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Autologous Chondrocyte Implantation: Scaffold-Based Solutions

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Abstract

Autologous chondrocyte implantation is a surgical technique utilized for repair of articular cartilage defects. The originally described technique for autologous chondrocyte implantation involves applying a liquid suspension of the cultured chondrocytes to a cartilage defect and sealing the defect with a periosteum or collagen patch. Scaffolds for housing chondrocytes were introduced to allow for increased ease of delivery and application, to avoid leakage of chondrocytes out of the defect, and to allow for an implant that more closely mimics the non-uniform tissue architecture of healthy articular cartilage. In this chapter we describe the design, clinical outcomes, and commercial availability of various scaffolds reported in the clinical literature for autologous chondrocyte implantation.

Keywords: scaffold, MACI, MACT, autologous chondrocyte implantation, 3rd generation ACI

1. Introduction

Autologous chondrocyte implantation (ACI) is a two-stage articular cartilage repair technique for treatment of articular cartilage defects. Originally described by Brittberg et al. [1], it involves an initial surgery to harvest chondrocytes from a non-weight bearing portion of the distal femur, typically the intercondylar notch or medial or lateral margin of the trochlea. The cartilage extracellular matrix is then enzymatically digested within the laboratory to isolate the chondrocytes. The harvested chondrocytes are then cultured in a laboratory. In the second stage, a liquid suspension of chondrocytes is applied to the cartilage defect and is sealed in place with a soft tissue membrane cover [1]. Originally periosteum was utilized as the cover, though a collagen membrane was later introduced to minimize periosteal donor site morbidity and risk of periosteal



patch hypertrophy [2]. Disadvantages of ACI with periosteum or collagen membrane covers with the use of a liquid cultured chondrocyte suspension include a high degree of technical difficulty, potential for leakage of chondrocytes, and non-uniform distribution of chondrocytes.

Scaffolds for housing chondrocytes were introduced for increased ease of delivery and application, to avoid leakage of chondrocytes out of the defect, and to allow homogeneous distribution of chondrocytes within the defect [3]. Additionally, there is some evidence that chondrocytes grown in monolayer culture do not fully regain their original phenotype [3, 4], which has prompted research in culture directly within a scaffold and design of implants that more closely mimics the non-uniform tissue architecture of healthy articular cartilage [3]. Use of a 3-dimensional structure for chondrocyte culture has been shown to maintain the chondrocyte differentiated phenotype [5]. Use of a scaffold is termed 'matrix-assisted autologous chondrocyte transplantation,' or the MACT procedure, and has been employed in clinical practice in Europe since 1998. The MACT procedure involves implantation of a chondrocyte seeded biocompatible scaffold in the articular defect [2]. The implant is fixed in place with fibrin glue with no membrane cover and allows for implantation with use of a mini-arthrotomy or arthroscopic implantation. The field of scaffold-based ACI has greatly expanded in recent years, with more than a dozen implants developed (Table 1). A wide variety of natural and synthetic materials have been utilized in MACT scaffolds; though clinical outcomes studies are generally favorable regardless of scaffold design, the number or published studies and length of follow-up vary widely among implants.

In this chapter, the design rationale, commercial availability, and clinical results of various scaffolds for use in MACT will be described. Of note, all implants described in this chapter follow a two-step implantation protocol (initial cartilage harvest and culturing of chondrocytes followed by a delayed implantation several weeks later). The single-stage implantation techniques with published outcomes data are either no longer commercially available (the CAIS implant) [6], or have yet to be marketed [7].

Scaffold content	Commercial name	Implantation steps
Porcine collagen I/III membrane	MACI	Two-steps
Three-dimensional collagen I based scaffold	NeoCart	Two-steps
Three-dimensional collagen I based scaffold	CaReS	Two-steps
Three-dimensional collagen I based scaffold	Novocart 3D	Two-steps
Hyaluronic acid based scaffold	Hyalograft C	Two-steps
Human fibrin and recombinant hyaluronic acid-based scaffold	BioCart II	Two-steps
Fibrin based gel	Chondron	Two-steps
Hydrogel of agarose and alginate	Cartipatch	Two-steps
Atelocollagen gel	Koken Atelocollagen Implant	Two-steps
Fibrin, polyglycolic/polylactic acid, polydioxanone	BioSeed-C	Two-steps

Table 1. Summary of MACT scaffolds.

2. Scaffolds utilized for autologous chondrocyte implantation

2.1. Porcine collagen I/III membranes

2.1.1. MACI

As of December 2016, matrix-assisted chondrocyte implantation (MACI; Vericel, Cambridge, MA) is currently the only FDA approved MACT technique for use in the United States. In this technique, chondrocytes are cultured ex-vivo in a monolayer and then seeded on one side of a porcine collagen I/III membrane (Table 2). At the second stage operation (reimplantation), the side seeded with chondrocytes (the roughened side) is placed against the subchondral bone surface and the graft is secured with fibrin glue [8]. The implantation may be performed arthroscopically or with a mini-arthrotomy, and recent work demonstrates MACI grafts may be safely applied with use of carbon dioxide insufflation arthroscopy [9]. Regardless of technique, gentle handling of the graft is recommended, as excessive or forceful handling of the graft causes a significant decrease in viable chondrocytes [10]. A histologic study of 56 MACI patients up to 6 months after surgery demonstrated that chondrocytes appeared well-integrated and maintained chondrocyte phenotype [11]. Hyalinelike cartilage production began as early as 21 days after implantation, and there was 75% hyaline-like cartilage regeneration at 6 months [11]. Another histologic study of 33 secondlook biopsies at median 15 months after surgery found a median ICRS histological grade of 57 which did not correlate with an arthroscopic ICRS grade of normal in 30% of cases and nearly normal in 51% of cases [12].

Several comparative studies have been performed with MACI, all of which demonstrated encouraging results (Table 3). However, it should be noted that use of MACI in clinical practice tends to be in larger defects (mean 5.64 cm²) than lesions treated in clinical trials (weighted mean 4.89 cm²) [13]. Approval by the FDA was based primarily on results of the SUMMIT trial, reported by Saris et al. [14]. In this randomized trial, 144 patients with high grade femoral condylar defects were randomized to MACI or microfracture and followed for 2 years; mean defect size was equivalent between groups (4.9 cm² MACI vs. 4.7 cm² microfracture) [14]. At final follow-up there was significantly better improvement in KOOS symptom scores with MACI, lower failure rates, yet no difference in repair quality as assessed by histology or MRI versus microfracture [14]. A randomized controlled trial was performed by Bartlett et al. with comparison of ACI-C (ACI with collagen cover) and MACI for treatment

Commercial name	Manufacturer	Structure	Expansion	Availability
MACI	Vericel, Cambridge, MA (Formerly provided by Verigen Transplantation Service, Copenhagen, Denmark)	Porcine- derived collagen I/III bilayer	Cells are expanded in monolayer then seeded onto porous side of collagen membrane	FDA approved for use in the USA. Available in Europe and Australia
FDA, U.S. Foo	od and Drug Administration.			

Table 2. MACT with porcine collagen I/III membrane scaffold.

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
1	Ebert et al. [20]	19 MACI-standard WB; 18 MACI- accelerated WB	2 years	Randomized trial of standard 8 week return to weight bearing versus accelerated 6 week return to weight bearing. No difference in symptom improvement.
1	Wondrasch et al. [21]	15 MACI-standard WB; 16 MACI- accelerate WB	5 years	Randomized trial of 6 versus 10 week return to weight bearing. No difference in symptom improvement between groups. MOCART score decreased from years 2 to 5 which did not correlate with symptom scores
1	Akgun et al. [18]	7 MACI; seven mesenchymal stem cell	2 years	Small randomized trial of MACI versus stem cells (also seeded onto a collagen scaffold. Stem cell group had greater symptom improvement at 6 months but similar improvement at final follow-up.
1	Basad et al. [15]	40 MACI; 20 microfracture	2 years	At 24 months, greater improvements seen with MACI in Tegner activity score, subjective symptoms scores and ICRS scores on 2nd look arthroscopy.
1	Saris et al. [14]	72 MACI; 72 microfracture	2 years	Greater improvement in KOOS scores, lower failure rate with MACI (12.5%) versus microfracture (31.9%). Similar MRI and histologic outcomes.

Table 3. Outcomes of MACT with collagen I/III membrane scaffold (MACI) from level 1 prospective clinical studies.

of high grade chondral defects. Mean defect size was 6.0 cm² for the MACI group and 6.1 cm² for the ACI-C group [8]. At 1 year follow-up both groups demonstrated significant improvement in Cincinnati knee scores and similar re-operation rates (9% for both groups) [8]. Basad et al. performed a randomized study of MACI versus microfracture with 2 years follow-up on high grade defects 4–10 cm² [15]. The MACI group in this study had greater improvements in symptom scores, activity scores, and ICRS surgeon grading of cartilage appearance at second look arthroscopy [15]. In a comparative imaging and clinical study of MACI versus osteochondral autograft transfer (OAT) by Salzmann et al., superior Lysholm symptoms scores were observed in the MACI group; patients in this study were matched for demographics, but MACI-treated lesions were >3 cm² and OAT-treated lesions were <3 cm² [16]. For treatment of chondromalacia patella, Macmull et al. noted a higher rate of good-excellent patient symptom scores with MACI (56.5%) than ACI-C (40%). Higher rates of clinical failure (poor patient-rated symptoms) were noted with lateral facet lesions, and the authors did not report distribution of lesions (medial facet, lateral facet, or multiple facets) by treatment group [17]. Finally, Akgun et al. report a small randomized trial of MACI versus autologous mesenchymal stem cells (also seeded onto a collagen scaffold) with 2 years follow-up [18]. The stem cell group had greater symptom improvement at 6 months but similar improvement at final follow-up; no clinical failures were noted in either group [18].

Several randomized trials of delayed versus accelerated weight-bearing after MACI have been performed (Table 3). A randomized trial of 6 week versus 8 week return to full weight bearing found no significant difference in failure rates or symptom improvement at 2 years (interim 12-month results reported in an earlier publication [19]); the study authors concluded accelerated weight bearing after MACI is safe [20]. Another trial of 6 week versus 10 week return to full weight bearing with 5 years follow-up after MACI similarly found no difference in symptom improvement between groups [21]. The authors note that MRI-based MOCART scores decreased from years 2 to 5 but did not correlate with symptom scores [21].

Several case series have reported also reported good results with MACI (Table 3). The series with the longest follow-up is reported by Gille et al.; of 19 cases with mean 16 years follow-up, 21% underwent knee arthroplasty (4/19), with durable symptom improvement in the remaining 15 patients [22]. In another series of MACI patients, Basad et al. report durable improvements in activity and symptoms scores and a failure rate of 18.5% at 5 years with MACI [23]. Behrens et al. similarly report 8/11 patients rated their current knee function as 'much better or better' than their pre-operative function at 5 years follow-up [24]. A larger case series by Ebert et al. of 41 patients and 5 years follow up (35/41, 85% with 5 years follow-up) reported significant improvements in knee function, a 12% rate of graft hypertrophy at 5 years, and a graft failure rate of 3% at 5 years [25]. Durable results are seen with arthroscopic implantation of MACI scaffolds, as Ebert et al. report stable clinical improvement at 5 years follow-up and a failure rate of 6.4% [26]. Ventura et al. note improvement in Lysholm symptom scores at 2 years but no change in Tegner activity scores in a series of 53 patients; a high rates of subchondral abnormalities were noted on MRI at 1 year (70% of cases) which did not correlate with clinical symptoms [27].

For the patellofemoral joint, Meyerkort et al. report durable improvement in symptoms at 5 years with MACI; clinical improvement did not correlate with MRI assessment of graft appearance at 5 years [28]. Gigante et al. published results of treatment of patellar defects with MACI and concomitant distal realignment; at 3 years, there was significant improvement in symptoms in most patients and one clinical failure (7%) [29].

As a salvage operation in young patients with medial compartment osteoarthritis, Bauer et al. report significant clinical improvement at 5 years with combined high tibial osteotomy and MACI; however, they note declining results and high graft failure over time for this salvage operation [30]. Finally, outcomes for MACI and concomitant bone grafting for treatment of osteochondral lesions with use of a bilayer 'sandwich' technique have also been reported. Vijayan et al. report outcomes with use of two MACI membranes and impaction bone grafting of osteochondral lesions greater than 8 mm depth; at a mean 5.2 year's follow-up, 12/14 patients had good to excellent results with one graft failure [31].

2.2. Three-dimensional collagen I based scaffolds

2.2.1. NeoCart

NeoCart (Histogenics Corporation, Waltham, Massachusetts) is an MACT implant that consists of a three-dimensional bovine collagen I scaffold (Table 4). Rather than being cultured

Commercial name	Manufacturer	Structure	Expansion	Availability
NeoCart	Histogenics Corporation, Waltham, Massachusetts	Bovine collagen type I matrix	Cells are expanded directly on 3D scaffold via a custom bioreactor	Ongoing phase III clinical trials; not yet approved by the FDA
CaReS	Arthro Kinetics (Ars Arthro, Esslingen, Germany)	Rat collagen type I matrix	Cells are mixed with collagen which forms a gel and cultured for 2 weeks	SFDA certified; not yet approved by the FDA
Novocart 3D	B. Braun-Tetec, Reutlingen, Germany	Collagen- chondroitin sulfate scaffold	Initial monolayer culture followed by seeding onto scaffold; re-implantation 3–4 weeks after harvest	Available in Europe, ongoing phase III clinical trials.

FDA, U.S. Food and Drug Administration; SFDA, State Food and Drug Administration of China; 3D, three-dimensional.

Table 4. MACT with three-dimensional collagen 1 scaffold.

in a monolayer, the scaffold is seeded initially with chondrocytes which then proliferate in a custom bioreactor [32]. The bioreactor is designed to incubate the scaffold in a low-oxygen tension environment with varying pressure to mimic the native intra-articular environment with the goal of preserving the chondrocyte phenotype [33]. At the time of implantation, the graft is fixed to the defect with a proprietary adhesive (CT3 bioadhesive, Histogenics). A randomized phase II trial by Crawford et al. of distal femoral lesions treated with NeoCart versus microfracture demonstrated superior improvement in IKDC and KOOS scores at 24 months with NeoCart and no difference in adverse events between groups (**Table 5**) [33]. A small case series (8 patients) with 2 years follow-up demonstrated significant symptom improvement from baseline and no cases of graft hypertrophy or arthrofibrosis (**Table 5**) [32]. Defect fill was noted to be moderate (33–66%) in 1/8 cases and poor (<33%) in 1/8 cases. A longitudinal

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
1	Crawford et al. [33]	21 NeoCart; 9 microfracture	2 years	Randomized trial of distal femoral lesions. Greater IKDC and KOOS improvement at 2 years with NeoCart.
3	Flohe et al. [35]	9 CaReS; 11 MACI	1 year	No difference in clinical outcomes between groups.
3	Petri et al. [36]	17CaReS; 10 microfracture	3 years	Comparative trial for patellofemoral defects. No difference in groups between IKDC, SF-36, or Cincinnati knee scores at 3 years follow-up.

Table 5. MACT clinical outcome studies with three-dimensional collagen 1 scaffold.

clinical and MRI-based study with 5 years follow-up by Anderson et al. demonstrate that clinical improvement and graft appearance on MRI both evolve over the first 24 months after surgery [34]. Both clinical scores and MRI appearance appeared stable from 24 to 60 months follow-up [34].

2.2.2. CaReS

The Cartilage Regeneration System (CaReS, Ars Arthro, Esslingen, Germany) utilizes a ratderived collagen I gel rather than the bovine collagen matrix utilized by NeoCart (Table 4). The harvested chondrocytes are similarly seeded into the collagen gel and cultured in this 3-dimensional environment with the intention of preserving cartilage phenotype. In a small comparative study of CaReS (9 patients) versus MACI (11 patients) with 1 year follow-up, Flohe et al. demonstrate significant improvement in symptoms with no difference between groups (Table 5) [35]. A small comparative study of microfracture (n = 10) vs. CaReS (n = 17) for patellofemoral lesions found significant improvements in symptoms from baseline with no difference in outcomes between groups [36]. In a multicenter clinical trial, Schneider et al. report outcomes of 116 at mean 30.6 month follow-up from 9 different centers; mean defect size in the trial was 5.4 cm² [37]. At final follow-up there was significant improvement in IKDC, VAS and SF-36 scores and a patient satisfaction rate of 80%. A total of 8 revision arthroscopies were performed for pain with 2 cases of implant hypertrophy and 2 cases of early failure [37]. In an imaging based outcome study, Welsch et al. compared 3T MRI results at 2 years for Hyalograft C versus CaReS and found greater T2 relaxation times for CaReS despite similar clinical outcomes between groups [38].

2.2.3. Novocart 3D

The Novocart 3D implant (B. Braun-Tetec, Reutlingen, Germany) is a collagen-chondroitin sulfate sponge (**Table 4**). After chondrocyte harvest, cells are initially cultured in a monolayer and then seeded onto the collagen-chondroitin sulfate scaffold at a density of 0.5–3.0 × 10⁶ cells/cm², after which the scaffold is cultivated in serum for 2 days before shipment for re-implantation [39]. Niethammer et al. performed several MRI-based studies of graft maturation and graft filling with Novocart 3D. In a 3 years prospective MRI study, graft maturation as assessed by T2 mapping required at least 1 year [40]. In a 2 years prospective MRI study, incomplete graft filling as assessed by MRI was common (55.7%) at 2 years and did not correlate with clinical results; the authors noted that graft thickness appeared to increase throughout the 2 years follow-up period [41]. A 2 years follow-up MRI study showed a 25% graph hypertrophy rate in Novocart 3D patients (11/44 patients), with higher hypertrophy rates in cases of acute traumatic defects or osteochondritis dissecans [42].

In a small non-randomized comparative study, Panagopoulos et al. report outcomes of Novocart 3D (n = 9) and ACI-P (periosteal cover) (n = 11) and mean 37.5 months follow-up (**Table 5**) [43]. No significant difference in Tegner, Lysholm, or IKDC scores was noted between groups. The patient population consisted of high demand athletes and soldiers, with low rates of return to pre-injury activity levels (6/19, 31.5%) [43]. In a comparative study of 40 pediatric (<20 years old) patients treated with Novocart 3D versus 40 matched adult historical controls who also

underwent Novocart for similar size/location lesions, both groups had significant improvement in VAS and IKDC scores at 36 months, but the pediatric group had greater improvement than the adult group at final follow-up [44]. A case series of 23 patients with 2 years follow-up by Zak et al. report improvement in symptoms scores as well as activity scores versus baseline with use of Novocart 3D [39]. At final follow-up, hypertrophy was noted via MRI in 16% and incomplete filling (>50%) in 20% of patients [39]. A large case series by Angele et al. of 433 patients with mean 6.9 months follow-up (max 2.5 years) found an 8.5% re-operation rate, a 6% graft failure rate in patients with >12 months follow-up [45]. Finally, in a case series with 2 years follow-up, Niethammer noted that clinical outcomes at 2 years were worse for patients who returned to sport/physical activities at earlier than 12 months after surgery [46].

2.3. Hyaluronic acid or fibrin based scaffolds

2.3.1. Hyalograft C

The Hyalograft C scaffold is based on the benzylic ester of hyaluronic acid (HYAFF 11; Fidia Advanced Biopolymers Laboratories, Padova, Italy) (**Table 6**). The resulting scaffold is a meshwork of 20 micrometer diameter fibers. The cells are cultured directly on the scaffold with resulting collagen II and aggrecan production [5]. The implant is naturally adhesive and does not require an additional adhesive at time of implantation. Clinical outcomes of Hyalograft C were encouraging, with superior results in comparison to microfracture [47] and comparable results to MACI [48] or traditional ACI with a periosteum cover (**Table 7**) [49]. However, production of this implant has been discontinued by the manufacturer in favor of further development of a single-stage delivery system (no published clinical outcomes data available for the single-stage system).

Commercial name	Manufacturer	Structure	Expansion	Availability
Hyalograft C	Anika Therapeutics (Fidia Advanced Biopolymers Laboratories, Padova, Italy)	Benzylic ester of hyaluronic acid (HYAFF) combined with expanded patient cells	Cells seeded and cultured directly on scaffold	No longer commercially available; production discontinued
BioCart II	Histogenics Corporation, Waltham, MA (merger with former supplier, ProChon Biotech)	Human fibrin and recombinant hyaluronic acid- based scaffold	Cells cultured in human serum and growth factor FGF2v1, then seeded onto scaffold	Available in Italy, Greece, and Israel; ongoing clinical trials in the United States; not yet approved by the FDA
Chondron	Sewon Cellontech, Seoul, Korea	Fibrin based gel	Cells cultured in serum; at time of surgery, suspension is mixed 1:1 with fibrin	Available in Korea

AIFA, Italian Medicines Agency; FDA, U.S. Food and Drug Administration.

Table 6. Hyaluronic acid or fibrin-based scaffolds.

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
2	Kon et al. [47]	21 Hyalograft C; 20 microfracture	7.5 years	Return to sport was a median 8 months for microfracture, 12.5 months for Hyalograft C. Symptom improvement with microfracture deteriorated with time whereas Hyalograft C was durable.
3	Kon et al. [48]	22 Hyalograft C; 39 MACI	5 years	All patients 40 years or older, treated with miniopen MACI or arthroscopic Hyalograft C. Overall failure rate 20%, similar symptom improvement seen in both treatment groups.
3	Ferruzzi et al. [49]	50 Hyalograft C; 48 ACI-P	2–5 years	Similar IKDC improvement at 2+ years. Greater symptom improvement in first 12 months in Hyalograft C (arthroscopic) group versus ACI-P (mini-open)

Table 7. MACT clinical outcome studies with hyaluronic acid or fibrin-based scaffolds.

2.3.2. BioCart II

An implant called BioCart II (Histogenics Corporation, Waltham, MA formerly supplied by ProChon Biotech prior to merger with Histogenics) is comprised of a scaffold of recombinant hyaluronan with fibrin to form a sponge (Table 6). Cells are initially cultured in human serum with recombinant fibroblast growth factor 2 variant (FGF2v1) and then seeded onto the scaffold prior to implantation with a mini-open approach. A small 1 year outcome study by Nehrer et al. of 8 patients demonstrated significant improvement in IKDC and Lysholm scores; 3 patients had a transient effusion post-operatively and there were no clinical failures (Table 7) [50]. A case series by Eshed et al. of patients who underwent MRI evaluation at mean 17.3 months after surgery (range 6-48 months) found continued maturation of cartilage with time (>1 year versus <1 year) and higher IKDC scores in patients with >12 months follow-up and without a history of prior cartilage surgeries [51].

2.3.3. Chondron

The Chondron scaffold is a fibrin-based gel (Sewon Cellontech Co. Ltd., Seoul, Korea) (Table 6). Chondrocytes are first cultured separately in a specialized serum (CRM kit, Sewon Cellontech, Korea). At the time of surgery the serum and cultured chondrocytes are mixed 1:1 with fibrin and injected directly onto the defect. In addition to typical preparation of the defect for ACI, several holes are drilled into the subchondral bone to improve adherence [52]. Choi et al. report a multicenter study of 98 patients with mean 24 month follow-up treated with Chondron (Table 7) [52]. Symptom improvement increased with time, with greater improvement noted with >25 months follow-up versus <25 months. Complication rates were low with one early repeat operation (1%) and two cases of symptomatic catching (2%) [52]. Similar findings were reported in a series by Kim et al., with no graft-related complications among 30 patients at 24 months follow up; a second look arthroscopy at 12 months showed nearly normal cartilage in 8/10 patients [53]. A small series by Konst et al. of 9 patients with osteochondral defects (mean depth 0.9 cm) treated with autologous bone grafting as well as Chondron showed satisfactory short term results at 12 months; there was one treatment failure which was converted to a unicompartmental knee arthroplasty [54].

2.4. Alginate based scaffolds

2.4.1. Cartipatch

Cartipatch (TBF Tissue Engineering, Mions, France) is a MACT implant with a scaffold composed of agarose and alginate (**Table 8**). Chondrocytes are first cultured in a monolayer and then mixed with a hydrogel of agarose and alginate. The hydrogel can be manipulated at 37°C and will solidify around 25°C, allowing formation of complex/irregular shapes with the scaffold. A multicenter randomized trial with 2 years follow-up was recently published by Clave et al. (**Table 9**) [55]. In this study, 30 patients were randomized to Cartipatch and 25 to mosaicplasty; all patients had isolated high grade femoral condylar defects 2.5–7.5 cm² in size. At 2 years, there was significantly greater improvement in IKDC scores with mosaicplasty than Cartipatch, though both groups had significant improvement over baseline. A total of 12 adverse events were reported for the Cartipatch groups and six in the mosaicplasty group [55]. An earlier case series by Selmi et al. reported 2 years outcomes of 17 patients treated with Cartipatch with a mean defect size of 3 cm² [56]. All patients had significant symptom improvement with no clinical failures; second look biopsies in 13 patients had mostly hyaline-like cartilage in 62% of cases (8/13) [56].

Commercial name	Manufacturer	Structure	Expansion	Availability
Cartipatch	Tissue Bank of France (TBF) Tissue Engineering, Mions, France	Alginate-agarose hydrogel combined with autologous cells	Two-step procedure; reduces cell leakage and implantation time	Ongoing phase III clinical trials; not yet approved by the FDA

FDA, U.S. Food and Drug Administration.

Table 8. Alginate hydrogel.

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
1	Clave et al. [55]	30 Cartipatch; 25 mosaicplasty	2 years	Both groups showed improvement in IKDC scores over baseline though mosaicplasty had greater symptom improvement than Cartipatch at 2 years for femoral lesions 2.5–7.5 cm ² .
4	Selmi et al. [56]	17 Cartipatch	2 years	Multicenter study. Significant symptom improvement in all patients, no clinical failures. Second look biopsies showed mostly hyaline-like cartilage in 8/13 patients (62%).

Table 9. MACT clinical outcome studies with alginate-based scaffolds.

2.5. Atelocollagen gel

2.5.1. Koken Atelocollagen Implant

The MACT technique with use of the Koken Atelocollagen Implant (Koken, Tokyo, Japan) is similar to the ACI-P (periosteum cover) technique, but chondrocytes are suspended in atelocollagen gel rather than a liquid to obtain uniform distribution of chondrocytes within the defect and theoretically reduce risk of leakage (Table 10). In this technique, after initial isolation of chondrocytes from cartilage biopsy, the chondrocyte suspension is mixed 1:4 with a 3% bovine atelocollagen solution (Koken, Tokyo, Japan) [57]. Chondrocytes are expanded in this mixture for 28 days; the final product (the Koken Atelocollagen Implant) is an opaque implant with a jelly-like consistency. The Koken Atelocollagen Implant is implanted with a mini-arthrotomy and requires a periosteum cover to contain the atelocollagen-based scaffold within the defect [57]. A multicenter trial in Japan reported by Tohyama et al. reports use of the Koken Atelocollagen Implant and periosteum cover in 27 patients (Table 11) [57]. Overall there was a significant improvement in Lysholm scores at final 2 years follow-up. On second look arthroscopy, 24% of repair sites were ICRS grade normal and 48% were nearly normal. There was one case of graft hypertrophy, two cases of graft detachment, and two cases of abnormal or severely normal ICRS grade on second look arthroscopy [57]. Recently, Tadenuma et al. report clinical and imaging outcomes of 8 patients (11 knees) at mean 5.9 years after surgery [58]. The authors note significant improvement in Lysholm scores over baseline with one clinical failure (9%) and one traumatic repeat injury 7 years after surgery (9%). The authors report a correlation between T1 values of the repair site on MRI and clinical outcomes but no correlation between T2 values and outcomes [58].

Commercial name	Manufacturer	Structure	Expansion	Availability
Koken Atelocollagen Implant	Koken, Tokyo, Japan	Atelocollagen gel (3% type 1 bovine collagen gel)	Chondrocyte suspension is initially mixed 1:4 with 3% atelocollagen solution. The mixture is cultured for 4 weeks and thickens to a jelly-like consistency over that time.	Available in Japan

Table 10. Atelocollagen based scaffold.

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
4	Tohyama et al. [57]	27 Koken Atelocollagen Implant	2 years	Multicenter study. Symptom scores (Lysholm) improved at 2 years from baseline. Two cases of graft detachment (7.4%). Two remaining cases were graded abnormal or severely abnormal on second look arthroscopy (8%, 2/25).
4	Tadenuma et al. [58]	11 Koken Atelocollagen Implant	5.9 years	Improved Lysholm scores at final follow-up with one clinical failure (9%). T1 scores on MRI at final follow-up correlated with clinical scores but T2 scores did not.

Table 11. MACT clinical outcome studies with alginate-based scaffolds.

2.6. Polyglycolic/polylactic acid and polydioxanone based scaffold

2.6.1. BioSeed-C

The BioSeed-C (BioTissue Technologies GmbH, Freiburg, Germany) MACT scaffold is comprised polyglycolic/polylactic acid (polyglactin, vicryl), and polydioxanone (**Table 12**). Harvested chondrocytes are first expanded in serum and then seeded into the polymer scaffold with fixation by fibrin. The scaffold is available in a standard rectangular shape (2 cm × 3 cm × 0.2 cm thickness) can be implanted arthroscopically or with a mini-arthrotomy. The defect must be contoured to a rectangular shape (more than one scaffold can be used as needed for larger defects) and corners of the scaffold are secured with transosseous resorbable suture loops [59].

In a comparative non-randomized study of ACI-P versus BioSeed-C with minimum 2 years follow up, Erggelet et al. report similar improvement in symptom scores (**Table 13**) [60]. The graft failure rate was similar between groups (3/42 ACI-P; 2/40 BioSeed-C), but reoperation rates were twice as high in the ACI-P group, primarily due to graft hypertrophy [60]. A smaller randomized study of ACI-P (n = 10) versus BioSeed-C (n = 9) with 2 years follow-up by Zeifang et al. found similar improvement in symptoms between groups (per IKDC score) at both 1 and 2 years [61]. In contrast to the findings reported by Erggelet et al. [60], re-operation rates were higher in the BioSeed C group (3/11 patients) versus ACI-P (1/10 patients) [61].

Commercial name	Manufacturer	Structure	Expansion	Availability
BioSeed-C	BioTissue AG (BioTissue Technologies, GmbH, Freiburg, Germany)	Fibrin, polyglycolic/polylactic acid and polydioxanone-based material combined with culture-expanded autologous chondrocytes and suspended in fibrin.	Chondrocytes cultured in serum then subsequently seeded into scaffold.	CE mark approval; not yet approved by the FDA.

CE, Conformité Européenne; FDA, U.S. Food and Drug Administration.

Table 12. Scaffolds with polyglycolic/polylactic acid and polydioxanone.

Level of evidence	Author	Implant and sample size	Mean follow-up	Outcome
2	Zeifang et al. [61]	11 BioSeed-C; 9 ACI-P	2 years	Similar IKDC symptom improvement in both groups at 1 year and 2 years. Higher re-operation rate in BioSeed C group.
3	Erggelet et al. [60]	40 BioSeed-C; 42 ACI-P	36 m ACI-P 24 m BioSeed-C	Twice as many re-operations required for ACI-P versus BioSeed-C. Three graft failures in ACI-P group and two in BioSeed-C group. Equivalent improvement in symptom scores between groups.

Table 13. MACT clinical outcome studies with polyglycolic/polylactic acid and polydioxanone based scaffold.

Several case series have also been reported for BioSeed-C (Table 13). Ossendorf et al. report a case series of 40 patients treated with BioSeed-C with 2 years follow-up; symptom scores were significantly improved at both 1 and 2 years after baseline [59]. Reoperations occurred in 12.5% of patients including synovectomy (n = 2), debridement (n = 1), total knee arthroplasty (n = 1), and graft removal (n = 1) [59]. The mid-term outcomes of the same patient cohort with 4-years follow-up were reported by Kreuz et al. [62]. The authors note a durable symptom improvement over 4 years and a high rate of graft filling (mostly or completely filled in 43/44 patients on MRI assessment) [62]. In the subgroup analysis of 19 patients in this cohort with baseline osteoarthritis and a high grade focal defect, Kreuz et al. noted symptom improvement at 6-12 months which remained stable at 4 years as well as two clinical failures that went on to total knee arthroplasty (10.5%) [63].

3. Conclusions

In conclusion, short and mid-term clinical outcomes studies of MACT therapies for cartilage defects of the knee have been encouraging. However, commercial availability of MACT procedures is highly variable with respect to geographic region. Recent approval was granted in December 2016 by the FDA for use of MACI in the United States. To date this is the only MACT therapy available in this region. Availability is greater for multiple MACT therapies in Europe, though European Medicine Agency marketing approval for MACI was recently suspended in June 2016.

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References

- [1] Brittberg M, Lindahl A, Nilsson A, Ohlsson C, Isaksson O, Peterson L. Treatment of deep cartilage defects in the knee with autologous chondrocyte transplantation. The New England Journal of Medicine. 1994;331:889-895
- [2] Ruta DJ, Villarreal AD, Richardson DR. Orthopedic surgical options for joint cartilage repair and restoration. Physical Medicine and Rehabilitation Clinics of North America. 2016;27:1019-1042

- [3] Kon E, Filardo G, Di Martino A, Marcacci M. ACI and MACI. The Journal of Knee Surgery. 2012;25:17-22
- [4] Kon E, Verdonk P, Condello V, et al. Matrix-assisted autologous chondrocyte transplantation for the repair of cartilage defects of the knee: Systematic clinical data review and study quality analysis. The American Journal of Sports Medicine. 2009;37(Suppl 1): 156S-166S
- [5] Grigolo B, Lisignoli G, Piacentini A, et al. Evidence for redifferentiation of human chondrocytes grown on a hyaluronan-based biomaterial (HYAff 11): Molecular, immunohistochemical and ultrastructural analysis. Biomaterials. 2002;23:1187-1195
- [6] Cole BJ, Farr J, Winalski CS, et al. Outcomes after a single-stage procedure for cell-based cartilage repair: A prospective clinical safety trial with 2-year follow-up. The American Journal of Sports Medicine. 2011;39:1170-1179
- [7] Chiang H, Liao CJ, Hsieh CH, Shen CY, Huang YY, Jiang CC. Clinical feasibility of a novel biphasic osteochondral composite for matrix-associated autologous chondrocyte implantation. Osteoarthritis and Cartilage. 2013;21:589-598
- [8] Bartlett W, Skinner JA, Gooding CR, et al. Autologous chondrocyte implantation versus matrix-induced autologous chondrocyte implantation for osteochondral defects of the knee: A prospective, randomised study. Journal of Bone and Joint Surgery. British Volume (London). 2005;87:640-645
- [9] Vascellari A, Rebuzzi E, Schiavetti S, Coletti N. Implantation of matrix-induced autologous chondrocyte (MACI®) grafts using carbon dioxide insufflation arthroscopy. Knee Surgery, Sports Traumatology, Arthroscopy. 2014;**22**:219-225
- [10] Hindle P, Hall AC, Biant LC. Viability of chondrocytes seeded onto a collagen I/III membrane for matrix-induced autologous chondrocyte implantation. Journal of Orthopaedic Research. 2014;32:1495-1502
- [11] Zheng MH, Willers C, Kirilak L, et al. Matrix-induced autologous chondrocyte implantation (MACI): Biological and histological assessment. Tissue Engineering. 2007;**13**:737-746
- [12] Enea D, Cecconi S, Busilacchi A, Manzotti S, Gesuita R, Gigante A. Matrix-induced autologous chondrocyte implantation (MACI) in the knee. Knee Surgery, Sports Traumatology, Arthroscopy. 2012;20:862-869
- [13] Foldager CB, Farr J, Gomoll AH. Patients scheduled for chondrocyte implantation treatment with MACI have larger defects than those enrolled in clinical trials. Cartilage. 2016;7:140-148
- [14] Saris D, Price A, Widuchowski W, et al. Matrix-applied characterized autologous cultured chondrocytes versus microfracture: Two-year follow-up of a prospective randomized trial. The American Journal of Sports Medicine. 2014;**42**:1384-1394
- [15] Basad E, Ishaque B, Bachmann G, Stürz H, Steinmeyer J. Matrix-induced autologous chondrocyte implantation versus microfracture in the treatment of cartilage defects of the knee: A 2-year randomised study. Knee Surgery, Sports Traumatology, Arthroscopy. 2010;18:519-527

- [16] Salzmann GM, Paul J, Bauer JS, et al. T2 assessment and clinical outcome following autologous matrix-assisted chondrocyte and osteochondral autograft transplantation. Osteoarthritis and Cartilage. 2009;17:1576-1582
- [17] Macmull S, Jaiswal PK, Bentley G, Skinner JA, Carrington RW, Briggs TW. The role of autologous chondrocyte implantation in the treatment of symptomatic chondromalacia patellae. International Orthopaedics. 2012;36:1371-1377
- [18] Akgun I, Unlu MC, Erdal OA, et al. Matrix-induced autologous mesenchymal stem cell implantation versus matrix-induced autologous chondrocyte implantation in the treatment of chondral defects of the knee: A 2-year randomized study. Archives of Orthopaedic and Trauma Surgery. 2015;135:251-263
- [19] Edwards PK, Ackland TR, Ebert JR. Accelerated weightbearing rehabilitation after matrix-induced autologous chondrocyte implantation in the tibiofemoral joint: Early clinical and radiological outcomes. The American Journal of Sports Medicine. 2013;**41**:2314-2324
- [20] Ebert JR, Edwards PK, Fallon M, Ackland TR, Janes GC, Wood DJ. Two-year outcomes of a randomized trial investigating a 6-week return to full weightbearing after matrix-induced autologous chondrocyte implantation. The American Journal of Sports Medicine. 2017;45:838-848
- [21] Wondrasch B, Risberg MA, Zak L, Marlovits S, Aldrian S. Effect of accelerated weightbearing after matrix-associated autologous chondrocyte implantation on the femoral condyle: A prospective, randomized controlled study presenting MRI-based and clinical outcomes after 5 years. The American Journal of Sports Medicine. 2015;43:146-153
- [22] Gille J, Behrens P, Schulz AP, Oheim R, Kienast B. Matrix-associated autologous chondrocyte implantation: A clinical follow-up at 15 years. Cartilage. 2016;7:309-315
- [23] Basad E, Wissing FR, Fehrenbach P, Rickert M, Steinmeyer J, Ishaque B. Matrix-induced autologous chondrocyte implantation (MACI) in the knee: Clinical outcomes and challenges. Knee Surgery, Sports Traumatology, Arthroscopy. 2015;23:3729-3735
- [24] Behrens P, Bitter T, Kurz B, Russlies M. Matrix-associated autologous chondrocyte transplantation/implantation (MACT/MACI) — 5-year follow-up. The Knee. 2006;13:194-202
- [25] Ebert JR, Robertson WB, Woodhouse J, et al. Clinical and magnetic resonance imagingbased outcomes to 5 years after matrix-induced autologous chondrocyte implantation to address articular cartilage defects in the knee. The American Journal of Sports Medicine. 2011;39:753-763
- [26] Ebert JR, Fallon M, Wood DJ, Janes GC. A prospective clinical and radiological evaluation at 5 years after arthroscopic matrix-induced autologous chondrocyte implantation. The American Journal of Sports Medicine. 2017;45:59-69
- [27] Ventura A, Memeo A, Borgo E, Terzaghi C, Legnani C, Albisetti W. Repair of osteochondral lesions in the knee by chondrocyte implantation using the MACI® technique. Knee Surgery, Sports Traumatology, Arthroscopy. 2012;20:121-126

- [28] Meyerkort D, Ebert JR, Ackland TR, et al. Matrix-induced autologous chondrocyte implantation (MACI) for chondral defects in the patellofemoral joint. Knee Surgery, Sports Traumatology, Arthroscopy. 2014;**22**:2522-2530
- [29] Gigante A, Enea D, Greco F, et al. Distal realignment and patellar autologous chondrocyte implantation: Mid-term results in a selected population. Knee Surgery, Sports Traumatology, Arthroscopy. 2009;17:2-10
- [30] Bauer S, Khan RJ, Ebert JR, et al. Knee joint preservation with combined neutralising high tibial osteotomy (HTO) and Matrix-induced Autologous Chondrocyte Implantation (MACI) in younger patients with medial knee osteoarthritis: A case series with prospective clinical and MRI follow-up over 5 years. The Knee. 2012;19:431-439
- [31] Vijayan S, Bartlett W, Bentley G, et al. Autologous chondrocyte implantation for osteo-chondral lesions in the knee using a bilayer collagen membrane and bone graft: A two-to eight-year follow-up study. Journal of Bone and Joint Surgery. British Volume (London). 2012;94:488-492
- [32] Crawford DC, Heveran CM, Cannon WD, Foo LF, Potter HG. An autologous cartilage tissue implant NeoCart for treatment of grade III chondral injury to the distal femur: Prospective clinical safety trial at 2 years. The American Journal of Sports Medicine. 2009;37:1334-1343
- [33] Crawford DC, DeBerardino TM, Williams RJ. NeoCart, an autologous cartilage tissue implant, compared with microfracture for treatment of distal femoral cartilage lesions: An FDA phase-II prospective, randomized clinical trial after two years. The Journal of Bone and Joint Surgery. American Volume. 2012;94:979-989
- [34] Anderson DE, Williams RJ, DeBerardino TM, et al. Magnetic resonance imaging characterization and clinical outcomes after NeoCart surgical therapy as a primary reparative treatment for knee cartilage injuries. The American Journal of Sports Medicine. 2017;45:875-883
- [35] Flohé S, Betsch M, Ruße K, Wild M, Windolf J, Schulz M. Comparison of two different matrix-based autologous chondrocyte transplantation systems: 1 year follow-up results. European Journal of Trauma and Emergency Surgery. 2011;37:397-403
- [36] Petri M, Broese M, Simon A, et al. CaReS (MACT) versus microfracture in treating symptomatic patellofemoral cartilage defects: A retrospective matched-pair analysis. Journal of Orthopaedic Science. 2013;18:38-44
- [37] Schneider U, Rackwitz L, Andereya S, et al. A prospective multicenter study on the outcome of type I collagen hydrogel-based autologous chondrocyte implantation (CaReS) for the repair of articular cartilage defects in the knee. The American Journal of Sports Medicine. 2011;39:2558-2565
- [38] Welsch GH, Mamisch TC, Zak L, et al. Evaluation of cartilage repair tissue after matrix-associated autologous chondrocyte transplantation using a hyaluronic-based or a collagen-based scaffold with morphological MOCART scoring and biochemical T2 mapping: Preliminary results. The American Journal of Sports Medicine. 2010;38:934-942

- [39] Zak L, Albrecht C, Wondrasch B, et al. Results 2 years after matrix-associated autologous chondrocyte transplantation using the Novocart 3D scaffold: An analysis of clinical and radiological data. The American Journal of Sports Medicine. 2014;42:1618-1627
- [40] Niethammer TR, Safi E, Ficklscherer A, et al. Graft maturation of autologous chondrocyte implantation: Magnetic resonance investigation with T2 mapping. The American Journal of Sports Medicine. 2014;**42**:2199-2204
- [41] Niethammer TR, Pietschmann MF, Ficklscherer A, Gülecyüz MF, Hammerschmid F, Müller PE. Incomplete defect filling after third generation autologous chondrocyte implantation. Archives of Medical Science. 2016;12:785-792
- [42] Niethammer TR, Pietschmann MF, Horng A, et al. Graft hypertrophy of matrix-based autologous chondrocyte implantation: A two-year follow-up study of NOVOCART 3D implantation in the knee. Knee Surgery, Sports Traumatology, Arthroscopy. 2014;22:1329-1336
- [43] Panagopoulos A, van Niekerk L, Triantafillopoulos I. Autologous chondrocyte implantation for knee cartilage injuries: Moderate functional outcome and performance in patients with high-impact activities. Orthopedics. 2012;35:e6-14
- [44] Niethammer TR, Holzgruber M, Gülecyüz MF, Weber P, Pietschmann MF, Müller PE. Matrix based autologous chondrocyte implantation in children and adolescents: A match paired analysis in a follow-up over three years post-operation. International Orthopaedics. 2017;41:343-350
- [45] Angele P, Fritz J, Albrecht D, Koh J, Zellner J. Defect type, localization and marker gene expression determines early adverse events of matrix-associated autologous chondrocyte implantation. Injury. 2015;46(Suppl 4):S2-S9
- [46] Niethammer TR, Müller PE, Safi E, et al. Early resumption of physical activities leads to inferior clinical outcomes after matrix-based autologous chondrocyte implantation in the knee. Knee Surgery, Sports Traumatology, Arthroscopy. 2014;22:1345-1352
- [47] Kon E, Filardo G, Berruto M, et al. Articular cartilage treatment in high-level male soccer players: A prospective comparative study of arthroscopic second-generation autologous chondrocyte implantation versus microfracture. The American Journal of Sports Medicine. 2011;39:2549-2557
- [48] Kon E, Filardo G, Condello V, et al. Second-generation autologous chondrocyte implantation: Results in patients older than 40 years. The American Journal of Sports Medicine. 2011;39:1668-1675
- [49] Ferruzzi A, Buda R, Faldini C, et al. Autologous chondrocyte implantation in the knee joint: Open compared with arthroscopic technique. Comparison at a minimum follow-up of five years. The Journal of Bone and Joint Surgery. American Volume. 2008;90(Suppl 4): 90-101
- [50] Nehrer S, Chiari C, Domayer S, Barkay H, Yayon A. Results of chondrocyte implantation with a fibrin-hyaluronan matrix: A preliminary study. Clinical Orthopaedics and Related Research. 2008;466:1849-1855

- [51] Eshed I, Trattnig S, Sharon M, et al. Assessment of cartilage repair after chondrocyte transplantation with a fibrin-hyaluronan matrix—Correlation of morphological MRI, biochemical T2 mapping and clinical outcome. European Journal of Radiology. 2012;81:1216-1223
- [52] Choi NY, Kim BW, Yeo WJ, et al. Gel-type autologous chondrocyte (Chondron) implantation for treatment of articular cartilage defects of the knee. BMC Musculoskeletal Disorders. 2010;**11**:103
- [53] Kim MK, Choi SW, Kim SR, Oh IS, Won MH. Autologous chondrocyte implantation in the knee using fibrin. Knee Surgery, Sports Traumatology, Arthroscopy. 2010;18:528-534
- [54] Könst YE, Benink RJ, Veldstra R, van der Krieke TJ, Helder MN, van Royen BJ. Treatment of severe osteochondral defects of the knee by combined autologous bone grafting and autologous chondrocyte implantation using fibrin gel. Knee Surgery, Sports Traumatology, Arthroscopy. 2012;20:2263-2269
- [55] Clavé A, Potel JF, Servien E, Neyret P, Dubrana F, Stindel E. Third-generation autologous chondrocyte implantation versus mosaicplasty for knee cartilage injury: 2-year randomized trial. Journal of Orthopaedic Research. 2016;34:658-665
- [56] Selmi TA, Verdonk P, Chambat P, et al. Autologous chondrocyte implantation in a novel alginate-agarose hydrogel: Outcome at two years. Journal of Bone and Joint Surgery. British Volume (London). 2008;**90**:597-604
- [57] Tohyama H, Yasuda K, Minami A, et al. Atelocollagen-associated autologous chondrocyte implantation for the repair of chondral defects of the knee: A prospective multicenter clinical trial in Japan. Journal of Orthopaedic Science. 2009;14:579-588
- [58] Tadenuma T, Uchio Y, Kumahashi N, et al. Delayed gadolinium-enhanced MRI of cartilage and T2 mapping for evaluation of reparative cartilage-like tissue after autologous chondrocyte implantation associated with Atelocollagen-based scaffold in the knee. Skeletal Radiology. 2016;45:1357-1363
- [59] Ossendorf C, Kaps C, Kreuz PC, Burmester GR, Sittinger M, Erggelet C. Treatment of posttraumatic and focal osteoarthritic cartilage defects of the knee with autologous polymer-based three-dimensional chondrocyte grafts: 2-year clinical results. Arthritis Research & Therapy. 2007;9:R41
- [60] Erggelet C, Kreuz PC, Mrosek EH, et al. Autologous chondrocyte implantation versus ACI using 3D-bioresorbable graft for the treatment of large full-thickness cartilage lesions of the knee. Archives of Orthopaedic and Trauma Surgery. 2010;**130**:957-964
- [61] Zeifang F, Oberle D, Nierhoff C, Richter W, Moradi B, Schmitt H. Autologous chondrocyte implantation using the original periosteum-cover technique versus matrix-associated autologous chondrocyte implantation: A randomized clinical trial. The American Journal of Sports Medicine. 2010;38:924-933

- [62] Kreuz PC, Müller S, Freymann U, et al. Repair of focal cartilage defects with scaffold-assisted autologous chondrocyte grafts: Clinical and biomechanical results 48 months after transplantation. The American Journal of Sports Medicine. 2011;39:1697-1705
- [63] Kreuz PC, Müller S, Ossendorf C, Kaps C, Erggelet C. Treatment of focal degenerative cartilage defects with polymer-based autologous chondrocyte grafts: Four-year clinical results. Arthritis Research & Therapy. 2009;11:R33



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