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Exploitation of Chinese Upland Cotton Cultivar Germplasm Resources to Mine Favorable QTL Alleles Using Association Mapping

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Additional information is available at the end of the chapter

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1. Introduction

Cotton (*Gossypium* spp.) is the most important natural textile fiber source globally. The worldwide economic impact of the cotton industry is estimated at approximately \$500 billion per year with an annual utilization of about 27 million metric tons of cotton fiber [1]. The tetraploid species *Gossypium hirsutum* L. (n=26, AD genome), commonly referred to as Upland cotton, accounts for 95% of the world's cotton production [2]. Most of the objectives in cotton breeding, such as yield, fiber quality, biotic and abiotic stress tolerance, are all complex traits, which are controlled by a large number of quantitative trait loci (QTLs). It is becoming progressively more difficult to improve these traits using conventional breeding methods due to their complex architecture and inheritance [3]. Fortunately, the development in applied genomics research has provided alternative tools to improve efficiency in plant breeding programs. Molecular markers tightly linked to the target genes or QTLs can be used for marker-assisted selection (MAS) and/or genomic selection (GS) [4-5]. In the past two decades, the availability of abundant molecular markers has made tagging QTL harboring functional genes through family-based linkage mapping a routine process, and a large number of QTLs for fiber quality properties [6-40], yield and its components [6, 8, 10-11, 14, 18-19, 27, 30-32, 36, 40-45], nematode resistance [46-49], Verticillium wilt resistance [50-53] as well as Fusarium wilt resistance [54-60] have been identified in cotton.

However, approximately 80% of the previously reported QTLs could not be confirmed in subsequent studies, and few have actually been applied in breeding programs [61-62]. This may be because most QTLs were population-specific, and the genetic variations detected

in a unique bi-parental population were not shared with other genetic populations, or shared but fixed in the parental lines. In addition, limited genetic recombination events in most populations used for linkage mapping make it difficult to map QTL with a high resolution, which severely limits their application in breeding programs. With the potential to exploit all recombination events that occurred in the evolutionary history of natural populations, linkage disequilibrium (LD) based association mapping (AM) has become a powerful approach for the dissection of complex traits and identification of causal variation with modest effects for target traits in many plant species [3, 63] including cotton [64-73]. Furthermore, although AM has been successfully used to detect the QTLs underlying quantitative traits in some crops, from a breeding standpoint, detecting associated loci is just the first step; analyzing the genetic effects of alleles and identifying favorable alleles will be more beneficial for target trait improvement. For example, Breseghello and Sorrells identified several potentially beneficial alleles for kernel size and milling quality by comparing the average phenotypic value with specific alleles and null alleles in a soft winter wheat population [74]; Jia et al. identified some putative resistant alleles for Sheath Blight resistance in a rice panel composed of 217 accessions from the USDA core collection, and found that the number of putative resistant alleles presented in an entry was highly and significantly correlated with the decrease of ShB rating [75].

China is the world's largest cotton-growing nation, but not a cotton domestication region. Most Upland cotton cultivars developed in China were derived from a few germplasm resources such as Deltapine (DPL), Stoneville (STV), Foster, King and Uganda, all of which were introduced from abroad [76]. More than 8800 accessions have been collected, developed, and maintained in Chinese cotton germplasm collection [77]. Among these collections, *G. hirsutum* varieties and breeding lines comprise about 83% of the total. Current and obsolete cultivars have been and continue to be the main resources for cotton breeding programs in China. Dissecting the genetic basis of main breeding objective traits will be of great benefits to germplasm evaluation and future molecular breeding. In this chapter, we reviewed QTLs underlying yield and its components, fiber quality properties and Fusarium wilt resistance detected by association mapping and their favorable alleles identified in 2 Chinese Upland cotton AM panels [71-73]. We hope that these results should provide useful information for further understanding the genetic basis of these traits, and would facilitate future breeding process by MAS in Upland cotton.

2. Favorable QTL alleles for yield and its components

In recent years, demand for cotton fiber in the world market has dramatically increased. While cotton acreage has declined worldwide in the past few years, mainly it was due to strong competition from other crops as well as production costs [1]. To improve lint yield of Upland cotton cultivars will remain critical for meeting worldwide demand and maintaining profitability for cotton growers. A total of 356 representative Upland cotton cultivars and breeding lines were selected from the cotton germplasm collection in Cotton Research Institute, Chinese Academy of Agricultural Sciences (CRI-CAAS), and from the collection in our laboratory, and

assembled to construct an AM panel. The population consisted of 348 cultivars developed in China, seven introduced from the U.S. including the genetic standard line TM-1, and one introduced from Uganda. According to their release year, the 348 Chinese cultivars could be divided into the following six groups: I (1930–1960, 26 lines); II (1961–1970, 26 lines); III (1971–1980, 39 lines), IV (1981–1990, 83 lines); V (1991–2000, 125 lines); and VI (2000–2005, 49 lines). The cultivars introduced from abroad, DPL 15, DPL 16, STV 2B, King, Foster 6 and Uganda 3, were used as a check group for genetic diversity and allele transmission evaluation, because they had been used as the main founder parents in China's Upland cotton breeding programs [71]. Yield and its components of the accessions were evaluated in three diverse environments, including lint yield per plant (LP), seed cotton yield per plant (SY), bolls per plant (BN), boll weight (BW), lint percentage (LP), lint index (LI) and seed index (SI). Three hundred and eighty-one pairs of SSR primers that amplify loci evenly covering the tetraploid cotton genome [78] were selected to genotype the 356 accessions.

The Bayesian model-based program STRUCTURE 2.3 was used to infer the population structure using 66 unlinked or weakly linked SSR markers as described in reference [79]. The length of the burn-in period and the number of Markov Chain Monte Carlo replications after burn-in were all assigned at 100,000 with an admixture and allele frequencies correlated model. Five independent run iterations were performed with the hypothetical number of subpopulations (k) ranging from 1 to 10. However, the result showed that the $\text{Ln}P(D)$ value corresponding to each hypothetical k kept increasing with k value and did not show any peak (Figure 1). Thus an ad hoc statistic Δk [80] was joined to correctly estimate the population structure. The Δk value showed a much higher likelihood at $k=2$ than at $k=3-10$ (Figure 1), suggesting that the total panel could be divided into 2 major subpopulations. Based on the correct k , all accessions were assigned to each of the two subpopulations (P1 and P2), for which the membership value (Q value) was >0.5 , and the population structure matrix (Q) was generated for further association mapping. The P1 group contained 115 accessions including 63 cultivars from Yellow River cotton growing region, 46 lines from North and Northwest China regions, and 6 cultivars from Yangtze River region. The P2 group consisted of 241 accessions including 116 lines from Yellow River region, 107 lines from Yangtze River region, 10 lines from the North and Northwest China regions, and 8 lines introduced from abroad (see reference [71] for details). The software SPAGeDi was used to calculate the pair-wise relatedness [81]. For the kinship coefficient values, 86.85% was less than 0.05, 8.56% had a range of 0.05–0.10, and the remaining 4.59% showed various degrees of genetic relatedness. Based on the results of the relatedness analysis, a K matrix was constructed for further association mapping.

Marker-trait association analysis were performed with the MLM model, considering both kinship (K) and population structure (Q), implemented in TASSEL software [82]. At the $\alpha=0.01$ ($-\log P=2$) level, a total of 195 significant associations were detected between 82 SSR markers and 7 lint yield traits. Among these, most of the associations (125 of 195) were detected in only one environment, and the proportion of phenotypic variation explained by markers ranged from 1.52% to 9.40%, with an average of 3.70% (see reference [71] for details). After Bonferroni correction [83], 55 associations were found to be significant ($P \leq 0.05/145$, $-\log P \geq 3.46$) between 26 SSR markers and 7 lint yield traits (Table 1). Most of the associations could be detected in

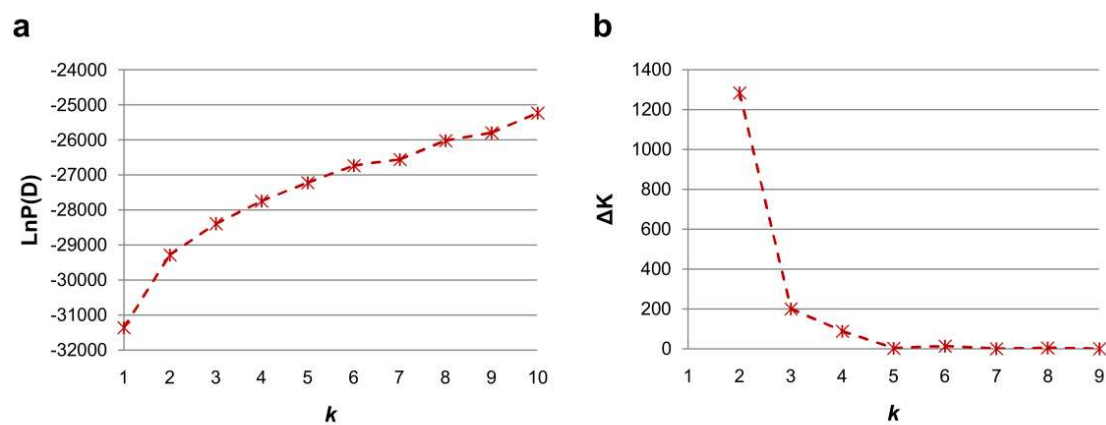


Figure 1. Estimated $\text{LnP}(D)$ and Δk over five repeats of STRUcTURE analysis (a) $\text{LnP}(D)$ for k from 1 to 10 for 356 accessions. Note that $\text{LnP}(D)$ value of each hypothetical k kept increasing with k value. (b) Δk for k from 2 to 9 for 356 accessions. The Δk value showed a much higher likelihood at $k=2$ than at $k=3-10$, suggesting that the total panel should be divided into two major subpopulations

more than one environment, and the proportion of phenotypic variation explained by markers ranged from 1.63% to 9.40%, with an average of 4.51%. The number of SSR markers associated with LY, SY, BN, BW, LP, LI and SI were 9, 4, 6, 4, 14, 17 and 1, respectively. Seventeen loci were co-associated with two or more different traits (Table 1), for example, NAU3269 (Chr. 5) and NAU3100 (Chr. 23) were simultaneously associated with FY, SY, BN, LP, and LI, and most of the lint yield-associated loci were associated with at least one of its components. These associations coincided with phenotypic correlations among these traits [71]. This might result from pleiotropy of a single causal gene or tight linkage of multiple causal genes. We found that 10 of 14 markers associated with LP were detected in all three environments, which was consistent with the results in phenotypic evaluation that LP possessed the highest broad-sense heritability ($h_B^2=75.77\%$) [71]. The phenotype of complex traits often result from the combined actions of multiple genes and environmental factors [3]; only those traits with high heritability can be stably detected. The resulting stably associated markers should be useful for cotton breeding with broad adaptability to different environments.

Traits	Marker loci	Chr.	Position	-LogP			$R^2(\%)$		
				E1	E2	E3	E1	E2	E3
LY	NAU3269	A05(Chr.05)	182.215	3.43	4.42	3.33	2.83	4.60	3.05
	NAU5166	A10(Chr.10)	23.423	ns	5.49	ns		5.40	
	NAU2935	A10(Chr.10)	51.592	ns	4.00	ns		4.72	
	JESPR204	A13(Chr.13)	59.714	4.12	3.00	ns	4.60	4.08	
	CIR246	D02(Chr.14)	112.473	ns	ns	3.55			4.44
	BNL3594	D06(Chr.25)	7.66	ns	4.51	ns		7.65	
	TMK19	D06(Chr.25)	70.38	4.30	2.77	3.97	4.21	3.21	4.28

Traits	Marker loci	Chr.	Position	-LogP			R ² (%)		
				E1	E2	E3	E1	E2	E3
SY	NAU3100	D09(Chr.23)	25.262	3.67	5.38	4.31	4.08	7.08	5.41
	NAU2776	D10(Chr.20)	9.452	ns	3.81	2.54		4.41	2.76
	NAU3269	A05(Chr.05)	182.215	2.25	4.16	2.43	1.85	4.41	2.19
	CIR246	D02(Chr.14)	112.473	ns	ns	4.23			5.36
	BNL3594	D06(Chr.25)	7.66	ns	4.61	ns		7.97	
	NAU3100	D09(Chr.23)	25.262	ns	4.03	2.82		5.61	3.84
BN	NAU6584	A03(Chr.03)	74.975	3.53	2.29	ns	3.93	2.91	
	NAU3269	A05(Chr.05)	182.215	2.08	3.52	3.45	1.74	3.64	3.44
	BNL3594	D06(Chr.25)	7.66	ns	3.62	ns		6.67	
	TMK19	D06(Chr.25)	70.38	3.58	ns	2.41	3.71		2.84
	NAU493	D07(Chr.16)	113.413	3.91	ns	ns	3.38		
	NAU3100	D09(Chr.23)	25.262	ns	3.48	ns		4.92	
BW	BNL1414	A09(Chr.09)	95.911	ns	3.26	3.72		4.18	4.45
	NAU4047	A12(Chr.12)	11.118	3.29	4.17	ns	3.32	4.61	
	NAU3398	A13(Chr.13)	3.311	3.47	3.46	ns	5.65	6.03	
	JESPR208	D09(Chr.23)	118.225	ns	4.01	3.69		5.15	4.45
	NAU3269	A05(Chr.05)	182.215	3.54	2.21	2.72	3.18	2.02	2.59
	NAU5166	A10(Chr.10)	23.423	2.33	4.36	2.74	1.80	4.10	2.44
LP	NAU2508	A10(Chr.10)	128.028	4.45	3.62	4.68	5.23	4.81	6.09
	NAU980	A11(Chr.11)	0	3.40	ns	3.75	5.65		6.72
	JESPR135	A11(Chr.11)	55.787	3.79	6.64	3.80	3.16	6.45	3.54
	NAU3398	A13(Chr.13)	3.311	3.50	ns	ns	4.99		
	JESPR204	A13(Chr.13)	59.714	3.73	2.96	2.22	4.41	3.96	3.11
	BNL3590	D03(Chr.17)	39.284	4.70	ns	2.89	4.91		3.38
LI	TMK19	D06(Chr.25)	70.38	5.97	4.58	3.67	6.04	5.18	4.19
	NAU3100	D09(Chr.23)	25.262	4.50	3.35	2.89	5.35	4.58	4.00
	NAU3917	D10(Chr.20)	31.125	ns	3.57	ns		4.03	
	BNL1404	D11(Chr.21)	33.571	4.48	6.20	3.51	3.81	6.01	3.23
	Gh508	D11(Chr.21)	54.48	2.15	3.37	3.81	1.63	3.06	3.54
	NAU2361	D11(Chr.21)	101.215	4.05	4.30	4.03	6.37	7.34	7.03
LI	NAU3269	A05(Chr.05)	182.215	4.98	2.77	2.38	4.92	2.63	2.32
	NAU980	A11(Chr.11)	0	8.85	2.34	2.80	8.96	4.76	5.72

Traits	Marker loci	Chr.	Position	-LogP			R ² (%)		
				E1	E2	E3	E1	E2	E3
	JESPR135	A11(Chr.11)	55.787	ns	5.43	2.34		5.20	2.09
	Gh369	A11(Chr.11)	84.701	3.77	2.09	ns	4.79	3.00	
	NAU1151	A12(Chr.12)	97.965	6.08	ns	ns	5.63		
	NAU3398	A13(Chr.13)	3.311	6.01	2.49	ns	8.19	4.15	
	JESPR204	A13(Chr.13)	59.714	ns	3.68	ns		4.81	
	CIR246	D02(Chr.14)	112.473	4.75	3.14	ns	5.77	4.17	
	BNL3590	D03(Chr.17)	39.284	3.30	2.70	3.47	3.67	3.13	4.26
	NAU2233	D05(Chr.19)	171.278	4.86	2.15	ns	5.96	3.04	
	TMK19	D06(Chr.25)	70.38	4.83	4.02	ns	5.19	4.55	
	NAU3100	D09(Chr.23)	25.262	5.90	2.74	3.47	7.19	3.84	4.93
	NAU2776	D10(Chr.20)	9.452	ns	ns	3.57			4.30
	NAU3917	D10(Chr.20)	31.125	ns	3.66	2.03		4.13	2.45
	BNL1404	D11(Chr.21)	33.571	ns	5.85	2.72		5.64	2.50
	Gh508	D11(Chr.21)	54.48	ns	3.67	3.41		3.38	3.27
	NAU2361	D11(Chr.21)	101.215	6.27	5.16	4.58	9.40	8.45	8.12
SI	NAU493	D07(Chr.16)	113.413	3.70	2.85	5.24	3.85	2.90	5.75

Table 1. SSR marker loci significantly associated with lint yield traits and their explained proportion of phenotypic variation in 3 different environments. LY: lint yield (g/plant); SY: seed cotton yield (g/plant); BN: bolls per plant; BW: boll weight (g); LP: lint percentage (%); LI: lint index (g/100 seeds); SI: seed index (g/100 seeds); E1: Jiangpu in 2009; E2: Dafeng in 2010; E3: Zhengzhou in 2010

To identify favorable alleles, the phenotypic allele effect was estimated through comparison between the average phenotypic value over accessions with the specific allele and that of all accessions:

$$a_i = \sum x_{ij} / n_i - \sum N_k / n_k$$

where a_i is the phenotypic effect of the i th allele; x_{ij} is the phenotypic value over the j th accession with the i th allele; n_i is the number of accessions with the i th allele; N_k is the phenotypic value over all accessions; n_k is the number of accessions. If the value of $a_i > 0$, the allele is considered to have a positive effect, if it is < 0 , it corresponds to a negative allele. The favorable alleles were then identified according to the breeding objective of each target trait [69, 71]. Phenotypic effects of each QTL allele for the 41 associated loci detected in more than one environment were measured, and 5, 2, 3, 4, 12, 14 and 1 favorable alleles for FY, SY, BN, BW, LP, LI and SI were identified, respectively. Phenotypic effects and representative accessions for each favorable allele are shown in Table 2. Among the favorable alleles, NAU3100-2 had the most positive phenotypic effect for FY and SY, and increased FY and SY by 3.61 g and 7.27 g, respectively; NAU6584-2, NAU3398-2, NAU5166-2 and NAU3917-2 increased BN, BW, LP and

LI by 0.89, 0.42 g, 4.93% and 0.94 g, respectively; while NAU493-1 decreased SI by 0.17 g. Lint yield of cotton is the result of series components and their interactions, such as boll number, boll weight, lint percentage, lint index, and seed index. Developing potentially high-yielding cultivars thus relies to some extent on selecting the appropriate yield components. As some of the QTLs were associated with more than one yield component, favorable alleles must be treated with caution. Positively co-associated genetic loci could simultaneously improve multiple target traits, while negative linkages must be broken.

Traits	Favorable allele	a_i	Accessions	Representative accessions
LY	NAU3269-2	0.27	133	Simian3, Zhongmiansuo9, Huakangmian1
	JESPR204-1	0.70	314	Simian3, Zhongmiansuo9, P164-2
	TMK19-2	1.02	240	Simian3, Zhongmiansuo9, P164-2
	NAU3100-2	3.61	87	Simian3, Zhongmiansuo9, Lumianyan16
	NAU2776-1	0.85	151	Zhongmiansuo9, P164-2, Lumianyan16
SY	NAU3269-2	0.42	133	Zhongmiansuo9, Zhongmiansuo19, Simian3
	NAU3100-2	7.27	87	Zhongmiansuo9, Han4849, Lumianyan16
BN	NAU6584-2	0.89	217	Lumianyan16, Zhongmiansuo44, Zhongmiansuo9
	NAU3269-2	0.08	133	Zhongmiansuo9, Wanmian73-10, Simian3
	TMK19-2	0.45	235	Zhongmiansuo44, Zhongmiansuo9, Wanmian17
BW	BNL1414-2	0.18	93	Zhongmiansuo18, Zhongmiansuo5, I40005
	NAU4047-2	0.03	221	Zhongmiansuo18, Zhongmiansuo5, I40005
	NAU3398-2	0.42	22	Zhongmiansuo5, I40005, Hua101
	JESPR208-2	0.20	86	Zhongmiansuo18, Zhongmiansuo5, I40005
LP	NAU3269-2	0.23	133	Simian3, Ekangmian9, Huakangmian1
	NAU5166-2	4.93	8	Simian3, Huakangmian1, Sumian4
	NAU2508-2	0.36	113	Nannongzao, 86-1, Yu668
	NAU980-3	2.79	9	Ekangmian6, Emian16, Ekangmian10
	JESPR135-1	0.13	343	XiangSC-24, Simian3, Ekangmian9
	JESPR204-1	0.36	309	XiangSC-24, Simian3, Ekangmian9
	BNL3590-1	0.26	327	XiangSC-24, Simian3, Ekangmian9
	TMK19-2	0.58	235	XiangSC-24, Simian3, Huakangmian1
	NAU3100-2	1.55	86	Simian3, Ekangmian9, Nannongzao
	BNL1404-1	0.13	343	XiangSC-24, Simian3, Ekangmian9
LI	Gh508-1	0.07	347	XiangSC-24, Simian3, Ekangmian9
	NAU2361-3	0.75	73	Ekangmian9, Yu668, Yumian21
	NAU3269-2	0.01	133	Huakangmian1, I40005, Ekangmian9

Traits	Favorable allele	a_i	Accessions	Representative accessions
	NAU980-3	0.84	9	I40005, Zhongmiansuo5, Hua101
	JESPR135-1	0.02	343	Huakangmian1, Emian23, I40005
	Gh369-3	0.11	8	Emian16, Ekangmian8, Yumian20
	NAU3398-2	0.80	22	Huakangmian1, I40005, Zhongmiansuo5
	CIR246-3	0.63	16	Hua101, Zhongmiansuo41, Yumian9
	BNL3590-1	0.06	327	Huakangmian1, Emian23, I40005
	NAU2233-1	0.01	212	Huakangmian1, Emian23, I40005
	TMK19-2	0.10	235	Huakangmian1, Emian23, I40005
	NAU3100-2	0.38	86	I40005, Zhongmiansuo5, Hua101
	NAU3917-2	0.94	6	Huakangmian1, Simian4, Sumian9
	BNL1404-1	0.03	343	Huakangmian1, Emian23, I40005
	Gh508-1	0.01	347	Huakangmian1, Emian23, I40005
	NAU2361-3	0.28	73	Emian23, I40005, Zhongmiansuo5
SI	NAU493-1	-0.17	230	Chaoyangmian1, Xuzhou1818, XiangSC-24

Table 2. Favorable QTL alleles, their phenotypic effects (a_i) and representative accessions. LY: lint yield (g/plant); SY: seed cotton yield (g/plant); BN: bolls per plant; BW: boll weight (g); LP: lint percentage (%); LI: lint index (g/100 seeds); SI: seed index (g/100 seeds); Representative accessions are the top-3 entries for the target trait value of accessions with the corresponding favorable allele

Allele frequencies of the 23 favorable alleles in the CK group and the six Chinese historically released cultivar groups are summarized in Table 3. Based on allele frequencies across the different groups, these favorable alleles could be categorized into three classes. The alleles in the first class, such as JESPR135-1, BNL1404-1 and Gh508-1, presented in the founder cultivars and with high frequency in all populations, might have been passed down stably from the original parents and were almost fixed in modern cultivars by selection. Alleles in the second class, such as BNL3269-2, BNL1414-2, NAU3100-2 and JESPR208-2, presented in the founder cultivars and with moderate to low frequency in most populations, should have been underutilized in modern breeding programs. Those in the third class, such as NAU5166-2, NAU980-3, Gh369-3 and CIR246-3, not presented in the founder cultivars and presented at low frequency in modern cultivars, might be from other original parents or could have been generated by mutations and/or recombinations. Favorable alleles, especially of the latter two classes, should have a great potential in future Upland cotton genetic improvement. We suggest that a multi-parent population should be constructed using cultivars that possess most of the favorable alleles, and in the meantime, a ranking system for MAS or genomic selection should be developed based on the results of AM. Favorable alleles that were passed down from the founder parents and have been almost fixed in modern cultivars formed the basis of lint yield of Chinese Upland cotton, and should be treated as fundamental elements in order to reject deleterious alleles at the corresponding loci. Alleles either absent in the founder cultivars or

present at moderate to low frequencies in most cultivar groups have been underutilized in modern breeding programs, and should be regarded as essential elements for increasing lint yield potential.

Favorable alleles	CK	I	II	III	IV	V	VI	Total
NAU6584-2	0.3333	0.5000	0.6923	0.7692	0.6747	0.4720	0.7755	0.6096
NAU3269-2	0.6667	0.5769	0.5385	0.6154	0.3976	0.3040	0.1020	0.3736
BNL1414-2	0.5000	0.3077	0.1923	0.4872	0.2410	0.2320	0.1633	0.2612
NAU5166-2	0.0000	0.0000	0.0000	0.0500	0.0000	0.0320	0.0417	0.0225
NAU2508-2	0.4000	0.1200	0.2400	0.2250	0.3012	0.3952	0.4694	0.3333
NAU980-3	0.0000	0.0200	0.0000	0.0256	0.0244	0.0369	0.0000	0.0233
JESPR135-1	1.0000	0.8462	1.0000	1.0000	0.9759	0.9840	1.0000	0.9775
Gh369-3	0.0000	0.0000	0.0000	0.0000	0.0241	0.0480	0.0000	0.0225
NAU4047-2	0.6667	0.5000	0.5769	0.6154	0.5663	0.6400	0.7551	0.6208
NAU3398-2	0.0000	0.0385	0.0000	0.1026	0.0843	0.0720	0.0204	0.0618
JESPR204-1	1.0000	0.8462	0.8077	0.7692	0.8675	0.9200	0.9388	0.8820
CIR246-3	0.0000	0.0000	0.0000	0.0256	0.0241	0.0800	0.0612	0.0449
BNL3590-1	0.5000	0.8077	0.8462	0.8462	0.9036	0.9440	0.8776	0.8904
NAU2233-1	0.6667	0.4231	0.5000	0.3590	0.5542	0.6560	0.8163	0.5955
TMK19-2	0.6667	0.5000	0.5769	0.6667	0.6024	0.7280	0.8163	0.6742
NAU493-1	0.6667	0.5000	0.7308	0.6923	0.6265	0.6800	0.5714	0.6461
NAU3100-2	0.3333	0.2308	0.0769	0.2308	0.2289	0.2400	0.3673	0.2444
JESPR208-2	0.5000	0.3077	0.1154	0.4359	0.2289	0.2240	0.1633	0.2444
NAU2776-1	0.1667	0.4231	0.3462	0.5385	0.4458	0.4480	0.3061	0.4242
NAU3917-2	0.0000	0.0000	0.0000	0.0513	0.0000	0.0240	0.0204	0.0169
BNL1404-1	1.0000	0.8462	1.0000	1.0000	0.9759	0.9840	1.0000	0.9775
Gh508-1	1.0000	0.9231	1.0000	0.9750	1.0000	0.9920	1.0000	0.9888
NAU2361-3	0.2143	0.0769	0.0417	0.1538	0.1125	0.2810	0.3673	0.2066

Table 3. Allele frequency for each favorable QTL allele in historically released Chinese Upland cotton cultivar groups. CK, I, II, III, IV, V and VI indicates the founder parent group (CK), and the Chinese cultivars released in 1930–1960, 1961–1970, 1971–1980, 1981–1990, 1991–2000 and 2000–2005, respectively

3. Favorable QTL alleles for fiber quality traits

With the acceleration of spinning speed, the demand for cotton fiber quality is increasing rapidly [9, 17]. It is important to elucidate the molecular genetics of Upland cotton fiber

qualities, and such information would enable the subsequent improvement of cotton cultivars by pyramiding favorable alleles of fiber quality traits. We performed a marker-trait association mapping for fiber quality traits using MLM model implemented in TASSEL [82] with the panel mentioned above [71]. At the $\alpha=0.01$ ($-\log P=2$) level, a total of 59 significant associations were detected between 41 SSR markers and 5 fiber quality traits (Table 4). Among these, almost all of the associations were detected in only one environment, and the proportion of phenotypic variation explained by markers ranged from 1.84% to 7.12%, with an average of 3.53% (Mei et al. unpublished data). If a more stringent threshold with Bonferroni correction [83] was adopted, only 9 associations were found to be significant (Table 4). The result might be caused by low diversity of fiber quality properties existed in the 356-accession panel. Although the yield potential of Chinese cultivars are equal to or a bit higher than those developed in the United States or Australia, the fiber quality traits of Chinese cultivars are not as good as those of American or Australian cultivars [73, 84]. Narrow variation in fiber qualities of Chinese cultivars had severely limited the marker-trait association detection power in association mapping.

Traits	Marker loci	Chr.	Position	-LogP			R ² (%)		
				E1	E2	E3	E1	E2	E3
FL	NAU845	A07(Chr.07)	54.589	4.16	ns	ns	5.91		
	NAU3398	A13(Chr.13)	3.311	ns	3.15	ns		5.64	
	BNL1707	A13(Chr.13)	104.694	2.39	ns	ns	2.27		
	JESPR152	D01(Chr.15)	110.252	2.59	ns	ns	3.25		
	BNL3359	D06(Chr.25)	140.397	4.3	ns	ns	5.67		
	JESPR297	D07(Chr.16)	43.109	ns	2.18	ns		2.08	
	NAU5418	D11(Chr.21)	75.156	ns	2.08	ns		4.47	
	BNL3537	D12(Chr.26)	39.496	ns	ns	2.06			2.77
	NAU2980	D13(Chr.18)	0	ns	ns	2.02			2.64
FS	NAU6584	A03(Chr.03)	74.975	2.3	ns	ns	3.16		
	NAU2561	A05(Chr.05)	56.73	2.96	ns	ns	3.08		
	NAU3269	A05(Chr.05)	182.215	3.53	ns	ns	3.88		
	NAU3427	A06(Chr.06)	5.598	2.25	ns	ns	2.91		
	NAU3654	A07(Chr.07)	74.363	ns	3.02	ns		4.36	
	NAU5166	A10(Chr.10)	23.423	2.09	ns	ns	1.95		

Traits	Marker loci	Chr.	Position	-LogP			R ² (%)		
				E1	E2	E3	E1	E2	E3
FF	BNL1707	A13(Chr.13)	104.694	5.59	ns	ns	6.04		
	BNL2646	D01(Chr.15)	75.614	ns	2.15	ns	2.56	3.44	
	NAU2901	D01(Chr.15)	104.568	2.21	ns	ns	2.09		
	NAU1070	D02(Chr.14)	13.885	3.3	ns	ns	4.3		
	BNL1606	D03(Chr.17)	50.883	ns	2.27	ns		2.92	
	NAU2233	D05(Chr.19)	171.278	2.15	ns	ns	3.42		
	BNL3594	D06(Chr.25)	7.66	ns	2.41	ns		5.4	
	BNL3359	D06(Chr.25)	140.397	3.16	ns	ns	4.23		
	NAU6468	D07(Chr.16)	83.101	2.44	ns	ns	2.36		
	JESPR304	A02(Chr.02)	22.517	ns	ns	2.16			3.41
	NAU862	A03(Chr.03)	9.309	2.24	ns	ns	2.21		
	BNL3452	A05(Chr.05)	188.717	2	ns	ns	3.14		
	NAU3427	A06(Chr.06)	5.598	3.82	ns	ns	4.72		
	NAU2935	A10(Chr.10)	51.592	2.55	ns	ns	3.2		
	JESPR135	A11(Chr.11)	55.787	2.34	ns	ns	2.15		
	JESPR204	A13(Chr.13)	59.714	3.81	ns	ns	5.36		
	BNL2646	D01(Chr.15)	75.614	2.54	ns	ns	3.8		
	BNL1604	D07(Chr.16)	36.152	ns	2.85	ns		5.43	
	JESPR297	D07(Chr.16)	43.109	ns	2.03	ns		1.89	
	BNL1694	D07(Chr.16)	52.03	2.11	ns	ns	3.2		
FU	BNL1404	D11(Chr.21)	33.571	2.72	ns	ns	2.58		
	Gh508	D11(Chr.21)	54.48	3.71	ns	ns	3.68		
	NAU3254	A01(Chr.01)	33.644	2.02	ns	ns	2.61		
	NAU2161	A03(Chr.03)	0	ns	ns	2.44			3.83
	NAU3654	A07(Chr.07)	74.363	ns	2.26	ns		3.29	0

Traits	Marker loci	Chr.	Position	-LogP			R ² (%)		
				E1	E2	E3	E1	E2	E3
FSCI	Gh486	A09(Chr.09)	10.554	ns	ns	2.24			4.23
	NAU5428	A11(Chr.11)	155.464	3.91	ns	ns	7.12		
	BNL1231	A11(Chr.11)	169.199	2.77	ns	ns	2.73		
	BNL3261	A12(Chr.12)	40.276	ns	2.08	ns		4.45	
	BNL1606	D03(Chr.17)	50.883	ns	2.48	ns		3.2	
	JESPR297	D07(Chr.16)	43.109	ns	ns	2.57			2.53
	Gh501	D13(Chr.18)	55.386	2.17	ns	ns	2.8		
	NAU2561	A05(Chr.05)	56.73	2.33	ns	ns	2.31		
	NAU5428	A11(Chr.11)	155.464	2.72	ns	ns	5.34		
	BNL1707	A13(Chr.13)	104.694	3.58	ns	ns	3.68		
	NAU2901	D01(Chr.15)	104.568	2.46	2.27	ns	2.36	2.17	
	JESPR152	D01(Chr.15)	110.252	3.45	ns	ns	4.34		
	NAU1070	D02(Chr.14)	13.885	3.19	ns	ns	4.15		1.99
	BNL1606	D03(Chr.17)	50.883	ns	2.52	ns		3.23	
	BNL3594	D06(Chr.25)	7.66	ns	2.29	ns	3.64	5.2	
	BNL3359	D06(Chr.25)	140.397	3.19	ns	ns	4.25		
	NAU6468	D07(Chr.16)	83.101	2	ns	ns	1.84		
	JESPR208	D09(Chr.23)	118.225	ns	2	ns		2.61	
	NAU2776	D10(Chr.20)	9.452	ns	2.11	ns		2.74	

Table 4. SSR loci significantly (-logP≥2.0) associated with fiber quality traits detected in the 356 Chinese Upland cotton cultivars. FL: fiber length; FS: fiber strength; FF: fiber fineness measured by micronaire value; FU: fiber uniformity ratio; FSCI: fiber spinning consistency index; E1: Jiangpu in 2009; E2: Dafeng in 2010; E3: Zhengzhou in 2010

To embrace the genetic diversity of fiber qualities as large as possible, a previous collection comprising 99 Upland cotton cultivars and breeding lines was used for further association mapping. The collection included 63 cultivars and 36 breeding lines with elite fiber quality traits (13 of which were PD lines introduced from the United States), and the fiber properties including fiber length, strength, and fineness were evaluated in 2004 and 2007. A total of 260

SSR markers, including the markers linked to fiber properties reported in previous studies [17-19], were used to screen the 99 accessions, and 97 of which showed polymorphisms [73]. Marker-trait association analysis was performed with the MLM model implemented in TASSEL [82]. At the $\alpha=0.01$ ($-\log P=2$) level, a total of 51 significant associations were detected between 33 SSR markers and 3 lint yield traits, and 7 of these associations could be detected in both 2 years. The proportion of phenotypic variation explained by markers ranged from 7.76% to 23.99%, with an average of 14.07%. After Bonferroni correction [83], 17 associations were still significant ($P \leq 0.05/97$, $-\log P \geq 3.29$). Compared with the results from our previous 356-accession panel, both the number of significant associations and the proportion of phenotypic variation explained by markers were improved in the current panel.

Traits	Marker loci	Chr.	Position	-logP		R ²	
				2004	2007	2004	2007
Fiber length	NAU934	A05(Chr.05)	186.594	2.32	ns	7.83	
	NAU1200	A05(Chr.05)		3.42	ns	17.4	
	NAU1037	A08(Chr.08)	53.526	ns	3.50		17.83
	BNL1317	A09(Chr.09)	88.456	2.62	2.68	12.18	14.12
	NAU2354	A09(Chr.09)	132.251	ns	2.06	10.4	13.32
	JESPR295	A12(Chr.12)	32.732	2.52	2.92	11.56	15.17
	NAU445	A12(Chr.12)	123.764	2.21	ns	10.22	
	JESPR153	A13(Chr.13)	63.046	3.00	ns	13.52	
	NAU2272	D02(Chr.14)	55.105	ns	2.64		16.26
	NAU1102	D05(Chr.19)		2.01	ns	9.32	
	NAU1302	D08(Chr.24)	72.101	ns	3.03		18.24
	BNL1521	D08(Chr.24)	73.477	ns	3.88		16.21
	JESPR78	D08(Chr.24)	75.81	ns	3.88		16.21
	NAU1336	D08(Chr.24)	78.13	ns	3.88		16.21
	NAU1262	D08(Chr.24)	88.599	ns	3.88		16.21
	JESPR127	D08(Chr.24)	97.939	ns	3.88		16.21
	NAU1004	D09(Chr.23)	13.67	2.64	ns	12.23	
	TMO06	D09(Chr.23)	109.242	3.78	3.88	16.86	19.56
	NAU904	D10(Chr.20)	53.078	ns	2.14		13.88
	NAU2443	D13(Chr.18)	62.214	2.30	ns	7.99	

Traits	Marker loci	Chr.	Position	-logP		R ²	
				2004	2007	2004	2007
Fiber strength	NAU934	A05(Chr.05)	186.594	3.82	ns	14.08	
	NAU1200	A05(Chr.05)		2.68	ns	14.5	
	NAU474	A07(Chr.07)	30.627	ns	2.43		17.44
	NAU1043	A07(Chr.07)		ns	2.14		15.86
	BNL1317	A09(Chr.09)	88.456	ns	2.37		12.58
	NAU2508	A10(Chr.10)	126.827	ns	3.29		23.99
	JESPR295	A12(Chr.12)	32.732	2.28	2.72	10.56	14.19
	NAU445	A12(Chr.12)	123.764	2.60	ns	11.96	
	JESPR153	A13(Chr.13)	63.046	5.42	ns	23.3	
	BNL1395	D07(Chr.16)	78.652	ns	2.12		13.62
	BNL1122	D07(Chr.16)	88.995	ns	2.23		14.19
	BNL2634	D07(Chr.16)		2.72	ns	12.47	
	NAU1369	D08(Chr.24)	57.434	2.74	ns	12.13	
	BNL1521	D08(Chr.24)	73.477	ns	2.34		9.26
	JESPR78	D08(Chr.24)	75.81	ns	2.34		9.26
	NAU1197	D08(Chr.24)	77.384	ns	2.01		7.76
	NAU1336	D08(Chr.24)	78.13	ns	2.34		9.26
	NAU1262	D08(Chr.24)	88.599	ns	2.34		9.26
	NAU1322	D08(Chr.24)	89.779	ns	2.04		10.82
	JESPR127	D08(Chr.24)	97.939	ns	2.34		9.26
	NAU780	D08(Chr.24)		ns	3.60		18.14
	NAU816	D08(Chr.24)		3.70	ns	13.6	
	TMO06	D09(Chr.23)	109.242	5.51	3.80	23.66	19.23
	NAU2443	D13(Chr.18)	62.214	6.13	3.16	22.85	13.02
Fiber fineness	NAU934	A05(Chr.05)	186.594	ns	3.074		12.62
	BNL1317	A09(Chr.09)	88.456	4.31	2.31	18.74	9.71
	NAU1162	A11(Chr.11)	99.954	2.05	ns	11.43	
	JESPR295	A12(Chr.12)	32.732	2.87	ns	12.95	

Traits	Marker loci	Chr.	Position	-logP		R ²	
				2004	2007	2004	2007
	BNL3436	D06(Chr.25)	105.073	2.24	ns	9.68	
	TMO06	D09(Chr.23)	109.242	2.85	ns	12.84	
	NAU1004	D09(Chr.23)	13.67	3.03	ns	13.67	

Table 5. SSR loci significantly ($-\log P \geq 2.0$) associated with fiber quality traits detected in the and their explained phenotypic variation in two years

Phenotypic effects of each QTL allele for the 33 significantly associated loci were measured, and 21, 24 and 7 favorable alleles for fiber length, strength and fineness were identified, respectively. Phenotypic effects and representative materials for each favorable allele are shown in Table 6. A wide variation of the favorable alleles processed in the 99 accessions was observed, with an average of 14.84 and a range of 7 to 21 (see reference [73] for details). Most of the representative materials were fiber-elite lines collected from China or introduced from the United States, and some of them, such as Yumian 1 and 7235, were introgression lines and contained more than half of the favorable alleles. For example, 7235 was developed from several hybridizations with multiple parents, such as a *G. anomalum* introgression line, Acala 3080 and PD4381 [85, 86]; Yumian 1 was an introgression line with *G. barbadense*, *G. arboreum*, and *G. raimondii* as putative donors, which is characterized by high lint yield and high fiber strength [24]. These fiber-elite materials had been used for tagging QTLs underlying fiber quality traits in previous family-based linkage mapping studies [13, 17-19, 24-25; 39]. With the genetic information provided by this and previous studies, MAS should be complemented in the future breeding programs.

Traits	Favorable allele	a_i	Accessions	Representative materials
Fiber length	NAU934-4	1.0498	17	I-62434, 7235, I-62478
	NAU1037-1	4.5458	1	7235
	NAU2354-2	0.3662	10	I-62478, I-62479, PD94045
	BNL1317-3	0.8522	21	I-62434, 7235, I-62431
	NAU445-3	0.7926	31	I-62434, 7235, I-62431
	JESPR295-2	0.8233	31	I-62434, 7235, I-62478
	JESPR153-3	0.8457	16	I-62434, I-62431, Yumian1
	NAU1200-1	0.5373	49	I-62434, I-62478, I-62431
	NAU2272-5	5.8050	1	7235
	NAU1102-3	0.4564	22	I-62431, I-62479, I-62429

Traits	Favorable allele	a_i	Accessions	Representative materials
Fiber strength	NAU1302-1	4.5458	1	7235
	BNL1521-3	5.8050	1	7235
	JESPR78-3	5.8050	1	7235
	NAU1336-3	5.8050	1	7235
	NAU1262-2	5.8050	1	7235
	JESPR127-2	4.5458	1	7235
	TMO06-3	1.3109	19	I-62434, 7235, Yumian1
	NAU904-3	2.3858	1	GA2001163
	NAU2443-2	0.8435	17	I-62434, 7235, I-62431
	NAU780-4	4.5458	1	7235
	NAU1004-2	0.9346	14	I-62434, 7235, TM-1
	NAU934-4	2.6759	17	I-62429, I-62478, PD93019
	NAU474-2	1.0231	31	PD94042, I-62478, PD93019
	BNL1317-3	2.3136	21	I-62429, 7235, I-62433
	NAU2508-1	3.7061	8	7235, PD6992, HS427
	NAU445-3	0.8072	31	I-62429, 7235, I-62433
	JESPR295-2	2.1342	31	I-62429, I-62478, 7235
	JESPR153-3	2.8625	16	I-62429, I-62433, I-62434
	NAU1200-1	0.3169	49	I-62429, PD94042, I-62478
	NAU1043-2	0.9147	29	I-62478, PD93019, Yuwu19
	BNL1395-1	2.2677	12	I-62478, 7235, Yumian1
	BNL1122-1	2.2677	12	I-62478, 7235, Yumian1
	NAU1369-2	1.4296	15	I-62429, I-62433, Yumian1
	BNL1521-3	8.9290	1	7235
	JESPR78-3	8.9290	1	7235
	NAU1197-3	5.6730	2	Xiangmian13, 7235
	NAU1336-3	8.9290	1	7235
	NAU1262-2	8.9290	1	7235
	NAU1322-2	1.7557	17	I-62429, 7235, I-62433

Traits	Favorable allele	a_i	Accessions	Representative materials
Fiber fineness	JESPR127-2	8.9290	1	7235
	TMO06-3	3.1629	19	I-62429, 7235, I-62433
	NAU2443-2	3.0749	17	I-62429, 7235, Yumian1
	BNL2634-3	0.8204	26	I-62429, I-62433, Yumian1
	NAU780-1	0.4073	71	I-62429, I-62478, Yumian1
	NAU816-3	1.6559	29	I-62429, PD94042, I-62478
	NAU934-4	-0.3064	17	Emian11, Emian6, CRI18
	BNL1317-3	-0.3617	21	Jinmian2, Jinmian9, Jimian11
	NAU1162-2	-0.2338	13	Sumian5, CRI16, Yu668
	JESPR295-3	-0.2148	9	Qiannong465, Cangzhou7315-38, Jin185
	BNL3436-3	-0.1132	60	Xiangmian11, Jinmian2, Qiannong465
	TMO06-3	-0.3587	19	Shiduan5, Tai8033-2, Sumian5
	NAU1004-2	-0.1530	14	Qiannong465, Wanmian73-10, Qinli514

Table 6. Favorable QTL alleles for fiber qualities, their phenotypic effects (a_i) and representative materials. Representative accessions are the top-3 entries for the target trait value of accessions with the corresponding favorable allele

4. Favorable QTL alleles for FOV race 7 resistance

Fusarium wilt (FW), caused by *Fusarium oxysporum* f. sp. *vasinfectum* (FOV), is a widespread disease that causes huge losses in cotton (*Gossypium* spp.) production worldwide [87-90]. Once established in soil, FOV can survive in the field for several years as Chlamydia spores, even in the absence of a host, and is nearly impossible to eliminate. Although both chemical controls and cultural practices have been employed in protecting plants from damage, the most effective and efficient control should be provided through host resistance [87, 89]. In the past several years, eight races (race 1–8) of FOV had been indentified worldwide that use both cotton and non-cotton differential hosts [88]. Three FOV races (race 3, 7 and 8) had been found in China; race 7 possesses the highest virulence and is the most widely distributed race [91]. Recently, DNA-based techniques were employed in conjunction with pathogenicity tests to validate these races and to test new isolates, and highly virulent isolates of FOV were identified in Australia [92] and the United States [93-94]. Host resistance to FOV races has been widely evaluated in cotton germplasm under both field nursery and greenhouse conditions, and many highly resistant cotton cultivars and breeding lines have been developed through conventional breeding [89, 95-96]. However, little is known about the mechanism and genetic basis of FOV resistance. Some early classical genetic studies have suggested that the inheritance of FOV

resistance in cotton is determined by a single gene [97-99], while other studies have suggested that FOV resistance is controlled by multiple genes [100-102]. Large-scale resistance evaluations in breeding programs are time-consuming and labor-intensive, and it is not easy to obtain the ideal genotype simply through phenotypic selection. The cultivar development process has been slow in the face of the emergence of new, highly virulent FOV isolates [89-90].

To accumulate useful information for understanding the genetic basis of FOV 7 resistance and identify favorable alleles for facilitating future molecular resistance breeding, we performed a marker-trait association mapping to detect QTLs underlying FOV 7 resistance in Upland cotton [72]. The 356-accession panel mentioned above [71] was used for association analysis and favorable allele mining. Meanwhile, a composite cross population (CP) with three parents was developed for linkage mapping. Three Upland cotton cultivars, Xuzhou 142, Yumian 21 and Shang 9901, were chosen as parents according to many years of FOV 7 resistance evaluation. Xuzhou 142, an obsolete cultivar selected from STV 2B in the 1970s with large boll size and high lint percentage that is severely infested by FOV 7, was selected as the susceptible parent. Yumian 21 was released in 1999 and is currently used as a resistant control in national cotton regional trials due to its extremely high resistance to FOV 7. The cultivar Shang 9901 is an anonymous breeding line with high yield potential and moderately high resistance to FOV 7. Thus serial QTLs for FOV 7 resistance were detected by joint population-based association mapping and family-based linkage mapping. Marker-trait association mapping was performed with the MLM model implemented in TASSEL software [82]. A total of 27 markers were significantly associated with FOV 7 resistance at the $\alpha=0.01$ level ($-\log P=2.0$), and were localized to 16 chromosomes (Table 7). Of these loci, 23 were detected under field nursery conditions, 10 were detected in the greenhouse and 6 were detected under both conditions. The proportion of phenotypic variation explained by the markers ranged from 1.48% to 12.99%, with an average of 4.21%. If a more stringent threshold by the Bonferroni correction ($P \leq 0.05/145$, $-\log P \geq 3.46$) is adopted [83], only 10 associations were significant (Table 7). Five (*qFW-A3-1*, *qFW-A12-1*, *qFW-D3-1*, *qFW-D5-1* and *qFW-D8-1*) of the 7 QTLs identified by linkage mapping could be detected by association mapping at the $\alpha=0.05$ level, while only 3 QTLs (*qFW-D3-1*, *qFW-D5-1* and *qFW-D8-1*) could be detected at the $\alpha=0.01$ level (see reference [72] for details).

The phenotypic effects of each QTL allele of the 27 associated loci were estimated in both the greenhouse and field nursery evaluations according to the method mentioned above [71], and therefore, favorable alleles for FOV 7 resistance were identified (Table 8). The phenotypic effects of the 27 loci in the greenhouse and field nursery averaged -1.31 and -3.45, with ranges of -3.81 to -0.01 and -14.88 to -0.20, respectively. Among the favorable alleles, NAU934-2 had the most negative phenotypic effect in the greenhouse assay and was able to decrease FW DI by 3.8, while NAU6966-3 had the most negative phenotypic effect in the field nursery and was able to decrease FW DI by 14.88. A wide variation of the favorable alleles processed in the 356 accessions was observed, with an average of 14.84 and a range of 7 to 21 (see [72] for details). Pearson correlation analysis between the number of favorable alleles and FW DI was carried out, and highly negative significance was found in both the greenhouse ($r=-0.344$, $P<0.001$) and

Marker loci	Chr.	Position(cM)	-Log ₁₀ P		R ² (%)	
			Gh	FN	Gh	FN
NAU2437	A01(Chr.01)	15.248	ns	2.19		5.21
NAU4073	A01(Chr.01)	85.025	2.03	ns	1.48	
JESPR304	A02(Chr.02)	22.517	ns	7.09		9.65
NAU934	A05(Chr.05)	19.421	ns	4.61		5.78
NAU3273	A05(Chr.05)	37.244	ns	2.85		2.81
NAU797	A05(Chr.05)	143.032	2.04	ns	3.22	
BNL3255	A08(Chr.08)	81.913	ns	3.20		4.41
STV031	A10(Chr.10)	12.751	2.09	2.82	3.34	4.34
NAU440	A10(Chr.10)	42.017	2.80	2.57	2.81	2.51
Gh369	A11(Chr.11)	84.701	ns	2.29		3.61
BNL1066	A11(Chr.11)	134.486	ns	2.12		3.94
BNL1707	A13(Chr.13)	104.694	2.29	ns	1.59	
NAU1070	D02(Chr.14)	13.885	2.47	5.89	3.10	7.56
CIR246	D02(Chr.14)	112.473	2.55	3.14	3.65	4.56
BNL3590	D03(Chr.17)	39.284	ns	2.77		3.66
BNL834	D03(Chr.17)	81.098	ns	5.07		5.46
NAU6966	D04(Chr.22)	36.51	ns	2.47		4.16
NAU3095	D05(Chr.19)	10.634	4.20	4.04	5.12	4.99
NAU2816	D05(Chr.19)	160.89	ns	2.62		4.20
BNL3594	D06(Chr.25)	7.66	7.87	4.12	12.99	7.92
NAU3911	D07(Chr.16)	28.658	ns	3.52		3.93
NAU6752	D07(Chr.16)	74.671	ns	3.35		3.44
NAU478	D08(Chr.24)	67.677	2.08	ns	3.86	
NAU3207	D08(Chr.24)	90.625	ns	5.21		5.70
NAU1350	D08(Chr.24)	108.529	ns	4.38		4.69
NAU5418	D11(Chr.21)	75.156	ns	3.71		6.98
NAU3084	D12(Chr.26)	0	ns	2.36		2.27

Table 7. SSR loci significantly ($-\log P \geq 2.0$) associated with FOV 7 resistance and their explained phenotypic variation in greenhouse and field nursery evaluations. Gh and FN indicate the results evaluated in greenhouse and field nursery, respectively; ns indicates non-significant at $\alpha=0.01$ level

the field nursery ($r=-0.488$, $P<0.001$) evaluations. The top-3 resistant accessions for each favorable allele were listed as representative materials in Table 8.

Favorable alleles	a_g	a_f	Accessions	Representative materials
NAU2437-4	-3.27	-8.49	26	Yumian21,Xingmian2,Shiyuan321
NAU4073-2	-1.46	-0.68	172	Xinluzao24,Xingmian2,Xinluzao28
JESPR304-1	-0.45	-2.69	316	Yumian21,Jimian14,Xinluzao24
NAU934-2	-3.81	-4.68	39	Xinluzao24,Zhongmiansuo3,Xinluzao28
NAU3273-1	-0.13	-0.79	341	Yumian21,Jimian14,Xinluzao24
NAU797-2	-1.51	-1.03	172	Yumian21,Jimian14,Xinluzao24
BNL3255-1	-0.01	-2.33	284	Yumian21,Jimian14,Xinluzao24
STV031-1	-2.09	-4.91	137	Yumian21,Xingmian2,Zhongmiansuo3
NAU440-1	-0.33	-0.60	347	Yumian21,Jimian14,Xinluzao24
Gh369-2	-1.02	-3.33	10	Xinluzao24,Xinluzao28,Emian13
BNL1066-4	-1.96	-4.14	15	Zhongmiansuo27,Jimian20,Emian13
BNL1707-2	-1.93	-4.79	112	Yumian21,Xinluzao24,Xingmian2
NAU1070-1	-1.73	-5.84	199	Jimian14,Xinluzao24,Xingmian2
CIR246-3	-2.77	-8.65	16	Ejing1,Ekangmian3,Zhongmiansuo49
BNL3590-1	-0.06	-1.04	315	Yumian21,Jimian14,Xinluzao24
BNL834-1	-0.24	-1.70	312	Yumian21,Jimian14,Xingmian2
NAU6966-3	-1.89	-14.88	37	Xinluzao24,Xinluzao28,GK44
NAU3095-1	-2.30	-4.55	192	Yumian21,Xinluzao24,Xingmian2
NAU2816-2	-1.70	-3.43	152	Xinluzao24,Jimian14,Zhongmiansuo3
BNL3594-1	-2.58	-5.23	201	Jimian14,Xingmian2,Zhongmiansuo9
NAU3911-1	-0.33	-1.91	282	Yumian21,Jimian14,Xingmian2
NAU6752-1	-0.14	-0.83	341	Yumian21,Jimian14,Xinluzao24
NAU478-1	-0.21	-0.20	239	Yumian21,Jimian14,Xingmian2
NAU3207-1	-0.17	-2.27	301	Yumian21,Xinluzao24,Xingmian2
NAU1350-1	-0.03	-0.62	348	Yumian21,Xinluzao24,Jimian14
NAU5418-1	-2.90	-2.44	47	Yumian21,Jimian14,Yumian20
NAU3084-1	-0.42	-1.13	326	Yumian21,Jimian14,Xinluzao24

Table 8. Favorable QTL alleles, phenotypic effects (a_g) and their representative materials. a_g and a_f indicate phenotypic effects evaluated in greenhouse and field nursery conditions, respectively; Representative materials are the top-3 accessions with low disease index

Wang and Roberts identified a major resistance gene (*Fov1*) against FOV 1 in *G. barbadense* cv. Pima-S7 and indicated that one or more minor genes in Acala NemX could delay wilt symptoms [54]. Ulloa et al. performed QTL mapping in an F_2 (Pima-S7 × Acala NemX) and a

recombinant inbred line (RIL; *G. hirsutum* TM-1 × *G. barbadense* Pima 3-79) population. The authors detected 6 QTLs (Fov1-C06, Fov1-C08, Fov1-C11₁, Fov1-C11₂, Fov1-C16 and Fov1-C19) conferring FOV 1 resistance in different genetic backgrounds [58]. Lopez-Lavalle et al. used an intraspecific cross between the Upland cotton MCU-5 and Siokra 1-4 to detected QTLs conferring resistance to Australian FOV races and found that MCU-5 resistance is complex, with 3 QTLs identified in F₃ and 8 ones in F₄. The QTLs were localized to chromosomes A6, D4 and D6 [60]. Very recently, Ulloa et al. investigated 3 intraspecific (*G. hirsutum* × *G. hirsutum* L. and *G. barbadense* × *G. barbadense* L.), 5 interspecific (*G. hirsutum* × *G. barbadense*) and one RIL population in 4 greenhouse and 2 field experiments and identified a set of 11 SSR markers across 6 linkage groups/chromosomes (3, 6, 8, 14, 17 and 25) associated with FOV 4 resistance [59]. Integrating information mentioned above with the QTL mapping results for FOV 7 in our study, it suggests that the inheritance of FOV resistance in cotton should be far more complex than has been elucidated to date. Interestingly, the map positions of some QTLs conferring resistance against different FOV races coincided, suggesting that these genes may play the same or similar roles in the process of different host-pathogen and/or environment interactions. While great differences in the resistance of cotton genotypes against FOV races have been demonstrated in many evaluations worldwide, a genotype with a high level of resistance against all FOV races has not been found. Given that FW is the result of interactions among the host, pathogen and environment, many genes must be involved in this process. The accumulation of resistance alleles, even with relatively minor individual effects, will result in higher levels of resistance. The Upland cotton cultivars from China and Africa have shown more resistance to Australian FOV races [90]. Therefore, the favorable alleles and their typical germplasm resources identified in this study should have great potential for developing highly resistant Upland cotton cultivars in future molecular breeding programs.

5. Discussion and future prospects

Compared to family-based linkage mapping, population-based association mapping offers several advantages such as: (1) the saving of time and money by using existing populations instead of creating cross-controlled populations, (2) analysis of more than two alleles per locus on average, and (3) high expected resolution owing to a short extent of linkage disequilibrium [103]. All these make association mapping an increasingly important tool for complex trait dissection [63, 104].

A suitable association mapping panel should embrace as much phenotypic and genotypic diversity as possible. For example, because the 99-accession panel [73] comprised higher diversity in fiber properties than the 356-accession panel [71] and 81-accession panel [69], both the number of significant associations detected and the proportion of phenotypic variation explained by markers were improved. However, the presence of high diversity from different genetic origins may induce LD between unlinked loci, and consequently result in spurious marker-trait associations [105]. Although several statistical strategies have been developed to account for issues related to population structure and relatedness in order to decrease false positive [106-107], whether these approaches will induce new false negative remain unclear.

Synthetic association populations, such as the multi-parent advanced generation inter-cross (MAGIC) population [108] and the nested association mapping (NAM) population in maize [109], had been used for genome-wide association studies (GWAS); with which both high resolution and better population structure control can be achieved [110-111]. It is certain that such populations had been or are being developed in Upland cotton. Two artificially controlled multiple parent random-mating populations with each comprising more than 800 lines had been developed in our library, which should provide another theoretically ideal panel for association mapping.

The other inherent constraint limiting the successful use of association mapping is rare alleles existing in natural populations. Given that the number of individuals with a specific genotype is quite small, the effect of rare alleles on mapping can go far beyond the effect of small population sizes [112]. Large population size had been considered as an important factor to improve the QTL detection power in association mapping studies [3, 112]. Many more marker-trait associations for fiber yield were detected in our 356-accession panel [71] than in the 81-accession panel [69] between the same markers and target traits at the same significance level. While family-based linkage mapping can make use of alleles that occur at low frequencies in natural populations by designing crosses to create artificial populations with inflated frequencies of those alleles. So specifically designed mapping populations such as recombinant inbred lines (RIL) and near isogenic lines (NIL) will remain important. Furthermore, joint linkage and association mapping was recommended as an alternative approach to overcome some of the inherent limitations of both linkage and association mapping [105, 112], and this approach has proven to be a powerful tool to detecting architecture of complex traits [72, 113-118].

Recently, two preliminary maps of the whole-genome scaffolds of *G. raimondii* (the putative diploid donor for tetraploid species) were separately released by two different groups [119-120], which will facilitate the tetraploid genome sequencing and assembly. Real genome-wide association mapping will be realized in the near future through resequencing or other high-throughput genotyping technologies [121], which will dramatically accelerate genetic diversity exploitation and favorable allele mining in Upland cotton germplasm resources.

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References

- [1] National Cotton Council, USA. <http://www.cotton.org> (accessed 10 October 2013).
- [2] Zhang HB, Li YN, Wang BH, Chee PW. Recent advances in cotton genomics. *Int J Plant Genomics* 2008; ID 742304. doi: 10.1155/2008/742304.
- [3] Mackay TFC, Stone EA, Ayroles JF. The genetics of quantitative traits: challenges and prospects. *Nat Rev Genet* 2009; 10: 565–577.
- [4] Xu Y, Crouch J. Marker-assisted selection in plant breeding: from publications to practice. *Crop Sci* 2008; 48: 391–407.
- [5] Jannink JL, Lorenz AJ, Iwata H. Genomic selection in plant breeding: from theory to practice. *Brief Funct Genomics* 2010; 9: 166–177.
- [6] Jiang CX, Writght RJ, EL-Zik KM, et al. Polyploid formation created unique avenues for response to selection in *Gossypium* (cotton). *Proc Natl Acad Sci USA* 1998; 95: 4419–4424.
- [7] Ulloa M, Meredith WR. Genetic linkage map and QTL analysis of agronomic and fiber quality traits in an intraspecific population. *J Cotton Sci* 2000; 4: 161–170.
- [8] Ulloa M, Saha S, Jenkin N, et al. Chromosomal assignment of RFLP linkage groups harboring important QTL on an intraspecific cotton (*Gossypium hirsutum* L.) joinmap. *J Hered* 2005; 96: 132–144.
- [9] Kohel RJ, Yu J, Park YH, et al. Molecular mapping and characterization of traits controlling fiber quality in cotton. *Euphytica* 2001; 121: 163–172.
- [10] Saranga Y, Menz M, Jiang CX, et al. Genomic dissection of genotype x environment interactions conferring adaptation of cotton to arid conditions. *Genome Res* 2001; 11: 1988–1995.
- [11] Ren LH, Guo WZ, Zhang TZ. Identification of quantitative trait loci (QTLs) affecting yield and fiber properties in chromosome 16 in cotton using substitution line. *Acta Botanica Sinica* 2002; 44: 815–820.

- [12] Paterson AH, Saranga Y, Menz M, et al. QTL analysis of genotype x environment interaction affecting cotton fiber quality. *Theor Appl Genet* 2003; 6: 384–396.
- [13] Zhang T, Yuan Y, Yu J, et al. Molecular tagging of a major QTL for fiber strength in Upland cotton and its marker-assisted selection. *Theor Appl Genet* 2003; 106: 262–268.
- [14] Wu MQ, Zhang XL, Nie YC, et al. Localization of QTLs for yield and fiber quality traits of tetraploid cotton cultivar. *Acta Genetica Sinica* 2003; 30: 443–452.
- [15] Mei M, Syed NH, Gao W, et al. Genetic mapping and QTL analysis of fiber related traits in cotton (*Gossypium*). *Theor Appl Genet* 2004; 108: 280–291.
- [16] Lacape JM, Nguyen TB, Courtois B, et al. QTL analysis of cotton fiber quality using multiple *Gossypium hirsutum* x *Gossypium barbadense* backcross generations. *Crop Sci* 2005; 45: 123–140.
- [17] Shen X, Guo W, Zhu X, et al. Molecular mapping of QTLs for fiber qualities in three diverse lines in Upland cotton using SSR markers. *Mol Breed* 2005; 15: 169–181.
- [18] Shen X, Zhang T, Guo W, et al. Mapping fiber and yield QTLs with main, epistatic, and QTL x Environment interaction effects in recombinant inbred lines of Upland cotton. *Crop Sci* 2006; 46: 61–66.
- [19] Shen X, Guo W, Lu Q, et al. Genetic mapping of quantitative trait loci for fiber quality and yield trait by RIL approach in Upland cotton. *Euphytica* 2007; 155: 371–380
- [20] Chee P, Draye X, Jiang CX, et al. Molecular dissection of interspecific variation between *Gossypium hirsutum* and *Gossypium barbadense* (cotton) by a backcross-self approach: I. Fiber elongation. *Theor Appl Genet* 2005; 111: 757–763.
- [21] Chee P, Draye X, Jiang CX, et al. Molecular dissection of phenotypic variation between *Gossypium hirsutum* and *Gossypium barbadense* (cotton) by a backcross-self approach: III. Fiber length. *Theor Appl Genet* 2005; 111: 772–781.
- [22] Park YH, Alabady MS, Sickler B, et al. Genetic mapping of new cotton fiber loci using EST-derived microsatellites in an interspecific recombinant inbred line (RIL) cotton population. *Mol Genet Genom* 2005; 274: 428–441.
- [23] Lin ZX, He DH, Zhang XL, et al. Linkage map construction and mapping QTL for cotton fibre quality using SRAP, SSR and RAPD. *Plant Breed* 2005; 124: 180–187.
- [24] Zhang ZS, Xiao Y, Luo M, et al. Construction of a genetic linkage map and QTL analysis of fiber-related traits in upland cotton (*Gossypium hirsutum* L.). *Euphytica* 2005; 144: 91–99.
- [25] Zhang ZS, Hu MC, Zhang J, et al. Construction of a comprehensive PCR-based marker linkage map and QTL mapping for fiber quality traits in upland cotton (*Gossypium hirsutum* L.). *Mol Breed* 2009; 24: 49–61.

- [26] Draye X, Chee P, Jiang CX, et al. Molecular dissection of interspecific variation between *Gossypium hirsutum* and *G. barbadense* (cotton) by a backcross-self approach: II. Fiber fineness. *Theor Appl Genet* 2005; 111: 764–771.
- [27] Saha S, Jenkins JN, Wu J, et al. Effects of chromosome specific introgression in upland cotton on fiber and agronomic traits. *Genetics* 2006; 172: 1927–1938.
- [28] Wang B, Guo W, Zhu X, et al. QTL mapping of fiber quality in an elite hybrid derived-RIL population of upland cotton. *Euphytica* 2006; 152: 367–378.
- [29] Frelichowski JE, Palmer M, Main D, et al. Genetic mapping of microsatellites derived from BAC-end sequences of *Gossypium hirsutum* Acala 'Maxxa' BAC-ends. *Mol Genet Genom* 2006; 275: 479–491.
- [30] He DH, Lin ZX, Zhang XL, et al. QTL mapping for economic traits based on a dense genetic map of cotton with PCR-based markers using the interspecific cross of *Gossypium hirsutum* x *Gossypium barbadense*. *Euphytica* 2007; 153: 181–197.
- [31] Qin H, Guo W, Zhang Y, et al. QTL mapping of yield and fiber traits based on a four-way cross population in *Gossypium hirsutum* L. *Theor Appl Genet* 2008; 117: 883–894.
- [32] Wu JX, Gutierrez OA, Jenkins JN, et al. Quantitative analysis and QTL mapping for agronomic and fiber traits in an RI population of upland cotton. *Euphytica* 2009; 165: 231–245.
- [33] Chen H, Qian N, Guo W, et al. Using three overlapped RILs to dissect genetically clustered QTL for fiber strength on Chro.D8 in Upland cotton. *Theor Appl Genet* 2009; 119: 605–12.
- [34] Luan M, Guo X, Zhang Y, et al. QTL mapping for agronomic and fiber traits using two interspecific chromosome substitution lines of Upland cotton. *Plant Breed* 2009; 128: 671–679.
- [35] Qin YS, Ye WX, Liu RZ, et al. QTL mapping for fiber quality properties in upland cotton (*Gossypium hirsutum* L.). *Sci Agric Sin* 2009; 42: 4145–4154.
- [36] An CF, Jenkins JN, Wu JX, et al. Use of fiber and fuzz mutants to detect QTL for yield components, seed, and fiber traits of upland cotton. *Euphytica* 2010; 172: 21–34.
- [37] Wang FR, Gong YC, Zhang CY, et al. Genetic effects of introgression genomic components from sea island cotton (*Gossypium barbadense* L.) on fiber related traits in upland cotton (*G. hirsutum* L.). *Euphytica* 2011; 181: 41–53.
- [38] Sun FD, Zhang JH, Wang SF, et al. QTL mapping for fiber quality traits across multiple generations and environments in upland cotton. *Mol Breed* 2012; 30: 569–582.
- [39] Zhang K, Zhang J, Ma J, et al. Genetic mapping and quantitative trait locus analysis of fiber quality traits using a three-parent composite population in upland cotton (*Gossypium hirsutum* L.). *Mol Breed* 2012; 29: 335–348

- [40] Ning Z, Chen H, Mei H, et al. Molecular tagging of QTLs for fiber quality and yield in the upland cotton cultivar Acala-Prema. *Euphytica* 2014; 195: 143–156.
- [41] He DH, Lin ZX, Zhang XL, et al. Mapping QTLs of traits contributing to yield and analysis of genetic effects in tetraploid cotton. *Euphytica* 2005; 144: 141–149.
- [42] Wang B, Guo W, Zhu X, et al. QTL mapping of yield and yield components for elite hybrid derived-RILs in upland cotton. *J Genet Genomics* 2007; 34: 35–45.
- [43] Qin YS, Liu RZ, Mei HX, et al. QTL Mapping for yield traits in Upland cotton (*Gossypium hirsutum* L.). *Acta Agronomica Sinica* 2009; 35: 1812–1821.
- [44] Chen H, Qian N, Guo WZ, et al. Using three selected overlapping RILs to fine-map the yield component QTL on Chro.D8 in Upland cotton. *Euphytica* 2010; 176: 321–329.
- [45] Liu R, Wang B, Guo W, et al. Quantitative trait loci mapping for yield and its components by using two immortalized populations of a heterotic hybrid in *Gossypium hirsutum* L. *Mol Breed* 2012; 29: 297–311.
- [46] Shen X, Becelaere GV, Kumar P, et al. QTL mapping for resistance to root-knot nematodes in the M-120 RNR Upland cotton line (*Gossypium hirsutum* L.) of the Auburn 623 RNR source. *Theor Appl Genet* 2006; 113: 1539–1549.
- [47] Wang C, Ulloa M, Roberts PA. Identification and mapping of microsatellite markers linked to a root-knot nematode resistance gene (*rkn1*) in Acala NemX cotton (*Gossypium hirsutum* L.). *Theor Appl Genet* 2006; 112: 770–777.
- [48] Ynturi P, Jenkins JN, McCarty Jr JC, et al. Association of root-knot nematode resistance genes with simple sequence repeat markers on two chromosomes in cotton. *Crop Sci* 2006; 46: 2670–2674.
- [49] Romano GB, Sacks EJ, Stetina SR, et al. Identification and genomic location of a reniform nematode (*Rotylenchulus reniformis*) resistance locus (Renari) introgressed from *Gossypium aridum* into upland cotton (*G. hirsutum*). *Theor Appl Genet* 2009; 120: 139–150.
- [50] Bolek Y, El-Zik Km, Pepper AE, et al. Mapping of Verticillium wilt resistance genes in cotton. *Plant Sci* 2005; 168: 1581–1590.
- [51] Wang HM, Lin ZX, Zhang XL, et al. Mapping and quantitative trait loci analysis of verticillium wilt resistance genes in cotton. *J Integr Plant Biol* 2008; 50: 174–182.
- [52] Jiang F, Zhao J, Zhou L, et al. Molecular mapping of Verticillium wilt resistance QTL clustered on chromosomes D7 and D9 in upland cotton. *Sci China C Life Sci* 2009; 52: 872–884.
- [53] Ning Z, Zhao R, Chen H, et al. Molecular tagging of a major quantitative trait locus for broad-spectrum resistance to Verticillium wilt in upland cotton cultivar Prema. *Crop Sci* 2013; 53: 1–9.

- [54] Wang C, Roberts PA. A Fusarium wilt resistance gene in *Gossypium barbadense* and its effect on root-knot nematode-wilt disease complex. *Phytopathology* 2006; 96: 727–734.
- [55] Chen X, Ge F, Wang D, et al. Mapping of QTLs conferring resistance to Fusarium wilt in cotton. *Molecular Plant Breeding* 2008; 6: 1127–1133.
- [56] Wang P, Su L, Qin L, et al. Identification and molecular mapping of a Fusarium wilt resistant gene in upland cotton. *Theor Appl Genet* 2009; 119: 733–739.
- [57] Wang P, Shi L, Su L, et al. Quantitative trait loci for resistance against Fusarium wilt based on three cotton F₂ populations. *Agricultural Sciences in China* 2010; 9: 1799–1806.
- [58] Ulloa M, Wang C, Hutmacher RB, et al. Mapping Fusarium wilt race 1 resistance genes in cotton by inheritance, QTL and sequencing composition. *Mol Genet Genomics* 2011; 286: 21–36.
- [59] Ulloa M, Hutmacher RB, Roberts PA, et al. Inheritance and QTL mapping of Fusarium wilt race 4 resistance in cotton. *Theor Appl Genet* 2013; 126: 1405–1418.
- [60] Lopez-Lavalle LAB, Gillespie VJ, Tate WA, et al. Molecular mapping of a new source of Fusarium wilt resistance in tetraploid cotton (*Gossypium hirsutum* L.). *Mol Breed* 2012; 30: 1181–1191.
- [61] Rong J, Feltus FA, Waghmare VN, et al. Meta-analysis of polyploid cotton QTL shows unequal contributions of subgenomes to a complex network of genes and gene clusters implicated in lint fiber development. *Genetics* 2007; 176: 2577–2588.
- [62] Lacape JM, Llewellyn D, Jacobs J, et al. (2010) Meta-analysis of cotton fiber quality QTLs across diverse environments in a *Gossypium hirsutum* x *G. barbadense* RIL population. *BMC Plant Biol* 2010; 10: 132.
- [63] Zhu C, Gore M, Buckler ES, et al. Status and prospects of association mapping in plants. *Plant Genome* 2008; 1: 5–20.
- [64] Kantartzi SK, Stewart JMcD. Association analysis of fibre traits in *Gossypium arboreum* accessions. *Plant Breed* 2008; 127: 173–179.
- [65] Abdurakhmonov IY, Kohel RJ, Yu JZ, et al. Molecular diversity and association mapping of fiber quality traits in exotic *G. hirsutum* L. germplasm. *Genomics* 2008; 92: 478–487.
- [66] Abdurakhmonov IY, Saha S, Jenkins JN, et al. Linkage disequilibrium based association mapping of fiber quality traits in *G. hirsutum* L. variety germplasm. *Genetica* 2009; 136: 401–417.
- [67] Zeng L, Meredith Jr WR, Gutierrez OA, et al. Identification of associations between SSR markers and fibre traits in an exotic germplasm derived from multiple cross among *Gossypium* tetraploid species. *Theor Appl Genet* 2009; 119: 93–103.

- [68] Rakshit A, Rakshit S, Singh J, et al. Association of AFLP and SSR markers with agronomic and fibre quality traits in *Gossypium hirsutum* L. J Genetics 2010; 89: 155–162.
- [69] Zhang T, Qian N, Zhu X, et al. Variations and transmission of QTL alleles for yield and fiber qualities in Upland cotton cultivars developed in China. PLoS ONE 2013; 8: e57220.
- [70] Wang XQ, Yu Y, Li W, et al. Association analysis of yield and fiber quality traits in *Gossypium barbadense* with SSRs and SRAPs. Genet Mol Res 2013; 12: 3353–3362.
- [71] Mei H, Zhu X, Zhang T. Favorable QTL alleles for Yield and Its Components Identified by Association Mapping in Chinese Upland Cotton Cultivars. PLOS ONE 2013; 8: e82193.
- [72] Mei H, Ai N, Zhang X, et al. QTLs conferring FOV 7 resistance detected by linkage and association mapping in Upland cotton. Euphytica 2014; Doi 10.1007/s10681-014-1063-y.
- [73] Cai C, Ye W, Zhang T, et al. Association analysis of fiber quality traits and exploration of elite alleles in Upland cotton cultivars/accessions (*Gossypium hirsutum* L.). J Integr Plant Biol 2014; 56: 51–62.
- [74] Breseghello F, Sorrells ME. Association mapping of kernel size and milling quality in wheat (*Triticum aestivum* L.) cultivars. Genetics 2006; 172: 1165–1177.
- [75] Jia L, Yan W, Zhu C, et al. Allelic analysis of sheath blight resistance with association mapping in rice. PLoS ONE 2012; 7: e32703.
- [76] Huang ZK., editor. The Cultivars and Their Pedigree of Cotton in China. China Agriculture Press, Beijing; 2007.
- [77] Du X, Sun J, Zhou Z, et al. Current situation and the future in collection, preservation, evaluation and utilization of cotton germplasm in China. J Plant Genetic Resources 2012; 13: 163–168.
- [78] Guo W, Cai C, Wang C, et al. A microsatellite-based, gene-rich linkage map reveals genome structure, function and evolution in *Gossypium*. Genetics 2007; 176: 527–541.
- [79] Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multi-locus genotype data. Genetics 2000; 155: 945–959.
- [80] Evanno G, Regnaut S, Goudet J. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. Mol Ecol 2005; 14: 2611–2620.
- [81] Hardy O, Vekemans X. SPAGeDi: a versatile computer program to analyse spatial genetic structure at the individual or population levels. Mol Ecol Notes 2002; 2: 618–620.
- [82] Bradbury PJ, Zhang Z, Kroon DE, et al. TASSEL: software for association mapping of complex traits in diverse samples. Bioinformatics 2007; 23: 2633–2635.

- [83] Lander ES, Botstein D. Mapping mendelian factors underlying quantitative traits using RFLP linkage maps. *Genetics* 1989; 121: 185–199.
- [84] Wang Y, Yang W, Xu H, et al. The main problems and recommendations in Chinese cotton production. *Chin Agr Sci Bull* 2009; 25: 86–90.
- [85] Qian S, Huang J, Peng Y, et al. Studies on the hybrid of *G. hirsutum* L. and *G. anomalum* Wawr. & Peyr. and application in breeding. *Sci Agric Sin* 1992; 25: 44–51.
- [86] Zhou B, Zhu X, Guo W, et al. Major gene plus poly-gene inheritance of super quality fiber properties in upland cotton introgressed from *Gossypium anomalum* Wawra & Peyritsch. *Cotton Sci* 2006; 18: 60–62.
- [87] Ma C., editor. Researches on Fusarium and Verticillium wilt in cotton. China Agriculture Press, Beijing; 2007.
- [88] Davis RM, Colyer PD, Rothrock CS, et al. Fusarium wilt of cotton: population diversity and implications for management. *Plant Dis* 2006; 90: 692–703.
- [89] Ulloa M, Hutmacher RB, Davis RM, et al. Breeding for Fusarium wilt race 4 resistance in cotton under field and greenhouse conditions. *J Cotton Sci* 2006; 10: 114–127.
- [90] Constable GA, Reid PE, Stiller WN. Breeding for resistance to a new strain of Fusarium wilt in Australia. In: Stephens M. (eds.) Proceedings of the World Cotton Research Conference-4, Sept 10-14, Lubbock TX; 2007.
- [91] Chen Q, Ji X, Sun W. Identification of cotton-wilt *Fusarium* races in China. *Scientia Agricultura Sinica* 1985; 6: 1–6.
- [92] Wang B, Brubaker CL, Burdon JJ. Fusarium species and Fusarium wilt pathogens associated with native *Gossypium* populations in Australia. *Mycol Res* 2004; 108: 35–44.
- [93] Kim Y, Hutmacher RB, Davis RM. Characterization of California isolates of *Fusarium oxysporum* f. sp. *vasinfectum*. *Plant Dis* 2005; 89: 366–372.
- [94] Holmes EA, Bennett RS, Spurgeon DW, et al. New genotypes of *Fusarium oxysporum* f. sp. *vasinfectum* from the Southeastern United States. *Plant Dis* 2009; 93: 1298–1304.
- [95] Ma C, Jian G, Zheng C. The advances in cotton breeding resistance to Fusarium and Verticillium wilts in China during past fifty years. *Scientia Agricultura Sinica* 2002; 35: 508–513.
- [96] Ulloa M, Percy R, Zhang J, et al. Registration of four Pima cotton germplasm lines having good levels of Fusarium wilt race 4 resistance with moderate yields and good fibers. *J Plant Registr* 2009; 3: 198–202.
- [97] Smith AL, Dick JB. Inheritance of resistance to Fusarium wilt in upland cotton and sea island cotton as complicated by nematodes under field conditions. *Phytopathology* 1960; 50: 44–48.

- [98] Netzer D, Tal Y, Marani A, et al. Resistance of interspecific cotton hybrids (*Gossypium hirsutum* x *Gossypium barbadense* containing *Gossypium harknessii* cytoplasm) to fusarium wilt. *Plant Dis* 1985; 69: 312–313.
- [99] Feng C, Zhang J, Liu J, et al. Inheritance of resistance to Fusarium wilt in several Chinese upland cotton varieties. *Acta Agronomica Sinica* 1996; 22: 550–554.
- [100] Kappelman AJ. Inheritance of resistance to Fusarium wilt in cotton. *Crop Sci* 1971; 11: 672–674.
- [101] Jiao B. Primary research of general combining ability and heritability of resistance to Fusarium wilt and Verticillium wilt in upland cotton. *Acta Agronomica Sinica* 1985; 11, 267–273.
- [102] Wang Z, Ma Z, Qu J. Analysis of gene effect of resistance to Fusarium and Verticillium wilt in upland cotton. *Journal of Heibei Agriculture University* 1989; 12: 21–25.
- [103] Flint-Garcia SA, Thuillet AC, Yu J, et al. Maize association population: a high-resolution platform for quantitative trait locus dissection. *Plant J* 2005; 44: 1054–1064.
- [104] Abdurakhmonov IY, Abdukarimov A. Application of association mapping to understanding the genetic diversity of plant germplasm resources. *Int J Plant Genomics* 2008; ID 574927. doi:10.1155/2008/574927.
- [105] Gupta PK, Rustgi S, Kulwal PL. Linkage disequilibrium and association studies in higher plants: present status and future prospects. *Plant Mol Biol* 2005; 57: 461–485.
- [106] Price A, Patterson N, Plenge R, et al. Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet* 2006; 38: 904–909.
- [107] Yu J, Pressoir G, Briggs WH, et al. A unified mixed-model method for association mapping that accounts for multiple levels of relatedness. *Nat Genet* 2006; 38: 203–208.
- [108] Cavanagh C, Morell M, Mackay I, et al. From mutations to MAGIC: resource for gene discovery, validation and delivery in crop plants. *Cur Opin Plant Bio* 2008; 11: 215–221.
- [109] Yu J, Holland JB, McMullen MD, et al. Genetic design and statistical power of nested association mapping in maize. *Genetics* 2008; 178: 539–551.
- [110] Buckler ES, Holland JB, Bradbury PJ, et al. The genetic architecture of maize flowering time. *Science* 2009; 325: 714–718.
- [111] Kover PX, Valdar W, Trakalo J, et al. A multiparent advanced generation inter-cross to fine-map quantitative traits in *Arabidopsis thaliana*. *PLoS Genetics* 2009; 5: e1000551.
- [112] Myles S, Peiffer J, Brown PJ, et al. Association mapping: critical consideration shift from genotype to experimental design. *Plant Cell* 2009; 21: 2194–2202.

- [113] Nemri A, Atwell S, Tarone AM, et al. Genome-wide survey of *Arabidopsis* natural variation in downy mildew resistance using combined association and linkage mapping. *Proc Natl Acad Sci USA* 2010; 107: 10302–10307.
- [114] Lu Y, Zhang S, Shah T, et al. Joint linkage-linkage disequilibrium mapping is a powerful approach to detecting quantitative trait loci underlying drought tolerance in maize. *Proc Natl Acad Sci USA* 2010; 107: 19585–19590.
- [115] Brachi B, Faure N, Horton M, et al. Linkage and association mapping of *Arabidopsis thaliana* flowering time in nature. *PLoS Genet* 2010; 6: e1000940.
- [116] Pagny G, Paulstephenraj PS, Poque S, et al. Family-based linkage and association mapping reveals novel genes affecting *Plum pox virus* infection in *Arabidopsis thaliana*. *New Phytologist* 2012; 196: 873–886.
- [117] Korir PC, Zhang J, Wu K, et al. (2013) Association mapping combined with linkage analysis for aluminum tolerance among soybean cultivars released in Yellow and Changjiang River Valleys in China. *Theor Appl Genet* 2013; doi 10.1007/s00122-013-2082-0.
- [118] Cadic E, Coque M, Vear F, et al. Combined linkage and association mapping of flowering time in Sunflower (*Helianthus annuus* L.). *Theor Appl Genet* 2013; doi 10.1007/s00122-013-2056-2.
- [119] Paterson AH, Wendel JF, Gundlach H, et al. Repeated polyploidization of *Gossypium* genomes and the evolution of spinnable cotton fibres. *Nature* 2012; 492: 423–427.
- [120] Wang K, Wang Z, Li F, et al. The draft genome of a diploid cotton *Gossypium raimondii*. *Nat Genet* 2012; 44: 1098–1103.
- [121] Davey JW, Hohenlohe PA, Etter PD, et al. Genome-wide genetic marker discovery and genotyping using next-generation sequencing. *Nat Rev Genet* 2011; 12: 499–510.

