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Occupational Exposure to Coal, Genotoxicity, and Cancer Risk

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Abstract

Coal is a heterogeneous mixture containing large quantities of organic and inorganic matter, including carbon, hydrogen, oxygen, sulfur, nitrogen, and organometallic forms. The presence of mineral matter in coal may result in a number of environmental and human health problems related to its mining, preparation, and combustion. During coal mining activities, large quantities of coal dust, ashes, polycyclic aromatic hydrocarbons (PAHs), and heavy metals are released into the environment, forming a complex mixture. This mixture becomes one of the most important occupational risks for the health and safety of workers due to its synergistic, additive, and enhancing effects. Once inside the organism, this cocktail-like mixture can interact with cellular mechanisms related to the production of reactive oxygen species (ROS) and can cause damage in important macromolecules such as DNA, lipids, and proteins. In this review, human populations exposed to coal and coal burning were analyzed. Data from different studies were evaluated in relation to the effect of complex mixture exposure on DNA damage and mechanisms, and the background factors, such as gender, age, or smoking habit. The high temperatures that occur in combustion processes affect the characteristics of the resulting particles. The coal fly ash is released by combustion and its composition varies depending on the coal type and the method of collection used such as electrostatic precipitators. Compounds such as PAHs once activated by the organisms have been shown to have mutagenic and carcinogenic activity due to its ability to form adducts with purines. Moreover, metals that commonly are evaporated during the cooling process increase its toxicity. The particles when inhaled can pass from the alveoli into the bloodstream and affect extrapulmonary organs. Several studies have described the inflammatory cascade that triggers exposure to coal and coal fly ash particles; they have a complex composition capable of generating a persistent inflammatory process, resulting in diseases widely described as emphysema, bronchitis, pneumoconiosis, as thma, and cancer. Several human biomonitoring studies have been conducted evaluating the inflammatory process and



© 2016 The Author(s). Licensee InTech. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. the release of cytokines, polymorphisms involved in detoxification mechanisms, different biomarkers associated with occupational exposure, DNA damage, and the influence of oxidative stress in disease development. The relationship between chronic exposure to coal and coal ash particles and cancer is still widely debated. This review gave us a broad assessment about the associated mechanisms between cancer and exposure to coal and different findings around the world.

Keywords: coal, biomonitoring, DNA damage, ROS, PAHs, diseases

1. Introduction

In the last decades, the human population genetics integrity has been compromised by the great industrial activity, which exposes people to a variety of chemicals and genotoxic agents. As a result, it is important to determine what is considered as an "acceptable" level of genetic damage in a concrete population, carry out assay genotoxicity as a routine and also monitor those who, by their occupation or lifestyle, are more exposed or with a bigger risk of having alterations on their genetics stability [1].

One method to quantify the exposure to those substances, as well as its possible impact on the organism, is the use of biological monitoring procedures, or biomonitoring, through biomarkers. Biomonitoring studies try to establish a connection between the environmental factors and the diseases. They detect first alterations in nonmalignant phases, so as to prevent health problems by recognizing the environmental cause of them.

The biological markers, or biomarkers, are the measurable changes (biochemical, physiological, or morphological) that associate to a toxic exposure or any early biochemical alteration, whose study on the biological fluids, tissues, or exhaled air that allow to assess the health risk exposure intensity. The identification of genotoxicity markers believed to cause genome damage is useful, since it can define a prepathogenesis state, such as cancer. It is of vital importance for different diseases prevention, which is the final goal of biomonitoring. In order to achieve it, there must be two stages: 1) detecting human exposure to environment carcinogenic agents; 2) determining genotoxic effects *in vivo* [2].

The combined use of genetic biomarkers and classic epidemiology tools (clinic history and questionnaires) has enabled the identification of early effects to the occupational exposure to distinct pollutant around the world [2–4]. Many biomarkers are used to assess genotoxic effects on human populations exposed to complex mixtures of chemicals. Although there are different possibilities, micronuclei (MN) frequency, chromosomal aberrations (CAs), and comet assay are the most commonly chosen biomarkers. MN originates from chromosome fragments or whole chromosomes that are not included in the main daughter nuclei during nuclear division [5, 6]. MN induction reflects clastogenic and aneugenic damage and is a predictive biomarker of cancer risk [7]. Comet assay detects DNA lesions in individual cells obtained under a variety of experimental conditions; the technique can also be used to evaluate DNA repair [8, 9].

The large inter-individual variability in the capacity to activate or inactivate potential genotoxic and carcinogenic compounds is probably influenced by polymorphisms of the genes encoding the metabolizing enzymes. Genes and proteins involved in metabolization/detoxification of xenobiotics, as well as those involved in DNA repair, are usually used as potential markers of susceptibility for the development of several diseases in which the etiology is related to exposure to environmental hazards. Polymorphisms in such genes have been linked with an increased risk of cancer in several case-control studies [10].

Biomonitoring studies in populations exposed to complex mixtures of chemicals considering individual susceptibility are quite complicated due to inadequate toxicity data, and the unpredictable nature of interaction effects that may be synergistic, additive, or enhancers.

2. Occupational exposure to coal

The coal reserves in a worldwide level is up to 847.5 billion of tons, enough amount to serve the current production for 119 years. This prediction is different from the ones related to oil and gas, which have available supplies for less time [11]. According to data from the International Energy Agency (IEA), coal is the most used resource for energy generation in the world, responsible for 41% of the total production. Nowadays, the main application of mineral coal is to generate energy through thermal power plants. These reserves are considered to have a 109-year lifespan and their coalfields are located in 75 countries. The main world coal producers are China, the United States, India, Australia, Indonesia, Russia, South Africa, Germany, Poland, and Kazakhstan, which are responsible for 91% of the world's production [12]. If those projections are right, the consequences of coal mining and combustion will have large effects in the environment. Thus, the exposed populations monitoring is fundamental with the aim of contributing to the state of knowledge about the health risk and motivate the establishment of control, hygiene, and prevention strategies.

It is well known that coal mining activities are one of the biggest resources of contamination due to the large quantity of substances liberated in the environment. The content of the coal dust and ashes produced by burning are not always homogeneous and this depends on the source and rank of the coal [13, 14]. Coal dust is constituted from carbon, hydrogen, oxygen, nitrogen, quartz (crystalline silica), and inorganic minerals, such as beryllium, cadmium, cobalt, chromium, iron, boron, copper, nickel, antimony, zinc, aluminum, titanium, magnesium, manganese, mercury, and lead [15]. As observed, coal is a mixture of a variety of chemicals, including hydrocarbons, which may raise polycyclic aromatic hydrocarbons (PAHs). All technological processes associated with open fire or temperatures between 400 and 600°C, that may lead to PAHs, should be considered potentially hazardous [16, 17].

In relation to coal mining residues exposure, studies in which biomarkers of effect, susceptibility, and exposure are used as epidemiological tools remain rare and a big part of them come from studies on underground coal mining [18, 19]. The effects generated by open coal mining are little explored, though. In open coal mining, the residues pass directly to the atmosphere, where complex mixtures are formed, and the coal exposure to environmental factors such as sunlight facilitates the processes of spontaneous combustion and, therefore, the release of PAHs [20].

Studies about the coal exposure and its harmful effects have been conducted around the world [21–23]. The main way for exposure of the coal mining workers to the potentially dangerous residues is through the inhaling of coal dust particles from mining and manipulation. It is a known fact that the coal mining continuous exposure can cause a variety of diseases, such as coal workers pneumoconiosis (CWP), silicosis, cancer, and chronic obstructive pulmonary disease (COPD), as emphysema and chronic bronchitis [24].

Many studies have established that some of those diseases could have been originated from the genotoxic damage generated by the inhalation of those mineral particles, able to interact with macrophages, epithelial cells, and other cells generating the production of large amount of reactive oxygen species (ROS) [24–26]. The continuous inhalation of coal dust and fly ashes particles is an important cell and non-cell source of ROS in the lung. This may be associated to the damage of target cells of that tissue and other cell lines, after spreading through the bloodstream [27].

Coal-induced DNA damage is related to macrophage activation and the recruitment of polymorphonuclear cells. This cell activation induces the release of inflammatory mediators, such as cytokines, ROS and reactive nitrogen species (RNS). The proinflammatory properties of ROS and RNS include endothelial cell damage, lipid peroxidation and oxidation, the release of chemostatic factors, the recruitment of neutrophils, and DNA damage [26, 28]. Interaction of ROS with DNA can result in DNA structural and transcriptional errors [29, 30]. Damage caused by ROS is recognized by DNA glycosylases, apurinic/apyrimidinic endonucleases of the base excision repair (BER) mechanism, and in some cases, by the nucleotide excision repair (NER) machinery, leading to DNA strand-breaks [31, 32].

Although chronic exposure may continue to damage the DNA, it has been suggested that inorganic elements can induce DNA single-strand breaks, possibly via the generation of ROS and that this type of damage is soon repaired. Metals are also known to modulate gene expression of enzymes [33]. In addition, PAHs can induce DNA lesions as single-strand breaks via DNA repair mechanisms, related with increased adduct formation and electrophilic metabolites [34–36]. Electrophilic metabolites covalently interact with the DNA [37, 38], and adducts are formed with purines, especially guanine, after metabolic activation by enzymatic complex P450 [39]. The International Agency for Research on Cancer (IARC) classified quartz, main constituent of coal, into IARC Group 1 (carcinogen), due to sufficient evidence for carcinogenicity in experimental animals and in humans [40, 41]. The other factor that could lead to different results in coal dust exposure, with positive and negative results, might be explained by the possible differences in composition, in which the proportion of the metals, PAHs, and silica (quartz) content may have an influence on the genotoxicity. Despite those findings, coal dust remains classified as non-carcinogen for human (Group 3) in IARC [40, 41]. The importance of coal as an energy source makes its characterization and estimation of risks of extreme importance to the safety of those individuals and the environment.

Several factors may explain conflicting results among different studies with human exposed to coal, e.g. cigarettes smoked, age, gender, nutritional status, and individual polymorphisms [6, 42]. Susceptibility is critical to an understanding of coal diseases, including cancer, and many xenobiotic agents act to alter susceptibility. Unknown individual susceptibility, inadequate toxicity data, and the unpredictable nature of interaction effects make the implementation of a human biomonitoring assessment for complex mixtures of chemicals extremely complicated.

3. Oxidative stress and genotoxic damage related with coal exposure

One important aspect to consider about the coal exposure is the amount of products generated during the coal combustion. The burning of coal, in order to generate electricity, produces flue gasses and particulate materials like coal fly ashes and residues as scoria and bottom ash. The finer particles (coal fly ash) are obtained by mechanical or electrostatic precipitation of the dust in suspension in the gases produced by combustion, while the coarser particles fall to the bottom by gravity and are removed at the bottom of the boiler [43, 44].

The combustion temperature is an important factor that determines the physical properties of the particles. In the combustion of conventional high temperature (>1400°C), the main aluminosilicate melts and condenses to form spherical particles. The coal fly ash particles produced are mostly irregularly shaped and contain a complex mixture consisting of unburned carbon; oxides; quartz; elements such as aluminum, silicon, calcium, iron, nickel, arsenic, chromium, copper, lead, cadmium, zinc [45, 46], and PAHs [47].

The coal fly ash has a relatively low toxicity as compared with coal or quartz [45]. Studies have determined the role of coal fly ash particle size and the release of iron, which leads to generation of radicals and oxidative stress. In this context, it was demonstrated the ability of coal fly ash release of bioavailable iron, which triggers processes and redox oxidant production [48]. In addition, it was shown that interleukin 8 (IL-8) levels in human lung epithelial cells are increased in response to coal fly ash and vary with the bioavailability of iron, as a function of source of coal and particle size [49]. The smaller size fraction had more stimulatory activity, which may be related to the fact that iron is more concentrated in this fraction. Particle size is a critical factor because a larger surface area allows more significant transport of metal and other adsorbed components, increasing the pulmonary toxicity of particulate matter (PM) [50].

The particles are classified according to their aerodynamic diameter (in micrometer) in coarse (PM 10), fine (PM 2.5), ultrafine (PM 0.1) [51]. The smaller particles are more harmful with respect to health effects because of their very high alveolar deposition fraction, large surface area, chemical composition, ability to induce inflammation, and potential to translocate to the circulation to extrapulmonary organs [52–54]. These particles could trigger persistent lung inflammation compared to the coarse particles in addition to the exposure to genotoxic compounds, which are contained in the particles [26, 55].

Depending on the toxicity, the chemical properties, and the concentration in air, coal and coal fly ash particles can constitute a risk to exposed workers. When these particles are inhaled and

deposited in the lungs, they can lead to health risks due to the leaching of genotoxic compounds and altered immunological mechanisms affecting the lung parenchyma causing diseases [56]. These nanometric particles are very small, which allows them to penetrate the biological organs and affect its normal function. More specifically, as the particle load in the lung increases the alveolar macrophages and epithelial cells are activated, leading to the release of inflammatory mediators, ROS, enzymes (elastases, proteases, collagenases), cytokines [tumor necrosis factor alpha (TNF- α), interleukins], and growth factors (TGF- β) that control and stimulates the fibrosis, genotoxic events, and cell death [45, 57, 58].

Persistent inflammatory processes have been accepted as a crucial factor in the pathogenesis. In Zhai et al. [59], was investigated whether systemic TNF- α , soluble TNF- α receptors (p55, p75), IL-6, and soluble IL-6 receptor could be markers of biological activities of Chinese CWP. Interestingly, those results suggest that serum levels of TNF receptors and IL-6 are associated with the fibrotic process of CWP and serum cytokine levels may be correlated with the severity of CWP. In the pathogenesis of these respiratory diseases related with coal exposure, oxidative damage plays a key role. Either acting in association or independently, the chemical and physical characteristics can lead to the generation of ROS and oxidative stress [60, 61].

These particles are chemically heterogeneous and can be a source of oxidants by themselves ("acellular" mechanisms), due to their composition, such as oxides, metals, and PAHs [26]. Soluble metals (transition) associated to the particle can increase the generation of ROS by Haber-Weiss reactions. PAHs may be metabolically activated and induce ROS and oxidative stress, also forming bulky adducts or strand breaks on DNA [50, 62, 63].

Another way of generating oxidants is via cellular. Once in the lungs, alveolar macrophages are activated and generate large amounts of ROS, and chemoattractant factors of other inflammatory cells such as monocytes and neutrophils are released, which amplify this response generating more oxidants [64]. The particle size is a critical factor, because very large particles are difficult to phagocytose, leading to the process of incomplete or "frustrated" phagocytosis aggravating the response [65, 66].

Considering three different scenarios with respect to exposure to particles, the generation of oxidative stress, inflammation, and oxidative DNA damage, several authors questioned whether the lung inflammation may be related to secondary genotoxic effects. They also questioned if phenomena of oxidative stress, inflammation and DNA damage are independent or interrelated, whether oxidative stress stimulates inflammatory processes, or inflammation mediated by particles cause oxidative stress, or even if it is possible that particles may cause both phenomena of oxidative stress and inflammation but for different mechanisms of action [26, 61].

In normal physiological conditions, there is a balance between ROS generation and antioxidant defenses. However, the continuous inhalation of particles may interfere in this equilibrium leading to oxidative stress process in the lung. Consequently, a high loading of particles alters the oxidant-antioxidant balance, leading to oxidative damage and the beginning of pathological processes [67]. The most important effects of ROS in the lung include damage to cell membranes by lipid peroxidation process, protein oxidation, and DNA damage in target cells [27].

As seen in **Figure 1**, oxidative DNA damage can have many consequences, from cell death and tissue destruction to cell proliferation. Furthermore, ROS can also act as regulators in signaling pathways intracellularly and transcription factors of a variety of genes including those of proinflammatory cytokines, adhesion molecules, and proto-oncogenes [68].

In vitro effects induced by coal exposure have been described in different cells such as murine alveolar type II epithelial cells (C10) [69] and in 7TD1 cells [70]. ROS generation and oxidative damage by coal fly ash particles have been described in different cell lines, in human peripheral blood mononuclear cells [71], in rat alveolar macrophages (NR8383) [72], in BEAS-2B human lung epithelial cells [73], and in rat lung epithelial (RLE) cells [74].

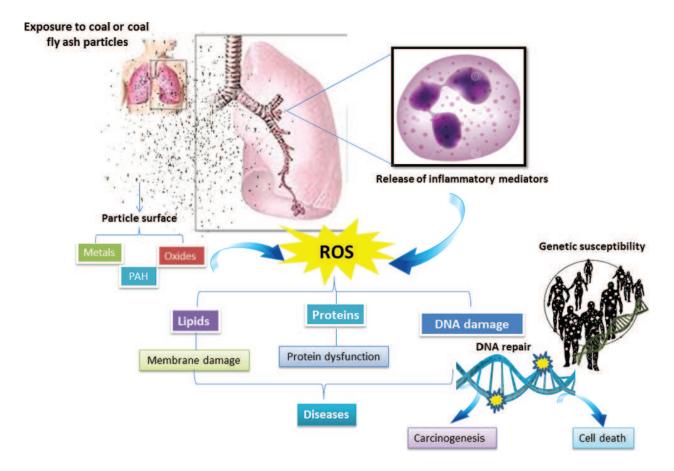


Figure 1. Main pathways associated with the generation of oxidative damage and the development of diseases induced by coal and coal fly ash particles.

ROS induce point mutations and CAs in cells. Many inhaled toxic substances contained in the particles contribute to oxidative modification that has as target of attack specific components of the cytoplasm and the nucleus. Such changes include DNA breakage, DNA oxidative modification, base modifications, alterations in the DNA sequence, poly-ADP ribosylation, activation of kinases, activation of proto-oncogenes, and inactivation of tumor suppressor

genes. Persistent generation of ROS generated by mineral particles indestructible or engulfed incompletely leads to damage to organelles keys [59, 61, 75]. The oxidation of C8 deoxyguanosine (dG), resulting in 7-dihydro-8-oxo-2'-deoxyguanosine (8-oxodG), is the most common oxidative lesion generated by ROS. The proportion of 8-oxodG/dG has been considered as a biomarker of oxidative stress and has been studied in relation to exposure to mineral particles *in vitro* and *in vivo* [76].

Human biomonitoring studies about the effects of exposure to coal and residues using different biomarkers have been conducted around the world. In this context, our group has obtained interesting findings in workers exposed to coal mining in Colombia and Brazil. In Rohr et al. [77], was found that Brazilian workers with occupational exposure to coal had significantly increased genetic damage in peripheral blood lymphocytes compared with unexposed individuals. Exposed workers presented lower average levels of thiobarbituric acid reactive substances (TBARS) and catalase activity (CAT). In addition, DNA damage evaluated by human buccal micronucleus cytome (BMCyt) assay was observed in mine workers, which could be a consequence of oxidative damage resulting from exposure to coal residue mixtures [78].

In Colombia, DNA damage in lymphocytes of coal open-cast mining workers using the cytokinesis-blocked micronucleus test and the comet assay were observed [79]. Also, in buccal mucosa samples, the micronucleus frequencies and nuclear buds were significantly higher in the exposed group than in non-exposed control group. Interestingly, blood samples of Colombian mining workers analyzed showed higher values of silicon and aluminum characteristic elements of coal particles, compared with the control group [80]. All these studies converge to a point: the compounds contained in the particles may be related to ROS generation, DNA damage, and formation of pro-mutagenic adducts.

These are important findings if we consider that oxidative DNA damage can lead to long-term risk of cancer and other diseases caused by air pollution by these particles. In **Table 1**, can be observed an overview of key studies on the genotoxicity in human population exposed to coal and coal combustion products. These studies demonstrated DNA damage using different methods, related with inorganic elements and oxidative stress.

| References C | Country | Exposure(s) | Biomarker | Outcome(s) |
|--------------|----------|-------------|-----------------------------|---|
| [81] S | Slovenia | | Sister-chromatid exchanges | Significantly higher levels of |
| | | | (SCE), unstable chromosome | chromosomal aberrations, SCE and |
| | | | and chromatid aberrations | micronuclei in exposed group |
| | | | and micronuclei in | compared with the control group. |
| | | | blood lymphocytes | |
| [82] B | Brazil | Underground | Oxidative stress | The results showed that subjects |
| | | workers | biomarkers (TBARS, GSH/ | directly and indirectly exposed to coal |
| | | directly | GSSG, α -tocopherol, | dust face an oxidative stress |
| | | exposed, | GST, GR, GPx, SOD, | condition. They also indicate |
| | | surface | CAT). | that people living in the |

| References | Country | Exposure(s) | Biomarker | Outcome(s) |
|------------|----------|--|--|--|
| | | workers indirectly exposed, residents living near the mines. | | vicinity of the mine plant are in health risk regarding coal mining-related diseases. |
| [22] | Turkey | Coal combustion products | Chromosomal aberrations (CAs), polyploidy, sister- chromatid exchanges (SCEs), and micronuclei (MN) in blood cells. | Significantly higher levels of CA, polyploidy, SCE, and MN in peripheral blood lymphocytes of workers compared with controls. |
| [83] | Turkey | Underground coal mining | SCE, CA, and micronuclei frequencies in peripheral lymphocytes. | Increase in sister chromatid exchanges, chromosomal aberrations, and micronucleus frequencies found in underground coal miners as compared to control group. |
| [21] | | coal emissions | GSTM1 and GSTT1 genotypes. Expression of p53 protein in sputum samples. | The GSTM1 null genotype may enhance susceptibility to lung cancer due to these indoor coal combustion emissions. Smoky coal use was strongly associated with overexpression of p53 in tumor cells among highly exposed women. |
| [79] | Colombia | Open cast mining | (MN) frequency and DNA damage (comet assay) in lymphocytes. | The biomarkers evaluated showed statistically significant higher values in the exposed group compared to the non-exposed control group. |
| [80] | | Open cast mining | Micronucleus (MN) frequencies, nuclear buds, karyorrhectic and karyolytic cells in buccal mucosa samples and content of inorganic elements in blood samples by PIXE. | MN frequencies and nuclear buds in buccal mucosa samples were significantly higher in the exposed group than in the non-exposed control group. In addition, karyorrhectic and karyolytic cells were also significantly higher in the exposed group (cell death). Blood samples showed higher values of silicon (Si) and aluminum (Al) in the exposed group. |
| [84] | Russian | Underground coal | Chromosomal and chromatid type | A higher frequency of chromosomal aberrations in the exposed group |

| References | 6 Country | Exposure(s) | Biomarker | Outcome(s) |
|------------|-----------|-------------------------------|--|--|
| | | mining | aberrations in blood lymphocytes | compared with the control group. |
| [77] | Brazil | Open coal mining | MN and nucleoplasmic bridge frequencies in peripheral lymphocytes, damage index and damage frequency (comet assay). | Increased MN and nucleoplasmic bridge frequencies in peripheral lymphocytes, increased damage index and damage frequency (comet assay). Lower average levels of TBARS and catalase activity (CAT), while the mean superoxide dismutase activity (SOD) levels were higher in the exposed group. |
| [78] | Brazil | Open coal mining | Buccal micronucleus cytome (BMCyt) DNA damage, cell death, and basal cell frequency in buccal cells. | The exposed group presented a significantly higher frequency of basal cells, micronuclei in basal and differentiated cells, and binucleated cells compared to the non-exposed group. No correlation between DNA damage and metal concentration in the blood of mine workers. |
| [19] | Peru | Underground coal mining | Chromosomal aberrations in peripheral lymphocytes | Miners occupationally exposed to underground mining activity have an increased frequency of chromosomal aberrations compared with the controls. |
| [85] | - | Coal fly ash particles | SCE frequencies in peripheral blood lymphocytes. | No increased SCE frequencies were found in PBLs of workers potentially exposed to coal fly ash when compared to the control group. No differences were observed between the exposed and control groups for frequencies of gene mutations at the HPRT locus in PBLs, for micronucleus frequencies using the cytokinesis block method, or for urinary mutagen excretion measured with <i>Salmonella</i> <i>typhimurium</i> tester strains TA98 and TA97. |
| [86] | Germany | Underground coal mining | Structural chromosomal aberrations in peripheral lymphocytes | Coal miners had significantly higher frequencies of chromosomal aberrations compared with controls. |
| [87] | Turkey | Underground coal | Sister chromatid exchange (SCE) and | SCE and MN frequencies in CWP patients were found significantly higher than |

| References Country | Exposure(s) | Biomarker | Outcome(s) |
|--------------------|-------------|-----------------------|---------------------------------------|
| | mining | micronucleus (MN) | in coal workers and unexposed groups. |
| | | frequency in | |
| | | lymphocytes of | |
| | | Turkish CWP patients. | |

Table 1. Overview of key studies on the genotoxicity in human population exposed to coal and coal combustion products.

4. Conclusions

The coal mining activities generate different types of compounds that are released into the environment. Once into the atmosphere, these compounds form a complex mixture that consists of metals, oxides, and PAHs. These compounds can interact with "acellular" and cellular mechanisms related with ROS production. The metals found in the coal fly ash and coal particles by different ways lead to the ROS formation. Important macromolecules as DNA, proteins, and lipids can suffer oxidative modifications. The PAHs contained in the particles also influence the particles toxicity. A second indirect way for excessive ROS formation is related to cellular mechanisms, which is consequence of oxidative burst of macrophages and neutrophils during phagocytosis of particles and inflammation produced.

If we think in exposed populations, we cannot ignore the social and environmental impact associated with coal mining. The continuous inhalation, the high load of particles in phagocytic cells, the oxidant-antioxidant imbalance which are linked to the origin of pathological processes; this whole scenario is worrisome to biologic level for these populations. In addition, in recent years, coal mining had a remarkable increase in demand; international mining companies have increased their investments in exploration around the world. For this reason, human biomonitoring studies in exposed populations become really necessary to contribute to knowledge state about the risk for those people in order to motivate the design of control, hygiene, and prevention strategies, besides epidemiological surveillance.

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References

- [1] Zalacain, M; Sierrasesumaga, L; Patino, A. (2005). The cytogenetic assay as a measure of genetic instability induced by genotoxic agents. An Sist Sanit Navar. 28(2);227–36.
- [2] Pastor, S; Lucero, L; Gutierrez, S; Durban, R; Gomez, C; Parron, T; et al. (2002). A followup study on micronucleus frequency in Spanish agricultural workers exposed to pesticides. Mutagenesis. 17(1);79–82.
- [3] Martino-Roth, M.G; Viegas, J; Roth, D.M. (2003). Occupational genotoxicity risk evaluation through the comet assay and the micronucleus test. Genet Mol Res. 2(4); 410–7.
- [4] Cassini, C; Calloni, C; Bortolini, G; Garcia, SC; Dornelles, MA; Henriques, JÁ; Erdtmann B; Salvador M. (2011). Occupational risk assessment of oxidative stress and genotoxicity in workers exposed to paints during a working week. Int J Occup Med Environ Health. 24(3);308–19.
- [5] Fenech, M; Morley, A. (1985). Solutions to the kinetic problem in the micronucleus assay. Cytobios. 43;233–46.
- [6] Mateuca, R; Lombaert, N; Aka, P.V; Decordier, I; Kirsch-Volders, M. (2006). Chromosomal changes: induction, detection methods and applicability in human biomonitoring. Biochimie. 88(11);1515–31.
- [7] Bonassi, S; Znaor, A; Ceppi, M; Lando, C; Chang, W.P; Holland, N; et al. (2007). An increased micronucleus frequency in peripheral blood lymphocytes predicts the risk of cancer in humans. Carcinogenesis. 28(3);625–31.

- [8] Tice, R.R; Agurell, E; Anderson, D; Burlinson, B; Hartmann, A; Kobayashi, H; et al. (2000). Single cell gel/comet assay: guidelines for in vitro and in vivo genetic toxicology testing. Environ Mol Mutagen. 35(3);206–21.
- [9] Collins, A.R. (2014). Measuring oxidative damage to DNA and its repair with the comet assay. Biochim Biophys Acta 1840;794–800. doi: 10.1016/j.bbagen.2013.04.022.
- [10] International Agency for Research on Cancer (IARC), Monographs on the Evaluation of Carcinogenic Risk to Humans, Reevaluation of Some Organic Chemicals, Hydrazine and Hydrogen Peroxide Agency for Research on Cancer, Toluene, World Health Organization. (1999). Lyon 71;829–864. Online: http://monographs.iarc.fr/ENG/ Monographs/vol71/mono71.pdf
- [11] Cerrejón-responsible mining. Uses of coal. (2013). Online, in Spanish: http:// www.cerrejon.com/site/mas-sobre-el-carbon/usos-del-carbon.aspx
- [12] International Energy Agency (IEA). (2016). Online: http://www.iea.org/
- [13] Aneel- National Electric Energy Agency. Atlas Electric Power in Brazil. Chapter 9 Mineral Coal. (2009). Online, in Portuguese: www.aneel.gov.br/arquivos/pdf/ atlas_par3_cap9.pdf
- [14] Caballero, A.L; Médico, O.A. (2013). Characterization and possible use of coal ash resulting from coal combustion in thermocentral of bed fluidized. Río Turbio (Argentina). National University of Austral Patagonia. Online, in Spanish: http:// www.redisa.uji.es/artSim2013/CaracterizacionDeResiduosSolidos/Caracterizacion %20Cenizas%20Combustion%20Carbon.pdf
- [15] Kalkreuth, W; Holz, M; Kern, M; Machado, G; Mexias, A; Silva, M.B; Willett, J; Finkelman, R; Burger, H. (2006). Petrology and chemistry of permian coals from the Paraná basin: Santa Terezinha, Leão-Butiá and Candiota Coal fields, Rio Grande do Sul, Brazil. Intern. J. Coal Geol. 68;79–116.
- [16] Kvitko, K; Bandinelli, E; Henriques, J; Heuser, V; Rohr, P; Da Silva, F; Schneider, N; Fernandes, S; Ancines, C; Da Silva, J. (2012). Susceptibility to DNA damage in workers occupationally exposed to pesticides, to tannery chemicals and to coal dust during mining. Genet. Mol. Biol. 35;1060–68.
- [17] Jarvis, I.W; Dreij, K; Mattsson, Å; Jernström, B; Stenius, U. (2014). Interactions between polycyclic aromatic hydrocarbons in complex mixtures and implications for cancer risk assessment. Toxicology. 3(321);27–39.
- [18] Agostini, J; Otto P; Wajntal A. (1996). Chromosome damage in underground coal miners: detection by conventional cytogenetic techniques and by submitting lymphocytes of unexposed individuals to plasma from at-risk groups. Braz J Genet. 19;641–6.
- [19] Santa Maria, S.R; Arana, M; ,Ramirez O. (2007). Chromosomal aberrations in peripheral lymphocytes from male native miners working in the Peruvian Andes. Genet Mol Biol. 30;1135–8.

- [20] León, G; Perez, L.E; Linares, J.C; Hartmann, A; ,Quintana M. (2007). Genotoxic effects in wild rodents (*Rattus rattus* and *Mus musculus*) in an open coal mining area. Mutat Res. 630;42–9.
- [21] Lan, Q; ,He X. (2004). Molecular epidemiological studies on the relationship between indoor coal burning and lung cancer in Xuan Wei, China. Toxicology. 198;301–5.
- [22] Celik, M; Donbak, L; Unal, F; Yüzbasioglu, D; Aksoy, H; Yilmaz, S. (2007). Cytogenetic damage in workers from a coal-fired power plant. Mutat Res. 627(2);158–63.
- [23] Karami, S; Boffetta, P; Brennan, P; Stewart, P.A; Zaridze, D; Matveev, V; Janout, V; Kollarova, H; Bencko, V; Navratilova, M; Szeszenia-Dabrowska, N; Mates, D; Gromiec, J.P; Sobotka, R; Chow, W.H; Rothman, N; Moore, L.E. (2011). Renal cancer risk and occupational exposure to polycyclic aromatic hydrocarbons and plastics. J Occup Environ Med. 53(2);218–223; 1076–2752.
- [24] Petsonk, E.L; Rose, C; ,Cohen R. (2013). Coal mine dust lung disease. New lessons from old exposure. Am J Respir Crit Care Med. 187(11);1178–85.
- [25] Vallyathan, V; Shi, X; Castranova, V. (1998). Reactive oxygen species: their relation to pneumoconiosis and carcinogenesis. Environ Health Perspect. 106(Suppl. 5);1151–5.
- [26] Knaapen, A.M; Borm, P.J; Albrecht, C; ,Schins R.P. (2004). Inhaled particles and lung cancer. Part A: Mechanisms. Int J Cancer 109;799–809.
- [27] Schins, R.P; ,Borm P.J. (1999). Mechanisms and mediators in coal dust induced toxicity: a review. Ann Occup Hyg. 43(1);7–33.
- [28] Ba, X; Aguilera-Aguirre, L; Rashid, QT; Bacsi, A; Radak, Z; Sur, S; Hosoki, K; Hegde, ML; Boldogh I. (2014). The role of 8-oxoguanine DNA glycosylase-1 in inflammation. Int J Mol Sci. 15(9);16975–97.
- [29] Schottenfield, D; Bebe-Dimmer, J. (2006). Chronic inflammation: a common and important factor in the pathogenesis of neoplasia. CA: Cancer J. Clin. 56;69–83.
- [30] Svilar, D; Goellner, EM; Almeida, KH; Sobol, RW. (2011). Base excision repair and lesion-dependent subpathways for repair of oxidative DNA damage. Antioxid Redox Signal. 14(12);2491–507.
- [31] Sander, M; ,Wilson S. (2005). Base Excision Repair, AP Endonucleases and DNA Glycosylases, John Wiley & Sons Ltd, Chichester.
- [32] Storr, S.J; Woolston, C.M; Zhang, Y; ,Martin S.G. (2013). Redox environment, free radical, and oxidative DNA damage. Antioxid Redox Signal. 18(18);2399–408.
- [33] Jomova, K; ,Valko M. (2011). Advances in metal-induced oxidative stress and human disease. Toxicology. 283;65–87.
- [34] Brescia, G; Celotti, L; Clonfero, E; Neumann, G.H; Forni, A; Foa, V; Pisoni, M; Ferri, G.M; Assennato, G. (1999). The influence of cytochrome P450 1A1 and glutathioneS-

transferase M1 genotypes on biomarker levels in coke-oven workers. Arch. Toxicol. 73;431–9.

- [35] Rojas, M; Cascorbi, I; Alexandrov, K; Kriek, E; Auburtin, G; Mayer, L; Kopp-Schneider, A; Roots, I; Bartsch, H. (2000). Modulation of benzo[a]pyrene diolepoxide-DNA adduct levels in human white blood cells by CYP1A1, GSTM1 and GSTT1 polymorphism.
 Carcinogenesis 21;35–41.
- [36] Pavanello, S; Pulliero, A; Lai, A; Gaiardo, A; Mastrangelo, G; Clonfero, E. (2005). Anti-B[a]PDE-DNA formation in lymphomonocytes of humans environmentally exposed to polycyclic aromatic hydrocarbons, G. Ital. Med. Lav. Ergon. 27;312–4.
- [37] Pereira-Neto, A.D; Moreira, J.C; Dias A.E.X.O; Arbilla G; Ferreira L.F.V; Oliveira A.S; Barek J. (2000). Assessment of human contamination by polycyclic aromatic hydrocarbons (PAHs) and nitrated derivatives (NPAHs): a methodological review. Quim. Nova 23;9. In Portuguese.
- [38] Singh, R; Sram, R.J; ,Binkova B; Kalina, I; Popov, T.A; ,Georgieva T; ,Garte S; Taioli, E; ,Farmer P.B. (2007). The relationship between biomarkers of oxidative DNA damage, polycyclic aromatic hydrocarbon DNA adducts, antioxidant status and genetic susceptibility following exposure to environmental air pollution in humans. Mutat. Res. 620;83–92.
- [39] Baird, W.M; Hooven, L.A; ,Mahadevan B. (2005). Carcinogenic polycyclic aromatic hydrocarbon-DNA adducts and mechanism of action. Environ. Mol. Mutagen. 45;106–14.
- [40] International Agency for Research on Cancer (IARC). (2012). Arsenic, metals, fibres, and dusts: a review of human carcinogens. Lyon. 100C;526. Online: http://monographs.iarc.fr/ENG/Monographs/vol100C/mono100C.pdf
- [41] International Agency for Research on Cancer (IARC). (1997). Silica, some silicates, coal dust and para-aramid fibrils, IARC Monographs on the Evaluation of the Carcinogenic Risks of Chemicals to Humans. Lyon. 68;528. Online: http://monographs.iarc.fr/ENG/ Monographs/vol68/mono68.pdf
- [42] Bonassi, S; Coskun, E; Ceppi, M; Lando, C; Bolognesi, C; Burgaz, S; et al. (2011). The HUman MicroNucleus project on eXfoLiated buccal cells (HUMN(XL)): the role of lifestyle, host factors, occupational exposures, health status, and assay protocol. Mutat Res. 728(3);88–97.
- [43] Silva, L; Ward, C.R; ,Hower J; ,Izquierdo M; ,Waanders F; ,Oliveira M; ,Li Z; Hatch, R; ,Querol X. (2010). Mineralogy and leaching characteristics of coal ash from a major Brazilian power plant. Coal Combust Gasificat Prod. 2;51–65.
- [44] Martinez, J. (2012). Coal fly ash and bottom ash or scoria. Technical file, Spain. On line, in Spanish: http://www.cedexmateriales.es/catalogo-de-residuos/24/cenizas-volantesde-carbon-y-cenizas-de-hogar-o-escorias/

- [45] Borm, P.J. (1997). Toxicity and occupational health hazards of coal fly ash (CFA). A review of data and comparison to coal mine dust. Ann Occup Hyg. 41;659–76.
- [46] Swaine, D.J; ,Goodarzi F. (1997). Environmental Aspects of Trace Elements in Coal. Kluwer, Dordrecht.
- [47] Ruwei, W; Jiamei, Z; Jingjing, L; Liu, G. (2013). Levels and patterns of polycyclic aromatic hydrocarbons in coal-fired power plant bottom ash and fly ash from Huainan, China. Arch Environ Contam Toxicol. 65(2);193–202.
- [48] Ball, B.R; Smith, K.R; Veranth, J.M; ,Aust A.E. (2000). Bioavailability of iron from coal fly ash: mechanisms of mobilization and of biological effects. Inhal Toxicol. 12(Suppl. 4);209–25.
- [49] Smith, K.R; Veranth, J.M; Hu, A.A; Lighty, J.S; ,Aust A.E. (2000). Interleukin-8 levels in human lung epithelial cells are increased in response to coal fly ash and vary with the bioavailability of iron, as a function of particle size and source of coal. Chem Res Toxicol. 13(2);118–25.
- [50] Mazzoli-Rocha, F; Fernandes, S; Einicker-Lamas, M; Zin, W.A. (2010). Roles of oxidative stress in signaling and inflammation induced by particulate matter. Cell Biol Toxicol. 26(5);481–98.
- [51] Donaldson, K; ,Stone V. (2003). Current hypotheses on the mechanisms of toxicity of ultrafine particles. Ann Ist Super Sanità; 39;405–10.
- [52] Oberdörster, G; Sharp, Z; Atudorei, V; Elder, A; Gelein, R; Kreyling, W; Cox, C. (2004). Translocation of inhaled ultrafine particles to the brain. Inhal Toxicol. 16(6–7);437–45.
- [53] Schins, R.P; Lightbody, J.H; Borm, P.J; ,Shi T; Donaldson, K; ,Stone V. (2004). Inflammatory effects of coarse and fine particulate matter in relation to chemical and biological constituents. Toxicol. Appl. Pharmacol. 195;1–11.
- [54] Mani, U; Prasad, A.K; ,Suresh Kumar V; Lal, K; Kanojia, R.K; Chaudhari, B.P; Murthy R.C. (2007). Effect of fly ash inhalation on biochemical and histomorphological changes in rat liver. Ecotoxicol Environ Saf. 68(1);126–33.
- [55] Donaldson, K; Tran, L; Jimenez, L.A; Duffin, R; Newby, D.E; Mills, N. (2005). Combustion-derived nanoparticles: a review of their toxicology following inhalation exposure. Part Fibre Toxicol. 2;10.
- [56] Carbone, F; Pagliara, R; Barone, A.C; Beretta, F; D'Anna, A. (2009). Characterization of nano-ashes generated during pulverized coal combustion. Report Ricerca Sistema Elettrico. Online: http://www.enea.it/it/Ricerca_sviluppo/documenti/ricerca-disistema-elettrico/centrali-carbone-rendimenti/rse109.pdf.
- [57] Gilmour, M.I; O'Connor, S; Dick, C; Miller, C.A; ,Linak W.P. (2004). Differential pulmonary inflammation and in vitro cytotoxicity of size-fractionated fly ash particles from pulverized coal combustion. J Air Waste Manag Assoc. 54;286–95.

- [58] Sambandam, B; Devasena, T; Islam, V.I; Prakhya, B.M. (2015). Characterization of coal fly ash nanoparticles and their induced in vitro cellular toxicity and oxidative DNA damage in different cell lines. Indian J Exp Biol. 53(9);585–93.
- [59] Zhai, R; Liu, G; Ge, X; Bao, W; Wu, C; Yang, C; Liang, D. (2002). Serum levels of tumor necrosis factor-alpha (TNF-alpha), interleukin 6 (IL-6), and their soluble receptors in coal workers' pneumoconiosis. Respir Med. 96(10);829–34.
- [60] Bonner, J.C. (2007). Lung fibrotic responses to particle exposure. Toxicol Pathol. 35(1); 148–53.
- [61] Møller, P; Danielsen, P.H; Karottki, D.G; ,Jantzen K; ,Roursgaard M; Klingberg, H; Jensen, D.M; Christophersen, D.V; Hemmingsen, J.G; Cao, Y; ,Loft S. (2014). Oxidative stress and inflammation generated DNA damage by exposure to air pollution particles. Mutat Res Rev Mutat Res. 762;133–66.
- [62] Risom, L; Møller, P; Loft, S. (2005). Oxidative stress-induced DNA damage by particulate air pollution. Mutat. Res. 592;119–37.
- [63] Valavanidis, A; Vlachogianni, T; Fiotakis, K; Loridas, S. (2013). Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. Int J Environ Res Public Health. 10;3886–907.
- [64] Fubini, B; ,Hubbard A. (2003). Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. Free Radic Biol Med. 34;1507–16.
- [65] Champion, J.A; ,Mitragotri S. (2006). Role of target geometry in phagocytosis. Proc Natl Acad Sci U S A. 103(13);4930–4.
- [66] Schinwald, A; ,Donaldson K. (2012). Use of back-scatter electron signals to visualise cell/nanowires interactions in vitro and in vivo; frustrated phagocytosis of long fibres in macrophages and compartmentalisation in mesothelial cells in vivo. Particl Fibre Toxicol. 9;34.
- [67] Porter, D.W; Leonard, S.S; Castranova V. (2006). Particles and Cellular Oxidative and Nitrosative Stress. Chapter 6. National Institute for Occupational Safety and Health. pp. 119–138
- [68] Porter, D.W; Leonard, S.S; Castranova V. (2006). Particles and Cellular Oxidative and Nitrosative Stress. Chapter 6. National Institute for Occupational Safety and Health. Particle toxicology. Chapter 6. pp 119–138.
- [69] Albrecht, C; Borm, P.J.A; Adolf, B; Timblin, C.R; ,Mossman B.T. (2002). In vitro and in vivo activation of extracellular signal-regulated kinases by coal dusts and quartz silica. Toxicol Appl Pharmacol. 184;37–45.

- [70] Gosset, P; Lassalle, P; Vanhée, D; Wallaert, B; Aerts, C; Voisin, C; Tonnel, A.B. (1991). Production of tumor necrosis factor-alpha and interleukin-6 by human alveolar macrophages exposed in vitro to coal mine dust. Am J Respir Cell Mol Biol. 5;431–6.
- [71] Dwivedi, S; Saquib, Q; Al-Khedhairy, A.A; Ali, A.Y; Musarrat, J. (2012). Characterization of coal fly ash nanoparticles and induced oxidative DNA damage in human peripheral blood mononuclear cells. Sci Total Environ. 15;331–8.
- [72] Diabaté, S; Mülhopt, S; Paur, H.R; Wottrich, R; Krug, H.F. (2002). In vitro effects of incinerator fly ash on pulmonary macrophages and epithelial cells. Int J Hyg Environ Health. 204;323–6.
- [73] Diabaté, S; Bergfeldt, B; Plaumann, D; Ubel, C; Weiss, C. (2011). Anti-oxidative and inflammatory responses induced by fly ash particles and carbon black in lung epithelial cells. Anal Bioanal Chem. 401(10);3197–212.
- [74] van Maanen, J.M; Borm, P.J; ,Knaapen A; van Herwijnen, M; Schilderman, P.A; Smith, K.R; Aust, A.E; Tomatis, M; Fubini, B. (1999). In vitro effects of coal fly ashes: hydroxyl radical generation, iron release, and DNA damage and toxicity in rat lung epithelial cells. Inhal Toxicol. 11;1123–41.
- [75] Liou, G.Y; ,Storz P. (2010). Reactive oxygen species in cancer. Free Radic Res. 44(5);479– 96.
- [76] Haghdoost S, Czene S, Näslund I, Skog S, Harms-Ringdahl M. (2005). Extracellular 8oxo-dG as a sensitive parameter for oxidative stress in vivo and in vitro. Free Radic Res. 39(2);153–62.
- [77] Rohr, P; Kvitko, K; da Silva, F.R; Menezes, A.P; Porto, C; Sarmento, M; Decker, N; Reyes, J.M; Allgayer, Mda C; Furtado, T.C; Salvador, M; Branco, C; da Silva, J. (2013a). Genetic and oxidative damage of peripheral blood lymphocytes in workers with occupational exposure to coal. Mutat Res. 758(1–2); 23–8.
- [78] Rohr, P; da Silva, J; da Silva, F.R; Sarmento, M; Porto, C; Debastiani, R; Dos Santos, C.E; Dias, J.F; Kvitko, K. (2013b). Evaluation of genetic damage in open-cast coal mine workers using the buccal micronucleus cytome assay. Environ Mol Mutagen. 54(1);65– 71.
- [79] León-Mejía, G; Espitia-Pérez, L; Hoyos-Giraldo, L.S; Da Silva, J; Hartmann, A; Henriques, J.A; Quintana, M. (2011). Assessment of DNA damage in coal open-cast mining workers using the cytokinesis-blocked micronucleus test and the comet assay. Sci Total Environ. 409(4);686–91.
- [80] León-Mejía, G; Quintana, M; Debastiani, R; Dias, J; Espitia-Pérez, L; Hartmann, A; Henriques, J.A; Da Silva, J. (2014). Genetic damage in coal miners evaluated by buccal micronucleus cytome assay. Ecotoxicol Environ Saf. 107;133–9.

- [81] Al-Sabti, K; Lloyd, D.C; Edwards, A; ,Stegnar P. (1992). A survey of lymphocyte chromosomal damage in Slovenian workers exposed to occupational clastogens. Mutat. Res. 280;215–23.
- [82] Avila Júnior,S; Possamai, FP; Budni, P; Backes, P; Parisotto, EB; Rizelio, VM; Torres, MA; Colepicolo, P; Wilhelm Filho, D. (2009). Occupational airborne contamination in south Brazil: 1. Oxidative stress detected in the blood of coal miners. Ecotoxicology 18(8);1150-7.
- [83] Donbak, L; Rencuzogullari, E; Yavuz, A; Topaktas, M. (2005). The genotoxic risk of underground coal miners from Turkey. Mutat Res. 588(2);82–7.
- [84] Minina, V.I; Kulemin, I.E; Tolotchko, T.A; Meĭer, A.V; Savtchenko, I.A; Volobaev, V.P; Gafarov, N.I; ,Semenikhina M.V. (2015). Genotoxic effects of occupational environment in Kuzbass miners. Med. Tr. Prom. Ekol. 5; 4–8.
- [85] Stierum, R; Hageman, G; Welle, I; Albering, H; Schreurs, J; Kleinjans, J. (1993). Evaluation of exposure reducing measures on parameters of genetic risk in a population occupationally exposed to coal fly ash. Mutat Res. 319;245–55.
- [86] Wolf, G; Arndt, D; Kotschy-Lang, N; Obe, G. (2004). Chromosomal aberrations in uranium and coal miners. Int J Radiat Biol. 80(2);147–53.
- [87] Ulker, O; Ustundag, A; Duydu, Y; Yucesoy, B; Karakaya, A. (2008). Cytogenetic monitoring of coal workers and patients with coal workers' pneumoconiosis in Turkey. Environ Mol Mutagen. 49;232–7.





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