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## High-Risk Human Papillomavirus and Colorectal Carcinogenesis

Ala-Eddin Al Moustafa, Noor Al-Antary and Amber Yasmeen

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http://dx.doi.org/10.5772/63295

#### **Abstract**

Colorectal, colon and rectal, cancer is the third most common malignancy in both men and women worldwide. Colorectal carcinogenesis is a complex, multistep process implicating environmental and lifestyle factors in addition to gene mutation and viral infections. On the other hand, it is well established that human papillomaviruses (HPVs) infection play a crucial role in certain types of human carcinomas including cervical and head and neck (HN); as roughly 96% and 30% of these cancers are positive for high-risk HPVs, respectively. Moreover, it has been reported that the presence of high-risk HPVs is associated with vascular invasion, lymph node metastases, and tumor size in cervical and HN cancers. Recently, several investigations pointed-out that high-risk HPVs are present in around 70% of human colorectal cancers. Likewise, our group has demonstrated that E6/E7 oncoproteins of HPV type 16 convert noninvasive and nonmetastatic human cancer cells to invasive and metastatic form. Accordingly, it is evident that high-risk HPVs are present in human colorectal cancers where they could play an important role in the development of these malignancies. In this chapter, we will discuss the presence and role of high-risk HPVs in human colorectal carcinogenesis and metastasis; particularly, the interaction between E5 and E6/E7 oncoproteins of high-risk HPVs in colorectal malignancies, which has been linked with the initiation and progression of these tumors.

**Keywords:** colorectal cancer, high-risk HPVs, E5 & E6/E7 oncoproteins, cancer initiation, cancer progression



#### 1. Introduction

Colorectal cancers (CRCs) colon and rectal, are the most common malignancies, accounting for approximately 1.36 million new cases worldwide every year [1]. These cancers are characterized by a marked propensity for local invasion and lymph node metastases. Thus, the overall 5-year-survival rate for patients diagnosed with colorectal cancers is approximately 60% worldwide and has not significantly improved over the past decade [2]. Colorectal carcinogenesis is a complex, multistep process involving environmental, demographic, and lifestyle factors in addition to gene alterations and viral infections. The highest incidence of CRCs is observed in Western Europe, North America, Australia as well as in some Middle-Eastern countries [3, 4]. It is notable also that although the rate of this disease is relatively lower in sub-Saharan African communities, South America, and Asia; however, CRCs are gradually increasing due to assimilating life style and dietary habits of Western countries [3–5]. Additionally, around two-thirds of CRC patients will develop distant metastases during the course of their illness, which is the main cause of cancer-related death of this disease [6].

Although, human papillomaviruses (HPVs) have been established as etiological agents of invasive cervical cancer, as generally 96% of these cancers are positive for high-risk HPVs [7– 9]. However, persistent infection with high-risk HPVs is necessary but not sufficient for the development of malignant lesions [10, 11]. Furthermore, it was pointed-out that high-risk HPVs have carcinogenic effects at several other anatomical sites in women and men such as head and neck (HN) as well as colorectal [12–15]. These studies and others showed that highrisk HPVs are present in roughly 30% and 70% of HN and colorectal cancers, respectively, especially in their invasive form [14, 15]. Accordingly, we recently investigated the incidence of high-risk HPVs in CRCs in the Syrian population; our data revealed that 54% of human CRCs in Syria are positive for high-risk-HPVs; this was accompanied by an expression/ overexpression of Fascin, Id-1, and P-cadherin genes [16], which are major regulators of cell invasion and metastasis [17–19]. Meanwhile, we revealed that E5 and E6/E7 oncoproteins of high-risk HPVs could cooperate together to enhance cancer progression through the deregulation of several key controller genes of the epithelial-mesenchymal transition (EMT) event [7, 20, 21]. It is clear that CRCs and especially their invasive forms are major health problems wherein high-risk HPVs infection can play important roles in the development of these malignancies as well as their metastasis via EMT. In this chapter, we will overview the presence and contributions of high-risk HPVs in CRC initiation and progression.

#### 2. Colorectal cancers

CRCs are the most prevalent cancers worldwide, along with lung and breast cancers, they are one of the deadliest diseases today [22]. For instance, in the United States, CRCs are the third leading cause of cancer death in both sexes and the second overall in men and women combined [23, 24]. At current rates, approximately 5–6% of individuals will develop colon or rectum cancer within their lifetime [23]. These malignances are most common in Europe with

432,000 new cases reported annually in men and women combined, and the second most common cause of cancer deaths in Europe [22, 25]. In general, it is the second leading cause of cancer-related mortality worldwide and the third most commonly diagnosed malignant disease [26].

The prognosis of patients with colorectal cancer has slowly but steadily improved during the past decades in many countries. A 5-year relative survival has reached almost 65% in high-income countries, such as Australia, Canada, the USA, and several European countries, but has remained less than 50% in low-income countries [27–29]. Relative survival decreases with age, and at young ages, it is slightly higher for women than for men [30]; taking into consideration that the stage at diagnosis is the most important prognostic factor.

Colorectal carcinogenesis is common in the elderly; as approximately 90% of new colorectal cancers are diagnosed in patients over 50 years with the median age of diagnosis being 69 years. Furthermore, the incidence of CRCs dramatically rises as one ages, regardless of sex and racial background [26]. Although, it is well-known that patients with colorectal cancer may have a range of symptoms that include occult blood loss, rectal bleeding, change in stool caliber, unintentional weight loss, or have signs of bowel obstruction or perforation.

There are many risk factors for the development of colorectal cancer, one of which is colonic polyps. Pathologic entities include tubular adenoma, tubulovillous adenoma, villous adenoma, hyperplastic polyp, sessile serrated adenoma, sessile serrated polyp, and traditional serrated adenoma. In addition, some hamartomatous polyps are considered premalignant lesions [31]. Among precancerous polyps adenomatous and advanced adenomatous polyps that have polyp size >10 mm, in addition to villous/tubulovillous histological features, or having high-grade dysplasia (HGD), are found to have an increased prevalence and incidence in the elderly [26], and have a potential to progress to invasive adenocarcinomas [26, 32]. HGD is associated with larger size, villous morphology, *TP53* mutation, and deletion of a region of chromosome 18q. Chromosomal instability can be demonstrated in late precursor adenomas. In this sequence, *APC* mutation is a common early event, while the serrated lesions commonly have *BRAF* or *KRAS* mutation [31]. Other risk factors include diet and lifestyle (such as consumption of red meat, smoking, excessive alcohol, weight gain, etc.) as well as advancing age [23, 26].

For the most part, colorectal cancer arises sporadically, however, few cases are associated with inherited syndromes such as familial adenomatous polyposis (FAP; <1% of CRC) where patients exhibit germline mutations in one allele of the adenomatous polyposis (*APC*) tumor suppressor gene, MUTYH-associated polyposis (MAP; rare recessive condition, carrier estimated at ~1%), and Lynch syndrome/hereditary nonpolyposis colon cancer (LS/HNPCC; 2–4% of CRCs) [7, 33].

The usual malignant tumor of the large bowel is a well-to-moderately differentiated adenocarcinoma secreting variable amounts of mucin [34]. In World Health Organization (WHO) classification, a number of histologic variants of this tumor are listed, such as mucinous adenocarcinoma, signet ring cell, medullary, micropapillary, serrated, cribriform comedotype, adenosquamous, spindle cell, and undifferentiated. The most widely used immunohistochemical markers for colorectal adenocarcinoma are cytokeratin (CK) 20, CK7, and CDX2. The most common immunophenotype of colorectal adenocarcinoma is positivity for CK20 and negativity for CK7 [35]. The CRCs are divided in to four grades. G1 are well-differentiated tumors (usually adenocarcinomas) that have more than 95% glandular structures. Further, G2 are designated as moderately differentiated tumors with 50–95% gland formation. G3 are poorly differentiated tumors with 5–50% gland formation; whereas G4 are highly aggressive and undifferentiated tumors with less than 5% gland formation. Recently, WHO also suggests dividing CRCs into low grade (G1 and G2) and high grade (G3 and G4) categories. The diagnosis of G3 and G4 is relatively consistent, but differentiation between G1 and G2 is associated with a significant degree of inter-observer variability [36, 37].

As we mentioned above, CRCs are characterized by a marked propensity for invasion and metastasis. About 20% of patients with newly diagnosed colorectal cancer present with distant metastases [38, 39]. The most common location is the liver [38, 40]; however, investigators identified lung metastases in 2·1% of patients newly diagnosed with CRC in a large cancer registry in France [41]. Frequency was nearly three times higher for patients with rectal cancer than for patients with colon cancer. Smaller studies [42–44] have shown isolated lung metastases in 9–18% of patients with rectal cancer; although distant metastases can be identified in other organs including the bone and the brain [38].

As we cited above, lifetime risk of CRCs is estimated to be 5–6% in the general population of Western countries [45, 46]. Although hereditary forms of CRC have been well established; however, most cases are sporadic [47]. Numerous epidemiological studies have identified lifestyle and environmental factors contributing to the occurrence of CRCs [48, 49]. In the past decades, *Helicobacter pylori* and Epstein Barr virus infections have been identified as potential causal factors of gastric cancer [50, 51] and personal communication. A number of studies aimed to assess the possible role of viral infections, such as infection with high-risk HPVs, human polyomaviruses, and human herpesviruses in colorectal carcinogenesis [7, 45, 52, 53]. Thus, in the next paragraph the presence and role of high-risk HPVs in human CRCs will be reviewed.

#### 3. Human papillomaviruses (HPVs)

Papillomaviruses were first identified in rabbits in 1933, and they were found to be involved in transmissible growth of benign papillomas [54]. HPVs were first identified in 1956, and they were associated with a variety of benign growths in humans [55]. However, it was later observed that HPVs, a highly prevalent sexually transmitted infection, have potentially serious health consequences in males and females. HPV infections have received considerable attention in recent years. So far, more than 150 HPV types have been isolated and characterized. While the involvement of HPV in causing benign warts was already known, the first evidence of the association between human cancer and certain HPV types was proposed more than thirty years ago by zur Hausen and his colleagues [56].

The common mode of transmission and acquisition of HPV is by horizontal transmission consequent to sexual activity. Occasionally, HPV may be transmitted through modes other

than sexual activity [57–61]. Thus, prevalence sites of HPVs include the epithelium of the vagina, vulva, penis, anal canal, cervix, perianal region, crypts of the tonsils, and oropharynx. Persistent HPV infection is essential for the development of cervical precancerous lesions and cancer. However, this may take a long time, usually a decade or more after the initial infection [62].

HPVs are small, double-stranded DNA viruses that generally infect cutaneous and mucosal epithelial tissues of the anogenital tract. The HPV DNA genome encodes approximately eight open reading frames (ORFs) [52, 62]. It is divided into three functional parts: the early (E) region, the late (L) region, and a long control region (LCR). The E region is important for replication, cellular transformation, and for the control of viral transcription, whereas the L region encodes the structural proteins (L1-L2) that take part in assembly [12]. The LCR is necessary for viral DNA replication and transcription. The seven proteins of the E region are E1, E2, E3, E4, E5, E6, and E7. E1 is necessary for viral DNA replication, while E2 has a role in viral gene transcription and replication. The function of E3 is still not understood. On the other hand, E4 protein interacts with the keratin cytoskeleton and intermediate filaments. Moreover, it facilitates virus assembly and release. The E5 protein interacts with the receptors of growth factors and stimulates cellular proliferation and inhibits apoptosis. E6 induces DNA synthesis, prevents cell differentiation, and interacts with tumor suppressor proteins and repair factors. In fact, E7 induces cell proliferation and interacts with negative regulators of cell cycle and tumor suppressor proteins. E5, E6, and E7 proteins act as oncogenes which are associated with carcinogenesis [12, 20, 63-66] (please see below).

As we mentioned above, over 150 different viral types have been identified, and about one-third of these infect epithelial cells in the genital tract [67]. HPVs are classified as either high risk or low risk. Infections with low-risk types are generally self-limiting and do not lead to malignancy. However, infections with high-risk HPVs (type 16, 18, 31, 33, 35, 39, 45, 51, 52, 55, 56, 58, 59, 68, 73, 82, and 83) are associated with the development of cervical cancers since more than 96% of these cancers are positive for high-risk HPVs [7, 9, 68–70].

It is well known that high-risk HPV early proteins, including E5, E6, and E7 oncoproteins, increase cellular alteration and probably lead to HPV induced carcinogenesis [20, 71–73]. More specifically, the E5 oncoprotein interacts with EGF-R1 signaling pathways (MAP Kinase and P13K-Akt) and proapoptotic proteins [74–76]; and therefore, it can play an important role in cell transformation and tumor formation. On the other hand, E6 and E7 of the high-risk HPV types, such as HPV16, are thought to work together in lesions caused by this virus, since, the two proteins are expressed from bicistronic mRNA [77] and initiated from the viral early promoter (p97). These proteins have functions that stimulate cell cycle progression and both can associate with regulators of the cell cycle [70, 72, 78].

Several studies have shown that the viral E6 protein complements the role of E7 and is thought to prevent the induction of apoptosis in response to unscheduled S-phase entry mediated by E7 [70, 79]. The E6 protein is also involved in the inactivation of p53-mediated growth suppression and/or apoptosis and can also associate with other proapoptotic proteins including Bak [80] and Bax [81]. In addition, E6 stimulates cell proliferation independently from E7 through its C-terminal PDZ-ligand domain [70, 82]. E6-PDZ binding is sufficient to mediate

suprabasal cell proliferation [83, 84] and may contribute to the development of metastatic tumours by disrupting normal cell adhesion. On the other hand, the E7 viral is involved with members of the pocket protein family such as pRb, which is well documented. E7 binding to pRb displaces E2F, irrespective of the presence of external growth factors and leads to the expression of proteins necessary for DNA replication [70, 71, 78, 85].

To address the role of E6/E7 genes in high-risk HPV-associated carcinogenesis *in vivo*, transgenic mice have been developed expressing E6/E7 of HPV type 16 individually and together under the human K14 promoter [86, 87]. These transgenic mice developed skin tumors, in general, and cervical cancer with chronic estrogen administration [87, 88]. On the other hand, and to examine the oncogenic properties of E5 *in vivo*, K14-E5 transgenic mice were generated in which the expression of E5 was directed to the basal layer of the stratified squamous epithelia. These mice exhibited the epidermal hyperplasia, aberrant differentiation of the epithelium, and were susceptible to spontaneous skin tumors [89]. Recently, it was reported that K14-E6/E7 transgenic mice have high susceptibility to colorectal cancers and precancerous lesions after dimethylbenz[a]anthracene-treatment, which is a chemical carcinogen that is known to induce squamous cell carcinomas in other sites [90]. These studies show clearly that high-risk HPVs play an important role in cancer initiation and/or progression of several anatomical sites, which could include colorectal, through their E5, E6, and E7 onco-proteins.

#### 4. High-risk HPVs in colorectal cancers

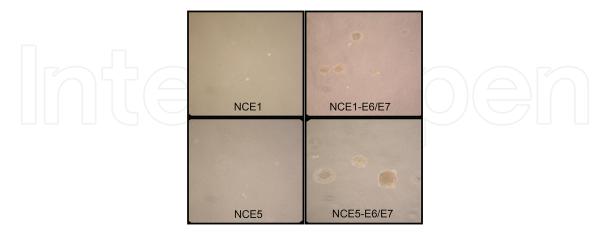
High-risk HPVs have been established as etiological agents of invasive cervical cancer, as more than 96% of these cancers are positive for high-risk HPVs which are the most common viral sexually transmitted infection worldwide [7–9]. Infection with high-risk HPVs is important for the development of premalignant lesions and/or progression of the disease [10, 11]. Additionally, it was revealed that high-risk HPVs have carcinogenic effects at several other anatomical regions in women and men such as HN as well as colorectal [12–15]. These studies showed that high-risk HPVs are present in roughly 30 and 70% of HN and colorectal cancers, respectively, especially in their invasive form [14, 15]. Therefore, several recent studies including one from our group pointed-out that high-risk HPVs are present in human CRCs, specifically types 16, 18, 31, 33, and 35 [7, 12, 15, 16, 52]. Moreover, six recent meta-analysis studies confirmed the presence of high-risk HPVs in human CRCs [70, 91–95]; however, the prevalence of high-risk HPVs varied from one geographic location to another [7, 52]. Meanwhile, it was stated that high-risk HPVs are present especially in the invasive form of these malignancies worldwide [15].

Nevertheless, it is important to mention that high-risk HPV infection alone is not sufficient to induce neoplastic transformation of human normal epithelial cells; the infected cells must undergo additional genetic changes and/or coinfection with another oncovirus to reach full transformation and consequently tumor formation. Based on this fact, we have developed a new model to study the cooperation effect between high-risk HPVs and other oncogenes in

human carcinogenesis using human normal epithelial (HNE) cells. In this model, we established that E6/E7 oncoproteins of high-risk type 16 cooperate with the ErbB-2 receptor to induce cellular transformation of HNE cells; this was accompanied by a delocalization of  $\beta$ -catenin from the undercoat membrane to the nucleus in HNE cells. Furthermore, we reported that cyclin D1 is the target of E6/E7/ErbB-2 cooperation via the conversion of  $\beta$ -catenin's role from a cell–cell adhesion molecule to a transcriptional regulator [96]. In parallel, we revealed that D-type cyclins (D1, D2, and D3) are essential for cell transformation induced by E6/E7/ErbB-2 cooperation in human HNE and mouse normal embryonic fibroblast (NEF) cells [96, 97]. Finally, we were able to show that the cooperation effect of E6/E7 with ErbB-2, in human normal epithelial and cancer cells, occurs via  $\beta$ -catenin tyrosine phosphorylation through pp60 (c-Src) kinase activation [98, 99]. Thus, the cooperation between E6/E7 oncoproteins of high-risk HPVs and other oncogenes could occur in colorectal carcinogenesis.

On the other hand, and to determine the role of high-risk HPVs infection in human cancer cells, we examined the effect of E6/E7 of HPV type 16 in two noninvasive human breast cancer cell lines. We reported that E6/E7 of HPV type 16 induce cell invasive and metastatic abilities of the two cell lines *in vitro* and *in vivo*, respectively, in comparison with their wild-type cells [100]. This is accompanied by an overexpression of Id-1, a family member of helix-loop-helix transcription factors which regulates cell invasion and metastasis of human cancer cells [101, 102]. We also demonstrated that E6/E7 oncoproteins upregulate Id-1 promoter activity in human cancer cells. These data suggest that high-risk HPVs could play an important role in the progression of human carcinomas via Id-1 deregulation. Thus, we believe that E6/E7 oncoproteins of high-risk HPVs could play a similar role in the progression of human CRCs.

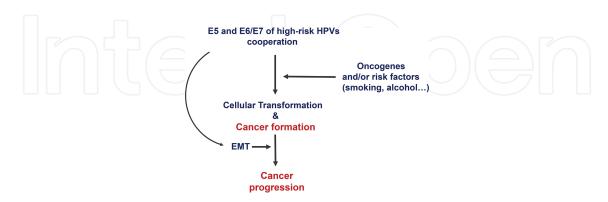
In order to investigate the role of high-risk HPV infection in human colorectal carcinogenesis, we examined the effect of E6/E7 of HPV type 16 in two human primary normal colorectal "mesenchymal" cell lines, NCM1 and NCM5, which were established in our laboratory [20].



**Figure 1.** E6/E7 oncoproteins of high-risk HPV type 16 induce cellular transformation in human primary normal color-ectal "mesenchymal" cell lines, NCE1, and NCE5 cells [103]. We note that NCE1 and NCE5 cells are unable to grow in soft agar. In contrast, NCE1 and NCE5 cells expressing E6/E7 oncoproteins form colonies in soft agar assay, which is an important characteristic of cancer cells.

We found that the expression of E6/E7 oncoproteins stimulate cell proliferation and induce cellular transformation (**Figure 1**) and migration of NCM1 and NCM5 cell lines. Moreover, our data revealed that E6/E7 of HPV type 16 provoke the upregulation of D-type cyclins and Cyclin E as well as Id-1 in these cell lines [103]. It is important to highlight that there are no other studies regarding the role of E6/E7 oncoproteins of high-risk HPVs in human colorectal cancers. Meanwhile, the function of E5 oncoprotein, in these malignancies, has not been investigated yet.

Additionally, we have recently investigated the incidence of high-risk HPVs in human CRCs in the Syrian population in a cohort of 78 cancer samples using PCR and tissue microarray analyses. We reported, for the first time, that high-risk HPVs are present in 42 samples (53.84%), which represent the majority of invasive colorectal cases; more significantly, our data pointedout that the most frequent high-risk HPV types in the Syrian population are 16, 33, 18, 35, and 31, respectively. Furthermore, the expression of E6 oncoprotein of high-risk HPVs was found to be correlated with Fascin, Id-1, and P-cadherin expression/overexpression in the majority of cancer tissue samples, which are major regulators of cell invasion and metastasis [17-19, 52]. Our data imply that high-risk HPVs are present in human CRCs, and their presence is associated with invasive and metastatic phenotype [16, 52, 104]. Collectively, these data suggest that high-risk HPVs are present in CRCs and therefore could play an important role in the initiation and progression of these cancers. Thus, we believe that high-risk HPVs can be associated with a subset of colorectal cancers. However, future large-scale multicenter casecontrol studies with data on risk factors such as lifestyle and sexual behavior are needed; meanwhile, molecular and cellular studies are necessary to determine the role of E5 and E6/E7 oncoproteins in human colorectal cancer and normal cells since it was proposed that E5 can cooperate with E6/E7 oncoproteins to enhance cancer progression of other human malignancies via the EMT event [20, 52]. Thus, we believe that E5 and E6/E7 of high-risk HPVs can cooperate with other oncogenes and/or risk factors such as smoking or alcohol to initiate colorectal cancer; in addition, E5 could cooperate with E6/E7 to enhance cancer progression of this malignancy via the EMT event (Figure 2).



**Figure 2.** E5 and E6/E7 of high-risk HPVs cooperation and colorectal carcinogenesis. We believe that E5 and E6/E7 of high-risk HPVs can cooperate with other oncogene overexpressions that are linked to lifestyle or/and environmental factors to induce cellular transformation and consequently tumor formation. On the other hand, E5 and E6/E7 together can enhance cancer progression of colorectal cancer via the initiation of the epithelial-mesenchymal transition (EMT) event .

Finally, we think it is important to talk about the prevention strategy of HPV infections and their related cancers, which is essentially based on HPV vaccines. These vaccines are made of virus-like particles (VLPs) that contain inactive L1 HPV proteins—proteins from and specific to each type of HPV viruses [105, 106]. Thus, the quadrivalent vaccine Gardasil (Merck and Co) was developed and approved by the FDA in 2006 for protection against low-risk HPV types 6 and 11, which cause genital warts—and rarely, nongenital warts [107] and high-risk HPV types 16 and 18 [108]. The quadrivalent vaccine will not protect against anogenital disease other than HPV types 6, 11, 16, and 18 [109, 110]. In 2010, the FDA approved the quadrivalent vaccine for the prevention of CRC [106]. The efficacy of prevention of rectal intraepithelial neoplasia in some group of patients is 77.5% [111]. In 2009, a bivalent vaccine (Cervarix; GlaxoSmithKline) was approved for the prevention of HPV infections from high-risk types 16 and 18 [112]. On December 10, 2014, the FDA approved a 9-valent HPV vaccine (Gardasil-9; Merck and Co) that was approved to be given in three intramuscular doses to males 9–15 years of age and females 9-26 years of age [106, 113]. The 9-valent HPV vaccine targets high-risk HPV type 16 (responsible for 50% of cervical carcinogenesis) [114], high-risk HPV type 18 (detected in 20% of cervical cancers) [115], and types 31, 33, 45, 52, and 58, which are responsible for 25% of cervical cancers. Immunizations against low-risk HPV types 6 and 11, which cause genital warts, are also included in the 9-valent vaccine [106, 116]. Approval of the 9-valent vaccine was based on a randomized control study with 14,000 females 16-26 years of age; it noted efficacy of 97% [106]. Therefore, this vaccine will have an important role in preventing HPV infections and their related cancers including colorectal malignancies and their metastasis.

#### 5. Conclusion and perspectives

This chapter presented substantial evidence that high-risk HPVs are present in human CRCs, thereby these viruses, through their E5 and E6/E7 oncoproteins, could play an important role in the initiation and progression of these malignances (Figure 2) [20, 100]. However, we believe that further studies are required to determine the function of E5 and E6/E7 oncogenes in human colorectal normal and cancer cells. Thus, developing new *in vitro* and *in vivo* models, such as cell lines and animal models, are necessary to identify the exact role of these oncoproteins and their potential cooperation in human colorectal carcinogenesis. Such studies can lead to the discovery of new targets to manage these malignances and other human carcinomas related to high-risk HPVs.

Alternatively, and with regards to colorectal malignancies as well as other human carcinomas prevention, we assume that the elimination of a number of known risk factors especially unprotected sexual activity, physical inactivity, smoking, alcohol, high consumption of red meat, and oncovirus infections such as high-risk HPVs could diminish the development of these malignancies and their metastases [20, 23, 26]. Additionally, prevention methodologies of high-risk HPVs using presently available vaccines could greatly reduce high-risk HPVs-associated cancers, including colorectal, and their progression to invasive form, which is responsible for the majority of cancer-related deaths.

#### Acknowledgements

We are grateful to Mrs. A. Kassab for her critical reading of the chapter. The research works from Dr. Al Moustafa's laboratory is supported by the College of Medicine of Qatar University.

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#### References

- [1] Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. International Journal of Cancer. 2015;136:E359–86.
- [2] Rose J, Augestad KM, Cooper GS. Colorectal cancer surveillance: what's new and what's next. World Journal of Gastroenterology. 2014;20:1887–97.
- [3] Beg S, Siraj AK, Prabhakaran S, Bu R, Al-Rasheed M, et al. Molecular markers and pathway analysis of colorectal carcinoma in the Middle East. Cancer. 2015;121:3799–808.
- [4] Tezcan G, Tunca B, Ak S, Cecener G, Egeli U. Molecular approach to genetic and epigenetic pathogenesis of early onset colorectal cancer. World Journal of Gastrointestinal Oncology. 2016;8:83–98.
- [5] Vainio H, Miller AB. Primary and secondary prevention in colorectal cancer. Acta Oncologica. 2003;42:809–15.
- [6] Mattar RE, Al-Alem F, Simoneau E, Hassanain M. Preoperative selection of patients with colorectal cancer liver metastasis for hepatic resection. World Journal of Gastroenterology. 2016;22:567–81.

- [7] Al Moustafa AE, Al-Awadhi R, Missaoui N, Adam I, Durusoy R, et al. Human papillomaviruses-related cancers. Presence and prevention strategies in the Middle east and north African regions. Human Vaccines & Immunotherapeutics. 2014a;10:1812—21.
- [8] de Sanjose S, Diaz M, Castellsague X, Clifford G, Bruni L, et al. Worldwide prevalence and genotype distribution of cervical human papillomavirus DNA in women with normal cytology: a meta-analysis. The Lancet Infectious Diseases. 2007;7:453–9.
- [9] Smith JS, Lindsay L, Hoots B, Keys J, Franceschi S, et al. Human papillomavirus type distribution in invasive cervical cancer and high-grade cervical lesions: a meta-analysis update. International Journal of Cancer. 2007;121:621–32.
- [10] Begum S, Gillison ML, Ansari-Lari MA, Shah K, Westra WH. Detection of human papillomavirus in cervical lymph nodes: a highly effective strategy for localizing site of tumor origin. Clinical Cancer Research: An Official Journal of the American Association for Cancer Research. 2003;9:6469–75.
- [11] Graflund M, Sorbe B, Sigurdardottir S, Karlsson M. HPV-DNA, vascular space invasion, and their impact on the clinical outcome in early-stage cervical carcinomas. International Journal of Gynecological Cancer: Official Journal of the International Gynecological Cancer Society. 2004;14:896–902.
- [12] Abramowitz L, Jacquard AC, Jaroud F, Haesebaert J, Siproudhis L, et al. Human papillomavirus genotype distribution in anal cancer in France: the EDiTH V study. International Journal of Cancer. 2011;129:433–9.
- [13] Shukla S, Bharti AC, Mahata S, Hussain S, Kumar R, et al. Infection of human papillomaviruses in cancers of different human organ sites. The Indian Journal of Medical Research. 2009;130:222–33.
- [14] Umudum H, Rezanko T, Dag F, Dogruluk T. Human papillomavirus genome detection by in situ hybridization in fine-needle aspirates of metastatic lesions from head and neck squamous cell carcinomas. Cancer. 2005;105:171–7.
- [15] Varnai AD, Bollmann M, Griefingholt H, Speich N, Schmitt C, et al. HPV in anal squamous cell carcinoma and anal intraepithelial neoplasia (AIN). Impact of HPV analysis of anal lesions on diagnosis and prognosis. International Journal of Colorectal Disease. 2006;21:135–42.
- [16] Ghabreau L, Segal ED, Yasmeen A, Kassab A, Akil N, et al. High-risk human papillomavirus infections in colorectal cancer in the Syrian population and their association with Fascin, P-cadherin and Id-1 expressions: a tissue microarray study. Clinical Cancer Investigation 2012; J 1: 26–30.
- [17] Ling MT, Wang X, Zhang X, Wong YC. The multiple roles of Id-1 in cancer progression. Differentiation; Research in Biological Diversity. 2006;74:481–7.

- [18] Oh SY, Kim YB, Suh KW, Paek OJ, Moon HY. Prognostic impact of fascin-1 expression is more significant in advanced colorectal cancer. The Journal of Surgical Research. 2012;172:102–8.
- [19] Van Marck V, Stove C, Jacobs K, Van den Eynden G, Bracke M. P-cadherin in adhesion and invasion: opposite roles in colon and bladder carcinoma. International Journal of Cancer. 2011;128:1031–44.
- [20] Al Moustafa AE. E5 and E6/E7 of high-risk HPVs cooperate to enhance cancer progression through EMT initiation. Cell Adhesion & Migration. 2015;9:392–3.
- [21] Yasmeen A, Alachkar A, Dekhil H, Gambacorti-Passerini C, Al Moustafa AE. Locking Src/Abl tyrosine kinase activities regulate cell differentiation and invasion of human cervical cancer cells expressing E6/E7 oncoproteins of high-risk HPV. Journal of Oncology. 2010;2010. doi: 10.1155/2010/530130
- [22] Binefa G, Rodriguez-Moranta F, Teule A, Medina-Hayas M. Colorectal cancer: from prevention to personalized medicine. World Journal of Gastroenterology. 2014;20:6786–808.
- [23] Chan AT, Giovannucci EL. Primary prevention of colorectal cancer. Gastroenterology. 2010;138:2029–43 e10.
- [24] Schoen RE, Pinsky PF, Weissfeld JL, Yokochi LA, Church T, et al. Colorectal-cancer incidence and mortality with screening flexible sigmoidoscopy. The New England Journal of Medicine. 2012;366:2345–57.
- [25] European Colorectal Cancer Screening Guidelines Working G, von Karsa L, Patnick J, Segnan N, Atkin W, Halloran S, et al. European guidelines for quality assurance in colorectal cancer screening and diagnosis: overview and introduction to the full supplement publication. Endoscopy. 2013;45:51–9.
- [26] Day LW, Velayos F. Colorectal cancer screening and surveillance in the elderly: updates and controversies. Gut and Liver. 2015;9:143–51.
- [27] Butler EN, Chawla N, Lund J, Harlan LC, Warren JL, et al. Patterns of colorectal cancer care in the United States and Canada: a systematic review. Journal of the National Cancer Institute Monographs. 2013;2013:13–35.
- [28] Marin-Gabriel JC, Fernandez-Esparrach G, Diaz-Tasende J, Herreros de Tejada A. Colorectal endoscopic submucosal dissection from a Western perspective: today's promises and future challenges. World Journal of Gastrointestinal Endoscopy. 2016;8:40–55.
- [29] Sankaranarayanan R. Screening for cancer in low- and middle-income countries. Annals of Global Health. 2014;80:412–7.
- [30] Ballester V, Rashtak S, Boardman L. Clinical and molecular features of young-onset colorectal cancer. World Journal of Gastroenterology. 2016;22:1736–44.

- [31] Lochhead P, Chan AT, Giovannucci E, Fuchs CS, Wu K, et al. Progress and opportunities in molecular pathological epidemiology of colorectal premalignant lesions. The American Journal of Gastroenterology. 2014;109:1205–14.
- [32] Grady WM, Pritchard CC. Molecular alterations and biomarkers in colorectal cancer. Toxicologic Pathology. 2014;42:124–39.
- [33] Sengupta N, Yau C, Sakthianandeswaren A, Mouradov D, Gibbs P, et al. Analysis of colorectal cancers in British Bangladeshi identifies early onset, frequent mucinous histotype and a high prevalence of RBFOX1 deletion. Molecular Cancer. 2013;12:1.
- [34] Rosi J, Ackerman L. Surgical Pathology, Mosby Elsevier editors. 10th edition; 2011.
- [35] Fleming M, Ravula S, Tatishchev SF, Wang HL. Colorectal carcinoma: pathologic aspects. Journal of Gastrointestinal Oncology. 2012;3:153–73.
- [36] Maguire A, Sheahan K. Controversies in the pathological assessment of colorectal cancer. World Journal of Gastroenterology. 2014;20:9850–61.
- [37] Al Moustafa AE. Role of high risk human papillomavisus in breast carcinogenesis. In: Satya Prakash Gupta editors. Book Cancer causing Viruses and their Inhibitors. Taylor and Francis Group, CRC Press; Germany, 2014, p. 245–262. ISBN 9781466589773
- [38] Brenner H, Kloor M, Pox CP. Colorectal Cancer. Lancet. 2014;383:1490–502.
- [39] Leufkens AM, van den Bosch MA, van Leeuwen MS, Siersema PD. Diagnostic accuracy of computed tomography for colon cancer staging: a systematic review. Scandinavian Journal of Gastroenterology. 2011;46:887–94.
- [40] Niekel MC, Bipat S, Stoker J. Diagnostic imaging of colorectal liver metastases with CT, MR imaging, FDG PET, and/or FDG PET/CT: a meta-analysis of prospective studies including patients who have not previously undergone treatment. Radiology. 2010;257:674–84.
- [41] Mitry E, Guiu B, Cosconea S, Jooste V, Faivre J, et al. Epidemiology, management and prognosis of colorectal cancer with lung metastases: a 30-year population-based study. Gut. 2010;59:1383–8.
- [42] Choi DJ, Kwak JM, Kim J, Woo SU, Kim SH. Preoperative chest computerized tomography in patients with locally advanced mid or lower rectal cancer: its role in staging and impact on treatment strategy. Journal of Surgical Oncology. 2010;102:588–92.
- [43] Kirke R, Rajesh A, Verma R, Bankart MJ. Rectal cancer: incidence of pulmonary metastases on thoracic CT and correlation with T staging. Journal of computer assisted tomography. 2007;31:569–71.
- [44] Tan KK, Lopes Gde L, Jr., Sim R. How uncommon are isolated lung metastases in colorectal cancer? A review from database of 754 patients over 4 years. Journal of Gastrointestinal Surgery: Official Journal of the Society for Surgery of the Alimentary Tract. 2009;13:642–8.

- [45] Chen H, Chen XZ, Waterboer T, Castro FA, Brenner H. Viral infections and colorectal cancer: a systematic review of epidemiological studies. International Journal of Cancer. 2015;137:12–24.
- [46] Jemal A, Bray F, Center MM, Ferlay J, Ward E, et al. Global cancer statistics. CA: A Cancer Journal for Clinicians. 2011;61:69–90.
- [47] Lynch HT, de la Chapelle A. Hereditary colorectal cancer. The New England Journal of Medicine. 2003;348:919–32.
- [48] Aykan NF. Red meat and colorectal cancer. Oncology Reviews. 2015;9:288.
- [49] Liang PS, Chen TY, Giovannucci E. Cigarette smoking and colorectal cancer incidence and mortality: systematic review and meta-analysis. International Journal of Cancer. 2009;124:2406–15.
- [50] Collins D, Hogan AM, Winter DC. Microbial and viral pathogens in colorectal cancer. The Lancet Oncology. 2011;12:504–12.
- [51] Wu Q, Yang ZP, Xu P, Gao LC, Fan DM. Association between Helicobacter pylori infection and the risk of colorectal neoplasia: a systematic review and meta-analysis. Colorectal Disease: The Official Journal of the Association of Coloproctology of Great Britain and Ireland. 2013;15:e352–64.
- [52] Al Moustafa AE, Ghabreau L, Akil N, Rastam S, Alachkar A, et al. High-risk HPVs and human carcinomas in the Syrian population. Frontiers in Oncology. 2014b;4:68.
- [53] Coelho TR, Gaspar R, Figueiredo P, Mendonca C, Lazo PA, et al. Human JC polyomavirus in normal colorectal mucosa, hyperplastic polyps, sporadic adenomas, and adenocarcinomas in Portugal. Journal of Medical Virology. 2013;85:2119–27.
- [54] Shope RE, Hurst EW. Infectious papillomatosis of rabbits: with a note on the histopathology. The Journal of Experimental Medicine. 1933;58:607–24.
- [55] Moore PS, Chang Y. Why do viruses cause cancer? Highlights of the first century of human tumour virology. Nature Reviews Cancer. 2010;10:878–89.
- [56] zur Hausen H. Papillomaviruses and cancer: from basic studies to clinical application. Nature Reviews Cancer. 2002;2:342–50.
- [57] Edwards S, Carne C. Oral sex and the transmission of viral STIs. Sexually Transmitted Infections. 1998;74:6–10.
- [58] Frega A, Cenci M, Stentella P, Cipriano L, De Ioris A, et al. Human papillomavirus in virgins and behaviour at risk. Cancer Letters. 2003;194:21–4.
- [59] Gervaz P, Allal AS, Villiger P, Buhler L, Morel P. Squamous cell carcinoma of the anus: another sexually transmitted disease. Swiss Medical Weekly. 2003;133:353–9.

- [60] Ho GY, Bierman R, Beardsley L, Chang CJ, Burk RD. Natural history of cervicovaginal papillomavirus infection in young women. The New England Journal of Medicine. 1998;338:423–8.
- [61] Sonnex C, Strauss S, Gray JJ. Detection of human papillomavirus DNA on the fingers of patients with genital warts. Sexually Transmitted Infections. 1999;75:317–9.
- [62] Prabhu SR, Wilson DF. Human papillomavirus and oral disease—emerging evidence: a review. Australian Dental Journal. 2013;58:2–10; quiz 125.
- [63] Antonsson A, Forslund O, Ekberg H, Sterner G, Hansson BG. The ubiquity and impressive genomic diversity of human skin papillomaviruses suggest a commensalic nature of these viruses. Journal of Virology. 2000;74:11636–41.
- [64] Hiller T, Iftner T. The human papillomavirus. In: Prendiville W, Davies P, editors. Human Papillomavirus and Cervical Cancer. The Health Professional's HPV Handbook. London and New York: Taylor & Francis; 2004, p. 11–26.
- [65] de Villiers EM, Fauquet C, Broker TR, Bernard HU, zur Hausen H. Classification of papillomaviruses. Virology. 2004;324:17–27.
- [66] Mammas IN, Sourvinos G, Spandidos DA. The paediatric story of human papillomavirus (Review). Oncology Letters. 2014;8:502–6.
- [67] Bernard HU. The clinical importance of the nomenclature, evolution and taxonomy of human papillomaviruses. Journal of Clinical Virology: The Official Publication of the Pan American Society for Clinical Virology. 2005;32 Suppl 1:S1–6.
- [68] Castellsague X, Diaz M, de Sanjose S, Munoz N, Herrero R, et al. Worldwide human papillomavirus etiology of cervical adenocarcinoma and its cofactors: implications for screening and prevention. Journal of the National Cancer Institute. 2006;98:303–15.
- [69] Zuna RE, Allen RA, Moore WE, Mattu R, Dunn ST. Comparison of human papillomavirus genotypes in high-grade squamous intraepithelial lesions and invasive cervical carcinoma: evidence for differences in biologic potential of precursor lesions. Modern Pathology: An Official Journal of the United States and Canadian Academy of Pathology, Inc. 2004;17:1314–22.
- [70] Abbas A, Yang G, Fakih M. Management of anal cancer in 2010. Part 1: overview, screening, and diagnosis. Oncology. 2010;24:364–9.
- [71] Doorbar J. Latent papillomavirus infections and their regulation. Current Opinion in Virology. 2013;3:416–21.
- [72] Grm HS, Massimi P, Gammoh N, Banks L. Crosstalk between the human papillomavirus E2 transcriptional activator and the E6 oncoprotein. Oncogene. 2005;24:5149–64.
- [73] Yuan CH, Filippova M, Duerksen-Hughes P. Modulation of apoptotic pathways by human papillomaviruses (HPV): mechanisms and implications for therapy. Viruses. 2012;4:3831–50.

- [74] Kim SH, Juhnn YS, Kang S, Park SW, Sung MW, et al. Human papillomavirus 16 E5 up-regulates the expression of vascular endothelial growth factor through the activation of epidermal growth factor receptor, MEK/ ERK1,2 and PI3K/Akt. Cellular and Molecular Life Sciences: CMLS. 2006;63:930–8.
- [75] Oh JM, Kim SH, Cho EA, Song YS, Kim WH, et al. Human papillomavirus type 16 E5 protein inhibits hydrogen-peroxide-induced apoptosis by stimulating ubiquitin-proteasome-mediated degradation of Bax in human cervical cancer cells. Carcinogenesis. 2010;31:402–10.
- [76] Suprynowicz FA, Disbrow GL, Krawczyk E, Simic V, Lantzky K, et al. HPV-16 E5 oncoprotein upregulates lipid raft components caveolin-1 and ganglioside GM1 at the plasma membrane of cervical cells. Oncogene. 2008;27:1071–8.
- [77] Stacey SN, Jordan D, Williamson AJ, Brown M, Coote JH, et al. Leaky scanning is the predominant mechanism for translation of human papillomavirus type 16 E7 oncoprotein from E6/E7 bicistronic mRNA. Journal of Virology. 2000;74:7284–97.
- [78] Munger K, Basile JR, Duensing S, Eichten A, Gonzalez SL, et al. Biological activities and molecular targets of the human papillomavirus E7 oncoprotein. Oncogene. 2001;20:7888–98.
- [79] Ghittoni R, Accardi R, Hasan U, Gheit T, Sylla B, Tommasino M. The biological properties of E6 and E7 oncoproteins from human papillomaviruses. Virus Genes. 2010;40:1–13.
- [80] Thomas M, Banks L. Inhibition of Bak-induced apoptosis by HPV-18 E6. Oncogene. 1998;17:2943–54.
- [81] Magal SS, Jackman A, Ish-Shalom S, Botzer LE, Gonen P, et al. Downregulation of Bax mRNA expression and protein stability by the E6 protein of human papillomavirus 16. The Journal of General Virology. 2005;86:611–21.
- [82] Thomas M, Laura R, Hepner K, Guccione E, Sawyers C, Lasky L, et al. Oncogenic human papillomavirus E6 proteins target the MAGI-2 and MAGI-3 proteins for degradation. Oncogene. 2002;21:5088–96.
- [83] Nguyen ML, Nguyen MM, Lee D, Griep AE, Lambert PF. The PDZ ligand domain of the human papillomavirus type 16 E6 protein is required for E6's induction of epithelial hyperplasia in vivo. Journal of Virology. 2003a;77:6957–64.
- [84] Nguyen MM, Nguyen ML, Caruana G, Bernstein A, Lambert PF, et al. Requirement of PDZ-containing proteins for cell cycle regulation and differentiation in the mouse lens epithelium. Molecular and Cellular Biology. 2003b;23:8970–81.
- [85] Doorbar J. The papillomavirus life cycle. Journal of Clinical Virology: The Official Publication of the Pan American Society for Clinical Virology. 2005;32 Suppl 1:S7–15.

- [86] Herber R, Liem A, Pitot H, Lambert PF. Squamous epithelial hyperplasia and carcinoma in mice transgenic for the human papillomavirus type 16 E7 oncogene. Journal of Virology. 1996;70:1873–81.
- [87] Song S, Pitot HC, Lambert PF. The human papillomavirus type 16 E6 gene alone is sufficient to induce carcinomas in transgenic animals. Journal of Virology. 1999;73:5887–93.
- [88] Riley RR, Duensing S, Brake T, Munger K, Lambert PF, Arbeit JM. Dissection of human papillomavirus E6 and E7 function in transgenic mouse models of cervical carcinogenesis. Cancer Research. 2003;63:4862–71.
- [89] Genther Williams SM, Disbrow GL, Schlegel R, Lee D, Threadgill DW, et al. Requirement of epidermal growth factor receptor for hyperplasia induced by E5, a high-risk human papillomavirus oncogene. Cancer Research. 2005;65:6534–42.
- [90] Stelzer MK, Pitot HC, Liem A, Schweizer J, Mahoney C, et al. A mouse model for human anal cancer. Cancer Prevention Research. 2010;3:1534–41.
- [91] Baandrup L, Thomsen LT, Olesen TB, Andersen KK, Norrild B, et al. The prevalence of human papillomavirus in colorectal adenomas and adenocarcinomas: a systematic review and meta-analysis. European Journal of Cancer. 2014;50:1446–61.
- [92] Damin DC, Ziegelmann PK, Damin AP. Human papillomavirus infection and colorectal cancer risk: a meta-analysis. Colorectal Disease: The Official Journal of the Association of Coloproctology of Great Britain and Ireland. 2013;15:e420–8.
- [93] De Vuyst H, Clifford GM, Nascimento MC, Madeleine MM, Franceschi S. Prevalence and type distribution of human papillomavirus in carcinoma and intraepithelial neoplasia of the vulva, vagina and anus: a meta-analysis. International Journal of Cancer. 2009;124:1626–36.
- [94] Lorenzon L, Ferri M, Pilozzi E, Torrisi MR, Ziparo V, French D. Human papillomavirus and colorectal cancer: evidences and pitfalls of published literature. International Journal of Colorectal Disease. 2011;26:135–42.
- [95] Machalek DA, Poynten M, Jin F, Fairley CK, Farnsworth A, et al. Anal human papillomavirus infection and associated neoplastic lesions in men who have sex with men: a systematic review and meta-analysis. The Lancet Oncology. 2012;13:487–500.
- [96] Al Moustafa AE, Foulkes WD, Wong A, Jallal H, Batist G, et al. Cyclin D1 is essential for neoplastic transformation induced by both E6/E7 and E6/E7/ErbB-2 cooperation in normal cells. Oncogene. 2004;23:5252–6.
- [97] Yasmeen A, Hosein AN, Yu Q, Al Moustafa AE. Critical role for D-type cyclins in cellular transformation induced by E6/E7 of human papillomavirus type 16 and E6/E7/ErbB-2 cooperation. Cancer Science. 2007a;98:973–7.

- [98] Al Moustafa AE, Kassab A, Darnel A, Yasmeen A. High-risk HPV/ErbB-2 interaction on E-cadherin/catenin regulation in human carcinogenesis. Current Pharmaceutical Design. 2008;14:2159–72.
- [99] Yasmeen A, Bismar TA, Dekhil H, Ricciardi R, Kassab A, et al. ErbB-2 receptor cooperates with E6/E7 oncoproteins of HPV type 16 in breast tumorigenesis. Cell Cycle. 2007b;6:2939–43.
- [100] Yasmeen A, Bismar TA, Kandouz M, Foulkes WD, Desprez PY, et al. E6/E7 of HPV type 16 promotes cell invasion and metastasis of human breast cancer cells. Cell Cycle. 2007c; 6:2038–42.
- [101] Fong S, Itahana Y, Sumida T, Singh J, Coppe JP, et al. Id-1 as a molecular target in therapy for breast cancer cell invasion and metastasis. Proceedings of the National Academy of Sciences of the United States of America. 2003;100:13543–8.
- [102] Sikder HA, Devlin MK, Dunlap S, Ryu B, Alani RM. Id proteins in cell growth and tumorigenesis. Cancer Cell. 2003;3:525–30.
- [103] Ricciardi R, Ghabreau L, Yasmeen A, Darnel AD, Akil N, et al. Role of E6/E7 oncoproteins of high-risk human papillomaviruses in human colorectal carcinogenesis. Cell Cycle. 2009;8:1964–5.
- [104] Al Moustafa AE, Yasmeen A, Ghabreau L, Akil N. Does the Syrian population have to wait for the new generation of human papillomaviruses vaccine? Human Vaccines & Immunotherapeutics. 2012;8:1867–8.
- [105] Dochez C, Bogers JJ, Verhelst R, Rees H. HPV vaccines to prevent cervical cancer and genital warts: an update. Vaccine. 2014;32:1595–601.
- [106] Handler MZ, Handler NS, Majewski S, Schwartz RA. Human papillomavirus vaccine trials and tribulations: Clinical perspectives. Journal of the American Academy of Dermatology. 2015;73:743–56; quiz 57–8.
- [107] Papadopoulos AJ, Schwartz RA, Lefkowitz A, Tinkle LL, Janniger CK, et al. Extragenital bowenoid papulosis associated with atypical human papillomavirus genotypes. Journal of Cutaneous Medicine and Surgery. 2002;6:117–21.
- [108] Grimes RM, Benjamins LJ, Williams KL. Counseling about the HPV vaccine: desexualize, educate, and advocate. Journal of Pediatric and Adolescent Gynecology. 2013;26:243–8.
- [109] Brown DR, Kjaer SK, Sigurdsson K, Iversen OE, Hernandez-Avila M, et al. The impact of quadrivalent human papillomavirus (HPV; types 6, 11, 16, and 18) L1 virus-like particle vaccine on infection and disease due to oncogenic nonvaccine HPV types in generally HPV-naive women aged 16–26 years. The Journal of Infectious Diseases. 2009;199:926–35.

- [110] Goldstone SE, Jessen H, Palefsky JM, Giuliano AR, Moreira ED, Jr., et al. Quadrivalent HPV vaccine efficacy against disease related to vaccine and non-vaccine HPV types in males. Vaccine. 2013;31:3849–55.
- [111] Palefsky JM, Giuliano AR, Goldstone S, Moreira ED, Jr., Aranda C, et al. HPV vaccine against anal HPV infection and anal intraepithelial neoplasia. The New England Journal of Medicine. 2011;365:1576–85.
- [112] Paavonen J, Naud P, Salmeron J, Wheeler CM, Chow SN, et al. Efficacy of human papillomavirus (HPV)-16/18 AS04-adjuvanted vaccine against cervical infection and precancer caused by oncogenic HPV types (PATRICIA): final analysis of a double-blind, randomised study in young women. Lancet. 2009;374:301–14.
- [113] Merck. Patient Information about GARDASIL®92014 (2014) Available at: http://www.merck.com/product/usa/pi\_circulars/g/gardasil\_9/gardasil\_9\_ppi.pdf
- [114] Ault KA. Epidemiology and natural history of human papillomavirus infections in the female genital tract. Infectious Diseases in Obstetrics and Gynecology. 2006;2006 Suppl: 40470.
- [115] Burger RA, Monk BJ, Kurosaki T, Anton-Culver H, Vasilev SA, et al. Human papillomavirus type 18: association with poor prognosis in early stage cervical cancer. Journal of the National Cancer Institute. 1996;88:1361–8.
- [116] Hariri S, Unger ER, Schafer S, Niccolai LM, Park IU, et al. HPV type attribution in high-grade cervical lesions: assessing the potential benefits of vaccines in a population-based evaluation in the United States. Cancer Epidemiology, Biomarkers & Prevention: A Publication of the American Association for Cancer Research, Cosponsored by the American Society of Preventive Oncology. 2015;24:393–9.



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