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Effect of Allergy on Root Resorption Induced by Orthodontic Tooth Movement

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Additional information is available at the end of the chapter

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1. Introduction

Orthodontic teeth movement results from the application of orthodontic forces to teeth through orthodontic appliances which transmit the forces to the surrounding tissues. However, an undesirable and unexpected result of orthodontic treatment is the orthodontically induced tooth resorption (Fig. 1). The development of excessive root resorption during orthodontic treatment is considered an adverse side effect of the orthodontic tooth movement. Many factors have been investigated to explain the orthodontically induced root resorption observed among orthodontic patients. However, there are still controversies over this issue.

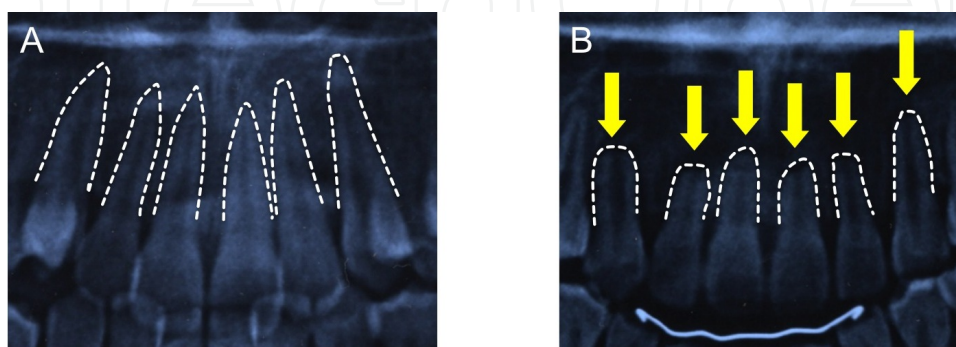


Figure 1. An undesirable and unexpected result of orthodontic treatment (A) Before treatment, (B) After treatment.

Davidovitch et al. [1] hypothesized that individuals who have a history of immune diseases including allergy, asthma, and systemic diseases may be at a high level of risk for developing excessive root resorption during the course of orthodontic treatment. To confirm the association between allergy and the orthodontically induced root resorption, we designed the epidemiological clinical study. Moreover, to extrapolate the direct effect of allergy on the orthodontically induced root resorption, which is difficult to prove in the clinical study design, we conducted experimental animal study using rats.

In this chapter, we refer at the beginning to the current knowledge of root resorption and then focus on the risk factors inherent in individuals to the external root resorption (ERR), especially allergic diseases as a possible risk factor of ERR.

2. Basic information

2.1. Prevalence

The presence of root resorption is common after orthodontic treatment [2]. Prevalence of root resorption following orthodontic treatment has been reported from 0% to 100% [3-5]. Thus, the prevalence varies widely due to differing methods of reporting root resorption, differing radiographic technique and interpretation, and individual susceptibility [2]. Close radiographic examination revealed that some loss of root length occurred in nearly every orthodontic patient. Moderate to severe apical root resorption (>2 mm, $<1/3$ of the root length) has been found in 10-17% of orthodontically treated patients [6,7] and excessive root resorption ($>1/3$ of the root length) in 1-5% [8,9]. Phillips [10] stated apical root resorption exceeding one-fourth of the original root length was shown in 1.5 % of maxillary central incisors and in 2.2 % of lateral incisors.

2.2. Site of root resorption

Regardless of patient-related or treatment-related factors, the maxillary incisors likely indicated more root resorption than the other teeth, followed by the mandibular incisors and first molars [11]. However, every tooth is capable of root resorption during orthodontic treatment.

2.3. Risk factors

2.3.1. Orthodontic treatment-related risk factors

Magnitude of force

Many studies have shown a distinct positive correlation between the applied orthodontic force and the amount of root resorption in both animal studies and clinical studies [12-16]. Owman-Mall et al. [17,18] reported that there were large individual variations in the amount of root resorption even when the same degree of orthodontic force was applied. They speculated that root resorption would not be very force-sensitive.

Duration of force

Several investigations revealed that the duration of force was one of the risk factors of orthodontically induced root resorption [19,20]. Some claimed that the duration of force application was an even more critical factor than the degree of orthodontic force [21,22]. Interaction between magnitude and duration of force would play an important role for orthodontic treatment-related risk factors. Large individual variations still existed even in well-controlled research design [17,18].

2.3.2. Patient-related risk factors (individual susceptibility)

Besides orthodontic treatment-related factors, patient-related risk factors have also been discussed previously. This includes systemic diseases [23], nutrition [24,25], age [26], trauma [10,27], previous root resorption [28,29], gender [20,28], nail biting habits [30], root morphology [31], endodontically treated teeth [32], gingivitis [9], allergy [33-35], and medications [36]. Al-Qawasmi et al. [37] identified an interleukin (IL)-1 β polymorphism in orthodontically treated individuals as having a role in the genetic influence on external apical root resorption.

3. The role of allergy related to orthodontically induced root resorption

Teeth are relocated under the dynamic balance of bone metabolism. The role of bone cells, including osteocytes, osteoblasts, and osteoclasts, are to resorb alveolar bone in sites of mechanically compressed tissues, thus enabling dental roots in these locations to move in the direction of the applied orthodontic force [38]. In these sites of tissue compression, one finds infiltrations of inflammatory cells followed by multinucleated osteoclasts and odontoclasts engaged in resorbing the dental roots [39,40]. Osteoclasts and their mononucleated progenitors are derived from the monocyte/macrophage lineage [41]. Osteoclasts and odontoclasts have a similar histochemical appearance; we defined here the multinucleated cells faced to tooth root as odontoclasts.

Patients with asthma and rhinitis share common physiology including heightened bronchial hyperresponsiveness and heightened reactivity to a variety of stimuli. Immunopathology of allergic rhinitis is also similar with a predominance of T-helper type 2 inflammation and tissue eosinophilia [42]. Allergy and asthma trigger the various associated biological, cellular, and molecular events with inflammation, such as increased vascular permeability, vasodilatation, cellular migration, increased mucus secretion, bronchoconstriction, structural changes of airway architecture, decline in pulmonary functions, release of intracellular mediators, increased formation of reactive oxygen species, cartilage degradation, and loss of function [43].

During orthodontic tooth movement, inflammation appears to be the main mechanism whereby immune cells and blood plasma reach the periodontal ligament of mechanically loaded teeth [44]. Apparently, strained sensory nerve endings in close proximity to periodontal ligament small blood vessels and capillaries release signal molecules such as substance P, vasoactive intestinal peptide, and calcitonin gene-related peptide [45]. Substance P stimulates

endothelial cells to adhere to circulating leukocytes and promote their migration to the extravascular space, where they secrete a variety of cytokines, such as interleukins and tumor necrosis factors (TNFs).

TNF- α and IL-1 β play an important role in bone pathophysiology [46,47] and have been suggested to be involved in orthodontic tooth movement [48]. These inflammatory cytokines induce local expression of receptor activator of nuclear factor- κ B ligand (RANKL), which is critical for the terminal differentiation of osteoclast precursor cells [49].

Furthermore, leukotrienes, which are metabolites of arachidonic acid, are potent lipid mediators that play an important role in a variety of allergic and inflammatory reactions and comprise several products of the 5-lipoxygenase (5-LOX) pathway [50–52]. Leukotrienes are produced by activated leukocytes in inflammatory diseases such as bronchial asthma and rheumatoid arthritis [53,54]. LTB₄ is known to play an important role in the allergic reactions induced by ovalbumin (OVA) in rodents [55] and has been suggested to modulate bone metabolism by increasing osteoclastic bone resorption [56].

In light of these facts, we hypothesize that patients with allergic diseases such as food allergy, allergic asthma, allergic rhinitis, and allergic conjunctivitis are susceptible to orthodontic root resorption. We show the scheme to explain the hypothesis of orthodontically induced root resorption due to the risk factor of allergy (Fig. 2).

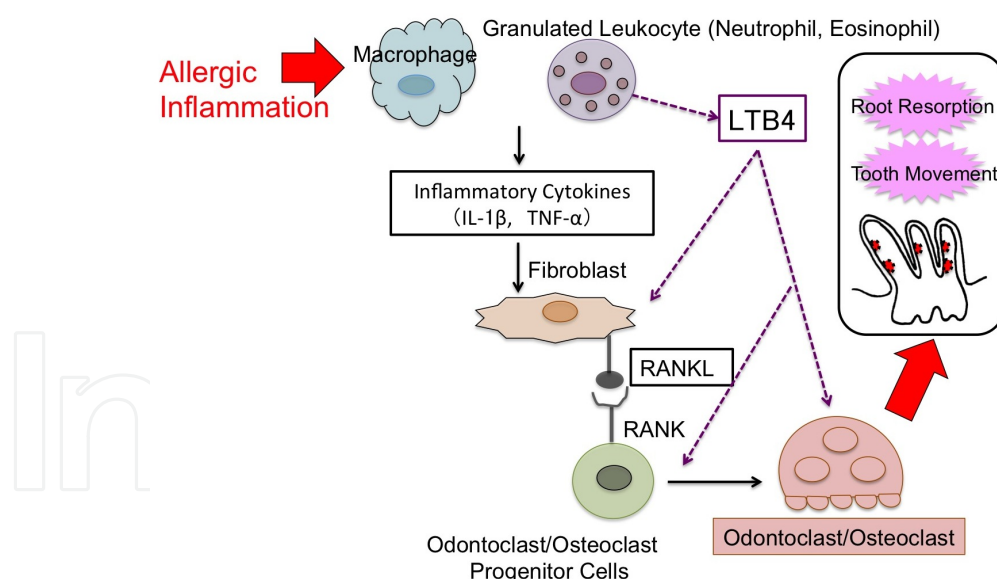


Figure 2. Scheme of possible explanation of the orthodontically induced root resorption due to the risk factor of allergy.

At first, we report the possible association between excessive root resorption during the orthodontic tooth movement and allergy using clinical study design. Next, we extrapolate the effect of allergy on orthodontically induced root resorption using animal models.

3.1. Epidemiological study

3.1.1. Sample

The records of 3500 patients were obtained from the Section of Orthodontics, Kyushu University Hospital, Fukuoka, Japan. All these patients have completed their course of orthodontic treatment. In this sample, 100 individuals were found to have developed excessive resorption of dental roots. In the root resorption group, 29 subjects were males and 71 were females (Table 1). A control group was selected from the remaining patients of this sample who did not display any significant radiographic evidence for dental root resorption in the light of root morphology at the end of their orthodontic treatment. Each individual in the control group was pair-matched with one in the root resorption group based on age, gender, treatment period, and the type of malocclusion (Table 1).

	Number	Age (y)	Treatment Period (y)
Root resorption group			
Male	29	18.9 ± 5.4	2.9 ± 0.8
Female	71	17.3 ± 6.5	3.1 ± 1.1
Total	100	17.8 ± 6.2	3.1 ± 1.1
Control group			
Male	29	17.9 ± 6.1	2.8 ± 0.8
Female	71	18.2 ± 4.8	2.9 ± 0.8
Total	100	18.1 ± 5.2	2.9 ± 0.8

Table 1. Comparison of Means and Standard Deviations of Age and Treatment Period in the Root Resorption and Control Groups

3.1.2. Methods

Determination of root resorption

Determination of the root resorption status of each patient was established by comparing the dental panoramic radiographs that had been taken before and after treatment. All teeth were measured along the tooth’s longitudinal axis for root length (defined as cement-enamel junction to apex of root). This method was similar to that used by Linge & Linge [7]. Individuals were assigned to the root resorption group when it was disclosed that one or more of their dental roots had been shortened by more than 25% of the original root length in the time that had elapsed between the two radiographs. The root shape was recorded subjectively as normal or abnormal (shortened, blunt, eroded, pointed, bent, bottle shaped) (Fig. 3) by examining dental panoramic radiographs that had been taken before the onset of treatment.

Questionnaire

The questionnaire sought information on the following subjects: personal details, medical history, dental history, and oral habits. Information on medical history included local or systemic diseases, details on medication uptake, hospitalizations, allergies, health during

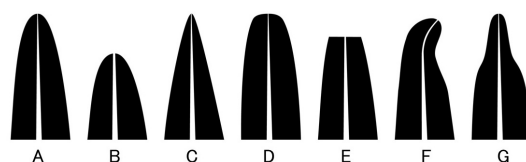


Figure 3. Morphology classification of the root (A) Normal, (B) Shortened, (C) Pointed, (D) Blunt, (E) Eroded, (F) Bent, (G) Bottle shaped.

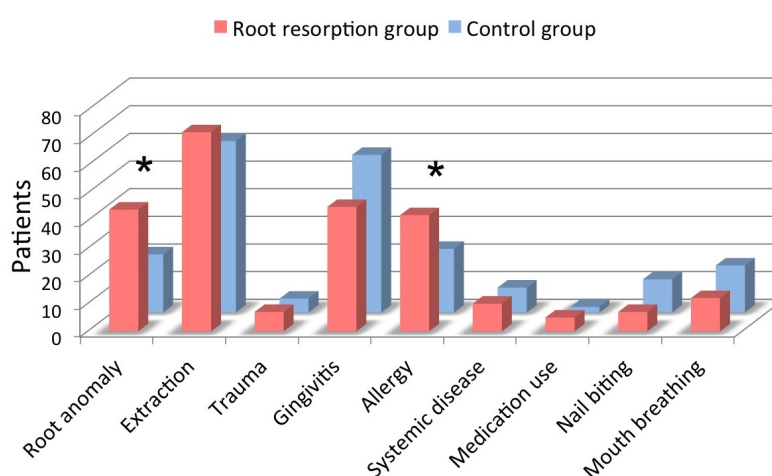
gestation and first year of life, childhood diseases, other conditions, and developmental events in infancy and childhood. Dental history questions referred to details on previous dental treatment and information about past oral injuries. Habits listed in the questionnaire were nail biting, tongue thrusting, and mouth breathing.

Statistical analysis

The validity of our hypothesis was tested by the logistic regression analysis using the Stat View 5.0 pro-gram (SAS Institute Inc, Cary, NC). Logistic regression produces odds ratios associated with each predictor variables (root anomaly, extraction, trauma, gingivitis, allergy, asthma, systemic disease, medication use, thumb sucking, nail biting, tongue thrusting, mouth breathing).

3.1.3. Results

The prevalence of excessive root resorption was 7.8%. The distribution of each risk factor in the root resorption and the control groups is shown in Figure 4. The logistic regression analysis is shown in Table 2. The incidence of allergy and asthma was significantly higher in the root resorption group ($P = 0.005$), with a mean odds ratio of 2.54 and 95% confidence interval of 1.34–4.92. The incidence of root anomaly was significantly higher in the root resorption group ($P = 0.001$), with a mean odds ratio of 2.95 and 95% confidence interval of 1.53–5.83.



*, $P < 0.05$.

Figure 4. Prevalence of each risk factor in the root resorption and control groups.

Risk Factors	P Value	Odds Ratio	95% Confidence Interval
Root anomaly	0.001	2.95	1.53-5.83
Extraction	0.168	1.56	0.83-2.98
Trauma	0.473	1.59	0.45-5.97
Gingivitis	0.120	0.62	0.34-1.13
Allergy	0.005	2.54	1.34-4.92
Systemic disease	0.900	0.93	0.32-2.70
Medication use	0.344	2.31	0.45-17.2
Nail biting	0.311	1.57	0.66-3.84
Mouth breathing	0.539	0.76	0.31-1.81

Table 2. Logistic Regression Analysis of Each Risk Factor

3.1.4. Discussion

This clinical study showed that allergy might be an aetiological factor in increased root resorption. The same association was found in earlier studies [1,57]. However, those studies were primarily performed on Caucasian subjects. Our finding, derived from an examination of the clinical records of Japanese subjects, supports the hypothesis that allergy is a high risk factor for the development of excessive root resorption during orthodontic treatment.

Our results also indicate that abnormal root shape is probably associated with excessive root resorption ($p=0.056$). This finding is in agreement with the results of the previous study [23,58-60]. If the same orthodontic force is applied to the dental crown, the root apex is exposed to increasing stress as the root becomes shorter. Additionally, when the same orthodontic force is applied to the root apex, the distribution of the stress is different according to the types of root anatomy, and the dental root with pointed or bent shape may be exposed to larger stress than roots with normal morphological features. These increased stresses may traumatize the apical periodontal ligament, followed by an inflammatory/repair process, which includes resorption of the root apex.

Patients with periodontal disease have circulating primed monocytes, and sera of patients with periodontal disease contain high levels of proinflammatory cytokines [8]. In this study, however, gingivitis did not have a significant association to orthodontic root resorption. This is most likely due to the sample selection of young patients; mean age of 17.8 ± 6.2 years, and the method used to detect the status of periodontal health; examination of intra-oral photographs.

In cases requiring tooth extraction, the remaining teeth are usually moved relatively great distance, particularly when maxillary incisors are retracted in order to reduce large overjet [58,59,61]. Additionally, biologically, tooth extraction and the ensuing wound healing attract vast numbers of immune cells to the extraction site. These inflammatory cells may directly spread from the wound site to tissues surrounding adjacent teeth or, indirectly, produce large

amounts of cytokines that enter the circulation and exit into the extravascular space in the periodontal ligament of neighboring mechanically stressed teeth, which regulate remodeling activities not only at extraction site but also in tissues surrounding adjacent teeth. However, in this study, we have not found a significant association between extraction of permanent teeth and orthodontic root resorption. Therefore, we conclude that healing of extraction sites is primarily a local event which does not promote the resorption of adjacent teeth.

One limitation of this study was using the panoramic radiographs in order to evaluate the amount of root resorption. Although this method of examination makes it possible to view the whole dentition, its main drawback is distortion of the tooth image, predominantly in the incisor region. Periapical radiographs would be preferable to determine the size and shape of dental roots, but these radiographs were not available to us this time.

3.2. Animal model study

In the experimental animal study, we used Brown–Norway (BN) rats, which are known to produce high levels of IgE after sensitization with OVA. BN rats have been used extensively as animal models of allergic asthma [62–64], atopic dermatitis [65], and food allergies [66,67].

This study aimed to determine whether systemic allergic inflammation had adverse effects on orthodontic tooth movement and bone and root resorption and, if so, to investigate the possible mechanisms of the acceleration. Furthermore, we examined the ability of aspirin, which has been reported to improve bone mineral density (BMD) in human epidemiological studies [68,69], to reverse tooth root resorption.

3.2.1. Allergen sensitization for rats and OF

Six-week-old male BN rats (weighing 110–140 g) were sensitized by a subcutaneous injection of saline (1 mL) containing 1 mg of OVA (grade V; Sigma-Aldrich, St. Louis, MO, USA) and 200 mg of aluminum hydroxide. A *Bordetella pertussis* vaccine containing 1×10^{10} heat-killed bacilli (Wako, Osaka, Japan) was given intraperitoneally as an adjuvant. After 7 days, OVA was injected for a booster effect. The serum IgE levels were measured using an IgE ELISA Kit (Shibayagi, Gunma, Japan). At 7 days after the second sensitization, an orthodontic appliance consisting of an Ni-Ti closed-coil spring (Sentalloy®, Ultra Light; Tomy International Inc., Tokyo, Japan) was inserted between the upper right first molar (M1) and the incisors. The orthodontic force (OF) level of the spring was set at approximately 0.1N (Fig. 5).

The animals were divided into four groups: OVA with OF group, OVA alone group (no OF), OF alone group (no OVA), and control group (no OVA or OF).

After 7 or 14 days of OF (at days 7 and 14, respectively), the animals were perfused transcardially. The two halves of the maxilla were decalcified in 10% ethylenediamine-tetraacetic acid and cut into 8- μ m-thick parasagittal sections. The sections were stained for tartrate-resistant acid phosphatase (TRAP) (Sigma-Aldrich) and counterstained with toluidine blue.

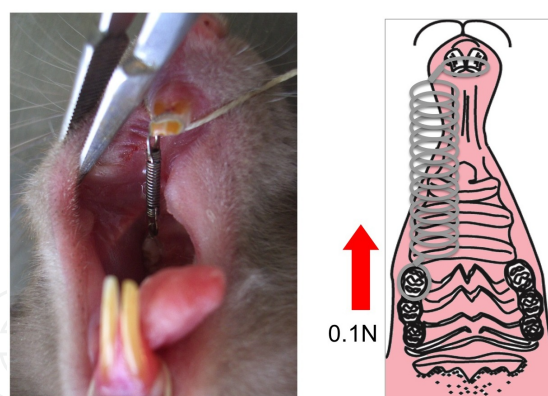


Figure 5. Maxillary first molar (M1) was moved medially with Ni-Ti Coil spring (0.1N) (Murata et al., 2013).

3.2.2. Measurements and analyses

We measured the area of ERR as well as the number of odontoclasts and osteoclasts within a defined area according to the method of Mavragani et al. [70]. TRAP-positive cells in resorption lacunae that faced the tooth roots were counted as odontoclasts, and other TRAP-positive cells (containing more than two nuclei) were counted as osteoclasts.

Proinflammatory cytokines were measured in the periodontal tissues, including the bone surrounding M1, after 24 hours of OF. The levels of TNF- α , IL-1 β , and IL-6 were evaluated using a TNF- α ELISA Kit (Shibayagi), IL-1 β ELISA Kit, and IL-6 ELISA Kit (Biosource, Camarillo, CA, USA), respectively, according to the manufacturers' protocols.

The lipid components were extracted in 100% methanol at -30°C overnight.

The periodontal tissues around M1 after 24 hours of OF were rapidly frozen in liquid nitrogen and homogenized. After mRNA isolation, quantitative real-time PCR or conventional RT-PCR was performed with gene-specific primer sets as described previously [71].

Micro-CT images were acquired at a resolution of 9 μm using a SkyScan 1076 (SkyScan, Antwerp, Belgium) to assess the degree of OF. At day 14 (10 weeks of age), the hemi-maxilla samples were excised. The hemi-maxilla samples were directed parallel to the occlusal plane and scanned for 2.70 mm. The distance from the distal surface of M1 to the mesial surface of the second molar (M2) was measured using DataViewer software (ver. 1.4.3; SkyScan).

Aspirin dissolved in feed water (250 $\mu\text{g}/\text{kg}/\text{day}$) was administered orally to OVA-sensitized rats with orthodontic force application. For histologic experiments, aspirin was administered from the initiation of OF. For ELISA and RT-PCR analyses, aspirin was administered from 24 hours before OF.

3.2.3. Statistical analysis

The total serum IgE was compared using Student's *t*-test. The means of multiple groups were compared by one-way ANOVA. Other values were compared by ANOVA followed by Tukey's multiple comparison test. Values of $P < 0.05$ were considered to indicate statistical significance.

3.2.4. Results

OVA sensitization, OF, and root and bone resorption

At day 14, the degree of tooth movement in the OVA with OF group was significantly larger than that in the OF alone group. Without orthodontic force application, no osteoclasts and/or odontoclasts were observed on the medial surface of the alveolar bone in any of the groups (Fig. 6A, 6B). On day 7, marked ERR was observed on the pressured side in the OF alone group, especially in the OVA with OF group (Figs. 7A, 7B). On day 14, these tendencies became further significant in both the OF alone and the OVA with OF groups (Figs. 8A, 8B). The OVA with OF group exhibited significantly broader and deeper TRAP-stained areas compared to the OF alone group on days 7 as well as 14 (Fig. 9A). The number of odontoclasts and osteoclasts was significantly greater in the OVA with OF group than in the OF alone group on day 7 (Figs. 9B, 9C). The number of osteoclasts peaked on day 7 in the OVA with OF group and then decreased by day 14. While the area of ERR increased from day 7 to day 14, the number of odontoclasts was maintained through days 7 to 14 (Figs. 9A, 9C). The alveolar bone resorption and osteoclast number were significantly greater in the OVA with OF group than in the OF alone group on days 7 and 14 (Figs. 9D, 9E).

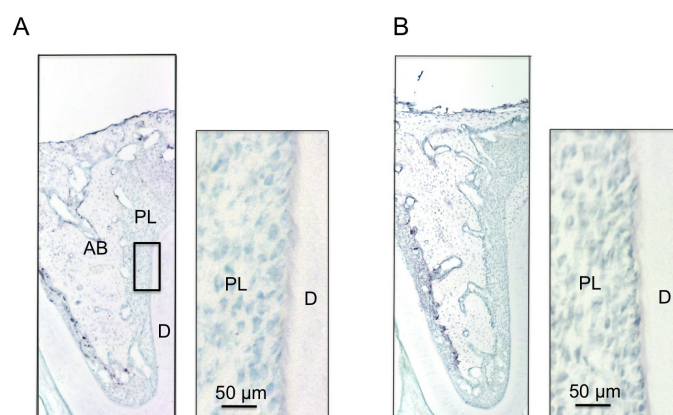


Figure 6. Histologic sections of periodontal tissues around the distopalatal root of the first molar stained for TRAP. (A) Control group (no treatment), (B) OVA-alone group (Murata et al., 2013).

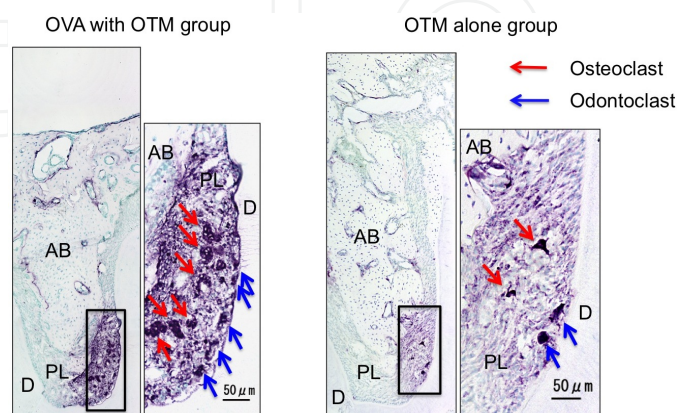


Figure 7. Histologic sections of periodontal tissues around the distopalatal root of the first molar stained for TRAP on day 7. (A) OVA with OTM group, (B) OTM-alone group (Murata et al., 2013).

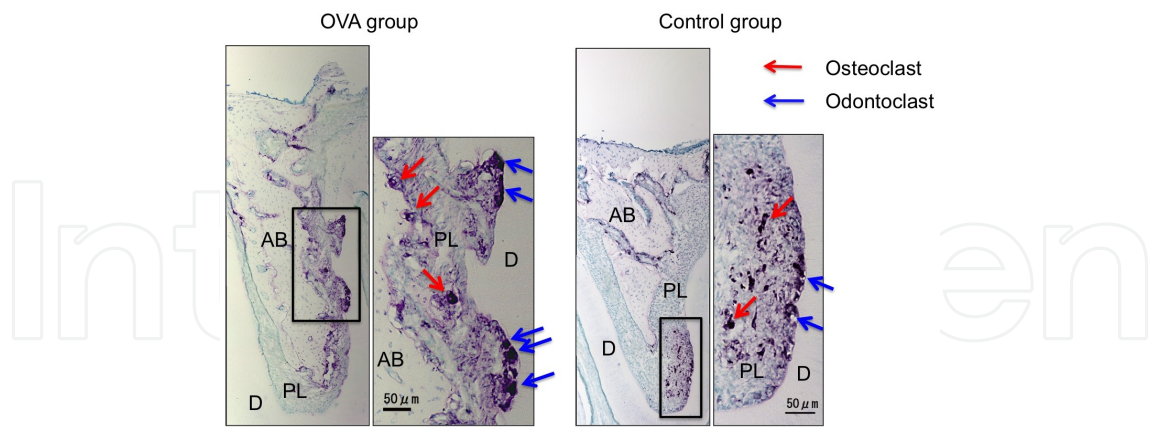


Figure 8. Histologic sections of periodontal tissues around the distopalatal root of the first molar stained for TRAP on day 14. (A) OVA with OTM group, (B) OTM-alone group (Murata et al., 2013).

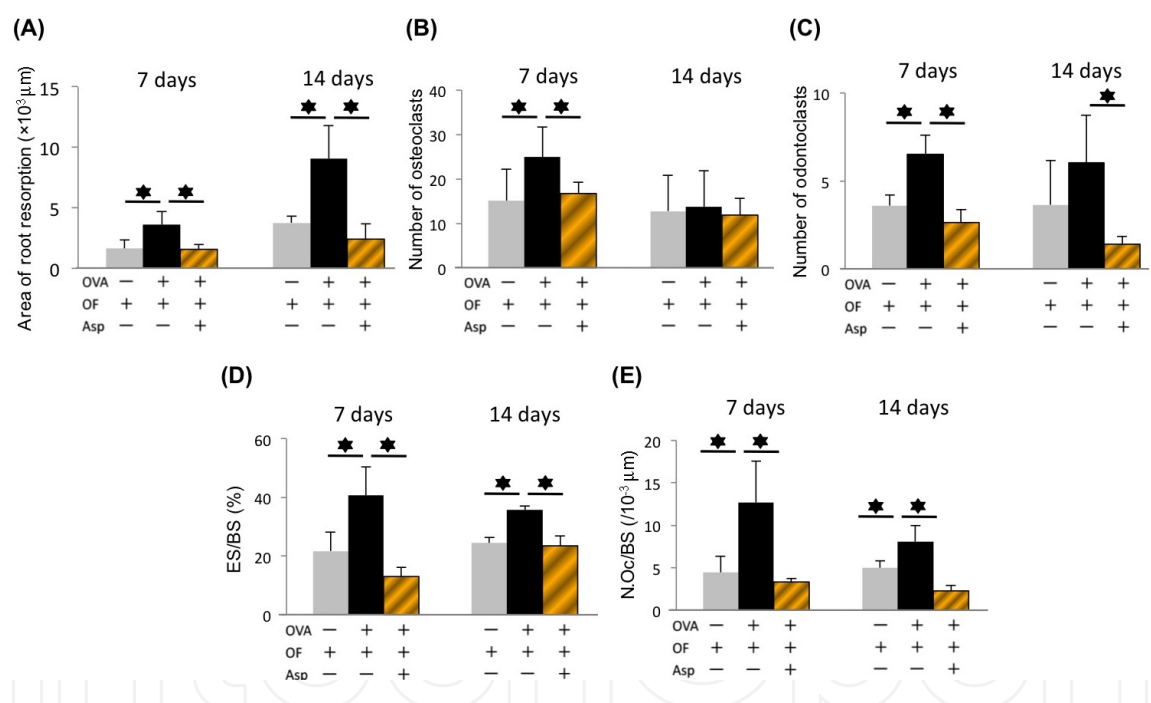


Figure 9. The numbers of osteoclasts/odontoclasts and amounts of root/bone resorption on the pressured side after tooth movement. (A) Area of root resorption (×103 μm2), (B) Number of osteoclasts, (C) Number of odontoclasts, (D) Eroded surface per alveolar bone surface (ES/ BS), (E) Number of osteoclasts per alveolar bone surface (N.Oc/BS). *: *P* < 0.05. (Murata et al., 2013).

Pro-inflammatory cytokines

The expression levels of RANKL, TNF-α, and IL-1β were significantly higher in the OVA with OF group than in the other groups after 24 hours of OF (Figs. 10A–10C), while the difference in IL-6 was not significant (Fig. 10D). Although no significant differences were observed, the OF alone group showed tendencies toward higher levels of TNF-α and IL-1β (Figs. 10B, 10C).

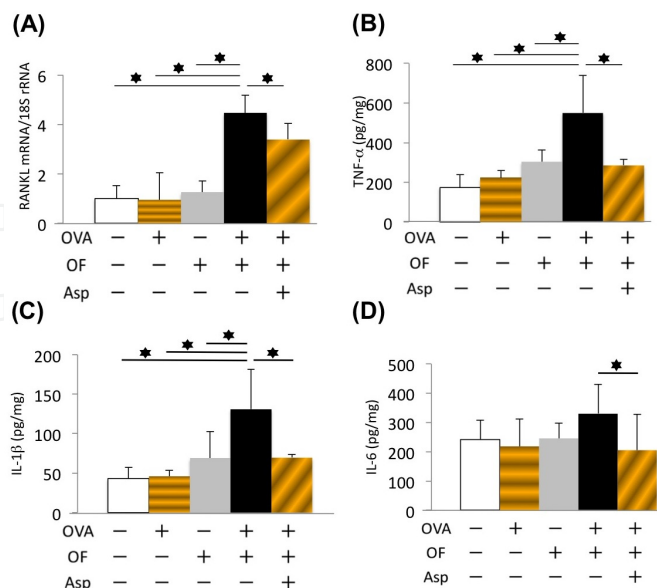


Figure 10. Concentrations of RANKL and inflammatory cytokines in rat periodontal tissues with or without OVA sensitization, after 24 h of orthodontic force application, and after aspirin administration. (A) RANKL, (B) TNF- α , (C) IL-1 β , (D) IL-6. *: $P < 0.05$. (Murata et al., 2013).

Leukotrienes

The LTB₄ level and mRNA levels of the leukotriene synthases LTA₄h were significantly increased in the OVA with orthodontic force group after 24 hours of orthodontic force compared with the other groups (Figs. 11A, 11B).

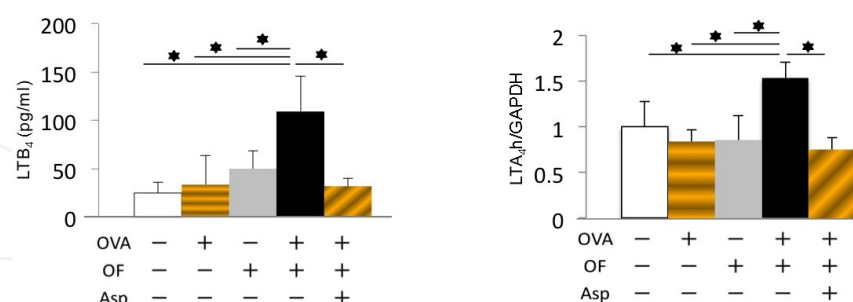


Figure 11. Effects of OVA sensitization, OF application for 24 h, and aspirin administration on the leukotriene synthases. (A) LTB₄, (B) LTA₄h. *: $P < 0.05$. (Murata et al., 2013).

Aspirin effects

Aspirin administration reversed the degree of tooth movement in the OVA with OF group to a similar level of the OF alone group on day 14. Histologic examinations revealed that the area of ERR and the numbers of odontoclasts and osteoclasts were decreased to the levels observed in the OF alone group (Figs. 9A–9E). The expression levels of RANKL mRNA and TNF- α ,

IL-1 β , and IL-6 proteins were also suppressed in the levels detected in the OF alone group (Figs. 10A–10D). Furthermore, the expression levels of LTB₄ and LTA₄h were suppressed by aspirin treatment (Fig. 11).

3.2.5. Discussion

We demonstrated that allergies affect ERR during orthodontic treatment using an animal model. It is notable that the degree of OF in the present study was greater in the OVA-sensitized animals, suggesting that allergies can affect OF. In the histological observations, we found that the OVA-sensitized rats showed increased numbers of odontoclasts and osteoclasts on the pressured side during OF. The present findings suggested that allergies enhanced both odontoclastogenesis and osteoclastogenesis under the condition of orthodontic force.

TNF- α , IL-1 β , and IL-6, which are well-known proinflammatory cytokines, were detected at high levels in the periodontal tissues from the OVA with OF group. Recently, Polzer et al. [72] suggested that IL-1 β contributes to TNF-mediated inflammatory osteopenia, reporting that TNF- α transgenic IL-1 β -deleted mice were protected against bone loss. The elevated expression levels of TNF- α and IL-1 β , together with RANKL as an essential cytokine for osteoclastogenesis, suggested boosts in the rates and magnitudes of ERR as well as orthodontic tooth movement due to the differentiation of osteoclasts and odontoclasts.

Lipid mediators derived from arachidonic acid through the lipoxygenase and cyclooxygenase pathways are known to act as important modulators of inflammation. Increased levels of leukotrienes are found in inflammatory diseases such as asthma, arthritis, and periodontal disease [73–75]. LTB₄ has been reported to stimulate osteoclast formation and bone resorption in the mouse calvaria [55] and to inhibit osteogenesis [76]. In our model, the expression levels of the leukotriene synthases LTB₄ and LTA₄h were increased in the periodontal tissues in OVA-sensitized rats with OF. The fact that mechanical loading with OVA sensitization elevated the expression of leukotriene synthases reveals a potential role for leukotrienes in bone and root resorption. Interestingly, these effects of mechanical OF suggest the involvement of the lipoxygenase pathway in response to mechanical loading. We assume that the present findings are compatible with our previous epidemiologic observations [33,34].

We further found that oral administration of aspirin was able to reverse the increases in ERR and degree of OF in the allergic model. The dose of aspirin in the present study, 250 μ g/kg/day, was much lower than that for general use as a painkiller. A recent study showed that low-dose aspirin inhibited ovariectomy-induced osteoporosis by inhibiting T cell activation [77]. Our findings raise the possibility that low-dose aspirin administration inhibits osteoclastogenesis under inflammatory conditions.

Through the epidemiological and animal model studies, we have demonstrated that allergies would be possible risk factors of ERR in the course of orthodontic treatments. In the clinical study, the evidence would be strengthened using a prospective study involving a much larger sample, such as the randomized controlled test. Moreover, we need to investigate the unsolved

mechanism of ERR, and the elucidation of the pharmacological targets for ERR during orthodontic tooth movement would be warranted.

4. Conclusions

In the clinical study, we found that allergy and root morphology abnormalities may be considered high-risk factors for the development of excessive root resorption during the course of orthodontic treatment.

We have proposed a model for studying the effects of allergen-induced inflammation and/or mechanical stress on ERR during OTM. This process was affected by pro-inflammatory cytokines, together with lipid mediators. Furthermore, our findings suggest that pro-inflammatory cytokines and leukotrienes, together with low-dose aspirin, may represent pharmacological targets for ERR during orthodontic treatment with allergic conditions.

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References

- [1] Davidovitch Z, Lee YJ, Counts AL, Park YG and Bursac Z. The immune system possibly modulates orthodontic root resorption. In: Davidovitch Z. (ed.) *Biological Mechanisms of Tooth Movement and Craniofacial Adaptation*. Oklahoma City: University of Oklahoma; 2000. p207-217.
- [2] Thilander B, Rygh P, Reitan K. Tissue reactions in orthodontics. In: Graber TM, Varnarsdall RL. (eds.) *Orthodontics: Current Principles and Techniques*. 3rd ed. St Louis: Mosby; 2000. p117-191.
- [3] Henry J, Weinmann J. The pattern of resorption and repair of human cementum. *Journal of the American Dental Association* 1951;42(3) 270-290.
- [4] Massler M, Perreault J. Root resorption in the permanent teeth of young adults. *Journal of Dentistry for Children* 1954;21 158-164.
- [5] Copeland S, Green L. Root resorption in maxillary central incisors following active orthodontic treatment. *American Journal of Orthodontics and Dentofacial Orthopedics* 1986;89(1) 51-55.
- [6] Hollender L, Rönnerman A, Thilander B. Root resorption, marginal bone support and clinical crown length in orthodontically treated patients. *European Journal of Orthodontics* 1980;2(4) 197-205.
- [7] Linge L, Linge BL. Patients characteristics and treatment variables associated with apical root resorption during orthodontic treatment. *American Journal of Orthodontics and Dentofacial Orthopedics* 1991;99(1) 35-43.
- [8] Levander E, Malmgren O. Evaluation of the risk of root resorption during orthodontic treatment: a study of upper incisors. *European Journal of Orthodontics* 1988;10(1) 30-38.
- [9] Davidovitch Z. Etiologic factors in force-induced root resorption. In: Davidovitch Z, Norton LA. (eds.) *Biological Mechanisms of Tooth Movement and Craniofacial Adaptation*. Boston: Harvard Society for the Advancement of Orthodontics; 1996. p349-355.
- [10] Phillips J. *Apical Root Resorption under Orthodontic Therapy*: University of Washington. Seattle; 1955.
- [11] Weltman B, Vig KWL, Fields HW, Shanker S, Kaizar EE. Root resorption associated with orthodontic tooth movement. *American Journal of Orthodontics and Dentofacial Orthopedics* 2010;137(4) 462-476.
- [12] Reitan K. Effects of force magnitude and direction of tooth movement on different alveolar bone types *Angle Orthod.* 1964;34(4) 244-255.

- [13] Dellinger EL. A histologic and cephalometric investigation of premolar intrusion in the *Macaca speciosa* monkey. *American Journal of Orthodontics*. 1967;53(5) 325-355.
- [14] Kvam E. Scanning electron microscopy of tissue changes on the pressure surface of human premolars following tooth movement Scandinavian. *Journal of Dental Research* 1972;80(5) 357-368.
- [15] King GJ, Fischlschweiger W. The effect of force magnitude on extractable bone resorption activity and cemental cratering in orthodontic tooth movement. *Journal of Dental Research* 1982;61(6) 775-779.
- [16] Pilon JJGM, Kuijpers-Jagtman AM, Maltha JC. Magnitude of orthodontic forces and rate of bodily tooth movement, an experimental study in beagle dogs. *American Journal of Orthodontics and Dentofacial Orthopedics* 1996;110(1) 16-23.
- [17] Owman-Moll P, Kurol J, Lundgren D. Effects of doubled orthodontic force magnitude on tooth movement and root resorption. An interindividual study in adolescents. *European Journal of Orthodontics* 1996;18(2) 141-150.
- [18] Owman-Moll P, Kurol J, Lundgren D. The effects of a four-fold increased force magnitude on tooth movement and root resorptions. An intraindividual study in adolescents. *European Journal of Orthodontics* 1996;18(3) 287-295.
- [19] DeShields RW. A study of root resorption in treated Class II, division I malocclusions. *Angle Orthodontist* 1969;39(4) 231- 245.
- [20] McFadden WM, Engstrom C, Engstrom H, Anholm JM. A study of the relationship between incisor intrusion and root shortening. *American Journal of Orthodontics and Dentofacial Orthopedics* 1989;96(5) 390-396.
- [21] Stenvik A, Mjör IA. Pulp and dentine reactions to experimental tooth intrusion. A histological study of the initial changes. *American Journal of Orthodontics* 1970;57(4) 370-385.
- [22] Harry MR, Sims MR. Root resorption in bicuspid intrusion. A scanning electron microscopy study. *Angle Orthodontist* 1982;52(3) 235-258.
- [23] Newman WG. Possible etiological factors in external root resorption. *American Journal of Orthodontics* 1975;67(5) 522-539.
- [24] Becks H. Root resorption and their relation to pathologic bone formation. Part 1. *International Journal of Orthodontia and Oral Surgery* 1936;22 445-482.
- [25] Becks H. Orthodontic prognosis: evaluation of routine dentomedical examinations to determine "good and poor risks." *American Journal of Orthodontics and Oral Surgery* 1936;25(7) 610-624.
- [26] Malmgren O, Goldson L, Hill C, Orwin A, Petrini L, Lundberg M. Root resorption after orthodontic treatment of traumatized teeth. *American Journal of Orthodontics* 1982;82(6) 487-491.

- [27] Linge B, Linge L. Apical root resorption in upper anterior teeth. *European Journal of Orthodontics* 1983;5(3) 173-183.
- [28] Massler M, Malone AJ. Root resorption in human permanent teeth: A roentgenographic study. *American Journal of Orthodontics* 1954;40(8) 619-633.
- [29] Kalley J, Phillips C. Factors related to root resorption in edgewise practice. *Angle Orthodontist* 1991;61(2) 125-132.
- [30] Odenrick L, Brattström V. The effect of nailbiting on root resorption during orthodontic treatment. *European Journal of Orthodontics* 1983;5(3) 185-188.
- [31] Levander E, Malmgren O, Eliasson S. Evaluation of root resorption in relation to two orthodontic treatment regimes. A clinical experimental study. *European Journal of Orthodontics* 1994;16(3) 223-228.
- [32] Mirabella AD, Aertun J. Prevalence and severity of apical root resorption of maxillary anterior teeth in adult orthodontic patients. *European Journal of Orthodontics* 1995;17(2) 93-99.
- [33] Nishioka M, Ioi H, Nakata S, Nakasima A, Counts AL. Association between orthodontic root resorption and factors of the immune system in Japanese. In: Davidovitch Z, Norton LA. (eds.) *Biological Mechanisms of Tooth Movement and Craniofacial Adaptation*. Boston: Harvard Society for the Advancement of Orthodontics; 2004. P131-136.
- [34] Nishioka M, Ioi H, Nakata S, Nakasima A, Counts AL. Root resorption and immune system factors in the Japanese. *Angle Orthod* 2006;76(1) 103-108.
- [35] Murata N, Ioi H, Ouchi M, Takao T, Oida H, Aijima R, Yamaza T, Kido MA. Effects of allergen sensitization on external root resorption. *Journal of Dental Research* 2013;92(7) 641-647.
- [36] Kameyama Y, Nakane S, Maeda H, Fujita M, Takesue M, Sato E. Inhibitory effect of aspirin on root resorption induced by mechanical injury of the soft periodontal tissue in rats. *Journal of Periodontal Research* 1994;29(2) 113-117.
- [37] Al-Qawasmi RA, Hartsfield JK Jr, Everett ET, Flury L, Liu L, Foroud TM, Macri JV, Roberts WE. Genetic predisposition to external apical root resorption. *American Journal of Orthodontics and Dentofacial Orthopedics* 2003;123(3) 242-252.
- [38] Reitan K. Tissue behavior during orthodontic tooth movement. *American Journal of Orthodontics* 1960;46(12) 881-890.
- [39] Reitan K. Initial tissue behavior during apical root resorption. *Angle Orthodontist* 1974;44(1) 68-82.
- [40] Brudvik P, Rygh P. The initial phase of orthodontic root resorption incident to local compression of the periodontal ligament. *European Journal of Orthodontics* 1993;15(4) 249-263.

- [41] Rossi M, Whitcomb S, Lindemann R. Interleukin-1 β and tumor necrosis factor- α production by human monocytes cultured with L-thyroxine and thyrocalcitonin: Relation to severe root resorption. *American Journal of Orthodontics* 1996;110(4) 399-404.
- [42] Khan DA. Allergic rhinitis and asthma: Epidemiology and common pathophysiology. *Allergy and Asthma Proceedings* 2014;35(5) 357-361.
- [43] Naik SR, Wala SM. Inflammation, Allergy and Asthma, Complex Immune Origin Diseases: Mechanisms and Therapeutic Agents. *Recent Patents on Inflammation & Allergy Drug Discovery* 2013;7(1) 62-95.
- [44] Storey E. The nature of tooth movement. *Am J Orthod* 1970;63(3) 292-314.
- [45] Davidovitch Z. Tooth movement. *Crit Rev Oral Biol Med* 1991;2(4) 411-35.
- [46] Redlich K, Hayer S, Ricci R, David JP, Tohidast-Akrad M, Kollias G, Steiner G, Smolen JS, Wagner EF, Schett G. Osteoclasts are essential for TNF- α -mediated joint destruction. *Journal of Clinical Investigation* 2002;110(10) 1419-1427.
- [47] Buchs N, Giovine FS di, Silvestri T, Vannier E, Duff GW, Miossec P. IL-1B and IL-1Ra gene polymorphisms and disease severity in rheumatoid arthritis: interaction with their plasma levels. *Genes and Immunity* 2001;2(4) 222-228.
- [48] Tian YL, Xie JC, Zhao ZJ, Zhang Y. Changes of interleukin-1 β and tumor necrosis factor- α levels in gingival crevicular fluid during orthodontic tooth movement. *Hua Xi Kou Qiang Yi Xue Za Zhi* 2006;24(3) 243-245.
- [49] Azuma Y, Kaji K, Katogi R, Takeshita S, Kudo A, Tumor necrosis factor- α induces differentiation of and bone resorption by osteoclasts, *J. Biol. Chem* 2000;275(7) 4858-4864.
- [50] Samuelsson B. Leukotrienes: mediators of immediate hypersensitivity reactions and inflammation. *Science* 1983;220(4597) 568-575.
- [51] Henderson WR Jr. The role of leukotrienes in inflammation, *Annals of Internal Medicine* 1994;121(9) 684-697.
- [52] Murphy RC, Gijon MA. Biosynthesis and metabolism of leukotrienes. *Biochemical Journal* 2007;405(3) 379-395.
- [53] Sala A, Folco G. Neutrophils, endothelial cells, and cysteinyl leukotrienes: a new approach to neutrophil-dependent inflammation? *Biochemical Biophysical Research Communication* 2001;283(5) 1003-1006.
- [54] Yu W, Xu L, Martin JG, Powell WS. Cellular infiltration and eicosanoid synthesis in brown Norway rat lungs after allergen challenge. *American Journal of Respiratory Cell and Molecular Biology* 1995;13(4) 477-486.

- [55] Garcia C, Boyce BF, Gilles J, Dallas M, Qiao M, Mundy GR, Bonewald LF. Leukotriene B4 stimulates osteoclastic bone resorption both in vitro and in vivo. *Journal of Bone and Mineral Research* 1996;11(11) 1619-1627.
- [56] Hikiji H, Ishii S, Yokomizo T, Takato T, Shimizu T. A distinctive role of the leukotriene B4 receptor BLT1 in osteoclastic activity during bone loss, *Proceedings of National Academy of Sciences of the United States of America* 2009;106(50) 21294-21299.
- [57] Owman-Moll P, Kurol J. Root resorption after orthodontic treatment in high- and low-risk patients: analysis of allergy as a possible predisposing factor. *European Journal of Orthodontics* 2000;22(6) 657-663.
- [58] Kjær I. Morphological characteristics of dentitions developing excessive root resorption during orthodontic treatment. *European Journal of Orthodontics* 1995;17(1) 25-34.
- [59] Mirabella AD, Årtun J. Risk factors apical root resorption of maxillary anterior teeth in adult orthodontic patients. *American Journal of Orthodontics and Dentofacial Orthopedics* 1995;108(1) 48-55.
- [60] Lee RY, Årtun J, Alonzo TA. Are dental anomalies risk factors for apical root resorption in orthodontic patients? *American Journal of Orthodontics and Dentofacial Orthopedics* 1999;116(2) 187-195.
- [61] Costopoulos G, Nanda R. An evaluation of root resorption incident to orthodontic intrusion. *American Journal of Orthodontics and Dentofacial Orthopedics* 1996;109(5) 543-548.
- [62] Herszberg B, Ramos-Barbon D, Tamaoka M, Martin JG, Lavoie JP. Heaves, an asthma-like equine disease, involves airway smooth muscle remodelling. *Journal of Allergy and Clinical Immunology*. 2006;118(2) 382-388.
- [63] Martin JG, Tamaoka M. Rat models of asthma and chronic obstructive lung disease. *Pulmonary Pharmacology & Therapeutics* 2006;19(6) 377-385.
- [64] Xu KF, Vlahos R, Messina A, Bamford TL, Bertram JF, Stewart AG. Antigen-induced airway inflammation in the Brown Norway rat results in airway smooth muscle hyperplasia. *Journal of Applied Physiology* 2002;93(5) 1833-1840.
- [65] Fujii Y, Takeuchi H, Sakuma S, Sengoku T, Takakura S. Characterization of a 2,4-dinitrochlorobenzene-induced chronic dermatitis model in rats. *Skin Pharmacology and Physiology* 2009;22(5) 240-247.
- [66] De Jonge JD, Ezendam J, Knippels LM, Odink J, Pourier MS, Penninks AH, Pieters R, van Loveren H, Bis(tributyltin)oxide (TBTO) decreases the food allergic response against peanut and ovalbumin in Brown Norway rats. *Toxicology* 2007;239(1-2) 68-76.

- [67] Jia XD, Li N, Wu YN, Yang XG. Studies on BN rats model to determine the potential allergenicity of proteins from genetically modified foods. *World Journal of Gastroenterology* 2005;11(34) 5381–5384.
- [68] Bauer DC, Orwoll ES, Fox KM, Vogt TM, Lane NE, Hochberg MC, Stone K, Nevitt MC. Aspirin and NSAID use in older women: effect on bone mineral density and fracture risk, Study of Osteoporotic Fractures Research Group. *Journal of Bone and Mineral Research* 1996;11(1) 29-35.
- [69] Carbone LD, Tylavsky FA, Cauley JA, Harris TB, Lang TF, Bauer DC, Barrow KD, Kritchevsky SB. Association between bone mineral density and the use of nonsteroidal anti-inflammatory drugs and aspirin: impact of cyclooxygenase selectivity. *Journal of Bone and Mineral Research* 2003;18(10) 1795-1802.
- [70] Mavragani M, Brudvik P, Selvig KA. Orthodontically induced root and alveolar bone resorption: inhibitory effect of systemic doxycycline administration in rats. *European Journal of Orthodontics* 2005;27(3) 215-225.
- [71] Wang B, Danjo A, Kajiya H, Okabe K, Kido MA. Oral epithelial cells are activated via TRP channels. *Journal of Dental Research* 2011;90(2) 163-167.
- [72] Polzer K, Neubert K, Meister S, Frey B, Baum W, Distler JH, Gückel E, Schett G, Voll RE, Zwerina J. Proteasome inhibition aggravates TNF-mediated bone resorption. *Arthritis & Rheumatology* 2011;63(3) 670-680.
- [73] Hallstrand TS, Henderson WR Jr. An update on the role of leukotrienes in asthma. *Current Opinion and Allergy and Clinical Immunology* 2010;10(1) 60-66.
- [74] Davidson EM, Rae SA, Smith MJ. Leukotriene B4, a mediator of inflammation present in synovial fluid in rheumatoid arthritis, *Annals of the Rheumatic Diseases* 1983;42(6) 677-679.
- [75] Emingil G, Cinarcik S, Baylas H, Coker I, Huseyinov. A Levels of leukotriene B4 in gingival crevicular fluid and gingival tissue in specific periodontal diseases. *Journal of Periodontology* 2001;72(8) 1025-1031.
- [76] Traianedes K, Dallas MR, Garrett IR, Mundy GR, Bonewald LF. 5-Lipoxygenase metabolites inhibit bone formation in vitro. *Endocrinology* 1998;139(7) 3178-3184.
- [77] Yamaza T, Miura Y, Bi Y, Liu Y, Akiyama K, Sonoyama W, Patel V, Gutkind S, Young M, Gronthos S, Le A, Wang CY, Chen W, Shi S. Pharmacologic stem cell based intervention as a new approach to osteoporosis treatment in rodents. *PLoS One* 2008;3(7) e2615.