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# Nitric Oxide Synthase in Male Urological and Andrologic Functions

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Additional information is available at the end of the chapter

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## Abstract

Nitric oxide (NO), a crucial signaling molecule, is synthesized by the nitric oxide synthase (NOS) enzyme. The significant effects of NOS are under exploration, and the roles of potential therapy targets for diseases of NOS are widely accepted. In this chapter, we summarized the important roles of NOS mainly on pathogenesis of prostate diseases, male infertility, erectile dysfunction and, addition, the potential therapeutic efficacies of NOS for those diseases.

**Keywords:** nitric oxide synthase, nitric oxide, prostate cancer, male infertility, erectile dysfunction, male reproduction

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## 1. Introduction

Urology and andrology are the branches of medicine that focus on urinary tract system and male reproductive organs. In recent years, incidences of diseases in urology and andrology system such as prostate cancer and male infertility are increasing and causing heavy burden to our society. Growing studies have been demonstrating that nitric oxide synthase (NOS), which synthesized nitric oxide (NO) by converting L-arginine to L-citrulline, locates in tissues of urinary and male reproductive system and acts as key regulators for sexual function, male reproduction, cancer progression and so on [1–3]. The aims of this chapter are to present the roles of NOS and the recent advances of regulation and therapy function with regard to sexual function, male infertility, prostate carcinoma, Peyronie disease, priapism and cryptorchidism.

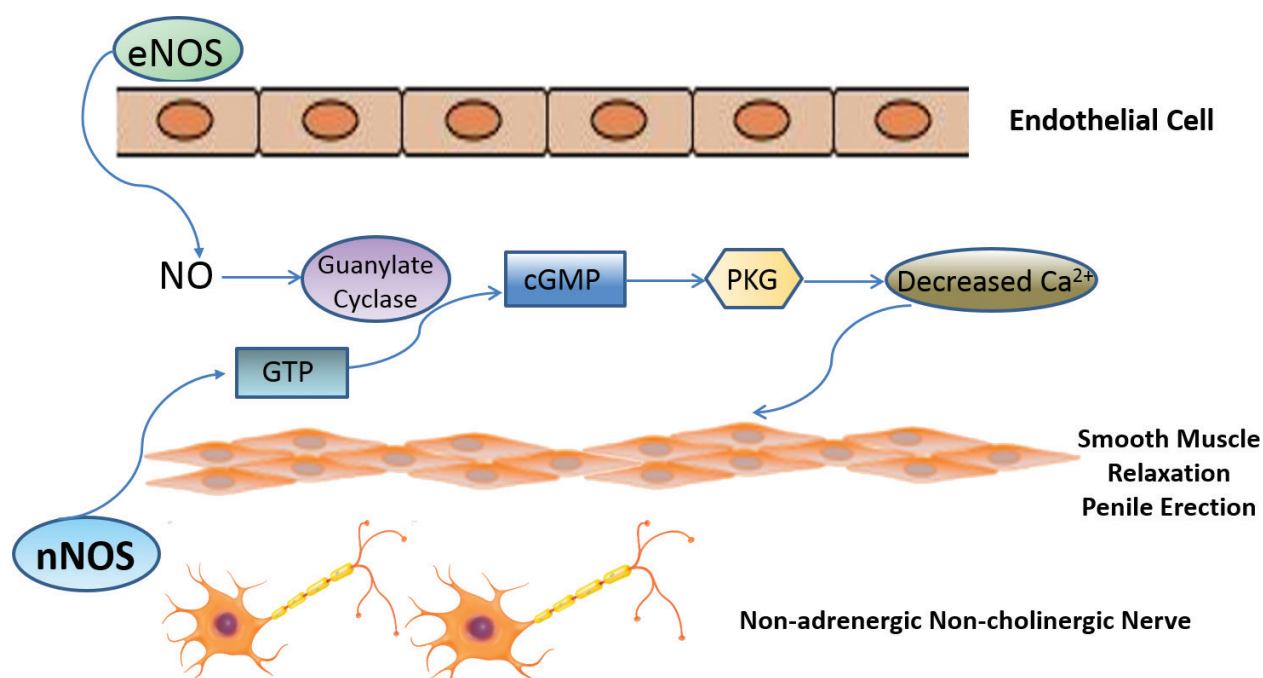
## 2. NOS and male sexual function

### 2.1. NOS and erectile dysfunction

Erectile dysfunction (ED) refers to the symptom that the penis cannot reach and (or) maintain the adequate erection to complete the satisfaction of sexual intercourse, and the course of disease will last at least 6 months or more. Penile erection is an integrated process of artery blood supply and cavernous blood storage launched by nerve, and during this process, neurotransmitter plays an important role [4]. NO is a main messenger, which involves in the induction and maintenance of erection through hemangiectasis and corpus cavernosum relaxation [5]. It has been clear that NO penetrates the smooth muscle cell membrane and catalyzes the formation of cGMP after combining with the ornithine enzyme on the iron ring and then changing the intracellular calcium concentration of smooth muscle to cause relaxation.

#### 2.1.1. NOS in penile tissue

The nNOS and iNOS were found in the central nervous system, especially the hypothalamic area, such as paraventricular nucleus and the medial optic zone, that control the erectile and sexual behavior and also regulate penile erection through spinal nerve centers [6, 7] (**Figure 1**). Specially, nNOS mainly distributes in the penile and pelvic nerve plexus in adult rats, whereas eNOS is in the penis and pelvic area of the urethra but less in the body part of the penis [8, 9].



**Figure 1.** Role of NOS and NO in penile erection. The nNOS and iNOS regulate penile erection through NO/cGMP/PKG pathway. Abbreviations: eNOS, endothelial nitric oxide synthase; nNOS, neuronal nitric oxide synthase; NO, nitric oxide; GTP, guanosine triphosphate; cGMP, cyclic guanosine monophosphate; PKG, protein kinase G.

### 2.1.2. *Current reviews on the effects of NOS on disease-related ED*

#### 2.1.2.1. *Diabetes-related ED*

A possible percentage ranges from 50% to 90% of diabetes patients suffer from ED [10], which counts around three times more than that in healthy cohort. Also, previous report showed that diabetes patients who firstly suffered from ED had a younger age in average compared with people without diabetes and more severe appeared of their symptoms [11]. It has now reached a consensus on the relationship between diabetes and ED, the damage of endothelial cell, ultrastructure changes in cavernous smooth muscle and matrix fibrosis are the common factors affecting the cavernous diastolic function and resulting in ED [12], and damage of eNOS-NO-cGMP pathway was considered to be the main molecular mechanism [13]. The expression of Recombinant Nitric Oxide Synthase Trafficker (NOSTRIN) in Dulbecco's modified eagle medium (DMED) corpus cavernosum increased while the expression of eNOS decreased. It is theorized that increased NOSTRIN may be an important mechanism for the reduction of eNOS expression, while further study is still needed [14].

#### 2.1.2.2. *Benign prostatic hyperplasia-related ED*

Approximately 49% of BPH patients suffer from ED. A significant correlation was reported between BPH/lower urinary tract obstruction (LUTS) and ED after excluding the effects of age and other etiologies on ED [15, 16]. BPH/LUTS seems to be one of the most harmful factors contribute to ED compared with diabetes, hypertension and (or) heart disease [17]. In fact, both parasympathetic innervation of prostate and cavernous nerve of penis are from the pelvic plexus [18]. Pathophysiology studies also showed that the mechanisms of BPH are similar to those of sexual dysfunction which include decrease of the ratio of endothelial NOS/NO, enhancement of endothelial presenilin-1 contraction effect, overreaction of autonomic nervous of bladder, prostate and penis, enhancement of signaling pathway Rho kinase expression/activity and (or) pelvic vascular sclerosis [19]. Since NOS has been found to play significant effects on BPH patients with ED, new treatment by exogenous NO donor and NOS activating enzyme would be promising [20]. However, only few animal experiments have been under exploration up to date, and the mechanisms for the occurrence of ED in BPH patients are still needed to be identified.

#### 2.1.2.3. *Hypertension-related ED*

Hypertension is another important risk factor for ED. Jensen et al. reported that nearly 27% of hypertensive patients suffered from ED [21]. The increase of plasma asymmetrical dimethylarginine (ADMA) concentration caused reduction of the NO expression in penile tissue by inhibiting the NOS activity, which might be a possible mechanism for hypertension-related ED [22].

### 2.1.3. *Possible therapy strategies of NOS on ED*

Increasing evidence has been indicating that L-Arg-NO-cGMP pathway might be a crucial mechanism of penile erection [23]. As a key enzyme in the synthesis of NO, NOS has always been one of the research focuses. Specially, nNOS acts as a key role in erection launch, whereas

eNOS enables cavernous body dilate and maintains the status of erection [24]. Although the effect of iNOS was absent in the direct regulation of penile erection, a special “double effect” in the elderly and the pathological state was reported [25]. Since the reduction of NOSs or the decrease of its activity might contribute to ED, the treatment on L-Arg-NO-cGMP for ED might be revolutionary breakthrough, as phosphodiesterase type 5 inhibitors (PDE5Is) was found to improve the erectile function by increasing the NO concentration but reducing the eGMP degradation [26]. However, nearly 20% of patients with ED still showed little benefit after receiving PDE5Is, especially in patients with diabetes or prostate cancer (Prostate carcinoma) after radical mastectomy [27]. Future NOSs gene transfer therapy from the molecular level would be another choice [28], which might have long-term curative effect, little side effect to the body and, even, completely cure ED. Therapies including increasing expression of NOSs (nNOS, iNOS, eNOS) or inhibiting the expression of protein inhibitor of NOS (PIN) might be promising and worthwhile exploration [29]. However, shortcomings such as short effect duration, possibility of inducing abnormal erection and other potential unknown side effects from the long-term excessive expression of NO are addressed.

## 2.2. NOS and libido

ED may cause low sexual desire or loss of libido in men [30]. Previous study reported that treatment for ED could somehow retrieve sexual desire [31]. It is believed that NO and NOS are beneficial for penile erection, and consequently, NO and NOS may enhance sexual motivation in indirect ways. NO could also affect libido in the direct ways.

Areas for male sexual behavior in brain distribute NO responsive guanylyl cyclase, which involves cellular events of NO [32], and previous studies showed that the NOS inhibitor N<sup>G</sup>-nitro-L-arginine methyl ester (NAME) administered to medial preoptic area by reverse dialysis caused reduced mounting of male rat [32, 33]. Chu et al. further reported that Impaza, a stimulator of eNOS, could raise the sexual incentive motivation rates of male rats through the NO-guanylyl cyclase pathway [34], and nNOS was also considered to affect the male sexual behavior by activating the cyclic guanosine monophosphate (cGMP) [35]. However, adverse result was reported by other researchers, and the conclusion was still controversial [36].

## 3. NOS and male infertility

Approximately 15% of couples suffer from infertility while male cause contributed to nearly 50% in these infertile couples [37]. Male reproduction is known to involve complicated aspects such as spermatogenesis, sperm dynamics, sperm morphology and acrosome reaction. Increasing evidences have been indicating that NOS and NO are associated with male infertility [38].

### 3.1. NOS and male reproductive system

The hypothalamic-pituitary axis plays core roles in reproduction and steroid hormone production in man. Gonadotropin-releasing hormone (GnRH), also known as luteinizing hormone-releasing hormone (LHRH), which is produced and secreted by the arcuate nucleus



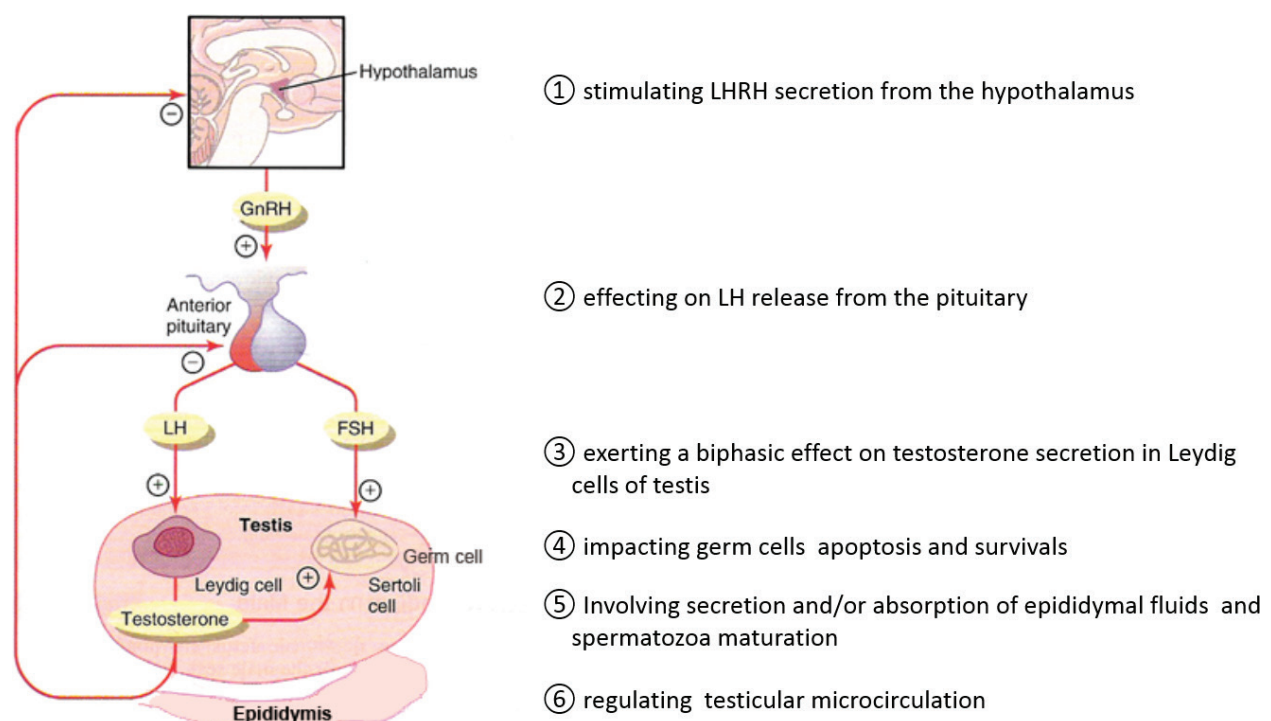
of the hypothalamus, could stimulate the anterior pituitary to episodically release follicle-stimulating hormone (FSH) and luteinizing hormone (LH). LH stimulates the Leydig cells to produce testosterone, and FSH exerts its effect directly on the Sertoli cells to promote spermatogenesis (Figure 2).

In vitro studies have shown that NO stimulates LHRH secretion from the hypothalamus and modulates LH release from the pituitary [39, 40]. Ceccatelli [41] reported that sodium nitroprusside, a NO donor, suppressed GnRH-stimulated LH release from pituitaries in male rats. Chatterjee et al. [42] showed that NOS inhibitor p-nitro-L-arginine methyl ester (L-NAME) enhanced GnRH-induced LH release from pituitaries in rats. Decreased level of GnRH and gonadotropin in chronic NO deficiency rats were also observed [43].

### 3.1.1. Testis

#### 3.1.1.1. Testicular microcirculation

The testis has a rich vascular system that plays a very important role in maintaining the normal functions and stable inner environment of the testis [44]. The regulation of testicular blood microcirculation is very complex, including self-regulation, neural regulation and humoral regulation [45]. NO is the major physiological regulator of basal blood vessel tone and is continually released from endothelium of testicular arteries [46]. A study showed that the regulation effect of NO on testicular blood flow was limited under basal conditions, but this limitation could be significant reversed after HCG treatment; in this case, NO showed



**Figure 2.** Regulation of hypothalamic-pituitary axis. Abbreviations: LHRH, luteinizing hormone-releasing hormone; GnRH, gonadotropin-releasing hormone; LH, luteinizing hormone; FSH, follicle-stimulating hormone.

the effects of increasing blood flow and inhibiting leukocyte accumulation on rat testicular arteries [46]. NO is also an important factor in regulating testicular vessel tension at different temperatures, at 34–37°C, disturbance of testicular arteries reaction appeared after L-NAME treatment [47]. Interestingly, NO content and NOS activity could be significantly increased at abnormal high temperatures caused by varicose spermatic veins in varicocele patients [48].

#### 3.1.1.2. *Leydig cells*

Leydig cells, also known as interstitial cells, are adjacent to the seminiferous tubules in the testicle. Leydig cells produce and release testosterone under the control of LH and act as auto-crine and/or paracrine hormones in gonad under the modulation of NO [49]. An immunohistochemistry study demonstrated that eNOS, nNOS and iNOS all expressed in cytoplasm of Leydig cells in rat testis [50]. Interestingly, a testis-specific subclass of nNOS, known as the truncated form of nNOS (TnNOS), has been recently identified as a major contributor to the formation of NO [51]. TnNOS has been found to be localized solely in the Leydig cells of the testes but neither in the Sertoli nor germ cells [51], which enable us to predict that NO may associate with functions of Leydig cells. Kozieł et al. found that NOS was able to act directly within the male gonad by means of regulating androgen secretion through Leydig cells [52]. Another study showed that stress-induced stimulation of the testicular NO signaling pathway led to the inhibition of steroidogenic enzymes [53]. But NOS seemingly exerted a biphasic effect on testosterone secretion [54]. At low concentrations, NO exerted a transient stimulatory effect on testosterone secretions mediated by cyclic GMP, whereas at high concentrations, it inhibited steroidogenesis by Leydig cells.

#### 3.1.1.3. *Sertoli cells*

Spermatogenesis is a complex process in which Sertoli cells closely involve, and NOS also plays a crucial role in this process through Sertoli cells. Zini et al. [55] showed that eNOS protein located in Sertoli cells and some parts of germ cells in seminiferous tubules, especially in degenerating germ cells and spermatids in histologically normal testes. The iNOS was also found in Sertoli cells, as well as a small subset of pachytene spermatocytes and elongated spermatids in the normal testis [56]. However, iNOS expression could be very intense in Sertoli cells in pathological conditions, for example, the absence of seminiferous tubules [57]. iNOS involves in germ cell death in testicular ischemia-reperfusion injury model, and inhibition of iNOS could improve impaired spermatogenesis [58]. In cryptorchidism model, the transgene expression of eNOS increased testicular germ cell apoptosis. In iNOS<sup>-/-</sup> mice, the numbers of spermatocytes, spermatids and Sertoli cells per tubule were significantly more than those with wild-type testes [59]. A possible conclusion could be drawn that NO plays an important role in both numerical and functional regulation of key somatic cells in the testis, which in turn impacts on germ cells and their survivals during the process of daily sperm production.

#### 3.1.1.4. *Epididymis*

The main functions of the epididymis are promoting spermatozoa mature and storing spermatozoa [60]. An immunostaining study in human epididymis showed that NOS almost exclusively located in the epithelium [55], and the greatest concentration was in the adluminal

region [55]. It suggests that NOS may involve secretion and/or absorption of epididymal fluids, or in another way diffuse into the tubule lumen to affect nearby spermatozoa. Another study showed a similar distribution of NOS protein in rat epididymis, speculating that epididymal NOS protein might contribute to spermatozoa maturation [61].

### 3.2. NOS and sperm function

Approximately 15% of couples suffer from infertility while male cause contributed to nearly 50% in these infertile couples [37]. Male reproduction is known to involve complicated aspects, such as spermatogenesis, sperm dynamics, sperm morphology and acrosome reaction. Increasing evidences have been indicating that NOS and NO are associated with male infertility [38].

#### 3.2.1. Sperm motility, morphology and viability

Sperm motility is an essential factor for male fertility. Low sperm motility, also referred as asthenozoospermia, is one of the major causes to male infertility [62]. Previous study indicated that nearly 80% of semen samples from infertile males were defective in sperm motility [63]. Hellstrom et al. reported for the first time that sodium nitroprusside, a NO releaser, was beneficial for maintenance of thaw-sperm motility by reducing lipid peroxidative damage to sperm membranes. Significantly improved motion parameters of sperm were observed in semen samples treated with sodium nitroprusside in concentrations of 50 and 100 nM compared to control samples, and this beneficial effect maintained for 5–6 hours after thaw [64]. However, NO concentration in normozoospermic fertile men was observed to be significantly lower than those of asthenospermia infertile men [65]. In fact, the effect of NO seems to be double-sided, low concentration of NO improves sperm motility, while high concentration contributes to adverse effect [66]. Herrero et al. reported that a significant decrease on sperm motility was observed in semen samples treated with sodium nitroprusside in a higher concentration of 300 mM, and this effect could be blocked by hemoglobin, a scavenger of NO, as sperm motility in samples furtherly treated with hemoglobin was significantly higher than those without. While when the incubating concentration of sodium nitroprusside reduced to 150 mM, no modifications of sperm motility were found [67]. Besides, the other NO releaser, S-nitroso-N-acetylpenicillamine (0.012–0.6 mM), along with sodium nitroprusside (0.25–2.5 mM), was found to decrease percentage of forward progressive sperm motility and straight line velocity in a concentration-dependent manner [68].

As to sperm morphology and viability, the effects of NO reveal controversial contributions. a positive correlation between NO with defects in sperm morphology has been found in male with normal sperm rate  $\geq 14\%$  but a negative correlation with defects in sperm morphology in male with normal sperm rate  $< 14\%$  [69]. However, later study failed to find any significant association between NO production and sperm morphology [70]. Researchers reported that semen treated with 0.25–2.5 mM sodium nitroprusside revealed significantly less sperm bound to the zona pellucida compared with the control group which treated without NO [71], whereas some other researchers reported no any significant effect of NO on sperm viability [66, 69, 72]. And meanwhile, low concentration of NO also plays a role in the maintenance of sperm viability after cryopreservation and post-thaw sperm [65, 66, 68].



### 3.2.2. *Capacitation, hyperactivation and acrosome reaction*

Capacitation is a process in which spermatozoa acquire the ability to bind to the egg's zone pellucida and fertilize an oocyte during their transit in the female genital tract [73, 74]. Capacitation involves in some molecular events, and it was clear that low level of NO from NO-releasing agents induces human sperm capacitation [75]. Indeed, it has been reported that NO-releasing compounds significantly benefit the capacitation, whereas NO inhibitors decrease this process [76]. In fact, NO produced by spermatozoa involves in a cascade of molecular events of capacitation, which is needed over the course of this process [77–79].

Hyperactivation can be treated as a subcategory of capacitation. Hyperactivation of spermatozoa exhibits high amplitude and asymmetric flagellar movement, non-linear motility and penetrate the oocyte with strong propulsive force [70, 80]. The effect of NO on hyperactivation was found to be similar to which on sperm motility, little concentrations of NO increased spermatozoa hyperactivation, whereas excessive concentrations decreased the hyperactivated spermatozoa motility [81, 82].

Acrosome reaction denotes the process that capacitated and hyperactivated motility sperm binds to the zona pellucida and continues to pass through the exocytotic release of proteolytic enzymes from the acrosome so that to bind to the mature ovum. The amount of NO influenced the acrosome reaction, and increased amount of sperm was observed to undergo the acrosome reaction with the presence of NO donor compound [83]. Meanwhile, significantly increased amount of sperm was found to bind to membrane of the ovum with their plasma membrane [84].

### 3.2.3. *Sperm mitochondria*

Mitochondria in sperm activate as a generator which supply sperm with energy for the process of motility, acrosome reaction, oocyte fusion, fertilization and so on [85]. NO has been reported to involve in functions of mitochondrial that include biogenesis, remodeling and mitochondrial respiration [86–88]. Specially, different levels of NO could cause different sperm mitochondrial functions, low concentrations of NO enhanced the sperm motility, while NO with higher levels cause mitochondrial hyperpolarization and sperm apoptosis [64, 89]. This might explain the adverse effects of various concentrations NO on sperm motility.

## 3.3. **Single-nucleotide polymorphisms of NOS and male infertility**

Genetic variations are crucial etiological factors contribute to male infertility. Up to date, some single-nucleotide polymorphisms (SNPs) have been identified to involve in sperm defects and male infertility in ethnic populations. Polymorphisms T786C and G894T of eNOS were reported to decrease sperm motility and quantity by increasing the seminal oxidative stress in Egyptian infertile male population [90]. Similar results of G894T were reported in Italian and Iranian infertile male populations [91, 92], and this SNP also found to be associated with higher level of sperm DNA fragmentation in Chinese infertile males [93]. The polymorphism 4a4b, which refers to a sequence variant with variable sequence of tandem 4a4b repeats in intron 4, was found to be associated with poor sperm morphology and male infertility in a

Korean and Chinese population [94, 95]. Associations between SNPs of NOS and male infertility are under exploration, which would be promising tools for diagnosis or further curing male infertility.

### **3.4. Possible therapy strategies of NOS on male infertility**

Increasing evidences has been showing that inappropriate concentration level of NO may contribute to male infertility in some extent by means of decreasing sperm motility and normal sperm morphology, reducing efficiency of capacitation and acrosome action. It is reasonable to consider possible therapy strategies to the utilization of NOS donors or inhibitors so that to adjust the concentration of NO to the “right” level. In fact, significantly higher fertile rate was observed in animal experiment in vitro, and further researches would be needed to warrant the potential benefits for human beings.

## **4. NOS and prostate carcinoma**

Prostate carcinoma is one of the most common cancers among men and second in cancer-related deaths in the United States. An estimated study predicted that there will be 180, 890 new prostate carcinoma cases and 26, 120 deaths due to the disease in the country in 2016 [96]. Etiological studies implicated that multiple reasons involved in prostate carcinoma susceptibility, such as dietary, environment, hormone status and genetic factors [97]. Growing studies indicated that NOS and NO system play crucial roles in progression of human prostate carcinoma [98–100].

The physiological functions of NO are dependent primarily on concentrations. Low concentration of NO acted as a signal transducer and affects many physiological processes including blood flow regulation, platelet activity, iron homeostasis, cell proliferation and neurotransmission, whereas, in high concentrations, it exerted a cytotoxic protective effect, for example, to against pathogens and perhaps tumors [101, 102].

### **4.1. Role of nitric oxide synthase in cancer biology**

The roles of NOS and NO on DNA damage, apoptosis, cell cycle, enhancement of cell proliferation, angiogenesis and metastasis are currently viewed, and NO was found to be associated with tumor environment, for example, the vasculature cells and other stromal cells [103–105]. Research also indicated that NOS2 expression was correlated with tumor vascularization, accumulations of p53 mutations and activation of epidermal growth factor receptor, even could be treated as an independent predictor of poor survival in women with estrogen receptor (ER)-negative breast tumors [106]. Low concentrations of NO acted as a promotional role in angiogenesis which stimulates tumor progression by providing blood flow access to the tumor and subsequently resulting in cell proliferation. On the contrary, high levels of NO tend to be cytotoxic to cancer cells [107]. While in animal models, iNOS overexpression produced either pro-tumor or anti-tumor effect on tumor growth, these alterable effects seem to be dependent on the tumor microenvironment and the tumor type itself [104, 108]. The effects

of NO possibly differ in expression level of iNOS, duration and timing of NO delivery, the microenvironment, the genetic background and the cell type (**Figure 3**) [109].

#### 4.2. NOS and proliferation of prostate carcinoma

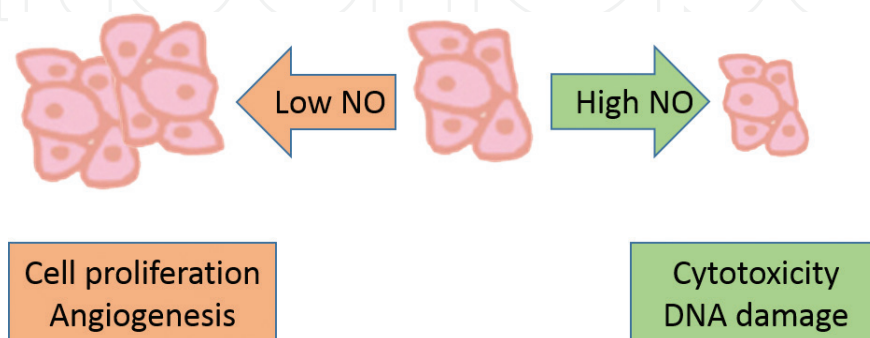
NO generated by eNOS or iNOS might be involved in prostate proliferation. At low concentrations, NO acted as a signaling molecule regulating smooth muscle relaxation and blood flow, neurotransmission, platelet activity, iron homeostasis, cell survival and proliferation, while at high concentrations acted as modulating immune-mediated anti-tumor activities [110, 111]. Concentration of NO less than 100nM had an effect of preventing certain cell types from apoptosis and thereby favors tumorigenesis and progression [112]. Higher expression of iNOS was detected in cancer specimens than that in normal tissues of prostate carcinoma patient. Aaltoma et al. also demonstrated a positive association between expression level of iNOS and rapid cancer cell proliferation rate, dedifferentiation and advanced stage cancer [113]. A recent study has shown that NO also regulated cell proliferation in a pathway of CPD-Arg-NO [114].

#### 4.3. Nitric oxide synthase and angiogenesis of prostate carcinoma

Angiogenesis is a critical molecular event in tumor progression [115, 116]. Epidermal growth factor receptor (EGFR) signaling pathway, tumor suppressor p53 and VEGF, which are collective mediators that exacerbate angiogenesis can be stimulated by NO [115, 117]. The involvement of eNOS in the NO-induced human endothelial and prostate carcinoma cell migration was further warranted [116]. Recent research also reported that NO played vital roles in maintaining blood supply for prostate carcinoma, and an anti-tumor vascular activity effect revealed with presence in inhibition of NOS [115].

#### 4.4. Single-nucleotide polymorphisms of NOS and susceptibility of prostate carcinoma

Several studies suggested that polymorphisms of some NOS genes were genetic susceptibility factors for prostate carcinoma, especially for aggressive diseases [118, 119]. A plethora of meta-analyses has identified eNOS gene polymorphisms as strong susceptibility factors for the progression toward prostate carcinoma [120]. Another study also reported that NOS3 gene



**Figure 3.** Roles of NO in prostate cancer. Abbreviation: NO, nitric oxide.

polymorphisms were genetic susceptibility factors for the progression of prostate carcinoma and poor patient outcomes [121]. A meta-analysis conducted by Zhao et al. suggested that eNOS gene 894G > T polymorphism contributed to aggravate the onset of prostate carcinoma in males [122]. Nikolic et al [123] also corroborated the involvement of eNOS or NOS3 gene in the pathogenesis of prostate carcinoma. NOS3 rs1799983 polymorphism augmented the risk of prostate carcinoma in various populations. As one of the possible mechanisms, the involvement of NO receptor component, sGC-1, in mediating the proliferation of prostate carcinogenesis, has been surmised [124].

#### **4.5. Possible therapy strategies of NOS on prostate carcinoma**

Anti-cancer agents such as gold lotion have successfully demonstrated their anti-carcinogenic potential through the regulation of both iNOS and eNOS [125–127]. Yu et al. [128] also elucidated the significance of eNOS as a seemingly promising strategy for targeting anti-androgen resistant prostate carcinoma. Arginine-releasing compounds such as carboxypeptidase-D increased NO production, which slackened progression of prostate carcinoma so that prolonged survival time [129]. NO-donor drugs also have been under increasing explorations. A few NO-donor drugs have been confirmed to have favorable anticancer activity and could be potential anticancer therapies [3, 130]. GIT-27NO, a novel NO donor, inhibited the growth of PC3 and LnCap prostate carcinoma cells xenografted into nude mice in a concentration-dependent manner [131]. And DETA-NONOate was revealed to inhibit epithelial-mesenchymal transition (EMT) and invasion of human prostate metastatic cells by producing large amount of NO [132]. It is sensible that novel NOS-based therapeutics may prove valuable in the future treatment of prostate carcinoma.

## **5. NOS and other urinary and male reproductive diseases**

### **5.1. Peyronie disease**

Peyronie disease (PD) is an intractable, sexually dysfunctional disease resulting in penile curvature, penile pain, penile deformity, difficulty with coitus, shortening, hinging, narrowing and ED. Mechanisms of PD have not been fully elucidated. A recent hypothesis was that the recurrent microtrauma of the tunica albuginea caused small damages that activated processes of wound healing and fibrotic plaque development during sexual intercourse [133]. Inflammatory cells and iNOS accumulated in the process of wound healing, the increased NO then led to the myofibroblasts and proliferation of fibroblasts and redundant collagen between the layers of the tunica albuginea (penile plaque) [134]. Although surgical therapy is now the first-option for PD patients, researchers are focusing on the nonsurgical treatments of PD, and NOs inhibitors might be a promising choice [135].

### **5.2. Priapism**

Priapism is defined as a persistent and painful erection that lasts longer than 4 hours without sexual stimulation and can lead to ED [136]. The relation between penile erection and production of NOS has been well investigated: nNOS and eNOS were the causes of both the initiation

and maintenance phases of penile erection [137]. However, decreased function in NO generated by decreased activation of eNOS resulted in PDE5 downregulation that was thought to be a derivate of NO and, therefore, reduced basal levels of PDE5 and caused priapism [138].

### 5.3. Cryptorchidism

Cryptorchidism denotes failure of the movement of the testis to the scrotum, and in most cases, it raises risk of testicular germ cell cancer and subfertility later in patients' life course. Testicular germ cell apoptosis which causes by exposure of testicular in elevated temperature and oxidative stress is the primary etiology of infertility. Animal models with cryptorchidism induced by surgery revealed that eNOS played a significant role in mouse spermatogenesis in cryptorchidism-induced apoptosis [139]. Contemporaneously, reduced rate of testicular atrophy was observed in heterozygous Hoxa 11 knockout mice which had congenital bilateral cryptorchidism when early treated with Nomega-nitro-L-arginine methyl ester (L-NAME), a NOS inhibitor [140].

## 6. Conclusion

It has been becoming evident that redox regulation driven by NOS and NO represents a promising tool for exploring fundamental diseases process and new development of strategies to treat urinary, male reproductive and sexual diseases.

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## References

- [1] Buzadzic B, Vucetic M, Jankovic A, Stancic A, Korac A, Korac B, Otasevic V. New insights into male (in)fertility: The importance of NO. *British Journal of Pharmacology*. 2015;**172**:1455–1467.
- [2] Toque HA, Caldwell RW. New approaches to the design and discovery of therapies to prevent erectile dysfunction. *Expert Opinion on Drug Discovery*. 2014;**9**:1447–1469.
- [3] Hirst D, Robson T. Nitric oxide in cancer therapeutics: Interaction with cytotoxic chemotherapy. *Current Pharmaceutical Design*. 2010;**16**:411–420.
- [4] Patel CK, Bennett N. Advances in the treatment of erectile dysfunction: What's new and upcoming? *F1000Research*. 2016;**5**.
- [5] Gur S, Kadowitz PJ, Sikka SC, Peak TC, Hellstrom WJ. Overview of potential molecular targets for hydrogen sulfide: A new strategy for treating erectile dysfunction. *Nitric oxide: Biology and Chemistry/official Journal of the Nitric Oxide Society*. 2015;**50**:65–78.
- [6] Bivalacqua TJ, Champion HC, Usta MF, Celtek S, Chitaley K, Webb RC, Lewis RL, Mills TM, Hellstrom WJ, Kadowitz PJ. RhoA/rho-kinase suppresses endothelial nitric oxide synthase in the penis: A mechanism for diabetes-associated erectile dysfunction. *Proceedings of the National Academy of Sciences of the United States of America*. 2004;**101**:9121–9126.
- [7] Azadzoi KM, Master TA, Siroky MB. Effect of chronic ischemia on constitutive and inducible nitric oxide synthase expression in erectile tissue. *Journal of Andrology*. 2004;**25**:382–388.
- [8] Seyam RM, Huynh HT, Brock GB. Neuronal and endothelial nitric oxide synthase isoforms: Quantification of protein and mRNA in the normal rat penis. *International Journal of Impotence Research*. 1999;**11**:301–308.
- [9] Mizusawa H, Hedlund P, Brioni JD, Sullivan JP, Andersson KE. Nitric oxide independent activation of guanylate cyclase by yc-1 causes erectile responses in the rat. *The Journal of Urology*. 2002;**167**:2276–2281.
- [10] Schramek P, Holzmann RM, Floth A. Disorders of erectile potency in diabetes mellitus. *Wiener klinische Wochenschrift*. 1988;**100**:460–463.
- [11] Chu NV, Edelman SV. Erectile dysfunction and diabetes. *Current Diabetes Reports*. 2002;**2**:60–66.
- [12] Pelliccione F, D'Angeli A, D'Andrea S, Barbonetti A, Pezzella A, Necozone S, Falone S, Amicarelli F, Francavilla F, Francavilla S. Tadalafil treatment had a modest effect on endothelial cell damage and repair ability markers in men with erectile dysfunction and vascular risk. *Asian Journal of Andrology*. 2014;**16**:290–294.
- [13] Nunes KP, Wynne BM, Cordeiro MN, Borges MH, Richardson M, Leite R, DeLima ME, Webb RC. Increased cavernosal relaxation by *Phoneutria nigriventer* toxin, pntx2–6, via activation at no/cgmp signaling. *International Journal of Impotence Research*. 2012;**24**:69–76.

- [14] Mookerjee RP, Wiesenthal A, Icking A, Hodges SJ, Davies NA, Schilling K, Sen S, Williams R, Novelli M, Muller-Esterl W, Jalan R. Increased gene and protein expression of the novel eNOS regulatory protein nostrin and a variant in alcoholic hepatitis. *Gastroenterology*. 2007;**132**:2533–2541.
- [15] Dogan Y, Uruc F, Aras B, Sahin A, Kivrak M, Urkmez A, Guner ND, Aydin S. The relationships between metabolic syndrome, erectile dysfunction and lower urinary tract symptoms associated with benign prostatic hyperplasia. *Turkish Journal of Urology*. 2015;**41**:7–12.
- [16] Lee LK, Goren A, Boytsov NN, Donatucci CF, McVary KT. Treatment satisfaction among men with concurrent benign prostatic hyperplasia and erectile dysfunction treated with tadalafil or other phosphodiesterase type-5 inhibitor combinations. *Patient Preference and Adherence*. 2016;**10**:1205–1215.
- [17] Mevcha A, Gulur DM, Gillatt D. Diagnosing urological disorders in ageing men. *The Practitioner*. 2010;**254**:25–26, 28–29, 22–23.
- [18] Ganzer R, Stolzenburg JU, Wieland WF, Brundl J. Anatomic study of periprostatic nerve distribution: Immunohistochemical differentiation of parasympathetic and sympathetic nerve fibres. *European Urology*. 2012;**62**:1150–1156.
- [19] Fusco F, D'Anzeo G, Sessa A, Pace G, Rossi A, Capece M, d'Emmanuele di Villa Bianca R. BPH/LUTS and ED: Common pharmacological pathways for a common treatment. *The Journal of Sexual Medicine*. 2013;**10**:2382–2393.
- [20] McVary K. Lower urinary tract symptoms and sexual dysfunction: Epidemiology and pathophysiology. *BJU International*. 2006;**97** Suppl 2:23–28; discussion 44–25.
- [21] Jensen J, Lendorf A, Stimpel H, Frost J, Ibsen H, Rosenkilde P. The prevalence and etiology of impotence in 101 male hypertensive outpatients. *American Journal of Hypertension*. 1999;**12**:271–275.
- [22] Maas R, Wenske S, Zabel M, Ventura R, Schwedhelm E, Steenpass A, Klemm H, Noldus J, Boger RH. Elevation of asymmetrical dimethylarginine (adma) and coronary artery disease in men with erectile dysfunction. *European Urology*. 2005;**48**:1004–1011; discussion 1011–1002.
- [23] Wessells H, Hruby VJ, Hackett J, Han G, Balse-Srinivasan P, Vanderah TW. Ac-nle-c[asp-his-dphe-arg-trp-lys]-nh<sub>2</sub> induces penile erection via brain and spinal melanocortin receptors. *Neuroscience*. 2003;**118**:755–762.
- [24] Burnett AL, Nelson RJ, Calvin DC, Liu JX, Demas GE, Klein SL, Kriegsfeld LJ, Dawson VL, Dawson TM, Snyder SH. Nitric oxide-dependent penile erection in mice lacking neuronal nitric oxide synthase. *Molecular Medicine (Cambridge, Mass)*. 1996;**2**:288–296.
- [25] Gonzalez-Cadavid NF, Rajfer J. The pleiotropic effects of inducible nitric oxide synthase (iNOS) on the physiology and pathology of penile erection. *Current Pharmaceutical Design*. 2005;**11**:4041–4046.

- [26] Chen L, Staubli SE, Schneider MP, Kessels AG, Ivic S, Bachmann LM, Kessler TM. Phosphodiesterase 5 inhibitors for the treatment of erectile dysfunction: A trade-off network meta-analysis. *European Urology*. 2015;**68**:674–680.
- [27] Wang X, Wang X, Liu T, He Q, Wang Y, Zhang X. Systematic review and meta-analysis of the use of phosphodiesterase type 5 inhibitors for treatment of erectile dysfunction following bilateral nerve-sparing radical prostatectomy. *PLoS One*. 2014;**9**:e91327.
- [28] Gonzalez-Cadavid NF, Rajfer J. Molecular pathophysiology and gene therapy of aging-related erectile dysfunction. *Experimental Gerontology*. 2004;**39**:1705–1712.
- [29] Magee TR, Kovanecz I, Davila HH, Ferrini MG, Cantini L, Vernet D, Zuniga FI, Rajfer J, Gonzalez-Cadavid NF. Antisense and short hairpin RNA (shRNA) constructs targeting pin (protein inhibitor of NOS) ameliorate aging-related erectile dysfunction in the rat. *The Journal of Sexual Medicine*. 2007;**4**:633–643.
- [30] Lewis RW, Fugl-Meyer KS, Corona G, Hayes RD, Laumann EO, Moreira ED, Jr., Rellini AH, Segraves T. Definitions/epidemiology/risk factors for sexual dysfunction. *The Journal of Sexual Medicine*. 2010;**7**:1598–1607.
- [31] Jannini EA, Screponi E, Carosa E, Pepe M, Lo Giudice F, Trimarchi F, Benvenga S. Lack of sexual activity from erectile dysfunction is associated with a reversible reduction in serum testosterone. *International Journal of Andrology*. 1999;**22**:385–392.
- [32] De Vente J, Hopkins DA, Markerink-Van Ittersum M, Emson PC, Schmidt HH, Steinbusch HW. Distribution of nitric oxide synthase and nitric oxide-receptive, cyclic GMP-producing structures in the rat brain. *Neuroscience*. 1998;**87**:207–241.
- [33] Ratnasooriya WD, Dharmasiri MG, Wadsworth RM. Reduction in libido and fertility of male rats by administration of the nitric oxide (no) synthase inhibitor n-nitro-l-arginine methyl ester. *International Journal of Andrology*. 2000;**23**:187–191.
- [34] Chu X, Zhavbert ES, Dugina JL, Kheyfets IA, Sergeeva SA, Epstein OI, Agmo A. Sildenafil and a compound stimulating endothelial no synthase modify sexual incentive motivation and copulatory behavior in male Wistar and Fisher 344 rats. *The Journal of Sexual Medicine*. 2008;**5**:2085–2099.
- [35] Chu X, Agmo A. Sexual incentive motivation in old male rats: The effects of sildenafil and a compound (impaza) stimulating endothelial no synthase. *Pharmacology, Biochemistry, and Behavior*. 2008;**89**:209–217.
- [36] Ratnasooriya WD, Jayakody JR, Dharmasiri MG. Sodium nitroprusside impairs sexual competence of male rats. *Human & Experimental Toxicology*. 2004;**23**:187–192.
- [37] Schlegel PN. Evaluation of male infertility. *Minerva Ginecologica*. 2009;**61**:261–283.
- [38] Lee NP, Cheng CY. Nitric oxide and cyclic nucleotides: Their roles in junction dynamics and spermatogenesis. *Advances in Experimental Medicine and Biology*. 2008;**636**:172–185.

- [39] McCann SM, Mastronardi C, Walczewska A, Karanth S, Rettori V, Yu WH. The role of nitric oxide in reproduction. *Brazilian Journal of Medical and Biological Research*. 1999;**32**:1367–1379.
- [40] Rettori V, Belova N, Dees WL, Nyberg CL, Gimeno M, McCann SM. Role of nitric oxide in the control of luteinizing hormone-releasing hormone release in vivo and in vitro. *Proceedings of the National Academy of Sciences of the United States of America*. 1993;**90**:10130–10134.
- [41] Ceccatelli S. Expression and plasticity of no synthase in the neuroendocrine system. *Brain Research Bulletin*. 1997;**44**:533–538.
- [42] Chatterjee S, Collins TJ, Yallampalli C. Inhibition of nitric oxide facilitates lh release from rat pituitaries. *Life Sciences*. 1997;**61**:45–50.
- [43] Barnes MJ, Lapanowski K, Rafols JA, Lawson DM, Dunbar JC. GnRH and gonadotropin release is decreased in chronic nitric oxide deficiency. *Experimental Biology and Medicine* (Maywood, NJ). 2001;**226**:701–706.
- [44] Damber JE, Bergh A. Testicular microcirculation—A forgotten essential in andrology? *International Journal of Andrology*. 1992;**15**:285–292.
- [45] Gu L, Jin B, Zhang X. Progress in testicular microcirculation structure and regulation. *Chinese Journal of Microcirculation*. 2015;**24**:65–67.
- [46] Lissbrant E, Lofmark U, Collin O, Bergh A. Is nitric oxide involved in the regulation of the rat testicular vasculature. *Biology of Reproduction*. 1997;**56**:1221–1227.
- [47] Sabanegh E, Dewire D, Inman S. The effect of nitric oxide blocked on rat testicular microcirculation. *Fertility and Sterility*. 1994;**62**:1–17.
- [48] Xu Y, Xu QY, Yang BH, Zhu XM, Peng YF. Relationship of nitric oxide and nitric oxide synthase with varicocele infertility. *Zhonghua nan ke xue [National Journal of Andrology]*. 2008;**14**:414–417.
- [49] Lamanna C, Assisi L, Vittoria A, Botte V, Di Fiore MM. D-aspartic acid and nitric oxide as regulators of androgen production in boar testis. *Theriogenology*. 2007;**67**:249–254.
- [50] Li MX, He LP, Guo ZQ, Liu YS, Long ZF. The expression of nitric oxide synthase in testes of male rat. *Zhonghua nan ke xue [National Journal of Andrology]*. 2002;**8**:250–252.
- [51] Wang Y, Newton DC, Miller TL, Teichert AM, Phillips MJ, Davidoff MS, Marsden PA. An alternative promoter of the human neuronal nitric oxide synthase gene is expressed specifically in Leydig cells. *American Journal of Pathology*. 2002;**160**:369–380.
- [52] Koziel E, Kotula M, Andronowska A, Pierscinski A, Bilinska B. Correlation between nadph-diaphorase and iNOS in bank vole Leydig cells in vitro and in testicular sections with the use of histochemistry and immunocytochemistry. *Folia Histochemica et Cytobiologica*. 2000;**38**:71–78.

- [53] Kostic TS, Andric SA, Maric D, Kovacevic RZ. Inhibitory effects of stress-activated nitric oxide on antioxidant enzymes and testicular steroidogenesis. *The Journal of Steroid Biochemistry and Molecular Biology*. 2000;**75**:299–306.
- [54] Valenti S, Cuttica CM, Fazzuoli L, Giordano G, Giusti M. Biphasic effect of nitric oxide on testosterone and cyclic GMP production by purified rat Leydig cells cultured in vitro. *International Journal of Andrology*. 1999;**22**:336–341.
- [55] Zini A, O'Bryan MK, Magid MS, Schlegel PN. Immunohistochemical localization of endothelial nitric oxide synthase in human testis, epididymis, and vas deferens suggests a possible role for nitric oxide in spermatogenesis, sperm maturation, and programmed cell death. *Biology of Reproduction*. 1996;**55**:935–941.
- [56] O'Bryan MK, Schlatt S, Gerdprasert O, Phillips DJ, de Kretser DM, Hedger MP. Inducible nitric oxide synthase in the rat testis: Evidence for potential roles in both normal function and inflammation-mediated infertility. *Biology of Reproduction*. 2000;**63**:1285–1293.
- [57] Costur P, Filiz S, Gonca S, Culha M, Gulecen T, Solakoglu S, Canberk Y, Caliskan E. Expression of inducible nitric oxide synthase (iNOS) in the azoospermic human testis. *Andrologia*. 2012;**44** Suppl 1:654–660.
- [58] Shiraishi K, Naito K, Yoshida K. Nitric oxide promotes germ cell necrosis in the delayed phase after experimental testicular torsion of rat. *Biology of Reproduction*. 2001;**65**:514–521.
- [59] Auharek SA, Avelar GF, Lara NL, Sharpe RM, Franca LR. Sertoli cell numbers and spermatogenic efficiency are increased in inducible nitric oxide synthase mutant mice. *International Journal of Andrology*. 2011;**34**:e621–629.
- [60] Jones RC. To store or mature spermatozoa? The primary role of the epididymis. *International Journal of Andrology*. 1999;**22**:57–67.
- [61] Burnett AL, Ricker DD, Chamness SL, Maguire MP, Crone JK, Bredt DS, Snyder SH, Chang TS. Localization of nitric oxide synthase in the reproductive organs of the male rat. *Biology of Reproduction*. 1995;**52**:1–7.
- [62] Luconi M, Forti G, Baldi E. Pathophysiology of sperm motility. *Frontiers in Bioscience: A Journal and Virtual Library*. 2006;**11**:1433–1447.
- [63] Curi SM, Ariagno JI, Chenlo PH, Mendeluk GR, Pugliese MN, Sardi Segovia LM, Repetto HE, Blanco AM. Asthenozoospermia: Analysis of a large population. *Archives of Andrology*. 2003;**49**:343–349.
- [64] Hellstrom WJ, Bell M, Wang R, Sikka SC. Effect of sodium nitroprusside on sperm motility, viability, and lipid peroxidation. *Fertility and Sterility*. 1994;**61**:1117–1122.



- [65] Balercia G, Moretti S, Vignini A, Magagnini M, Mantero F, Boscaro M, Ricciardo-Lamonica G, Mazzanti L. Role of nitric oxide concentrations on human sperm motility. *Journal of Andrology*. 2004;**25**:245–249.
- [66] Doshi SB, Khullar K, Sharma RK, Agarwal A. Role of reactive nitrogen species in male infertility. *Reproductive Biology and Endocrinology: RB&E*. 2012;**10**:109.
- [67] Herrero MB, Cebral E, Boquet M, Viggiano JM, Vitullo A, Gimeno MA. Effect of nitric oxide on mouse sperm hyperactivation. *Acta Physiologica, Pharmacologica et Therapeutica Latinoamericana. organo de la Asociacion Latinoamericana de Ciencias Fisiologicas y [de] la Asociacion Latinoamericana de Farmacologia*. 1994;**44**:65–69.
- [68] Rosselli M, Dubey RK, Imthurn B, Macas E, Keller PJ. Effects of nitric oxide on human spermatozoa: Evidence that nitric oxide decreases sperm motility and induces sperm toxicity. *Human Reproduction (Oxford, England)*. 1995;**10**:1786–1790.
- [69] Wu TP, Huang BM, Tsai HC, Lui MC, Liu MY. Effects of nitric oxide on human spermatozoa activity, fertilization and mouse embryonic development. *Archives of Andrology*. 2004;**50**:173–179.
- [70] Miraglia E, Rullo ML, Bosia A, Massobrio M, Revelli A, Ghigo D. Stimulation of the nitric oxide/cyclic guanosine monophosphate signaling pathway elicits human sperm chemotaxis in vitro. *Fertility and Sterility*. 2007;**87**:1059–1063.
- [71] Bolanos JP, Delgado-Esteban M, Herrero-Mendez A, Fernandez-Fernandez S, Almeida A. Regulation of glycolysis and pentose-phosphate pathway by nitric oxide: Impact on neuronal survival. *Biochimica et Biophysica Acta*. 2008;**1777**:789–793.
- [72] Kruger TF, Menkveld R, Stander FS, Lombard CJ, Van der Merwe JP, van Zyl JA, Smith K. Sperm morphologic features as a prognostic factor in in vitro fertilization. *Fertility and Sterility*. 1986;**46**:1118–1123.
- [73] Chang MC. Fertilizing capacity of spermatozoa deposited into the fallopian tubes. *Nature*. 1951;**168**:697–698.
- [74] Austin CR. The capacitation of the mammalian sperm. *Nature*. 1952;**170**:326.
- [75] Zini A, De Lamirande E, Gagnon C. Low levels of nitric oxide promote human sperm capacitation in vitro. *Journal of Andrology*. 1995;**16**:424–431.
- [76] Herrero MB, de Lamirande E, Gagnon C. Nitric oxide regulates human sperm capacitation and protein-tyrosine phosphorylation in vitro. *Biology of Reproduction*. 1999;**61**:575–581.
- [77] O'Flaherty C, de Lamirande E, Gagnon C. Positive role of reactive oxygen species in mammalian sperm capacitation: Triggering and modulation of phosphorylation events. *Free Radical Biology & Medicine*. 2006;**41**:528–540.
- [78] de Lamirande E, O'Flaherty C. Sperm activation: Role of reactive oxygen species and kinases. *Biochimica et Biophysica Acta*. 2008;**1784**:106–115.

- [79] de Lamirande E, Lamothe G. Reactive oxygen-induced reactive oxygen formation during human sperm capacitation. *Free Radical Biology and Medicine*. 2009;**46**:502–510.
- [80] Kothari S, Thompson A, Agarwal A, du Plessis SS. Free radicals: Their beneficial and detrimental effects on sperm function. *Indian Journal of Experimental Biology*. 2010;**48**:425–435.
- [81] Otasevic V, Korac A, Vucetic M, Macanovic B, Garalejic E, Ivanovic-Burmazovic I, Filipovic MR, Buzadzic B, Stancic A, Jankovic A, Velickovic K, Golic I, Markelic M, Korac B. Is manganese (ii) pentaazamacrocyclic superoxide dismutase mimic beneficial for human sperm mitochondria function and motility. *Antioxidants and Redox Signaling*. 2013;**18**:170–178.
- [82] Miraglia E, De Angelis F, Gazzano E, Hassanpour H, Bertagna A, Aldieri E, Revelli A, Ghigo D. Nitric oxide stimulates human sperm motility via activation of the cyclic GMP/protein kinase g signaling pathway. *Reproduction (Cambridge, England)*. 2011;**141**:47–54.
- [83] Donnelly ET, Lewis SE, Thompson W, Chakravarthy U. Sperm nitric oxide and motility: The effects of nitric oxide synthase stimulation and inhibition. *Molecular Human Reproduction*. 1997;**3**:755–762.
- [84] Sikka SC. Relative impact of oxidative stress on male reproductive function. *Current Medicinal Chemistry*. 2001;**8**:851–862.
- [85] Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: An update. *American Journal of Reproductive Immunology (New York, NY: 1989)*. 2008;**59**:2–11.
- [86] Petrovic V, Buzadzic B, Korac A, Vasilijevic A, Jankovic A, Korac B. No modulates the molecular basis of rat interscapular brown adipose tissue thermogenesis. *Comparative Biochemistry and Physiology. Toxicology and Pharmacology: CBP*. 2010;**152**:147–159.
- [87] Vasilijevic A, Vojcic L, Dinulovic I, Buzadzic B, Korac A, Petrovic V, Jankovic A, Korac B. Expression pattern of thermogenesis-related factors in interscapular brown adipose tissue of alloxan-treated rats: Beneficial effect of l-arginine. *Nitric Oxide: Biology and Chemistry/Official Journal of the Nitric Oxide Society*. 2010;**23**:42–50.
- [88] Vucetic M, Otasevic V, Korac A, Stancic A, Jankovic A, Markelic M, Golic I, Velickovic K, Buzadzic B, Korac B. Interscapular brown adipose tissue metabolic reprogramming during cold acclimation: Interplay of hif-1alpha and ampka. *Biochimica et Biophysica Acta*. 2011;**1810**:1252–1261.
- [89] Mishra DP, Shaha C. Estrogen-induced spermatogenic cell apoptosis occurs via the mitochondrial pathway: Role of superoxide and nitric oxide. *The Journal of Biological Chemistry*. 2005;**280**:6181–6196.

- [90] Mostafa T, Rashed LA, Nabil N, Fouad H, Sabry D, El-Saied DM. Endothelial nitric oxide synthase gene polymorphism relationship with semen parameters and oxidative stress in infertile oligoasthenoteratozoospermic men. *Urology*. 2015;**85**:1058–1061.
- [91] Buldreghini E, Mahfouz RZ, Vignini A, Mazzanti L, Ricciardo-Lamonica G, Lenzi A, Agarwal A, Balercia G. Single nucleotide polymorphism (SNP) of the endothelial nitric oxide synthase (eNOS) gene (glu298asp variant) in infertile men with asthenozoospermia. *Journal of Andrology*. 2010;**31**:482–488.
- [92] Safarinejad MR, Shafiei N, Safarinejad S. The role of endothelial nitric oxide synthase (eNOS) t-786c, g894t, and 4a/b gene polymorphisms in the risk of idiopathic male infertility. *Molecular Reproduction and Development*. 2010;**77**:720–727.
- [93] Yan L, Guo W, Wu S, Liu J, Zhang S, Shi L, Ji G, Gu A. Genetic variants in nitric oxide synthase genes and the risk of male infertility in a Chinese population: A case-control study. *PLoS One*. 2014;**9**:e115190.
- [94] Yun YJ, Park JH, Song SH, Lee S. The association of 4a4b polymorphism of endothelial nitric oxide synthase (eNOS) gene with the sperm morphology in Korean infertile men. *Fertility and Sterility*. 2008;**90**:1126–1131.
- [95] Ying HQ, Pu XY, Liu SR, A ZC. Genetic variants of eNOS gene may modify the susceptibility to idiopathic male infertility. *Biomarkers: Biochemical Indicators of Exposure, Response, and Susceptibility to Chemicals*. 2013;**18**:412–417.
- [96] Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA: A Cancer Journal for Clinicians*. 2016;**66**:7–30.
- [97] Gronberg H. Prostate cancer epidemiology. *Lancet*. 2003;**361**:859–864.
- [98] Tong X, Li H. Enos protects prostate cancer cells from trail-induced apoptosis. *Cancer Letters*. 2004;**210**:63–71.
- [99] Nanni S, Grasselli A, Benvenuti V, Aiello A, Pantisano V, Re A, Gaetano C, Capogrossi MC, Bacchetti S, Pontecorvi A, Farsetti A. The role of nuclear endothelial nitric oxide synthase in the endothelial and prostate microenvironments. *Hormone Molecular Biology and Clinical Investigation*. 2011;**5**:91–96.
- [100] Nanni S, Aiello A, Re A, Guffanti A, Benvenuti V, Colussi C, Castro-Vega LJ, Felsani A, Londono-Vallejo A, Capogrossi MC, Bacchetti S, Gaetano C, Pontecorvi A, Farsetti A. Estrogen-dependent dynamic profile of eNOS-DNA associations in prostate cancer. *PLoS One*. 2013;**8**:e62522
- [101] Estrada C, Murillo-Carretero M. Nitric oxide and adult neurogenesis in health and disease. *The Neuroscientist: A Review Journal Bringing Neurobiology, Neurology and Psychiatry*. 2005;**11**:294–307.
- [102] Burke AJ, Sullivan FJ, Giles FJ, Glynn SA. The yin and yang of nitric oxide in cancer progression. *Carcinogenesis*. 2013;**34**:503–512.

- [103] Hiraku Y, Kawanishi S, Ichinose T, Murata M. The role of iNOS-mediated DNA damage in infection- and asbestos-induced carcinogenesis. *Annals of the New York Academy of Sciences*. 2010;**1203**:15–22.
- [104] Korde Choudhary S, Sridharan G, Gadbail A, Poornima V. Nitric oxide and oral cancer: A review. *Oral Oncology*. 2012;**48**:475–483.
- [105] Vannini F, Kashfi K, Nath N. The dual role of iNOS in cancer. *Redox Biology*. 2015;**6**: 334–343.
- [106] Glynn SA, Boersma BJ, Dorsey TH, Yi M, Yfantis HG, Ridnour LA, Martin DN, Switzer CH, Hudson RS, Wink DA, Lee DH, Stephens RM, Ambs S. Increased nos2 predicts poor survival in estrogen receptor-negative breast cancer patients. *The Journal of Clinical Investigation*. 2010;**120**:3843–3854.
- [107] Lee SY, Rim Y, McPherson DD, Huang SL, Kim H. A novel liposomal nanomedicine for nitric oxide delivery and breast cancer treatment. *Bio-Medical Materials and Engineering*. 2014;**24**:61–67.
- [108] Bogdan C. Nitric oxide synthase in innate and adaptive immunity: An update. *Trends in Immunology*. 2015;**36**:161–178.
- [109] Vahora H, Khan MA, Alalami U, Hussain A. The potential role of nitric oxide in halting cancer progression through chemoprevention. *Journal of Cancer Prevention*. 2016;**21**:1–12.
- [110] Yu H, Payne TJ, Mohanty DK. Effects of slow, sustained, and rate-tunable nitric oxide donors on human aortic smooth muscle cells proliferation. *Chemical Biology & Drug Design*. 2011;**78**:527–534.
- [111] Ambs S, Hussain SP, Harris CC. Interactive effects of nitric oxide and the p53 tumor suppressor gene in carcinogenesis and tumor progression. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*. 1997;**11**:443–448.
- [112] Lechner M, Lirk P, Rieder J. Inducible nitric oxide synthase (iNOS) in tumor biology: The two sides of the same coin. *Seminars in Cancer Biology*. 2005;**15**:277–289.
- [113] Aaltoma SH, Lipponen PK, Kosma VM. Inducible nitric oxide synthase (iNOS) expression and its prognostic value in prostate cancer. *Anticancer Research*. 2001;**21**:3101–3106.
- [114] Thomas LN, Merrimen J, Bell DG, Rendon R, Too CK. Prolactin- and testosterone-induced carboxypeptidase-d correlates with increased nitrotyrosines and ki67 in prostate cancer. *The Prostate*. 2015;**75**:1726–1736.
- [115] Ng QS, Goh V, Milner J, Stratford MR, Folkes LK, Tozer GM, Saunders MI, Hoskin PJ. Effect of nitric-oxide synthesis on tumour blood volume and vascular activity: A phase i study. *The Lancet Oncology*. 2007;**8**:111–118.

- [116] Polytarchou C, Hatziapostolou M, Poimenidi E, Mikelis C, Papadopoulou A, Parthymou A, Papadimitriou E. Nitric oxide stimulates migration of human endothelial and prostate cancer cells through up-regulation of pleiotrophin expression and its receptor protein tyrosine phosphatase beta/zeta. *International Journal of Cancer*. 2009;**124**: 1785–1793.
- [117] Cianchi F, Cortesini C, Fantappie O, Messerini L, Sardi I, Lasagna N, Perna F, Fabbroni V, Di Felice A, Perigli G, Mazzanti R, Masini E. Cyclooxygenase-2 activation mediates the proangiogenic effect of nitric oxide in colorectal cancer. *Clinical Cancer Research: An Official Journal of the American Association for Cancer Research*. 2004;**10**:2694–2704.
- [118] Ozturk E, Dikensoy E, Balat O, Ugur MG, Balci SO, Aydin A, Kazanci U, Pehlivan S. Association of endothelial nitric oxide synthase gene polymorphisms with endometrial carcinoma: A preliminary study. *Journal of the Turkish German Gynecological Association*. 2011;**12**:229–233.
- [119] Lee KM, Kang D, Park SK, Berndt SI, Reding D, Chatterjee N, Chanock S, Huang WY, Hayes RB. Nitric oxide synthase gene polymorphisms and prostate cancer risk. *Carcinogenesis*. 2009;**30**:621–625.
- [120] Safarinejad MR, Safarinejad S, Shafiei N, Safarinejad S. Effects of the t-786c, g894t, and intron 4 vntr (4a/b) polymorphisms of the endothelial nitric oxide synthase gene on the risk of prostate cancer. *Urologic Oncology*. 2013;**31**:1132–1140.
- [121] Brankovic A, Brajuskovic G, Nikolic Z, Vukotic V, Cerovic S, Savic-Pavicevic D, Romac S. Endothelial nitric oxide synthase gene polymorphisms and prostate cancer risk in Serbian population. *International Journal of Experimental Pathology*. 2013;**94**:355–361.
- [122] Zhao C, Yan W, Zu X, Chen M, Liu L, Zhao S, Liu H, Hu X, Luo R, Xia Y, Qi L. Association between endothelial nitric oxide synthase 894g>t polymorphism and prostate cancer risk: A meta-analysis of literature studies. *Tumour Biology: The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014;**35**:11727–11733.
- [123] Nikolic ZZ, Pavicevic D, Romac SP, Brajuskovic GN. Genetic variants within endothelial nitric oxide synthase gene and prostate cancer: A meta-analysis. *Clinical and Translational Science*. 2015;**8**:23–31.
- [124] Wu JH, Yang K, Ma HS, Xu Y. Association of endothelial nitric oxide synthase gene rs1799983 polymorphism with susceptibility to prostate cancer: A meta-analysis. *Tumour Biology: The Journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014;**35**:7057–7062.
- [125] Cai C, Hsieh CL, Gao S, Kannan A, Bhansali M, Govardhan K, Dutta R, Shemshedini L. Soluble guanylyl cyclase alpha1 and p53 cytoplasmic sequestration and down-regulation in prostate cancer. *Molecular Endocrinology*. 2012;**26**:292–307.



- [126] Lai CS, Li S, Miyauchi Y, Suzawa M, Ho CT, Pan MH. Potent anti-cancer effects of citrus peel flavonoids in human prostate xenograft tumors. *Food and Function*. 2013;**4**:944–949.
- [127] Oktem G, Bilir A, Selvi N, Yurtseven ME, Vatansever S, Ates U, Uysal A, Omay SB. Chemotherapy influences inducible nitric oxide synthase (iNOS) and endothelial nitric oxide synthase (eNOS) activity on 3d breast cancer cell line. *Oncology Research*. 2006; **16**:195–203.
- [128] Yu S, Jia L, Zhang Y, Wu D, Xu Z, Ng CF, To KK, Huang Y, Chan FL. Increased expression of activated endothelial nitric oxide synthase contributes to antiandrogen resistance in prostate cancer cells by suppressing androgen receptor transactivation. *Cancer Letters*. 2013;**328**:83–94.
- [129] Thomas LN, Morehouse TJ, Too CK. Testosterone and prolactin increase carboxypeptidase-d and nitric oxide levels to promote survival of prostate cancer cells. *The Prostate*. 2012;**72**:450–460.
- [130] Gao Y, Li L, Zhang J, Su F, Gong Z, Lai Y, Zhang Y. Physicochemical characterization and an injection formulation study of water insoluble zcvi(4)-2, a novel no-donor anti-cancer compound. *Archives of Pharmacal Research*. 2012;**35**:1177–1186.
- [131] Donia M, Mijatovic S, Maksimovic-Ivanic D, Miljkovic D, Mangano K, Tumino S, Biondi A, Basile F, Al-Abed Y, Stosic-Grujicic S, Nicoletti F. The novel no-donating compound git-27no inhibits in vivo growth of human prostate cancer cells and prevents murine immunoinflammatory hepatitis. *European Journal of Pharmacology*. 2009; **615**: 228–233.
- [132] Baritaki S, Huerta-Yepez S, Sahakyan A, Karagiannides I, Bakirtzi K, Jazirehi A, Bonavida B. Mechanisms of nitric oxide-mediated inhibition of emt in cancer: Inhibition of the metastasis-inducer snail and induction of the metastasis-suppressor RKIP. *Cell Cycle*. 2010;**9**:4931–4940.
- [133] Bivalacqua TJ, Musicki B, Kutlu O, Burnett AL. New insights into the pathophysiology of sickle cell disease-associated priapism. *The Journal of Sexual Medicine*. 2012;**9**:79–87.
- [134] B. GPT. Inflammatory mechanisms and oxidative stress in Peyronie's disease therapeutic rationale and related emerging treatment strategies. *Inflammation and Allergy – Drug Targets*. 2012;**11**:48–57.
- [135] Shaw EJ, Mitchell GC, Tan RB, Sangkum P, Hellstrom WJ. The non-surgical treatment of Peyronie disease: 2013 update. *The World Journal of Men's Health*. 2013;**31**:183–192.
- [136] Shigehara K, Namiki M. Clinical management of priapism: A review. *The World Journal of Men's Health*. 2016;**34**:1–8.

- [137] Hurt KJ, Musicki B, Palese MA, Crone JK, Becker RE, Moriarity JL, Snyder SH, Burnett AL. Akt-dependent phosphorylation of endothelial nitric-oxide synthase mediates penile erection. *Proceedings of the National Academy of Sciences of the United States of America*. 2002;**99**:4061–4066.
- [138] Anele UA, Morrison BF, Burnett AL. Molecular pathophysiology of priapism: Emerging targets. *Current Drug Targets*. 2015;**16**:474–483.
- [139] Ishikawa T, Kondo Y, Goda K, Fujisawa M. Overexpression of endothelial nitric oxide synthase in transgenic mice accelerates testicular germ cell apoptosis induced by experimental cryptorchidism. *Journal of Andrology*. 2005;**26**:281–288.
- [140] DeFoor WR, Kuan CY, Pinkerton M, Sheldon CA, Lewis AG. Modulation of germ cell apoptosis with a nitric oxide synthase inhibitor in a murine model of congenital cryptorchidism. *The Journal of Urology*. 2004;**172**:1731–1735; Discussion 1735.