

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

6,900

Open access books available

185,000

International authors and editors

200M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Neuroinflammation on the Epigenetics of Neural Stem Cells

Nando Dulal Das and Young Gyu Chai

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/54981>

1. Introduction

Under physiological conditions, neuronal stem cells (NSCs) can undergo both self-renewal and differentiation stages. The formation of new neurons, neurogenesis, is a vital process by which the brain maintains its lifelong plasticity in response to extrinsic and intrinsic changes. However, the exact mechanisms that regulate NSC self-renewal and differentiation are largely unknown. NSCs become stimulated after neuronal injury and can migrate at pathological sites (Nakatomi et al., 2002; Russo et al., 2011) that dictate the potential of NSCs therapeutic use in pathological conditions of the central nervous system. In this chapter, we describe the effect of neuroinflammation in NSCs and discuss whether the inflammatory mediators can epigenetically affect the capacity of NSCs and alter their proliferation and differentiation ability. The mechanism by which the inflammatory environment influences the NSC niche and thus, alters the self-renewal, survival, migration, and differentiation of the NSCs is currently unknown (Martino and Pluchino, 2006). Several studies have focused the effects of inflammation on the regenerative capacity of NSCs subjected to microglial activation after an acute injury or after LPS treatment. Overall, the connection between brain inflammation and NSC neurogenesis and the role of the niche in the modulation of neuronal differentiation under alternative conditions are under intense investigation.

To gain further insight into these phenomena, we describe epigenetic mechanisms, including DNA methylation and histone modification in NSCs inflammation. DNA methylation and histone modification are known to play significant roles in the modulation of stem cell proliferation and differentiation (Li and Zhao, 2008). Regarding DNA methylation, methylated CpG-binding protein (MBD) deficiency results the suppression of NSCs differentiation. Therefore, to identify the downstream target genes of MBDs has potential in NSCs differentiation study. Histone modifications are another important epigenetic mark. There are many

types of post-translational modifications of the residues at histone tails, including methylation of lysines and arginines, acetylation, phosphorylation, ubiquitination, SUMOylation, and ADP-ribosylation. Among the histone modifications, histone H3 lysine (K) methylation is a central epigenetic modification with both activating and repressive roles in eukaryotic chromatin (Reinberg et al., 2004).

Next, we will focus on epigenetic involvement in neurodegenerative diseases and NSCs. Actually, inflammatory stimuli induce beneficial effects (e.g., phagocytosis of debris and apoptotic cells), and inflammation is linked to tissue repair processes, uncontrolled inflammation may result in production of neurotoxic factors that amplify underlying disease states and pathogenesis of Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), multiple sclerosis (MS), and a growing number of other nervous system pathologies (Glass et al., 2010). Here, endogenous NSCs cannot fully compensate the neuronal loss in such neurodegenerative diseases. The possible reasons include the lack of trophic support and inhibitory signals within the brain microenvironment (Croft and Przyborski, 2009), indicative of oxidative stress (Kelly et al., 2011) and age-related neuroinflammation. In summary, the recent development of stem cells technology open new areas of research aimed at stimulating neuronal regeneration in the brain during aging, neuroinflammation and neurodegenerative diseases (Russo et al., 2011). Here, we overview the therapeutic approach of NSC and how these stem cells are responsible for brain homeostasis, induction of neurogenesis in several diseased states. Finally, this chapter indicates the possibility of combination therapy of epigenetic drug with NSC transplantation in these neurodegenerative diseases.

2. Epigenetics and neuroinflammation

Alterations in cell signaling by environmental changes can remodel epigenetic marks (Borrelli et al., 2008; Weaver et al., 2007). Epigenetics thus presents potential explanations for sustained changes in transcriptional activity associated with cell differentiation, learning and memory, age-related neurodegeneration and effects of early experience, repeated drug exposure, chronic stress, and environmental toxins. The implicit hypothesis is that environmental signals alter chromatin modifications, which then serve as the mechanism for the transcriptional 'plasticity' that mediates sustained variation in neural function (Meaney and Ferguson-Smith, 2010). Although most extensively studied in embryonic stem cells, such 'bivalent' domains, which are also found in the adult brain (Sanz et al., 2008), suggest a developmentally 'poised' state awaiting environmental direction. Indeed, such states may mark the potential for plasticity. The same epigenetic mark can recruit effectors that activate as well as others that repress transcription. We describe the detail epigenetic changes in NSC later in this chapter. We summarize the recent evidence that physiological and environmental signals influence adaptive transcriptional responses in neurons through the epigenetic modification of chromatin. We highlight to the regulation of histone modifications and DNA methylation in response to neuroinflammation and related signaling. In addition, mechanisms that induce chromatin modifications in association with multiprotein complexes on neuronal gene promoters are mentioned.

3. NSC and inflammation

To maintain brain homeostasis, NSCs are highly controlled under physiological conditions in which the stem cell niche is vital for the NSC self-renewal, proliferation, differentiation, and migration. NSCs become activated after neuronal injury and migrate to the site of injury, indicating that some regulators at the injury site can guide the migration of precursor cells. Damaged neurons can be repaired by the activation of endogenous neuronal stem cells, which migrate to regions of the brain injury, differentiate into neuronal cells, and integrate into neuronal circuits (Belmadani et al., 2006; Russo et al., 2011). The mechanism by which the inflammatory environment influences the NSC niche and thus, alters the self-renewal, survival, migration, and differentiation of the NSCs is currently unknown (Martino and Pluchino, 2006). Alterations of NSC functions either pro-neurogenic or anti-neurogenic in inflammation may depend on the NSC niche and activation of brain microglial cells. It is reported that activated microglia in inflammatory conditions can inhibit neurogenesis (Butovsky et al., 2006). On the contrary, activated microglia also showed helpful for neurogenesis (Hanisch and Kettenmann, 2007). Inflammatory cytokines and nitric oxide (NO) released by microglial cells can inhibit the adult neurogenesis. Activation of microglia with LPS results the production of inflammatory mediators *in vitro*, including TNF- α and IL-6, that inhibit the generation of neurons from NSCs (Monje et al., 2003). However, modification of microglial status by other cytokines, such as IL-4 or low dose interferon- γ (IFN- γ) changes their phenotype to strongly promote neurogenesis (Butovsky et al., 2006). However, the positive effects are at least partly dependent on microglia production of insulin-like growth factor-1 (IGF-1), a potent proneurogenic growth factor. Though controversial, this raises the possibility that some types of controlled inflammation may be exploited in CNS regeneration or in combating neurological diseases that have pronounced chronic proinflammatory components (Rolls et al., 2009).

Several studies have focused the effects of inflammation on the regenerative capacity of NSCs subjected to microglial activation after an acute injury or after LPS treatment. It is reported that TLR4 is expressed by NSCs, and LPS suppresses the proliferation of NSCs under culture conditions via an NF- κ B-dependent mechanism (Monje et al., 2003; Rolls et al., 2007). In addition, TLR4 can directly modulate the self-renewal and cell-fate decision of neuronal progenitor cells (Rolls et al., 2007). Overall, the effects of proinflammatory signaling on NSCs go beyond simple changes in the abundance of new neurons (Carpentier and Palmer, 2009). It is shown that the neurons generated during the period of inflammation are morphologically normal, with normal cell body location, polarity, and branching, yet they display an accentuated inhibitory or excitatory responses in immature versus mature neurons, respectively (Jakubs et al., 2008). So, the functions of new neurons are severely affected by immune signaling. Moreover, the connection between brain inflammation and NSC neurogenesis and the role of the niche in the modulation of neuronal differentiation under alternative conditions are under intense investigation.

4. Epigenetic significance in NSC-inflammation

Epigenetic refers to any heritable influence (in the progeny of cells or individuals) on chromosome or gene function that is not accompanied by a change in DNA sequence (Yoder et al., 1997). It includes processes such as DNA methylation, histone modification and noncoding RNA expression. Appropriate gene function either activation or repression at inflammatory stages of NSC progression could be achieved by such epigenetic regulation. Here, we cover recent reports involving the role of epigenetic mechanisms in NSC-inflammation and its fate on NSC mechanisms. One of the important epigenetic mechanisms, DNA methylation in the genome is established by a family of DNA methyltransferases (DNMTs). Maintenance of methylation patterns is achieved by a function of DNMT1 during DNA replication, while de novo methylation is primarily catalyzed by DNMT3a and DNMT3b. DNA methylation is responsible for the regulation of gene expression, where two mechanisms are involved. First, methylation of CpG dinucleotides affects DNA structure and can directly interfere with the binding of TFs to their target sequences (Takizawa et al., 2001); second, a more pervasive effect, methyl-CpG-binding domain (MBD)-containing protein family members can bind to genes with methylated CpG dinucleotides, thereby suppressing the genes' expression (Nan et al., 1997). Though, DNA methylation is actively involved in the acquisition of multipotentiality in NSC from early-, mid- to late-gestation. Here, we mainly focus on two well-studied pathways that act synergistically to promote astrocytic differentiation of NSC are those activated by the interleukin-6 (IL-6) family of cytokines such as leukemia inhibitory factor (LIF), ciliary neurotrophic factor (CNTF) and cardiotrophin-1 (CT-1) and bone morphogenetic protein (BMP) signaling (Juliandi et al., 2010). In early- and mid-gestational NSCs, astrocytic gene promoters such as glial fibrillary acidic protein (GFAP) are hypermethylated, a status that impedes binding of the STAT3-p300/CBP-SMADs complex to its target sequence and thus prevents these NSCs from differentiating into astrocytes even when the cells are stimulated by astrocyte-inducing cytokines (Takizawa et al., 2001). These IL-6 family cytokines have been shown to be expressed in NSCs and neurons in the fetal mouse brain (Barnabe-Heider et al., 2005); but how DNA methylation of IL-6 does affect of NSC differentiation in inflammation is not disclosed. On the contrary, the STAT3 binding site-containing GFAP promoter in NSCs at late gestation is barely methylated, so that upon LIF stimulation these NSCs can differentiate into astrocytes (Takizawa et al., 2001). Overall, the genome-wide DNA methylation status of NSCs in well-defined inflammatory conditions may give us possible clue of gene specific methylation status of NSC whether DNA methylation can play an important role in defining the NSC fate from neurogenesis to astrocytogenesis in inflammatory conditions. Notch signaling is a conserved pathway from insects to mammals, which contributes to cell-to-cell communication (Louvi and Artavanis-Tsakonas, 2006) and controls cell fate determination in the CNS (Lundkvist and Lendahl, 2001). Upon Notch activation by its ligand, the Notch intracellular domain (NICD) is released from the plasma membrane and is translocated into the nucleus, where it converts a particular repressor complex into an activator complex (Nakayama et al., 2008). It is confirmed that Notch ligands are indeed expressed in neuronally committed NPCs and young neurons, and that these ligands activate Notch signaling in the residual NSCs. Further, forced expression of NICD in midgestational NSCs induced the

upregulation of nuclear factor 1A (NF1A), which in turn accelerated demethylation of astrocytic gene promoters by preventing DNMT1 from binding to them and thus allowed precocious astrocytic differentiation in response to LIF stimulation (Namiyama et al., 2009).

It has shown that methyl binding domain (MBD) proteins expressed predominantly in neurons, and not in astrocytes or oligodendrocytes, in the CNS (Kishi and Macklis, 2004); may regulate in NSC differentiation. It was found that exon1 of GFAP are hypermethylated in all neural cell types and that only in neurons, methyl-CpG-binding protein 2 (MeCP2), a member of the MBD family, is highly expressed and binds to this methylated exon1 region (Setoguchi et al., 2006) that is linked to block the astrocyte differentiation. Indeed, ectopic expression of MeCP2 directs NSCs to become neurons and inhibits astrocytic differentiation, even in the presence of astrocyte-inducing cytokines such as LIF and BMP2 (Tsujimura et al., 2009). MBD1-deficient NSCs generate fewer neurons than do wild type NSCs, suggesting an important role for MBD1 in neuronal fate specification (Zhao et al., 2003).

Histone proteins within the chromosome play a significant role in chromatin structure, gene transcription and epigenetic information. Multiple modifications decorate each histone tail within the nucleosome, and some amino acids on the histone tail can be modified in several different ways. Covalent modifications of histone tails include methylation, acetylation, phosphorylation, ubiquitylation, sumoylation, glycosylation, biotinylation, carbonylation and adenosine diphosphate (ADP)-ribosylation (Strahl and Allis, 2000). Among these, modifications by histone acetylation and methylation are the most common. Acetylation and deacetylation of lysine residue in histone tails is mediated by histone acetyl transferases (HATs) and histone deacetylases (HDACs), respectively (Hsieh and Gage, 2005). Histone acetylation by HATs is responsible for open chromatin (euchromatin) formation that leads to transcriptional activation. Conversely, HDACs result decrease of histone acetylation and formation of condensed chromatin (heterochromatin) that causes transcriptional silencing. Adult hippocampal-derived NSCs differentiate predominantly into neurons, at the expense of astrocytes and oligodendrocytes, when treated by the antiepileptic and HDAC inhibitor valproic acid (VPA) *in vitro*, even in conditions that favor glia-specific differentiation (Hsieh et al., 2004). VPA-mediated HDAC inhibition upregulates the neuron-specific gene *NeuroD*, a neurogenic basic helix-loop-helix transcription factor (TF), is resulting in the induction and suppression, respectively, of neuronal and glial differentiation. In the developing rat brain and in cultured E14 NSCs, VPA treatment has also been shown to promote neurogenesis by activating the Ras-ERK pathway (Jung et al., 2008).

Histone methylation is involved in the regulation of a variety of nuclear processes dedicated to the maintenance of active and silent states of gene expression, which is essential for cellular regulation, homeostasis and fate determination (Cloos et al., 2008). There are five lysine residues in the histone N termini that are prominently methylated. H3K4 and H3K36 methylation primarily transduce activating functions, whereas H3K9, H3K27, and H4K20 methylation is mainly associated with repressed chromatin. Histone lysine methylation can result in mono-, di-, or trimethyl states and each distinct methylation state confers different biological read-outs. Histone lysine trimethyl states, particularly those with repressive functions, appear relatively robust because they are stably propagated during several cell divisions (Lachner et

al., 2004). Among the histone modifications, histone H3 lysine (K) methylation is a central epigenetic modification with both activating and repressive roles in eukaryotic chromatin (Reinberg et al., 2004). JmjC domain proteins demethylate histone lysine and arginine residues in an oxidative reaction that requires Fe (II) and α -ketoglutarate as cofactors. Depending on their target specificity, JmjC domain proteins promote transcriptional repression or activation, thereby impacting important processes such as hormone response, stem cell renewal, germ cell development, and cellular proliferation and differentiation (Beyer et al., 2008). Interestingly, a range of JmjC proteins is induced in different cancers and has been linked to cell proliferation (Cloos et al., 2006) and the suppression of senescence (Pfau et al., 2008). Members of the JMJD2 family that target H3K9me3/me2 and H3K36me3/me2 are highly expressed in prostate cancer (Wissmann et al., 2007).

Recently, we reported the effect of lipopolysaccharides (LPS) on NSCs epigenetics, where we used an immortalized neuroectodermal stem cell line, NE-4C. The NE-4C cell line was cloned from the anterior brain vesicles of E9 mouse embryos lacking functional p53 (Livingstone et al., 1992). Non-induced NE-4C cells grow as homogeneous, epithelial-like populations, and in response to all-trans retinoic acid (RA) treatment, they differentiate into neurons on a highly reproducible schedule (Jelitai et al., 2004). We found that histone demethylase, Jmjd2b is functional in long-term LPS treatment and regulates the histone demethylation of the promoters of its target genes that may be crucial in multiple signaling pathways and biological processes in murine NSCs (NE-4C cells). MetaCore pathway analysis revealed the gene networks and canonical pathways affected in Jmjd2b-attenuated NE-4C cells that involved neurophysiological processes (receptor-mediated axon growth repulsion, GABA-A receptor life cycle), the Notch1-mediated pathway for NF- κ B activity modulation, and TGF- β -dependent induction. Several extrinsic factors affect the histone methylation status of NSCs. In the postnatal mouse brain, MLL1 is required for neurogenesis and its deficiency in NSCs in the subventricular zone (SVZ) leads to a glial lineage preference. One of the key downstream regulators of SVZ neurogenesis, Dlx2, is not expressed in MLL1-deficient NSCs. This is due to a change in histone methylation of Dlx2, from a single high level of H3K4 trimethylation (H3K4me3) to a bivalent poised state marked by both activating H3K4me3 and repressive H3K27me3 (Lim et al., 2009).

We found that Jmjd2b is functional in long-term LPS treatment and regulates the histone demethylation of the promoters of its target genes that may be crucial in multiple signaling pathways and biological processes in NE-4C cells. Jmjd2b is a newly identified member of the histone demethylase Jmjd2 family that is characterized by the catalytic Jumonji C (JmjC) domain. Jmjd2b specifically targets the trimethylated lysine 9 of histone H3 (H3K9) for demethylation at pericentric heterochromatin and euchromatin (Fodor et al., 2006). It is reported that JMJD2B is critical to breast cancer cell survival under conditions of normoxia and hypoxia, which occurs partially via the regulation of cell cycle progression, is highly expressed in ER α -positive primary breast cancers, and is an adverse prognostic factor in hypoxic breast cancers (Yang et al., 2010). In this study, MetaCore pathway analysis was used to reveal the gene networks and canonical pathways affected in Jmjd2b-kd cells. Among the network, generation of neurons, neurogenesis, cell differentiation, and cellular developmental

processes were most significantly affected in Jmjd2b-attenuated NE-4C cells. The significantly downregulated genes were clustered in different networks and canonical pathways. We found that Jmjd2b-kd NE-4C cells downregulated various key genes involved in neurophysiological processes (receptor-mediated axon growth repulsion, GABA-A receptor life cycle), the Notch1-mediated pathway for NF- κ B activity modulation, and TGF- β -dependent induction. Jmjd2b encodes a histone demethylase that has been recently shown to be a HIF-1 α target gene (Yang et al., 2009). Jmjd2b attenuation significantly inhibited p53, iNOS, Bcl2 and TGF- β expression in Jmjd2b-kd NE-4C cells. A GeneGo analysis of Jmjd2b-kd NE-4C cells revealed that Jmjd2b attenuation affected the generation of neurons, neurogenesis, system development, cell differentiation and cellular development processes. Several genes involved in the receptor-mediated axon growth repulsion (semaphorin 3a, pleiotrophin-OSF1, ephrin A receptor 2), the GABA-A receptor life cycle (GABA-A receptor beta 2), the NOTCH1-mediated pathway for NF- κ B activity modulation (c-Rel, Jagged 1, p53/p52) and the TGF- β -dependent induction (TGF- β 2, Jagged1, N-cadherin, Lef1) were directly or indirectly affected by Jmjd2b attenuation. We predict that Jmjd2b recruitment may be necessary for the expression of regulated genes from several pathways that are crucial for various neurological functions. These results suggest that LPS has an inflammatory effect on NE-4C cells via epigenetic modulation.

It has also been reported that the mRNA expression of NeuroD, a neural progenitor cell marker, was significantly decreased in the hippocampus of aged mice compared with that in young mice. In light of previous results, we examined the presence of H3K9me3 at the NeuroD promoter but did not observe a reduction of the H3K9me3 level in Jmjd2b-kd NE-4C cells. We predicted that other histone modifications might be involved at the promoter site of NeuroD for its expression. However, the functions of most histone demethylases, including Jmjd2b, are not clear under inflammatory conditions, and the mechanism by which Jmjd2b epigenetically regulates gene expression in NSC inflammation has not been well shown. Therefore, the clarification of the function of Jmjd2b may help to identify novel therapeutic targets for brain inflammation.

5. Epigenetic regulations of proinflammatory cytokines in NSC

Cytokines are the secreted molecules that mediate communication between immune cells and between immune system and host. Cytokines encompass a broad class of signaling molecules that have the potential to influence an immense variety of signals that regulate NSC function, including growth factor production, electrical activity, synaptic function, and axonal path finding (Carpentier and Palmer, 2009). We will focus our discussion on the epigenetic regulations of inflammatory cytokines in NSC. Though, several recent reports shown that important cytokines include TNF- α , IL-6, and IL-1 β have prominent inhibitory effect on adult neurogenesis in vivo. TNF- α can induce apoptosis in NSCs or newborn neurons via TNFR1. TNFR1 signaling, but not that of TNFR2, has been demonstrated to inhibit neurogenesis in the normal hippocampus (Iosif et al., 2006). In addition, neurogenesis is severely affected by another strong inflammatory mediator, NO. It has been reported that the SVZ cell proliferation rate is significantly increased after the inhibition of neuronal NOS activity (Sun et al., 2005). Notably, the

pathological concentration of NO has a skewing effect on NSC differentiation when the astroglial fate is very dominant (Covacu et al., 2006). At present, not many studies have been reported regarding the epigenetic involvement for cytokine regulations in NSC; recently, we reported LPS could affect NSC in vitro via epigenetic regulation (Das et al., 2012). The in vitro treatment of NE-4C cells with LPS (1 $\mu\text{g/ml}$ for 96 h) significantly increased Jmjd2b expression and decreased the levels of H3K9me3. It has been reported that IL-1 β suppresses the proliferation of hippocampal progenitor cells (Koo and Duman, 2008). The decreased proliferation of neural stem cells is responsible for decreased neural differentiation, and increased proliferation could correspond to the promotion of neurogenesis. We predicted that H3K9me3 is involved in Jmjd2b-attenuated NE-4C cells. A ChIP analysis showed that Jmjd2b-attenuated samples experienced an increase in the H3K9me3 on inflammatory signaling-mediated genes. An induced presence of H3K9me3 has been observed at the promoters of the Notch1, IL-1 β , and IL-2 genes in Jmjd2b-kd NE-4C cells, suggesting that Jmjd2b can fine-tune the local chromatin state to enhance the transcription of these genes (Das et al., 2012).

6. Potential of NSC in neurodegenerative diseases

Neurogenesis by endogenous NSC cannot fully overcome the neuronal loss observed in neurodegenerative diseases. One reason for this limited response is the lack of trophic support and inhibitory signals within the brain microenvironment (Croft and Przyborski, 2009), indicative of oxidative stress and age-related neuroinflammation. These observations stimulated a search for agents that could increase neurogenesis and enhance neuroprotection (Russo et al., 2011). Now we will discuss the various neurodegenerative diseases including Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), multiple sclerosis (MS), epilepsy, and stroke in inflammatory contexts. Each of the neurodegenerative diseases considered here is distinguished by a disease-specific mechanism for induction of inflammatory responses. The distinct pathways for production of inducers of inflammation—such as Ab, α -synuclein, mutant SOD1, and myelin peptide mimetic—and the specific anatomical locations at which these processes occur are likely determinants of the specific pathological features of each disease. In particular, TLRs and other pattern recognition receptors expressed on microglia and astrocytes are likely to play significant roles in initiating inflammatory responses. Later the downstream signal transduction pathways like NF- κ B and AP-1 appear to play general roles in mediating the production of amplifiers and effector molecules, such as cytokines (e.g., TNF- α , IL-1 β , and IL-6), ROS, and NO which involving in neurotoxicity for all of the neurodegenerative diseases (Glass et al., 2010).

Now we will focus on epigenetic involvement for such neurodegenerative diseases. Genes that are epigenetically regulated in Alzheimer's disease are *S100A2* (a member of the S100 family of calcium-binding proteins) and *SORBS3* (a sorbin and SH3 domain containing the cell-adhesion Protein) that display significant different level of DNA methylation (Siegmund et al., 2007). *S100A2* has been previously identified as a metastatic inductor in non-small-cell lung cancer (Bulk et al., 2009), but its role in Alzheimer's disease pathogenesis remains unknown.

Most importantly, S100B, another member of the S100 family, which acts as a neurotrophic and pro-survival neuronal factor, might have a role in Alzheimer's disease pathogenesis and how does the exogenous and endogenous NSC express and epigenetically regulate such neurotrophic factors is still unknown. Among other epigenetic regulations in the pathogenesis of PD; DNA hypomethylation of TNF- α can directly lead to specific vulnerability of the substantia nigra could be the direct consequence of PD (Pieper et al., 2008) and that may potentiate why the cytokine mediated inflammation is one of the major causes for PD. It is also reported that TNF- α overexpression induces apoptosis in neuronal cells and TNF- α levels are high in the CSF of patients with Parkinson's disease (Mogi et al., 1996). Multiple sclerosis is an inflammatory chronic disease characterized by a demyelinating process, which is followed by neurodegeneration. Although little is known about the epigenetics of this disease, some evidence suggests hypomethylation was proven at the promoter region of *PADI2* (peptidyl arginine deiminase, type II), also found to be overexpressed in multiple sclerosis. *PADI2* catalyzes the citrullination of myelin basic protein that can change the properties of myelin (Mastronardi et al., 2007; Urdinguio et al., 2009). Epilepsy is described as a common chronic neurological disorder characterized by recurrent spontaneous seizures. Sporadic epilepsy can arise as a result of traumatic brain injury, stroke, abnormalities in brain wiring, toxic-metabolic etiologies, inflammation, autoimmunity, or an imbalance in the ratio of inhibitory to excitatory synaptic transmission (Hwang et al, 2012; Berg et al, 2010). Spontaneous seizures activate REST and promote deacetylation of core histone protein H4 (a mark of gene repression) at the RE1 site of the *glra2* promoter (gene encoding the AMPAR subunit GluA2) recruits mSin3A and CoREST, HDACs-1/2, G9a and MeCP2, while promoting an increase in acetylation of H4 (a mark of open chromatin) at the promoter of brain-derived neurotrophic factor BDNF (Tsankova et al, 2004). Although, GluA2 expression was decreased, leading to an increase in GluA2-lacking, Ca²⁺-permeable AMPARs at CA3 synapses and neuronal death in CA3. Alterations of these proteins contribute to the pathophysiology of recurrent seizures. In epileptic adult rats transplanted fetal NSC (E14 rat) cells differentiated into neurons (13%, mostly GABAergic) and astrocytes (57%) and showed a reduction of motor seizure by 43% and severe convulsive seizure by 90% (Waldau et al., 2010). But how does the epigenetic regulators in exogenous NSC play crucial role in epilepsy yet to disclose. Ischemic insults also trigger activation of REST in mature hippocampal neurons destined to die and that the increase in REST correlates with a decrease in histone acetylation and gene silencing of GluA2. This is significant in that the GluA2 subunit prevents Ca²⁺ influx via AMPA receptors (AMPA receptors), is essential to synaptogenesis, long lasting forms of synaptic plasticity and neuronal death (Hwang et al., 2012; Liu and Zukin, 2007). Since, REST is a master transcriptional regulator of neuronal genes in pluripotent stem cells and neural progenitors and that loss of REST during the late stages of neural differentiation by ubiquitin based proteosomal degradation is required for acquisition of the neural phenotype (Hwang et al., 2012). The more study needs to answer how does REST perform at transplanted NSC in stroke model and whether other synaptic proteins correlate with REST for neuronal death in a clinically relevant ischemic stroke model.

Finally, more research will be required to understand the epigenetic mechanisms that underlie the neuroprotective roles of NSC in neurodegenerative diseases.

7. Conclusion

Due to self-renewal ability and differentiation to various neural cell types, NSC has great potential for clinical treatment of neurological diseases and dysfunctions. This regenerative capacity of NSC hold a great promise to open new areas of research aimed at stimulating neuronal regeneration in the brain during aging, neuroinflammation and neurodegenerative diseases. Epigenetic regulation along with other mechanisms can control these properties of NSCs. However, our knowledge about the precise mechanisms that control NSC function in neuroinflammation is still in its infancy and many avenues remain to be explored. The acute innate proinflammatory signaling cascade strongly suppresses the production and retention of new neurons in the adult brain. Here, the related immune signaling and epigenetic role might involve that must be addressed. We are just at the beginning of understanding the field. There are not reproducible comprehensive profiles of the DNA methylomes and histone modifications of NSCs in proper inflammatory stages that could generate some biomarkers to test in disease-associated conditions. Although more neurodevelopmental diseases caused by mutations in epigenetic genes are being identified, we still do not understand how the disturbance of DNA methylation and histone modification would directly affect NSC fate except the regulation of some neurotrophic factors. Moreover, a clear epigenetic interpretation that control the stimulation of neurogenesis during neuroinflammation, and the integration of NSC in diseased brain could assist to develop novel therapeutic approaches with a potential application in neuroinflammatory diseases.

Acknowledgements

This work was supported by the National Research Foundation of Korea Grant funded by the Korean Government (No.2012-0009212).

Author details

Nando Dulal Das* and Young Gyu Chai

*Address all correspondence to: nando.hu@gmail.com; ygchai@hanyang.ac.kr

Division of Molecular & Life Science, Hanyang University, Gyeonggi-do, Ansan, Korea

The authors declare that they have no conflict of interest.

References

- [1] Barnabe-Heider, F., Wasylnka, J.A., Fernandes, K.J., Porsche, C., Sendtner, M., Kaplan, D.R., and Miller, F.D. (2005). Evidence that embryonic neurons regulate the onset of cortical gliogenesis via cardiotrophin-1. *Neuron* 48, 253-265.
- [2] Belmadani, A., Tran, P.B., Ren, D., and Miller, R.J. (2006). Chemokines regulate the migration of neural progenitors to sites of neuroinflammation. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 26, 3182-3191.
- [3] Berg, A.T, Berkovic, S.F., Brodie, M.J., Buchhalter, J., Cross, J.H., Van Emde, B.W. *et al.* (2010). Revised terminology and concepts for organization of seizures and epilepsies: report of the ILAE commission on classification and terminology, 2005–2009. *Epilepsia* 51, 676–685.
- [4] Beyer, S., Kristensen, M.M., Jensen, K.S., Johansen, J.V., and Staller, P. (2008). The histone demethylases JMJD1A and JMJD2B are transcriptional targets of hypoxia-inducible factor HIF. *The Journal of biological chemistry* 283, 36542-36552.
- [5] Borrelli, E., Nestler, E.J., Allis, C.D., and Sassone-Corsi, P. (2008). Decoding the epigenetic language of neuronal plasticity. *Neuron* 60, 961-974.
- [6] Bulk, E., Sargin, B., Krug, U., Hascher, A., Jun, Y., Knop, M., Kerkhoff, C., Gerke, V., Liersch, R., Mesters, R.M., *et al.* (2009). S100A2 induces metastasis in non-small cell lung cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research* 15, 22-29.
- [7] Butovsky, O., Ziv, Y., Schwartz, A., Landa, G., Talpalar, A.E., Pluchino, S., Martino, G., and Schwartz, M. (2006). Microglia activated by IL-4 or IFN-gamma differentially induce neurogenesis and oligodendrogenesis from adult stem/progenitor cells. *Molecular and cellular neurosciences* 31, 149-160.
- [8] Carpentier, P.A., and Palmer, T.D. (2009). Immune influence on adult neural stem cell regulation and function. *Neuron* 64, 79-92.
- [9] Cloos, P.A., Christensen, J., Agger, K., and Helin, K. (2008). Erasing the methyl mark: histone demethylases at the center of cellular differentiation and disease. *Genes & development* 22, 1115-1140.
- [10] Cloos, P.A., Christensen, J., Agger, K., Maiolica, A., Rappsilber, J., Antal, T., Hansen, K.H., and Helin, K. (2006). The putative oncogene GASC1 demethylates tri- and dimethylated lysine 9 on histone H3. *Nature* 442, 307-311.
- [11] Covacu, R., Danilov, A.I., Rasmussen, B.S., Hallen, K., Moe, M.C., Lobell, A., Johansson, C.B., Svensson, M.A., Olsson, T., and Brundin, L. (2006). Nitric oxide exposure diverts neural stem cell fate from neurogenesis towards astroglialogenesis. *Stem cells* 24, 2792-2800.

- [12] Croft, A.P., and Przyborski, S.A. (2009). Mesenchymal stem cells expressing neural antigens instruct a neurogenic cell fate on neural stem cells. *Experimental neurology* 216, 329-341.
- [13] Das, N.D., Choi, M.R., Jung, K.H., Park, J.H., Lee, H.T., Das, A., Kim, S.H., and Chai, Y.G. (2012). Functional Analysis of Histone Demethylase Jmjd2b on Lipopolysaccharide-Treated Murine Neural Stem Cells (NSCs). *Neurotoxicity research*.
- [14] Glass, C.K., Saijo, K., Winner, B., Marchetto, M.C., and Gage, F.H. (2010). Mechanisms underlying inflammation in neurodegeneration. *Cell* 140, 918-934.
- [15] Hanisch, U.K., and Kettenmann, H. (2007). Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nature neuroscience* 10, 1387-1394.
- [16] Hsieh, J., and Gage, F.H. (2005). Chromatin remodeling in neural development and plasticity. *Current opinion in cell biology* 17, 664-671.
- [17] Hsieh, J., Nakashima, K., Kuwabara, T., Mejia, E., and Gage, F.H. (2004). Histone deacetylase inhibition-mediated neuronal differentiation of multipotent adult neural progenitor cells. *Proceedings of the National Academy of Sciences of the United States of America* 101, 16659-16664.
- [18] Hwang, J-Y., Aromolaran, K.A., Zukin, R.S., (2012). Epigenetic Mechanisms in Stroke and Epilepsy *Neuropsychopharmacology Reviews*, 1–16.
- [19] Iosif, R.E., Ekdahl, C.T., Ahlenius, H., Pronk, C.J., Bonde, S., Kokaia, Z., Jacobsen, S.E., and Lindvall, O. (2006). Tumor necrosis factor receptor 1 is a negative regulator of progenitor proliferation in adult hippocampal neurogenesis. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 26, 9703-9712.
- [20] Jakubs, K., Bonde, S., Iosif, R.E., Ekdahl, C.T., Kokaia, Z., Kokaia, M., and Lindvall, O. (2008). Inflammation regulates functional integration of neurons born in adult brain. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 28, 12477-12488.
- [21] Jelitai, M., Anderova, M., Marko, K., Kekesi, K., Koncz, P., Sykova, E., and Madarasz, E. (2004). Role of gamma-aminobutyric acid in early neuronal development: studies with an embryonic neuroectodermal stem cell clone. *Journal of neuroscience research* 76, 801-811.
- [22] Juliandi, B., Abematsu, M., and Nakashima, K. (2010). Epigenetic regulation in neural stem cell differentiation. *Development, growth & differentiation* 52, 493-504.
- [23] Jung, G.A., Yoon, J.Y., Moon, B.S., Yang, D.H., Kim, H.Y., Lee, S.H., Bryja, V., Arenas, E., and Choi, K.Y. (2008). Valproic acid induces differentiation and inhibition of proliferation in neural progenitor cells via the beta-catenin-Ras-ERK-p21Cip/WAF1 pathway. *BMC cell biology* 9, 66.
- [24] Kelly, L., Grehan, B., Chiesa, A.D., O'Mara, S.M., Downer, E., Sahyoun, G., Massey, K.A., Nicolaou, A., and Lynch, M.A. (2011). The polyunsaturated fatty acids, EPA

and DPA exert a protective effect in the hippocampus of the aged rat. *Neurobiology of aging* 32, 2318 e2311-2315.

- [25] Kishi, N., and Macklis, J.D. (2004). MECP2 is progressively expressed in post-migratory neurons and is involved in neuronal maturation rather than cell fate decisions. *Molecular and cellular neurosciences* 27, 306-321.
- [26] Koo, J.W., and Duman, R.S. (2008). IL-1beta is an essential mediator of the antineurogenic and anhedonic effects of stress. *Proceedings of the National Academy of Sciences of the United States of America* 105, 751-756.
- [27] Lachner, M., Sengupta, R., Schotta, G., and Jenuwein, T. (2004). Trilogies of histone lysine methylation as epigenetic landmarks of the eukaryotic genome. *Cold Spring Harbor symposia on quantitative biology* 69, 209-218.
- [28] Lim, D.A., Huang, Y.C., Swigut, T., Mirick, A.L., Garcia-Verdugo, J.M., Wysocka, J., Ernst, P., and Alvarez-Buylla, A. (2009). Chromatin remodelling factor Mll1 is essential for neurogenesis from postnatal neural stem cells. *Nature* 458, 529-533.
- [29] Liu, S.J., Zukin, R.S. (2007). Ca²⁺-permeable AMPA receptors in synaptic plasticity and neuronal death. *Trends in Neuroscience* 30, 126-134.
- [30] Livingstone, L.R., White, A., Sprouse, J., Livanos, E., Jacks, T., and Tlsty, T.D. (1992). Altered cell cycle arrest and gene amplification potential accompany loss of wild-type p53. *Cell* 70, 923-935.
- [31] Louvi, A., and Artavanis-Tsakonas, S. (2006). Notch signalling in vertebrate neural development. *Nature reviews Neuroscience* 7, 93-102.
- [32] Lundkvist, J., and Lendahl, U. (2001). Notch and the birth of glial cells. *Trends in neurosciences* 24, 492-494.
- [33] Martino, G., and Pluchino, S. (2006). The therapeutic potential of neural stem cells. *Nature reviews Neuroscience* 7, 395-406.
- [34] Mastronardi, F.G., Noor, A., Wood, D.D., Paton, T., and Moscarello, M.A. (2007). Peptidyl argininedeiminase 2 CpG island in multiple sclerosis white matter is hypomethylated. *Journal of neuroscience research* 85, 2006-2016.
- [35] Meaney, M.J., and Ferguson-Smith, A.C. (2010). Epigenetic regulation of the neural transcriptome: the meaning of the marks. *Nature neuroscience* 13, 1313-1318.
- [36] Mogi, M., Harada, M., Narabayashi, H., Inagaki, H., Minami, M., and Nagatsu, T. (1996). Interleukin (IL)-1 beta, IL-2, IL-4, IL-6 and transforming growth factor-alpha levels are elevated in ventricular cerebrospinal fluid in juvenile parkinsonism and Parkinson's disease. *Neuroscience letters* 211, 13-16.
- [37] Monje, M.L., Toda, H., and Palmer, T.D. (2003). Inflammatory blockade restores adult hippocampal neurogenesis. *Science* 302, 1760-1765.

- [38] Nakatomi, H., Kuriu, T., Okabe, S., Yamamoto, S., Hatano, O., Kawahara, N., Tamura, A., Kirino, T., and Nakafuku, M. (2002). Regeneration of hippocampal pyramidal neurons after ischemic brain injury by recruitment of endogenous neural progenitors. *Cell* 110, 429-441.
- [39] Nakayama, K., Nagase, H., Hiratochi, M., Koh, C.S., and Ohkawara, T. (2008). Similar mechanisms regulated by gamma-secretase are involved in both directions of the bi-directional Notch-Delta signaling pathway as well as play a potential role in signaling events involving type 1 transmembrane proteins. *Current stem cell research & therapy* 3, 288-302.
- [40] Namihira, M., Kohyama, J., Semi, K., Sanosaka, T., Deneen, B., Taga, T., and Nakashima, K. (2009). Committed neuronal precursors confer astrocytic potential on residual neural precursor cells. *Developmental cell* 16, 245-255.
- [41] Nan, X., Campoy, F.J., and Bird, A. (1997). MeCP2 is a transcriptional repressor with abundant binding sites in genomic chromatin. *Cell* 88, 471-481.
- [42] Pfau, R., Tzatsos, A., Kampranis, S.C., Serebrennikova, O.B., Bear, S.E., and Tschlis, P.N. (2008). Members of a family of JmjC domain-containing oncoproteins immortalize embryonic fibroblasts via a JmjC domain-dependent process. *Proceedings of the National Academy of Sciences of the United States of America* 105, 1907-1912.
- [43] Pieper, H.C., Evert, B.O., Kaut, O., Riederer, P.F., Waha, A., and Wullner, U. (2008). Different methylation of the TNF-alpha promoter in cortex and substantia nigra: Implications for selective neuronal vulnerability. *Neurobiology of disease* 32, 521-527.
- [44] Reinberg, D., Chuikov, S., Farnham, P., Karachentsev, D., Kirmizis, A., Kuzmichev, A., Margueron, R., Nishioka, K., Preissner, T.S., Sarma, K., *et al.* (2004). Steps toward understanding the inheritance of repressive methyl-lysine marks in histones. *Cold Spring Harbor symposia on quantitative biology* 69, 171-182.
- [45] Rolls, A., Shechter, R., London, A., Ziv, Y., Ronen, A., Levy, R., and Schwartz, M. (2007). Toll-like receptors modulate adult hippocampal neurogenesis. *Nature cell biology* 9, 1081-1088.
- [46] Rolls, A., Shechter, R., and Schwartz, M. (2009). The bright side of the glial scar in CNS repair. *Nature reviews Neuroscience* 10, 235-241.
- [47] Russo, I., Barlati, S., and Bosetti, F. Effects of neuroinflammation on the regenerative capacity of brain stem cells. *J Neurochem* 116, 947-956.
- [48] Russo, I., Barlati, S., and Bosetti, F. (2011). Effects of neuroinflammation on the regenerative capacity of brain stem cells. *Journal of neurochemistry* 116, 947-956.
- [49] Sanz, L.A., Chamberlain, S., Sabourin, J.C., Henckel, A., Magnuson, T., Hugnot, J.P., Feil, R., and Arnaud, P. (2008). A mono-allelic bivalent chromatin domain controls tissue-specific imprinting at Grb10. *The EMBO journal* 27, 2523-2532.

- [50] Setoguchi, H., Namihira, M., Kohyama, J., Asano, H., Sanosaka, T., and Nakashima, K. (2006). Methyl-CpG binding proteins are involved in restricting differentiation plasticity in neurons. *Journal of neuroscience research* 84, 969-979.
- [51] Siegmund, K.D., Connor, C.M., Campan, M., Long, T.I., Weisenberger, D.J., Binisz-kiewicz, D., Jaenisch, R., Laird, P.W., and Akbarian, S. (2007). DNA methylation in the human cerebral cortex is dynamically regulated throughout the life span and involves differentiated neurons. *PloS one* 2, e895.
- [52] Strahl, B.D., and Allis, C.D. (2000). The language of covalent histone modifications. *Nature* 403, 41-45.
- [53] Sun, Y., Jin, K., Childs, J.T., Xie, L., Mao, X.O., and Greenberg, D.A. (2005). Neuronal nitric oxide synthase and ischemia-induced neurogenesis. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism* 25, 485-492.
- [54] Takizawa, T., Nakashima, K., Namihira, M., Ochiai, W., Uemura, A., Yanagisawa, M., Fujita, N., Nakao, M., and Taga, T. (2001). DNA methylation is a critical cell-intrinsic determinant of astrocyte differentiation in the fetal brain. *Developmental cell* 1, 749-758.
- [55] Tsankova, N.M., Kumar, A., Nestler, E.J. (2004). Histone modifications at gene promoter regions in rat hippocampus after acute and chronic electroconvulsive seizures. *Journal of Neuroscience* 24, 5603-5610.
- [56] Tsujimura, K., Abematsu, M., Kohyama, J., Namihira, M., and Nakashima, K. (2009). Neuronal differentiation of neural precursor cells is promoted by the methyl-CpG-binding protein MeCP2. *Experimental neurology* 219, 104-111.
- [57] Urdinguio, R.G., Sanchez-Mut, J.V., and Esteller, M. (2009). Epigenetic mechanisms in neurological diseases: genes, syndromes, and therapies. *Lancet neurology* 8, 1056-1072.
- [58] Waldau, B., Hattiangady, B., Kuruba, R., Shetty, A.K. (2010). Medial ganglionic eminencederived neural stem cell grafts ease spontaneous seizures and restore GDNF expression in a rat model of chronic temporal lobe epilepsy. *Stem Cells* 28, 1153-1164.
- [59] Weaver, I.C., D'Alessio, A.C., Brown, S.E., Hellstrom, I.C., Dymov, S., Sharma, S., Szyf, M., and Meaney, M.J. (2007). The transcription factor nerve growth factor-inducible protein a mediates epigenetic programming: altering epigenetic marks by immediate-early genes. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 27, 1756-1768.
- [60] Wissmann, M., Yin, N., Muller, J.M., Greschik, H., Fodor, B.D., Jenuwein, T., Vogler, C., Schneider, R., Gunther, T., Buettner, R., *et al.* (2007). Cooperative demethylation

by JMJD2C and LSD1 promotes androgen receptor-dependent gene expression. *Nature cell biology* 9, 347-353.

- [61] Yang, J., Jubb, A.M., Pike, L., Buffa, F.M., Turley, H., Baban, D., Leek, R., Gatter, K.C., Ragoussis, J., and Harris, A.L. (2010). The histone demethylase JMJD2B is regulated by estrogen receptor alpha and hypoxia, and is a key mediator of estrogen induced growth. *Cancer research* 70, 6456-6466.
- [62] Yang, J., Ledaki, I., Turley, H., Gatter, K.C., Montero, J.C., Li, J.L., and Harris, A.L. (2009). Role of hypoxia-inducible factors in epigenetic regulation via histone demethylases. *Annals of the New York Academy of Sciences* 1177, 185-197.
- [63] Yoder, J.A., Walsh, C.P., and Bestor, T.H. (1997). Cytosine methylation and the ecology of intragenomic parasites. *Trends in genetics : TIG* 13, 335-340.
- [64] Zhao, X., Ueba, T., Christie, B.R., Barkho, B., McConnell, M.J., Nakashima, K., Lein, E.S., Eadie, B.D., Willhoite, A.R., Muotri, A.R., *et al.* (2003). Mice lacking methyl-CpG binding protein 1 have deficits in adult neurogenesis and hippocampal function. *Proceedings of the National Academy of Sciences of the United States of America* 100, 6777-6782.