We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

186,000

200M

Download

154
Countries delivered to

Our authors are among the

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.

For more information visit www.intechopen.com



Utility of Modified Ultrafiltration in Congenital Heart Disease Patients Operated with Cardiopulmonary Bypass

Pedro José Curi-Curi, Juan Calderón-Colmenero, Samuel Ramírez-Marroquín and Jorge Luis Cervantes-Salazar

Additional information is available at the end of the chapter

http://dx.doi.org/10.5772/intechopen.77122

Abstract

Modified ultrafiltration is used in cardiac surgery with cardiopulmonary bypass in order to diminish systemic inflammatory response syndrome. We aimed to show its utility for removing pro-inflammatory agents in operated pediatric patients with congenital heart disease and its impact at operative care. A clinical case-control trial was designed, including patients with simple congenital heart disease operated on with cardiopulmonary bypass in a 1-year period. We randomized them to a problem group (with modified ultrafiltration, n = 15) and a control group (without it, n = 16), and blood samples to measure interleukins (6 and 10); 3d and 4d complement fraction concentrations were taken at the following times: baseline, before cardiopulmonary bypass, after it, after modified ultrafiltration, and from the ultrafiltration concentrate. Operative clinical end points of success were defined as hemodynamic stability, absence of morbidity, and lack of mortality. We observed a higher significant interleukin 6 concentration in the problem group patients at baseline, as well as a higher removal of this pro-inflammatory agent at the ultrafiltration concentrate. Modified ultrafiltration has a positive impact over simple congenital heart disease surgery with cardiopulmonary bypass because of removing interleukin 6. We recommend its routine use when hemodynamic conditions are favorable.

Keywords: cardiopulmonary bypass, congenital heart disease, interleukin



1. Introduction

Cardiopulmonary bypass (CPB) allowed the correction of several congenital heart diseases such as intracardiac malformations, but it is well-known that this is not a harmless procedure because it can lead to a systemic inflammatory response syndrome (SIRS), with activation of complement, cytokines, coagulation, and fibrinolysis pathways. Factors that contribute to the development of SIRS include blood contact with the synthetic surface of cardiopulmonary bypass components, as well as leukocyte and endothelial activation after tissue ischemia and reperfusion [1–5]. If there is a severe inflammatory response, it could also develop a multiorganic dysfunction syndrome that increases morbidity and mortality of the patients at pediatric intensive care units (PICUs). Some of the methods used to quantify the magnitude of SRIS due to the use of CPB include measurement of blood cytokine concentrations (interleukins 1 and 6), complement activation products (C3d and C4d), and also coagulation activated factors (Von Willebrand, fibrinogen and factor VIII) [6].

There are several operative strategies for diminishing SRIS and its clinical repercussion, such as the use of steroids, modified tubular surfaces for CPB, and ultrafiltration. Despite the single or combined use of these strategies [7–12], ultrafiltration is the one that probably removes a larger amount of pro-inflammatory agents, as well as water (volume) [13]. The two ultrafiltration technique modalities widely accepted for pediatric cardiac surgery are conventional ultrafiltration (CUF) and modified ultrafiltration (MUF). CUF is applied in CPB during the heart re-warming period and MUF right after ending CPB.

Currently, there is no enough evidence that favor the routine use of MUF [14–19], and we can still find some controversies regarding the benefits of this technique [20–22]. In addition, most reports of the study are focalized in adult cohorts of patients, and there is few information provided for pediatric population that show the real impact of MUF in the re-motion of proinflammatory agents due to CPB use. Therefore, we aimed to study the real utility of MUF for re-motion of pro-inflammatory agents induced by CPB in operated pediatric patients with simple congenital heart disease. We made a special emphasis in hemodynamic variables, morbidity, and mortality at the operative period.

2. Materials and methods

2.1. Study design

A prospective, randomized, analytic, and clinical case-control trial was designed at the Department of Pediatric Cardiac and Congenital Heart Surgery of a single center during a 1-year period of time. Inclusion criteria were age \leq 18 years, and simple congenital heart disease that required elective surgical treatment with CPB use for at least 30 min. Exclusion criteria were preoperative renal failure, preoperative cardiogenic shock requiring the use of inotropics, preoperative sepsis, and preoperative mechanical ventilatory support of \leq 48 h, preoperative lactate seric levels of \geq 3 mmol/l, and cardiac reoperation. Patients were randomized into two

study groups: problem group (with MUF) and control group (without MUF). With the use of an electronic URNA software, a statistical person randomized the patients and told the perfussionist, which was the only surgical team person informed about the results of randomization. All patients included in this study were operated on with informed consent signed by their parents or tutors. The study was also approved by our institutional research and ethics committee.

2.2. Modified ultrafiltration technique

Patients randomized to problem group (with MUF), when informed to the perfussionist, were prepared for CPB with an additional MUF set. Once CPB was ended and hemodynamic stability of the patient was provided, the surgeon was told not to remove the venous canula, and the venous line was clamped just before its connection to the reservoir. Arterial and venous line pathways were released in order to begin MUF with a 10–20-ml/kg/min flow. MUF continuous flow was achieved, pumping the venous residual reservoir volume by means of the arterial line to the patient. A 150–200-mmHg venous vacuum was applied when needed. MUF lasted 10–20 min in order to reach a desired hematocrit level and obtain also a suitable volume and electrolyte balance. MUF was stopped in case of hemodynamic instability. Once ended, MUF volume was restored to the patient from the hemofilter and venous canula, allowing the surgeon for decanulation of the patient.

2.3. Biochemical and clinical operative analysis

Biochemical and clinical results were compared between the two study groups at the operative period. Biochemical results were the concentration of cytokine (interleukins 6 and 10) and complement activated products (C3d and C4d). These concentrations were measured from blood samples at the following times: T0 (baseline, at the beginning of anesthesia induction), T1 (before CPB), T2 (immediately after CPB), and T3 (immediately after MUF, in the problem group). The same agents were measured in the MUF fluid concentrate of the problem group after the procedure (T4). Clinical operative results were evaluated in terms of hemodynamic instability (>20% post CPB variation with respect to previous CPB values of at least three of the following five hemodynamic variables: heart rate, systolic, diastolic and mean blood pressure, and central venous pressure), operative morbidity and mortality. Operative clinical end points of success were defined as hemodynamic stability, absence of morbidity, and lack of mortality.

2.4. Laboratory analysis of the fluid samples

All patient samples were obtained from central or peripheral blood and collected in tubes without heparin (vacutainer, Beckton Dickinson). A 3-ml blood sample was obtained for each of the study times (T0, T1, T2, and T3). The same volume of T4 samples was obtained from the ultrafiltration fluid concentrate. All of the samples were centrifugated at 3000 rpm for 15 min, 4° C, and cryopreserved in aliquots of 15 ml at -75° C. Interleukin concentrations (IL-6 and IL-10) were measured by means of an ELISA-Sandwich technique with the use of monoclonal antibodies (Peprotech, NJ, EUA). Complement activation products (C3d and C4d) were

measured with the same technique, using commercial kits (Bachem, San Carlos, CA, EUA). Optical density was determined at 450 nm in the ELISA lector. Concentrations of IL-6, IL-10 (pg/ml), as well as C3d and C4d (ng/ml) were calculated by means of a GraphPad Software v. 4.2.

2.5. Statistical analysis

Information was registered in evaluation sheets, stored in an electronic Excel page and analyzed by means of a Prisma Graphics v3.1 statistical software. Continuous variables are presented as a mean, standard deviation, and variability ranges (minimum and maximum). Categorical data are presented by means of frequency and percentages in relation to the population at risk. Comparison between the two study groups was made by means of a Student's t-test for continuous variables. A chi-squared (X²) test was used for comparing categorical variables with a 95% confidence interval (CI). A p-value <0.05 was considered as statistically significant.

3. Results

A total of 31 patients were enrolled and randomized to this trial: 15 to the problem group (with MUF) and 16 to the control group (without MUF).

3.1. Preoperative characteristics

Table 1 shows the type of congenital diseases that were operated by means of CPB in both groups of study. There are no differences in the total number of congenital heart disease in the studied groups, but control group (without MUF) showed more patients with AV channel than the problem group (with MUF).

Congenital heart disease type	Total series (n = 31) n (%)	Problem group (with MUF) (n = 15) n (%)	Control group (without MUF) (n = 16) n (%)	p	
Ventricular septal defect	13 (42%)	8 (52%)	5 (31%)	NS	
Balanced AV channel	8 (26%)	1 (7%)	7 (44%)	0.04	
Congenital mitral valve disease	4 (13%)	3 (20%)	1 (6%)	NS	
Subaortic membrane	3 (10%)	1 (7%)	2 (13%)	NS	
Right ventricular outflow tract obstruction	1 (3%)	1 (7%)	0 (0%)	NS	
Double chamber right ventricle	1 (3%)	1 (7%)	0 (0%)	NS	
Atrial septal defect	1 (3%)	0 (0%)	1 (6%)	NS	
Total	31 (100%)	15 (100%)	16 (100%)	NS	

Table 1. Congenital heart disease type in the studied groups.

Variable	Total series n (%) or mean \pm SD (range)	Problem group (with MUF) n (%) or mean \pm SD (range)	Control group (without MUF) n (%) or mean \pm SD (range)	
Age (years)	$4.26 \pm 4.11 \ (0.38 - 17.18)$	$37 \pm 14 \ (18-76)$	31 ± 11 (18–56)	NS
Gender				
Male	12 (39%)	8 (53%)	4 (25%)	NS
Female	19 (61%)	7 (47%)	12 (75%)	NS
Anthropometric data				
Weight (kg)	$14.9 \pm 10.8 (4-47)$	$14.1 \pm 10.4 (4 – 38.3)$	$15.9 \pm 11.6 \ (5.3-47)$	NS
Height (cm)	$90 \pm 31.1 (12 – 159)$	$94.2 \pm 31.2 (55-158)$	$86 \pm 31.5 (12 – 159)$	NS
Body surface area (m ²)	$0.56 \pm 0.27 \ (0.25 – 1.32)$	$0.58 \pm 0.31 (0.25 – 1.32)$	$0.53 \pm 0.18 \; (0.28 – 0.78)$	NS
Circulating blood volume (ml)	$1032 \pm 627 (343 – 2660)$	$1164 \pm 756 \ (343-2660)$	$867 \pm 385 \ (452 - 1560)$	NS
Cardiovascular background	1			
Previous surgery	0 (0%)	0 (0%)	0 (0%)	NS
Previous catheterization	2 (6%)	0 (0%)	2 (6%)	NS
Pathologic background				
Preoperative infection	1 (3%)	0 (0%)	1 (6%)	NS
Pulmonary artery hypertension	4 (13%)	0 (0%)	4 (25%)	NS
None	26 (84%)	15 (100%)	11 (69%)	NS
Syndromes				
Down	3 (10%)	0 (0%)	3 (19%)	NS
None	28 (90%)	15 (100%)	13 (81%)	NS
NYHA/Ross pre-operative	functional class			
I	8 (26%)	4 (27%)	4 (25%)	NS
II	21 (68%)	9 (60%)	12 (75%)	NS
III	2 (6%)	2 (13%)	0 (0%)	NS
Operative risk				
RACHS-1 score	$2.4 \pm 0.5 \ (1-3)$	2.4 ± 0.5 (2–3)	$2.4 \pm 0.6 \ (1-3)$	NS
Basic aristoteles	$7.2 \pm 1.5 (3-9)$	$7 \pm 1.2 (6-9)$	$7.4 \pm 1.9 (3-9)$	NS
Complete aristoteles	8.1 ± 1.8 (4–11)	$7.8 \pm 1.5 \ (6-10)$	8.4 ± 2.1 (4–11)	NS
Preoperative morbidity				
Mechanic ventilation	0 (0%)	0 (0%)	0 (0%)	NS
Preoperative inotropic support	0 (0%)	0 (0%)	0 (0%)	NS
Preoperative infection	1 (3%)	0 (0%)	1(6%)	NS
None	30 (97%)	15 (100%)	15 (94%)	NS
Preoperative laboratory exa	ams			
Lactate	$1.2 \pm 0.3 \; (0.6 – 1.7)$	$1.2 \pm 0.3 \; (0.7 – 1.7)$	$1.1 \pm 0.3 \; (0.6 – 1.5)$	NS

Variable	Total series n (%) or mean \pm SD (range)	Problem group (with MUF) n (%) or mean \pm SD (range)	Control group (without MUF) n (%) or mean \pm SD (range)	
Creatinine	$0.4 \pm 0.1 \; (0.2 – 0.7)$	$0.4 \pm 0.1 \ (0.2 – 0.7)$	$0.4 \pm 0.1 (0.3 – 0.5)$	NS
Perfusion variables				
Oxigenator type				
Baby Rx	14 (52%)	7 (47%)	7 (58%)	NS
Terumo SX10	6 (22%)	4 (27%)	2 (17%)	NS
Terumo SX18	1 (4%)	1 (7%)	0 (0%)	NS
Mini max	5 (19%)	2 (13%)	3 (25%)	NS
Safe Mini	1 (4%)	1 (7%)	0 (0%)	NS
Arterial filter use	18 (67%)	12 (80%)	6 (50%)	NS
Surgical variables				
CPB time (min)	$81.9 \pm 26.9 \ (40 – 131)$	$76.5 \pm 23.7 \ (40122)$	$87 \pm 29.4 (41131)$	NS
Aortic cross clamp time (min)	$53.7 \pm 23.6 \ (12-96)$	$49.5 \pm 21.8 \ (18-90)$	$57.6 \pm 25.2 \ (12-96)$	NS
Temperature (°C)	$27 \pm 1.6 \ (24 – 30)$	$27 \pm 1.5 \ (24-29)$	$27.3 \pm 1.8 \ (24-30)$	NS
Anterograde cardioplegia	29 (94%)	14 (93%)	15 (94%)	NS
Blood cardioplegia	29 (94%)	14 (93%)	15 (94%)	NS

Table 2. Preoperative characteristics of the studied groups.

Table 2 shows the rest of preoperative characteristics in both studied groups. Note that there are no statistical differences in all variables analyzed between the two groups.

Although more random patients with AV channel in the control group, the rest of the preoperative data showed that both groups are absolutely comparable.

3.2. Biochemical operative results

Table 3 compares the concentration of pro-inflammatory agents between groups before surgical correction (T0). Note a baseline elevated concentration of IL-6 in the problem group (with

Pro-inflammatory agent	T0 Problem group (with MUF) n = 15 Mean \pm DE	T0 Control group (without MUF) n = 16 Mean ± DE	p
C3d (ng/ml)	368.66 ± 331.87	413.248 ± 316.804	NS
C4d (ng/ml)	199.57 ± 201.56	213.89 ± 116.72	NS
IL-6 (pg/ml)	672.249 ± 433.186	246.874 ± 365.69	0.0061
IL-10 (pg/ml)	239.698 ± 381.517	299.618 ± 370.148	NS

The words and numbers in "bold" highlight the variables that have a statistical significance (p<0.005).

Table 3. Comparison between concentrations of pro-inflammatory agents in both groups of study (with and without MUF) at baseline (T0).

MUF), without differences in both groups for the rest of pro-inflammatory agents (IL-10, C3d, and C4d).

On the other hand, **Table 4** shows a lack of statistically significant difference in the concentrations of pro-inflammatory agents at the control group before surgical correction (T0) and after CPB (T2).

Finally, **Table 5** shows the comparison between the concentration of pro-inflammatory agents in the problem group before surgical correction (T0) and after MUF (T4). There is a statistically significant removal of IL-6, but no difference in the concentrations of the rest of pro-inflammatory agents analyzed (IL-10, C3d, and C4d).

3.3. Clinical operative results

Table 6 summarizes the comparison of clinical end point variables in both groups of study (with and without MUF). There is a statistically significant decrease of hemoglobin (Hb) in the problem group after MUF compared with the baseline level, which is not observed in the control group.

Both groups show an increase in lactate levels and heart rate after surgery when comparing these values with the baseline ones before CPB. Control group (without MUF) showed a statistically significant increase in the central venous pressure after CPB compared with the ones before CPB. There were no differences before and after CPB in the other hemodynamic variables (systolic, diastolic, and mean blood pressures), nor in operative morbidity and mortality. Successful clinical operative endpoints were achieved in both groups of study.

Pro-inflammatory agent	T0 Group control (sin UFM) n = 16 Media \pm SD	T2 Control group (without MUF) n = 16 Media \pm DE	p
C3d (ng/ml)	413.248 ± 316.804	264.33 ± 198.12	NS
C4d (ng/ml)	213.89 ± 116.72	210.65 ± 141.13	NS
IL-6 (pg/ml)	246.874 ± 365.69	289.499 ± 301.913	NS
IL-10 (pg/ml)	299.618 ± 370.148	387.26 ± 306.07	NS

Table 4. Comparison between concentrations of pro-inflammatory agents at T0 (baseline) and T2 (after CPB) for the control group (without MUF).

Pro-inflammatory agent	T0 Grupo problema (con UFM) n = 15 Media ± SD	T4 Problem group (with MUF) n = 15 Media \pm DE	p
C3d (ng/ml)	368.66 ± 331.87	379.99 ± 264.64	NS
C4d (ng/ml)	199.57 ± 201.56	172.89 ± 139.64	NS
IL-6 (pg/ml)	672.249 ± 433.186	366.31 ± 280.25	0.0293
IL-10 (pg/ml)	239.698 ± 381.517	230.453 ± 352.27	NS

The words and numbers in "bold" highlight the variables that have a statistical significance (p<0.005).

Table 5. Comparison between concentrations of pro-inflammatory agents at baseline (T0) and after MUF (T4) for the problem group (with MUF).

	Problem group (with MUF)			Control group (without MUF)			Problem versus control groups (with vs. without MUF)		
Operative clinical end point	Control group	Problem group	p	Control group	Problem group	p	Problem group	Control group	p
variable	Before CPB	After MUF	_	Before CPB	After MUF	_	After MUF	After CPB	_
	n/total n (%) or	n/total n (%) or	_	n/total n (%) or	n/total n (%) or	_	n/total n (%) or	n/total n (%) or	_
	Mean \pm SD	Mean \pm SD	_	$\overline{\text{Mean} \pm \text{SD}}$	Mean \pm SD	_	Mean ± SD	Mean \pm SD	_
Laboratory examinations									
Hematocrit (%)	38 ± 7	34 ± 6	NS	37 ± 5	34 ± 7	NS	34 ± 6	34 ± 7	NS
Hemoglobin (g/dl)	14 ± 5	$\textbf{11} \pm \textbf{2}$	0.0344	12 ± 2	11 ± 2	NS	11 ± 2	11 ± 2	NS
CPB hematocrit (%)						$26 \pm 5*$	$24\pm4^*$	NS	
Lactate (mmol/l) 1.2 ± 0.3	3.5 ± 1.4	0.0001	$\textbf{1.1} \pm \textbf{0.3}$	$\textbf{3.3} \pm \textbf{1.2}$	0.0001	3.5 ± 1.4	3.3 ± 1.2	NS	
Hemodynamic variables									
Heart rate (beats per minute)	97 ± 15	$\textbf{113} \pm \textbf{18}$	0.012	97 ± 16	$\textbf{112} \pm \textbf{15}$	0.0116	113 ± 18	112 ± 15	NS
Systolic blood pressure (mmHg)	85 ± 16	89 ± 12	NS	83 ± 10	90 ± 20	NS	89 ± 12	90 ± 20	NS
Diastolic blood pressure (mmHg)	53 ± 15	52 ± 12	NS	49 ± 7	49 ± 12	NS	52 ± 12	49 ± 12	NS
Mean blood pressure (mmHg)	64 ± 18	61 ± 12	NS	64 ± 13	64 ± 17	NS	61 ± 12	64 ± 17	NS
Central venous pressure (mmHg)	10 ± 8	12 ± 7	NS	8 ± 1	10 ± 3	0.0203	12 ± 7	10 ± 3	NS
Operative morbidity and mortality									
Morbidity						3 (20%)	1 (6%)		NS
Mortality						0 (0%)	0 (0%)		NS

Table 6. Comparison between operative clinical end point variables in both groups of study (with and without MUF).

Shades: The words and numbers in "bold" highlight the variables that have a statistical significance (p<0.005).

*CPB measured values (due to hemodilution).

4. Discussion

Cardiopulmonary bypass (CPB) is able to trigger a systemic inflammatory response syndrome (SRIS) due to several factors that include (1) cell activation secondary to contact with CPB synthetic surfaces, (2) mechanic stress, (3) tissue ischemia and reperfusion, (4) hypotension, (5) non-pulsatile flow, (6) hemodilution relative anemia, (7) blood and blood products transfusion, (8) heparin and protamine administration, and (9) hypothermic effects. CPB activates the vessels endothelium and releases pro-inflammatory agents such as tumoral necrosis factor α (TNF- α), interleukins, and endotoxins. These agents activate the intracellular transcription factor as well, which increases endothelial pro-inflammatory cytokines and the molecular expression of leukocyte adhesion.

It is a well-known fact that younger age increases the inflammatory effects of CPB even more. Some reasons include an increased metabolic demand in these patients, hyperactivity of their pulmonary vessels, immaturity of their organs/systems, and altered homeostasis. Risk is particularly high in neonates and young infants due to a mismatch between CPB and patient's size, with CPB circuit volume usually 200–300% higher than that of the patient. In addition, an increased metabolic demand requires elevated pump flow up to 200 ml/kg/min in neonates. Combining a relative major size of CPB with an increased perfusion rate leads to a greater blood exposure to synthetic surfaces of the circuit components [23]. In our series, there was no age difference between the studied groups, and it is important to highlight that none of the groups included neonate patients for the reasons already discussed.

One of the most involved cytokines in SRIS development is, indeed, IL-6. Increased concentrations of IL-6 have been reported in patients with postoperative complications and a correlation with the posterior left ventricular wall dyskinesia detected by means of transesophageal echocardiography has been established. IL-6 is also an endogenous pyrogen agent that activates acute phase reactant proteins. Concentration of IL-6 increases independently of the oxygenator type, degree of hypothermia, or heparin use in the CPB circuit surfaces [24, 25]. Although in our study IL-6 concentrations were significantly higher before surgery in the problem group than in the control group, this agent is also the one that is significantly more removed by MUF. This is probably the most relevant fact of our study because it shows that the benefit of MUF in congenital heart disease surgery is the removal of IL-6, an important proinflammatory agent, particularly in patients that SRIS is enhanced because of the immaturity of their immune system. Another effect that is important to discuss is the fact that if MUF benefits patients with simple congenital heart disease surgery as were the ones included in our study, it would indeed improve operative outcomes in those operated on for complex congenital heart disease [26]. This single fact justifies the routine use of MUF in all patients with congenital heart disease that are operated on with CPB.

There are several additional methods, despite ultrafiltration, that had been developed in order to diminish SRIS secondary to CPB at surgical correction of congenital heart disease in pediatric population. Some of them are steroids (e.g., dexamethasone 10–30 mg/kg, 6–12 h before CPB), and modified tubular synthetic surfaces in the CPB circuit. However, none of these methods are as useful for this purpose as MUF, which is established right after ending the CPB and before

decanulation of the patient [27]. Since 1973, different types of hemofilters have been developed in order to remove priming volume (water) following the principle of pressure gradient, particularly those made of polycarbonate. These filters have been replaced by the ones made out of poliarileter-sulfonate in 1986, and later by the current generation of polyamide hemofilters. These are the most practical ones because of its greater biocompatibility, reduced surface, and more ultrafiltration effectiveness due to a less than physiological pressure.

The effectiveness of ultrafiltration for removing pro-inflammatory agents depends also on the type of hemofilter and on the modality of ultrafiltration procedure used. Berdat et al. studied the effectiveness of poliariletersulfonate filters versus polyamide ones in the two ultrafiltration modalities for the removal of pro-inflammatory agents such as IL-6, IL-10, and $\text{TNF}\alpha$ [10]. They prove that IL-6 was better removed by conventional ultrafiltration (CUF) with poliarileter-sulfonate filter, while $\text{TNF}\alpha$ was better removed by modified ultrafiltration (MUF) and poliarileter-sulfonate filter. The rest of the pro-inflammatory agents were not modified neither for the ultrafiltration modality nor for the hemofilter type. Therefore, it seems that MUF with poliarileter-sulfonate hemofilter is the better strategy for removing pro-inflammatory agents in pediatric patients with congenital heart surgery. Our results are based on the ultrafiltration modality rather than the type of filter, since the material of hemofilters that we used was variable.

It has been reported that MUF is not only useful for removing extracellular fluid excess but also cytokines and other inflammatory agents triggered by CPB and surgical trauma. There is some controversy in the study regarding the efficacy of filters in the removal of cytokines, as well as in the differences between the two ultrafiltration modalities [28]. In addition, the comparative results between both ultrafiltration modalities are difficult to interpret due to variations in the ultrafiltration technique, equipment, definitions and objectives, and measurements of cytokines. Finally, it is still not known if the clinical benefits of MUF are due to the removal of cytokines and other inflammatory agents, or to the isolated reduction of tissue edema [29–33].

5. Conclusion

Based on the results of this study [34], we can say that although the baseline concentrations of IL-6 in the patients of the problem group were higher in relation to those of the control group, the removal of this pro-inflammatory agent by MUF was statistically significant. This indicates that MUF is a procedure that can benefit pediatric patients with congenital heart disease undergoing CPB because it is able to decrease the concentration of IL-6. Therefore, we consider that the use of MUF in pediatric patients should be routinely recommended as long as hemodynamic conditions allow it.

Acknowledgements

We thank the Cardio Slim Foundation for the financial support provided to carry out this study.

Conflict of interest

The authors declare no potential conflicts of interest with respect to the research, authorship, or publication of this manuscript.

Author details

Pedro José Curi-Curi^{1*}, Juan Calderón-Colmenero², Samuel Ramírez-Marroquín¹ and Jorge Luis Cervantes-Salazar¹

- *Address all correspondence to: pcuricuri001@gmail.com
- 1 Department of Congenital Heart Disease and Pediatric Cardiac Surgery, "Ignacio Chávez" National Cardiology Institute, Mexico City, Mexico
- 2 Department of Pediatric Cardiology, "Ignacio Chávez" National Cardiology Institute, Mexico City, Mexico

References

- [1] Brix-Christensen V. The systemic inflammatory response after cardiac surgery with cardiopulmonary bypass in children. Acta Anaesthesiologica Scandinavica. 2001;45:671-679
- [2] Seghaye MC. The clinical implications of the systemic inflammatory reaction related to cardiac operations in children. Cardiology in the Young. 2003;13:228-239
- [3] Kozik D, Tweddell J. Characterizing the inflammatory response to cardiopulmonary bypass in children. The Annals of Thoracic Surgery. 2006;81:S2347-S2354
- [4] Kirklin JK, Westaby S, Blackstone EH, Kirklin JW, Chenowelh DE, Pacifico AD. Complement and the damaging effects of cardiopulmonary bypass. The Journal of Thoracic and Cardiovascular Surgery. 1983;86:845-847
- [5] Seghaye M, Duchateau J, Grabitz RG, Nitsch G, Marcus C, Messmer BJ, von Bernuth G. Complement, leukocytes and leukocyte elastase in full-term neonates undergoing cardiac operation. Journal of Thoracic and Cardiovascular Surgery. 1994;108:29-36
- [6] Seghaye M, Grabitz RG, Duchateau J, Bussea S, Däbritz S, Koch D, et al. Inflammatory reaction and capillary leak syndrome related to cardiopulmonary bypass in neonates undergoing cardiac operations. The Journal of Thoracic and Cardiovascular Surgery. 1996;112:687-697
- [7] Ashraf SS, Tian Y, Zacharrias S, Cowan D, Martin P, Watterson K. Effects of cardiopulmonary bypass on neonatal and paediatric inflammatory profiles. European Journal of Cardio-Thoracic Surgery. 1997;12:862-868

- [8] McBride WT, McBride SJ. The balance of pro- and anti-inflammatory cytokines in cardiac surgery. Current Opinion in Anesthesiology. 1998;11:15-22
- [9] McBride WT, Armstrong MA, Gilliland H, McMurray TJ. The balance of pro- and antiinflammatory cytokines in plasma and bronchoalveolar lavage (BAL) at paediatric cardiac surgery. Cytokine. 1996;8:724-729
- [10] Chevv M, Brandslund I, Brix-Christensen V, et al. Tissue injury and the inflammatory response to pediatric cardiac surgery with cardiopulmonary bypass. Anesthesiology. 2001;94:745-753
- [11] Finn A, Moat N, Rebuck N, Klein N, Slrobel S, Elliott M. Changes in neutrophil CDllb/CD18 and L-selectin expression and release of interleukin 8 and elastase in paediatric cardiopulmonay bypass. Agents and Actions. 1993;38:C44-C46
- [12] Brix-Christensen V, Petersen TK, Ravn HB, Hjortdal VE, Andersen NR, Tonnesen E. Cardiopulmonary bypass elicits a pro- and anti-inflammatory response and impaired chemotaxis in neonatal pigs. Acta Anaesthesiologica Scandinavica. 2001;45:407-413
- [13] Hennein HA, Kiseltepe U, Barst S, Bocchieri KA, Remick DG, et al. Venovenous modified ultrafiltration after cardiopulmonary bypass in children: A prospective randomized study. The Journal of Thoracic and Cardiovascular Surgery. 1999;117:496-505
- [14] Kern FH, Morana NJ, Sears JJ, Hickey PR. Coagulation defects in neonates during cardio-pulmonary bypass. The Annals of Thoracic Surgery. 1992;54:541-546
- [15] Boga M, Islamoglu F, Badak I, Cikirikcioglu M, Bakalim T, Yagdi T. The effects of modified hemofiltration on inflammatory mediators and cardiac performance in coronary artery bypass grafting. Perfusion. 2000;**15**:143-150
- [16] Tassani P, Richter JA, Eising GP, Barabkay A, Braun SL, Haehnel CH, et al. Influence of combined zero-balanced and modified ultrafiltration on the systemic inflammatory response during coronary artery bypass grafting. Journal of Cardiothoracicand Vascular Anesthesia. 1999;13:285-291
- [17] Pearl JM, Manning PB, McNamara JL, Saucier MM, Thomas DW. Effect of modified ultrafiltration on plasma thromboxane B2, leukotriene B4, endothelin-1 in infants undergoing cardiopulmonary bypass. The Annals of Thoracic Surgery. 1999;68:1369-1375
- [18] Grunenfelder J, Sund G, Schoeberlein A, Maly FE, Guntli S, Fischer K, et al. Modified ultrafiltration lowers adhesion molecule and cytokine levels after cardiopulmonary bypass without clinical relevance in adults. European Journal of Cardio-Thoracic Surgery. 2000;17:77-83
- [19] Chew MS. Does modified ultrafiltration reduce the systemic inflammatory response to cardiac surgery with cardiopulmonary bypass? Perfusion. 2004;19:S57-S60
- [20] Hauser GJ, Ben-Ari J, Colvin MP, Dalton HJ, Hertzog JH, Bearb M, et al. Interleukin-6 levels in serum and lung lavage fluid of children undergoing open heart surgery correlate with postoperative morbidity. Intensive Care Medicine. 1998;24:481-486

- [21] Gilliland HE, Armstrong MA, McMurray TJ. The inflammatory response to pediatric cardiac surgery: Correlation of granulocyte adhesion molecule expression with postoperative oxygenation. Anesthesia and Analgesia. 1999;89:1188-1191
- [22] Andreasson S, Góthberg S, Berggren H, Bengtsson A, Eriksson E, Risberg B. Hemofiltration modifies complement activation after extracorporeal circulation in infants. The Annals of Thoracic Surgery. 1993;56:1515-1517
- [23] Wang W, Chiu I, Chao-Min W, Pei-Lin L, Huand HM, Chung IC, et al. Modified ultrafiltration in pediatric cardiopulmonary bypass. Perfusion. 1998;13:304-310
- [24] Ming-Jiuh W, Chiu I-S, Chao-Ming W, Pei-Lin L, Chung-I C, Chi-Hsiang H, Shu-Hsun C. Efficacy of ultrafiltration in removing inflammatory mediators during pediatric cardiac operations. The Annals of Thoracic Surgery. 1996;61:651-656
- [25] Naik S, Knight A, Elliott MJ. A prospective randomized study of a modified technique of ultrafiltration during pediatric open-heart surgery. Circulation. 1991;84(suppl III):422-431
- [26] Bando K, Turrentine MW, Vijay P, Sharp TG, Lalone BJ, et al. Effect of modified ultrafiltration in high-risk patients undergoing operations for congenital heart disease. The Annals of Thoracic Surgery. 1998;66:821-828
- [27] Draasima AM, Hazekamp MG, Frank M, Anes N, Schoof PH, Huysmans HA. Modified ultrafiltration after cardiopulmonary bypass in pediatric cardiac surgery. The Annals of Thoracic Surgery. 1997;64:521-525
- [28] Daggett CW, Lodge AJ, Scarborough JE, Chai PJ, Jaggers J, Ungerleider RM. Modified ultrafiltration versus conventional ultrafiltration: A randomized prospective study in neonatal piglets. The Journal of Thoracic and Cardiovascular Surgery. 1998;H5:336-342
- [29] Davies MJ, Nguyen K, Gaynor JW, Elliott MJ. Modified ultrafiltration improves left ventricular systolic function in infants after cardiopulmonary bypass. The Journal of Thoracic and Cardiovascular Surgery. 1998;5:361-370
- [30] Journois D, Israel-Biet D, Pouard P, Rolland B, Silvester W, Vouhe P, Safran D, et al. High-volume, zero-balanced hemofiltration to reduce delayed inflammatory response to cardiopulmonary bypass in children. Anesthesiology. 1996;85:965-976
- [31] Chew MS, Brix-Christensen V, Ravn H, Brandslund I, Ditlevsen E, Pedersen J, et al. Effect of modified ultrafiltration on the inflammatory response in paediatric open-heart surgery: A prospective, randomized study. Perfusion. 2002;17:327-333
- [32] Ramamoorthy C, Lynn AM. The use of modified ultrafiltration during pediatric cardiovascular surgery is not a benefit. Journal of Cardiothoracic and Vascular Anesthesia. 1998; 12(4):483-485
- [33] García-Montes JA, Calderón-Colmenero J, Juanico A. Cuidados Intensivos en el niño cardiópata en Attie F. In: Calderón-Colmenero J, Zabal C, Buendía A, editors. Cardiología Pediátrica. México DF: Editorial Médica Panamericana; 2013. p. 203

[34] Curi-Curi PJ, del Villar MRS, Gómez-García L, Vergara BG, Calderón-Colmenero J, Ramírez-Marroquín S, Cervantes-Salazar JL. Impacto intraoperatorio de la ultrafiltración modificada en pacientes pediátricos sometidos a cirugía cardíaca con circulación extracorpórea. Cirugía Cardiovascular. 2016;**23**(4):179-186

IntechOpen

IntechOpen