Microbes and Infections. 2017;**6**:e87

[36] Fernandez-Ruiz M, Puig-Asensio M, Guinea J, Almirante B, Padilla B, Almela M, et al. *Candida tropicalis* bloodstream infection: Incidence, risk factors and outcome in a population-based surveillance. The Journal of Infection. 2015;**71**:385-394

[37] Cortés JA, Cuervo SI, Hernandez LF, Potdevin G, Urdaneta AM. Ojo de buey en tomografia hepatica. Biomédica. 2004;**24**:7-12. DOI: 10.7705/ biomedica.v24i1.1243

[38] Li D, Xia R, Zhang Q, Bai C, Li Z, Zhang P. Evaluation of candidemia in epidemiology and risk factors among cancer patients in a cancer center of China: An 8-year case-control study. BMC Infectious Diseases. 2017;**17**:536

[39] Tang HJ, Liu WL, Lin HL, Lai CC. Epidemiology and prognostic factors of candidemia in cancer patients. PLoS One. 2014;**9**:e99103

[40] Ortegon L, Puentes-Herrera M, Corrales IF, Cortes JA. Colonization and infection in the newborn infant: Does chlorhexidine play a role in infection prevention? Archivos Argentinos de Pediatría. 2017;**115**:65-70

[41] Chen J, Jiang Y, Wei B, Ding Y, Xu S, Qin P, et al. Epidemiology of and risk factors for neonatal candidemia at a tertiary care hospital in western China. BMC Infectious Diseases. 2016;**16**:700

[42] Benjamin DK Jr, Stoll BJ, Gantz MG, Walsh MC, Sanchez PJ, Das A, et al. Neonatal candidiasis: Epidemiology, risk factors, and clinical judgment. Pediatrics. 2010;**126**:e865-e873

[43] Benjamin DK Jr, Stoll BJ, Fanaroff AA, McDonald SA, Oh W, Higgins RD, et al. Neonatal candidiasis among extremely low birth weight infants: Risk factors, mortality rates, and neurodevelopmental outcomes at 18 to 22 months. Pediatrics. 2006;**117**:84-92

[44] Orozco PA, Cortes JA, Parra CM. Colonization by yeasts in newborns and healthcare personnel in a neonatal intensive care unit at a university hospital in Bogota, Colombia. Revista Iberoamericana de Micología. 2009;**26**: 108-111

[45] Fu J, Wang X, Wei B, Jiang Y, Chen J. Risk factors and clinical analysis of candidemia in very-low-birth-weight neonates. American Journal of Infection Control. 2016;**44**:1321-1325

[46] Cleminson J, Austin N, McGuire W. Prophylactic systemic antifungal agents to prevent mortality and morbidity in very low birth weight infants. Cochrane Database of Systematic Reviews. 2015;**10**:CD003850

[47] van Schalkwyk E, Iyaloo S, Naicker SD, Maphanga TG, Mpembe RS, Zulu TG, et al. Large outbreaks of fungal and bacterial bloodstream infections in a neonatal unit, South Africa, 2012–2016. Emerging Infectious Diseases. 2018;**24**: 1204-1212

[48] Asmundsdottir LR, Erlendsdottir H, Haraldsson G, Guo H, Xu J, Gottfredsson M. Molecular epidemiology of candidemia: Evidence of clusters of smoldering nosocomial infections. Clinical Infectious Diseases. 2008;**47**:e17-e24

[49] Escribano P, Sanchez-Carrillo C, Munoz P, Bouza E, Guinea J. Reduction in percentage of clusters of *Candida albicans* and *Candida parapsilosis* causing candidemia in a general Hospital in Madrid, Spain. Journal of Clinical Microbiology. 2018;**56**(7): e00574-18

[50] Pappas PG, Kauffman CA, Andes DR, Clancy CJ, Marr KA, Ostrosky-Zeichner L, et al. Clinical practice guideline for the management of candidiasis: 2016 update by the Infectious Diseases Society of America. Clinical Infectious Diseases. 2016;**62**: e1-e50

[51] DiazGranados CA, Martinez A, Deaza C, Valderrama S. An outbreak of Candida spp. bloodstream infection in a tertiary care center in Bogota, Colombia. The Brazilian Journal of Infectious Diseases. 2008;**12**:390-394

[52] Li D, Li X, Xia R, Zhang W, Zheng S, Zhang Q, et al. Molecular surveillance of candidemia due to *Candida albicans* among cancer patients during 2009 to 2013 by microsatellite typing. Microbial Pathogenesis. 2015;**81**:28-32

[53] Bogomin F, Gago S, Oladele RO, Denning DW. Global and multinational prevalence of fungal diseases—Estimate precision. Journal of Fungi. 2017;**3**:57

[54] Arendrup MC. Epidemiology of invasive candidiasis. Current Opinion in Critical Care. 2010;**16**:445-452

[55] Nucci M, Queiroz-Telles F, Alvarado-Matute T, Tiraboschi IN, Cortes J, Zurita J, et al. Epidemiology of candidemia in Latin America: A laboratory-based survey. PLoS One. 2013;**8**:e59373

[56] Lockhart SR, Iqbal N, Cleveland AA, Farley MM, Harrison LH, Bolden CB, et al. Species identification and antifungal susceptibility testing of Candida bloodstream isolates from population-based surveillance studies in two U.S. cities from 2008 to 2011. Journal of Clinical Microbiology. 2012; 50:3435-3442

[57] Puig-Asensio M, Padilla B, Garnacho-Montero J, Zaragoza O, Aguado JM, Zaragoza R, et al. Epidemiology and predictive factors for early and late mortality in Candida bloodstream infections: A populationbased surveillance in Spain. Clinical Microbiology and Infection. 2014;**20**: 0245-0254

[58] Wang H, Xu YC, Hsueh PR. Epidemiology of candidemia and antifungal susceptibility in invasive Candida species in the Asia-Pacific region. Future Microbiology. 2016;**11**: 1461-1477

[59] Lortholary O, Renaudat C, Sitbon K, Madec Y, Denoeud-Ndam L, Wolff M, et al. Worrisome trends in incidence and mortality of candidemia in intensive care units (Paris area, 2002–2010). Intensive Care Medicine. 2014;**40**: 1303-1312

[60] Tan BH, Chakrabarti A, Li RY, Patel AK, Watcharananan SP, Liu Z, et al. Incidence and species distribution of candidaemia in Asia: A laboratory-based surveillance study. Clinical Microbiology and Infection. 2015;**21**: 946-953

[61] Lagrou K, Maertens J, Van Even E, Denning DW. Burden of serious fungal infections in Belgium. Mycoses. 2015;**58** (Supp. 5):1-5

[62] Arendrup MC, Fuursted K, Gahrn-Hansen B, Schonheyder HC, Knudsen JD, Jensen IM, et al. Semi-national surveillance of fungaemia in Denmark 2004–2006: Increasing incidence of fungaemia and numbers of isolates with reduced azole susceptibility. Clinical Microbiology and Infection. 2008;**14**: 487-494

[63] Poikonen E, Lyytikainen O, Anttila VJ, Koivula I, Lumio J, Kotilainen P, et al. Secular trend in candidemia and the use of fluconazole in Finland, 2004–2007. BMC Infectious Diseases. 2010;**10**:312

[64] Ruhnke M, Groll AH, Mayser P, Ullmann AJ, Mendling W, Hof H, et al. Estimated burden of fungal infections in Germany. Mycoses. 2015;**58**(Supp. 5): 22-28

[65] Sinko J, Sulyok M, Denning DW. Burden of serious fungal diseases in Hungary. Mycoses. 2015;**58**(Supp. 5): 29-33

[66] Dorgan E, Denning DW, McMullanR. Burden of fungal disease in Ireland.Journal of Medical Microbiology. 2015;64:423-426

[67] Nordoy I, Hesstvedt L, Torp Andersen C, Mylvaganam H, Kols NI, Falch BM, et al. An estimate of the burden of fungal disease in Norway. Journal of Fungi (Basel). 2018;4:E29

[68] Sabino R, Verissimo C, Brandao J, Martins C, Alves D, Pais C, et al. Serious fungal infections in Portugal. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:1345-1352

[69] Mares M, Moroti-Constantinescu VR, Denning DW. The burden of fungal diseases in Romania. Journal of Fungi (Basel). 2018;**4**:E31

[70] Klimko N, Kozlova Y, Khostelidi S, Shadrivova O, Borzova Y, Burygina E, et al. The burden of serious fungal diseases in Russia. Mycoses. 2015;**58** (Supp. 5):58-62

[71] Arsenijevic VA, Denning DW. Estimated burden of serious fungal diseases in Serbia. Journal of Fungi (Basel). 2018;4:E76

[72] Rodriguez-Tudela JL, Alastruey-Izquierdo A, Gago S, Cuenca-Estrella M, Leon C, Miro JM, et al. Burden of serious fungal infections in Spain. Clinical Microbiology and Infection. 2015;**21**: 183-189

[73] Ericsson J, Chryssanthou E, Klingspor L, Johansson AG, Ljungman P, Svensson E, et al. Candidaemia in Sweden: A nationwide prospective observational survey. Clinical Microbiology and Infection. 2013;**19**: E218-E221

[74] Osmanov A, Denning DW. Burden of serious fungal infections in Ukraine. Mycoses. 2015;**58**(Supp. 5):94-100 [75] Dufresne SF, Cole DC, Denning DW, Sheppard DC. Serious fungal infections in Canada. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:987-992

[76] Corzo-Leon DE, Armstrong-James D, Denning DW. Burden of serious fungal infections in Mexico. Mycoses. 2015;**58**(Supp. 5):34-44

[77] Cleveland AA, Harrison LH, Farley MM, Hollick R, Stein B, Chiller TM, et al. Declining incidence of candidemia and the shifting epidemiology of Candida resistance in two US metropolitan areas, 2008–2013: Results from population-based surveillance. PLoS One. 2015;**10**:e0120452

[78] Colombo AL, Nucci M, Park BJ, Nouer SA, Arthington-Skaggs B, da Matta DA, et al. Epidemiology of candidemia in Brazil: A nationwide sentinel surveillance of candidemia in eleven medical centers. Journal of Clinical Microbiology. 2006;**44**: 2816-2823

[79] Colombo AL, Guimaraes T, Sukienik T, Pasqualotto AC, Andreotti R, Queiroz-Telles F, et al. Prognostic factors and historical trends in the epidemiology of candidemia in critically ill patients: An analysis of five multicenter studies sequentially conducted over a 9-year period. Intensive Care Medicine. 2014;**40**: 1489-1498

[80] Riera FO, Caeiro JP, Denning DW. Burden of serious fungal infections in Argentina. Journal of Fungi (Basel). 2018;**4**:E151

[81] Giacomazzi J, Baethgen L, Carneiro LC, Millington MA, Denning DW, Colombo AL, et al. The burden of serious human fungal infections in Brazil. Mycoses. 2016;**59**:145-150

[82] Alvarez Duarte E, Denning DW. Serious fungal infections in Chile. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:983-986

[83] Alvarez-Moreno CA, Cortes JA, Denning DW. Burden of fungal infections in Colombia. Journal of Fungi (Basel). 2018;4

[84] Zurita J, Denning DW, Paz YMA, Solis MB, Arias LM. Serious fungal infections in Ecuador. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:975-981

[85] Medina N, Samayoa B, Lau-Bonilla D, Denning DW, Herrera R, Mercado D, et al. Burden of serious fungal infections in Guatemala. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:965-969

[86] Gugnani HC, Denning DW. Estimated burden of serious fungal infections in Jamaica by literature review and modelling. The West Indian Medical Journal. 2015;**64**:245-249

[87] Bustamante B, Denning DW,Campos PE. Serious fungal infections in Peru. European Journal of Clinical Microbiology & Infectious Diseases.2017;36:943-948

[88] Denning DW, Gugnani HC. Burden of serious fungal infections in Trinidad and Tobago. Mycoses. 2015;**58**(Supp. 5): 80-84

[89] Macedo-Vinas M, Denning DW. Estimating the burden of serious fungal infections in Uruguay. Journal of Fungi (Basel). 2018;4:E37

[90] Gugnani HC, Denning DW, Rahim R, Sadat A, Belal M, Mahbub MS. Burden of serious fungal infections in Bangladesh. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:993-997

[91] Ben R, Denning DW. Estimating the burden of fungal diseases in Israel. The

Israel Medical Association Journal. 2015; 17:374-379

[92] Wadi J, Denning DW. Burden of serious fungal infections in Jordan. Journal of Fungi (Basel). 2018;4:E15

[93] Kemaykin VM, Tabinbaev NB, Khudaibergenova MS, Olifirovich AA, Abdrakhmanova LM, Denning DW, et al. An estimate of severe and chronic fungal diseases in the Republic of Kazakhstan. Journal of Fungi (Basel). 2018;**4**:E34

[94] Huh K, Ha YE, Denning DW, Peck KR. Serious fungal infections in Korea. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:957-963

[95] Velayuthan RD, Samudi C, Lakhbeer Singh HK, Ng KP, Shankar EM, Denning DW. Estimation of the burden of serious human fungal infections in Malaysia. Journal of Fungi (Basel). 2018;4:E38

[96] Jabeen K, Farooqi J, Mirza S, Denning D, Zafar A. Serious fungal infections in Pakistan. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:949-956

[97] Batac MCR, Denning D. Serious fungal infections in the Philippines. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:937-941

[98] Taj-Aldeen SJ, Chandra P, Denning DW. Burden of fungal infections in Qatar. Mycoses. 2015;**58**(Supp. 5):51-57

[99] Chayakulkeeree M, Denning DW. Serious fungal infections in Thailand. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:931-935

[100] Tilavberdiev SA, Denning DW, Klimko NN. Serious fungal diseases in the Republic of Uzbekistan. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:925-929

[101] Xiao M, Sun ZY, Kang M, Guo DW, Liao K, Chen SC, et al. Five-year National Surveillance of invasive candidiasis: Species distribution and azole susceptibility from the China hospital invasive fungal surveillance net (CHIF-NET) study. Journal of Clinical Microbiology. 2018;**56**(7):e00577-18

[102] Chekiri-Talbi M, Denning DW. Burden of fungal infections in Algeria. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:999-1004

[103] Bamba S, Zida A, Sangare I, Cisse M, Denning DW, Hennequin C. Burden of severe fungal infections in Burkina Faso. Journal of Fungi (Basel). 2018;4: E35

[104] Mandengue CE, Denning DW. The burden of serious fungal infections in Cameroon. Journal of Fungi (Basel). 2018;**4**:E44

[105] Zaki SM, Denning DW. Serious fungal infections in Egypt. European Journal of Clinical Microbiology & Infectious Diseases. 2017;**36**:971-974

[106] Kalua K, Zimba B, Denning DW. Estimated burden of serious fungal infections in Malawi. Journal of Fungi (Basel). 2018;4:E61

[107] Sacarlal J, Denning DW. Estimated burden of serious fungal infections in Mozambique. Journal of Fungi (Basel). 2018;4:E75

[108] Faini D, Maokola W, Furrer H, Hatz C, Battegay M, Tanner M, et al. Burden of serious fungal infections in Tanzania. Mycoses. 2015;**58**(Supp. 5): 70-79

[109] Lortholary O, Desnos-Ollivier M, Sitbon K, Fontanet A, Bretagne S, Dromer F, et al. Recent exposure to caspofungin or fluconazole influences the epidemiology of candidemia: A prospective multicenter study involving 2,441 patients. Antimicrobial Agents and Chemotherapy. 2011;**55**:532-538

[110] Maldonado NA, Cano LE, De Bedout C, Arbelaez CA, Roncancio G, Tabares AM, et al. Association of clinical and demographic factors in invasive candidiasis caused by fluconazoleresistant Candida species: A study in 15 hospitals, Medellin, Colombia 2010–2011. Diagnostic Microbiology and Infectious Disease. 2014;**79**: 280-286

[111] Garnacho-Montero J, Diaz-Martin A, Garcia-Cabrera E, Ruiz Perez de Pipaon M, Hernandez-Caballero C, Aznar-Martin J, et al. Risk factors for fluconazole-resistant candidemia. Antimicrobial Agents and Chemotherapy. 2010;**54**:3149-3154

[112] Mellinghoff SC, Panse J, Alakel N, Behre G, Buchheidt D, Christopeit M, et al. Primary prophylaxis of invasive fungal infections in patients with haematological malignancies: 2017 update of the recommendations of the Infectious Diseases Working Party (AGIHO) of the German Society for Haematology and Medical Oncology (DGHO). Annals of Hematology. 2018; **97**:197-207

[113] Leonart LP, Tonin FS, Ferreira VL, Penteado STS, Wiens A, Motta FA, et al. A network meta-analysis of primary prophylaxis for invasive fungal infection in haematological patients. Journal of Clinical Pharmacy and Therapeutics. 2017;**42**:530-538

[114] Satoh K, Makimura K, Hasumi Y, Nishiyama Y, Uchida K, Yamaguchi H. *Candida auris* sp. nov., a novel ascomycetous yeast isolated from the external ear canal of an inpatient in a Japanese hospital. Microbiology and Immunology. 2009;**53**:41-44 [115] Lee WG, Shin JH, Uh Y, Kang MG, Kim SH, Park KH, et al. First three reported cases of nosocomial fungemia caused by *Candida auris*. Journal of Clinical Microbiology. 2011;**49**: 3139-3142

[116] Chowdhary A, Sharma C, Duggal S, Agarwal K, Prakash A, Singh PK, et al. New clonal strain of *Candida auris*, Delhi, India. Emerging Infectious Diseases. 2013;**19**:1670-1673

[117] Magobo RE, Corcoran C, Seetharam S, Govender NP. *Candida auris*-associated candidemia, South Africa. Emerging Infectious Diseases. 2014;**20**:1250-1251

[118] Emara M, Ahmad S, Khan Z,
Joseph L, Al-Obaid I, Purohit P, et al. *Candida auris* candidemia in Kuwait,
2014. Emerging Infectious Diseases.
2015;21:1091-1092

[119] Calvo B, Melo AS, Perozo-Mena A, Hernandez M, Francisco EC, Hagen F, et al. First report of *Candida auris* in America: Clinical and microbiological aspects of 18 episodes of candidemia. The Journal of Infection. 2016;**73**: 369-374

[120] McCarthy M. Hospital transmitted *Candida auris* infections confirmed in the US. BMJ. 2016;**355**:i5978

[121] Mizusawa M, Miller H, Green R, Lee R, Durante M, Perkins R, et al. Can multidrug-resistant *Candida auris* be reliably identified in clinical microbiology laboratories? Journal of Clinical Microbiology. 2017;55:638-640

[122] Arendrup MC, Prakash A, Meletiadis J, Sharma C, Chowdhary A. Comparison of EUCAST and CLSI reference microdilution MICs of eight antifungal compounds for *Candida auris* and associated tentative epidemiological cutoff values. Antimicrobial Agents and Chemotherapy. 2017;**61**(6): e00485-17

[123] Clancy CJ, Nguyen MH. Emergence of *Candida auris*: An international call to arms. Clinical Infectious Diseases. 2017;**64**:141-143

[124] Ruiz Gaitan AC, Moret A, Lopez Hontangas JL, Molina JM, Aleixandre Lopez AI, Cabezas AH, et al. Nosocomial fungemia by *Candida auris*: First four reported cases in continental Europe. Revista Iberoamericana de Micología. 2017;**34**:23-27

[125] Morales-Lopez SE, Parra-Giraldo CM, Ceballos-Garzon A, Martinez HP, Rodriguez GJ, Alvarez-Moreno CA, et al. Invasive infections with multidrug-resistant yeast *Candida auris*, Colombia. Emerging Infectious Diseases. 2017;**23**:162-164

[126] Borman AM, Szekely A, Johnson EM. Comparative pathogenicity of United Kingdom isolates of the emerging pathogen *Candida auris* and other key pathogenic Candida species. mSphere. 2016;**1**:e00189-16

[127] Lockhart SR, Etienne KA, Vallabhaneni S, Farooqi J, Chowdhary A, Govender NP, et al. Simultaneous emergence of multidrug-resistant *Candida auris* on 3 continents confirmed by whole-genome sequencing and epidemiological analyses. Clinical Infectious Diseases. 2017;**64**:134-140

[128] Gonzalez de Molina FJ, Leon C, Ruiz-Santana S, Saavedra P, Group CIS. Assessment of candidemia-attributable mortality in critically ill patients using propensity score matching analysis. Critical Care. 2012;**16**:R105

[129] Jia X, Li C, Cao J, Wu X, Zhang L. Clinical characteristics and predictors of mortality in patients with candidemia: A six-year retrospective study. European Journal of Clinical Microbiology & Infectious Diseases. 2018;**37**:1717-1724

[130] Cortes JA, Reyes P, Gomez CH, Cuervo SI, Rivas P, Casas CA, et al. Clinical and epidemiological characteristics and risk factors for mortality in patients with candidemia in hospitals from Bogota, Colombia. The Brazilian Journal of Infectious Diseases. 2014;**18**:631-637

[131] Barchiesi F, Orsetti E, Mazzanti S, Trave F, Salvi A, Nitti C, et al. Candidemia in the elderly: What does it change? PLoS One. 2017;**12**:e0176576

[132] Horn DL, Ostrosky-Zeichner L, Morris MI, Ullmann AJ, Wu C, Buell DN, et al. Factors related to survival and treatment success in invasive candidiasis or candidemia: A pooled analysis of two large, prospective, micafungin trials. European Journal of Clinical Microbiology & Infectious Diseases. 2010;**29**:223-229

[133] Bassetti M, Righi E, Ansaldi F, Merelli M, Scarparo C, Antonelli M, et al. A multicenter multinational study of abdominal candidiasis: Epidemiology, outcomes and predictors of mortality. Intensive Care Medicine. 2015;**41**: 1601-1610

[134] Raza A, Zafar W, Mahboob A, Nizammudin S, Rashid N, Sultan F. Clinical features and outcomes of Candidaemia in cancer patients: Results from Pakistan. The Journal of the Pakistan Medical Association. 2016;**66**: 584-589

[135] Cornely OA, Gachot B, Akan H, Bassetti M, Uzun O, Kibbler C, et al. Epidemiology and outcome of fungemia in a cancer Cohort of the Infectious Diseases Group (IDG) of the European Organization for Research and Treatment of Cancer (EORTC 65031). Clinical Infectious Diseases. 2015;**61**: 324-331

[136] Andes DR, Safdar N, Baddley JW, Playford G, Reboli AC, Rex JH, et al. Impact of treatment strategy on outcomes in patients with candidemia and other forms of invasive candidiasis: A patient-level quantitative review of randomized trials. Clinical Infectious Diseases. 2012;**54**:1110-1122

[137] Lopez-Cortes LE, Almirante B, Cuenca-Estrella M, Garnacho-Montero J, Padilla B, Puig-Asensio M, et al. Empirical and targeted therapy of candidemia with fluconazole versus echinocandins: A propensity scorederived analysis of a population-based, multicentre prospective cohort. Clinical Microbiology and Infection. 2016; 22(733):e1-e8

[138] Reboli AC, Rotstein C, Pappas PG, Chapman SW, Kett DH, Kumar D, et al. Anidulafungin versus fluconazole for invasive candidiasis. The New England Journal of Medicine. 2007;**356**: 2472-2482

